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## Article

# Composting Waste from the White Wine Industry

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**Abstract:** The white wine industry generates a large amount of wastes and composting is an alternative for recycling these residues with agronomic and environmental advantages. With this aim, grape marc and grape stalks were composted in static and turned piles, with 3 and 6 turns, to investigate the effects of pile conditions during composting in order to improve final compost quality. Thermophilic temperatures were attained soon after pile construction, and the highest maximum temperatures were achieved in the turned piles (70.5–71.8 °C). However, pile moisture content decreased below recommended values after day 42 in these piles. The extremely high temperatures and low moisture content in turned piles hampered OM mineralization rates and the amount of potentially mineralizable organic matter (OM<sub>0</sub>) (391–407 g kg<sup>-1</sup>), whereas the structure of the static pile provided adequate porosity to increase OM decomposition and OM<sub>0</sub> (568 g kg<sup>-1</sup>). This study shows that composting grape marc with stalks, for a period of 140 days, resulted in stabilized and matured compost (NH<sub>4</sub><sup>+</sup>-N / NO<sub>3</sub><sup>-</sup>-N <0.5), with good chemical characteristics for application as soil organic amendment in vineyards, without the need for rewetting or turning the piles, thus reducing the agronomic, and environmental cost of the composting process.

**Keywords:** compost quality; grape marc; grape stalks; organic matter mineralization

## 1. Introduction

Mediterranean vineyards are threatened by serious risks of soil deterioration due to loss of organic matter, loss of biodiversity, erosion, fertilizer contamination and compaction [1–3]. In addition, water availability will decrease due to the impact of climate change that imposes more severe warm and drought conditions on the vineyards [4–6]. Low soil moisture content due to heatwaves during the summer may have serious consequences on the quantity and quality of wine produced [7]. Under this scenario, the sustainable soil management practices are crucial to improve soil quality and increase soil water content [8,9]. Recently, several authors studied the effect of vineyards compost application to minimize threats such as extended drought or soil degradation [10–12]. The major role of organic matter in the formation of stable aggregates was described, as well as its effects on water holding capacity, aeration and resistance to root growth [13,14]. Badalikova et al. [12], for example, reported that bulk density decreased (2–10%), and consequently porosity and soil moisture content increased, 2 years after the incorporation of 30 t ha<sup>-1</sup> compost into the vines in furrows 25–30 cm deep. In addition to enhance the structure of soils and thereby improving water

availability, compost application reduces soil-born pathogen and increases nutrient use efficiency [2,15].

Composting is defined as a biological process, which degrades organic matter under conditions that allow the development of thermophilic temperatures, resulting a stabilized and sanitized product, free of weed seeds and pathogenic microorganisms [16]. The OM degradation depends on the composition of the feedstock, but also on environmental conditions such as moisture content or oxygen levels [17,18]. Excess moisture restrict the oxygen movement and anaerobic conditions may decrease the speed of composting. For this reason, the moisture content values for efficient composting are in the range 50-60% [19]. Turning the composting piles during the bio-oxidative phase provide oxygen to the decomposition process, but increases nutrient losses [20]. For this reason, the construction of the composting piles with materials that provide the adequate aeration, contribute to decrease the number of turnings and consequently increase the agronomic value of the compost [18].

Winery waste is a major environmental problem for the wine industry because the winemaking process involves the generation of a significant amount of wastes [9,21]. Winery waste compost can be a suitable way for winery wastes valorization allowing them to be reintroduced into the soil according to the principles of circular economy, which is considered a climate change mitigation strategy [11,21]. Moreover, several studies indicated that soil treatment with compost from winery wastes increase soil organic matter content and microbial diversity providing a slow release of nutrients, and high to moderate values of potassium which is considered a quality factor in vines [22–24].

The primary white winery waste is grape marc obtained after the pressing process. Usually, stalks are separated from the grapes by being fed into a de-stemmer. After pressing, grape marc without stalks have approximately 58% of skins and 42% of seeds (w/w) [25]. Wine lees are produced after the fermentation and clarification steps, and, sludge is obtained from the wastewater used for the cleaning process of the wine machinery, after dewatered aerobically in a wastewater treatment plant. Stalks and sludge may be recycled and valorised through the composting process [23]. Grape marc and lees are considered products with added value and are sent to distilleries, but, alternatively, these wastes may also be sent for composting [26].

Grape marc as a soil amendment show some disadvantages such as low pH and the presence of phytotoxic compounds such as ethanol and polyphenols which may inhibit root growth [27,28]. However, grape marc is an interesting soil fertilizer due to its richness in potassium and high organic matter content [29]. A suitable option is composting to reduce the presence of phytotoxic compounds [25]. However, the decomposition of grape marc alone may be hampered by the low pH that negatively affect microbial activity [24]. A better option is to mix grape marc with grape stalks to act as bulking agent and improve the porosity of the composting piles [30]. These authors reported that grape marc and stalks composted in the proportion of 25%:75% (grape marc: stalk, w:w) stayed in the thermophilic phase time enough to satisfy sanitation requirements, achieving a high quality compost.

The Vinho Verde Region located in the NW of Portugal is the largest wine region in Portugal with 16022 ha [31]. Wine tourism has a fundamental role in the development of Vinho Verde Region, and sustainability is pointed out as being important to improve wine tourism as wine consumers increasingly appreciate environment friendly practices [32]. However, the vineyard industry has not implemented strategies to promote a circular approach. Indeed, there is a lack of knowledge in wine industries about the composting process and application of winery waste compost in the vineyards. This requires an additional effort of growers and technical advisers [33]. Therefore, composting experiments at industrial scale are very important to develop winery waste management strategies. With this purpose, this work aims to investigate the composting process of grape marc with grape stalks, under field conditions, for compost application in vineyards as soil organic amendments.

## 2. Materials and Methods

Composting was carried out at Quinta da Torre (42°01' N; 8°29' O) with white winery waste from Anselmo Mendes winery located at the Vinho Verde Region (NW of Portugal). Grape stalks after

destemming, and grape marc (skin and seeds) obtained after pressing the juice, were collected in the same container before the fermentation process. At the composting site, this feedstock material was turned over with a backhoe loader to thoroughly mix the grape marc and stalks. Subsequently, three composting piles were built outdoors to approximately 2 m wide, 1.6 m high and 8 m long and covered with TenCate Toptex® (GEOSIN) to avoid rainfall and simultaneously allow gas exchange. The composting process was carried out for a period of 140 days. Composting treatments included turned piles with a backhoe loader, after 7, 28 and 56 days (PT3) and after 7, 14, 28, 42, 56 and 84 days (PT6) from the beginning of the composting process, and a static pile (PT0). The pile cover was removed 56 days after the commencement of the composting process to allow rain to wet the piles.

Environmental air temperature was automatically measured with a thermistor below a reflector board at 80 cm height and composting pile temperatures were monitored with thermistors placed in the core of three different parts of each pile. The temperatures were recorded every hour with a Data Logger DL2 from Delta Devices. Four replicate samples of each pile were collected for chemical analysis at the start of the process and at 7, 14, 28, 42, 56, 84 and 140 days after the beginning of composting.

Fresh samples were used to determine dry matter content (DM), pH and electrical conductivity (EC) by standard procedures [34]. Mineral N was extracted from 100 g of fresh compost using KCl 1:5 solution and  $\text{NH}_4^+\text{-N}$  and  $\text{NO}_3^-\text{-N}$  content were determined by molecular absorption spectrophotometry with a segmented flow analyser (SanPlus System, Skalar, Breda, the Netherlands). The samples were dried at 65 °C until constant weight and subsequently were milled using a rotor mill with a 2 mm sieve to determine organic matter (OM), total N, P, K, Ca and Mg. The OM content was determined by the loss of mass at 550 °C for 6 hours using a muffle furnace. The C content in the soil was calculated by dividing OM content by a factor of 1.8 [20]. Total N was determined using the modified Kjeldahl method based on a sulphuric acid digestion with a cooper selenium catalyst, using a Kjeldahl digestion unit and a compact distillation unit. The total P was measured by UV visible spectrophotometry (UV visible espectro, Thermo Scientific) after digestion with sulfuric acid and the total K, Ca and Mg were quantified by atomic spectrophotometry (Analyst 200, Perkin Elmer) after nitro-perchloric acid digestion.

The germination test was performed incubating the compost extract (1:10, w/v) with 50 seeds of radish (*Raphanus sativus* L.) and cress (*Lepidium sativum* L.) per treatment at 25 °C for 96 hours. The germination index was calculated by multiplying relative seed germination (%) and relative root growth (%) [35].

Total boron content was determined by Azomthine-H method after boiled at 550°C. The heavy metals (Cu, Zn, Pb, Cd, Cr and Ni) were extracted by digestion with aqua regia and measured by ICP-OES method (EN 14084). Total Hg was determined by USA method (EPA 7473). Prevalence of *Salmonella* spp. and *Escherichia coli* were detected according to ISO 6579 [36,37], respectively.

Losses of OM and mass reduction were calculated according to the equation 1 [38] and equation 2 [39], respectively.

$$\text{OM loss (g kg}^{-1}\text{)} = 1000 - 1000 [x_1(1000 - x_2)]/[x_2(1000 - x_1)] \quad (1)$$

$$\text{Mass reduction (g kg}^{-1}\text{)} = (1 - x_1/x_2) \times 1000 \quad (2)$$

where  $x_1$  and  $x_2$  are the initial and final ash content (g kg<sup>-1</sup>), respectively.

The first order kinetic model (equation 3) was fitted to OM mineralization determined by the OM lost [38].

$$\text{OM}_m = \text{OM}_0(1 - e^{-kt}) \quad (3)$$

where  $\text{OM}_m$  is estimated mineralized OM (g kg<sup>-1</sup> DM) at time  $t$  (day),  $\text{OM}_0$  is the amount of potentially mineralizable OM, and  $k$  is the mineralization constant rate (day<sup>-1</sup>).

Analysis of variance (ANOVA) was performed by the general linear model procedure. A probability level of  $P = 0.05$  was applied to assess differences between mean chemical characteristics from each composting treatment. All statistical calculations were performed using SPSS v. 17.0 for windows (SPSS Inc.).

### 3. Results and Discussion

#### 3.1. Temperature

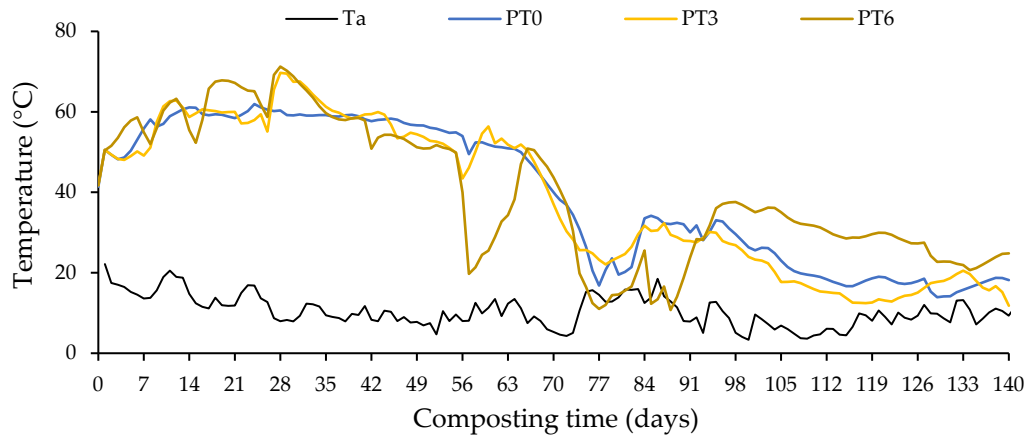
After pile construction, the temperature of the composting material rose quickly due to the rapid breakdown of the readily available OM. All composting piles reached thermophilic temperatures ( $> 50^{\circ}\text{C}$ ) in the second day of composting, and these temperatures were maintained for approximately 50 days during the composting process (Figure 1). Turning promoted aeration, increasing oxygen levels and causing microorganisms to improve their activity. Therefore, composting progressed faster and piles temperature increased after turning [40,41]. The maximum temperatures of  $70.5^{\circ}\text{C}$  and  $71.8^{\circ}\text{C}$  were registered on day 28 at PT3 and PT6, respectively. The temperature was over  $60^{\circ}\text{C}$  for 18 days in the turned piles. In contrast, PT0 temperature did not exceed  $62.2^{\circ}\text{C}$ . The maximum temperatures measured on turned piles were similar to previous published results with winery wastes [23,42,43]. For example, Carmona et al. [43] registered maximum temperatures between  $65$  and  $73^{\circ}\text{C}$  in composting piles with grape marc and grape stalks.

The temperature was above  $55^{\circ}\text{C}$  at least 31 days in all compost treatments. The sanitation requirements of 28 days at  $55^{\circ}\text{C}$  with the pile turned 3 times [44] was fulfilled for PT3 and PT6. The temperatures returned to thermophilic values within 1 to 3 days after the turnings on days 7, 14, 28 and 42. However, the lag time period for the temperatures to rise increased to 9 and 13 days after the turnings on days 56 and 84, respectively, as the composting piles became more recalcitrant to decomposition [45]. After the thermophilic phase, the OM became more stabilized, microbial activity decreased and the temperature approached to ambient levels about 100 days after starting composting, except for PT6, where temperatures remained above ambient air temperature for another month.

#### 3.2. Moisture Content

The moisture content (MC) was initially slightly above  $600\text{ g kg}^{-1}$  in all piles (Table 1), and remained between  $487\text{ g kg}^{-1}$  and  $621\text{ g kg}^{-1}$  during the composting process in PT0. However, the MC of PT3 and PT6 decreased to  $417\text{ g kg}^{-1}$  and  $335\text{ g kg}^{-1}$ , respectively, 56 days after composting commencement, because of increased evaporative water losses in turned piles compared to the static piles [46,47]. The wet conditions allowed the MC value for PT0 to persist within an adequate range for optimal composting of  $500\text{--}600\text{ g kg}^{-1}$  [19] without the need of rewetting. However, the MC in turned piles decreased to values below or near  $400\text{ g kg}^{-1}$ , decreasing microbial activity. A similar decrease on moisture content below  $400\text{ g kg}^{-1}$  on turned piles after 60 days of composting was reported for composting the solid fraction of dairy cattle slurry [48]. Therefore, it may be recommendable to reduce pile turning, because pile rewetting requires availability of water resources and increases the process complexity and cost of the turned piles. Moreover, turning increases fossil energy use and may increase the emissions of pollutant gases such as ammonia and nitrous oxide [19,40,49].





**Figure 1.** Temperature during composting of grape stalks and grape marc for the static pile (PT0), pile turned 3 times (PT3) and pile turned 6 times (PT6). Ta is the ambient air temperature.

**Table 1.** Moisture content (MC), pH and electrical conductivity (EC) during composting of grape stalks and grape marc in the static pile (PT0), pile turned 3 times (PT3) and pile turned 6 times (PT6).

Characteristic (unit)	Pile conditions	Sample day							
		0	7	14	28	42	56	84	140
MC (g kg <sup>-1</sup> )	PT0	621	581	609	563	611	487	611	567
	PT3	621	638	577	585	481	417	440	422
	PT6	621	606	552	530	439	335	402	408
	LSD	0	17	57	27	66	61	43	13
pH	PT0	3.9	5.3	4.6	5.0	4.4	7.8	8.0	7.7
	PT3	3.9	4.2	4.4	4.4	6.2	6.8	7.8	7.8
	PT6	3.9	5.0	5.5	5.6	7.3	7.7	8.1	8.1
	LSD	0.0	0.7	1.3	0.7	0.8	0.1	0.2	0.1
EC (dS m <sup>-1</sup> )	PT0	2.2	1.8	2.0	2.0	2.7	1.6	1.1	1.5
	PT3	2.2	2.0	1.9	2.1	2.3	1.7	1.6	1.6
	PT6	2.2	2.2	2.2	2.3	2.0	1.7	1.6	1.3
	LSD	0.0	0.2	0.5	0.2	0.4	0.2	0.2	0.1

LSD = least significant difference ( $P < 0.05$ ) between mean values within columns.

### 3.3. pH and Electrical Conductivity

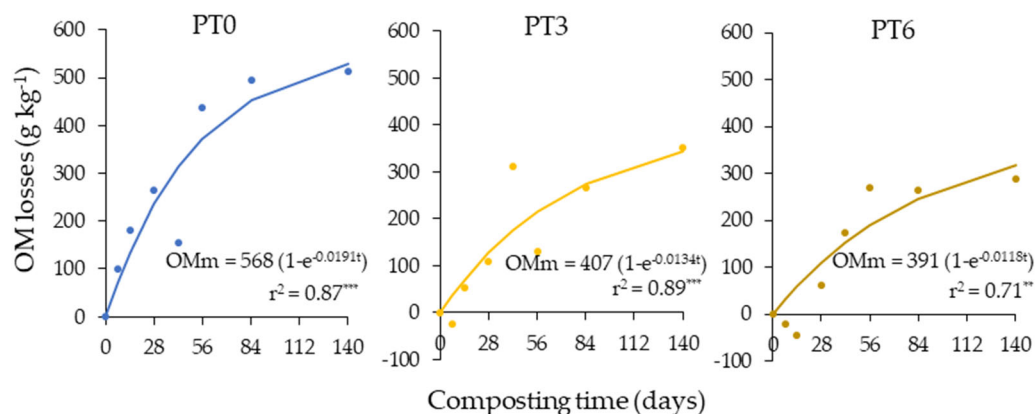
The pH value of the raw material increased during 84 days probably due to the degradation of organic acids compounds [50] and degradation of organic N leading to the formation of amines and ammonia [51]. During the first 42 days of composting the pH value was generally low decreasing the risk of NH<sub>3</sub> volatilization during the thermophilic phase of composting [52,53]. After 28 days of composting, the pH value was significantly lower for PT0 compared to PT3 and PT6, probably due to low oxygen levels that delayed the breakdown of organic acids in PT0 [47]. The final pH values was similar for all piles (7.7-8.1) and similar to pH values reported by other authors after composting grape marc with different biowastes [22,25,30].

High salt concentration may cause phytotoxicity problems on seed germination, seedling emergence and root growth. Therefore, the EC value of compost is important in evaluating the suitability and safety of compost for agricultural purposes. The EC values of the composting piles ranged from 1.3 dS m<sup>-1</sup> to 2.7 dS m<sup>-1</sup> during the composting process, with a trend towards a slight decrease in EC after 42 days of composting. This trend may be related to the transformation of OM into humic substances, increasing the cation exchange capacity (CEC) and, therefore, causing a

reduction of the EC since it quantifies the non-adsorbed and soluble salts [54]. The final compost EC values ( $1.4\text{--}1.6\text{ dS m}^{-1}$ ) were in the range of EC values reported by Patti et al. [55] for grape marc composts from four different vineyards ( $1.2\text{--}2.9\text{ dS m}^{-1}$ ), and were well below the maximum value of  $3\text{ dS m}^{-1}$  recommended for application to soil [56].

### 3.4. Organic Matter Decomposition

Organic matter was gradually decomposed and mineralized as the available C and N sources were used by microorganisms, to produce energy (60-70%) and to incorporate into their cells (30-40%) [57]. Here, mineralization of OM during composting, determined by the OM lost, was fitted to a first-order kinetic model (Figure 2) used to describe OM biodegradation by numerous researchers [29,38]. This model shows that 60-70% of the total mineralization of OM occurred during the thermophilic period (56 days). The highest OM mineralization rate was found for the static pile ( $k = 0.0191\text{ day}^{-1}$ ). In contrast, the slowest rate of composting ( $k = 0.0118\text{ day}^{-1}$ ) occurred in PT6. Thus, the largest amount of potentially mineralizable OM ( $\text{OM}_0$ ) was found for PT0 ( $568\text{ g kg}^{-1}$ ), whereas for PT3 and PT6 it decreased to  $407\text{ g kg}^{-1}$  and  $391\text{ g kg}^{-1}$ , respectively.



**Figure 2.** Organic matter (OM) losses ( $\text{g kg}^{-1}$ ) during composting of grape stalks and grape marc for the static pile (PT0), pile turned 3 times (PT3) and pile turned 6 times (PT6). \*\*  $P < 0.01$ ; \*\*\*  $P < 0.001$

After 140 days of composting, the estimated mineralized OM ( $\text{OM}_m$ ) was relatively higher for PT0 ( $529\text{ g kg}^{-1}$ ), compared to PT3 ( $345\text{ g kg}^{-1}$ ) and PT6 ( $316\text{ g kg}^{-1}$ ). Consequently, maximum overall feedstock mass reduction after 140 days of composting was found for PT0 ( $490\text{ g kg}^{-1}$ ), compared to PT3 ( $317\text{ g kg}^{-1}$ ) and PT6 ( $284\text{ g kg}^{-1}$ ).

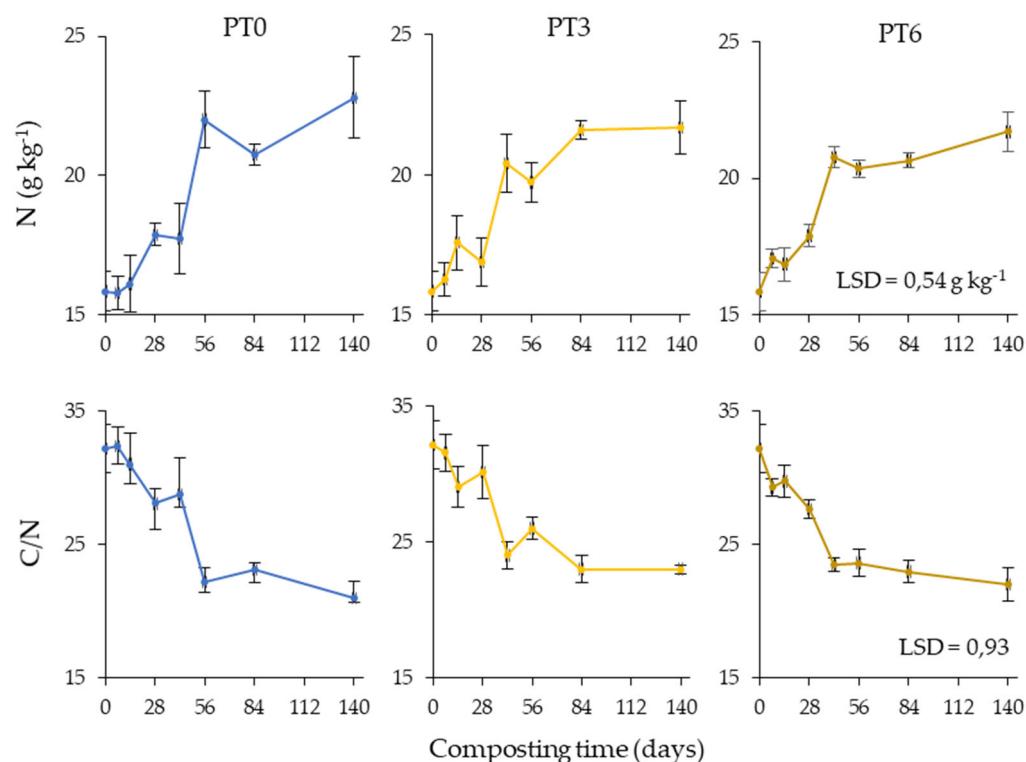
Although different raw materials with different characteristics such as moisture content and C/N ratio have different effects on OM mineralization rates [58], OM losses are usually above the values found here for PT3 and PT6. For example, OM losses between 520 and  $600\text{ g kg}^{-1}$  were found during co-composting of dairy cattle slurry with straw and gorse [59], and reached  $670\text{ g kg}^{-1}$  for cattle manure and  $720\text{ g kg}^{-1}$  for pig manure [19]. The lower composting rates found here are explained by the large amount of cellulose, hemi-celluloses and lignin resistant to microbial biodegradation of both, grape marc and stalks.

It is commonly accepted that the rate of OM mineralization increases for turned piles compared to static piles because the turned piles have higher oxygen levels causing the microorganisms to increase their activity [41,53]. However, here, OM mineralization increased for the static pile, compared to the turned piles, because of the extremely high temperatures during the thermophilic phase of composting and lower moisture content of turned piles. The temperatures generated during the composting in the turned piles exceeded largely the adequate values of  $45\text{--}55\text{ }^{\circ}\text{C}$  for maximum decomposition rates [60]. Therefore, slower mineralization rates in turned piles may be associated with high thermophilic temperatures, which may have caused a reduction in microbial activity [61]. Moreover, Bueno et al. [62] indicated that the OM decomposition is less influenced by the aeration than by the moisture content, and the moisture content of the turned piles, after 42 days of

composting ( $< 500 \text{ g kg}^{-1}$ ) was a limiting factor for the biodegradation of the grape marc residues. These results indicate that the structure of the static pile provided adequate porosity to support aerobic decomposition and to increase the decomposition rates of the feedstock material, compared to the turned piles, under the climatic conditions of the experimental site.

### 3.5. Nitrogen Transformations

Total N increased in the composting piles from  $15.8 \text{ g kg}^{-1}$  to  $21.7\text{--}22.8 \text{ g kg}^{-1}$  after 140 days (Figure 3), which is important from an agricultural point of view in terms of N fertilizer. These results are close to those reported by Bustamante et al. [63] during composting of grape marc with grape stalk (from  $14.5$  to  $23.7 \text{ g kg}^{-1}$ ). The final N content slightly increased in the static pile ( $22.8 \text{ g kg}^{-1}$ ) in comparison to the turned piles ( $21.7 \text{ g kg}^{-1}$ ). This increase in N content is explained by increased OM losses in the static pile, compared to turned piles, because the higher rate of C loss compared to N loss, increases final compost N content [57].



**Figure 3.** Total N and C/N ratio during composting of grape stalks and grape marc in the static pile (PT0), pile turned 3 times (PT3) and pile turned 6 times (PT6). LSD = least significant difference ( $P < 0.05$ ) between mean values for each characteristic. Vertical bars indicate standard deviation of mean.

The C/N ratio declined from 32 at the beginning of the composting process to values of 21 to 23 in the final composts (Table 2). It is generally recognized that the final C/N values  $< 20$  indicate that the composts reached an acceptable degree of stability [24,64]. However, the C/N ratio is not always a good indicator of stability because it depends on the type of raw material. In the case of grape marc, which is made up mainly by recalcitrant compounds such as cellulose, hemi-celluloses and lignin [65], a final C/N value for composts greater than 20 is expected, because the increased proportion of lignin reduces the availability of organic carbon in the composting process [20]. The relatively high C/N ratio at the beginning of the composting process may have contributed to the retention of  $\text{NH}_4^+$  released by OM decomposition [17,19] and to decrease  $\text{NH}_3$  volatilization [52,66].

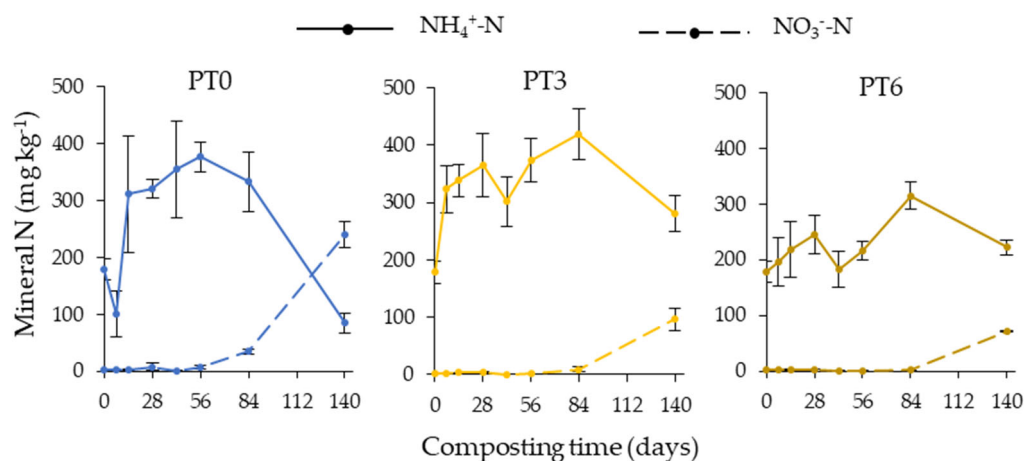


**Table 2.** Relative seed germination (RSG; %), relative root growth (RRG; %) and germination index (GI; %) of final composts.

		RSG (%)	RRG (%)	GI (%)
Cress	TP0	101	86	87
	TP3	94	87	82
	TP6	102	78	82
	LSD	11	24	24
Radish	TP0	96	118	113
	TP3	102	113	115
	TP6	106	118	125
	LSD	7	17	20

$$GI (\%) = RSG (\%) \times RRG (\%).$$

The mineral N was only 1.1% of the total N in the raw material. During the thermophilic phase of composting the  $\text{NH}_4^+\text{-N}$  content increased (Figure 4) due to the transformation of organic N into ammonium N [19,67]. The highest content of  $\text{NH}_4^+\text{-N}$  (MS) ( $314\text{--}418 \text{ mg kg}^{-1}$ ) occurred 56 and 84 days after the start of composting in the static and turned piles, respectively. The nitrification was detected by the formation of  $\text{NO}_3^-\text{-N}$  approximately 84 days after the beginning of the composting process when the temperature reached approximately  $35^\circ\text{C}$ . Indeed, the nitrification process takes place mostly below  $35^\circ\text{C}$  [51,67] because the nitrifying bacteria are suppressed at temperatures above  $40^\circ\text{C}$  [68]. After 140 days of composting, the lower content of  $\text{NH}_4^+\text{-N}$  ( $84 \text{ mg kg}^{-1}$ ) in PT0 compared to turned piles ( $222\text{--}280 \text{ mg kg}^{-1}$ ) indicated a more dynamic N transformation in PT0 during the nitrification period (Figure 4). The increased nitrification in PT0 compared to turned piles may be explained by the reduced moisture content in PT3 (42–44%) and PT6 (40–41%) compared to PT0 (57–61%), which impaired the activity of nitrifying microorganisms [69,70].

**Figure 4.** Mineral N contents during composting of grape stalks and grape marc in the pile static pile (PT0), pile turned 3 times (PT3) and pile turned 6 times (PT6). Vertical bars indicate standard deviation of mean.

### 3.6. Compost Quality

The main criteria for compost quality includes compost maturation and stability, chemical characteristics such as pH, EC, OM and nutrient contents, absence of phytotoxicity, and low levels of toxic compounds such as heavy metals [19]. Compost stability is related to the degree of OM decomposition. Labile organic compounds are degraded and, as composting progresses, other more recalcitrant compounds are partially breakdown and humified leading to the formation of stable compounds responsible for the organic fertility of the soil [71]. Here, final composts showed a high

level of stability 140 days after the start of composting, as suggested by the stable temperatures near ambient levels at the end of the composting process [64], in addition to  $\text{NH}_4^+\text{-N}$  contents well below the maximum recommended value of  $400 \text{ mg kg}^{-1}$  for stabilized composts [72]. However, the lower limit for OM losses of  $420 \text{ g kg}^{-1}$  recommended for mature composts [73], suggests that whereas composts from PT0 ( $529 \text{ g kg}^{-1}$ ) achieved a high degree of stabilization, the same was not true for PT3 ( $345 \text{ g kg}^{-1}$ ) and PT6 ( $316 \text{ g kg}^{-1}$ ).

The nitrification activity is an important indicative of compost maturation since the nitrifying bacteria are mainly active during the maturation stage [18,67]. Therefore, the ratio  $\text{NH}_4^+\text{-N} / \text{NO}_3^-\text{-N}$  is a useful index of compost maturation with values under 0.5 indicating mature composts [74]. Here, the ratio  $\text{NH}_4^+\text{-N} / \text{NO}_3^-\text{-N}$  below the upper limit value of 0.5 was only achieved for the static pile (0.35) 140 days after composting, because the OM mineralization in the turned piles was delayed, compared to the static pile.

The presence or absence of phytotoxic compounds on composts is usually assessed by germination tests that quantify seed growth [71]. Several types of seeds have been used in germination tests, with cress being the most common [75]. Conversely, Komilis et al. [76] studied the interaction between different types of seeds and composts and concluded that a germination test with radish is a valid test to assess compost stability and phytotoxicity. Here, two germination tests were set up with cress and radish to assess the presence of phytotoxic compounds on grape marc compost (Table 2). The germination index (GI) above 80% found for this compost with cress indicated the absence of phytotoxicity [35]. These results are in agreement with [25] that found GI values over 80% with grape marc compost with or without stalks during a bioassay with cress. In contrast, the high GI (113-118%) found for these composts with radish indicated a beneficial effect on seed growth, which is in line with Moldes et al. [25] that found GI values above 125% for composted grape marc with or without stalks during a germination test with rye grass seeds. Therefore, the metabolic degradation of the phytotoxic compounds of grape marc such as ethanol, acetic acid or lactic acid during composting enable the grape marc to be used as soil fertilizer unlike the raw grape marc [25]. Moreover, the absence of *Salmonella* spp. evaluated in 25 g compost samples and the counting of *Escherichia Coli*  $< 1000 \text{ UFC g}^{-1}$  meet the sanitation requirements for the final composts [44].

Final compost nutrient contents (Table 3) indicate nutrient rich composts similar to values previously reported for winery waste composts by other authors [22,55,77]. This, in addition to high compost OM content ( $860\text{-}884 \text{ g kg}^{-1}$ ), suitable pH value (7.7-8.1), low electrical conductivity ( $1.35\text{-}1.58 \text{ dS m}^{-1}$ ) and low heavy metal contents (Table 4), below the Portuguese and the European limits established for compost [44,78], indicates good quality compost. The humified nature of mature compost that promotes water retention and slow release of N, and the balance between nutrients in the final grape marc composts suggests their suitability as soil amendments for vineyards [23,79,80] However, it is crucial to ensure a moisture content during the composting process that does not harm microbial activity and, consequently, does not slow down the composting process.

Table 3. Chemical characteristics of final composts.

Pile conditions	OM ( $\text{g kg}^{-1}$ )	N ( $\text{g kg}^{-1}$ )	C/N	$\text{NH}_4^+\text{-N}$ ( $\text{mg kg}^{-1}$ )	$\text{NO}_3^-\text{-N}$ ( $\text{mg kg}^{-1}$ )	P	K ( $\text{g kg}^{-1}$ )	Ca	Mg
PT0	860	22.8	21	36.4	103.8	4.3	24.5	6.7	2.3
PT3	884	21.7	23	161.6	55.6	3.5	20.9	6.6	2.1
PT6	869	21.7	22	131.3	42.3	3.5	24.9	5.3	2.2

Nutrient content is expressed on a dry matter basis.

Table 4. Heavy metal content of final composts.

Pile conditions	B	Cu	Zn	Pb	Cd	Cr	Ni	Hg
	(mg kg <sup>-1</sup> )				(µg kg <sup>-1</sup> )			
PT0	24.0	46.9	21.8	0.27	0.11	1.85	0.66	2.6
PT3	22.9	45.7	20.7	0.28	0.15	1.88	0.64	3.2
PT6	25.2	38.7	23.3	0.31	0.12	4.93	1.52	3.1

Nutrient content is expressed on a dry matter basis .

4. Conclusions

Composting is an efficient method to recycle grape marc and stalks with environmental and agronomic advantages. The grape skins, seeds and stalks provide a feedstock material with adequate porosity to support aerobic decomposition for effective composting at industrial scale (piles with 2 m wide and 1.6 m high) without the need of turning strategies or rewetting. The moisture content in the static pile persists within an adequate range for optimal composting, whereas the turned piles require availability of water resources increasing the complexity and energy cost of the composting process. Therefore, the minimum intervention during the composting process may be recommended to produce stable, mature and sanitized compost, with high organic matter content and rich in nutrients. These characteristics, in addition to the adequate compost pH, low electrical conductivity and low levels of heavy metals, indicates that these composts are suitable for application in vineyards and other crops, to improve soil sustainability according to the principles of circular economy.

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References

1. Cataldo, E.; Salvi, L.; Sbraci, S.; Storch, P.; Mattii, G.B. Sustainable viticulture: effects of soil management in *Vitis vinifera*. *Agronomy* **2020**, *10*, 2-15. <https://doi.org/10.3390/agronomy11112359>

2. Giffard, B. Winter; S., Guidoni, S.; Nicolai, A.; Castaldini, M.; Leyer I. et al. Vineyard Management and its impacts on soil biodiversity, functions, and ecosystems services. *Front. Ecol. Evol.* **2022**, *10*, 1-21. <https://doi.org/10.3389/fevo.2022.850272>

3. Karimi, B.; Cahurel, J.; Gontier, L.; Chovelon, M., Mahé; H., Ranjard, L. A meta-analysis of the ecotoxicological impact of viticultural practices on soil biodiversity. *Environ. Chem. Lett.* **2020**, *18*, 1947-1966. <https://doi.org/10.1007/s10311-020-01050-5>

4. Fraga, H.; Malheiro, A.C.; Moutinho-Pereira, J.; Santos J.A. An overview of climate change on European viticulture. *Food Energy Secur.* **2012**, *1*, 94-110. <https://doi.org/10.1002/fes3.14>

5. Leeuwen, C.; Destrac-Irvine, A.; Dubernet, M.; Duchêne, E.; Gowdy, M.; Marguerit, E.; Pieri, P.; Parker, A.; Rességuier, L.; Ollat, N. An update on the impact of climate change in viticulture and potential adaptations. *Agronomy* **2019**, *9*, 514. <https://doi.org/10.3390/agronomy9090514>

6. Santos, J.A.; Fraga, H.; Malheiro, A.C.; Moutinho-Pereira, J.; Dinis, L., Correia, C.; Moriondo, M.; Leolini, L.; Dibari, C.; Costafreda-Aumedes, S.; Kartschall, T.; Menz, C.; Molitor, D.; Junk, L.; Beyer, M.; Schultz, H.R. A review of the potential climate change impacts and adaptation options for European viticulture. *Appl. Sci.* **2020**, *10*, 3092. <https://doi.org/10.3390/app10093092>

7. Webb, L.B.; Whetton, P.H.; Barlow, E.W.R. Climate change and winegrape quality in Australia. *Clim. Res.* **2008**, *36*, 99-111. <https://doi.org/10.3354/Cr00740>
8. Ramos, M.C. Effects of compost amendment on the available soil water and grape yield in vineyards planted after land levelling. *Agric. Water Manage.* **2017**, *191*, 67-76. <https://doi.org/10.1016/j.agwat.2017.05.013>
9. Maicas, S.; Mateo, J.J. Sustainability of wine production. *Sustainability* **2020**, *12*, 559. <https://doi.org/10.3390/su12020559>
10. Burg, P.; Masan, V.; Cizskova, A.; Badalikova, B. Impact of compost in vineyards on changes of physical properties of soil. *Engineering for Rural Development* **2019**, *22*, 575-581. <https://doi.org/10.22616/ERDev2019.18.N263>
11. Cortés, A.; Oliveira, L.F.S.; Ferrari, V.; Taffarel, S.R.; Feijoo, G.; Moreira, M.T. Environment assesment of viticulture waste valorization through composting as a biofertilisation strategie for cerealand fruit crops. *Environ. Pollut.* **2020**, *264*. <https://doi.org/10.1016/j.envpol.2020.114794>
12. Badalikova, B.; Burg, P.; Masán, V.; Prudil, J.; Jobbágy, J.; Cizková, A.; Kristof, K.; Vasinka, M. Deep placement of compost into vineyard soil affecting physical properties of soil, yield and quality of grapes. *Sustainability* **2022**, *14*, 7823. <https://doi.org/10.3390/su14137823>
13. Benbi, D.K.; Biswas, C.R.; Bawa, S.S.; Kumar, K. Influence of farmyard manure, inorganic fertilizers and weed control practices on some soil physical properties in a long-term experiment. *Soil Use Manage.* **1998**, *14*, 52-54. <https://doi.org/10.1111/j.1475-2743.1998.tb00610>
14. Mamman, E.; Ohu, J.O.; Crowther, T. Effects of soil compaction and organic matter on the early growth of maize (*Zea mays*) in a vertisol. *Int. Agrophys.* **2007**, *21*, 367-375.
15. Wall, D.H.; Nielsen, U.N.; Six, J. Soil biodiversity and human health. *Nature* **2015**, *528*, 69. <https://doi.org/10.1038/nature15744>
16. Bertoldi, M.; Vallini, G.; Pera, A. The biology of composting: A review. *Waste Manage. Res.* **1983**, *1*, 157-176.
17. Liang, Y.; Leonard, J.J.; Feddes, J.J.R.; McGill, W.B. Influence of carbon and buffer amendments on ammonia volatilization in composting. *Bioresour. Technol.* **2006**, *97*, 748-761. <https://doi.org/10.1016/j.biortech.2005.03.041>
18. Brito, L.M.; Mourão, I.; Coutinho, J.; Smith, S.R. Simple technologies for on-farm composting of cattle slurry solid fraction. *Waste Manage.* **2012**, *32*, 1332-1340. <https://doi.org/10.1016/j.wasman.2012.03.013>
19. Bernal, M.P.; Alburquerque, J.A.; Moral, R. Composting of animal manures and chemical criteria for compost maturity assessment. A review. *Bioresour. Technol.* **2009**, *100*, 5444-5453. <https://doi.org/10.1016/j.biortech.2008.11.027>
20. Brito, L.M.; Mourão, I.; Coutinho, J.; Smith, S.R. Co-composting of invasive *Acacia longifolia* with pine bark for horticultural use. *Environ. Technol.* **2015**, *36*, 1632-1642. <https://doi.org/10.1080/09593330.2014.1002863>
21. Devesa-Rey, R.; Vecino, X.; Varela-Alende, J.L.; Barral, M.T.; Cruz, J.M.; Moldes, A.B. Valorization of winery waste vs. the cost of not recycling. *Waste Manage.* **2011**, *31*, 2327-2335. <https://doi.org/10.1016/j.wasman.2011.06.001>
22. Diaz, M.J.; Madejon, E.; López, F.; López, R.; Cabrera, F. Optimization of the rate vinasse/grape marc for co-composting process. *Process Biochem.* **2002**, *37*, 1143-1150. [https://doi.org/10.1016/S0032-9592\(01\)00327-2](https://doi.org/10.1016/S0032-9592(01)00327-2)
23. Bertrand, E.; Sort, X.; Soliva, M.; Trillas, I. Composting winery waste: sludges and grape stalks. *Bioresour. Technol.* **2004**, *95*, 203-208. <https://doi.org/10.1016/j.biortech.2003.07.012>
24. Paradelo, R.; Moldes, A.B.; Barral, M.T. Evolution of organic matter during the mesophilic composting of lignocellulosic winery wastes. *J. Environ. Manage.* **2013**, *116*, 18-26. <https://doi.org/10.1016/j.jenvman.2012.12.001>
25. Moldes, A.B.; Vázquez, M.; Dominguez, J.M.; Díaz-Fierros, F.; Barral, M. Evaluation of mesophilic biodegraded grape marc as soil fertilizer. *Appl. Biochem. Biotechnol.* **2007**, *14*, 27-37. <https://doi.org/10.1007/s12010-007-9208-2>
26. Oliveira, M.; Duarte, E. Integrated approach to winery waste: waste generation and data consolidation. *Front. Environ. Sci. Eng.* **2016**, *10*, 168-176. <https://doi.org/10.1007/s11783-014-0693-6>
27. Bustamante, M.A.; Moral, R.; Paredes, C.; Pérez-Espinosa, A.; Moreno-Caselles, J.; Pérez-Murcia, M.D. Agrochemical characterization of the solid by-product and residues from the winery and distillery industry. *Waste Manage.* **2008**, *372-380*. <https://doi.org/10.1016/j.wasman.2007.01.013>
28. Dwier, K.; Hosseinian, F.; Rod, M. The market potential of grapes waste alternatives. *J. Food Res.* **2014**, *3*, 91-106. <https://doi.org/10.5539/jfr.v3n2p91>
29. Bustamante, M.A.; Paredes, C.; Marhuenda-Egea, F.C.; Perez-Espinosa, A.; Bernal, M.P.; Moral, R. Co-composting of distillery wastes with animal manures: carbon and nitrogen transformations in the evolution of compost stability. *Chemosphere* **2008**, *72*, 551-557. <https://doi.org/10.1016/j.chemosphere.2008.03.030>
30. Fernández, F.J.; Sánchez-Arias, V.; Villasenor, J.; Rodríguez, L. Evaluation of carbon degradation during co-composting of exhausted grape marc with different biowastes. *Chemosphere* **2008**, *73*, 670-677. <https://doi.org/10.1016/j.chemosphere.2008.07.007>
31. CVRVV Comissão de Viticultura da Região dos Vinhos Verdes **2022**.

32. Pergula, M.; Persiani, A.; Palese, A.M.; Di Meo, V.; Pastore, V.; D'Adamo, C.; Celano, G. Composting: the way for a sustainable agriculture. *Appl. Soil Ecol.* **2018**, *123*, 744-750. <https://doi.org/10.1016/j.apsoil.2017.10.016>
33. Acampora, A.; Preziosi, M.; Merli, R.; Lucchetti, M.C. Environment management systems in the wine industry: best practices toward a circular economy. 23<sup>rd</sup> International Sustainable Development Research Society Conference, Bogota, Colombia, **2017**.
34. CEN European Standards-Soil Improvers and Growing Media European Committee for Standardization Brussels, Belgium 1999.
35. Zucconi, F.; Pera, A.; Forte, M.; De Bertoldi, M. Evaluating toxicity of immature compost. *Biocycle* **1981**, *22*, 54-57.
36. Mooijman, K. A. The new ISO 6579-1: A real horizontal standard for detection of Salmonella, at last! *Food Microbiol.* **2018**, *71*, 2-7. <http://dx.doi.org/10.1016/j.fm.2017.03.001>.
37. Yoruk, N. G. Most probable number technique in Escherichia coli count using ISO 16649-3, ISO 7251, and rapid test enumeration device (tempo EC) methods in milk and dairy products. *J. Food Saf.* **2018**, *38*, 1-7. <https://doi.org/10.1111/jfs.12502>
38. Paredes, C.; Roig, A.; Bernal, M.P.; Sánchez-Monedero, M.A.; Cegarra, J. Evolution of organic matter and nitrogen during co-composting of olive mill wastewater with solid organic wastes. *Biol. Fertil. Soils* **2000**, *20*, 222-227.
39. Tang, J.C.; Shibata, A.; Zhou, Q.; Katayama, A. Effect of temperature on reaction rate and microbial community in composting of cattle manure with rice straw. *J. Biosci. Bioeng.* **2007**, *104*, 321-328. <https://doi.org/10.1263/jbb.104.321>
40. Hao, X.; Chang, C. Gaseous NO, NO<sub>2</sub>, and NH<sub>3</sub> loss during cattle feedlot manure composting. *Phyton-Ann Rei Bot.* **2001**, *41*, 81-93.
41. Tiquia, S.M.; Richard, T.L.; Honeyman, M.S. Carbon, nutrient, and mass loss during composting. *Nutr. Cycl. Agroecosys.* **2002**, *62*, 15-42. <https://doi.org/10.2134/jeq1997.00472425002600010027>
42. Bustamante, M.A.; Moral, R.; Paredes, C.; Vargas-Garcia, M.C.; Suárez-Estela, F.; Moreno, J. Evolution of the pathogen content during co-composting of winery and distillery wastes. *Bioresour. Technol.* **2008**, *99*, 7299-7306. <https://doi.org/10.1016/j.biortech.2007.12.051>
43. Carmona, E.; Moreno, M.T.; Avilés, M.; Ordovás, J. Composting of wine industry wastes and their use as a substrate for growing soilless ornamental plants. *Span. J. Agric. Res.* **2012**, *10*, 482-491. <https://doi.org/10.5424/sjar/2012102-320-11>
44. Portuguese Decree-law 103/2015. Decreto-Lei nº 103/2015. Diário da República, 1<sup>a</sup> série - nº 114 de 15 de Junho, 3756-3788, **2015**.
45. Brito, L.M.; Mourão, I.; Coutinho, J.; Smith, S. Composting for management and resource recovery of invasive Acacia species. *Waste Manage. Res.* **2013**, *31*, 1125-1132. <https://doi.org/10.1177/0734242X13502384>.
46. Kalamdhad, A.S.; Kazmi, A.A. Effects of turning frequency on compost stability and some chemical characteristics in a rotary drum compost. *Chemosphere* **2009**, *74*, 1327-1334. <https://doi.org/10.1016/j.chemosphere.2008.11.058>.
47. Ma, Q.; Li, Y.; Xue, J.; Cheng, D.; Li, Z. Effects of turning frequency on ammonia emission during the composting of chicken manure and soybean straw. *Molecules* **2022**, *27*, 472. <https://doi.org/10.3390/molecules27020472>.
48. Brito, L.M.; Amaro, A.L.; Fernandes, A.S. Influence of aeration on the process of composting dairy cattle slurry manure. *Revista de Ciências Agrárias* **2009**, *33*, 298-311.
49. El Kader, N.A.; Robin, P.; Paillat, J.; Leterme, P. Turning, compacting, and the addition of water as factors affecting gaseous emissions in farm manure composting. *Bioresour. Technol.* **2007**, *98*, 2619- 2628. <https://doi.org/10.1016/j.biortech.2006.07.035>.
50. Peigné, J.; Girardin, P. Environmental impacts of farm scale composting practices. *Water, Air Soil Poll.* **2004**, *153*, 45-68. <https://doi.org/10.1023/B:WATE.0000019932.04020.b6>
51. Sánchez-Monedero, M.A.; Roig, A.; Paredes, C.; Bernal, M.P. Nitrogen transformation during organic waste composting by the Rutgers system and its effect on pH, EC and maturity of the composting mixtures. *Bioresour. Technol.* **2001**, *78*, 301-308. [https://doi.org/10.1016/S0960-8524\(01\)00031-1](https://doi.org/10.1016/S0960-8524(01)00031-1).
52. Raviv, M.; Medina, S.; Krasnovsky, A.; Ziadna, H. Organic matter and nitrogen conservation in manure compost for organic agriculture. *Compost Sci. Util.* **2004**, *12*, 6-10. <https://doi.org/10.1080/1065657X.2004.10702151>.
53. Brito L.M.; Coutinho, J.; Smith S.R. Methods to improve the composting process of the solid fraction of dairy cattle slurry. *Bioresour. Technol.* **2008**, *99*, 8955-8960. <https://doi.org/10.1016/j.biortech.2008.05.005>.
54. Barros, E.S.C.; de Amorim, M.C.C.; Olszewski, N.; Silva, T.S. Composting of winery waste and characteristics of the final compost according to Brazilian legislation. *Environ. Sci. Health B.* **2021**, *5*, 447-457. <https://doi.org/10.1080/03601234.2021.1900694>
55. Patti, A.F.; Issa, G.; Smernik, R.; Wilkinson, K. Chemical composition of composted grape marc. *Water Sci. and Technol.* **2009**, *60*, 1265-1271. <https://doi.org/10.2166/wst.2009.564>.



56. Soumaré, M.; Demeyer, A.; Tack, F.M.G.; Verloo, M.G. Chemical characteristics of Malian and Belgian solid waste composts. *Bioresour. Technol.* **2002**, *81*, 97-101. [https://doi.org/10.1016/S0960-8524\(01\)00125-0](https://doi.org/10.1016/S0960-8524(01)00125-0).
57. Barrington, S.; Choinière, D.; Trigui, M.; Knight, W. Effect of carbon source on compost nitrogen and carbon losses *Bioresour. Technol.* **2002**, *83*, 189-194. [https://doi.org/10.1016/S0960-8524\(01\)00229-2](https://doi.org/10.1016/S0960-8524(01)00229-2)
58. Chinakwe, E.C.; Nwogwugwu, U.N.; Ibekwe, V.L.; Nwachukwu, I.N.; Ihejirika, C.E.; Ofoegbu, C.J.; Chinakwe, P.O.; Mejeha, O.K. Changes in microbial population numbers during composting of some organic wastes in greenhouse. *J. Adv. Microbiol.* **2019**, *17*, 1-10. <https://doi.org/10.9734/jamb/2019/v17i130132>.
59. Brito, L.M.; Mourão, I.; Coutinho, J. Physicochemical Dynamics of composting screw pressed cattle slurry amended with Italian Reygrass straw or gorse bulking agent. *Compost Sci. Util.* **2010**, *18*, 119-126. <https://doi.org/10.1080/1065657X.2010.10736944>.
60. Stentiford E.T. Composting control, principles and practice. In DeBertoldi M.; Sequi P.; Lemmes B.; Papi T. *The Science of Composting*, Eds.; Chapman and Hall, London, **1996**; pp 49-59.
61. Chang, R.; Li, Y.; Li, J.; Chen, Q.; Zhao, H. Influences of the thermophilic period on biodegradation and nitrogen loss in stimulated vegetables waste composting. *Global Ecol. Conserv.* **2019**, *18*, 623. <https://doi.org/10.1016/j.gecco.2019.e00623>.
62. Bueno, P.; Tapias, R.; López, F.; Díaz, M.J. Optimizing composting parameters for nitrogen conservation in composting. *Bioresour. Technol.* **2008**, *99*, 5069-5077. <https://doi.org/10.1016/j.biortech.2007.08.087>.
63. Bustamante, M.A.; Paredes, C.; Morales, J.; Mayoral A.M.; Moral, R. Study of composting process of winery distillery wastes using multivariate techniques. *Bioresour. Technol.* **2009**, *100*, 4766-4772. <https://doi.org/10.1016/j.biortech.2009.04.033>.
64. Raj, D.; Antil, R.S. Evaluation of maturity and stability parameters of composts prepared from agro-industrial wastes. *Bioresour. Technol.* **2011**, *102*, 2868-2873. <https://doi.org/10.1016/j.biortech.2010.10.077>.
65. Antonic, B.; Janciková, S.; Dordevic, D.; Tremlová, B. Grape pomace valorization: A systematic review and meta-analysis. *Foods* **2020**, *9*, 1627. <https://doi.org/10.3390/foods9111627>.
66. Sommer, S.G. Effect of composting on nutrient loss and nitrogen availability of cattle deep litter. *Eur. J. Agron.* **2001**, *14*, 123-133. [https://doi.org/10.1016/S1161-0301\(00\)00087-3](https://doi.org/10.1016/S1161-0301(00)00087-3)
67. Cáceres, R.; Malinska, K.; Marfà, O. Nitrification within composting: a review. *Waste Manage.* **2018**, *72*, 119-137. <https://doi.org/10.1016/j.wasman.2017.10.049>
68. Hellmann, B.; Zelles, L.; Palojarvi, A.; Bai, Q. Emission of climate-relevant trace gases and succession of microbial communities during open-windrow composting. *Appl. Environ. Microb.* **1997**, *63*, 1011-1018. <https://doi.org/10.1128/aem.63.3.1011-1018.1997>.
69. Sáez, J.A.; Clemente, R.; Bustamante, M.A.; Yañez, D.; Bernal, M.P. Evaluation of slurry management strategy and the integration of the composting technology in a pig farm. Agronomical and environmental implications. *J. Environ. Manage.* **2017**, *192*, 57-67. <https://doi.org/10.1016/j.jenvman.2017.01.040>.
70. Hwang, S.; Hanaki, K. Effects of oxygen concentration and moisture content of refuse on nitrification, denitrification and nitrous oxide production. *Bioresour. Technol.* **2000**, *71*, 159-165. [https://doi.org/10.1016/S0960-8524\(99\)90068-8](https://doi.org/10.1016/S0960-8524(99)90068-8)
71. Wu, L.; Ma, L.Q.; Martinez, G.A. Comparison of methods for evaluating stability and maturity of biosolids compost. *J. Environ. Qual.* **2000**, *29*, 424-429. <https://doi.org/10.2134/jeq2000.00472425002900020008>.
72. Zucconi, F.; De Bertoldi, M. Composts specifications for the production and characterization of composts from municipal solid waste. In: de Bertoldi, M., Ferranti, M.P., L'Hermite, P., and Zucconi, F., *Compost: Quality and use*, Eds.; Elsevier Applied Science, London, 1987, pp. 30-50.
73. Antil, R.S.; Bar-Tal, A.; Fine, P.; Hadas, A. Predicting nitrogen and carbon mineralization of composted manure and sewage sludge in soil. *Compost Sci. Util.* **2011**, *19*, 33-43. <https://doi.org/10.1080/1065657X.2011.10736974>.
74. Buchanan M.; Brinton, W.; Shields, F.; West, J.; Thompson, W. Compost Maturity Index. CCQC - California Compost Quality Council, Nevada City, USA 2001.
75. Tiquia, S. Reduction of compost phytotoxicity during the process of decomposition. *Chemosphere* **2010**, *79*, 506-512. <https://doi.org/10.1016/j.chemosphere.2010.02.040>.
76. Komilis, D.P.; Tziouvaras, I.S. A statistical analysis to assess the maturity and stability of six composts. *Waste Manage.* **2009**, *29*, 1504-1513. <https://doi.org/10.1016/j.wasman.2008.10.016>
77. Gómez-Brandón, M.; Lores, M.; Insam, H.; Dominguez, J. Strategies for recycling and valorization of grape marc. *Crit. Rev. Biotechnol.* **2019**, *39*, 437-450. <https://doi.org/10.1080/07388551.2018.1555514>
78. Commission of European Communities, Commission Staff working Document on the Management of Biowaste in the European Union, 2008.

79. Biala, J. The use of recycled organic composts in viticulture-A review of the international literature and experience. 6th International Congress on Organic Viticulture, Basel, Switzerland, 2000.
80. Evanylo, G.; Sherony, C.; Spargo, J.; Starner, D.; Brosius, M.; Hearing, K. Soil and water environmental effects of fertilizer, manure, and compost-based fertility practices in an organic vegetable cropping system. *Agr. Ecosyst. Environ.* **2008**, *127*, 50-58. <https://doi.org/10.1016/j.agee.2008.02.014>.

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