

Article

Not peer-reviewed version

---

# Suppressive Effects of Volatile Compounds from *Bacillus* spp. on *Magnaporthe oryzae* *Triticum* (MoT) Pathotype, Causal Agent of Wheat Blast

---

[Musrat Zahan Surovy](#) <sup>\*</sup>, [Shahinoor Rahman](#), [Michael Rostás](#), [Tofazzal Islam](#), [Andreas Von Tiedemann](#) <sup>\*</sup>

Posted Date: 18 April 2023

doi: 10.20944/preprints202304.0468.v1

Keywords: Bacillus; biocontrol; volatile organic compound; GC-MS; sporulation



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

## Article

# Suppressive effects of volatile compounds from *Bacillus* spp. on *Magnaporthe oryzae* *Triticum* (MoT) pathotype, causal agent of wheat blast

Musrat Zahan Surovy <sup>1,2\*</sup>, Shahinoor Rahman <sup>3</sup>, Michael Rostás <sup>3</sup>, Tofazzal Islam <sup>2</sup>  
and Andreas von Tiedemann <sup>1\*</sup>

<sup>1</sup> Division of Plant Pathology and Crop Protection, Department of Crop Sciences, Georg-August-University of Goettingen, Grisebachstrasse 6, 37077 Goettingen, Germany; msurovy@gwdg.de and atiedem@gwdg.de

<sup>2</sup> Institute of Biotechnology and Genetic Engineering (IBGE), Bangabandhu Sheikh Mujibur Rahman Agricultural University, Salna, Gazipur 1706, Bangladesh; msurovy@gwdg.de and tofazzalislam@bsmrau.edu.bd

<sup>3</sup> Division of Agricultural Entomology, Department of Crop Sciences, Georg-August-University of Goettingen, Grisebachstrasse 6, 37077 Goettingen, Germany; shahinoor\_ent@yahoo.com and michael.rostas@uni-goettingen.de

\* Correspondence: M.Z.S. (msurovy@gwdg.de) and A.v. T. (atiedem@gwdg.de)

**Abstract:** *Magnaporthe oryzae Triticum* (MoT) pathotype is the causal agent of wheat blast, which has caused significant economic losses and threatens wheat production in South America, Asia and Africa. Three bacterial strains from rice and wheat seeds (*B. subtilis* BTS-3, *B. velezensis* BTS-4, and *B. velezensis* BTLK6A) were used to explore the antifungal effects of volatile organic compounds (VOCs) of *Bacillus* spp. as a potential biocontrol mechanism against MoT. All bacterial treatments significantly inhibited both mycelial growth and sporulation of MoT *in vitro*. We found that this inhibition was caused by *Bacillus* VOCs in a dose-dependent manner. In addition, biocontrol assays using detached wheat leaves infected with MoT showed reduced leaf lesions and sporulation compared to the untreated control. VOCs from *B. velezensis* BTS-4 alone or a consortium (mixture of *B. subtilis* BTS-3, *B. velezensis* BTS-4, and *B. velezensis* BTLK6A) treatments consistently suppressed MoT *in vitro* and *in vivo*. Compared to the untreated control, VOCs from BTS-4 and *Bacillus* consortium reduced MoT lesions *in vivo* by 85% and 81.25%, respectively. A total of thirty-nine VOCs (from 9 different VOC groups) from 4 *Bacillus* treatments were identified by gas chromatography-mass spectrometry (GC-MS), of which 11 were produced in all *Bacillus* treatments. Alcohols, fatty acids, ketones, aldehydes, and S-containing compounds were detected in all four bacterial treatments. *In vitro* assays using pure VOCs revealed that hexanoic acid, 2-methylbutanoic acid, and phenylethyl alcohol are potential VOCs emitted by *Bacillus* spp. that are suppressive for MoT. The minimum inhibitory concentrations for MoT sporulation were 250 mM for phenylethyl alcohol and 500 mM for 2-methylbutanoic acid and hexanoic acid. Therefore, our results indicate that VOCs from *Bacillus* spp. are effective compounds to suppress the growth and sporulation of MoT. Understanding the MoT sporulation reduction mechanisms by *Bacillus* VOCs may provide novel options to manage the further spread of wheat blast by spores.

**Keywords:** *Bacillus*; biocontrol; volatile organic compound; GC-MS; sporulation

## 1. Introduction

Wheat is a major cereal crop worldwide [1], and according to the United States Department of Agriculture (USDA), 779.03 million tons of wheat were produced globally in 2021 [2]. Wheat blast is a devastating fungal disease which is caused by *Magnaporthe oryzae Triticum* (MoT) pathotype [3–5]. It can cause significant reductions in wheat yield and grain quality [6,7]. Wheat blast first emerged in Brazil in 1985 and then gradually spread to Argentina, Bolivia and Paraguay [5,6,8]. Outbreaks of wheat blast in Bangladesh and Zambia in recent years confirmed its expansive potential [9,10]. Recently, it has also been recognized as a potential future threat to European wheat production [11].

Fungicides are commonly used to control blast and other ear-related wheat diseases [12]. However, the failure of fungicides to fully control MoT has led to repetitive applications, increasing the selection pressure on the pathogen to develop resistance [13,14]. The emergence of new fungicide-resistant strains of MoT severely threatens wheat production [7,15], which is why the use of resistant cultivars is an important strategy to control wheat blast. However, MoT can break cultivar resistance, and so far, no cultivars are entirely resistant to MoT [5,16].

Biological control is a potential alternative to manage plant pathogens [17–19]. Over the past three decades, multidisciplinary research in biocontrol has investigated effective strategies to control wheat pathogens [20]. The mechanisms exerted by biological control agents (BCAs) include direct (hyperparasitism, antibiosis) and indirect (competition, induction of host plant resistance) modes of action [19,21–23]. Direct suppression of target fungi by BCAs can be achieved by the production of antibiotics, volatile organic compounds (VOCs), or other bioactive secondary metabolites [21] that have an antagonistic effect on pathogens [19]. Inhibitory activity and biocontrol potential of some pure microbial non-volatile secondary metabolites against the wheat blast fungus MoT have been reported recently [24–27].

Bacterial VOCs are low molecular weight (<300 Da) [28,29], hydrophobic, low toxicity, and naturally-occurring substances [30]. They are diffusible in the environment and have been shown to be effective in biofumigation [31,32]. Recent discoveries of antifungal bacterial volatiles has triggered research interests to explore the potential use of VOCs in the control of fungal pathogens. The efficiency of bacterial VOCs depends on the adaptability of bacteria to diverse environmental conditions, nutritional properties, and bacterial colonization patterns in specific hosts [21,33]. To date, relatively few bacterial biocontrol agents have been employed in agriculture due to their low field efficacy, safety concerns, or issues related to international market trading [34]. VOCs from *Pseudomonas* [30,35], *Bacillus* [36,37], *Burkholderia* [38,39], and *Serratia* [40] have been reported for their active antifungal activity. VOCs from *Trichoderma* fungi [41] are effective against wheat crown and root rot, and those from *Bacillus* spp. against wheat fusarium crown and root rot [42] and fusarium head blight [43].

The potential antagonistic activity of *B. subtilis* BTS-3, *B. velezensis* BTS-4, and *B. velezensis* BTLK6A against MoT was evaluated previously through *in vitro* and *in vivo* screening. Those studies decoded the bacterial genes responsible for antagonism and induced systemic resistance (ISR) in host plants by whole genome sequencing [44,45]. Additionally, the genes responsible for acetoin, a volatile organic compound, was confirmed in the tested *Bacillus* spp. However, it has been reported that *Bacillus* spp. can produce diversified antifungal volatile organic compounds (VOCs) to suppress phytopathogenic growth [36,37,42,43]. So, it is essential to identify all potential VOCs from our selected *Bacillus* spp. and their mechanisms of action to suppress the growth of MoT. Therefore, to further understand the mechanisms underlying the antagonistic action of these bacterial VOCs, this study investigates the role of *Bacillus* VOCs in suppression of MoT without direct contact. More specifically, the objectives of the current study were to – (i) investigate the effects of *Bacillus* VOCs on mycelial growth and sporulation of MoT *in vitro*; (ii) demonstrate the suppressive effects of *Bacillus* volatiles against wheat leaf infection with MoT; (iii) identify and characterize *Bacillus* VOCs through GC-MS; and (iv) confirm suppressive effects of selected pure VOCs against MoT.

## 2. Materials and Methods

### 2.1. Bacterial strains and culture conditions

Three bacterial strains - *B. subtilis* BTS-3 (NCBI accession WOVJ000000000), *B. velezensis* BTS-4 (NCBI accession WOVK000000000) and *B. velezensis* BTLK6A (NCBI accession WOYD000000000) - were used in this study [23]. BTS-3 and BTS-4 bacterial strains were isolated from the 'Rangabinni' rice seeds, and BTLK6A from the 'Kanchan' wheat seeds of Bangladesh [23,44,46,47]. All *Bacillus* strains were stored as pure cultures in 20% glycerol at -20°C. Bacterial strains were streaked into Petri dishes (90 mm) containing ca. 20 ml Luria broth agar, LBA (10 g tryptone, 5 g yeast extract, 10 g NaCl, 15 g agar, 1000 ml H<sub>2</sub>O) and incubated for 24–48 h at 25°C. Then, three single colonies were inoculated into

a 50 ml Erlenmeyer flask containing 25 ml LB (10 g tryptone, 10 g NaCl, 5 g yeast extract, 1000 ml H<sub>2</sub>O) and incubated on a rotary shaker (100 rpm) for 24-48 h at 25°C. After incubation, the bacterial cultures were transferred to 2 ml Eppendorf tubes and centrifuged for 10 min (13,000 rpm). The supernatant was discarded and washed (3 times) with sterilized distilled water (SDW). The bacterial densities were then adjusted ( $1 \times 10^9$ ,  $1 \times 10^8$ , and  $1 \times 10^7$  CFU/ml) for further use and stored in 20% glycerol at -80°C for long-term preservation.

## 2.2. Fungal strain and culture condition

MoT fungal pathogen BTGP 6(f) was isolated from blast-infected wheat ear [48] and grown on V8 agar (V8A) following the protocol described by Surovy *et al.* [49]. The conidial suspension was prepared from 7 d old MoT cultures by adding 0.01% sterile Tween 20 solution (10 ml) per plate. The suspension was filtered through two-layer cheesecloth, and conidial density was adjusted ( $1 \times 10^5$  conidia/ml) using a hemocytometer (Fuchs-Rosenthal, 0.0625 mm<sup>2</sup>).

## 2.3. Volatile assays

### 2.3.1. Bi-partitioned Petri dish assay

Bi-partitioned Petri dishes (90 mm diameter) were used to assess the potential of *Bacillus* VOCs against MoT. LBA media (10 ml) was poured into one side, and 10 ml of V8A was poured into the other side of the Petri dishes. The bacterial suspension (100 µl) was pipetted in LBA, spread with a glass spreader, and incubated for 24 h at 25°C. Three different bacterial densities ( $1 \times 10^9$ ,  $1 \times 10^8$ , and  $1 \times 10^7$  CFU/ml) were used for this experiment. Twenty-four hours after bacterial incubation, a 2 mm 7 d old MoT mycelial plug was placed on the side containing V8A. The Petri dishes were tightly closed with parafilm to avoid evaporation of bacterial VOCs and incubated at the same conditions described earlier (see above 2.2) for 5 d. Mycelial radial growth (mm) of MoT was recorded 5 d after incubation. Subsequently, 10 ml of sterilized 0.01% Tween 20 was added per plate and MoT conidia were dislodged from mycelia using a paint brush (da Vinci, Germany; size 3/0). The conidial suspension was filtered through a two-layer cheesecloth, and conidia were counted (conidia/plate) with a hemocytometer (Fuchs-Rosenthal, 0.0625 mm<sup>2</sup>). Six replications were maintained in each experiment, and three repetitive experiments were performed.

### 2.3.2. Upside-down Petri dish assay

Bacterial strains at different densities ( $1 \times 10^9$ ,  $1 \times 10^8$ , and  $1 \times 10^7$  CFU/ml) were grown in Petri dishes containing LBA for 24 h at 25°C. Twenty-four hours after bacterial incubation, 10 µl ( $1 \times 10^5$  conidia/ml) of MoT conidial suspension was drop inoculated in another Petri dish containing V8A. These two plates, one containing bacteria and one MoT, were placed face-to-face on top each other, tightly sealed with parafilm to avoid the loss of VOCs, and incubated for 5 d at 25°C. Mycelial radial growth of MoT (mm) and the total number of MoT conidia/plate were recorded as described in 2.3.1. Six replications were maintained in each experiment, and three repetitive experiments were performed.

## 2.4. Detached leaf assay

Wheat cultivar BR 18 was used for the detached leaf assay. The seeds were surface sterilized with sodium hypochlorite (3% NaOCl) for 1 min and subsequently washed (3 times) with sterilized distilled water (SDW). Treated seeds were placed in Petri dishes containing moistened filter paper. After germination, they were then sown in plastic pots (7×7×8 cm; 10 seeds per pot) containing a mixture of sand, compost, and peat (1:2:1). Plants were grown in a greenhouse maintaining a 14/10 h light-dark cycle, 25 °C (±2) temperature and 65-70% relative humidity. At growth stage 13 (GS 13, three leaves emerged), the second leaf was cut into small pieces (ca. 2 cm) and surface sterilized with 3% NaOCl. The extra water from the surface-sterilized leaves was removed with a sterile paper towel. Leaf pieces were then placed on water agar (15 g agar, 1,000 ml H<sub>2</sub>O) containing benzimidazole (30



mg/l). Ten leaf pieces were placed in each Petri dish. MoT conidial suspension ( $1 \times 10^5$  conidia/ml) was drop-inoculated (10  $\mu$ l) on each leaf piece. The bacterial suspension (100  $\mu$ l,  $1 \times 10^9$  CFU/ml) was incubated in LBA for 24 h before the preparation of leaf pieces. After incubation, freshly grown (at 25°C) bacterial culture plates were placed open and upside-down on the Petri dishes containing leaf pieces. Plates were sealed tightly with parafilm to avoid the loss of VOCs. Five days after incubation, the lesion growth and total number of conidia in each lesion were recorded. The lesion size (cm<sup>2</sup>) was determined by using ImageJ software (version 1.53m). A single leaf section was placed in a 2 ml Eppendorf tube containing 1 ml water, briefly vortexed, and MoT conidia per lesion were counted using a hemocytometer (Fuchs-Rosenthal, 0.0625 mm<sup>2</sup>). Thirty leaf pieces were used for each bacterial treatment, and three repetitive experiments were performed.

### 2.5. Identification and quantification of *Bacillus volatiles*

*Bacillus* VOCs collection was performed as described previously by Sarenqimuge *et al.* [50]. As an internal standard, 200 ng of tetralin (1,2,3,4 tetrahydronaphthalene, Sigma-Aldrich, Germany) was added to each sample before GC-MS analysis. An aliquot of 30  $\mu$ l sample was transferred to another GC vial with a glass insert and placed into the tray of the GC-MS autosampler. A 2  $\mu$ l sample was injected in a pulsed splitless mode for analysis. The oven temperature was retained at 40°C for 3 min and gradually increased (8°C/min) to a final temperature of 220°C for 10 min. Helium was used as a carrier gas (flow rate was 1.5 ml/min). A homogenous series of n-Alkenes (C<sub>7-20</sub>) was used to determine retention indices. The MassHunter instrument (Agilent Technologies: GC 7890B, MS 5977B) was used for data processing; MSD ChemStation software with the NIST17 and Willey11 mass spectral libraries was used to tentatively identify bacterial VOCs by their mass spectra and retention indices. The identities of the ten bioactive compounds tested in 2.6 were confirmed by GC-MS analysis of commercially available standards. The VOCs quantification was done by comparing the peak area of individual compounds to the peak area of internal standard (tetralin). From each treatment, five replicates were analyzed, and LB without bacteria was used as a control.

### 2.6. Bioassay with pure volatile compounds

Pure VOCs (Supplementary table S4) were tested against MoT at four different concentrations (5M, 1M, 500 mM, and 250 mM), with DMSO as a diluent. Five sterilized paper discs were glued (Tesa stick, tesa SE, Hugo-Kirchberg-Str.1, D-22848 Norderstedt) on the Petri dish lid, and 20  $\mu$ l of each pure compound was pipetted on each paper disc (total 100  $\mu$ l per Petri dish). A 2 mm MoT mycelial block was placed in the center of a V8A plate and the two plates were sealed tightly to avoid the loss of VOCs. Five days after incubation, mycelial radial growth of MoT and the total number of conidia per plate were recorded as described in 2.3.1.

### 2.7. Statistical analysis

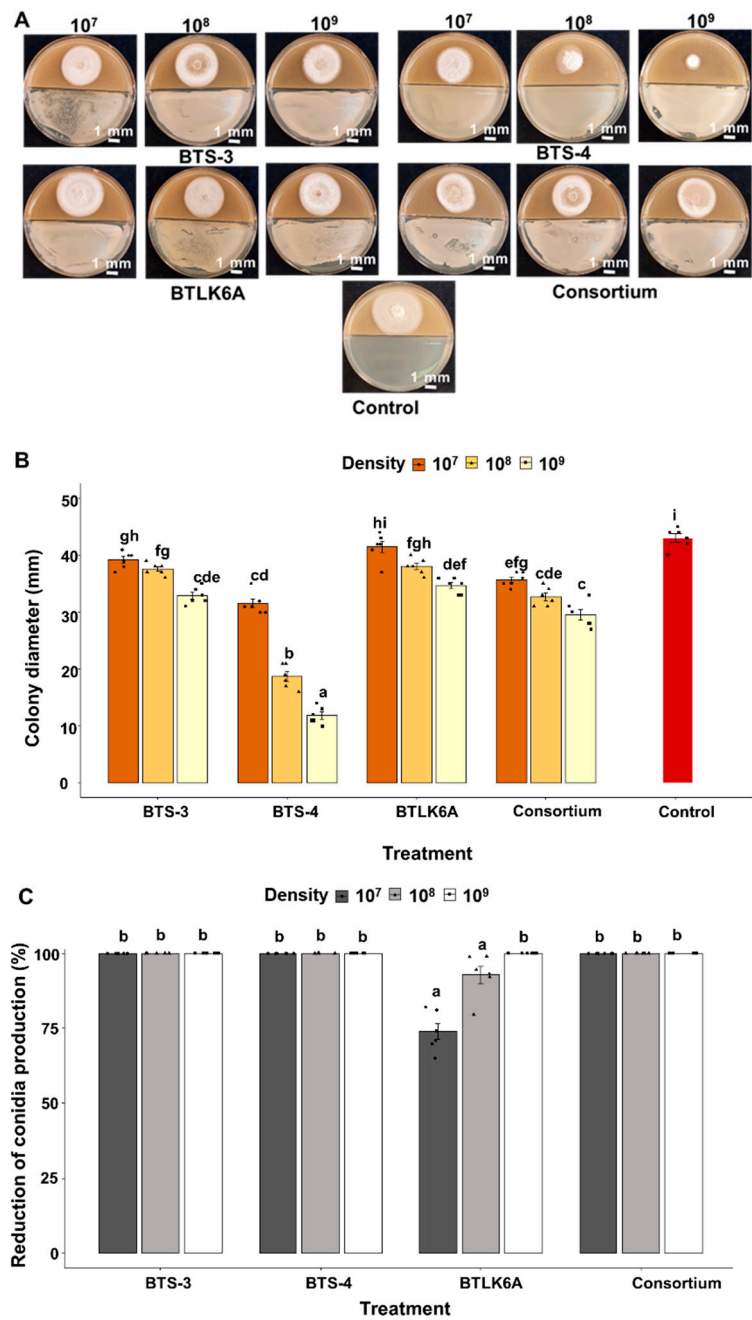
All data were analyzed by using linear models (LMs) in R software (version 4.0.5, accessed 31 March 2021) integrated into R studio (version 1.2.5001, accessed 31 March 2021). The function 'test dispersion' and stimulatedResiduals' of "DHARMA" package were used to test the dispersion and residuals of the models. These functions of "DHARMA" package uses a simulation-based method to create readily interpretable scaled residuals for fitted linear model. Analysis of variance (ANOVA) was calculated for normally distributed data followed by Tukey multiple comparisons ( $p < 0.05$ ) by using the "emmeans" package. For non-normally distributed data sets, Kruskal-Wallis test was performed by using "kruskal.test" function followed by Dunn multiple comparison analyses by using "FSA" and "rcompanion" packages ( $p < 0.05$ ). The "ggplot2" package was used to visualize bar graphs, and "ggVennDiagram" function was used to plot the number of VOCs produced in different *Bacillus* treatments in a Venn diagram. The "ComplexHeatmap" function was used to visualize the *Bacillus* VOCs profiles, the effects of pure VOCs on MoT mycelial growth, and MoT sporulation in a Heatmap. MetaboAnalyst 5.0 [51] was used for volcano plot analysis.

## 3. Results

3.1. Bioassay with volatiles

3.1.1. Effects of Bacillus VOCs on MoT growth

Three different bacterial densities ( $1 \times 10^7$ ,  $1 \times 10^8$ , and  $1 \times 10^9$  CFU/ml) were used in a bi-partitioned Petri dish assay to evaluate the effects of *Bacillus* VOCs on MoT mycelial growth. Different bacterial VOCs significantly inhibited MoT mycelial growth in a density-dependent manner ( $F = 161.44$ ,  $p < 0.001$ ) (Figure 1 A). Compared to the control (43 mm), the highest reduction (72.5%) in mycelial growth was recorded in BTS-4 ( $1 \times 10^9$  CFU/ml, 11.8 mm diameter), and the lowest (3%) in BTLK6A ( $1 \times 10^7$  CFU/ml and 41.5 mm diameter). Inhibition of MoT mycelial growth was consistently higher at the bacterial density of  $1 \times 10^9$  CFU/ml for all four *Bacillus* treatments (Figure 1 A, B).



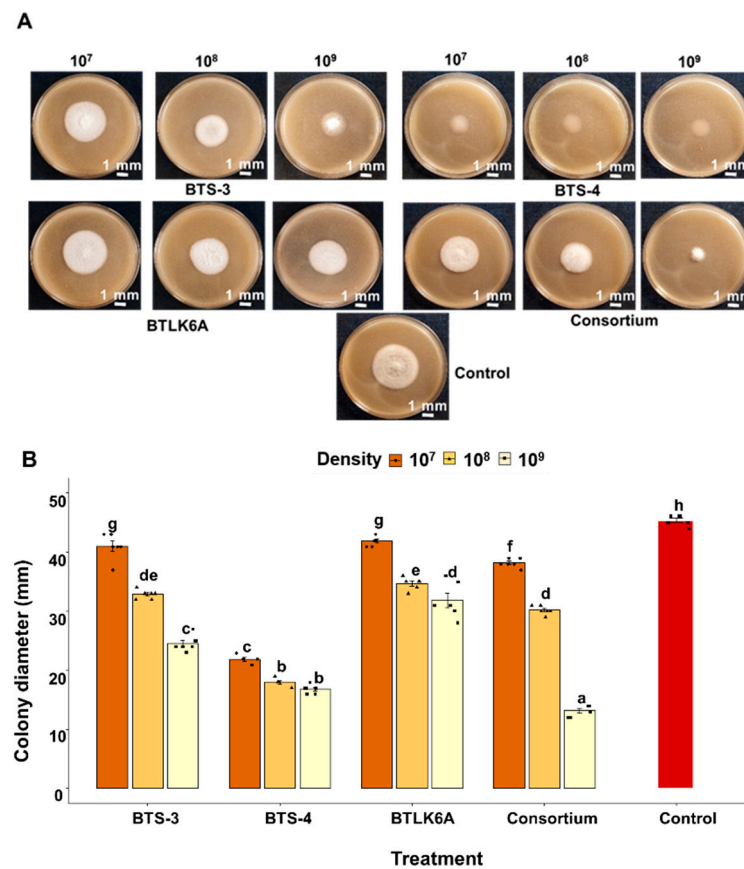
**Figure 1.** Effect of *Bacillus* spp. VOCs on MoT in bi-partitioned Petri dish assay *in vitro*. (A) Photographs showing the effects of *Bacillus* VOCs on mycelial growth of MoT; (B) Effects of *Bacillus* VOCs on mycelial growth of MoT *in vitro*; (C) Effects of *Bacillus* VOCs on reduction of MoT sporulation compared to control *in vitro*. Data were recorded after 5 d of MoT incubation at 25°C. BTS-

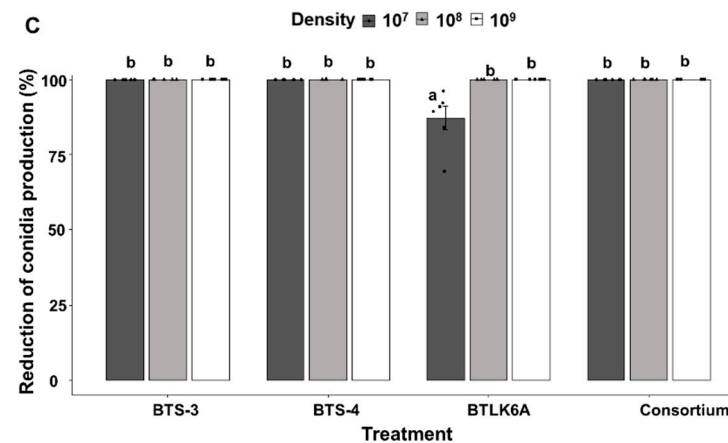
3: *B. subtilis* BTS-3; BTS-4: *B. velezensis* BTS-4; BTLK6A: *B. velezensis* BTLK6A; consortium: a mixture of BTS-3, BTS-4 and BTLK6A; control: without any bacterial treatment (Tukey test;  $n = 6$ ;  $p \leq 0.05$ ). Black dots in (B) and (C) represent data points for each replicate.

In addition to the assessment of MoT mycelial growth, the sporulation rate was evaluated after 5 d of bacterial treatments. Treatments with *Bacillus* VOCs significantly suppressed MoT sporulation ( $F = 23.092$ ,  $p \leq 0.001$ ). All three densities ( $1 \times 10^7$ ,  $1 \times 10^8$ , and  $1 \times 10^9$  CFU/ml) of BTS-3, BTS-4, and the *Bacillus* consortium treatments produced non-spore forming white mycelia (100% reduction of conidia compared to control). In case of BTLK6A, no sporulation was observed at  $1 \times 10^9$  CFU/ml. However, at  $1 \times 10^7$  and  $1 \times 10^8$  CFU/ml densities, black/grey-colored MoT sporulating colonies were observed as in the untreated control. However, the number of conidia was comparatively less in  $1 \times 10^7$  ( $2.37 \times 10^5$  conidia/plate, 73% reduction compared to control) and  $1 \times 10^8$  CFU/ml ( $6.60 \times 10^4$  conidia/plate, 92% reduction compared to control) of BTLK6A treatments compared to the control ( $9.10 \times 10^5$  conidia/plate) (Figure 1 C).

### 3.1.2. Effects of *Bacillus* spp. VOCs against germination of MoT conidia

An upside-down Petri dish assay was performed to evaluate the effect of *Bacillus* VOCs on MoT conidia germination. All MoT conidia germinated, and mycelial growth ensued after exposure to *Bacillus* VOCs. However, MoT mycelial growth was very slow in the *Bacillus* consortium treatment ( $1 \times 10^9$  CFU/ml). Additionally, less intense, flat mycelial growth was observed with all BTS-4 treatments (Figure 2 A). Similar to the bi-partitioned Petri dish assay, the bacterial VOCs also significantly inhibited MoT mycelial growth (developed from MoT conidia) ( $F = 372.63$ ,  $p \leq 0.001$ ).





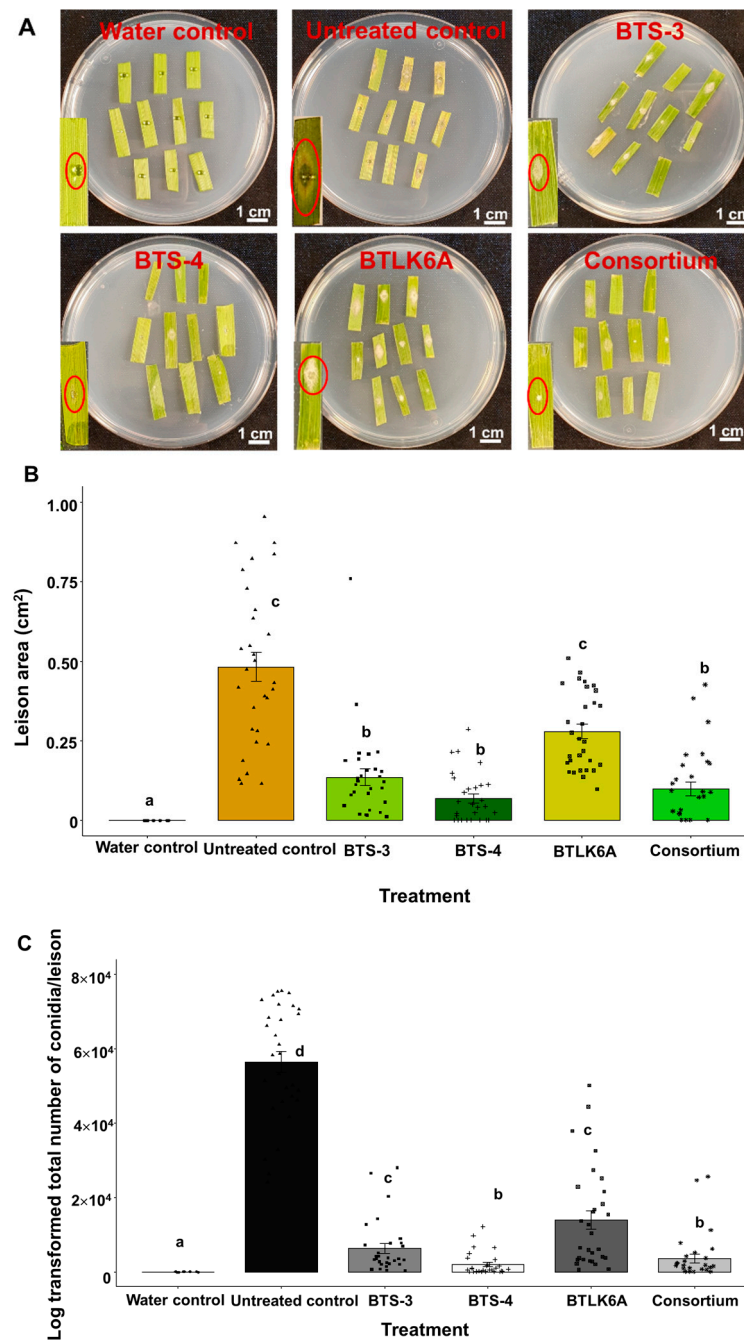
**Figure 2.** Effects of *Bacillus* spp. VOCs on MoT conidial germination in upside-down Petri dish assay *in vitro*. (A) Photographs showing the effects of *Bacillus* VOCs on MoT conidial germination and mycelial growth; (B) Effects of bacterial VOCs on mycelial growth of MoT *in vitro*; (C) Effects of bacterial VOCs on reduction of new MoT sporulation compared to control *in vitro*. Data were recorded after 5 d of MoT incubation at 25°C. BTS-3: *B. subtilis* BTS-3; BTS-4: *B. velezensis* BTS-4; BTLK6A: *B. velezensis* BTLK6A; consortium: a mixture of BTS-3, BTS-4 and BTLK6A; control: without any bacterial treatment (Tukey test;  $n = 6$ ;  $p \leq 0.05$ ). Black dots in (B) and (C) represent data points for each replicate.

Mycelial growth reduction was higher in the upside-down Petri dish assay than bi-partitioned Petri dish assay. The highest inhibition of mycelial growth was recorded with the treatment of *Bacillus* consortium ( $1 \times 10^9$  CFU/ml), with radial growth of 13.2 mm, followed by BTS-4 ( $1 \times 10^9$  CFU/ml, 16.8 mm). The lowest reduction was documented for BTLK6A ( $1 \times 10^7$  CFU/ml, 42.9 mm) (Figure 2 B). The highest sporulation was recorded for the control ( $9.73 \times 10^5$  conidia/plate). However, complete suppression of sporulation of MoT was recorded for all bacterial treatments except for BTLK6A ( $1.23 \times 10^5$  conidia/plate at  $1 \times 10^7$  CFU/ml, 87% reduction of sporulation compared to control) (Figure 2 C).

### 3.2. Effects of *Bacillus* VOCs in detached leaf assay

To investigate the capacity of *Bacillus* VOCs to reduce leaf infection with MoT, a detached leaf assay was performed using four different *Bacillus* VOC treatments (Figure 3 A). In our experiment, we found that *Bacillus* VOCs significantly reduced the development of blast disease symptoms in detached leaves and suppressed MoT sporulation under laboratory conditions, but with varying effects. *Bacillus* VOCs significantly reduced the lesion size ( $F = 37.14$ ,  $p \leq 0.001$ ) and MoT conidia production ( $F = 28.49$ ,  $p \leq 0.001$ ) (Figure 4 A). The largest lesion ( $0.48 \text{ cm}^2$ ) was recorded in the untreated control, followed by BTLK6A, where BTLK6A VOCs reduced the lesion size by 43.75% ( $0.27 \text{ cm}^2$ ) compared to the control. The smallest lesion was observed in BTS-4, with a >85% reduction in lesion size ( $0.07 \text{ cm}^2$ ) compared to the control, followed by the *Bacillus* consortium (81.25% reduction,  $0.09 \text{ cm}^2$ ) and BTS-3 (72.9% reduction,  $0.13 \text{ cm}^2$ ). Therefore, there were no significant differences existed between BTS-3, BTS-4, and *Bacillus* consortium treatments (Figure 3 B).





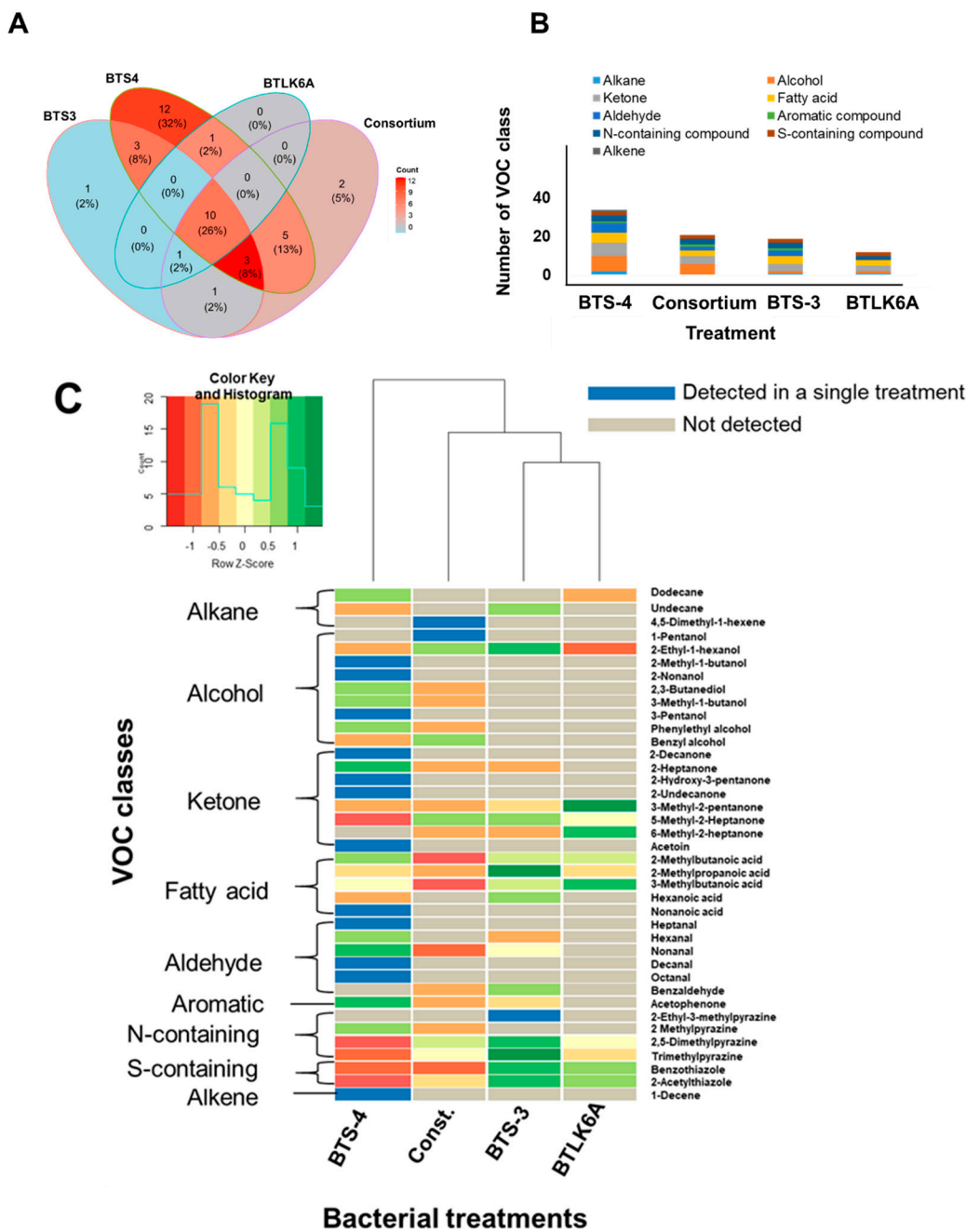
**Figure 3.** Effects of *Bacillus* VOCs on lesion development and MoT sporulation in a detached wheat leaf assay (cv. BR-18). (A) VOCs from *Bacillus* spp. significantly reduced leaf blast lesion size *in vivo*. (B) *Bacillus* VOCs significantly reduced leaf lesion area (cm<sup>2</sup>) caused by MoT *in vivo* (Kruskal Wallis test; n = 30;  $p \leq 0.05$ ); (C) Reduction of MoT sporulation by VOCs from different *Bacillus* spp. *in vivo* (Tukey test; n = 30;  $p \leq 0.05$ ). Data were recorded after 5 d of MoT incubation at 25°C. BTS-3: *B. subtilis* BTS-3; BTS-4: *B. velezensis* BTS-4; BTLK6A: *B. velezensis* BTLK6A; consortium: a mixture of BTS-3, BTS-4 and BTLK6A; untreated control: only MoT inoculated. Black dots in (B) and (C) represent data points for each replicate.

MoT sporulation in VOC-treated leaf lesions was lower compared to the control. A single lesion on a control leaf segment yielded  $5.6 \times 10^4$  conidia/lesion, significantly different from all other bacterial treatments. The BTS-4 treated leaf segments had the lowest number of conidia ( $1.9 \times 10^3$  conidia/lesion), followed by the *Bacillus* consortium ( $3.5 \times 10^3$  conidia/lesion). However, the numbers of conidia produced in BTS-4 and consortium-treated leaf lesions were not significantly different.

There was no MoT sporulation in the water-treated control as there was no MoT infection present (Figure 3 C).

3.3. Identification and quantification of *Bacillus* volatile organic compounds (VOCS)

The VOCs produced from the four different *Bacillus* treatments [BTS-3, BTS-4, BTLK6A, and consortium (a mixture of all three *Bacillus* strains)] were identified and quantified using GC-MS. Thirty-nine VOCs were identified in total, of which 11 were produced by all four bacterial treatments (Figure 4 A, Supplementary table S5).

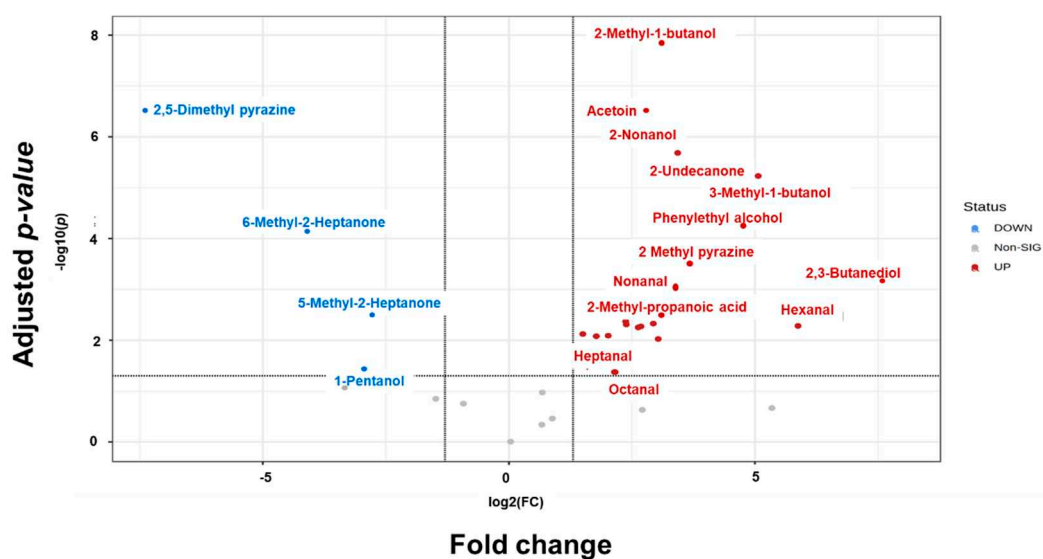


**Figure 4.** Identification and quantification of VOCs from different *Bacillus* treatments by GC-MS analysis. (A) Venn diagram representing the number of VOCs produced from different *Bacillus* spp.; (B) Number of different VOC classes produced from different *Bacillus* treatments; (C) Heatmap based on Euclidean distance showing the VOCs produced from each *Bacillus* treatment. Each line in the color heatmap indicates a single compound; red to green color code indicates low to high relative concentrations (based on row Z-scores) of the compounds; blue color indicates compounds detected

only in a single treatment; grey color indicates undetected volatile. BTS-3: *B. subtilis* BTS-3; BTS-4: *B. velezensis* BTS-4; BTLK6A: *B. velezensis* BTLK6A; const.: consortium (a mixture of *B. subtilis* BTS-3, *B. velezensis* BTS-4, and *B. velezensis* BTLK6A) ( $n = 5$ ;  $p \leq 0.05$ ). The VOCs were quantified from 40 ml LB inoculated with 100  $\mu$ l ( $1 \times 10^9$  CFU/ml) bacteria and incubated for 4 d at 25°C.

The greatest diversity of VOCs were released by BTS-4 (34), followed by the *Bacillus* consortium (22), and lastly by BTLK6A (12). Among the 39 VOCs, 12 unique VOCs were produced by BTS-4, two by the *Bacillus* consortium, and only one by BTS-3. A total of 9 different classes of volatiles were identified- alkanes 7.70%, alcohols 23.07%, ketones 20.51%, fatty acids 12.82%, aldehydes 15.38%, aromatic 2.56%, N-containing 10.25%, S-containing 5.12%, and alkene compounds 2.56% (Figure 4 B). Alcohol, fatty acid, ketone, S-containing, and aldehyde compounds were identified in all four *Bacillus* treatments. The highest number of diversified VOC classes was detected in BTS-4 (9), followed by BTS-3 (8), consortium (7), and BTLK6A (6). The number of alcoholic VOCs was higher for BTS-4, followed by the *Bacillus* consortium. Fatty acid VOCs were also higher in BTS-4, followed by BTS-3 and *Bacillus* consortium treatments (Figure 4 B). The concentrations of bacterial VOCs produced by different treatments differed significantly. Heatmap analysis represents the VOCs clustering and the relationships between different bacterial treatments (Figure 4 C).

*In vitro* and *in vivo* experimental data indicated that VOCs from BTS-4 and *Bacillus* consortium treatments had considerable potential to control MoT. Therefore, we investigated the relationship between VOCs produced from BTS-4 and consortium treatments to determine the effectiveness of BTS-4 and *Bacillus* consortium volatiles against MoT. Figure 5 displays the fold change ( $p \leq 0.05$ ) in VOCs production from BTS-4 compared to the *Bacillus* consortium. In BTS-4, 21 VOCs were upregulated, 4 were down-regulated, and 9 were not significantly different from the *Bacillus* consortium treatment (Figure 5).



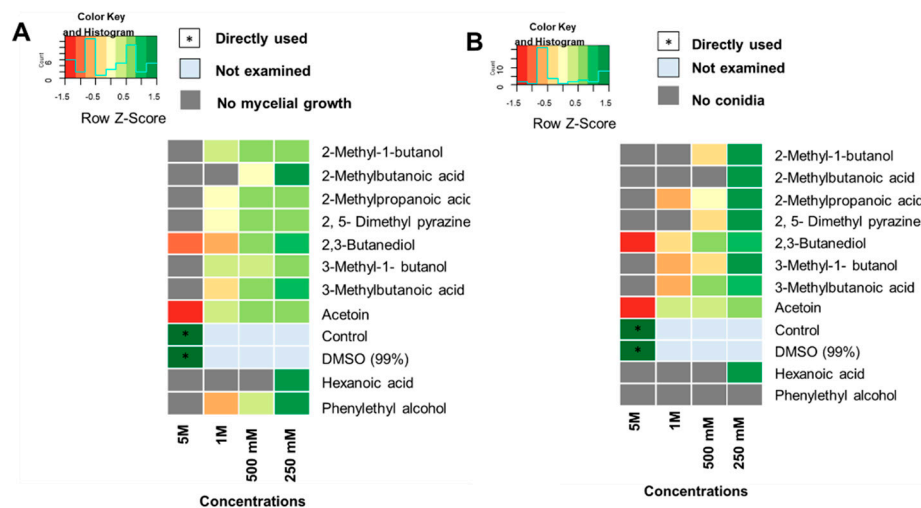
**Figure 5.** Fold change (FC) of VOCs produced by *B. velezensis* BTS-4 in relation to the *Bacillus* consortium (a mixture of *B. subtilis* BTS-3, *B. velezensis* BTS-4, and *B. velezensis* BTLK6A) treatment through volcano plot analysis. The log2 fold change threshold was 2.0. The false discovery rate (FDR) was maintained with  $p$  threshold at 0.05. The VOCs were quantified from 40 ml LB inoculated with 100  $\mu$ l ( $1 \times 10^9$  CFU/ml) bacteria and incubated for 4 d at 25°C. Non-SIG: Non-significant.

### 3.4. Effect of pure VOCs on mycelial growth and sporulation of MoT

From 39 identified VOCs, ten bioactive VOCs (2-methyl propionic acid, 3-methyl-1-butanol, 3-methyl butanoic acid, 2,3-butanediol, and 2-methyl butanoic acid, phenyl ethyl alcohol, hexanoic acid, 2-methyl-1-butanol, 2,5-dimethyl pyrazine, and acetoin) were tested *in vitro* against MoT based on previous literature [50]. DMSO and water were used as positive controls. Of the selected compounds, four VOCs (2-methyl butanoic acid, 2-methyl propanoic acid, 2,5- dimethyl pyrazine

and 3-methyl butanoic acid) were produced by all four bacterial treatments; three (2,3-butanediol, 3-methyl-1-butanol, and phenyl ethyl alcohol) were produced by BTS-4 and *Bacillus* consortium, two VOCs (2-methyl-1-butanol and acetoin) only in BTS-4; and one (hexanoic acid) in BTS-3 and BTS-4 treatments.

The efficacy of single pure volatile compounds against MoT mycelial growth was assessed in an *in vitro* bioassay. Pure VOCs were used at four different concentrations (5M, 1M, 500mM, and 250mM). At 5M, all compounds except acetoin and 2,3-butanediol inhibited the mycelial growth of MoT (Figure 6 A). Among the pure compounds, hexanoic acid suppressed MoT growth up to 500mM concentration (Supplementary figure S2 and Supplementary Table S6).



**Figure 6.** Effect of selected pure antifungal VOCs against MoT mycelial growth and sporulation *in vitro* represented by heatmaps based on Euclidean distance (A) effect of pure antifungal VOCs on MoT mycelial growth; (B) effect of pure antifungal VOCs on MoT sporulation. Each row in the color heatmap indicates a single compound. Red to green color code indicates low to high relative growth or sporulation of MoT (based on row Z score). \* indicates directly used single concentration (diluent) ( $n = 6$ ,  $p \leq 0.05$ ).

In parallel with the reduction of MoT mycelial growth, the selected pure VOCs significantly reduced sporulation from MoT mycelia *in vitro* (Supplementary figure S3 and Supplementary table S6). No sporulation was observed in any of the four treatments with phenylethyl alcohol (PEA). Similarly, no sporulation was recorded for hexanoic acid or 2-methylbutanoic acid up to 500mM concentration (Figure 6 B). Figure 6 A and 6 B contain heatmaps showing the relative effects of potential VOCs on the reduction of mycelial growth and sporulation of MoT. The lowest VOC concentration inhibitory to MoT sporulation was recorded for PEA (250 mM), followed by 2-methylbutanoic acid and hexanoic acid at 500 mM (Figure 6 B).

#### 4. Discussion

In this study, we used four bacterial treatments (BTS-3, BTS-4, BTLK6A, and a consortium of *Bacillus* spp.) to assess the effects of *Bacillus* spp. VOCs to control the emerging fungal pathogen *M. oryzae* *Triticum* (MoT) pathotype. All *Bacillus* treatments produced diverse VOCs and exhibited strong antagonism against MoT, by suppressing mycelial growth and sporulation *in vitro*. It is well documented that *Bacillus* VOCs exert antifungal activity against various phytopathogens [52,53]. Additionally, some studies have reported that the volatiles from *B. megaterium* [54], endophytic *Chryseobacterium* [37], *Pseudomonas* sp. [55] suppress the mycelial growth of the rice blast pathogen (*M. oryzae* *Oryzae* pathotype). However, so far, no information has been available about the suppression of MoT mediated by *Bacillus* volatiles. To the best of knowledge, this is the first report on *Bacillus* VOCs significantly inhibiting the mycelial growth of this important pathogen.



It has been previously reported that bacterial consortia are more effective at controlling certain fungal pathogens than single bacterial strains [56,57]. In this study, *B. velezensis* BTS-4 and the *Bacillus* consortium performed better than the other *Bacillus* treatments in suppressing MoT mycelial growth and conidial germination *in vitro*. Additionally, density of *Bacillus* spp. had a significant positive correlation with MoT inhibition. At  $1 \times 10^9$  CFU/ml, the MoT inhibition rate was higher than at  $1 \times 10^7$  and  $1 \times 10^8$  CFU/ml. Higher density of *Bacillus* spp. led to more *Bacillus* colony growth, higher VOCs production, and a significant reduction in growth and sporulation of MoT. *B. velezensis* can inhibit the growth of *Colletotrichum gloeosporioides* at a density of  $1 \times 10^7$  CFU/ml [52] and *S. sclerotiorum* at a density of  $1 \times 10^8$  CFU/ml [58]. Furthermore, *B. subtilis* has been documented to control *Alternaria solani* at a density of  $1 \times 10^8$  CFU/ml [59] and *B. amyloliquefaciens* VOCs can suppress *Fusarium oxysporum* f. sp. *cubense* *in vitro* also at density of  $1 \times 10^8$  CFU/ml [60].

These findings suggest that the VOCs from *Bacillus* spp. may lead to the functional degradation of MoT mycelia and thus suppress sporulation from MoT mycelia. Deformed hyphae with vacuolation, excessive branching, degeneration of hyphal cells, or combinations of excessive branching with vacuolation were recorded (Supplementary figure S1). Likewise, the VOCs from *B. velezensis* and *B. atrophaeus* also cause vacuolation and cavities in the mycelial cytoplasm of *B. cinerea* [58] and VOCs of *B. subtilis* may cause expanded uneven flaccid hyphae and suppression of *A. solani* sporulation [59].

Furthermore, *Bacillus* VOCs significantly reduced leaf blast lesion size and further MoT sporulation from the lesions in an *in vivo* detached leaf assay. Conidia are the main dispersal unit of MoT epidemiology. Reduced or no conidia formation will result in reduced wheat infection by MoT. Therefore, understanding the MoT sporulation reduction mechanisms of *Bacillus* VOCs is the first step in controlling MoT epidemics. Earlier reports reveal that the volatiles of *Bacillus* spp. may reduce the lesion size and sporulation of *A. solani* in potato leaves [59] as well as sporulation of *Sclerotinia sclerotiorum* on tomato, tobacco, and soybean leaves *in vivo* [61]. However, our results confirm that exposure to *Bacillus* VOCs does not entirely prevent MoT infection or sporulation but rather slows down the development of blast symptoms compared to the untreated control. *Bacillus* VOCs also significantly reduced blast lesion development in detached spike assay (data not shown).

GC-MS was used to determine the active VOCs from different *Bacillus* treatments. Thirty-nine VOCs were identified in total, of which 11 VOCs were produced in all four *Bacillus* treatments. The emitted bacterial VOCs were alcohols, alkenes, alkynes, ketones, aldehydes, fatty acids, aromatic, N-containing, and S-containing compounds. VOCs identified in our analysis have demonstrated broader antifungal activity against phytopathogens. The mixture of alcoholic volatiles 2-methyl-1-butanol and 3-methyl-1-butanol was very effective in suppressing the growth of *Phyllosticta citricarpa* [62] and *Aspergillus flavus* [63] and 2-ethyl-1-hexanol strongly inhibited the growth of *Colletotrichum acutatum* [64] and *B. cinerea* [65].

The 6-methyl-2-heptanone disrupts mycelial integrity, collapses conidial vesicles, and down-regulates the conidial germination gene of pathogenic fungus [66]. The S-containing volatile compound benzothiazole inhibits cystospore germination and mycelial growth of *Phytophthora parasitica* var. *nicotianae* [67]. Hexanal, an aldehyde group volatile, induces systemic resistance in mango plants by inducing defense-related enzymes [phenylalanine ammonia lyase (PAL), peroxidase (PO), polyphenol oxidase (PPO), superoxide dismutase (SOD) and catalase (CAT)] thus significantly reducing *Lasiodiplodia theobromae* infection [68].

In our study, ten VOCs were selected to test their potential in inhibiting MoT based on their reported bioactivity in the literature. All selected VOCs except acetoin and 2,3-butanediol significantly reduced MoT mycelial growth and sporulation. Acetoin and 2,3-butanediol play a role in inducing systemic resistance in plants [69,70] and do not seem to be directly involved in the suppression of MoT mycelial growth and sporulation. 2,5-dimethyl pyrazine stopped MoT sporulation at 1M concentration and has also been cited to control *Sclerotinia* sp., *Pythium* sp., *Rhizoctonia* sp. [71], and *Anthraco* sp. [72]. The activity of 2-methyl propanoic acid against MoT was not promising; although it effectively controls rubber white root rot disease, it negatively affects seedling growth [73].



Hexanoic acid, phenylethyl alcohol, and 2-methyl butanoic acid potentially inhibited the mycelial growth and sporulation of MoT. Phenylethyl alcohol slows phytopathogenic growth by inhibiting the synthesis of RNA, DNA and protein; and upregulates genes related to the phagosome, peroxisome, proteasome, and autophagy [74]. Considering this information, it can be deduced that the phenylethyl alcohol first causes MoT mycelial alternations and later induces autophagy triggering programmed cell death. Fatty acids have also been reported to exert inhibitory activity against some fungal pathogens, but saturated fatty acids have robust antifungal activity compared to other fatty acids [75]. This study found that hexanoic acid (a saturated fatty acid) inhibited mycelial growth and sporulation up to 500 mM. It has been documented that the minimum inhibitory concentrations (MICs) of hexanoic acid against *Micosporum gypseum* range from 0.02 to 75 µg/ml [76]. At a concentration of 10 mM, *Candida albicans* growth is inhibited by hexanoic acid through changes in intracellular hydrostatic pressure and subsequent disruption of the cell plasma membrane [77]. Additionally, hexanoic acid enhances plant jasmonic acid (JA) signaling and induces callose deposition during fungal infection [78].

Organic fertilizers promoted the growth of *B. amyloliquefaciens*, induced the release of 2-nonanone and nonanal, and suppressed *R. solanacearum* [79]. Encapsulation of *Bacillus* VOCs might be an effective way to use bacterial VOCs under field conditions; thus, it facilitates the slow and steady release of volatiles. Effective control of MoT using bacterial VOCs requires more detailed studies considering field environmental conditions and compatibility with other control strategies. Therefore, our study suggests that *Bacillus* VOCs are potential biologicals to suppress MoT, with fundamental and practical implications for wheat production through reducing the severity of wheat blast. As *Bacillus* spp. are rich in production of both volatile and non-volatile antimicrobial compounds [47], further studies are warranted to identify non-volatile antimicrobial secondary metabolites from the investigated *Bacillus* spp. that might work together to effectively control wheat blast. Field evaluation of wheat blast suppression by these *Bacillus* and their metabolites is required before recommending them for practical application in biorational management of wheat blast.

#### Supplementary Materials:

**Author Contributions:** M.Z.S.: processed samples, planned experiments, collected data, designed and performed data analysis, visualized data, and wrote original draft; M.Z.S and S.R.: performed volatile extraction, analyzed and visualized volatile data; M.R.: coordinated, designed volatile extraction experiments, and revised the paper; T.I.: Conceived the idea, provides bacterial and fungal strains, and reviewed the paper; A.vT.: conceived and coordinated the project, reviewed and edited the paper.

**Funding:** M.Z.S. received scholarship (personal reference no. 91730381) from the German academic exchange service (DAAD) to pursue her PhD degree at the Georg-August-University of Goettingen, Germany. The Division of Plant Pathology and Crop Protection, and the University Library (SUB) of the Georg-August-University of Goettingen, Germany paid for the lab consumables and open-access publication fees.

**Data Availability Statement:** All data supporting the results included in the manuscript.

**Acknowledgments:** We would like to thank Dr. Birger Koopmann, Dr. Dipali Rani Gupta, and Nur Uddin Mahmud for processing material transfer (bacterial and fungal strains); Sarenqimuge Sarenqimuge for her suggestions and support during volatile experiments. We also acknowledge the efforts contributed by the Crop Protection Master's students Bishnu Prasad Marahatta and Youssef Beik Saade for their preliminary work on MoT and *Bacillus* strains. Institute of Biotechnology and Genetic Engineering (IBGE), Bangladesh, has provided authorization and information for studying MoT and bacterial strains for scientific activities to the Division of Plant Pathology and Crop Protection, Georg-August-University of Goettingen, Germany.

**Institutional Review Board Statement:** No animals and/or human participants were involved in this research.

**Consent Statement:** This manuscript has not been published and is not under consideration by any other journals. We read and understood the journal policies and believe that the manuscript and study not violates any of these. All authors have been actively and personally involved in the work leading to the manuscript and will hold themselves individually and jointly responsible for its content. All authors agreed to this submission.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

- Awika, J. M. Major cereal grains production and use around the world. In: *Advances in Cereal Science: Implications to Food Processing and Health Promotion*; Awika, J. M.; Piironen, V.; Bean, S. (eds); ACS Symposium Series 1089, American Chemical Society held in Anaheim CA. **2011**; Volume 1089, Chapter 1, pp. 1-13. doi: 10.1021/bk-2011-1089.ch001.
- World Agricultural Production. com. Available online: [www.worldagriculturalproduction.com/crops/wheat.aspx](http://www.worldagriculturalproduction.com/crops/wheat.aspx) (accessed on 12 April 2023).
- Cruz, C. D.; Valent, B. Wheat blast disease: danger on the move. *Trop. Plant Pathol.* **2017**, *42*, 210-222. doi:10.1007/s40858-017-0159-z.
- Ceresini, P. C.; Castroagudín, V. L.; Rodrigues, F. A.; Rios, J. A.; Aucique-Pérez, C. E.; Moreira, S. I.; Croll, D.; Alves, E.; de Carvalho, G.; Maciel, J. L. N.; McDonald, B. A. Wheat blast: From its origins in South America to its emergence as a global threat. *Mol. Plant Pathol.* **2019**, *20*(2), 155- 172. doi: 10.1111/mpp.12747.
- Islam, T.; Ansary, M. W. R.; Rahman, M.M. *Magnaporthe oryzae* and its pathotypes: a potential plant pandemic threat to global food security. In: *Plant Relationships*; Scott, B.; Mesarich, C. (eds); Springer, Cham. *The Mycota*, **2023**; Volume 5, pp. 425–462. doi:10.1007/978-3-031-16503-0\_18.
- Islam, M. T.; Kim, K. H.; Choi, J. Wheat blast in Bangladesh: The current situation and future impacts. *The Plant Pathol. J.* **2019**, *35*(1), 1-10. doi: [10.5423/PJ.RW.08.2018.0168](https://doi.org/10.5423/PJ.RW.08.2018.0168).
- Islam, M. T.; Gupta, D. R.; Hossain, A.; Roy, K. K.; He, X.; Kabir, M. R.; Singh, P. K.; Khan, M. A. R.; Rahman, M.; Wang, G. L. Wheat blast: a new threat to food security. *Phytopathol. Res.* **2020**, *2*, 28. doi:10.1186/s42483-020-00067-6.
- Islam, M. T.; Croll, D.; Gladieux, P.; Soanes, D. M.; Persoons, A.; Bhattacharjee, P.; Hossain, M. S.; Gupta, D. R.; Rahman, M. M.; Mahboob, M. G.; Cook, N.; Salam, M. U.; Surovy, M. Z.; Sancho, V. B.; Maciel, J. L. N.; Nhani Júnior, A.; Castroagudín, V. L.; de Assis Reges, J.; Ceresini, P. C.; Ravel, S.; Kellner, R.; Fournier, E.; Tharreau, D.; Lebrun, M.-H.; McDonald, B. A.; Stitt, T.; Swan, D.; Talbot, N. J.; Saunders, D. G. O.; Win, J.; Kamoun, S. Emergence of wheat blast in Bangladesh was caused by a South American lineage of *Magnaporthe oryzae*. *BMC Biol.* **2016**, *14*, 84. doi: 10.1186/s12915-016-0309-7.
- Tembo, B.; Mulenga, R. M.; Sichilima, S.; M'siska, K. K.; Mwale, M.; Chikoti, P. C.; Singh, P. K.; He, X.; Pedley, K. F.; Peterson, G. L.; Singh, R. P.; Braun, H. J. Detection and characterization of fungus (*Magnaporthe oryzae* pathotype *Triticum*) causing wheat blast disease on rain-fed grown wheat (*Triticum aestivum* L.) in Zambia. *PLoS ONE* **2020**, *15*(9), e0238724. doi:10.1371/journal.pone.0238724.
- Latorre, S. M.; Were, V. M.; Foster, A. J.; Langner, T.; Malmgren, A.; Harant, A.; Asuke, S.; Reyes-Avila, S.; Gupta, D. R.; Jensen, C.; Ma, W.; Mahmud, N. U.; Meheub, M. S.; Mulenga, R. M.; Muzahid, A. N. M.; Paul, S. K.; Rabby, S. M. F.; Rahat, A. M.; Ryder, L.; Shrestha, R.-K.; Sichilima, S.; Soanes, D. M.; Singh, P. K.; Bentley, A. R.; Saunders, D. G. O.; Tosa, Y.; Croll, D.; Lamour, K. H.; Islam, T.; Tembo, B.; Win, J.; Talbot, N. J.; Burbano, H. A.; Kamoun, S. Genomic surveillance uncovers a pandemic clonal lineage of the wheat blast fungus. *Plos Biology* **2023**, *21*(4), e3002052. doi: 10.1371/journal.pbio.3002052.
- Barragan, A. C.; Latorre, S. M.; Mock, P. G.; Harant, A.; Win, J.; Malmgren, A.; Burbano, H. A.; Kamoun, S.; Langer, T. Wild grass strains of *Magnaporthe* (*Syn. Pyricularia*) spp. from Germany can cause blast disease on cereal crops. *bioRxiv*. **2022**. doi: 10.1101/2022.08.29.505667.
- Pagani, A.P.S.; Dianese, A.C.; Café-Filho, A.C. Management of wheat blast with synthetic fungicides, partial resistance and silicate and phosphite minerals. *Phytoparasitica* **2014**, *42*, 609–617.
- Poloni, N. M.; Carvalho, G.; Vicentini, S. N. C.; Dorigan, A. F.; Maciel, J. L. N.; McDonald, B. A.; Moreira, S. I.; Hawkins, N.; Fraaije, B. A.; Kelly, D. E. K.; Kelly, S. L.; Ceresini, P. C. Widespread distribution of resistance to triazole fungicides in Brazilian populations of the wheat blast pathogen. *Plant Pathol.* **2021**, *70*, 436-448. doi: 10.1111/ppa.13288.
- Corkley, L.; Fraaije, B.; Hawkins, N. Fungicides resistance management: Maximizing the effective life of plant protection products. *Plant Pathol.* **2021**, *71*(1), 150-169. doi:10.1111/ppa.13467.
- Castroagudín, V. L.; Ceresini, P. C.; de Oliveira, S. C.; Reges, J. T. A.; Maciel, J. L. N.; Bonato, A. L. V.; Dorigan, A. F.; McDonald, B. A. Resistance to QoI fungicides is widespread in Brazilian populations of the wheat blast pathogen *Magnaporthe oryzae*. *Phytopathol.* **2015**, *105*(3), 284- 294. doi: 10.1094/PHYTO-06-14-0184-R.
- Ceresini, P. C.; Castroagudín, V. L.; Rodrigues, F. A.; Rios, J. A.; Eduardo Aucique-Pérez, C.; Moreira, S. I.; Alves, E.; Croll, D.; Maciel, J. L. N. Wheat blast: Past, present, and future. *Annu. Rev. Phytopathol.* **2018**, *56*, 427-456. doi: [10.1146/annurev-phyto-080417-050036](https://doi.org/10.1146/annurev-phyto-080417-050036).

17. Chet, I.; Inbar, J. Biological control of fungal pathogens. *Appl. Biochem. Biotechnol.* **1994**, *48*(1), 37-43. doi:10.1007/BF02825358.
18. Raaijmakers, J. M.; Mazzola, M. Diversity and natural functions of antibiotics produced by beneficial and plant pathogenic bacteria. *Annu. Rev. phytopathol.* **2012**, *50*, 403-424.
19. Chakraborty, M.; Mahmud, N.U.; Ullah, C.; Rahman, M.; Islam, T. Biological and biorational management of blast diseases in cereals caused by *Magnaporthe oryzae*. *Crit. Rev. Biotechnol.* **2021**, *18*, 1–29. doi: 10.1080/07388551.2021.1898325.
20. Pal, K. K.; McSpadden Gardener, B. Biological control of plant pathogens. *Plant Health Instr.* **2006**, *2*, 1. doi: 10.1094/PHI-A-2006-1117-02.
21. Köhl, J.; Kolnaar, R.; Ravensberg, W. J. Mode of action of microbial biological control agents against plant diseases: Relevance beyond efficacy. *Front. Plant Sci.* **2019**, *10*, 845. doi:10.3389/fpls.2019.00845.
22. Malik, M. S.; Haider, S.; Rehman, A.; Rehman, S. U.; Jamil, M.; Naz, I.; Anees, M. Biological control of fungal pathogens of tomato (*Lycopersicon esculentum*) by chitinolytic bacterial strains. *J. Basic Microbiol.* **2022**, *62*, 48-62. doi:10.1002/jobm.202100512.
23. Surovy, M. Z.; Rahman, S.; Dame, Z. T.; Islam, T. Discovery of bioactive natural products from *Bacillus* Species: Chemistry, biosynthesis and biological activities. In: *Bacilli in Agrobiotechnology. Bacilli in Climate Resilient Agriculture and Bioprospecting*. Islam, M. T.; Rahman, M.; Pandey, P (eds.). Springer, Cham, **2022a**; pp. 47-87. doi : 10.1007/978-3-030-85465-2\_3.
24. Chakraborty, M.; Mahmud, N. U.; Muzahid, A. N. M.; Rabby, S. F.; Islam, T. Oligomycins inhibit *Magnaporthe oryzae* *Triticum* and suppress wheat blast disease. *PLoS One* **2020a**, *15*(8), e0233665. doi:10.1371/journal.pone.0233665.
25. Chakraborty, M.; Mahmud, N. U.; Gupta, D. R.; Tareq, F. S.; Shin, H. J.; Islam, T. (2020b). Inhibitory effects of linear lipopeptides from a marine *Bacillus subtilis* on the wheat blast fungus *Magnaporthe oryzae* *Triticum*. *Front. Microbiol.* **2020b**, *11*, 665. doi:10.3389/fmicb.2020.00665.
26. Chakraborty, M.; Rabby, S. M. F.; Gupta, D. R.; Rahman, M.; Paul, S. K.; Mahmud, N. U.; Rahat, A. A. M.; Jankuloski, L.; Islam, T. (2022). Natural protein kinase inhibitors staurosporine and chelerythrine suppress wheat blast disease caused by *Magnaporthe oryzae* *Triticum*. *Microorganisms* **2022**, *10*(6), 1186. doi: 10.3390/microorganisms10061186.
27. Paul, S. K.; Chakraborty, M.; Rahman, M.; Gupta, D. R.; Mahmud, N. U.; Rahat, A. A. M.; Sarker, A.; Hannan, M. A.; Rahman, M. M.; Akanda, A. M.; Ahmed, J. U.; Islam, T. Marine natural product antimycin A suppresses wheat blast disease caused by *Magnaporthe oryzae* *Triticum*. *J. Fungi* **2022**, *8*(6), 618. doi:10.3390/jof8060618.
28. Heenan-Daly, D.; Coughlan, S.; Dillane, E.; Prestwich, B. D. Volatile compounds from *Bacillus*, *Serratia*, and *Pseudomonas* promote growth and alter the transcriptional landscape of *Solanum tuberosum* in a passively ventilated growth system. *Front. Microbiol.* **2021**, *12*, 628437. doi: 10.3389/fmicb.2021.628437.
29. Fincheira, P.; Quiroz, A.; Tortella, G.; Diez, M. C.; Rubilar, O. Current advances in plant-microbe communication via volatile organic compounds as an innovative strategy to improve plant growth. *Microbiol. Res.* **2021**, 126726. doi: 10.1016/j.micres.2021.126726.
30. Schulz-Bohm, K.; Martin-Sánchez, L.; Garbeva, P. Microbial volatiles: Small molecules with an important role in intra and inter-kingdom interactions. *Front. Microbiol.* **2017**, *8*, 2484. doi: 10.3389/fmicb.2017.02484.
31. Brilli, F.; Loreto, F.; Baccelli, I. Exploiting plant volatile organic compounds (VOCs) in agriculture to improve sustainable defense strategies and productivity of crops. *Fron. in Plant Sci.* **2019**, *10*, 264. doi:10.3389/fpls.2019.00264.
32. Bui, H. X.; Desaeager, J. A. Volatile compounds as potential bio-fumigants against plant-parasitic nematodes – a mini review. *J. Nematol.* **2021**, *53*, e2021- 14. doi: 10.21307/jofnem-2021-014.
33. Kandel, S. L.; Joubert, P. M.; Doty, S. L. Bacterial endophyte colonization and distribution within plants. *Microorganisms.* **2017**, *5*(4), 77. doi: 10.3390/microorganisms5040077.
34. Mawar, R.; Manjunatha, B. L.; Kumar, S. Commercialization, diffusion and adoption of bioformulations for sustainable disease management in indian arid agriculture: Prospects and challenges. *Circ. Econ. Sust.* **2021**, *1*, 1367-1385. doi: 10.1007/s43615-021-00089-y.
35. Mülner, P.; Bergna, A.; Wagner, P.; Sarajlić, D.; Gstöttenmayr, B.; Dietel, K. et al. Microbiota associated with *Sclerotia* of soilborne fungal pathogens – a novel source of biocontrol agents producing bioactive volatiles. *Phytobiomes J.* **2019**, *3*, 125–136. doi: 10.1094/PBIOMES-11-18-0051-R.

36. Zhang, X.; Li, B.; Wang, Y.; Guo, Q.; Lu, X.; Li, S.; et al. Lipopeptides, a novel protein, and volatile compounds contribute to the antifungal activity of the biocontrol agent *Bacillus atrophaeus* CAB-1. *Appl. Microbiol. Biotechnol.* **2013**, *97*, 9525–9534. doi: 10.1007/s00253-013-5198-x.
37. Kumar, M.; Charishma, K.; Sahu, K. P.; Sheoran, N.; Patel, A.; Kundu, A.; Kumar, A. Rice leaf associated *Chryseobacterium* species: An untapped antagonistic *flavobacterium* displays volatile mediated suppression of rice blast disease. *Biol. Con.* **2021**, *1(161)*, 104703. doi: 10.1016/j.biocontrol.2021.104703.
38. Tenorio-Salgado, S.; Tinoco, R.; Vazquez-Duhalt, R.; Caballero-Mellado, J.; Perez-Rueda, E. Identification of volatile compounds produced by the bacterium *Burkholderia tropica* that inhibit the growth of fungal pathogens. *Bioengineered* **2013**, *4*, 236–243. doi: 10.4161/bioe.23808.
39. Groenhagen, U.; Baumgartner, R.; Bailly, A.; Gardiner, A.; Eberl, L.; Schulz, S.; et al. Production of bioactive volatiles by different *Burkholderia ambifaria* strains. *J. Chem. Ecol.* **2013**, *39*, 892–906. doi: 10.1007/s10886-013-0315-y.
40. Minerdi, D.; Bossi, S.; Maffei, M. E.; Gullino, M. L.; Garibaldi, A. *Fusarium oxysporum* and its bacterial consortium promote lettuce growth and expansin A5 gene expression through microbial volatile organic compound (MVOC) emission. *FEMS Microbiol. Ecol.* **2011**, *76*, 342–351. doi: 10.1111/j.1574-6941.2011.01051.x.
41. Foroutan, A. Evaluation of *Trichoderma* strains for biological control of wheat Fusarium foot and root rot. *Rom. Agric. Res.* **2013**, *30(30)*, 335–342.
42. Sasani, M.; Ahmadzadeh, M. Evaluation of antagonistic effect of several strain of *Bacillus* bacteria on control of crown and root rot of wheat with *Fusarium pseudograminearum*. *Biol. Con. Pests Plant Dis.* **2021**, *9(2)*, 115–26. doi: 10.22059/JBIOC.2021.326484.307.
43. Chen, L.; Heng, J.; Qin, S.; Bian, K. A comprehensive understanding of the biocontrol potential of *Bacillus velezensis* LM2303 against Fusarium head blight. *PLoS One* **2018**, *1*, 13(6):e0198560. doi: 10.1371/journal.pone.0198560.
44. Surovy, M. Z.; Gupta, D. R.; Chanclud, E.; Win, J.; Kamoun, S.; Islam, M. Plant probiotic bacteria suppress wheat blast fungus *Magnaporthe oryzae* *Triticum* pathotype. *Figshare* **2017**. doi: 10.21307/jofnem-2021-014.
45. Surovy, M. Z.; Dutta, S.; Mahmud, N. U.; Gupta, D. R.; Farhana, T.; Paul, S. K.; Joe, W.; Dunlap, C. A.; Oliva, R.; Rahman, M.; Islam, T. Probiotic *Bacillus* species: Promising biological control agents for managing worrisome wheat blast disease. *Preprints* **2022b**. doi: 10.20944/preprints202211.0382.v1.
46. Chanclud, E.; Win, J.; Malone, J.; Surovy, M. Z.; Gupta, D. R.; Islam, T.; Kamoun, S. Genome sequences of candidate wheat blast biocontrol bacteria. *figshare* **2017**. doi:10.6084/m9.figshare.5558641.v1.
47. Dutta, S.; Surovy, M. Z.; Gupta, D. R.; Mahmud, N. U.; Chanclud, E.; Win, J.; Kamoun, S.; Islam, T. Genomic analyses reveal that biocontrol of wheat blast by *Bacillus* spp. may be linked with production of antimicrobial compounds and induced systemic resistance in host plants. *figshare* **2018**. doi:10.6084/m9.figshare.5852661.v1.
48. Gupta, D. R.; Surovy, M. Z.; Mahmud, N. U.; Chakraborty, M.; Paul, S. K.; Hossain, M. S.; Bhattacharjee, P.; Meheub, M. S.; Rani, K.; Yeasmin, R.; Rahman, M.; Islam, M. T. Suitable methods for isolation, culture, storage and identification of wheat blast fungus *Magnaporthe oryzae* *Triticum* pathotype. *Phytopathol. Res.* **2020**, *2*, 30. doi: 10.1186/s42483-020-00070-x.
49. Surovy, M. Z.; Islam, T.; von Tiedemann, A. Role of seed infection for the near and far distance dissemination of wheat blast caused by *Magnaporthe oryzae* pathotype *Triticum*. *Front. Microbiol.* **2023**, *14*, 1040605. doi: 10.3389/fmicb.2023.1040605.
50. Sarenqimuge, S.; Rahman, S.; Wang, Y.; von Tiedemann, A. Dormancy and germination of microsclerotia of *Verticillium longisporium* are regulated by soil bacteria and soil moisture levels but not by nutrients. *Front. Microbiol.* **2022**, *13*, 979218. doi: 10.3389/fmicb.2022.979218.
51. Metaboanalyst 5.0. Available online: <https://www.metaboanalyst.ca/home.xhtml> (accessed on 22 November 2022).
52. Choub, V.; Won, S. J.; Ajuna, H. B.; Moon, J. H.; Choi, S. L.; Lim, H. I.; Ahn, Y. S. Antifungal activity of volatile organic compounds from *Bacillus velezensis* CE 100 against *Colletotrichum gloeosporioides*. *Horticulturae*. **2022**, *8*, 557. doi:10.3390/horticulturae8060557.
53. la Cruz-López, D.; Cruz-López, L.; Holguín-Meléndez, F.; Guillén-Navarro, G. K.; Huerta-Palacios, G. Volatile organic compounds produced by cacao endophytic bacteria and their inhibitory activity on *Moniliophthora roreri*. *Curr. Microbiol.* **2022**, *79(2)*, 1–1. doi: 10.1007/s00284-021-02696-2.
54. Munjal, V.; Nadakkakath, A. V.; Sheoran, N.; Kundu, A.; Venugopal, V.; Subaharan, K.; Rajamma, S.; Eapen, S. J.; Kumar, A. Genotyping and identification of broad spectrum antimicrobial volatiles in black pepper



- rot endophytic biocontrol agent, *Bacillus megaterium* BP17. *Biol. Cont.* **2016**, *92*, 66-76. doi:10.1016/j.biocontrol.2015.09.005.
55. Patel, A.; Kumar, A.; Sheoran, N.; Kumar, M.; Sahu, K. P.; Ganeshan, P.; Ashajyothi, M.; Gopalakrishnan, S.; Gogol, R. Antifungal and defense elicitor activities of pyrazines identified in endophytic *Pseudomonas putida* BP25 against fungal blast infected by *Magnaporthe oryzae* in rice. *J. Plant Dis. Prot.* **2021**, *128*, 261-272. doi:10.1007/s41348-020-00373-3.
  56. Cruz-Magalhães, V.; Guimarães, R. A., da Silva, J. C P.; de Faria, A. F.; Pedroso, M. P.; Campos, V. P.; Marbach, P. A. S.; de Medeiros, F. H. V.; Souza, J. T.D. The combination of two *Bacillus* strains suppresses *Meloidogyne incognita* and fungal pathogens, but does not enhance plant growth. *Pest Managem. Sci.* **2021**, *78*(2), 722-732. doi: 10.1002/ps.6685.
  57. Murty, L. D.; Shim, W. B. *Streptomyces* and *Bacillus* species utilize volatile organic compounds to impact *Fusarium oxysporum* f.sp. *vasinfectum* race 4 (Fov4) virulence and suppress *Fusarium* wilt in Pima cotton. *BioRxiv* **2021**. doi: 10.1101/2021.10.27.466178.
  58. Toral, L.; Rodriguez, M.; Martinez-Checa, F.; Montano, A.; Cortés-Delgado, A.; Smolinska, A.; Llamas, I.; Sampedro, I. Identification of volatile organic compounds in extremophilic bacteria and their effective use in biocontrol of postharvest fungal phytopathogens. *Front. Microbiol.* **2021**, *12*, 773092. doi: 10.3389/fmicb.2021.773092.
  59. Zhang, D.; Yu, S.; Yang, Y.; Zhang, J.; Zhao, D.; Pan, Y.; Fan, S.; Yang, Z.; Zhu, J. Antifungal effects of volatiles produced by *Bacillus subtilis* against *Alternaria solani* in potato. *Front. Microbiol.* **2020**, *11*, 1196. doi: 10.3389/fmicb.2020.01196.
  60. Yuan, J.; Raza, W.; Shen, Q.; Huang, Q. Antifungal activity of *Bacillus amyloliquefaciens* NJN-6 volatile compounds against *Fusarium oxysporum* f. sp. *cubense*. *Appl. Environ. Microbiol.* **2012**, *78*, 5942-5944. doi: 10.1128/AEM.0135.
  61. Massawe, V.; Hanif, A.; Farzand, A.; Ochola, S. O. Volatile organic compounds of endophytic *Bacillus* spp. have biocontrol activity against *Sclerotinia sclerotiorum*. *Phytopathol.* **2018**, 1-13. doi: 10.1094/PHYTO-04-18-0118-R.
  62. Toffano, L.; Fialho, M. B.; Pascholati, S. F. Potential of fumigation of orange fruits with volatile organic compounds produced by *Saccharomyces cerevisiae* to control citrus black spot disease at postharvest. *Biol. Control* **2017**, *108*, 77-82. doi: 10.1016/j.biocontrol.2017.02.009.
  63. Jaibangyang, S.; Nasanit, R.; Limtong, S. Effects of temperature and relative humidity on aflatoxin B1 reduction in corn grains and antagonistic activities against aflatoxin-producing *Aspergillus flavus* by a volatile organic compound-producing yeast, *Kwoniella heveanensis* DMKU-CE82. *BioControl* **2021**, *66*, 433-443. Doi: 10.1007/s10526-021-10082-x.
  64. Che, J.; Liu, B.; Liu, G.; Chen, Q.; Lan, J. Volatile organic compounds produced by *Lysinibacillus* sp. FJAT-4748 possess antifungal activity against *Colletotrichum acutatum*. *Biocon. Sci. Technol.* **2017**, *27*(12), 1349-1362. doi: 10.1080/09583157.2017.1397600.
  65. Huang, R.; Che, H. J.; Zhang, J.; Yang, L.; Jiang, D. H.; Li, G. Q. Evaluation of *Sporidiobolus pararoseus* strain YCXT3 as biocontrol agent of *Botrytis cinerea* on post-harvest strawberry fruits. *Biol. Con.* **2012**, *62*, 53-63. doi:10.1016/j.biocontrol.2012.02.010.
  66. Zhang, D.; Qiang, R.; Zhao, J. et al. Mechanism of a volatile organic compound (6-Methyl-2-Heptanone) emitted from *Bacillus subtilis* ZD01 against *Alternaria solani* in potato. *Front. Microbiol.* **2021**, *12*, 808337. doi: 10.3389/fmicb.2021.808337.
  67. Fang, Y.; Zhang, L.; Jiao, Y.; Liao, J.; Luo, L.; Ji, S.; Li, J.; Dai, K.; Zhu, S.; Yang, M. Tobacco rotated with rapeseed for soil-borne *Phytophthora* pathogen biocontrol mediated by rapeseed root exudates. *Front. Microbiol.* **2016**, *7*, 894. doi: 10.3389/fmicb.2016.00894.
  68. Seethapathy, P.; Gurudevan, T.; Subramanian, K. S.; Kuppasamy, P. Bacterial antagonists and hexanal-induced systemic resistance of mango fruits against *Lasiodiplodia theobromae* causing stem-end rot. *J. Plant Interact.* **2016**, *11*(1), 158-166. doi: 10.1080/17429145.2016.1252068.
  69. Wu, L.; Li, X.; Ma, L.; Borris, R.; Wu, Z.; Gao, X. Acetoin and 2,3-butanediol from *Bacillus amyloliquefaciens* induce stomatal closure in *Arabidopsis thaliana* and *Nicotiana benthamiana*. *J. Expt. Bot.* **2018**, *69*(22), 5625-5635. doi: 10.1093/jxb/ery326.
  70. Park, K. Y.; Seo, S. Y.; Oh, B.-R.; Seo, J.-W.; Kim, Y. J. 2,3-Butanediol induces systemic acquired resistance in the plant immune response. *J. Plant Biol.* **2018**, *61*, 424-434. doi:10.1007/s12374-018-0421-z.



71. Vlassi, A.; Nesler, A.; Perazzolli, M.; Lazazzara, V.; Büschl, C.; Parich, A.; Puopolo, G.; Schuhmacher, R. Volatile organic compounds from *Lysobacter capsici* AZ78 as potential candidates for biological control of soilborne plant pathogens. *Front. Microbiol.* **2020**, *11*, 1748. doi: 10.3389/fmicb.2020.01748.
72. Janamatti, A. T.; Kumar, A.; Kaur, C. et al. Fumigation by bacterial volatile 2,5-dimethylpyrazine enhances anthracnose resistance and shelf life of mango. *Eur. J. Plant Pathol.* **2022**, *164*, 209–227. doi: 10.1007/s10658-022-02551-5.
73. Siri-udom, S.; Suwannarach, N.; Lumyong, S. Applications of volatile compounds acquired from *Muscodor heveae* against white root rot disease in rubber trees (*Hevea brasiliensis* Müll. Arg.) and relevant allelopathy effects. *Fun. Biol.* **2017**, *121*(6–7), 573–581. doi: 10.1016/j.funbio.2017.03.004.
74. Liu, P.; Cheng, Y.; Yang, M.; Liu, Y.; Chen, K.; Long, C.-a.; Deng, X. Mechanisms of action for 2-phenylethanol isolated from *Kloeckera apiculata* in control of *Penicillium* molds of citrus fruits. *BMC Microbiol.* **2014**, *14*, 242. doi : 10.1186/s12866-014-0242-2.
75. Liu, P.; Cheng, Y.; Yang, M.; Liu, Y.; Chen, K.; Long, C.-a.; Deng, X. Mechanisms of action for 2-phenylethanol isolated from *Kloeckera apiculata* in control of *Penicillium* molds of citrus fruits. *BMC Microbiol.* **2014**, *14*, 242. doi : 10.1186/s12866-014-0242-2.
76. Chadeganipour, M.; Haims, A. Antifungal activities of pelargonic and capric acid on *Microsporum gypseum*. *Mycoses* **2001**, *44*, 109–112. doi: 10.1046/j.1439-0507.2001.00609.x.
77. Bergsson, G.; Arnfinnsson, J.; Steingrímsson, O.; Thormar, H. *In vitro* killing of *Candida albicans* by fatty acids and monoglycerides. *Antimicrob. Agents Chemother.* **2001**, *45*, 3209–3212. doi: 10.1128/AAC.45.11.3209-3212.2001.
78. Vicedo, B.; Flors, V.; Leyva, M. O.; Finiti, I.; Kravchuk, Z.; Real, M. D.; García-Agustín, P.; González-Bosch. Hexanoic acid-induced resistance against *Botrytis cinerea* in tomato plants. *Mol. Plant Microbe Interact.* **2009**, *22*(11), 1455–1465. doi:10.1094 / MPMI -22-11-1455.
79. Raza, W.; Wei, Z.; Ling, N.; Huang, Q.; Shen, Q. Effect of organic fertilizers prepared from organic waste materials on the production of antibacterial volatile organic compounds by two biocontrol *Bacillus amyloliquefaciens* strains. *J. Biotechnol.* **2016**, *277*, 43–53. doi:10.1016/j.jbiotec.2016.04.014.

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.