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Article

# Temperature but Not Photoperiod Can Predict Development and Survival of An Invasive Apple Pest

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Simple Summary: The apple leafcurling midge is an invasive pest of apple. The pest can pupate on fruit and contaminate fresh fruit for export, causing biosecurity problems to many uninvaded regions such as Australia, California, China, India, Japan, and Taiwan. To generate critical knowledge for effective pest risk analysis, forecast and control of this pest, we tested the effects of five temperatures and five daylengths on its development and survival and developed a thermal model for prediction of the number of generations per year under different climate conditions. We show that temperature but not daylength influenced its development and survival. Although the pest could not complete its lifecycle at 10°C, the high tolerance of its mature larvae and pupae to this temperature might allow it to survive lengthy shipment in cold containers and reach the countries that trade with the midge-infested regions. The lower pupation and emergence rates at ≥ 25°C may reduce the probability of its population build-up in regions where the average maximum temperature during the summer is over 25°C. The model developed in this study can accurately predict the number of generations a year and the time of adult emergence in each generation under different thermal locations.

**Abstract:** The apple leaf-curling midge, *Dasineura mali* Kieffer (Diptera: Cecidomyiidae), is an invasive pest of apple. To provide crucial information for its pest risk analysis, forecast and management, we investigated the effects of temperatures (5, 10, 15, 20 and 25°C) and daylengths (10, 11, 12, 13, 14 and 15 hrs) on its development and survival. The midge could not complete its development at 10°C or lower. After transferred from 20°C to 10°C, mature larvae could survive for about two months and develop to adults. Pupation and emergence rates were significantly higher at 20°C than at 10°C and 25°C. Daylength had no effect on these parameters. The low temperature threshold and thermal requirement to complete development from eggs to adults were 3.7°C and 627 degree-days, respectively. The midge had significantly lower thermal requirement for completion of its lifecycle at 20°C (614.5 degree-days) than at 15°C (650.1 degree-days) and 25°C (634.8 degree-days). The thermal model developed in this study provided accurate predictions of the number of *D. mali* generations and adult emergence time in each generation in different regions of New Zealand. We suggest that the model could be used to predict population dynamics of this pest in other parts of the world.

Keywords: biosecurity; temperature; photoperiod; phenology; pest risk analysis

# 1. Introduction

The apple leaf-curling midge, *Dasineura mali* Kieffer (Diptera: Cecidomyiidae), native to Europe, is a monophagous herbivore that attacks apple trees in Europe, South and North America, and New Zealand [1–5]. Its larvae feed on young leaves and shoots, reducing photosynthesis, resulting in poor shoot development, and potentially affecting the long-term yield [6]. Although the midge does not feed on fruit, its cocooned larvae can contaminate fresh fruit, causing quarantine problems [7,8]. *Dasineura mali* infestations in commercial orchards have raised concerns in the apple industry in New Zealand and some regions that trade with New Zealand, such as Australia, California, China, India, Japan, and Taiwan [9,10].

Dasineura mali adults emerge from the soil in the early morning and swarm around the emergence sites for mating, after which time females lay their eggs on growing apple shoots [11].

Eggs hatch in 3–5 days depending on the temperature and larval feeding on young leaves causes them to roll or produce galls alongside their outer edges; mature larvae ( $3^{rd}$  instar) leave the curled leaves and burrow in the soil ( $\leq 10$  mm in depth) where they spin cocoons and pupate [12] but some can spin cocoons on fruit on their way to the ground [9,13]. The number of generations of *D. mali* per year varies depending on climate conditions and availability of food, for example, there are three generations in Canada [5,14], three to six generations in Europe [15] and four to five generations in New Zealand [12,16–18].

Knowledge on the development, survival, and emergence of an insect species under different environmental conditions is vital to its pest risk analysis, forecast and management. Temperature and daylength are two main environmental factors that influence these biological parameters [19]. The low temperature threshold (temperature below which no measurable development takes place) and thermal constant (the number of degree-days above the lower threshold temperature for completion of development) are the two most important thermal parameters that determine how the temperature affects the developmental rate of ectotherms [5,20] and help predict pest phenology [21,22]. Many studies show that both temperature and daylength affect survival and development of cecidomyiid species [23–35] although daylength does not appear to have any effect on these parameters in several cecidomyiid species [26,29]. To date, little is known about how temperature and daylength influence *D. mali*, making its risk analysis, forecast and control difficult.

To generate critical information for development of measures for effective pest risk analysis, forecast and control of *D. mali*, we carried out experiments under a series of temperature and daylength conditions. We tested how temperature and day length affected the development and survival of this pest. Based on our findings, we then calculated developmental thresholds and thermal requirements and developed a model for prediction of the number of *D. mali* generations per year under different climate conditions in New Zealand. Our findings would help countries that trade with *D. mali*-infested regions perform pest risk analysis and develop forecast and management measures.

# 2. Materials and Methods

# 2.1. Insects

We established a breeding colony in the laboratory from the field-collected mature larvae (3<sup>rd</sup> instar) in a mature organic apple orchard in Plant Growth Unit, Massey University, Palmerston North, New Zealand. The larvae were transferred to the rearing medium (vermiculite) in Petri dishes (5.5 cm in diameter × 1.3 cm in height) and maintained at  $20 \pm 1^{\circ}$ C,  $65 \pm 5\%$  RH and 15:9 h (light:dark) for pupation. Newly emerged adults were released into an aluminium-framed experimental cage (43 × 42 × 40 cm) in which 10 apple seedlings (≈ 20 cm height, bred from rootstock MM 106 FSV) were maintained for oviposition. The cage had a fine metal mesh (aperture size = 0.25 mm) on the back and both sides and Perspex on the top and front and aluminium alloy on the bottom.

#### 2.2. Effect of Temperature on Development and Survival

We investigated the effect of temperature on the development, survival and emergence of D. mali at five constant temperatures: 5, 10, 15, 20 and 25°C with a day length of 15 hrs in the laboratory. For each test temperature, at the onset of photophase we released 30 newly emerged D. mali adults from the above colony with a sex ratio of about 1:1 into an above-mentioned experimental cage for ovipositon, where 10 apple seedlings were maintained as above. We removed seedlings 12 hrs after oviposition, caged them individually using transparent permeable plastic bags (42 cm in length  $\times$  23 cm in width with aperture = 0.25 mm) and maintained them at the test temperature. Because midge adults laid few eggs at 5 and 10°C, we allowed adults to lay eggs on seedlings at 20°C for 12 hrs and then individually caged the oviposited seedlings as above and transferred them to these two temperature conditions.

To determine the egg incubation duration and hatch rate, we made daily examination of two infested shoots at each temperature under a stereomicroscope (Olympus, Japan). The seedlings were

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maintained at the same temperature until larvae became mature (3<sup>rd</sup> instar, orange in colour). We started examining larval maturation by gently opening curled leaves when they turned brown. We counted and removed mature larvae from leaves once a day until all larvae became mature. Egg hatch rate (no. of newly hatched larvae/no. of eggs) and larval survival rate (no. of mature larvae/no. of newly hatched larvae) were calculated. Developmental duration of eggs and immature larvae were recorded.

Mature larvae collected from 10 seedlings under each test temperature were transferred into Petri dishes with the rearing medium as mentioned above and maintained at the same temperature until emergence. There were 18, 20 and 20 dishes with 20, 30 and 25 larvae per dish established at 15, 20 and 25°C, respectively. Because no eggs hatched at 5°C and no larvae completed development at 10°C, we collected mature larvae from 20°C, and immediately transferred them to 10°C in 12 dishes with 25 mature larvae each and maintained them at this temperature until adults emerged. Newly emerged adults were counted and sexed daily according to Gagné and Harris [36]. After all adults had emerged, midge cocoons from each dish were examined under the microscope to determine the pupation rate of larvae (no. of pupae/no. of mature larvae) and emergence rate of adults from the pupae (no. of emerged adults/no. of pupae).

#### 2.3. Effect of Daylength on Development and Survival

In this experiment we tested the effect of day length on the development, survival and emergence of *D. mali* under six daylength conditions: 10, 11, 12, 13, 14 and 15 h light based on a 24-h cycle at 20°C in the laboratory. This experiment was carried out and data were recorded as in the temperature experiment. There were 13, 17, 15, 25, 25 and 20 dishes with 25 mature larvae per dish at above day lengths, respectively.

#### 2.4. Low Temperature Thresholds and Thermal Constants

Based on the developmental time at test temperatures, we used the method of Campbell et al. **[37]** to estimate the low temperature threshold (t) and thermal constant (degree-days, K) for D. mali: r = a + bT, where r is the developmental rate [1/developmental duration (days)], T is the temperature, and a and b are estimates of the r intercept and slope, respectively. The low temperature threshold and thermal constant were calculated as t = -a/b and K = 1/b, respectively. The standard errors of t and

K were calculated as  $SE_t = \frac{\bar{r}}{b} \sqrt{\frac{s^2}{N \times r^2} + \left[\frac{SE(b)}{b}\right]^2}$  and  $SE_k = \frac{SE(b)}{b^2}$ , respectively, where SE(b) is the SE(b) is the SE(b) is the residual mean square of SE(b) is the sample mean, and SE(b) is the sample size. At a given constant temperature, thermal requirement for SE(b) is the sample mean, and SE(b) is the sample size. At a given constant temperature, thermal requirement for SE(b) is the sample mean, and SE(b) is the sample size. At a given constant temperature, thermal requirement for SE(b) is the sample mean, and SE(b) is the sample m

#### 2.5. Estimation of the Number of Generations in the Field

Based on the low temperature threshold and degree-days (see Results), we estimated the number of generations ALCM might have in four regions in New Zealand, Hawke's Bay (latitude 39.6°S), Palmerston North (latitude 40.4°S), Nelson (latitude 41.3°S) and Central Otago (latitude 45.3°S), by developing a degree-day model [21]:  $N = (\frac{1}{K}) \sum_{i=1}^{i=n} (T_i - t)$ , where  $T_i$  is the mean daily air temperature (1981–2010) [38] at day i after the midge emergence from the overwintered generation, and n is the number of days during the growing season. To avoid overestimation or underestimation, we calculated the degree-days since the mean date of the first adult emergence peak, i.e., 30 September (2004–2017) in Hawker's Bay, 02 October (2004–2017) in Nelson and 11 October (2001–2017) in Central Otago [18] and 10 October (2005–2007) in Palmerston North [39]. Hawke's Bay, Nelson and Central Otago are the three main apple growing regions for export in New Zealand [40].

## 2.6. Statistical Analysis

We performed analyses using SAS v. 9.4 software (SAS Institute Inc., NC, USA). Data on the development of different stages at different temperatures and daylengths and the thermal

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requirement (degree-days) were not normally distributed (Shapiro-Wilk test, UNIVARIATE Procedure) and thus analyzed using a generalized linear model (GLMMIX Procedure) followed by a Tukey-Kramer test for multiple comparisons between temperature. The relationship between the developmental rate and temperature was estimated using a general linear model (GLM Procedure). The survival data were analyzed using a generalized linear model (GENMOD Procedure) with a Binomial distribution and *logit* function. Multiple comparisons between temperatures or between day lengths were performed using the Contrast statement.

#### 3. Results

# 3.1. Effect of Temperature on Development and Survival

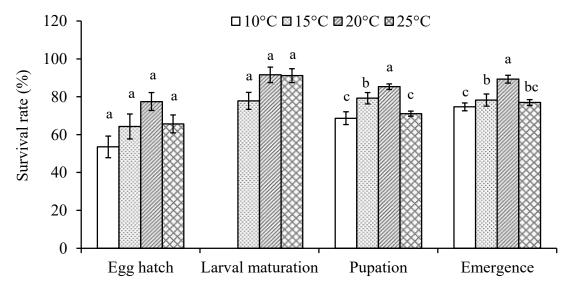
Eggs failed to hatch at 5°C. At 10°C eggs hatched but larvae failed to complete their development. Mature larvae transferred from 20°C to 10°C successfully pupated and developed to adults in about 60 days (**Table 1**). The midge developed significantly faster with the increase of temperature ( $F_{3,127} = 70.72$  for egg,  $F_{2,993} = 379.87$  for larva,  $F_{3,1171} = 3135.28$  for larva-adult,  $F_{2,993} = 1250.65$  for total,  $F_{2,470} = 586.28$  for male and  $F_{2,523} = 664.01$  for female; P < 0.0001) (**Table 1**).

Egg hatch and larval survival rates were higher at 20°C but there was no significant difference between temperatures ( $x_3^2 = 6.59$ , P = 0.0863 for egg hatch rate;  $x_2^2 = 3.49$ , P = 0.1747 for larval survival rates) (**Figure 1**). Significantly more mature larvae pupated and adults emerged at 20°C than at 10 and 25°C ( $x_3^2 = 47.65$  and 33.20 for pupation and emergence rates, respectively; P < 0.0001) (**Figure 1**).

**Table 1.** Mean (± SE) developmental duration (days) of *Dasineura mali* at different temperatures under daylength of 15 h.

	10°Ca	15°C	20°C	25°C
Egg	9.89±0.11a	4.50±0.10b	2.85±0.06c	2.03±0.04d
Larva <sup>b</sup>		21.57±0.02a	15.99±0.02b	11.94±0.01c
Larva-adult <sup>c</sup>	60.01±0.37a	31.41±0.08b	18.64±0.06c	15.77±0.07d
Totald		57.41±0.80a	37.63±0.06b	29.78±0.07c
Maled		57.38±0.11a	37.62±0.09b	29.81±0.10c
Female <sup>d</sup>		57.42±0.12a	37.65±0.09b	29.72±0.10c

Means followed by the same letters in row are not significantly different (P > 0.05). <sup>a</sup> Eggs and mature larvae ( $3^{rd}$  instar) were transferred from  $20^{\circ}$ C. <sup>b</sup> from  $1^{st}$  instar to  $3^{rd}$  instar. <sup>c</sup> from  $3^{rd}$  instar to adult. <sup>d</sup> from egg to adult.



**Figure 1.** Mean ( $\pm$  SE) survival rate of four life stages in *Dasineura mali* at different temperatures. Columns with the same letters for each category are not significantly different (P > 0.05).

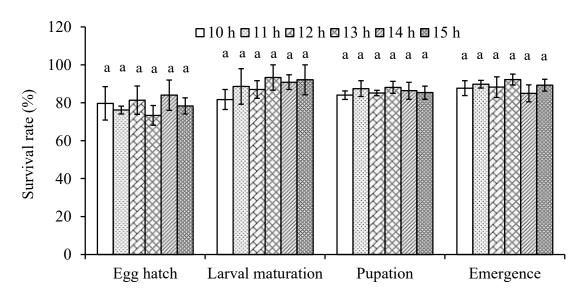
# 3.2. Effect of Daylength on Development and Survival

Our results show that daylength has no significant effect on the development of *D. mali* ( $F_{5,231}$  = 0.53 for egg,  $F_{5,207}$  = 0.18 for larva,  $F_{5,1743}$  = 1.55 for larva-adult,  $F_{5,1743}$  = 0.77 for complete lifecycle,  $F_{5,806}$  = 0.42 for male and  $F_{5,937}$  = 0.59 for female; P > 0.05) (**Table 2**). Similarly, daylength had no effect on the midge survival ( $x_2^2$  = 2.12, 7.18, 4.66 and 5.98 for egg hatch, larval survival, pupation and emergence rates, respectively, P > 0.05) (**Figure 2**).

**Table 2.** Mean (± SE) developmental duration (days) of *Dasineura mali* at different daylengths at 20°C.

	10 h	11 h	12 h	13 h	14 h	15 h
Egg	2.85±0.06 a	2.92±0.04a	2.80±0.06a	2.84±0.06a	2.79±0.06a	2.85±0.06a
Larvaª	15.94±0.0	15.91±0.09	15.89±0.10	15.91±0.10	15.87±0.09	15.97±0.10
	6a	a	a	a	a	a
Larva-	18.83±0.0	18.50±0.06	18.24±0.07	18.73±0.11	19.18±0.08	18.64±0.06
adult <sup>b</sup>	9a	a	a	a	a	a
Total <sup>c</sup>	37.83±0.0	37.49±0.06	37.24±0.07	37.73±0.11	38.18±0.08	37.64±0.06
	9a	a	a	a	a	a
Male <sup>c</sup>	38.00±0.1	35.57±0.09	37.28±0.11	38.21±0.14	38.14±0.11	37.62±0.09
	2a	a	a	a	a	a
Female <sup>c</sup>	37.66±0.1	37.45±0.07	37.21±0.10	37.15±0.14	38.22±0.13	37.65±0.09
	2a	a	a	a	a	a

Means followed by the same letters in row are not significantly different (P > 0.05). <sup>a</sup> from 1<sup>st</sup> instar to 3<sup>rd</sup> instar. <sup>b</sup> from 3<sup>rd</sup> instar to adult. <sup>c</sup> from egg to adult.

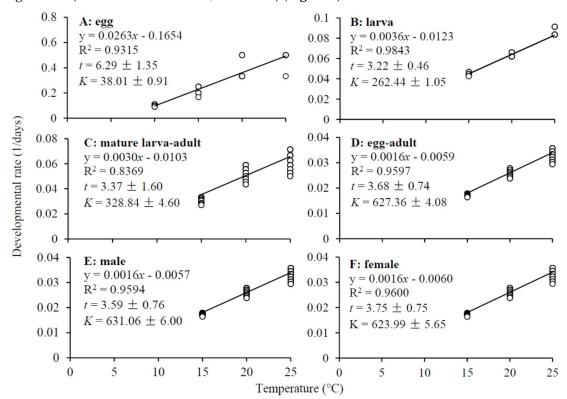


**Figure 2.** Mean ( $\pm$  SE) survival rate of four life stages in *Dasineura mali* at different daylengths ranging from 10 to 15 hours at 20°C. Columns with the same letters for each category are not significantly different (P > 0.05).

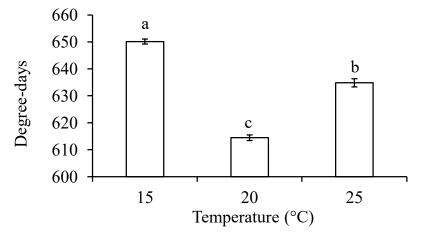
#### 3.3. Low Temperature Thresholds and Thermal Constants

The low temperature threshold was 6.7°C for egg development, which was twice as high as that for larval (3.2°C) and pupal (3.4°C) development (Figure 3A–C). Thermal requirement for *D. mali* developing from eggs to adults was about 630 degree-days with a low temperature threshold of about 3.7°C (Figure 3D–F). Thermal requirement for the midge to complete development from eggs to

adults was significantly different between three test temperatures with the lowest thermal constant being at  $20^{\circ}$ C (ANOVA:  $F_{2,993} = 151.09$ , P < 0.0001) (**Figure 4**).



**Figure 3.** Linear regressions of developmental rate (r) on temperature (T), low temperature threshold (t) and thermal constant (K) for development of *Dasineura mali*. All linear regressions are significant (P < 0.0001).

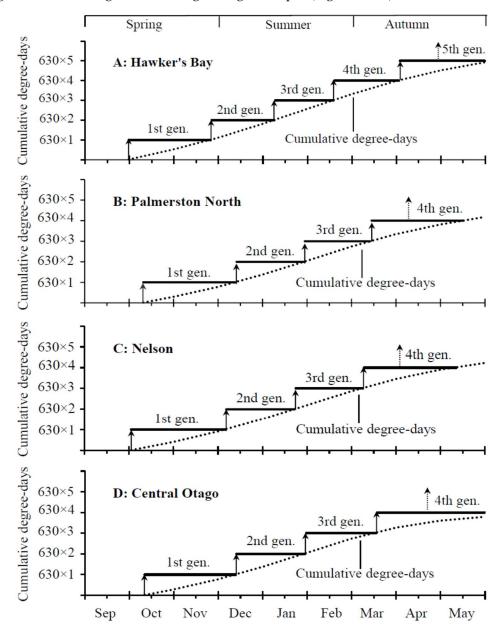


**Figure 4.** Thermal requirement for completion of development from eggs to adults in *Dasineura mali* at different temperatures. Columns ( $\pm$  SE) with the same letters are not significantly different (P > 0.05).

## 3.4. Estimation of the Number of Generations in the Field

Our model predicts that *D. mali* should have five generations a year in Hawker's Bay with four completed generations between October and late March/early May, and mature larvae produced by the 4<sup>th</sup> generation would enter overwintering during late April (**Figure 5**A). However, four generations should occur in Palmerston North, Nelson and Central Otago, with three completed

generations during the growing seasons (October–mid-March) and mature larvae made by adults of the 3<sup>rd</sup> generation entering overwintering during mid-April (Figure 5B–D).



**Figure 5.** Predicted number of generations of *Dasineura mali* in different regions of New Zealand. Arrows with a solid line indicate the time of adult emergence from the overwintered, 1<sup>st</sup>, 2<sup>nd</sup>, 3<sup>rd</sup> and 4<sup>th</sup> generations, and arrows with a dotted line indicate the time of mature larvae entering overwintering.

# 4. Discussion

Our results show that *D. mali* could not complete its lifecycle at  $10^{\circ}$ C but the high tolerance of its mature larvae and pupae to this temperature (**Table 1**) may allow it to survive lengthy shipment in cold containers and reach the countries that trade with midge-infested regions. The midge had significantly higher thermal requirement for completion of its lifecycle at  $25^{\circ}$ C than at  $20^{\circ}$ C (**Figure 4**), suggesting that the temperature  $\geq 25^{\circ}$ C may be less suitable for its development and explaining its summer outbreaks in New Zealand where average maximum temperature rarely exceeds  $25^{\circ}$ C [41]. Furthermore, the significantly lower pupation and emergence rates at  $25^{\circ}$ C (**Figure 1**) or higher [42] may lower the probability of its population build-up in regions where the average maximum temperature during the summer is over  $25^{\circ}$ C. The susceptibility to higher temperatures has also been

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reported in other cecidomyiid species such as the sorghum midge, *Contarinia sorghicola* (Coquillett) **[24]** and blackcurrant leaf midge, *D. tetensi* (Rübsaamen) **[43]**.

The low temperature threshold of 6.7°C for egg development in *D. mali* may explain why its eggs failed to hatch at 5°C. The low temperature thresholds for the development of other *D. mali* stages were about 3.5°C (Figure 3B–D), lower than the mean air temperatures during the winter in New Zealand [38], probably because this midge is native to colder Europe. Using thermal parameters of several midge pests, various authors have developed models for prediction of their population dynamics in the field, providing powerful tools for decision makings in pest management [23,28,43–46]. For example, the accurate prediction of adult emergence time is critical to the applications of pheromone traps and insecticides [47–50].

Based on our findings in the present study (**Figure 3**D) and long-history climate data [38], we have developed a thermal model to predict the number of generations and adult emergence time of each generation in four locations in New Zealand. We show that the model provides an accurate prediction of five generations in Hawker's Bay and four generations in Palmerston North, Nelson, and Central Otago (**Figure 5**), agreeing to the field data reported [16,18,39]. Like other gall-forming insects [51–53], the availability of host plants for *D. mali* is also a crucial factor for the occurrence of the number of generations per year. For example, the lack of food in the autumn [16] may prevent it from having the fifth generation in Palmerston North, Nelson, and Central Otago.

Like some midge species [26,29], our results demonstrate that day length had no effect on *D. mali* development (**Table 2**) and survival (**Figure 2**). Previous studies indicate that the effect of short-day length, if any, on development and survival of midge species is usually associated with low temperatures [29,30,33] or lack of food [16]. Therefore, day length is unlikely to affect these life parameters of *D. mali* during the apple growing season in New Zealand when the temperature is suitable. However, it is still unknown whether combinations of darkness and low temperature during fruit storage and shipment affect development of its mature larvae, which warrants further investigations.

In conclusion, we show that temperature rather than day length significantly affects D. mali development and survival. We suggest that countries or regions with temperatures of  $\geq 25^{\circ}$ C during the growing seasons may be less suitable for D. mali population build-up in apple orchards. The thermal model developed in this study can accurately predict the number of generations a year and the time of adult emergence in each generation in different locations of New Zealand. We recommend that this model be used to predict population dynamics of the pest in other parts of the world. Information generated in the present study can be used for pest risk analysis, forecast and control.

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Data Availability Statement: All data are included in the manuscript.

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Conflicts of Interest: The authors declare no conflict of interest.

#### References

- Marrison, L.G. Apple leaf-curling midge in New Zealand. New Zeal. J. Agri. 1953, 86, 565–566.
- 2. Carl, K. P. Observations on the apple leaf-curling midge, *Dasineura mali* Kieffer and a new species from apple leaf-galls, *Macrolabis sp.* (Dipt. Cecidomyiidae). Anz. Schädlingskund. Pfl., Umweltschutz 1980, 53, 99\_102
- 3. Gagné, R. J. The plant-feeding gall midges of North America. Cornell University Press, Ithaca, New York, 1989, 356 p.
- CABI/EPPO. Dasineura mali. Distribution Maps of Plant Pests No. 711, 2008. https://doi.org/10.1079/DMPP/2008327922 (accessed on 25 February 2023)

- Cossentine, J.; Brauner, A.; Franklin, J.; Robertson, M.; Buhl, P.; Blatt, S.; Gariepy, T. D.; Fraser, H.; Appleby, M.; Grigg-Mcguffin, K.; Mason, P. Parasitism and phenology of *Dasineura mali* (Diptera: Cecidomyiidae) in Canadian apple (Rosaceae) orchards. *Can. Entomol.* 2020, 152, 355–373. https://doi.org/10.4039/tce.2020.15
- 6. Allison, P. A.; Meekings J. A.; Tomkins, R.; Wilson, D. J. Effects of leaf damage by apple leafcurling midge (*Dasyneura mali*) on photosynthesis of apple leaves. Proceedings of the 48<sup>th</sup> New Zealand Plant Protection, Hastings, New Zealand, 8–10 August 1995, pp. 121–124. https://doi.org/10.30843/nzpp.1995.48.11504
- 7. Tomkins, A. R.; Wilson, D. J.; Hutchings, S. O.; June, S. A survey of apple leafcurling midge (*Dasyneura mali*) management in Waikato orchards. Proceedings of the 47th New Zealand Plant Protection, Waitangi, New Zealand, 9–11 August 1994, pp. 346–349. https://doi.org/10.30843/nzpp.1994.47.11052
- 8. Bayer CropScience. Apple leaf curling midge *Dasineura mali*. 2019. https://www.cropscience.bayer.co.nz/pests/insects/apple-leaf-curling-midge (accessed on 25 February 2023)
- 9. Lowe, S. Apple leafcurling midge. Pipmark Technical Bulletin, New Zealand apple and pear marketing board, 1993, 5 p.
- 10. New Zealand Government. Australia measures affecting the importation of apples from New Zealand (WT/DS367). First written submission of New Zealand to WTO, 20 June 2008, Geneva.
- 11. Heath, J. J.; Zhang, A.; Roelofs, W. L.; Smith, R. F. Flight activity and further evidence for a female-produced sex pheromone of the apple leaf midge, *Dasineura mali*, in Nova Scotia. *Northeast. Nat.* 2005, 12, 93–102. https://doi.org/10.1656/1092-6194(2005)012[0093:FAAFEF]2.0.CO;2
- 12. Todd, D. H. A preliminary account of *Dasyneura mali* Kieffer (Cecidomyiidae: Dipt.) and an associated hymenopterous parasite in New Zealand. *New Zeal. J. Sci. Tech. A* 1956, *37*, 462–464.
- 13. Smith, J. T. Aspects of the ecology and management of apple leafcurling midge (*Dasineura mali*) (Diptera: Cecidomyiidae) on the Waimea Plains, Nelson, New Zealand. PhD thesis, Lincoln University, New Zealand, 2000, 192 p.
- 14. Laroche, M.; Provost, C. Suivi des populations de cécidomyie du pommier et méthodes de captures. Rapport final, MAPAQ, Prime-vert, 2015, 13 p.
- 15. Cross, J. V.; Solomon, M. G.; Brabandreier, D.; Blommers, L.; Easterbrook, M. A.; Jay, C. N.; Jenser, G.; Jolly, R. L.; Kuhlmann, U.; Lilley, R.; Olivella, E.; Toepfer, S.; Vidal, S. Biocontrol of pests of apples and pears in Northern and Central Europe: 2. Parasitoids. *Biocont. Sci. Tech.* 1999, 9, 277–314. https://doi.org/10.1080/09583159929569
- 16. Shaw, P. W.; Wallis, D. R.; Alspach, P. A.; Sandanayaka, W. R. M. Phenology of apple leafcurling midge (*Dasineura mali*) in relation to parasitism by *Platygaster demades*. *New Zeal. Plant Prot.* 2005, *58*, 306–310. https://doi.org/10.30843/nzpp.2005.58.4268
- 17. Wearing, C. H.; Marshall, R. R.; Attfield, B.; Colhoun, C. Phenology and distribution of the apple leafcurling midge (*Dasineura mali* (Kieffer)) (Diptera: Cecidomyiidae) and its natural enemies on apples under biological and integrated pest management in Central Otago, New Zealand. *New Zeal. Entomol.* 2013, 36, 87–106. https://doi.org/10.1080/00779962.2012.712887
- 18. Lo, P. L.; Walker, J. T. S. Annual and regional variability in adult *Dasineura mali* (apple leafcurling midge) emergence in New Zealand. *New Zeal. Plant Prot.* 2017, 70, 131–136. https://doi.org/10.30843/nzpp.2017.70.38
- 19. Tauber, M. J.; Tauber, C. A.; Masaki, S. Seasonal adaptations of insects. Oxford University Press, Oxford, 1986, 411 p.
- 20. Higley, L. G.; Pedigo, L. P.; Ostile, K. R. DEGDAY: a program for calculating degree-days, and assumptions behind the degree-day approach. *Environ. Entomol.* 1986, *15*, 999–1016. https://doi.org/10.1093/ee/15.5.999
- 21. Muñiz, M.; Zalom, F. G. Developmental rate and number of generation estimates for *Ceratitis capitata* (Weiedemann) in fruit growing regions of California. *IOBC/wprs Bull*. 1997, 20, 55–66. http://hdl.handle.net/10261/11643
- 22. Rowley, C.; Cherrill, A.; Leather, S. R.; Pope, T. W. Degree-day based phenological forecasting model of saddle gall midge (*Haplodiplosis marginata*) (Diptera: Cecidomyiidae) emergence. *Crop Prot.* 2017, 102, 154–160. https://doi.org/10.1016/j.cropro.2017.08.025
- 23. Baxendale, F. P.; Teetes, G. L.; Sharpe P. J. H. Temperature-dependent model for sorghum midges (Diptera, Cecidomyiidae) spring emergence. *Environ. Entomol.* 1984, 13, 1566–1571. https://doi.org/10.1093/ee/13.6.1566
- 24. Baxendale, F. P.; Teetes, G. L.; Sharpe P. J. H.; Wu, H. Temperature-dependent model for development of nondiapausing sorghum midges (Diptera, Cecidomyiidae). *Environ. Entomol.* 1984, 13, 1572–1576. https://doi.org/10.1093/ee/13.6.1572
- 25. Moore, A. D. Effects of temperature and length of photophase on development and diapause in *Cystiphora schmidti* (Rübsaamen) (Diptera: Cecidomyiidae). *J. Aust. Entomol. Soc.* 1987, 26, 349–354.
- 26. Wellso, S. G. Aestivation and phenology of the Hessian fly (Diptera: Cecidomyiidae) in Indiana. *Environ. Entomol.* 1991, 20, 795–801. https://doi.org/10.1093/ee/20.3.795

- 27. Gillespie, D. R.; Opit, G.; Roitberg, B. Effects of temperature and relative humidity on development, reproduction, and predation in *Feltiella acarisuga* (Vallot) (Diptera: Cecidomyiidae). *Biol. Cont.* 2000, 17, 132–138. https://doi.org/10.1006/bcon.1999.0782
- 28. Hellqvist, S. Phenology of the blackcurrant leaf midge (*Dasineura tetensi*) in northern Sweden. *Acta Agri. Scand.*, *Section B Soil Plant Sci.* 2001, *51*, 84–90. https://doi.org/10.1080/090647101753483804
- 29. Gillespie, D. R.; Quiring, D. M. J. Effects of photoperiod on induction of diapause in *Feltiella acarisuga* (Diptera: Cecidomyiidae). *Can. Entomol.* 2002, 134, 69–75. https://doi.org/10.4039/Ent13469-1
- 30. Wise, I. L.; Lamb, R. J. Diapause and emergence of *Sitodiplosis mosellana* (Diptera: Cecidomyiidae) and its parasitoid *Macroglenes penetrans* (Hymenoptera: Pteromalidae). *Can. Entomol.* 2004, 136, 77–90. https://doi.org/10.4039/n03-032
- 31. Son, Y.; Lee, J. H.; Chung, Y. J. Temperature-dependent post-diapause development and prediction of spring emergence of the pine needle gall midge (Dipt., Cecidomyiidae). *J. Appl. Entomol.* 2007, 131, 674–683. https://doi.org/10.1111/j.1439-0418.2007.01214.x
- 32. Des Marteaux, L. E.; Habash, M. B.; Schmidt, J. M.; Hallett, R. H. A method for induction and quantification of diapause entry in the swede midge (Diptera: Cecidomyiidae). *Can. Entomol.* 2012, 144, 792–800. https://doi.org/10.4039/tce.2012.81
- 33. Yamane, M.; Yano, E.; Matsumoto,Y.; Yoshioka, S.; Kawai, T.; Toyonishi, H.; Nakamura, T. Effect of photoperiod and temperature on the induction of diapause in a Japanese strain of *Aphidoletes aphidimyza* (Diptera: Cecidomyiidae). *Appl. Entomol. Zool.* 2012, 47, 17–26. https://doi.org/10.1007/s13355-011-0084-4
- 34. Yukawa, J.; Ichinose, M.; Kim, W.; Uechi, N.; Gyoutoku, N.; Fujii, T. Lower development threshold temperatures and thermal constants for four species of *Asphondylia* (Diptera: Cecidomyiidae) in Japan and their larval developmental delay caused by heat stress. *Appl. Entomol. Zool.* 2016, *51*, 71–80. https://doi.org/10.1007/s13355-015-0372-5
- 35. Cheng, W.; Long, Z.; Zhang, Y.; Liang, T.; Zhu-Salzman, K. Effects of temperature, soil moisture and photoperiod on diapause termination and post-diapause development of the wheat blossom midge, *Sitodiplosis mosellana* (Géhin) (Diptera: Cecidomyiidae). *J. Insect Physiol.* 2017, 103, 78–85. https://doi.org/10.1016/j.jinsphys.2017.10.001
- 36. Gagné, R. J.; Harris, M. O. The distinction between *dasineura* spp. (Diptera: Cecidomyiidae) from apple and pear. *Proc. Entomol. Soc. Wash.* 1998, 100, 445–448.
- 37. Campbell, A.; Frazier, B. D.; Gilbert, N.; Guitierrez, A. P.; Mackauer, M. Temperature requirements of some aphids and their parasites. *J. Appl. Ecol.* 1974, 11, 431–438. https://doi.org/10.2307/2402197
- 38. NIWA. Mean monthly temperatures (°C) 1981-2010. https://niwa.co.nz/education-and-training/schools/resources/climate/meanairtemp (accessed on 28 February 2023)
- 39. He. X. Z.; Wang, Q. Phenological dynamics of *Dasineura mali* Kieffer (Diptera: Cecidomyiidae) and its parasitoid *Platygaster demades* Walker (Hymenoptera: Platygasteridae) in apple orchards. *J. Econ. Entomol.* 2011, 104, 1640–1646. https://doi.org/10.1603/EC11090
- 40. Anon. Statistical annual 2016. Pipfruit New Zealand Incorporated. 2017, 27 p.
- 41. New Zealand Meteorological Service. Daily climatological observations: monthly reports. Massey University, New Zealand, 2005–2007.
- 42. Sandanayaka, W. R. M.; Ramankutty, P. Temperature dependent emergence and survival of *Platygaster demades* (Hymenoptera: Platygastridae), parasitoid of apple leaf curling midge. *Biol. Cont.* 2007, 42, 41–47. https://doi.org/10.1016/j.biocontrol.2007.04.004
- 43. Cross, J. V.; Crook, D. J. Predicting spring emergence of blackcurrant leaf midge (*Dasineura tetensi*) from air temperatures. *Entomol. Exp. Appl.* 1999, 91, 421–430. https://doi.org/10.1046/j.1570-7458.1999.00510.x
- 44. Gordon, S. C.; Barrie, I. A.; Woodford, J. A. T. Predicting spring oviposition by raspberry cane midge from accumulated derived soil temperatures. *Ann. Appl. Biol.* 1989, 114, 419–427. https://doi.org/10.1111/j.1744-7348.1989.tb03357.x
- 45. Oakley, J. N.; Cumbleton, P. C.; Corbett, S. J.; Saunders, P.; Green, D. I.; Young, J. E. B.; Rodgers, R. Prediction of orange wheat blossom midge activity and risk of damage. *Crop Prot.* 1998, 17, 145–149. https://doi.org/10.1016/S0261-2194(97)00097-5
- 46. Hallett, R. H.; Goodfellow, S. A.; Weiss, R. M.; Olfert, O. MidgEmerge, a new predictive tool, indicates the presence of multiple emergence phenotypes of the overwintered generation of swede midge. *Entomol. Exp. Appl.* 2009, 130, 81–97. https://doi.org/10.1111/j.1570-7458.2008.00793.x
- 47. Burnip, G. M.; Gibb, A. R.; Suckling, D. M. Target and non-target impacts from diazinon applied against *Dasineura mali* in a Canterbury apple orchard. Proceedings of the 51st New Zealand Plant Protection, Hamilton, New Zealand, 11–13 August 1998, pp. 143–147. https://doi.org/10.30843/nzpp.1998.51.11676
- 48. Walker, J. T. S.; Wearing, C. H.; Bradley, S. J.; Shaw, P. W.; Burnip, G. M.; Tokins, A. R.; Richardson, C. A.; Hodson, A. J. Integrated fruit production (IFP) for New Zealand pipfruit: pest management recommendations. Proceedings of the 51st New Zealand Plant Protection, Hamilton, New Zealand, 11–13 August 1998, pp. 166–172. https://doi.org/10.30843/nzpp.1998.51.11639

- 11
- 49. Cross, J. V.; Hall, D. R.; Shaw, P.; Anfora, G. Exploitation of the sex pheromone of apple leaf midge *Dasineura mali* Kieffer (Diptera: Cecidomyiidae): Part 2. Use of sex pheromone traps for pest monitoring. *Crop Prot*. 2009, 28, 128–133. https://doi.org/10.1016/j.cropro.2008.09.004
- 50. Jacquemin, G.; Chavalle, S.; De Proft, M. Forecasting the emergence of the adult orange wheat blossom midge, *Sitodiplosis mosellana* (Géhin) (Diptera: Cecidomyiidae) in Belgium. *Crop Prot.* 2014, 58, 6–13. https://doi.org/10.1016/j.cropro.2013.12.021
- 51. Price, P. W.; Roininen, H.; Tahvanainen, J. Plant age and attack by the bud galler, *Euura mucronata*. *Oecologia*. 1987, 73, 334–337. https://doi.org/10.1007/BF00385248
- 52. Yukawa, J. Synchronization of gallers with host plant phenology. *Popul. Ecol.* 2000, 42, 105–113. https://doi.org/10.1007/PL00011989
- 53. Yukawa, J.; Akimoto, K. Influence of synchronization between adult emergence and host plant phenology on the population density of *Pseudasphondylia neolitseae* (Diptera: Cecidomyiidae) inducing leaf galls on *Neolitsea sericea* (Lauraceae). *Popul. Ecol.* 2006, 48, 13–21. https://doi.org/10.1007/s10144-005-0233-0

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