

Review

Not peer-reviewed version

Treatment of *Enterococcus faecalis* Infective Endocarditis: A Continuing Challenge

[Laura Herrero-Hidalgo](#) , Beatriz Fernández-Rubio , Rafael Luque-Márquez , [Luis E López-Cortés](#) ,
[María Victoria Gil-Navarro](#) , [Aristides De Alarcón](#) *

Posted Date: 2 March 2023

doi: 10.20944/preprints202303.0033.v1

Keywords: Enterococcus faecalis; treatment; new alternatives



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Review

Treatment of *Enterococcus faecalis* Infective Endocarditis: A Continuing Challenge

Laura Herrero-Hidalgo ¹, Beatriz Fernández-Rubio ¹, Rafael Luque-Márquez ²,
Luis E. López-Cortés ³, Maria V. Gil-Navarro ¹ and Arístides de Alarcón ^{2,*}

¹ Unidad Clínica de Farmacia. Grupo de Resistencias bacterianas y antimicrobianos (CIBERINFEC). Instituto de Biomedicina de Sevilla (IBiS). Hospital Universitario Virgen del Rocío/CSIC/Universidad de Sevilla. 4013 Seville, Spain.

² Unidad Clínica de Enfermedades Infecciosas, Microbiología y Parasitología (UCEIMP) Grupo de Resistencias bacterianas y antimicrobianos (CIBERINFEC). Instituto de Biomedicina de Sevilla (IBiS). Hospital Universitario Virgen del Rocío/CSIC/Universidad de Sevilla. 4013 Seville, Spain.

³ Unidad Clínica de Enfermedades infecciosas y Microbiología. Grupo de Resistencias bacterianas y antimicrobianos (CIBERINFEC). Instituto de Biomedicina de Sevilla (IBiS). Hospital Universitario Virgen Macarena/SCIC/Universidad de Sevilla. 4109 Seville, Spain.

* Correspondence: aa2406ge@yahoo.es

Abstract: Today, enterococci (mainly *Enterococcus faecalis*) are one of the main causes of infective endocarditis in the world, generally affecting an elderly and fragile population, with a high mortality rate. enterococci are intrinsically resistant to many commonly used antimicrobial agents. All enterococci exhibit decreased susceptibility to penicillin and ampicillin, as well as high-level resistance to most cephalosporins and all semi-synthetic penicillins, as the result of expression of low-affinity penicillin-binding proteins, that precludes an unacceptable number of therapeutic failures with monotherapy with these drugs. For years, the synergistic combination of penicillins and aminoglycosides was the cornerstone of treatment, but the emergence of strains with high resistance to aminoglycosides led to the search for new alternatives, such as dual beta-lactam therapy. The development of multi-drug resistant strains of *Enterococcus faecium* is a matter of considerable concern due to its probable spread to *E. faecalis* and have forced the search of new alternatives with the combination of daptomycin, fosfomycin or tigecycline. Some of them have scarce clinical experience and others are still under investigation and will be analyzed in this review. In addition, the need for prolonged treatment (6-8 weeks) to avoid relapses has led to the consideration of viable options such as outpatient parenteral therapies or long-acting administrations with the new lipoglycopeptides (dalbavancin or oritavancin), and sequential oral treatments, which will also be discussed.

Keywords: Enterococcus faecalis; treatment; new alternatives

1. Introduction

Enterococcal infective endocarditis (IE) represents the third leading causal agent worldwide, being responsible for approximately 10-15% of all cases¹⁻³. *Enterococcus faecalis* infective endocarditis accounts for around 90% of enterococcal IE and has experienced important transformation in the last two decades. Compared with patients with non-enterococcal IE, patients with enterococcal IE tend to be older and to have higher rates of cancer, aortic valve affection and previous history of urinary tract or abdominal infections^{4,5}. In the past, enterococcal IE was mainly community-acquired, but nowadays there is a significant increase in the incidence of healthcare acquisition,⁶⁻⁸ especially in aged patients (>70 years) with many comorbidities, one of them being colorectal neoplasms (advanced adenomas and carcinomas)^{9,10}. Today, *E. faecalis* is the most common causative organism isolated in IE in transcatheter aortic valve implantation (TAVI)¹¹.

Diagnosis of *E. faecalis* IE is challenging due to its often subacute course, with nonspecific constitutional symptoms and chronic anemia difficult to interpret in an elderly and frequently immunosuppressed population with a large number of comorbidities. In fact, it is not uncommon that the first symptom to lead to diagnosis to be left ventricular failure, and the rate of cardiac surgery reported is about 40%, usually lower than clinically indicated due to the sometimes poor clinical condition of the patient¹²⁻¹⁴.

Treatment of enterococcal infections has long been recognized as an important clinical challenge, particularly in the setting of IE. The success of enterococcal population for surviving in multiple environments alongside a wide range of inhabitants, and the ease by which they acquire mobile genetic elements, including plasmids from other bacteria is astonishing. Furthermore, the enterococci are frequently present within as bacterial biofilm (specially *E. faecalis*), which provides stability and protection to the bacterial population along with an opportunity for a variety of bacterial interactions^{15,16}. The frequent lack of bactericidal activity of traditional agents (penicillin or ampicillin), the toxicity incurred with the addition of aminoglycosides, and the increased reports of high-level resistances to them¹⁷, in parallel with the production of bacterial biofilms over prosthetic devices^{18,19}, has led to a much higher rate of relapses (7-10%) compared with other etiologies^{5,12,14,20}. These relapses can occur still several months after the end of the antimicrobial therapy^{21,22}, generating continuous uncertainty for the clinician and the need for a prolonged follow-up. Moreover, the emergence of multidrug-resistant isolates in *E. faecium*, brings a new concern for which we have yet no solid therapeutic evidence^{23,24}.

In this review, we will focus on the therapeutic options for *E. faecalis* IE in which we still have many therapeutic options, from “classical” guidelines to new alternatives.

2. Mechanisms of resistance

Enterococci exhibit significant resistance to a wide variety of antimicrobial agents. This resistance is almost certainly relevant in most natural ecological settings in which enterococci dwell. As normal commensals of the human gastrointestinal tract, enterococci are routinely exposed to a myriad of antibiotics in the course of contemporary medical treatment, and enterococcal resistance plays a key role in the ecological dynamics that occur during and after antibiotic therapy. In addition, their resistance has confounded the best efforts of contemporary medicine to cope with infections caused by enterococci. Intrinsic resistance—that which is encoded within the core genome of all members of the species—differs from acquired resistance, in that the latter is present in only some members of the species and is obtained via the horizontal exchange of mobile genetic elements (or via selection upon antibiotic exposure). Resistance for many antibiotics have emerged, including those that are (or once were) clinically useful as therapeutics to treat enterococcal infections, as well as those to which enterococci, as commensals of humans, are incidentally exposed in the course of therapy for infections caused by other bacteria.

A complete description of the mechanisms of resistance is beyond the scope of this review, and we will briefly review the most important ones for treatment. A complete description of the various types is provided in **Table 1**.

Table 1. Antimicrobial resistance in enterococci.

Antimicrobial class	Mechanism of resistance Genes/operons	Species	Comments
Beta-lactams	Production of bet-lactamase	<i>E. faecalis</i> (rare in <i>E. faecium</i>)	*: High level resistance to penicillin, ampicillin, and piperacillin
- High level resistance* but susceptibility to beta-lactamase inhibitors	plasmid non-induced or plasmid-mediated		
- High level resistance **	-Low-affinity PBP4	<i>E. faecalis</i>	**: High level resistance to penicillin, ampicillin, and piperacillin and carbapenems
	-Overproduction of PBP5	<i>E. faecium</i>	
Aminoglycosides			

-	Low-level resistance	Defective aerobic transport across the cell membrane	<i>Enterococcus</i> spp	Structural resistance in all species
-	High-level resistance	-Modification of the aminoglycoside by different enzymes -Alteration of the ribosomal target site		Self- transferable plasmids: phosphotransferase, nucleotidyl transferase, acetyltransferase Chromosomal mutation. Confers resistance to Streptomycin
<i>Glycopeptides (principal phenotypes)</i>				
-	VanA (R to high levels of vancomycin and teicoplanin)	Transposons inserted into the chromosome or on plasmids. Induced by either vancomycin or teicoplanin	<i>E. faecium</i> , <i>E. faecalis</i>	
-	VanB (R to variable levels of vancomycin and susceptible to teicoplanin)	Transposons inserted into the chromosome or on plasmids. Induced only by teicoplanin	<i>E. faecium</i> , <i>E. faecalis</i>	
-	VanC (Low level R to vancomycin and susceptible to teicoplanin)	Located in the chromosome and non-transferable.	<i>E. gallinarum</i> , <i>E. casseliflavus</i> , <i>E. flavescens</i>	Synthesis of peptidoglycan precursors with low affinity for pentaglycopeptides ending in D-Ala-D-lac instead of D-Ala-D-Ala
-	VanD (R to intermediate levels of vancomycin and low levels of teicoplanin)	Located in the chromosome and non-transferable. Expressed constitutively.	<i>E. faecium</i>	
-	VanE (R to low levels of vancomycin and susceptible to teicoplanin)	Located in the chromosome and not transferable.	<i>E. faecalis</i>	Synthesis of peptidoglycan precursors with low affinity for pentaglycopeptides ending in D-Ala-D-Ser instead of D-Ala-D-Ala
-	VanG (R to low levels of vancomycin and susceptible to teicoplanin)	Located on the chromosome and transferable.	<i>E. faecalis</i>	
-	VanN (R to low levels of vancomycin and susceptible to teicoplanin)	Expressed constitutively. Could be transferred by conjugation.	<i>E. faecium</i>	
-	VanM (R to high levels of vancomycin and teicoplanin)	Plasmid-encoded resistance. Could be transferred by conjugation	<i>E. faecium</i>	
<i>Quinolones</i>				
		- Alterations in the GyrA or GyrB subunits of DNA gyrase (gene <i>gyrA/gyrB</i>) and/or the <i>parC</i> subunit of DNA topoisomerase IV (gene <i>parC</i>)	<i>E. faecalis</i>	Encodes protein that protects DNA gyrase from inhibition by fluoroquinolones
		- <i>qnr'</i> -like gene	<i>E. faecalis</i> , <i>E. faecium</i>	
		- Efflux pump	<i>E. faecalis</i> , <i>E. faecium</i>	
<i>Oxazolidinones</i>				
		- rRNA genes	<i>E. faecalis</i> , <i>E. faecium</i>	Mutations reducing affinity in ribosomal subunit.
		- <i>cfr</i>	<i>E. faecalis</i> , <i>E. faecium</i>	Methylation of 23S rRNA
<i>Daptomycin</i>				
		- YycFGHIJ / LiaFSR		Stress-sensing response system
		- Mutation in gene encoding cardiolipin synthase	<i>E. faecalis</i> , <i>E. faecium</i>	Alteration in membrane charge and fluidity

Macrolides and clindamycin	Methylation of an adenoma residue in the 23S rRNA (gene <i>ermB</i>)	<i>Enterococcus</i> spp	Located on plasmids and chromosomal transposons. Constitutive or inducible
Tetracyclines	- Active efflux (gene <i>tetL</i>)	<i>Enterococcus</i> spp	Located on plasmid and transposons. Constitutive or inducible
	- Protection of the ribosome (genes <i>tetN</i> and <i>tetM</i>)		
	- Overexpression of gene <i>tet(L)</i> and <i>tet (M)</i>	<i>E. faecium</i>	Reported with Tigecycline
Chloramphenicol	Chloramphenicol acetyltransferase (gene <i>cat</i>)	<i>Enterococcus</i> spp	Produces acetylation of chloramphenicol. Plasmid-mediated
Sulphonamides and trimethoprim	No gene identified.	<i>Enterococcus</i> spp	Enterococci can use exogenous folates

2.1. Beta-lactams

Enterococci exhibit an intrinsic natural resistance to beta-lactams, due to the low affinity of their penicillin-binding proteins (PBP) for these antibiotics^{25,26}. This intrinsic resistance differs among the different beta-lactams, with penicillins generally having the highest activity against enterococci, carbapenems having slightly lower activity and cephalosporins with the lowest activity, except new-generation cephalosporins ceftobiprole and ceftaroline²⁷⁻²⁹. Piperacillin activity is similar to that of penicillin, but oxacillin, ticarcillin, ertapenem or aztreonam have limited or no activity against enterococci. The most active penicillin *in vitro* is ampicillin, with a minimum inhibitory concentration (MIC) that ranges from 1 to 16 mg/L (much higher than most streptococci), usually one dilution lower than those for penicillin. But despite an apparent good *in vitro* inhibitory activity (e.g., MIC = 1 for ampicillin), previous *in vitro* and *in vivo* studies promptly demonstrated that beta-lactam monotherapy was associated with a poor outcome in patients with endovascular infections^{30,31}. Indeed, the bactericidal activity that is required for curation is rarely achieved with these compounds as a result of the phenomenon known as “tolerance” (lack of killing), making the success of beta-lactam monotherapy unpredictable. Moreover, certain enterococcal strains are killed only at a specific concentration of the beta-lactam, above which the killing effect decreases and has been named as the “paradoxical response”³², although the true significance in the real life of this *in vitro* effect is unknown³³.

Although rare and not reported in Europe yet, resistance to beta-lactams antibiotics in *E. faecalis* can be mediated by the production of a no-inducible beta-lactamase enzyme and may respond to a beta-lactamase inhibitor combination (e.g., ampicillin-sulbactam) plus an aminoglycoside when treating endocarditis and maintaining also sensitivity to carbapenems^{34,35}. Non-beta-lactamase-mediated resistance to ampicillin is quite rare in *E. faecalis*. However, ampicillin plus beta-lactamase inhibitors and imipenem resistance has also been and appears to be associated with mutations of the *pbp4* gene that produce increased expression of low-affinity PBP4 or amino acid changes within the enzyme itself^{36,37}. Conversely, resistance to beta-lactams in most clinical isolates of *E. faecium* is associated with mutations or overproduction of PBP5, with ampicillin MICs > 256 mg/l in some strains³⁸. Molecular epidemiological data suggest that highly ampicillin-resistant strains fall to relatively few lineages that have spread widely, largely in hospitals, causing clinical infections and colonization of patients exposed to a variety of antibiotics. It must be remarked that in many centers of the USA and Europe, rates of high-level ampicillin resistance in *E. faecium* exceed 70-80%. Theoretically, isolates with this phenomenon but MICs < 64 mg/L may respond to high-dose ampicillin therapy (18-30 g per day) that can achieve this threshold.

2.2. Aminoglycosides

Enterococci, due to its outer bacterial wall is relatively impervious to aminoglycosides and are considered structurally resistant to clinically achievable concentrations of these antibiotics³⁹. Most species show low-level aminoglycoside resistance (gentamicin MIC < 1024 and streptomycin MIC <

512 mg/L). However, the combination with cell wall-active agents that blocks peptidoglycan synthesis raise the permeability of the enterococcal wall and markedly increases the uptake of these antibiotics, thus promoting synergy between beta-lactams or vancomycin with gentamycin or streptomycin with good results⁴⁰⁻⁴².

However, the existence of high-level resistance (HLAR) precludes the use of this combination. Acquired resistance include alterations of the aminoglycoside's ribosomal target due to chromosomal mutation (streptomycin)⁴³ and plasmid-mediated resistance genes that encode various aminoglycoside-modifying enzymes, which results in the development of a very high resistance (MICs usually > 2000 mcg/ml)⁴⁴.

The inactivating enzymes may be phosphotransferases, acetyltransferases or nucleotidyltransferases. The most commonly found enzyme is the bifunctional enzyme AAC(6')-Ie-APH(2'') that confers resistance to all available aminoglycosides, except streptomycin. Other enzymes frequently found in HLAR enterococci include ANT(6')-Ia and APH (2'')-Ic which confer resistance to streptomycin and gentamycin respectively. In general, resistance to streptomycin is restricted only to this drug, while resistance to gentamycin implies resistance to all other aminoglycosides except for streptomycin.

2.3. Glycopeptides

Enterococci are considered susceptible to vancomycin and to teicoplanin, that have a long elimination half-life which permits once-daily dosing and has the advantage of not having associated renal toxicity.

Strains of enterococci are considered sensitive to vancomycin if MIC is < 4 mg/L, intermediate when MIC is between 8 and 16, and fully resistant when >16 mg/L. The resistance is attributable to the acquisitions of operons that alter the nature of peptidoglycan precursors, substituting a D-lactate of the terminal D-alanine in the UDP-MurNac pentapeptide. Glycopeptides bind to the terminal D-alanine of the cell wall precursor, preventing PBP access, but pentapeptide stems terminating in D-lactate have a 1000-fold-lower affinity for vancomycin. Different genotypes with resistance to vancomycin and teicoplanin have been described, being the operon *vanA* the most commonly encountered in the clinical setting (Table 1).

The isolation of vancomycin-resistant enterococci (VRE) has steadily increased worldwide since 1986 and nowadays is prevalent (60-80%) among *E. faecium* isolates in USA⁴⁵. In Europe VRE isolates are common in farm animals, feed, and wastewater, and also as colonizers in healthy humans⁴⁶, but are much less frequent in hospitalized patients (although with high variability between countries), probably due to the widespread use of the glycopeptide avoparcin as a growth promoter⁴⁷. However, even after the ban of avoparcin, the European continent has continued to experience an important increase in the isolation of VRE (mostly *E. faecium*) from hospitals, indicating that other factors are promoting the dissemination of VRE isolates, such as hospital clonal clusters, like CC17⁴⁸.

In the case of *E. faecium* and some strains of *E. faecalis*, *vanA* and *vanB* genes play a major role. Fortunately, *E. faecalis* vancomycin-resistant are usually susceptible to beta-lactams, as are *E. gallinarum* and *E. casseliflavus* (which are intrinsically vancomycin-resistant).

2.4. Daptomycin

Daptomycin is a lipopeptide antibiotic approved for the treatment of complicated skin and soft tissue infections and *S. aureus* bacteremia in adult patients, including those with right-sided infective endocarditis. The mechanism of action involves the interaction of the antibiotic with the cytoplasmic membrane via the calcium-dependent insertion, resulting in a variety of alterations in cell membrane characteristics. Daptomycin has dose-dependent bactericidal activity against most Gram-positive agents, including vancomycin and ampicillin-resistant enterococci⁴⁹. The Clinical and Laboratory Standards Institute (CLSI), has recently determined a new "susceptible" breakpoint of ≤ 2 mg/L for *E. faecalis* and a separate "susceptible dose-dependent" breakpoint of ≤ 4 mg/L for *E. faecium*, but indications do not include VRE⁵⁰. However, EUCAST (European Committee on Antimicrobial Susceptibility Testing) daptomycin breakpoint have not been set due to various uncertainties,

particularly the inability of, even the highest published doses (12 mg/kg/day), to achieve adequate exposure against all wild-type isolates of *E. faecalis* and *E. faecium*⁵¹. In fact, emergence of daptomycin-resistant strains with treatment failures has been described with standard dose monotherapy (6 mg/Kg)^{52,53}. Resistance to daptomycin occurs through a variety of mutations that have different effects depending on the species. Much of it is attributed to mutations in several genes including the stress-sensing response system YycFGHIJ and LiaFSR, as also alterations in phospholipid biosynthesis enzymes such as a cardiolipin synthetase *cls* and glycerophosphoryl diester phosphodiesterase *gdpD*^{54,55}.

2.5. Quinolones

The activity of fluoroquinolones against enterococci is moderate and resistance is frequent among clinical isolates. Ciprofloxacin and levofloxacin have marginal activity against enterococci and moxifloxacin is more potent against Gram-positive bacteria but exhibits only intermediate activity versus enterococci⁵⁶. Mutations in the *parC* gene encoding the parC subunit of topoisomerase IV are the first step in the acquisition of resistance, which may be followed by additional mutations in the *gyrA* gene encoding the GyrA subunit of DNA gyrase, thereby increasing the level of resistance⁵⁷. In general, most resistant strains have mutations in the two genes that are related to aminoacidic changes in the Ser83 position of DNA gyrase and in the Ser80 position of topoisomerase IV. Low-level resistance may also be due to alterations in the uptake of these antimicrobials into the bacteria, although specific efflux pumps have not been identified⁵⁸.

2.6. Oxazolidinones

Linezolid is the most common used agent of this class. This drug selectively binds to the 50S ribosomal subunit, thereby resulting in inhibition of bacterial protein synthesis. In general, this resistance is still rare (overall 1-2%), but has been described in *E. faecium* and, most frequently, in *E. faecalis* with higher prevalence in USA and Africa⁵⁹. The resistance is due to mutations in the 23S subunit of ribosomal RNA and ribosomal protein-coding regulatory genes such as *rplC*, *rplD*, and *rplV*, mutations leading to amino acid substitutions on several ribosomal proteins. Enterococci possess multiple copies of the gene encoding this subunit and the higher the number of mutated alleles in this gene, the higher the level of resistance: with a single mutated gene, the MIC of linezolid is 4-8 mg/L, while with 5 mutated alleles, the MIC rises to 64 mg/L⁶⁰. Moreover, enterococci strains have also exhibited the acquisition (via plasmid) of more generic resistance genes such as *cfr* or *cfr(B)*, which encodes a chromosomal methylase that modifies bacterial 23S rRNA⁶¹. This enzyme confers resistance to a variety of antimicrobial classes, including phenicols, lincosamides, oxazolidinones and streptogramin A, as well as decreased susceptibility to the 16-membered macrolides spiramycin and josamycin. Finally, plasmid-mediated resistance has also been attributed to the acquisition of *optrA*, which encodes a putative ABC (ATP-binding cassette) transporter⁶². Most of the reported cases are from patients who had received linezolid for long periods and were selected in the presence of the antibiotic, although clonal dissemination has also been described⁶³.

2.7. Tigecycline

This bacteriostatic drug inhibits protein synthesis upon an interaction with the bacterial 30S ribosomal unit, blocking bacterial protein synthesis at the elongation state, that confers a broad-spectrum therapeutic effect against multi-drug-resistant Gram-positive bacteria including VRE and MRSA in addition to beta-lactamase-producing enterobacteriales and anaerobes. Only a few numbers of resistances have been reported for *E. faecalis* and *E. faecium*, although the emergence of resistant strains seems to be increasing in Europe and Asia⁵⁹. Mutations in various efflux pumps is the main mechanism that is associated with tigecycline-resistance in the enterococci. Other resistance-related mechanisms are deletions in ribosomal protein gene *rpsJ* and elimination of transcriptional regulation of the ribosomal protection protein⁶⁴.

2.8. Therapeutic Choices

The management of enterococcal IE has long been recognized as a challenging clinical problem. Endovascular infections, such as IE are entities in which bactericidal therapy appears to be of paramount importance for eradication of infecting organisms and clinical cure. But unlike the clinical success initially observed with penicillin in the treatment of staphylococcal and streptococcal IE, failures rates with this compound in enterococcal IE was unacceptable (up to 20%)^{30,31}. As it was referred above, the poor performance of penicillin monotherapy has been attributed to the “natural tolerance” of many enterococcal isolates to beta-lactams, which means that they do not achieve a bactericidal effect, even though they inhibit enterococcal growth and are successful in other infections, such as catheter-related bacteremia and those from urinary tract^{25,26}.

The actual recommendations stated in international guidelines^{65,66} are provided in **Tables 2** and **3**. We will focus on them and will consider new alternatives (**Table 4**).

Table 2. American Heart Association guidelines for the treatment of Enterococcus infective endocarditis⁶⁵.

Indication	Recommendation	Dosage and route	Duration (weeks)	Class and Level of evidence	Comments
Strains susceptible to Penicillin and Gentamicin in patients who can tolerate β -Lactam therapy	Ampicillin or penicillin G plus gentamicin	2 g IV every 4 h	4-6	IIa/B	4-wk therapy recommended for patients with native valve and symptoms of illness <3 months.
		18–30 million U/24 h IV either continuously or in 6 equally divided doses	4-6		6-wk therapy recommended for native valve symptoms >3 months
	Ampicillin plus ceftriaxone	3 mg/kg ideal body weight in 2–3 equally divided doses IV	4-6	IIa/B	and for patients with prosthetic valve or prosthetic material.
					Recommended for patients with initial creatinine clearance <50 mL/min or who develop creatinine.
	Ampicillin plus ceftriaxone	2 g IV every 4 h	6	IIa/B	clearance <50 mL/min during therapy with gentamicin-containing regimen.
		2 g IV every 12 h	6		
Strains susceptible to Penicillin and resistant to Aminoglycosides or Streptomycin-susceptible/ Gentamicin-resistant in patients able to tolerate β -Lactam therapy	Ampicillin or penicillin G plus Streptomycin [†]	2 g IV every 4 h	4-6	IIa/B	Use is reasonable only for patients with availability of rapid Streptomycin serum concentrations. Patients with creatinine clearance <50 mL/min or who develop creatinine
		18–30 million U/24 h IV either continuously or in 6 equally divided doses	4-6		clearance <50 mL/min during treatment should be treated with double- β -lactam regimen.
	Streptomycin [†]	15 mg/kg ideal body weight per 24h IV or IM in 2 equally divided doses	4-6	IIa/B	Patients with abnormal cranial nerve VIII function should be treated with double- β -lactam regimen.
Patients unable to tolerate β -Lactam or Penicillin-resistant <i>Enterococcus</i> species and Aminoglycoside-susceptible strains	Vancomycin plus gentamicin [‡]	30 mg/kg per 24 h IV in 2 equally divided doses	6	IIa/B (IIb/C for β -Lactamase-producing strain)	For β -lactamase-producing strain, if able to tolerate a β -lactam antibiotic, ampicillin-sulbactam plus aminoglycoside therapy may be used [§]
		3 mg/kg per 24 h IV in 3 equally divided doses	6		
<i>Enterococcus</i> species caused by strains resistant to Penicillin, Aminoglycosides, and Vancomycin	Linezolid or Daptomycin	600 mg IV or orally every 12 h	> 6	IIb/C	Linezolid use may be associated with potentially severe bone marrow suppression,
		10–12 mg/kg IV per dose	> 6		neuropathy, and numerous drug interactions. Patients with IE caused by these strains should

be treated by a care team including specialists in infectious diseases, cardiology, cardiac surgery, clinical pharmacy, and, in children, pediatrics. Cardiac valve replacement may be necessary for cure.

#: Streptomycin dose should be adjusted to obtain a serum peak concentration of 20 to 35 µg/mL and a trough concentration of <10 µg/mL. Doses recommended for patients with normal renal and hepatic function. Dose of vancomycin should be adjusted to obtain a serum trough concentration of 10 to 20 µg/mL. ¶: Dose of gentamicin should be adjusted to obtain serum peak and trough concentrations of 3 to 4 and <1 µg/mL, respectively. §: Ampicillin-sulbactam dosing is 3 g/6 hour IV.

Table 3. European Society of Cardiology Guidelines for the management of Enterococcus infective endocarditis⁶⁶.

Indication	Recommendation	Dosage and route	Duration (weeks)	Class and Level of evidence	Comments
Beta-lactam and gentamicin-susceptible strains	Amoxicillin/ampicillin plus Gentamycin	200 mg/kg/day IV in 4–6 doses	4–6	I/B	6-week therapy recommended for patients with > 3 months symptoms or PVE. Some experts recommend giving gentamicin for only 2 weeks (IIa/B).
	Paediatric doses: Ampicillin 300 mg/kg/day	3 mg/kg/day IV or IM in 1 dose	2–6		
	Ampicillin plus Ceftriaxone	200 mg/kg/day IV in 4–6 doses	6	I/B	This combination is active against <i>E. faecalis</i> strains with and without HLAR¶, being the combination of choice in patients with HLAR <i>E. faecalis</i> endocarditis.
	Paediatric doses: Ceftriaxone 100 g/kg/12 h IV or IM	4 g/day IV or IM in 2 doses	6		This combination is not active against <i>E. faecium</i>
Intolerance to beta-lactams or beta-lactam resistant# strains	Vancomycin plus Gentamycin	30 mg/kg/day IV in 2 doses	6	I/C	
	Paediatric doses: Vancomycin 40 mg/kg/day IV in 2–3 equally divided doses	3 mg/kg/day IV or IM in 1 dose	6		
Strains with multi-resistance to aminoglycosides, beta-lactams and vancomycin	Ampicillin plus daptomycin	200 mg/kg/day IV in 4–6 doses	> 6	I/C	
		10–12 mg/kg IV per dose	> 6		
	Linezolid	600 mg/day IV or PO	> 6	IIa/C	monitor haematological toxicity
	quinupristin–dalfopristin	7.5 mg/kg IV every 8 hours	> 6	IIa/C	Quinupristin–dalfopristin is not active against <i>E. faecalis</i>

¶: High-level resistance to gentamicin (MIC .500 mg/L): if susceptible to streptomycin, replace gentamicin with streptomycin 15 mg/kg/day in two equally divided doses. #: Beta-lactam resistance: (i) if due to beta-lactamase production, replace ampicillin with ampicillin–sulbactam or amoxicillin with amoxicillin–clavulanate; (ii) if due to PBP5 alteration, use vancomycin-based regimens.

Table 4. Alternatives for treatment in *Enterococcus* infective endocarditis.

Recommendation	Dosage and route	Duration (weeks)	References	Comments
Teicoplanin with/without gentamycin	6–10 mg/Kg/d IV or IM per dose	6–8 2	92,93,94	Good results in experimental models and humans as sequential therapy for IE

3 mg/kg/day IV or IM				
Daptomycin plus ceftaroline or ceftobiprole	10–12 mg/kg IV per dose 400 mg IV every 12 h 500 mg IV every 8 h	6-8	82,83,84, 106,107	Synergistic effects in vitro and experimental models. Very few experience in human IE.
Daptomycin plus imipenem	10–12 mg/kg IV per dose 1 gr IV every 6 h	6-8	107	Synergistic effects in vitro and experimental models
Daptomycin plus Fosfomycin with/without gentamycin	10–12 mg/kg IV per dose 3 g IV every 6 h 3 mg/kg/day IV or IM	6-8	109, 11.112, 113 114,115,116	Synergistic effects in vitro and experimental models. Good results for <i>S. aureus</i> IE
Tedizolid	200 mg IV or PO/24 h		126, 127,128, 129	Less toxic and more active in vitro than linezolid. There is no experience among IE in humans
Ampicillin plus Ciprofloxacin/ofloxacin	2 g IV every 4 h 500 mg/8-12 h	6-8	133,134,135	Very few experience in <i>E. faecalis</i> bacteremia
Tigecycline Plus Daptomycin	50-100 mg/12 h 10–12 mg/kg IV per dose	6-8	142,143,144	Always Consider the use of higher doses of tigecycline
Dalbavancin	1500 mg in a single dose IV, then 1000 mg every two weeks	6-8	150,151,152	Good results in humans as sequentially therapy for IE
Oritavancin	1200 mg in a single dose IV, then 800 mg every week	6-8	158,159,160	Very scarce experience reported in human IE
Amoxicillin plus moxifloxacin	1 g. every 6 h 400 mg every 12 h	4-6	211,212	Oral treatment must be considered as a sequentially strategy in selected patients.
Amoxicillin plus Linezolid	1 g. every 6 h 600 mg every 12 h	4-6	211,212	Oral treatment must be considered as a sequentially strategy in selected patients.
Amoxicillin plus Rifampicin	1 g. every 6 h 600 mg every 12 h	4-6	211,212	Oral treatment must be considered as a sequentially strategy in selected patients.
Moxifloxacin plus Linezolid	400 mg every 12 h 600 mg every 12 h	4-6	211,212	Oral treatment must be considered as a sequentially strategy in selected patients.

2.9. Beta with-lactams aminoglycosides (A + G)

The combination of penicillin plus streptomycin was empirically found to cure the patients who were not improving with penicillin alone and was subsequently shown to have synergistic bactericidal activity in vitro. The development of high resistance to streptomycin (which abolishes synergism) led to the use of gentamycin, an aminoglycoside for which resistance was rare at the time and showed similar results in terms of bactericidal effect and clinical efficacy. Treatment of enterococcal IE with the combination of penicillin plus streptomycin or gentamycin has been evaluated in many studies and became the standard of care many decades ago for patients with IE due to enterococci in the absence of HLAR strains^{30, 40-42, 67,68}.

2.10. Dual beta-lactam therapy (A + C)

Another option that is especially recommended in elderly people, patients with previous renal impairment, and specially, in IE caused by HLAR strains, is the ampicillin + ceftriaxone (A + C)

regimen, which is synergistic in vitro and has also proven effective both in experimental studies and in real life. Its similar efficacy and lower toxicity has led to be included as an alternative regimen in the current guidelines^{65,66}.

In 1995, a French group described an unexpected in vitro synergy between amoxicillin and cefotaxime, an antimicrobial discovered in 1981⁶⁹. Enterococci are naturally resistant to cephalosporins, which act only on Peptide Binding Proteins (PBP) 2 and 3, triggering the production of more efficient PBPs 1, 4 and 5 under treatment. However, aminopenicillins (ampicillin and amoxicillin) and ureidopenicillins (piperacillin) act effectively on these other PBPs, which means that the joint use of both classes of drugs results in a complete blockage and synergy of action in inhibiting bacterial growth. These results were considered by a Spanish group that found identical synergy between ampicillin and another cephalosporin (ceftriaxone) discovered years later with a very favorable pharmacokinetic profile that allowed less frequent administration, due to its high plasma half-life. Gavalda *et al* in 1999 demonstrated a reduction of 1 to 4 dilutions in the MIC of ampicillin of 10 strains of HLRA *E. faecalis* when using the fixed dose of 4 mg/L of ceftriaxone by the double-disk technique and of at least one dilution when using micro dilution in Mueller-Hinton medium for its determination⁷⁰. Using time-death curves, an ampicillin concentration of 2 mg/L and varying concentrations of 5-60 mg of ceftriaxone, they achieved a > 2 log reduction of the initial inoculum at 24 h of incubation in all strains, and this effect increased to > 3 log (synergy) in 7 of the 10 strains when 10 mg/L doses of ceftriaxone were used, and in 6 strains when the concentration was 5 mg/L. Similar results were reported much later, showing that even ampicillin concentrations at 1 mg/L + 2 mg/L ceftriaxone were synergistic⁷¹. Using a humanized model in the experimental animal, the authors found that by administering the equivalent of 2 g of IV ceftriaxone, drug concentrations at 12 h were around 50 mg/L and 20 mg/L at 24 hours (antibiotic bound to proteins). As ceftriaxone bound to protein at 90%, the administration of 2 g IV/12 of ceftriaxone, together with the administration of ampicillin, could always achieve free-drug concentrations above 2-4 mg/L, guaranteeing this synergistic effect during the entire dosing interval. These facts were effectively translated into a clear decrease in the colony count in the vegetations of the experimental animals treated with this regimen, and even the complete sterilization of the vegetations in some animals infected with certain strains.

The translation of this elegant experimentation on the animal model was published eight years later by the same group in a multicenter trial in 13 Spanish hospitals, in which 21 patients with HLRA strains and 22 with non-HLRA strains at high risk of nephrotoxicity were treated⁷². Cure was obtained in 100% of patients with HLRA strains who completed the protocol with this regimen. Due to the small sample size and the failures obtained in non-HLRA strains, the guideline was promptly considered but only for HLRA strains, due to the lack of existing alternatives. However, in 2003 the same group demonstrated, again in the experimental animal model, the efficacy of this pattern also in non-HLRA strains⁷³. And again ten years later, the demonstration of its clinical efficacy in these non-HLRA strains compared to the "standard" treatment (ampicillin + gentamicin) came from a Spanish multicenter trial, in which 150 patients treated with the A + C regimen (ampicillin + ceftriaxone) were compared to 87 patients treated with A + G (ampicillin + gentamicin), showing equal mortality (22% *vs* 21% during hospitalization and 8% *vs* 7% at 3 months) and recurrences (3% *vs* 4%), but with lower nephrotoxicity (23% *vs* 0%; $p < 0.01$)⁷⁴. Following these excellent results, the American and European guidelines included this regimen as "preferred" in HLRA strains and "alternative" in non-HLRA strains^{65,66}. However, in Europe and especially in Spain, the A + C regimen progressively gained followers until it became the majority⁷⁵, and this was easily explained by its lower toxicity, since the profile of the patient with enterococcal IE is precisely that of an elderly patient, with abundant comorbidities and in many cases previously weakened by a previous hospitalization, and it is precisely in this group where the physician must be especially cautious about the side effects of treatment^{4-6,13,14}.

One concern with ceftriaxone use is that it has been pointed as an independent risk factor for *Clostridium difficile* infection⁷⁶ although no such complication has been reported from Spain in which this guideline is prevalent. Another concern is that some clinical and observational studies implicate the use of ceftriaxone as major risk factor for occurrence of vancomycin-resistant *E. faecium* infection,

including bacteremia^{77,78}. In animal studies, ceftriaxone use promotes gastrointestinal colonization by VRE, probably due to the high biliary excretion of ceftriaxone that could select for drug-resistant enterococci living there^{79,80}. Unlike ceftriaxone, other cephalosporins antibiotics, such as cefepime⁷⁹, and ceftaroline⁸¹ do not appear to promote VRE colonization, and the combination of ampicillin plus ceftaroline have demonstrated efficacy similar to ampicillin plus ceftriaxone in several pharmacodynamic studies, although no clinical data are yet available^{82,83}. Ceftobiprole have high affinity for enterococcal PBPs and have demonstrated efficacy against VanB-resistant *E. faecalis* in addition to synergy when used in combination with aminoglycosides, but this combination requires further exploration in human subjects⁸⁴.

2.11. Glycopeptides

Vancomycin is an alternative therapy recommended in European and American guidelines but should be administered only if a patient is unable to tolerate penicillin or ampicillin. Vancomycin reduced CFU/ml in vegetations significantly more than ampicillin monotherapy in the rabbit experimental model⁸⁵. However, combinations of penicillin or ampicillin with gentamicin are preferable to combined vancomycin-gentamicin because of the potential increased risk of ototoxicity and nephrotoxicity with the vancomycin plus gentamicin combination during six weeks. Moreover, combinations of penicillin or ampicillin and gentamicin are more active than combinations of vancomycin and gentamicin *in vitro* and in animal models of experimental IE⁸⁶. Rarely, strains of *E. faecalis* produce an inducible β -lactamase. These β -lactamase-producing strains are susceptible to ampicillin-sulbactam and to vancomycin. Intrinsic penicillin resistance is today (especially in the USA) common in *E. faecium*, but scarce in *E. faecalis*.

Teicoplanin is particularly interesting due to the lack of renal toxicity and the *in vitro* data that demonstrate advantage over vancomycin against *E. faecalis*, with MIC₉₀ values usually lower than that for vancomycin⁸⁷⁻⁸⁹. Furthermore, its long-elimination half-life permits once-daily dosing and exhibit concentration-dependent activity with excellent results in experimental studies combined with gentamicin^{90,91}. Several observational studies (overall 56 patients) support the use of monotherapy with teicoplanin at doses of 6-10 mg/kg/d (two of them also introduced a loading dose), mainly as a continuation treatment for *E. faecalis* IE when adverse events have developed with standard treatments, or to facilitate outpatient treatments⁹²⁻⁹⁴. The largest study conducted embraced the use of monotherapy with teicoplanin for treating *E. faecalis* IE as continuation therapy. The reported mortality related to IE was low (8%), but the population treated with teicoplanin suffered from less severe IE than the standard therapy group⁹³. Within the patients treated with teicoplanin as a continuation or salvage therapy, 16 died (32%) in a minimal follow-up period of 3 months. Only three relapses were reported in these studies. Then, favorable results and very few toxicities lead us to consider it as a reasonable alternative. Theoretically, teicoplanin has also activity against enterococci with VanB mediated resistance, but development of resistance during therapy is concerning⁹⁵ and cannot be a recommendation in this setting.

2.12. Daptomycin

Although there are no prospective randomized-controlled studies evaluating the efficacy of daptomycin for the treatment of IE, several reports including a total of 26 patients were published shortly after its approval, within an "off-label" use⁹⁶⁻⁹⁸. The treatment scheme was considerably heterogeneous, included initial and salvage therapy, monotherapy and combined regimens, and the mean doses ranged between 8.5 and 10.125 mg/kg/day. Mortality rates reported were low (0–22%), although only one study⁹⁷ attained more than a one-month follow-up. In one study, the salvage treatment of five *E. faecalis* IE episodes was reported⁹⁶, of which four needed a treatment change due to treatment failure. Daptomycin patients had longer duration of bacteraemia (6 versus 1 day) and greater need of therapy switch due to complications (66.7% versus 0%) compared with conventional antibiotic regimens (ampicillin or vancomycin \pm gentamicin), although there were no differences regarding duration of hospital stay or mortality. So, the stated final conclusions differed, with two supporting daptomycin as an alternative treatment in this scenario^{97,98}, and one showing some

concerns⁹⁶. Among 22 patients with enterococcal IE treated with daptomycin in a European registry (18 *E. faecalis*), the success rate was 73%, but no information regarding dosage or combination therapy was given⁹⁹. An observational prospective single centre study found similar outcomes in patients with enterococcal endocarditis treated with daptomycin-based regimen versus standard regimens, although daptomycin was used in combination with another antibiotic (mostly a beta-lactam) at high doses (>10 mg/kg/day)¹⁰⁰. Microbiological failures of daptomycin were promptly reported when “standard” doses (6 mg/Kg/day, approved dose for *S. aureus* bacteremia and right-sided IE) were administered, and high doses (8-12 mg/Kg/day) are now recommended for enterococcal and *S. aureus* severe infections⁵⁰⁻⁵³. It is important to note that the daptomycin MIC₉₀ for enterococci is higher than that of staphylococci (4 mg/L and 0.5 mg/L, respectively), supporting the concept that higher doses of daptomycin may be needed for the management of enterococcal IE, and *in vitro* studies have demonstrated that a high percentage (33%) of *E. faecalis* incubated with daptomycin at a subinhibitory concentration (2 mg/L) can develop MIC \geq 8 mg/L¹⁰¹. Daptomycin display a dose-dependent bactericidal effect and high-dose regimens have demonstrated an enhanced pharmacodynamic profile and the most bactericidal regimen against VRE¹⁰². However, microbiological failures also have been described with high doses in patients with prior daptomycin exposures, prostheses, or immunocompromised patients with long hospitalization courses^{53,103,104}. Therefore, this alternative could be considered in resistant isolates or when adverse events appear, but not so simplify antibiotic treatment. Taking also account the synergistic activity between daptomycin and beta-lactams¹⁰⁵⁻¹⁰⁸ or tigecycline, a combination regimen with high doses seems to be preferable, whereas single therapy with this drug should be used with caution.

2.13. Fosfomycin

In vitro data have demonstrated synergy with fosfomycin in combination with ceftriaxone, rifampin, tigecycline and teicoplanin¹⁰⁹⁻¹¹². Current oral fosfomycin use is limited to uncomplicated urinary tract infections due to limited absorption and intravenous formulation are yet unavailable in the USA. However, fosfomycin has demonstrated utility against MSSA and MRSA endocarditis in combination with daptomycin or imipenem¹¹³⁻¹¹⁶ and a study of in vitro and in vivo with the guinea pig model using intraperitoneal fosfomycin demonstrated promising activity against both planktonic and biofilm-forming *E. faecalis* when the drug was used in combination with gentamicin and daptomycin¹¹⁷. Thus, new therapeutic options with this drug could be considered in the future for *E. faecalis* IE.

2.14. Linezolid and Tedizolid

Linezolid after a few promising studies has been recommended for the treatment of endocarditis as result of multi-drug resistant enterococci¹¹⁸ and has been recommended in the USA for the treatment of Enterococcus species caused by strains resistant to penicillin, aminoglycosides, and vancomycin⁶⁵. Regrettably, widespread use from the year 2000 has result in an emerging of linezolid-resistant VRE in 2001¹¹⁹ and increasing of these strains especially in hospitals from various countries (Denmark, Spain, Germany...)¹²⁰. However, the use of linezolid is a matter of controversy because of the lack of bactericidal effect and the lack of randomized clinical trials or robust cohorts. Linezolid has displayed efficacy in the treatment of VRE faecium bacteremia with an open-label nonrandomized, compassionate-use program reporting microbiological and clinical cure rates of 85.3% and 79% respectively with 10 out of 13 patients with VRE IR (76.9%) achieving clinical cure in the subgroup of endocarditis¹²¹. A systematic review attempted to evaluate the clinical efficacy of linezolid in the treatment of enterococcal IE. This study found that 7 out of 8 cases improved or were cured with linezolid: four of the included cases were caused by *E. faecalis* (two VRE) and the rest of them were cases of IE vancomycin-resistant *E. faecium*¹²². But clinical evidence is supported only by these case reports with heterogeneous results and numerous cases of enterococcal infections resistant to linezolid have been reported¹²³⁻¹²⁵. Another drawback is the need of a prolonged treatment (six weeks) of *E. faecalis* IE that usually occurs in elderly population, which is more likely to the myelotoxicity and neuropathy produced by this drug.

Gastrointestinal disorders and myelotoxicity are less frequent with tedizolid, and a favorable action against MRSA IE have been reported in experimental models (rats and rabbits). Against VRE tedizolid has a fourfold lower MIC when compared to linezolid and has activity against linezolid-resistant strains with a *cfr* mutation, probably due to additional interactions with the ribosomal¹²⁶⁻¹³¹. Thus, compared to linezolid, tedizolid has the potential to be a first-line agent for the treatment of serious VRE infections, but until now, no clinical data have been published in patients with IE.

2.15. Quinolones

Fluorquinolones have been used in the treatment of some enterococcal infections such as chronic enterococcal prostatitis with relapsing bacteremia. Similar to the tetracyclines, these antibiotics have also been used as part of combination therapies in endocarditis. The combination of ampicillin plus ciprofloxacin was tested in an experimental model of rabbit endocarditis with *E. faecalis* and the regimen caused a significant decrease in bacterial counts compared to each compound alone, although it was less effective than the combination of beta-lactams and aminoglycosides¹³³, and this effect was previously reported *in vitro*¹³⁴. Additionally, the use of ampicillin plus ofloxacin was shown to be also synergistic *in vitro*, achieving bactericidal activity, and to successfully clear the bacteremia in a patient with *E. faecalis* IE exhibiting HLAR¹³⁵. Nonetheless, the lack of clinical experience and the increased rates of resistance to some of these compounds usually preclude the use of these antibiotics for *E. faecalis* IE, particularly as monotherapy.

Delafloxacin is a new quinolone active *in vitro* against MSSA, MRSA, CoNS and streptococci and interestingly retains activity against fluoroquinolone-resistant strains. Specific features in the delafloxacin molecule determines enhanced activity in acidic environment due to its anionic character, which eventually leads to improved activity¹³⁶. Delafloxacin has been recently approved for acute bacterial skin and skin structure infections¹³⁷ and for the treatment of community-acquired bacterial pneumonia¹³⁸. Delafloxacin can be given intravenously or orally due to its good bioavailability (60-70%)¹³⁷. However, according to EUCAST, there is insufficient evidence that enterococci are a good target for therapy with delafloxacin and no clinical data have been published on the use of delafloxacin for IE.

2.16. Tigecycline

Tigecycline is a broad-spectrum antibiotic derived from minocycline which is approved for skin and soft tissue infections, including those with vancomycin-susceptible *E. faecalis*. In the management of soft tissue infections (including those with vancomycin-susceptible *E. faecalis*), tigecycline showed a microbiological eradication rate of 87.5%, similar to vancomycin plus aztreonam¹³⁹. In complicated abdominal infections, tigecycline exhibit similar rates of microbiological curation for vancomycin-susceptible *E. faecalis* (78.8%) than imipenem¹⁴⁰. Moreover, some *in vitro* models suggest that synergism of tigecycline with vancomycin, gentamicin and rifampin can be achieved for certain strains of *E. faecalis* and *E. faecium*¹⁴¹ and successful therapy of endocarditis was reported with the combination of tigecycline plus daptomycin in several cases¹⁴²⁻¹⁴⁴. However, a serious drawback of the use of tigecycline monotherapy is the low levels obtained with this antibiotic¹⁴⁵ and the emergence of resistance during therapy has been reported in experimental studies¹⁴⁶. Thus, the use of this compound as monotherapy for severe enterococcal infection is discouraged, although its role in combination therapies with bactericidal effects warrants further investigation.

2.17. Dalbavancin and Oritavancin

Considered a subclass of the glycopeptide antibiotics, the new lipoglycopeptides have similar mechanisms of action of binding to the carboxyl terminal d-alanyl-d-alanine residue of the growing peptide chains but differ from their parent glycopeptides by the addition of lipophilic tails. This addition allows for these agents to have prolonged half-lives, giving them unique dosing profiles.

Dalbavancin has a long-acting parenteral administration due to its high-protein binding (93%) and prolonged elimination half-life (14.4 days), that allows prolonged intervals between doses (7-14 days)¹⁴⁷. A promising activity against Gram-positive biofilms has also been reported¹⁴⁸. Although it has not been approved to treat patients with bloodstream infections or IE, there are in vitro studies showing a good susceptibility (MIC₉₀: 0.06 mg/L) of most *E. faecalis* isolates (97.6%) collected from patients with a diagnosis of bacterial endocarditis¹⁴⁹.

Dalbavancin was approved in the USA and Europe to treat acute bacterial skin and skin structure infections caused by Gram-positive cocci isolates, including vancomycin-susceptible *E. faecalis*, but it must be remarked that it is inactive against *VanA* phenotypes. Off-label utilization of dalbavancin was extensively done in patients with osteomyelitis, joint infections, and articular prostheses, and less in cardiovascular infections¹⁵⁰. A retrospective cohort in Austria evaluated 27 adults with Gram-positive bacteremia with IE treated at least with one dose of dalbavancin with excellent results¹⁵¹. In most patients dalbavancin was used as sequential treatment after clearance of bacteremia to allow OPAT and the same scheme was used in a Spanish cohort that included 34 patients (3 of them with *E. faecalis* IE)¹⁵². Two dosing strategies are used with similar results: a 1000 mg loading dose and the 500 mg/week or a 1500 mg loading dose and then 1000 mg every two weeks and in these cohorts, six patients were successfully treated.

Thus, limited available evidence suggests that dalbavancin might be a good option to treat *E. faecalis* IE in the context of OPAT, but studies with full use (not only sequentially) of this drug throughout treatment are needed.

Oritavancin is an interesting drug, very active against Gram-positive cocci including enterococci, and that also retains some activity in presence of *VanA* and *VanB*-mediated vancomycin resistance. Among two collections of more than 10,000 isolates, oritavancin showed potent in-vitro activity against staphylococci (including MRSA), streptococci and enterococci^{153,154}. Although higher MIC were registered against vancomycin-resistant *E. faecalis* (*vanA* phenotype) than against vancomycin-susceptible strains, all *VanA*-resistant *E. faecalis* were inhibited at 0.5 mg/L or less. Its high protein-binding (85-90%) and the prolonged terminal half-life (200-300 h) permits the administration of a single dose of 1,200 mg with good therapeutical levels beyond two weeks¹⁵⁵ and a good activity in biofilms¹⁵⁶. After the initial dose of 1,200 mg, sequentially doses of 800 mg can be administered weekly for infections that will require a more prolonged treatment such as osteomyelitis¹⁵⁷.

Elimination of oritavancin mainly occurs through the reticuloendothelial system, thus no adjustments of dosage are needed in the cases of renal or hepatic failure. Notably, oritavancin is a weak inhibitor of CYP2C9 and CYP2C19, and an inducer of CYP3A4 and CYP2D6, thus drug-drug interactions (e.g., patients treated with warfarin) should always be considered. Furthermore, attention should be paid to the possible alterations of some coagulation tests in the first hours/days after oritavancin administration because of its interaction with the phospholipid reagent. Prolonged prothrombin and active partial thromboplastin times have been reported and for this reason, the use of intravenous unfractionated heparin sodium is contraindicated for up to 5 days after oritavancin administration owing to the inability to reliably monitor coagulation tests. The results of chromogenic factor Xa and the thrombin time assays are not affected.

Oritavancin was also approved in the USA and Europe to treat adults with acute bacterial skin and skin structure infections caused by Gram-positive cocci isolates, including vancomycin-susceptible *E. faecalis*, but several reports with its utilization in osteoarticular infections, bacteremia and very few endocarditis have been reported¹⁵⁸⁻¹⁶¹.

In conclusion oritavancin seems also an important option for outpatient therapy and early discharge in patients with *E. faecalis* IE, but very limited number of papers are available, especially in this setting, and there is no experience with quantity and dosing schedule and duration of treatment. Thus, further studies are necessary for optimizing and refining its place in the treatment of IE.

2.18. Duration of treatment

One of the difficulties in the treatment of *E. faecalis* IE lies in its prolonged duration (4-6 weeks) and the fact of having to combine two antimicrobials to achieve synergy, which must also be

administered in several doses/day. Treatments of less than six weeks have been associated with a greater number of recurrences¹⁶², although in certain patients (endocarditis on native valve and absence of paravalvular extension) four weeks are probably enough¹⁶³. A shorter treatment with aminoglycosides has already been mentioned by some authors¹⁶⁴ and was later confirmed by a Swedish study¹⁶⁵ in which no differences were found between 2 and 4 weeks of administration. This same fact was endorsed by a Danish study¹⁶⁶ which, when administered over a short period (2 weeks), also found no difference between intermittent administration and administration in a single daily dose. No differences between 2 or 4 weeks were found in complications, relapse, in-hospital mortality, or 1-year event free survival, but patient patients receiving 2 weeks treatment with gentamycin therapy experienced a significantly reduction in renal function at discharge, compared to those receiving the full course, as measured by estimated glomerular filtration. Although this study was limited by a small size and insufficient power to establish the optimal duration of aminoglycoside treatment, other studies have shown that gentamycin nephrotoxicity increases with duration of treatment¹⁶⁷ and its discontinuation occurred frequently after 14-18 days of treatment^{74,75}. It is also be noted that the decrease in renal function is only partly reversible^{168,169}. Then, a 2-week treatment course of aminoglycosides seems reasonable taking account the clinical picture of most patients with *E. faecalis* IE (elderly, fragile and with high degree of comorbidity)^{5,12,14, 165, 166, 170}.

2.19. Outpatient Parenteral Antimicrobial Therapy (OPAT) with the standard guidelines

Outpatient parenteral antimicrobial treatment constitutes one of the most recent advances in the treatment of infective endocarditis. After the first two weeks, the septic process is usually well controlled (lack of fever and negative blood cultures), the risk of embolism is dramatically reduced^{171,172}, and the possible complications derived from the structural damage of the heart have been properly assessed by echocardiographic studies. Then the possibility of further complications is greatly reduced, and home treatment brings undeniable advantages in terms of comfort for the patient and their environment, cost savings as well as avoiding nosocomial infections¹⁷³. In 2019, the GAMES group (Grupos de Apoyo al Manejo de la Endocarditis en España) established much less strict criteria than the original recommendations for this therapeutic approach^{174,175}, but which proved to be equally safe and allow outpatient treatment of *S. aureus* (including methicillin-resistant strains, MRSA) and *Enterococcus* spp¹⁷⁶.

The "standard" guidelines (A + G) recommended in both American and European guidelines make this option almost impossible on an outpatient basis, since they recommend the administration of aminoglycosides every 8-12 hours and monitoring the levels. We would therefore need two routes (one of which would be for the continuous or intermittent administration of ampicillin by means of a perfusion pump) or a well-trained family member capable of performing (two or three times a day) these manipulations, in addition to the logistics inherent to the monitoring of levels. However, these recommendations are based on experimental studies¹⁷⁷⁻¹⁸⁰, sometimes contradictory^{181,182} possibly due to differences related to the different pharmacokinetics of the experimental model and humans. On the other hand, there are abundant studies that show the efficacy of single-dose administration of aminoglycosides, with excellent activity due to their prolonged post-antibiotic effect and much less renal toxicity¹⁸³. Therefore, the translation of these experimental results to humans, as reflected in the guidelines, is far from being justified by a hypothetical greater efficacy, considering the known increase in toxicity of aminoglycosides in intermittent administration for 4-6 weeks¹⁸⁴. Therefore, a valid option would be the administration (continuous or intermittent every 4 hours) of ampicillin by pump, in addition to a single dose of gentamicin, which could be done at the time of refilling the pump.

The A + C regimen arises from the finding of synergy between the two drugs, when free drug levels of ceftriaxone between 5 - 10 mg/L are achieved. Based on experimental data derived from the humanized rabbit model, doses of 2 g/12 h in humans are recommended⁷³. For OPAT programs, this implies the difficulty of an administration of ceftriaxone every 12 hours, which is not always easy due to the availability of the patient or caregiver to manipulate the system. Our group, based on theoretical concentrations of 30 mg/L (total drug) at 24 h when a single dose of 4 g was used, began

to administer this regimen (A + C24) in enterococcal IE, with good initial results¹⁸⁵. In parallel and given the absence of pharmacokinetic data with this dose, we conducted a clinical trial with healthy volunteers where we analyzed the pharmacokinetics of ceftriaxone administration at a dose of 2 g. IV/12 h (C12) and 4 g. IV/24 h (C24). Unfortunately, we observed that administration of a single dose of 4 g. did not achieve the target levels above 5 mg/L of free drug in any subject at 24 hours, in very few at 18 hours, and only in half at 12 hours¹⁸⁶. As previously reported in trials in healthy volunteers, administration every 12 hours did not achieve these levels at 24 hours in most individuals, although it did at 18 hours in half of them^{187,188}. However, the good clinical results reported with this regimen (A + C12) suggested that these "target" concentrations of ceftriaxone might not be necessary during the whole time but at least during 75% of the dosing interval, which could be achieved, however, only with administration every 12 hours. Administration every 24 h, however, showed clearly poor pharmacokinetics, and further dose escalation (6-7 g) did not seem to be a good solution either, as we tested in Monte Carlo simulation models. The binding of ceftriaxone to plasma albumin is very high (90%) and dose-dependent, but saturable, and so administering a very high dose of the drug to over 100 mcg/ml would result in high levels of free drug after infusion. This effect could be convenient in certain scenarios, i.e., infections of the central nervous system¹⁸⁹, where only the free fraction of the drug would cross the blood-brain barrier, but it would also mean that a large amount of this drug would be rapidly excreted by the kidney. So that, after a prolonged period (24 h) the amount of free drug available would be the result of its constant release from binding to albumin, like when lower doses are administered. It should be noted, however, that our data came from measurements in healthy volunteers, and it is possible that in real patients with enterococcal IE, usually elderly and with a lower renal clearance rate, ceftriaxone levels would be higher during interdoses^{190,191}. Unfortunately, our fears were confirmed by an unexpected failure rate (5/17; 29.4%) when we used the A + C24 regimen in the following patients. Although it is true that there are other factors that predispose to failure, such as the fact that the infection settles on prosthetic material (almost double the recurrence rate), or not performing surgery when there is a structural complication (e.g. an abscess of the ring), in our cohort we observed that patients who had complied with the A + C12 regimen while hospitalized for at least 3 weeks before switching to outpatient A + C24 had a much lower number of recurrences¹⁹².

A recently reported pharmacokinetic/pharmacodynamic study simulating human doses of ampicillin and ceftriaxone has remarked the usefulness of this combination administering ceftriaxone every 12 hours^{193,194}. This prompted us to abandon this regimen and consider other alternatives. An attractive option could be the co-administration of ampicillin + ceftriaxone in the same infusion. We know that ampicillin is stable in infusion for more than 24 h. at room temperature, and there are abundant data reported on adequate plasma levels in both continuous and intermittent infusion¹⁹⁵⁻¹⁹⁷. However, there were doubts about the stability of ceftriaxone at room temperature and its interaction with other beta-lactams¹⁹⁸. However, using a very precise technique¹⁹⁹, we demonstrated the stability of this combination that we believe it would be an excellent alternative (much less toxic) to the A + G regimen, which is still used as a reference. Indeed, continuous or intermittent administration (every 4 hours) of both drugs would always maintain levels above the required thresholds, without requiring any extraordinary manipulation other than periodic pump refilling^{200,201}.

Other alternatives for OPAT in *E. faecalis* IE have been previously analyzed in a previous report²⁰⁰.

2.20. Oral treatment

Current guidelines for management of infective endocarditis (IE) usually advise 4–6 weeks of IV antibiotics. This is based on historical data from animal models, which set a precedent for high peak serum antimicrobial levels, thought to be only achievable with IV therapy. However, there has been increasing recent interest in oral antibiotics as an alternative to prolonged parenteral therapy. Intravenous antibiotics offer immediate complete bioavailability, allowing peak serum levels to be achieved rapidly. This is desirable in critically unwell septic patients and is also a necessity in patients

who are unable to take medications enterally, or where there are concerns about absorption. In addition, antimicrobial susceptibility may require antibiotics that can only be given IV, such as aminoglycosides or glycopeptides. However, antibiotics given orally with a good bioavailability and with favorable pharmacokinetic and pharmacodynamic properties with standard doses, would provide effective antimicrobial therapy for the treatment of endocarditis caused by susceptible microorganisms. The advantages compared with outpatient parenteral treatment is that oral antibiotic therapy may reduce costs and minimize challenges associated with OPAT including logistics, monitoring and risks of complications associated with intravenous catheters (e.g., bleeding, local and systemic infections, and venous thrombosis). Reports of oral treatment in IE are scarce and mainly focused on streptococci and staphylococci from right-sided IE²⁰³⁻²¹⁰. More recently, a Danish trial^{211,213} evaluated the efficacy and safety of sequential switching to oral antibiotic treatment in stable patients with IE after at least 10 days of parenteral therapy and, among patients who had undergone valve surgery for at least 7 days after the operation. From the 201 patients that were randomized to oral therapy, 51 (25.4%) were *E. faecalis* IE and the obtained primary outcome (a composite of all-cause mortality, unplanned cardiac surgery, embolic events, or relapses) seemed similar across the four different bacterial types (*E. faecalis*, streptococcus, *S. aureus* and coagulase-negative staphylococci). Regarding *E. faecalis*, four patients with oral treatment (4/51; 7.8%) presented the primary outcome compared with seven (7/46; 15.2%) in the parenteral treatment. To address the risk of subtherapeutic antibiotics levels related to potentially reduced gastrointestinal uptake, as well as the risk of variations in pharmacokinetics of the orally administered antibiotics, all oral regimens included two antibiotics from different drug classes and with different antibacterial effects, and different metabolism processes. Indeed, in seven patients in the orally treated group, the plasma concentration of one of the two administered antibiotics was not at the most effective level, as assessed by peak levels and time above the MIC, although the plasma concentration of the other simultaneously administered antibiotic was appropriate. The primary outcome did not occur in any of these patients and no antibiotic regimen was changed due to this finding.

However, despite its favorable results that have maintained over time^{214,215}, it should be remarked that only 400 (20%) of the 1954 screened patients underwent randomization, and that patients were orally treated a median of 17 days after intravenous treatment. Furthermore, 15 patients in the oral group previously went to heart valve surgery. Other aspect is the combinations of high doses of amoxicillin, linezolid, rifampin, or moxifloxacin may represent a challenge in elderly patients most prone to associated side effects. Thus, precaution is necessary when interpreting the POET trial regarding *E. faecalis* IE and further clinical trials focused on this group seems necessary for establishing a robust recommendation.

3. Conclusions and future perspective

Although aminoglycoside containing regimens have been the standard of enterococcal IE treatment, the rise in resistance and availability of less nephrotoxic agents have led to novel treatment options. Double beta-lactam therapies have emerged as a novel strategy in the treatment of serious high-inoculum enterococcal infections due to their favorable side effect profiles and tolerability during long-term use. Currently, ampicillin plus ceftriaxone is the only beta-lactam therapy supported by robust clinical data and the main option for HLAR strains and elderly population, although other beta-lactam combinations supported by in vitro studies could be possible and must be explored. However, no recommendation can be done for non-HLAR strains that could also benefit from treatment with ampicillin plus a short course of gentamicin (2 weeks) and randomized trials will be necessary for a solid recommendation. Combinations of beta-lactams with daptomycin or fosfomycin are promising but needs further investigation. Other alternatives such as teicoplanin (with or without gentamicin), are interesting, especially in patients allergic to beta-lactams, because its lower renal toxicity and once-daily administration that favors OPAT. In this setting, the development of long-acting lipoglycopeptides (dalbavancin and oritavancin) has represented a considerable advance in security and shortening hospitalization times. Finally, oral treatment

combining amoxicillin with linezolid or fluoroquinolones is also an alternative for continuation treatments but requires further investigation in this type of IE.

References

1. Murdoch DR, Corey GR, Hoen B, Miró JM, Fowler VG Jr, Bayer AS, et al. Clinical presentation, etiology, and outcome of infective endocarditis in the 21st century: the International Collaboration on Endocarditis-Prospective Cohort Study. *Arch Intern Med*. 2009; 169:463–73.
2. Habib G, Erba PA, Iung B, Donal E, Cosyns B, Laroche C, Popescu BA et al. Clinical presentation, aetiology and outcome of infective endocarditis. Results of the ESC-EORP EURO-ENDO (European infective endocarditis) registry: a prospective cohort study. *Eur Heart J*. 2019 Oct 14;40(39):3222–3232.
3. Muñoz P, Kestler M, De Alarcón A, Miró JM, Bermejo J, Rodríguez-Abella H et al. Current Epidemiology and Outcome of Infective Endocarditis: A Multicenter, Prospective, Cohort Study. *Medicine (Baltimore)*. 2015 Oct;94(43): e1816.
4. Fernández-Hidalgo N, Escolà-Vergé L, Pericàs JM. Enterococcus faecalis endocarditis: what's next? *Future Microbiol*. 2020 Mar; 15:349–364.
5. Pericàs JM, Llopis J, Muñoz P, Gálvez-Acebal J, Kestler M, Valerio M et al. A Contemporary Picture of Enterococcal Endocarditis: a comparison of 516 cases with 308 cases of non-enterococcal endocarditis from the GAMES Cohort (2008–2016). *J Am Coll Cardiol*. 2020 Feb 11;75(5):482–494.
6. Dahl A, Fowler VG, Miro JM, Bruun NE. Sign of the Times: Updating Infective Endocarditis Diagnostic Criteria to Recognize Enterococcus faecalis as a Typical Endocarditis Bacterium. *Clin Infect Dis*. 2022 Sep 29;75(6):1097–1102.
7. Fernández-Guerrero ML, Herrero L, Bellver M, Gadea I, Roblas RF, de Górgolas M. Nosocomial enterococcal endocarditis: a serious hazard for hospitalized patients with enterococcal bacteraemia. *J Intern Med*. 2002; 252: 510–515.
8. Lomas JM, Martínez-Marcos FJ, Plata A, Ivanova R, Gálvez J, Ruiz J et al. Healthcare-associated infective endocarditis: an undesirable effect of healthcare universalization. *Clin Microbiol Infect*. 2010 Nov;16(11):1683–90.
9. Pericàs JM, Ambrosioni J, Muñoz P, de Alarcón A, Kestler M, Mari-Hualde A et al. Prevalence of Colorectal Neoplasms Among Patients With Enterococcus faecalis Endocarditis in the GAMES Cohort (2008–2017). *Mayo Clin Proc*. 2021 Jan;96(1):132–146.
10. Escolà-Vergé L, Peghin M, Givone F, Pérez-Rodríguez MT, Suárez-Varela M, Meije Y et al. Prevalence of colorectal disease in Enterococcus faecalis infective endocarditis: results of an observational multicenter study. *Rev Esp Cardiol (Engl Ed)*. 2020 Sep;73(9):711–717.
11. Khan A, Aslam A, Satti KN, Ashiq S. Infective endocarditis post-transcatheter aortic valve implantation (TAVI), microbiological profile and clinical outcomes: A systematic review. *PLoS One*. 2020 Jan 17;15(1): e0225077.
12. McDonald JR, Olaison L, Anderson DJ, Hoen B, Miro JM, Eykyn S et al. Enterococcal endocarditis: 107 cases from the international collaboration on endocarditis merged database. *Am J Med*. 2005 Jul;118(7):759–66.
13. Fernández Guerrero ML, Goyenechea A, Verdejo C, Roblas RF, de Górgolas M. Enterococcal endocarditis on native and prosthetic valves: a review of clinical and prognostic factors with emphasis on hospital-acquired infections as a major determinant of outcome. *Medicine (Baltimore)*. 2007 Nov;86(6):363–377.
14. Martínez-Marcos FJ, Lomas-Cabezas JM, Hidalgo-Tenorio C, de la Torre-Lima J, Plata-Ciézar A, Reguera-Iglesias JM et al. Enterococcal endocarditis: a multicenter study of 76 cases. *Enferm Infecc Microbiol Clin*. 2009 Dec;27(10):571–9.
15. Conwell M, Dooley JSG, Naughton PJ. Enterococcal biofilm-A nidus for antibiotic resistance transfer? *J Appl Microbiol*. 2022 May;132(5):3444–3460.
16. García-Solache M, Rice LB. The Enterococcus: a Model of Adaptability to Its Environment. *Clin Microbiol Rev*. 2019 Jan 30;32(2): e00058–18.
17. C H Heath, T K Blackmore, D L Gordon. Emerging resistance in Enterococcus spp. *Med J Aust*. 1996 Jan 15;164(2):116–20.
18. Casalta JP, Thuny F, Fournier PE, Lepidi H, Habib G, Grisoli D, Raoult D. DNA persistence and relapses questions on the treatment strategies of Enterococcus infections of prosthetic valves. *PLoS One*. 2012;7(12): e53335.

19. Lecomte R, Laine JB, Issa N, Revest M, Gaborit B, Le Turnier P et al. Long-term Outcome of Patients with Non-operated Prosthetic Valve Infective Endocarditis: Is Relapse the Main Issue? *Clin Infect Dis*. 2020 Aug 22;71(5):1316-1319.
20. Calderón-Parra J, Kestler M, Ramos-Martínez A, Bouza E, Valerio M et al. Clinical Factors Associated with Reinfection versus Relapse in Infective Endocarditis: Prospective Cohort Study. *J Clin Med*. 2021 Feb 13;10(4):748
21. Danneels P, Hamel JF, Picard L, Rezig S, Martinet P, Lorleac'h A et al. Impact of *Enterococcus faecalis* Endocarditis Treatment on Risk of Relapse. *Clin Infect Dis*. 2023 Jan 13;76(2):281-290.
22. López-Cortés LE, Fernández-Cuenca F, Luque-Márquez R, de Alarcón A. Enterococcal Endocarditis: Relapses or Reinfections? *Clin Infect Dis*. 2021 Jan 27;72(2):360-361
23. Bonten MJ, Willems R, Weinstein RA. Vancomycin-resistant enterococci: why are they here, and where do they come from? *Lancet Infect Dis*. 2001 Dec;1(5):314-25.
24. Mascini EM, Bonten MJ. Vancomycin-resistant enterococci: consequences for therapy and infection control. *Clin Microbiol Infect*. 2005 Jul;11 Suppl 4:43-56.
25. R Fontana, M Ligozzi, F Pittaluga, G Satta. Intrinsic penicillin resistance in enterococci. *Microb Drug Resist*. 1996;2(2):209-13
26. Sonne M, Jawetz E. Comparison of the action of ampicillin and benzylpenicillin on enterococci in vitro. *Appl Microbiol*. 1968 Apr;16(4):645-8.
27. Arias CA, Singh KV, Panesso D, Murray BE. Evaluation of ceftobiprole medocaril against *Enterococcus faecalis* in a mouse peritonitis model. *J. Antimicrob. Chemother*. 2007. 60, 594–598
28. Arias CA, Singh KV, Panesso D, Murray BE. Time-kill and synergism studies of ceftobiprole against *Enterococcus faecalis*, including beta-lactamase-producing and vancomycin-resistant isolates. *Antimicrob. Agents Chemother*. 2007. 51, 2043–2047.
29. Jacqueline C, Caillon J, Le Mabecque V, Miègeville AF, Ge Y, Biek D et al. In vivo activity of a novel antimethicillin-resistant *Staphylococcus aureus* cephalosporin, ceftaroline, against vancomycin-susceptible and -resistant *Enterococcus faecalis* strains in a rabbit endocarditis model: a comparative study with linezolid and vancomycin. *Antimicrob. Agents Chemother*. 2009. 53, 5300–5302
30. Wilson WR, J E Geraci. Treatment of streptococcal infective endocarditis. *Am J Med*. 1985 jun 28;78(6B):128-37.
31. M L Fernández Guerrero, A Núñez García. Enterococcal endocarditis, a model of therapeutic difficulty. *Rev Clin Esp* . 1995 Oct;195 Suppl 4:41-5.
32. Shah, P. Paradoxical effect of antibiotics. I. The "Eagle effect." *J. Antimicrob. Chemother* 1980; 10:259-260.
33. Fontana R, Grossato A, Ligozzi M, Tonin EA. In vitro response to bactericidal activity of cell wall-active antibiotics does not support the general opinion that enterococci are naturally tolerant to these antibiotics. *Antimicrob Agents Chemother*. 1990 Aug;34(8):1518-22.
34. Tomayko JF, Zscheck KK, Singh KV, Murray BE. Comparison of the beta-lactamase gene cluster in clonally distinct strains of *Enterococcus faecalis*. *Antimicrob Agents Chemother*. 1996; 40:1170-4.
35. Coudron PE, Markowitz SM, Wong ES. Isolation of a beta-lactamase producing strain of *Enterococcus faecium*. *Antimicrob Agents Chemother*. 1992; 36:1125-6
36. Duez C, Zorzi W, Sapunarić F, Amoroso A, Thamm I, Coyette J. The penicillin resistance of *Enterococcus faecalis* JH2-2R results from an overproduction of the low-affinity penicillin-binding protein PBP4 and does not involve a *psr*-like gene. *Microbiology*. 2001; 147:2561–2569.
37. Rice LB, Desbonnet C, Tait-Kamradt A, Garcia-Solache M, Lonks J, Moon TM et al. Structural and regulatory changes in PBP4 trigger decreased beta-lactam susceptibility in *Enterococcus faecalis*. *mBio*. 2018; 9: e00361-18.
38. Rice LB, Bellais S, Carias LL, Hutton-Thomas R, Bonomo RA, Caspers P, et al . Impact of specific *pbp5* mutations on expression of beta-lactam resistance in *Enterococcus faecium*. *Antimicrob Agents Chemother*. 2004; 48:3028–3032.
39. Murray BE. The life and times of the *Enterococcus*. *Clin Microbiol Rev*. 1990; 3: 46-65.
40. Geraci JE, Martin WJ. Subacute enterococcal endocarditis: clinical, pathologic and therapeutic considerations in 33 patients. *Circulation*. 1954; 10:173–194.
41. Jawetz E, Sonne M. Penicillin-streptomycin treatment of enterococcal endocarditis: a reevaluation. *N Engl J Med*. 1966; 274:710–715.

42. Moellering RC, Weinberg AN. Studies on antibiotic synergism against enterococci. II. Effect of various antibiotics on the uptake of ¹⁴C-labelled streptomycin by enterococci. *J Clin Invest.* 1971; 50:2580–2584.
43. Shaw KJ, Rather PN, Hare RS, Miller GH. Molecular genetics of aminoglycoside resistance genes and familial relationships of the aminoglycoside-modifying enzymes. *Microbiol Rev.* 1993; 57:138–163.
44. Chow JW. Aminoglycoside resistance in enterococci. *Clin Infect Dis.* 2001; 31:586–9.
45. Cetinkaya Y, Falk P, Mayhall CG. Vancomycin-resistant enterococci. *Clin Microbiol Rev.* 2000; 13:686–707.
46. Kühn I, Iversen A, Finn M, Greko C, Burman LG, Blanch AR, et al. Occurrence and relatedness of vancomycin-resistant enterococci in animals, humans, and the environment in different European regions. *Appl Environ Microbiol.* 2005; 71:5383–90.
47. Phillips I, Casewell M, Cox T, De Groot B, Friis C, Jones R, et al. Does the use of antibiotics in food animals pose a risk to human health? A critical review of published data. *J Antimicrob Chemother.* 2004; 53:28–52.
48. Mendes RE, Castanheira M, Farrell DJ, Flamm RK, Sader HS, Jones RN. Longitudinal (2001–14) analysis of enterococci and VRE causing invasive infections in European and US hospitals, including a contemporary (2010–13) analysis of oritavancin in vitro potency. *J Antimicrob Chemother.* 2016 Dec;71(12):3453–3458.
49. Carpenter CF, Chambers HF. Daptomycin: another novel agent for treating infections due to drug-resistant gram-positive pathogens. *Clin Infect Dis.* 2004 Apr 1;38(7):994–1000.
50. Satlin MJ, Nicolau DP, Humphries RM, Kuti JL, Campeau SA, Lewis JS et al. Development of Daptomycin Susceptibility Breakpoints for *Enterococcus faecium* and Revision of the Breakpoints for Other Enterococcal Species by the Clinical and Laboratory Standards Institute. *Clin Infect Dis.* 2020 Mar 3;70(6):1240–1246.
51. Turnidge J, GKahlmeter G, Cantón R, MacGowan A, Giske CG and the European Committee on Antimicrobial Susceptibility Testing. Daptomycin in the treatment of enterococcal bloodstream infections and endocarditis: a EUCAST position paper. *Clin Microbiol Infect.* 2020 Aug;26(8):1039–1043.
52. Kamboj M, Cohen N, Gilhuley K, Babady NE, Seo SK, Sepkowitz KA. Emergence of daptomycin-resistant VRE: experience of a single institution. *Infect Control Hosp Epidemiol.* 2011 Apr;32(4):391–4.
53. Kelesidis T, Humphries R, Uslan DZ, Pegues DA. Daptomycin non-susceptible enterococci: an emerging challenge for clinicians. *Clin Infect Dis.* 2011 Jan 15;52(2):228–34.
54. Miller WR, Bayer AS, Arias CA. Mechanism of action and resistance to daptomycin in *Staphylococcus aureus* and enterococci. *Cold Spring Harb Perspect Med.* 2016; 6: a026997.
55. Mishra NN, Bayer AS, Tran TT, Shamoo Y, Mileykovskaya E, Dowhan W, Guan Z, Arias CA. 2012. Daptomycin resistance in enterococci is associated with distinct alterations of cell membrane phospholipid content. *PLoS One.* 2012; 7: e43958.
56. Tankovic J, Bachoual R, Ouabdesselam S, Boudjadja A, Soussy CJ. In-vitro activity of moxifloxacin against fluoroquinolone-resistant strains of aerobic gram-negative bacilli and *Enterococcus faecalis*. *J Antimicrob Chemother.* 1999; 43:19–23.
57. Kanematsu E, Deguchi T, Yasuda M, Kawamura T, Nishino Y, Kawada Y. Alterations in the GyrA subunit of DNA gyrase and the ParC subunit of DNA topoisomerase IV associated with quinolone resistance in *Enterococcus faecalis*. *Antimicrob Agents Chemother.* 1998; 42:433–5.
58. Oyamada Y, Ito H, Fujimoto K, Asada R, Niga T, Okamoto R et al. Combination of known and unknown mechanisms confers high-level resistance to fluoroquinolones in *Enterococcus faecium*. *J Med Microbiol* 2006; 55:729–736.
59. Dadashi M, Sharifian P, Bostanshirin N, Hajikhani B, Bostanghadiri N, Khosravi-Dehaghi N et al. The Global Prevalence of Daptomycin, Tigecycline, and Linezolid-Resistant *Enterococcus faecalis* and *Enterococcus faecium* Strains from Human Clinical Samples: A Systematic Review and Meta-Analysis. *Front Med (Lausanne).* 2021 Sep 10; 8:720647
60. Marshall SH, Donskey CJ, Hutton-Thomas R, Salata RA, Rice LB. Gene dosage and linezolid resistance in *Enterococcus faecium* and *Enterococcus faecalis*. *Antimicrob Agents Chemother.* 2002; 46:3334–3336
61. Kehrenberg C, Schwarz S, Jacobsen L, Hansen LH, Vester B. A new mechanism for chloramphenicol, florfenicol and clindamycin resistance: methylation of 23S ribosomal RNA at A2503. *Mol Microbiol.* 2005; 57:1064–1073.
62. Wang Y, Lv Y, Cai J, Schwarz S, Cui L, Hu Z, et al. A novel gene, *optrA*, that confers transferable resistance to oxazolidinones and phenicols and its presence in *Enterococcus faecalis* and *Enterococcus faecium* of human and animal origin. *J Antimicrob Chemother.* 2015; 70:2182–2190.
63. Vester B. The *cfr* and *cfr*-like multiple resistance genes. *Res Microbiol* 2018; 169:61–66.

64. Yaghoubi S, Zekiy AO, Krutova M, Gholami M, Kouhsari E, Sholeh M, et al. Tigecycline antibacterial activity, clinical effectiveness, and mechanisms and epidemiology of resistance: narrative review. *Eur J Clin Microbiol Infect Dis*. 2022 Jul;41(7):1003-1022.
65. Baddour LM, Wilson WR, Bayer AS, Fowler VG Jr, Tleyjeh IM, Rybak MJ, et al. Diagnosis, Antimicrobial Therapy, and Management of Complications: A Scientific Statement for Healthcare Professionals from the American Heart Association. *Circulation*. 2015 Oct 13;132(15):1435-86
66. Habib G, Lancellotti P, Antunes MJ, Bongiorni MG, Casalta JP, Del Zotti F et al. 2015 ESC Guidelines for the management of infective endocarditis: The Task Force for the Management of Infective Endocarditis of the European Society of Cardiology (ESC). Endorsed by: European Association for Cardio-Thoracic Surgery (EACTS), the European Association of Nuclear Medicine (EANM). *Eur Heart J*. 2015 Nov 21;36(44):3075-3128.
67. Wilson WR, Wilkowske CJ, Wright AJ, Sande MA, Geraci JE. Treatment of streptomycin-susceptible and streptomycin-resistant enterococcal endocarditis. *Ann Intern Med*. 1984 Jun;100(6):816-23.
68. Serra P, Brandimarte C, Martino P, Carlone S, Giunchi G. Synergistic treatment of enterococcal endocarditis: in vitro and in vivo studies. *Arch Intern Med*. 1977 Nov;137(11):1562-7.
69. Mainardi JL, Gutmann L, Acar JF, Goldstein FW. Synergistic effect of amoxicillin and cefotaxime against *Enterococcus faecalis*. *Antimicrob Agents Chemother*. 1995 Sep;39(9):1984-7
70. Gavalda J, Torres C, Tenorio C, López P, Zaragoza M, Capdevila JA, Almirante B, Ruiz F, Borrell N, Gomis X, Pigrau C, Baquero F, Pahissa A. Efficacy of ampicillin plus ceftriaxone in treatment of experimental endocarditis due to *Enterococcus faecalis* strains highly resistant to aminoglycosides. *Antimicrob Agents Chemother*. 1999 Mar;43(3):639-46.
71. Liao CH, Huang YT, Tsai HY, Hsueh PR. In vitro synergy of ampicillin with gentamicin, ceftriaxone, and ciprofloxacin against *Enterococcus faecalis*. *Int J Antimicrob Agents*. 2014 Jul;44(1):85-6.
72. Gavalda J, Len O, Miró JM, Muñoz P, Montejo M, Alarcón A, et al. A. Brief communication: treatment of *Enterococcus faecalis* endocarditis with ampicillin plus ceftriaxone. *Ann Intern Med*. 2007 Apr 17;146(8):574-9.
73. Gavalda J, Onrubia PL, Gómez MT, Gomis X, Ramírez JL, Len O et al. Efficacy of ampicillin combined with ceftriaxone and gentamicin in the treatment of experimental endocarditis due to *Enterococcus faecalis* with no high-level resistance to aminoglycosides. *J Antimicrob Chemother*. 2003 Sep;52(3):514-7.
74. Fernández-Hidalgo N, Almirante B, Gavalda J, Gurgui M, Peña C, de Alarcón A et al. Ampicillin plus ceftriaxone is as effective as ampicillin plus gentamicin for treating *Enterococcus faecalis* infective endocarditis. *Clin Infect Dis*. 2013 May;56(9):1261-8
75. Pericàs JM, Cervera C, del Rio A, Moreno A, García de la Maria C, Almela M et al. Changes in the treatment of *Enterococcus faecalis* infective endocarditis in Spain in the last 15 years: from ampicillin plus gentamicin to ampicillin plus ceftriaxone. *Clin Microbiol Infect*. 2014 Dec;20(12):O1075-83.
76. Owens RC Jr, Donskey CJ, Gaynes RP, Loo VG, Muto CA. Antimicrobial associated risk factors for *Clostridium difficile* infection. *Clin Infect Dis*. 2008; 46 (Suppl 1): S19–31.
77. Amberpet R, Sistla S, Parija SC, Thabab MM. Screening for intestinal colonization with vancomycin resistant enterococci and associated risk factors among patients admitted to an adult intensive care unit of a large teaching hospital. *J Clin Diagn Res*. 2016; 10: DC06–9.
78. McKinnell JA, Kunz DF, Chamot E, et al. Association between vancomycin-resistant enterococci bacteremia and ceftriaxone usage. *Infect Control Hosp Epidemiol*. 2012; 33:718–24.
79. Lakticová V, Hutton-Thomas R, Meyer M, Gurkan E, Rice LB. Antibiotic-induced enterococcal expansion in the mouse intestine occurs throughout the small bowel and correlates poorly with suppression of competing flora. *Antimicrob Agents Chemother*. 2006; 50:3117–23.
80. Rice LB, Hutton-Thomas R, Lakticova V, Helfand MS, Donskey CJ. Beta-lactam antibiotics and gastrointestinal colonization with vancomycin-resistant enterococci. *J Infect Dis*. 2004; 189:1113–8.
81. Panagiotidis G, Bäckström T, Asker-Hagelberg C, Jandourek A, Weintraub A, Nord CE. Effect of ceftaroline on normal human intestinal microflora. *Antimicrob Agents Chemother*. 2010; 54:1811–4.
82. Luther MK, Rice LB, LaPlante KL. Ampicillin in combination with ceftaroline, cefepime, or ceftriaxone demonstrates equivalent activities in a high-inoculum *Enterococcus faecalis* infection model. *Antimicrob Agents Chemother*. 2016; 60:3178–82.

83. Werth BJ, Shireman LM. Pharmacodynamics of ceftaroline plus ampicillin against *Enterococcus faecalis* in an in vitro pharmacokinetic/pharmacodynamic model of simulated endocardial vegetations. *Antimicrob Agents Chemother*. 2017 Mar 24;61(4): e02235-16.
84. Arias CA, Singh KV, Panesso D, Murray BE. Time-kill and synergism studies of ceftobiprole against *Enterococcus faecalis*, including beta-lactamase-producing and vancomycin-resistant isolates. *Antimicrob Agents Chemother*. 2007 Jun;51(6):2043-7.
85. Ramos MC, Grayson ML, Eliopoulos GM, Bayer AS. Comparison of daptomycin, vancomycin, and ampicillin-gentamicin for treatment of experimental endocarditis caused by penicillin-resistant enterococci. *Antimicrob. Agents Chemother*. 1992; 36, 1864–1869.
86. Chen HY, Williams JD. The activity of vancomycin and teicoplanin alone and in combination with gentamicin or ampicillin against *Streptococcus faecalis*. *Eur. J. Clin. Microbiol*. 1984; 3, 436–438.
87. Chandrasekar PH, Price S, Levine DP. In-vitro evaluation of ceftipime (HR 810), teicoplanin and four other antimicrobials against enterococci. *J Antimicrob Chemother* . 1985 Aug;16(2):179-82.
88. Pohlod DJ, Saravolatz LD, Somerville MM. In-vitro susceptibility of Gram-positive cocci to LY146032 teicoplanin, sodium fusidate, vancomycin, and rifampicin. *J Antimicrob Chemother*. 1987; 20: 197-202
89. Spencer RC, Goering R.A critical review of the in-vitro activity of teicoplanin. *Int J Antimicrob Agents*. 1995; 5: 169-177.
90. Sullam PM, Täuber MG, Hackbarth CJ, Sande MA. Therapeutic efficacy of teicoplanin in experimental enterococcal endocarditis. *Antimicrob Agents Chemother*. 1985 Jan;27(1):135-6.
91. López P, Gavalda J, Martin MT, Almirante B, Gomis X, Azuaje C et al. Efficacy of teicoplanin-gentamicin given once a day on the basis of pharmacokinetics in humans for treatment of enterococcal experimental endocarditis. *Antimicrob Agents Chemother*. 2001 May;45(5):1387-93.
92. Escolà-Vergé L; Fernández-Hidalgo N; Rodríguez-Pardo D.; Pigrau C; González-López JJ, Bartolomé R. Almirante B. Teicoplanin for treating enterococcal infective endocarditis: A retrospective observational study from a referral centre in Spain. *Int J Antimicrob Agents*. 2019; 53: 165–170.
93. De Nadaï, T.; François, M.; Sommet, A; Dubois, D.; Metsu, D.; Grare, .; Marchou, B; Delobel, P; Martin-Blondel, G. Efficacy of teicoplanin monotherapy following initial standard therapy in *Enterococcus faecalis* infective endocarditis: A retrospective cohort study. *Infection*. 2019; 47: 463–469.
94. Presterl E, Graninger W, Georgopoulos A. The efficacy of teicoplanin in the treatment of endocarditis caused by Gram-positive bacteria. *J. Antimicrob. Chemother*. 1993; 31: 755–766.
95. Hayden MK, G M Trenholme, J E Schultz, D F Sahm. In vivo development of teicoplanin resistance in a VanB *Enterococcus faecium* isolate. *J Infect Dis*. 1993 May;167(5):1224-7.
96. Cerón I, Bermejo J, Bouza E; Eworo A, Cruz AF, Cuerpo G et al. Efficacy of daptomycin in the treatment of enterococcal endocarditis: A 5 year comparison with conventional therapy. *J. Antimicrob. Chemother*. 2014; 69: 1669–1674.
97. Carugati M, Bayer AS, Miró JM, Park LP, Guimarães AC, Skoutelis A, et al. High-Dose Daptomycin Therapy for Left-Sided Infective Endocarditis: A Prospective Study from the International Collaboration on Endocarditis. *Antimicrob Agents Chemother*. 2013; 57: 6213–6222.
98. Bassetti M, Russo A, Givone F, Ingani M, Graziano E, Bassetti M. Should High-dose Daptomycin be an Alternative Treatment Regimen for Enterococcal Endocarditis? *Infect. Dis. Ther*. 2019, 8, 695–702.
99. Cervera C, Castañeda X, Pericàs JM, Del Río A, de la Maria CG, Mestres C, et al. Clinical utility of daptomycin in infective endocarditis caused by Gram-positive cocci. *Int J Antimicrob Agents*. 2011; 38(5):365–70.
100. Peghin M, Russo A, Givone F, Ingani M, Graziano E, Bassetti M. Should High dose daptomycin be an alternative treatment regimen for enterococcal endocarditis? *Infect Dis Ther*. 2019; 8(4), 695–702.
101. Pericàs JM, García-de-la-Maria C, Brunet M, Armero Y, García-González J, Casals G. Early in vitro development of daptomycin non-susceptibility in high-level aminoglycoside-resistant *Enterococcus faecalis* predicts the efficacy of the combination of high-dose daptomycin plus ampicillin in an in vivo model of experimental endocarditis. *Journal of Antimicrobial Chemotherapy*. 2017; 72 (6): 1714–1722,
102. Hall AD, Steed ME, Arias CA, Murray BE, Rybak MJ. Evaluation of standard- and high-dose daptomycin versus linezolid against vancomycin-resistant *Enterococcus* isolates in an in vitro pharmacokinetic/pharmacodynamic model with simulated endocardial vegetations. *Antimicrob Agents Chemother*. 2012 Jun;56(6):3174-80.

103. Arias CA, Panesso D, McGrath DM, Qin X, Mojica MF, Miller C et al. Genetic Basis for In Vivo Daptomycin Resistance in Enterococci. *N Engl J Med*. 2011 Sep 8; 365(10): 892–900.
104. Storm JC, Diekema DJ, Kroeger JS, Johnson SJ, Johannsson B. Daptomycin exposure precedes infection and/or colonization with daptomycin non-susceptible enterococcus. *Antimicrob Resist Infect Control*. 2012 May 29;1(1):19.
105. Werth BJ, Shireman LM. Pharmacodynamics of ceftaroline plus ampicillin against *Enterococcus faecalis* in an in vitro pharmacokinetic/pharmacodynamic model of simulated endocardial vegetations. *Antimicrob Agents Chemother*. 2017; 61.
106. Sakoulas G, Nonejuie P, Nizet V, Pogliano J, Crum-Cianflone N, Haddad F. Treatment of high-level gentamicin-resistant *Enterococcus faecalis* endocarditis with daptomycin plus ceftaroline. *Antimicrob Agents Chemother*. 2013; 57:4042–5.
107. Smith JR, Barber KE, Raut A, Aboutaleb M, Sakoulas G, Rybak MJ. β -Lactam combinations with daptomycin provide synergy against vancomycin-resistant *Enterococcus faecalis* and *Enterococcus faecium*. *J Antimicrob Chemother*. 2015; 70:1738–43.
108. Sierra-Hoffman M, Iznola O, Goodwin M, Mohr J. Combination therapy with ampicillin and daptomycin for treatment of *Enterococcus faecalis* endocarditis. *Antimicrob Agents Chemother*. 2012; 56:6064.
109. Rice LB, Eliopoulos GM, Moellering RC Jr. In vitro synergism between daptomycin and fosfomycin against *Enterococcus faecalis* isolates with high-level gentamicin resistance. *Antimicrob Agents Chemother*. 1989; 33:470–3.
110. Rice LB, Eliopoulos CT, Yao JD, Eliopoulos GM, Moellering RC Jr. In vivo activity of the combination of daptomycin and fosfomycin compared with daptomycin alone against a strain of *Enterococcus faecalis* with high-level gentamicin resistance in the rat endocarditis model. *Diagn Microbiol Infect Dis*. 1992; 15:173–6.
111. Farina C, Russello G, Chinello P, Pasticci B, A Raglio A, Ravasio V, et al. In vitro activity effects of twelve antibiotics alone and in association against twenty-seven *Enterococcus faecalis* strains isolated from Italian patients with infective endocarditis: high in vitro synergistic effect of the association ceftriaxone-fosfomycin. *Chemotherapy*. 2011; 57:426–33.
112. Tang HJ, Chen CC, Zhang CC, Su BA, Li CM, Weng TC et al. In vitro efficacy of fosfomycin-based combinations against clinical vancomycin-resistant *Enterococcus* isolates. *Diagn Microbiol Infect Dis*. 2013; 77:254–7.
113. Pujol M, Miró JM, Shaw E, Aguado JM, San-Juan R, Puig-Asensio M et al. Daptomycin Plus Fosfomycin Versus Daptomycin Alone for Methicillin-resistant *Staphylococcus aureus* Bacteremia and Endocarditis: A Randomized Clinical Trial. *Clin Infect Dis*. 2021 May 4;72(9):1517-1525.
114. García-de-la-Mària C, Gasch O, García-Gonzalez J, Soy D, Shaw E, Ambrosioni J et al. The Combination of Daptomycin and Fosfomycin Has Synergistic, Potent, and Rapid Bactericidal Activity against Methicillin-Resistant *Staphylococcus aureus* in a Rabbit Model of Experimental Endocarditis. *Antimicrob Agents Chemother*. 2018 May 25;62(6): e02633-17
115. García-de-la-Mària C, Gasch O, Castañeda X, García-González J, Soy D, Cañas MA et al. Cloxacillin or fosfomycin plus daptomycin combinations are more active than cloxacillin monotherapy or combined with gentamicin against MSSA in a rabbit model of experimental endocarditis. *J Antimicrob Chemother*. 2020 Dec 1;75(12):3586-3592.
116. Pericàs JM, Moreno A, Almela M, García-de-la-Mària C, Marco F, Muñoz P et al. Efficacy and safety of fosfomycin plus imipenem versus vancomycin for complicated bacteraemia and endocarditis due to methicillin-resistant *Staphylococcus aureus*: a randomized clinical trial. *Clin Microbiol Infect*. 2018 Jun;24(6):673-676.
117. Oliva A, Furustrand T, Tiffin U, Maiolo EM, Jeddari S, Bétrisey B, Trampuz A. Activities of fosfomycin and rifampin on planktonic and adherent *Enterococcus faecalis* strains in an experimental foreign-body infection model. *Antimicrob Agents Chemother* 2014; 58:1284–93.
118. Arias CA, Murray BE. Emergence and management of drug resistant enterococcal infections. *Expert Rev Anti Infect Ther*. 2008; 6(5):637–55.
119. Scheetz MH, Knechtel SA, Malczynski M, Postelnick MJ, Qi C. Increasing incidence of linezolid-intermediate or -resistant, vancomycin-resistant *Enterococcus faecium* strains parallels increasing linezolid consumption. *Antimicrob Agents Chemother*. 2008 Jun;52(6):2256-9.

120. Bender JK, Cattoir V, Hegstad K, Sadowy E, Coque TM, Westh H, et al. Update on prevalence and mechanisms of resistance to linezolid, tigecycline and daptomycin in enterococci in Europe: towards a common nomenclature. *Drug Resist Updat.* (2018) 40:25-39.
121. Birmingham MC, Rayner VR, Meagher AK, Flavin SM, Batts DH, Schentag JJ. Linezolid for the treatment of multidrug-resistant, gram-positive infections: experience from a compassionate-use program. *Clin Infect Dis.* 2003 Jan 15;36(2):159-68.
122. Falagas ME, Manta KG, Ntziora F, Vardakas KZ. Linezolid for the treatment of patients with endocarditis: a systematic review of the published evidence. *J Antimicrob Chemother.* 2006; 58 (2):273–80.
123. Stevens MP, Edmond MB. Endocarditis due to vancomycin-resistant enterococci: case report and review of the literature. *Clin Infect Dis.* 2005; 41(8):1134–42.
124. Tsigrelis C, Singh KV, Coutinho TD, Murray BE, Baddour LM. Vancomycin-resistant *Enterococcus faecalis* endocarditis: linezolid failure and strain characterization of virulence factors. *J Clin Microbiol.* 2007; 45(2):631–5.
125. Berdal JE, Eskesen A. Short-term success, but long-term treatment failure with linezolid for enterococcal endocarditis. *Scand J Infect Dis.* 2008; 40(9):765–6.
126. Tsigrelis C, Singh KV, Coutinho TD, Murray BE, Baddour LM. Vancomycin-resistant *Enterococcus faecalis* endocarditis: linezolid failure and strain characterization of virulence factors. *J Clin Microbiol.* 2007 Feb;45(2):631-5.
127. Silva-Del Toro SL, Greenwood-Quaintance KE, Patel R. In vitro activity of tedizolid against linezolid-resistant staphylococci and enterococci. *Diagn Microbiol Infect Dis.* 2016 May;85(1):102-4.
128. Zhanel GG, Love R, Adam H, Golden A, Zelenitsky S, Schweizer F et al. Tedizolid: a novel oxazolidinone with potent activity against multidrug-resistant gram-positive pathogens. *Drugs.* 2015 Feb;75(3):253-70.
129. Singh KV, Arias CA, Murray BE. Efficacy of Tedizolid against Enterococci and Staphylococci, Including cfr+ Strains, in a Mouse Peritonitis Model. *Antimicrob Agents Chemother.* 2019 Mar 27;63(4):e02627-18.
130. Barber KE, Smith JR, Raut A, Rybak MJ. Evaluation of tedizolid against *Staphylococcus aureus* and enterococci with reduced susceptibility to vancomycin, daptomycin or linezolid. *J Antimicrob Chemother.* 2016 Jan;71(1):152-5.
131. Iqbal K, Rohde H, Huang J, Tikiso T, Amann LF, Zeitlinger M, Wicha SG. A pharmacokinetic-pharmacodynamic (PKPD) model-based analysis of tedizolid against enterococci using the hollow-fibre infection model. *J Antimicrob Chemother.* 2022 Aug 25;77(9):2470-2478.
132. Landman D, Quale JM, Mobarakai N, Zaman MM. Ampicillin plus ciprofloxacin therapy of experimental endocarditis caused by multidrug-resistant *Enterococcus faecium*. *J Antimicrob Chemother.* 1995; 36:253–258.
133. Fernández-Guerrero M, Rouse MS, Henry NK, Geraci JE, Wilson WR. In vitro and in vivo activity of ciprofloxacin against enterococci isolated from patients with infective endocarditis. *Antimicrob Agents Chemother.* 1987 Mar;31(3):430-3.
134. Van Nieuwkoop C, Visser LG, Groeneveld JH, Kuijper EJ. Chronic bacterial prostatitis and relapsing *Enterococcus faecalis* bacteraemia successfully treated with moxifloxacin. *J Infect.* 2008; 56:155–156.
135. Markham A. Delafloxacin: First Global Approval. *Drugs.* 2017 Sep;77(13):1481-1486.
136. Lee YR, Burton CE, Bevel KR. Delafloxacin for the Treatment of Acute Bacterial Skin and Skin Structure Infections. *J Pharm Technol.* 2019 Jun;35(3):110-118.
137. Bassetti M, Melchio M, Giacobbe DR. Delafloxacin for the treatment of adult patients with community-acquired bacterial pneumonia. *Expert Rev Anti Infect Ther.* 2022 May;20(5):649-656.
138. Ellis-Grosse EJ, Babinchak T, Dartois N, Rose G, Loh E. The efficacy and safety of tigecycline in the treatment of skin and skin-structure infections: results of 2 double-blind phase 3 comparison studies with vancomycin-aztreonam. *Clin Infect Dis.* 2005; 5:S341–S353.
139. Babinchak T, Ellis-Grosse E, Dartois N, Rose GM, Loh E. The efficacy and safety of tigecycline for the treatment of complicated intra-abdominal infections: analysis of pooled clinical trial data. *Clin Infect Dis.* 2005; 5:S354–S367.
140. Entenza JM, Moreillon P. Tigecycline in combination with other antimicrobials: a review of in vitro, animal and case report studies. *Int J Antimicrob Agents.* 2009; 34(8):e1–e9.
141. Jenkins I. Linezolid- and vancomycin-resistant *Enterococcus faecium* endocarditis: successful treatment with tigecycline and daptomycin. *J Hosp Med.* 2007 Sep;2(5):343-4.

142. Polidori M, Nuccorini A, Tascini C, Gemignani G, Iapoce R, Leonildi A, Tagliaferri E, Menichetti F. Vancomycin-resistant *Enterococcus faecium* (VRE) bacteremia in infective endocarditis successfully treated with combination daptomycin and tigecycline. *J Chemother*. 2011 Aug;23(4):240-1.
143. Schutt AC, Bohm NM. Multidrug-resistant *Enterococcus faecium* endocarditis treated with combination tigecycline and high-dose daptomycin. *Ann Pharmacother*. 2009; 43(12):2108–12.
144. Peleg AY, Potoski BA, Rea R, et al. *Acinetobacter baumannii* bloodstream infection while receiving tigecycline: a cautionary report. *J Antimicrob Chemother*. 2007; 59:128–131.
145. Lefort A, Lafaurie M, Massias L, Petegnief Y, Saleh-Mghir A, Muller-Serieys C, et al. Activity and diffusion of tigecycline (GAR-936) in experimental enterococcal endocarditis. *Antimicrob Agents Chemother*. 2003; 47(1):216–22.
146. Molina KC, Miller MA, Mueller SW, Van Matre ET, Krsak M, Kiser TH. Clinical Pharmacokinetics and Pharmacodynamics of Dalbavancin. *Clin Pharmacokinet*. 2022 Mar;61(3):363-374.
147. Oliva A, Stefani S, Venditti M, Di Domenico EG. Biofilm-Related Infections in Gram-Positive Bacteria and the Potential Role of the Long-Acting Agent Dalbavancin. *Front Microbiol*. 2021 Oct 22; 12:749685.
148. Sader HS, Mendes RE, Pfaller MA, Flamm RK. Antimicrobial activity of dalbavancin tested against Gram-positive organisms isolated from patients with infective endocarditis in US and European medical centres. *J. Antimicrob. Chemother*. 74(5), 1306–1310 (2019).
149. Gatti M, Andreoni M, Pea F, Viale P. Real-World Use of Dalbavancin in the Era of Empowerment of Outpatient Antimicrobial Treatment: A Careful Appraisal Beyond Approved Indications Focusing on Unmet Clinical Needs. *Drug Des Devel Ther*. 2021 Aug 3;15:3349-3378.
150. Tobudic S, Forstner C, Burgmann H, Lagler H, Ramharter M, Steininger C et al. Dalbavancin as Primary and Sequential Treatment for Gram-Positive Infective Endocarditis: 2-Year Experience at the General Hospital of Vienna. *Clin Infect Dis*. 2018, 67, 795–798.
151. Hidalgo-Tenorio C., Vinuesa D.; Plata A, Dávila PM, Iftime S et al. DALBACEN cohort: Dalbavancin as consolidation therapy in patients with endocarditis and/or bloodstream infection produced by gram-positive cocci. *Ann Clin Microbiol Antimicrob*. 2019, 18, 1–10.
152. Arhin FF, Draghi DC, Pillar CM, Parr TR Jr, Moeck G, Sahm DF. Comparative in vitro activity profile of oritavancin against recent gram-positive clinical isolates. *Antimicrob Agents Chemother*. 2009 Nov;53(11):4762-71.
153. Pfaller MA, Sader HS, Flamm RK, Castanheira M, Mendes RE. Oritavancin in vitro activity against gram-positive organisms from European and United States medical centers: results from the SENTRY Antimicrobial Surveillance Program for 2010-2014. *Diagn Microbiol Infect Dis*. 2018 Jun;91(2):199-204.
154. Saravolatz LD, Stein GE. Oritavancin: A Long-Half-Life Lipoglycopeptide. *Clin Infect Dis*. 2015 Aug 15;61(4):627-32
155. Yan Q, Karau MJ, Patel R. In vitro activity of oritavancin against planktonic and biofilm states of vancomycin-susceptible and vancomycin-resistant enterococci. *Diagn Microbiol Infect Dis*. 2018 Aug;91(4):348-350.
156. Scoble PJ, Reilly J, Tillotson GS. Real-World Use of Oritavancin for the Treatment of Osteomyelitis. *Drugs Real World Outcomes*. 2020 Jun;7(Suppl 1):46-54.
157. Morrisette T, Miller MA, Montague BT, Barber GR, McQueen RB, Krsak M. On- and off-label utilization of dalbavancin and oritavancin for Gram-positive infections. *J Antimicrob Chemother*. 2019 Aug 1;74(8):2405-2416.
158. Bassetti M, Labate L, Vena A, Giacobbe DR. Role of oritavancin and dalbavancin in acute bacterial skin and skin structure infections and other potential indications. *Curr Opin Infect Dis*. 2021 Apr 1;34(2):96-108.
159. Stewart CL, Turner MS, Frens JJ, Snider CB, Smith JR. Real-World Experience with Oritavancin Therapy in Invasive Gram-Positive Infections. *Infect Dis Ther*. 2017 Jun;6(2):277-289.
160. Johnson JA, Feeney ER, Kubiak DW, Corey GR. Prolonged Use of Oritavancin for Vancomycin-Resistant *Enterococcus faecium* Prosthetic Valve Endocarditis. *Open Forum Infect Dis*. 2015 Oct 29;2(4): ofv156.
161. Pericàs JM, Cervera C, Moreno A, Garcia-de-la-Mària C, Almela M, Falces C, et al. Outcome of *Enterococcus faecalis* infective endocarditis according to the length of antibiotic therapy: Preliminary data from a cohort of 78 patients. *PLoS One*. 2018 Feb 20;13(2): e0192387.
162. Ramos-Martínez A, Pericàs JM, Fernández-Cruz A, Muñoz P, Valerio M, Kestler M, et al. Four weeks versus six weeks of ampicillin plus ceftriaxone in *Enterococcus faecalis* native valve endocarditis: A prospective cohort study. *PLoS One*. 2020 Aug 3;15(8):e0237011.

163. Herzstein J, Ryan JL, Mangi RJ, Greco TP, Andriole VT. Optimal therapy for enterococcal endocarditis. *Am J Med.* 1984 Feb;76(2):186-91.
164. Olaison L, Schadewitz K; Swedish Society of Infectious Diseases Quality Assurance Study Group for Endocarditis. Enterococcal endocarditis in Sweden, 1995-1999: can shorter therapy with aminoglycosides be used? *Clin Infect Dis.* 2002 Jan 15;34(2):159-66.
165. Dahl A, Rasmussen RV, Bundgaard H, Hassager C, Bruun LE, Lauridsen TK et al. Enterococcus faecalis infective endocarditis: a pilot study of the relationship between duration of gentamicin treatment and outcome. *Circulation.* 2013 Apr 30;127(17):1810-7.
166. Buchholtz K, Larsen CT, Hassager C, Bruun NE. Severity of gentamicin's nephrotoxic effect on patients with infective endocarditis: a prospective observational cohort study of 373 patients. *Clin Infect Dis.* 2009 Jan 1; 48(1):65-71.
167. Cosgrove SE, Vigliani GA, Fowler VG Jr et al. Initial low-dose gentamicin for Staphylococcus aureus bacteremia and endocarditis is nephrotoxic. *Clin Infect Dis.* 2009; 48(6): 713-721.
168. Buchholtz K, Larsen CT, Hassager C, Bruun NE. Infective endocarditis: long-term reversibility of kidney function impairment. A 1-y post-discharge follow-up study. *Scand J Infect Dis.* 2010; 42 (6-7): 484-490.
169. Chirouze C, Athan E, Alla F, Chu VH, Ralph Corey G, Selton-Suty C et al. Enterococcal endocarditis in the beginning of the 21st century: analysis from the International Collaboration on Endocarditis-Pro Prospective Cohort Study. *Eur Heart J.* 2019 Oct 14;40(39):3222-3232.
170. Steckelberg JM, Murphy JG, Ballard D, Bailey K, Tajik AJ, Taliencio CP et al. Emboli in infective endocarditis: the prognostic value of echocardiography. *Ann Intern Med.* 1991 Apr 15;114(8):635-40.
171. García-Cabrera E, Fernández-Hidalgo N, Almirante B, Ivanova-Georgieva R, Nouredine M, Plata A et al. Neurological complications of infective endocarditis: risk factors, outcome, and impact of cardiac surgery: a multicenter observational study. *Circulation.* 2013 Jun 11;127(23):2272-84.
172. Williams DN, Baker CA, Kind AC, Sannes MR. The history and evolution of outpatient parenteral antibiotic therapy (OPAT). *Int J Antimicrob Agents.* 2015 Sep;46(3):307-12.
173. Andrews MM, von Reyn CF. Patient selection criteria and management guidelines for outpatient parenteral antibiotic therapy for native valve infective endocarditis. *Clin Infect Dis.* 2001 Jul 15;33(2):203-9.
174. Pericàs JM, Llopis J, González-Ramallo V, Goenaga MÁ, Muñoz P, García-Leoni ME et al. Outpatient Parenteral Antibiotic Treatment for Infective Endocarditis: A Prospective Cohort Study From the GAMES Cohort. *Clin Infect Dis.* 2019 Oct 30;69(10):1690-1700.
175. Pericàs JM, Llopis J, Muñoz P, González-Ramallo V, García-Leoni ME, de Alarcón A et al. Outpatient Parenteral Antibiotic Treatment vs Hospitalization for Infective Endocarditis: Validation of the OPAT-GAMES Criteria. *Open Forum Infect Dis.* 2022 Aug 30;9(9): ofac442.
176. Dubé L, Caillon J, Jacqueline C, Bugnon D, Potel G, Asseray N. The optimal aminoglycoside and its dosage for the treatment of severe Enterococcus faecalis infection. An experimental study in the rabbit endocarditis model. *Eur J Clin Microbiol Infect Dis.* 2012 Oct;31(10):2545-7.
177. Fantin B, Carbon C. Importance of the aminoglycoside dosing regimen in the penicillin-netilmicin combination for treatment of Enterococcus faecalis-induced experimental endocarditis. *Antimicrob Agents Chemother.* 1990 Dec;34(12):2387-91.
178. Marangos MN, Nicolau DP, Quintiliani R, Nightingale CH. Influence of gentamicin dosing interval on the efficacy of penicillin-containing regimens in experimental Enterococcus faecalis endocarditis. *J Antimicrob Chemother.* 1997 Apr;39(4):519-22.
179. Hessen MT, Pitsakis PG, Levison ME. Postantibiotic effect of penicillin plus gentamicin versus Enterococcus faecalis in vitro and in vivo. *Antimicrob Agents Chemother.* 1989 May;33(5):608-11.
180. Gavalda J, Cardona PJ, Almirante B, Capdevila JA, Laguarda M, Pou L, Crespo E, Pigrau C, Pahissa A. Treatment of experimental endocarditis due to Enterococcus faecalis using once-daily dosing regimen of gentamicin plus simulated profiles of ampicillin in human serum. *Antimicrob Agents Chemother.* 1996 Jan;40(1):173-8.
181. Schwank S, Blaser J. Once-versus thrice-daily netilmicin combined with amoxicillin, penicillin, or vancomycin against Enterococcus faecalis in a pharmacodynamic in vitro model. *Antimicrob Agents Chemother* 1996 Oct;40(10):2258-61.
182. Pagkalis S, Mantadakis E, Mavros MN, Ammari C, Falagas ME. Pharmacological considerations for the proper clinical use of aminoglycosides. *Drugs.* 2011 Dec 3;71(17):2277-94.

183. Goenaga-Sánchez MA, Kortajarena-Urkola X, Bouza-Santiago E, Muñoz-García P, Verde-Moreno E, Fariñas-Álvarez MC et al. Aetiology of renal failure in patients with infective endocarditis. The role of antibiotics. *Med Clin (Barc)*. 2017 Oct 23;149(8):331-338.
184. Gil-Navarro MV, Lopez-Cortes LE, Luque-Marquez R, Galvez-Acebal J, de Alarcón-González A. Outpatient parenteral antimicrobial therapy in *Enterococcus faecalis* infective endocarditis. *J Clin Pharm Ther*. 2018 Apr;43(2):220-223.
185. Herrera-Hidalgo L, de Alarcón A, López-Cortes LE, Luque-Márquez R, López-Cortes LF, Gutiérrez-Valencia A, Gil-Navarro MV. Is Once-Daily High-Dose Ceftriaxone plus Ampicillin an Alternative for *Enterococcus faecalis* Infective Endocarditis in Outpatient Parenteral Antibiotic Therapy Programs? *Antimicrob Agents Chemother*. 2020 Dec 16;65(1): e02099-20.
186. Patel IH, Miller K, Weinfeld R, Spicehandler J. Multiple intravenous dose pharmacokinetics of ceftriaxone in man. *Chemotherapy*. 1981;27 Suppl 1:47-56.
187. Perry TR, Schentag JJ. Clinical use of ceftriaxone: a pharmacokinetic-pharmacodynamic perspective on the impact of minimum inhibitory concentration and serum protein binding. *Clin Pharmacokinet*. 2001;40(9):685-94.
188. Grégoire M, Dailly E, Le Turnier P, Garot D, Guimard T, Bernard L et al. High-Dose Ceftriaxone for Bacterial Meningitis and Optimization of Administration Scheme Based on Nomogram. *Antimicrob Agents Chemother*. 2019 Aug 23;63(9): e00634-19.
189. Luderer JR, Patel IH, Durkin J, Schneek DW. Age and ceftriaxone kinetics. *Clin Pharmacol Ther*. 1984 Jan;35(1):19-25.
190. Patel IH, Sugihara JG, Weinfeld RE, Wong EG, Siemsen AW, Berman SJ. Ceftriaxone pharmacokinetics in patients with various degrees of renal impairment. *Antimicrob Agents Chemother*. 1984 Apr;25(4):438-42.
191. Herrera-Hidalgo L, Lomas-Cabezas JM, López-Cortés LE, Luque-Márquez R, López-Cortés LF, Martínez-Marcos FJ et al. Ampicillin Plus Ceftriaxone Combined Therapy for *Enterococcus faecalis* Infective Endocarditis in OPAT. *J Clin Med*. 2021 Dec 21;11(1):7.
192. Jimenez-Toro I, Rodriguez CA, Zuluaga AF, Otalvaro JD, Perez-Madrid H, Vesga O. Pharmacokinetic/Pharmacodynamic Index Linked to In Vivo Efficacy of the Ampicillin-Ceftriaxone Combination against *Enterococcus faecalis*. *Antimicrob Agents Chemother*. 2023 Feb 16;67(2): e0096622.
193. Jimenez-Toro I, Rodriguez CA, Zuluaga AF, Otalvaro JD, Vesga O. A new pharmacodynamic approach to study antibiotic combinations against enterococci in vivo: Application to ampicillin plus ceftriaxone. *PLoS One*. 2020 Dec 8;15(12): e0243365.
194. Maher M, Jensen KJ, Lee D, Nix DE. Stability of Ampicillin in Normal Saline and Buffered Normal Saline. *Int J Pharm Compd*. 2016 Jul-Aug;20(4):338-342.
195. W C Hellinger, M S Rouse, P M Rabadan, N K Henry, J M Steckelberg, W R Wilson. Continuous intravenous versus intermittent ampicillin therapy of experimental endocarditis caused by aminoglycoside-resistant enterococci. *Antimicrob Agents Chemother*. 1992 Jun;36(6):1272-5.
196. Lewis PO, Jones A, Amodei RJ, Youssef D. Continuous Infusion Ampicillin for the Outpatient Management of Enterococcal Endocarditis: A Case Report and Literature Review. *J Pharm Pract*. 2020 Jun;33(3):392-394.
197. Nickolai DJ, Lammel CJ, Byford BA, Morris JH, Kaplan EB, Hadley WK, Brooks GF. Effects of storage temperature and pH on the stability of eleven beta-lactam antibiotics in MIC trays. *J Clin Microbiol*. 1985 Mar;21(3):366-70.
198. Herrera-Hidalgo L, Gil-Navarro MV, Dilly Penchala S, López-Cortes LE, de Alarcón A, Luque-Márquez R, López-Cortes LF, Gutiérrez-Valencia A. Ceftriaxone pharmacokinetics by a sensitive and simple LC-MS/MS method: Development and application. *J Pharm Biomed Anal*. 2020 Sep 10; 189:113484.
199. Herrera-Hidalgo L, López-Cortes LE, Luque-Márquez R, Gálvez-Acebal J, de Alarcón A, López-Cortes LF, Gutiérrez-Valencia A, Gil-Navarro MV. Ampicillin and Ceftriaxone Solution Stability at Different Temperatures in Outpatient Parenteral Antimicrobial Therapy. *Antimicrob Agents Chemother*. 2020 Jun 23; 64(7): e00309-20.
200. Fernández-Rubio B, Herrera-Hidalgo L, Luque-Márquez R, de Alarcón A, López-Cortés LE, Luque-Pardos S et al. Stability of Ampicillin plus Ceftriaxone Combined in Elastomeric Infusion Devices for Outpatient Parenteral Antimicrobial Therapy. *Antibiotics*. 2023; 12: 432. <https://doi.org/10.3390/antibiotics12030432>
201. Herrera-Hidalgo L, de Alarcón A, López-Cortes LE, Luque-Márquez R, López-Cortes LF, Gutiérrez-Valencia A, Gil-Navarro MV. *Enterococcus faecalis* Endocarditis and Outpatient Treatment: A Systematic Review of Current Alternatives. *Antibiotics (Basel)*. 2020 Sep 30;9(10):657.

202. Phillips B, Watson GH. Oral treatment of subacute bacterial endocarditis in children. *Arch Dis Child*. 1977; 52: 235–7.
203. Chetty S, Mitha AS. High-dose oral amoxycillin in the treatment of infective endocarditis. *S Afr Med J*. 1988; 73: 709–10.
204. Stambouliau D, Bonvehi P, Arevalo C, Bologna R, Cassetti I, Scilingo V, Efron E. Antibiotic management of outpatients with endocarditis due to penicillin-susceptible streptococci. *Rev Infect Dis*. 1991; 13 Suppl 2: S160–3.
205. Tissot-Dupont H, Gouriet F, Oliver L, Jamme M, Casalta JP, Jimeno MT et al. High-dose trimethoprim sulfamethoxazole and clindamycin for *Staphylococcus aureus* endocarditis. *Int J Antimicrob Agents*. 2019; 54: 143–8.
206. Mzabi A, Kerneis S, Richaud CPodglajen I, Fernandez-Gerlinger MP, Mainardi JL. Switch to oral antibiotics in the treatment of infective endocarditis is not associated with increased risk of mortality in non-severely ill patients. *Clin Microbiol Infect*. 2016; 22:607–12.
207. Demonchy E, Dellamonica P, Roger PM, Bernard E, Cua E, Pulcini C. Audit of antibiotic therapy used in 66 cases of endocarditis. *Med Mal Inf*. 2011; 41: 602–7.
208. Parker RH, Fossieck BE. Intravenous followed by oral antimicrobial therapy for staphylococcal endocarditis. *Ann Intern Med*. 1980; 93: 832–4.
209. Dworkin RJ, Lee BL, Sande MA, Chambers HF. Treatment of right-sided *Staphylococcus aureus* endocarditis in intravenous drug users with ciprofloxacin and rifampicin. *Lancet*. 1989; 334: 1071–3.
210. Heldman AW, Hartert TV, Ray SC, Daoud EG, Kowalski TE, Pompili VJ, et al. Oral antibiotic treatment of right sided staphylococcal endocarditis in injection drug users: prospective randomized comparison with parenteral therapy. *Am J Med*. 1996; 101:68–76.
211. Colli A, Campodonico R, Gherli T. Early switch from vancomycin to oral linezolid for treatment of Gram-positive heart valve endocarditis. *Ann Thorac Surg*. 2007; 84: 87–91.
212. Iversen K, Høst N, Bruun NE, Elming H, Pump B, Christensen JJ et al. Partial oral treatment of endocarditis. *Am Heart J*. 2013 Feb;165(2):116-22.
213. Iversen K, Ihlemann N, Gill SU, Madsen T, Elming H, Jensen KT et al. Partial Oral versus Intravenous Antibiotic Treatment of Endocarditis. *N Engl J Med*. 2019 Jan 31;380(5):415-424.
214. Pries-Heje MM, Wiingaard C, Ihlemann N, Gill SU, Bruun NE, Elming H et al. Five-Year Outcomes of the Partial Oral Treatment of Endocarditis (POET) Trial. *N Engl J Med*. 2022 Feb 10;386(6):601-602.
215. Bundgaard JS, Iversen K, Pries-Heje M, Ihlemann N, Bak TS, Østergaard L et al. The impact of partial-oral endocarditis treatment on anxiety and depression in the POET trial. *J Psychosom Res*. 2022 Mar; 154:110718.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.