

Article

Autoantibody Correlation Signatures in Fibromyalgia and Myalgic Encephalomyelitis/Chronic Fatigue Syndrome: Association with Symptom Severity

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Abstract: (1) Background: Recent studies provide some evidence for the contribution of antibody-mediated autoimmune mechanisms to the nature of fibromyalgia (FM) and myalgic encephalomyelitis/chronic fatigue syndrome (ME/CFS). Much attention was paid to the autoantibodies (AAb) targeting G protein-coupled receptors as natural components of the immune system. However, natural AAb network is much more extensive, and has not been previously investigated in these disorders; (2) Methods: The enzyme immunoassays ELI-Viscero-Test and ELI-Neuro-Test were used to determine changes in serum content of a 33 natural AAb to neural, organ-specific and non-tissue-specific autoantigens a) in 11 FM patients with comorbid ME/CFS; b) in 11 ME/CFS patients without FM; c) in 11 healthy controls. Individual autoantibody profiles and their correlation with some clinical symptoms were analyzed. (3) Results: both patients with ME/CFS and ME/CFS+FM were characterized by more frequent and pronounced deviations in the immunoreactivity to GABA-receptors than healthy controls. Although the level of other natural AAb did not differ between study groups, AAb correlation signatures were changing in patients compared to healthy controls. Both in patients and healthy controls the level of natural AAb to various neural and tissue-specific antigens correlated with the severity of fatigue, bodily pain, depression, anxiety, physical and mental-health related quality of life. Notably, that widely different correlation patterns were observed between study groups. (4) Conclusions: Findings from this pilot study provide some evidence that the homeostasis of autoimmune relationships, which are possibly a physiological part of our immune system, may break down in FM and ME/CFS. The correlation of disease-induced perturbations in individual AAb profiles with some clinical symptoms may arise from the immune system's ability to reflect qualitative and quantitative changes in antigenic composition of the body.

Keywords: fibromyalgia; myalgic encephalomyelitis/chronic fatigue syndrome; autoantibodies; autoimmunity

1. Introduction

Fibromyalgia (FM) is a common cause of chronic, widespread, musculoskeletal pain with unclear etiopathogenesis. However, it was demonstrated in a recent breakthrough study that IgG from the patients of FM produced in mice painful sensory hypersensitivities by sensitizing peripheral nociceptive afferents[1]. IgG from patients in this study labeled satellite glial cells and neurons in vivo and in vitro, as well as human dorsal root

ganglia as well as myelinated fiber tracts and a small number of macrophages and endothelial cells in mouse dorsal root ganglia. The same author group further showed that a subset of FM patients had elevated levels of anti-spinal ganglia autoantibodies (AAb) (detected in a cell culture assay), which were associated with more severe symptoms[2]. However, a large variation between individual serum samples was observed, suggesting that only a subset of FM patients have autoreactive IgG to spinal ganglia.

Myalgic encephalomyelitis/chronic fatigue syndrome (ME/CFS) is a disabling clinical condition characterized by unexplained and persistent post exertional fatigue accompanied by a variety of symptoms related to cognitive, immunological, endocrinological, and autonomous dysfunction[3]. The common core symptoms of fatigue, sleep problems and cognitive difficulties lead to significant comorbidity between FM and ME/CFS, but it can vary depending on the diagnostic criteria used[4]. While in one study 34% of 313 patients diagnosed with ME/CFS had comorbid FM according to the old American College of Rheumatology (ACR) old diagnostic criteria of 1990, the other research group reported that the prevalence of FM among patients with ME/CFS could reach 50% when ACR 2010 newer diagnostic criteria were applied[4]. The recent meta-analysis confirmed prominent clinical overlap between FM and ME/CFS[5]. Although the exact pathogenesis of ME/CFS is still unknown, according to the most widely held hypothesis, it is a complex multifactorial syndrome with immunological, metabolic, mitochondrial, autonomic and adrenal dysfunction as the key pathogenetic mechanisms[6,7]. Emerging evidence suggests that disorders of autoimmunity play an important role in postinfectious ME/CFS and that targeting autoantibodies could be a promising treatment approach[8].

At the same time, increased level of any AAb in the serum of patients is not an ultimate sign of autoimmune disease, but can indicate changes in expression and/or excretion of the corresponding antigens[8]. From this point of view, not only increased, but also a decreased levels of AAb are relevant. Recently, a network of natural AAb against adrenergic, muscarinic and other G-protein coupled receptors (GPCR) has been described which was shown to be dysregulated in a wide spectrum of diseases (not only autoimmune ones)[9]. We assume that AAb to GPCR are only a special case of a complex natural AAb network. The purpose of our study was to compare these networks between patients suffered from ME/CFS with and without comorbid FM and health controls.

2. Materials and Methods

2.1. Patients and controls

Patients were included in the study if they met all three the most commonly used sets of ME/CFS diagnostic criteria (Fukuda et al. (1994) CFS criteria[10], the Canadian Consensus criteria of ME/CFS (2003) [11], and US Institute of Medicine, now called the National Academy of Medicine (IOM/NAM) criteria (2015)[12]). Patients were assigned to the ME/CFS(+)FM or ME/CFS(-)FM groups depending on whether they met ACR 2016 diagnostic criteria for FM[13]. The third group consisted of apparently healthy controls (HC). Individuals with any autoimmune disease and those who had any acute illness during last 3 months were excluded from the study. The study was approved by the Ethics Committee of Saint Petersburg State University. All participants gave informed consent.

2.3. Questionnaires for Symptom Scoring

Both patients and HC were assessed for the clinical symptoms (including depression and anxiety) and baseline health status using the following instruments: the Short Form 36 Health Survey (SF-36), the Multidimensional Fatigue Inventory (MFI), DePaul Symptom Questionnaire-Short form (DSQ-SF), and Hospital Anxiety and Depression Scale (HADS). The SF-36 includes the following subject-reported evaluations about current health status: physical and social functioning, physical and emotional limitations, vitality, pain, and general and mental health[14]. The MFI comprises of a 20-item self-reported

questionnaire focused on general, physical and mental fatigue, reduced activity, and reduced motivation[15]. Cognitive symptoms were tested based on the self-reported DSQ-SF questionnaire data. In particular, composite scores were calculated for cognitive symptoms by averaging the scores for the frequency and severity (ranged from 0 to 4) of two symptoms of cognitive dysfunction, included in DSQ-SF questionnaire (“Problems remembering things” and “Difficulty paying attention for a long period of time”). HADS is a reliable scale for identifying and assessing the severity of symptoms of anxiety disorders and depression, both among patients with somatic diseases and among patients with mental disorders[16]. The score 0–7 in each subscale (depression and anxiety symptoms) represents “normal,”; 8-10 points - "borderline results or doubtful case of anxiety / depression"; 11 points or more - "probable case of anxiety / depression"

2.2. Autoantibody quantification - ELISA

We defined individual normalized levels of AAb against 21 organ-specific and non-organ-specific antigens and 12 neural antigens using standardized ELISA test systems for semi-quantitative serum AAb evaluation (ELI-Viscero-test-24 and ELI-N-test-12 by Medical Research Center “Immunculus”, Moscow, Russia). The antigens used in the test systems are listed in Table 1.

Table 1. List of antigens, included in the test systems ELI-Viscero-test-24 and ELI-N-test-12

| № | Antigen | Abbreviation |
|----|--|--------------|
| 1 | Double stranded deoxyribonucleic acid | dsDNA |
| 2 | β2-glycoprotein-I | β2-GP |
| 3 | Fc-fragments of IgG | Fc-IgG |
| 4 | Collagen type IV | Collagen |
| 5 | Membrane antigen of cardiomyocytes | CoM |
| 6 | β1-adrenergic receptors of cardiomyocytes | β1 Adr Re |
| 7 | Platelet membrane antigen | TrM-03 |
| 8 | Cytoplasmic antigen of neutrophils | ANCA |
| 9 | Membrane antigen of renal glomerular cells | KiM-05 |
| 10 | Membrane antigen of pulmonary alveolocytes | LuM-02 |
| 11 | Membrane antigen of gastric wall cells | GaM-02 |
| 12 | Membrane antigen of small intestine wall cells | ItM-07 |
| 13 | Membrane antigen of colon wall cells | ScM |
| 14 | Cytoplasmic antigen of hepatocytes | HeS-08 |
| 15 | Membrane antigen of hepatocyte mitochondria | HMMP |
| 16 | Human insulin | Insulin |
| 17 | Insulin receptors | Ins-Re |
| 18 | Thyroglobulin | TG |
| 19 | Thyrotropin receptor | TSH-Re |
| 20 | Membrane antigen of adrenal medulla cells | Adr-D\C |
| 21 | Membrane antigen of sperm and prostate cells | SPr-06 |
| 22 | Neurofilament protein 200 | NF-200 |
| 23 | Glial fibrillary acidic protein | GFAP |
| 24 | S100 protein | S100 |
| 25 | Myelin basic protein | MBP |
| 26 | Voltage-dependent calcium channel | V-Ca-Channel |
| 27 | N-cholinergic receptors | Ach-Re |
| 28 | Glutamate receptors | Glu-Re |
| 29 | γ-aminobutyric acid receptors | GABA-Re |

| | | |
|----|---------------------|-------------|
| 30 | Dopamine receptors | Dopa-Re |
| 31 | Serotonin receptors | 5HT-Re |
| 32 | μ-opioid receptors | μ-Opioid-Re |
| 33 | β-endorphin | β-Endorphin |

The pooled control serum was a preparation of polyclonal immunoglobulins of the IgG class, synthesized by B-lymphocytes in response to antigenic stimuli that occurred throughout the life of donors. Immunoglobulins in the control serum were obtained from the blood serum of more than 5000 healthy donors and brought to a concentration close to physiological (16 mg/mL). Thus, pooled control serum contained population-normalized IgG class polyclonal antibodies to each of the studied antigens and was used as a universal standard for all tested antigens. Depending on the studied antigen, the pooled control serum was diluted to a final concentration, which was derived on the basis of studies of the level of autoantibodies of a large cohort of individual serum samples from healthy donors. The content of AAb to the studied antigens was evaluated in the conventional units of optical density and compared to their content in a control pool of sera from healthy donors (taken for 100%). Then the average individual immunoreactivity (AIR) of the studied samples for each of the antigens was calculated in comparison with pooled control serum according to the formula:

$$\text{AIR} = \frac{\frac{R(ag1) * 100}{R(k1)} - 100 + \frac{R(ag2) * 100}{R(k2)} - 100 + \dots + \frac{R(agN) * 100}{R(kN)} - 100}{24}$$

AIR – the average reactivity of an individual patient's serum to all studied antigens, expressed as a percentage of the average reactivity of the pooled control serum with the same antigens.

R (ag1, 2,...N) – reactivity (in units of optical density) of the patient's serum with studied antigens

R(k1, k2, ...N) – reactivity (in units of optical density) of pooled control serum with studied antigens

The normal (physiological) levels of individual AIR are restricted by the ranges –30% ... to 0% (or conditional units (CU)) of the control sample AIR.

To construct immunoreactivity profiles, the deviation (as a percentage of the individual AIR) of the patient's serum reactivity with each of the antigens was calculated using specialized software according to the formula:

$$R(dev)agN = \left(\frac{R(agN) * 100}{R(kN)} \right) - 100 - \text{AIR}$$

R(dev)agN – deviation (as a percentage of the AIR) of the patient's serum reactivity with antigen N;

R (agN) – reactivity (in units of optical density) of the patient's serum with studied antigens

R(kN) – reactivity (in units of optical density) of pooled control serum with studied antigens

The normal (physiological) R(dev)agN for each AAb is restricted by the range from –15% ... to +10% (or conditional units (CU)) from the individual AIR.

2.3. Statistical Analysis

Statistical processing was performed with the Statistica 10.0 software package. To compare the prevalence of non-physiological AAb deviations from the individual AIR Chi-squared test was applied. To compare mean R(dev)agN values between patients and HC Mann–Whitney U test was applied. In order to compare AAb correlation signatures between groups, Spearman correlation analysis was performed in each group. We used Chord diagrams to visualize the patterns of Spearman's rank correlation coefficients between AAb. Spearman correlation analysis was also performed to study relationship between the severity of different symptoms and R(dev)agN values. Differences were considered significant at $p < 0.05$.

3. Results

3.1. Subject Characteristics

The study involved 11 patients with ME/CFS+FM, 11 patients with ME/CFS who did not suffer from FM and 11 healthy controls. Patient characteristics are shown in Table 2.

Table 2. Study population characteristics. BMI – body mass index, FM – fibromyalgia, HC – healthy controls, IQR – interquartile range, ME/CFS – myalgic encephalomyelitis/chronic fatigue syndrome.

| Subject Characteristics | | ME/CFS(+) FM (n=11) | ME/CFS(-) FM (n=11) | HC (n=11) | p-values ¹ | |
|-------------------------|----------------------------------|---------------------------|---------------------------|--------------------|--------------------------|--------------------------|
| | | | | | ME/CFS(+) FM vs HC | ME/CFS(-) FM vs HC |
| Gender | Female | 8 | 9 | 8 | 1.000 | 0.655 |
| | Male | 3 | 2 | 3 | | |
| Age | Median (IQR) | 38.0 (31.0-47.0) | 30.0 (27.0-45.0) | 33.0 (27.0-49.0) 1 | 0.519 | 0.562 |
| BMI | Median (IQR) | 18.8 (17.9-19.6) | 23.8 (19.8-32.3) | 21.4 (19.3-25.4) | 0.010 | 0.193 |
| Illness | | | | | | |
| Duration | Median (IQR) | 6.0 (3.0-21.5) | 7.0 (6.0-10.5) | N/A | | N/A |
| Onset of disease | Infection-triggered | 4 | 8 | N/A | N/A | N/A |
| | Severe stress(-es)- triggered | 5 | 2 | | | |

1 – For categorical variable p-values were derived from Chi-squared test; for continuous variables, p-values were derived from Mann-Whitney U test.

ME/CFS(+)
FM and ME/CFS(-)
FM did not differ from controls in terms of gender and age. While BMI in ME/CFS(-)
FM group was similar to the BMI in HC, it was significantly lower in ME/CFS(+)
FM group. Regarding race and ethnicity, all patients and HC were Caucasians and not Hispanic.

The illness duration varied, with a range of 1 to 35 years and median value of 6.0 years in ME/CFS(+)
FM group and with a range of 2,5 to 35 years and median value of 7.0 years in ME/CFS(+)
FM group . 4 patients (33%) with ME/CFS and comorbid FM and 8 patients (73%) without comorbid FM reported an infection-triggered onset of disease.

All scales from SF-36 and MFI were significantly different between ME/CFS(+)
FM and healthy controls as well as between ME/CFS(-)
FM and healthy controls (Table 3). Both physical and mental component scores (PCS and MCS, respectively) derived from the SF-36 short survey, were, as expected, higher in the control group ($p < 0.05$), indicating better health.

Depression and anxiety levels were significantly higher in the groups of ME/CFS(-)
FM and ME/CFS(+)
FM. The median HADS-D subclass score indicated probable comorbid depression only in ME/CFS(+)
FM group, and the median HADS-A score corresponded to borderline anxiety in both groups of patients.

Table 3. Clinical symptoms, depression, anxiety, and baseline health status assessment. FM – fibromyalgia, HADS – Hospital Anxiety and Depression Scale, HC – healthy controls, IQR – interquartile range, PCS – physical component score; MCS – mental component score, ME/CFS – myalgic encephalomyelitis/chronic fatigue syndrome, MFI – multidimensional Fatigue Inventory.

| Subject Characteristics | | ME/CFS(+) FM (n=11) | ME/CFS(-) FM (n=11) | HC (n=11) | p-values | |
|-------------------------|--|---------------------------|---------------------------|-----------|-----------------------|-----------------------|
| | | | | | ME/CFS(+) FM vs HC | ME/CFS(-) FM vs HC |

| | | | | | | |
|------------------------------|----------------------|------------------|-------------------|--------------------|-------|-------|
| SF-36 Scales Median (IQR) | Physical Functioning | 40.0 (30.0-55.0) | 45.0 (30.0-75.0) | 100.0 (95.0-100.0) | 0.000 | 0.000 |
| | Role physical | 0.0 (0.0-0.0) | 0.0 (0.0-0.0) | 100.0 (50.0-100.0) | 0.000 | 0.000 |
| | Bodily pain | 41.0 (31.0-41.0) | 74.0 (22.0-100.0) | 100.0 (84.0-100.0) | 0.000 | 0.028 |
| | General Health | 35.0 (20.0-45.0) | 30.0 (25.0-40.0) | 87.0 (62.0-92.0) | 0.000 | 0.000 |
| | Vitality | 10.0 (0.0-25.0) | 10.0 (0.0-20.0) | 80.0 (55.0-85.0) | 0.000 | 0.000 |
| | Social functioning | 25.0 (0.0-50.0) | 25.0 (0.0-37.5) | 100.0 (75.0-100.0) | 0.000 | 0.000 |
| | Role emotional | 33.3 (0.0-100.0) | 0.0 (0.0-100.0) | 100.0 (66.7-100.0) | 0.040 | 0.028 |
| | Mental health | 52.0 (28.0-56.0) | 40.0 (28.0-56.0) | 68.0 (52.0-80.0) | 0.002 | 0.001 |
| | PCS | 30.6 (26.3-35.0) | 35.1 (30.0-45.5) | 57.2. (50.8-58.7) | 0.000 | 0.000 |
| MFI Scales Median (IQR) | MCS | 34.5 (21.1-43.3) | 26.0 (21.9-37.0) | 53.1 (44.5-55.1) | 0.001 | 0.000 |
| | General Fatigue | 19.0 (18.0-20.0) | 19.0 (19.0-20.0) | 7.0 (6.0-9.0) 1 | 0.000 | 0.000 |
| | Mental Fatigue | 16.0 (13.0-18.0) | 14.0 (9.0-15.0) | 6.0 (5.0-10.0) | 0.000 | 0.010 |
| HADS Median (IQR) | Physical Fatigue | 16.0 (15.0-20.0) | 18.0 (16.0-20.0) | 6.0 (5.0-9.0) | 0.000 | 0.000 |
| | Reduced Activity | 19.0 (16.0-20.0) | 19.0 (17.0-20.0) | 9.0 (5.0-11.0) | 0.000 | 0.000 |
| | Reduced Motivation | 14.0 (9.0-17.0) | 13.0 (11.0-14.0) | 9.0 (5.0-11.0) | 0.003 | 0.001 |
| | Depression subscale | 13.0 (10.0-15.0) | 11.0 (9.0-16.0) | 3.0 (0.0-4.0) | 0.000 | 0.000 |
| | Anxiety subscale | 10.0 (7.0-14.0) | 10.0 (6.0-12.0) | 4.0 (1.0-5.0) | 0.001 | 0.003 |

A significantly higher proportion of patients in ME/CFS(-)FM and ME/CFS(+)FM groups presented with abnormal levels of AAbs against GABA receptors than healthy controls (Table 4). At the same time, the proportion of patients with abnormal peaks of any other AAb in groups of patients did not differ significantly from the controls.

Table 4 Number of patients and HC with abnormal deviation (as a percentage of the AIR) of the patient's serum reactivity towards the studied antigens. FM – fibromyalgia, HC – healthy controls, ME/CFS - myalgic encephalomyelitis/chronic fatigue syndrome.

| Autoantibodies | ME/CFS(+)FM (n=11) | | ME/CFS(-)FM (n=11) | | HC (n=11) | | p-values | |
|---------------------------|-----------------------|---------|-----------------------|---------|-----------|---------|-------------------|-------------------|
| | | | | | | | ME/CFS(+)FM vs HC | ME/CFS(-)FM vs HC |
| Anti-dsDNA AAb | 2 | (18,2%) | 3 | (27,3%) | 2 | (18,2%) | 1.00 | 0.66 |
| Anti- β 2-GP AAb | 0 | (0,0%) | 3 | (27,3%) | 1 | (9,1%) | 1.00 | 0.57 |
| Anti-Fc-IgG AAb | 1 | (9,1%) | 1 | (9,1%) | 2 | (18,2%) | 0.61 | 0.61 |
| Anti Collagen AAb | 4 | (36,4%) | 1 | (9,1%) | 3 | (27,3%) | 0.68 | 0.34 |
| Anti-CoM AAb | 1 | (9,1%) | 2 | (18,2%) | 2 | (18,2%) | 0.61 | 1.00 |
| Anti β 1 Adr Re AAb | 2 | (18,2%) | 2 | (18,2%) | 3 | (27,3%) | 1.00 | 0.66 |
| Anti TrM-03 AAb | 1 | (9,1%) | 0 | (0,0%) | 3 | (27,3%) | 0.34 | 0.11 |
| Anti ANCA AAb | 1 | (9,1%) | 2 | (18,2%) | 2 | (18,2%) | 0.61 | 1.00 |
| Anti KiM-05 AAb | 1 | (9,1%) | 1 | (9,1%) | 1 | (9,1%) | 1.00 | 1.00 |

| | | | | | | | | |
|------------------------------|---|---------|---|---------|---|---------|------|------|
| Anti LuM-02 AAb | 4 | (36,4%) | 5 | (45,5%) | 6 | (54,5%) | 0.43 | 0.70 |
| Anti GaM-02 AAb | 1 | (9,1%) | 3 | (27,3%) | 2 | (18,2%) | 0.61 | 0.66 |
| Anti ItM-07 AAb | 5 | (45,5%) | 3 | (27,3%) | 5 | (45,5%) | 1.00 | 0.42 |
| Anti ScM AAb | 4 | (36,4%) | 3 | (27,3%) | 1 | (9,1%) | 0.17 | 0.34 |
| Anti HeS-08 AAb | 0 | (0,0%) | 4 | (36,4%) | 1 | (9,1%) | 1.00 | 0.53 |
| Anti HMMP AAb | 1 | (9,1%) | 0 | (0,0%) | 2 | (18,2%) | 1.00 | 0.24 |
| Anti Insulin AAb | 1 | (9,1%) | 2 | (18,2%) | 1 | (9,1%) | 1.00 | 0.61 |
| Anti Ins-Re AAb | 1 | (9,1%) | 0 | (0,0%) | 1 | (9,1%) | 1.00 | 1.00 |
| Anti TG AAb | 0 | (0,0%) | 3 | (27,3%) | 0 | (0,0%) | 1.00 | 0.11 |
| Anti TSH-Re AAb | 1 | (9,1%) | 1 | (9,1%) | 1 | (9,1%) | 1.00 | 1.11 |
| Anti-Adr-D\C AAb | 1 | (9,1%) | 3 | (27,3%) | 2 | (18,2%) | 0.61 | 0.66 |
| Anti-SPr-06 AAb | 2 | (18,2%) | 4 | (36,4%) | 3 | (27,3%) | 0.66 | 0.68 |
| Anti-NF-200 AAb | 0 | (0,0%) | 1 | (9,1%) | 1 | (9,1%) | 1.00 | 1.00 |
| Anti-GFAP AAb | 1 | (9,1%) | 0 | (0,0%) | 0 | (0,0%) | 1.00 | 1.00 |
| Anti-S100 AAb | 1 | (9,1%) | 1 | (9,1%) | 1 | (9,1%) | 1.00 | 1.00 |
| Anti-MBP AAb | 0 | (0,0%) | 0 | (0,0%) | 2 | (18,2%) | 1.00 | 0.48 |
| Anti-V-Ca-Channel AAb | 0 | (0,0%) | 1 | (9,1%) | 1 | (9,1%) | 1.00 | 1.00 |
| Anti-Ach-Re AAb | 0 | (0,0%) | 1 | (9,1%) | 0 | (0,0%) | 1.00 | 1.00 |
| Anti-Glu-Re AAb | 1 | (9,1%) | 0 | (0,0%) | 2 | (18,2%) | 1.00 | 0.48 |
| Anti-GABA-Re AAb | 6 | (54,5%) | 5 | (45,5%) | 0 | (0,0%) | 0.0 | 0.06 |
| Anti-Dopa-Re AAb | 2 | (18,2%) | 2 | (18,2%) | 2 | (18,2%) | 1.00 | 1.00 |
| Anti-5HT-Re AAb | 1 | (9,1%) | 1 | (9,1%) | 1 | (9,1%) | 1.00 | 1.00 |
| Anti- μ -Opioid-Re AAb | 1 | (9,1%) | 2 | (18,2%) | 1 | (9,1%) | 1.00 | 0.61 |
| Anti- β -Endorphin AAb | 1 | (9,1%) | 3 | (27,3%) | 2 | (18,2%) | 0.61 | 0.66 |

In order to assess the extent of abnormalities in the immunoreactivity profiles of patients, we calculate median absolute deviations of the participants' serum reactivity towards each of the analyzed antigens from AIR (median R(dev)agNs) in each of the three study groups, and then compare the obtained values between patients and controls. No significant differences have been observed.

Based on the concept of antibodyome as a functional and physiological network of AAbs, which reflects the exposome and could be disturbed in the disease process[17], we performed a correlation analysis between R(dev)agNs and some clinical characteristics (BMI, age, SF-36 and MFI-20 subscales, depression, anxiety, composite score for cognitive symptoms frequency and severity) in each of the three study groups. The significant correlations are shown in Tables 5,6,7.

Table 5 Clinical correlations of the absolute AAb deviations from AIR in the group of patients suffered from ME/CFS without comorbid FM. (Spearman correlation coefficient r and p-value)

| Subject Characteristics | Autoanti-bodies | r | p |
|---------------------------|--------------------------------------|--------|-------|
| Bodily pain | Anti KiM-05 AAb | +0,611 | 0,046 |
| | Anti ScM AAb | +0,752 | 0,008 |
| | Anti HMMP AAb | +0,795 | 0,003 |
| SF-36 Scales Median (IQR) | Anti ItM-07 AAb | +0,724 | 0,012 |
| Physical component score | No significant correlation was found | | |
| Mental component score | No significant correlation was found | | |

| | | | | |
|----------------------------|--|--------------------------------------|--------|-------|
| MFI Scales Median (IQR) | General Fatigue | Anti ItM-07 AAb | -0,659 | 0,027 |
| | | Anti-SPr-06 AAb | +0,697 | 0,017 |
| | Mental Fatigue | No significant correlation was found | | |
| | Physical Fatigue | No significant correlation was found | | |
| | Reduced Activity | Anti-V-Ca- Channel AAb | -0,620 | 0,042 |
| | | Anti-Ach- Re AAb | +0,623 | 0,041 |
| | Reduced Motivation | Anti β En- dorphin AAb | +0,665 | 0,026 |
| | Depression subscale | Anti- GABA-Re AAb | -0,639 | 0,034 |
| | HADS | Anti-SPr-06 AAb | -0,639 | 0,034 |
| | | Anti-Dopa- Re AAb | -0,762 | 0,006 |
| DSQ-SF | Composite score of cog- nitive symptoms | Anti-MBP AAb | 0,820 | 0,002 |
| | | No significant correlation was found | | |
| | Illness duration | Anti-Dopa- Re AAb | -0,652 | 0,03 |
| | | Anti-GFAP AAb | +0,668 | 0,025 |
| | | Anti TG AAb | +0,728 | 0,011 |
| | Age | Anti-Fc- IgG AAb | -0,741 | 0,009 |
| | | Anti GaM- 02 AAb | -0,636 | 0,035 |
| | | Anti HeS- 08 AAb | -0,800 | 0,003 |
| | | No significant correlation was found | | |
| | BMI | No significant correlation was found | | |

Table 6 Clinical correlations of the absolute AAb deviations from AIR in the group of patients suffered from ME/CFS with comorbid FM. (Spearman correlation coefficient r and p-value)

| Subject Characteristics | | Autoanti- bodies | r | p |
|------------------------------|--------------------------|--------------------------------------|--------|-------|
| SF-36 Scales Median (IQR) | Bodily pain | Anti-CoM AAb | +0,685 | 0,04 |
| | Physical component score | No significant correlation was found | | |
| | Mental component score | Anti-GFAP AAb | -0,774 | 0,005 |
| | | Anti ItM-07 AAb | +0,612 | 0,046 |
| | General fatigue | Anti-V-Ca- Channel AAb | +0,675 | 0,023 |

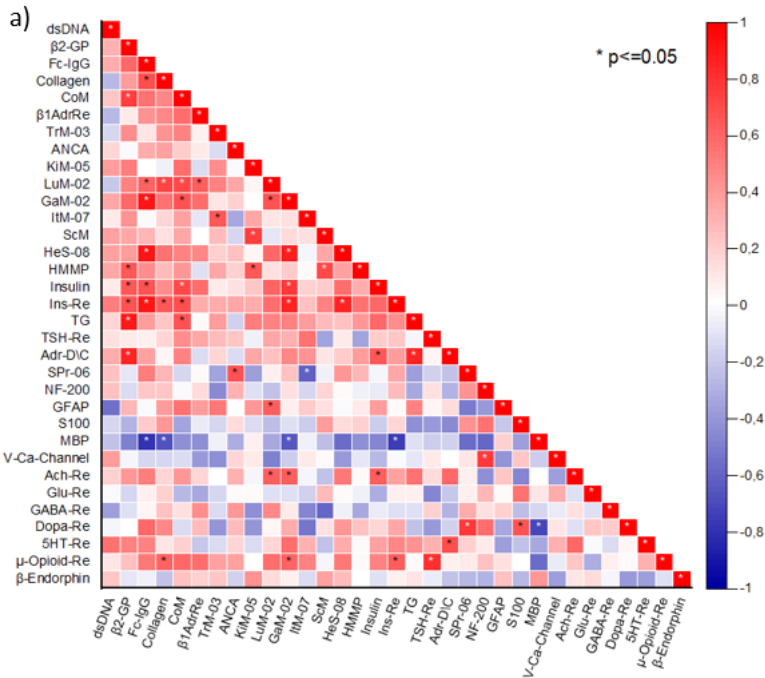
| | | | | |
|--------|---------------------------------------|--------------------------------------|--------|-------|
| HADS | Mental Fatigue | Anti β En-dorphin AAb | +0,773 | 0,005 |
| | Physical Fatigue | Anti-V-Ca-Channel AAb | +0,646 | 0,032 |
| | | Anti TG AAb | -0,785 | 0,004 |
| | | Anti TG AAb | -0,613 | 0,045 |
| | Reduced Activity | Anti- SPr-06 AAb | -0,832 | 0,001 |
| | | Anti β En-dorphin AAb | +0,629 | 0,038 |
| | Reduced Motivation | Anti-5HT-Re AAb | -0,772 | 0,005 |
| | | Anti- β 2-GP AAb | -0,629 | 0,038 |
| | Depression subscale | Anti ItM-07 AAb | -0,627 | 0,039 |
| | Anxiety subscale | Anti-dsDNA AAb | +0,700 | 0,017 |
| DSQ-SF | Composite score of cognitive symptoms | Anti-MBP AAb | +0,617 | 0,043 |
| Age | Illness duration | No significant correlation was found | | |
| BMI | | Anti-dsDNA AAb | +0,662 | 0,026 |

Table 7 Clinical correlations of the absolute AAb deviations from AIR in HC group. (Spearman correlation coefficient r and p-value)

| Subject Characteristics | | Autoantibodies | r | p |
|------------------------------|--------------------------|--------------------------------------|--------|-------|
| SF-36 Scales Median (IQR) | Bodily pain | Anti-CoM AAb | +0,774 | 0,005 |
| | | Anti- μ -Opioid-Re AAb | -0,719 | 0,013 |
| | Physical component score | Anti-CoM AAb | +0,636 | 0,035 |
| | | Anti-Dopa-Re AAb | -0,637 | 0,035 |
| | Mental component score | Anti-GABA-Re AAb | -0,664 | 0,026 |
| MFI Scales Median (IQR) | General Fatigue | No significant correlation was found | | |
| | Mental fatigue | Anti-CoM AAb | -0,647 | 0,031 |
| | Physical Fatigue | No significant correlation was found | | |
| | Reduced Activity | Anti-Adr-D\C AAb | -0,644 | 0,033 |
| | Reduced Motivation | No significant correlation was found | | |
| HADS | Depression subscale | Anti LuM-02 AAb | -0,617 | 0,043 |
| | | Anti-Adr-D\C AAb | -0,683 | 0,020 |
| | | Anti- μ -Opioid-Re AAb | +0,817 | 0,002 |
| | Anxiety subscale | Anti-Ach-Re AAb | +0,696 | 0,017 |
| | | Anti-Dopa-Re AAb | +0,646 | 0,032 |

| | | | | |
|--------|---------------------------------------|----------------------------|--------|-------|
| DSQ-SF | Composite score of cognitive symptoms | Anti- μ -Opioid-Re AAb | +0,861 | 0,001 |
| Age | | Anti Ins-Re AAb | -0,657 | 0,028 |
| BMI | | Anti-CoM AAb | -0,737 | 0,010 |
| | | Anti ItM-07 AAb | -0,618 | 0,043 |
| | | Anti HeS-08 AAb | -0,706 | 0,015 |
| | | Anti TG AAb | -0,679 | 0,022 |
| | | Anti-S100 AAb | +0,632 | 0,037 |

To identify changes in the AAb relationships in patients suffered from ME/CFS with and without concomitant FM, we analyzed AAb correlations in the two patients groups separately. A number of correlations between the absolute R(dev)agNs values in both groups were revealed. Disease-specific changes among the studied AAb were identified (Figure 1a, 1b).



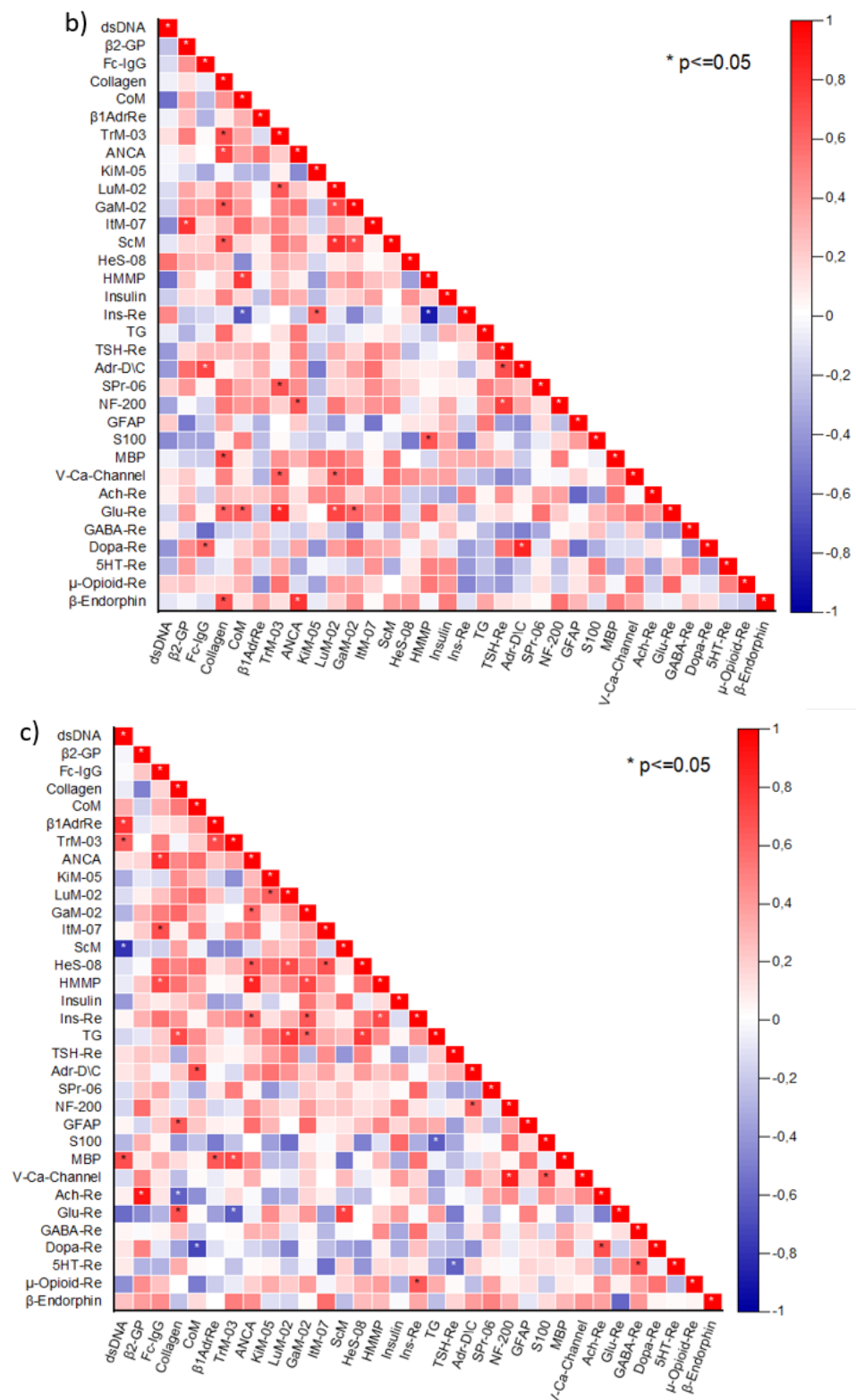
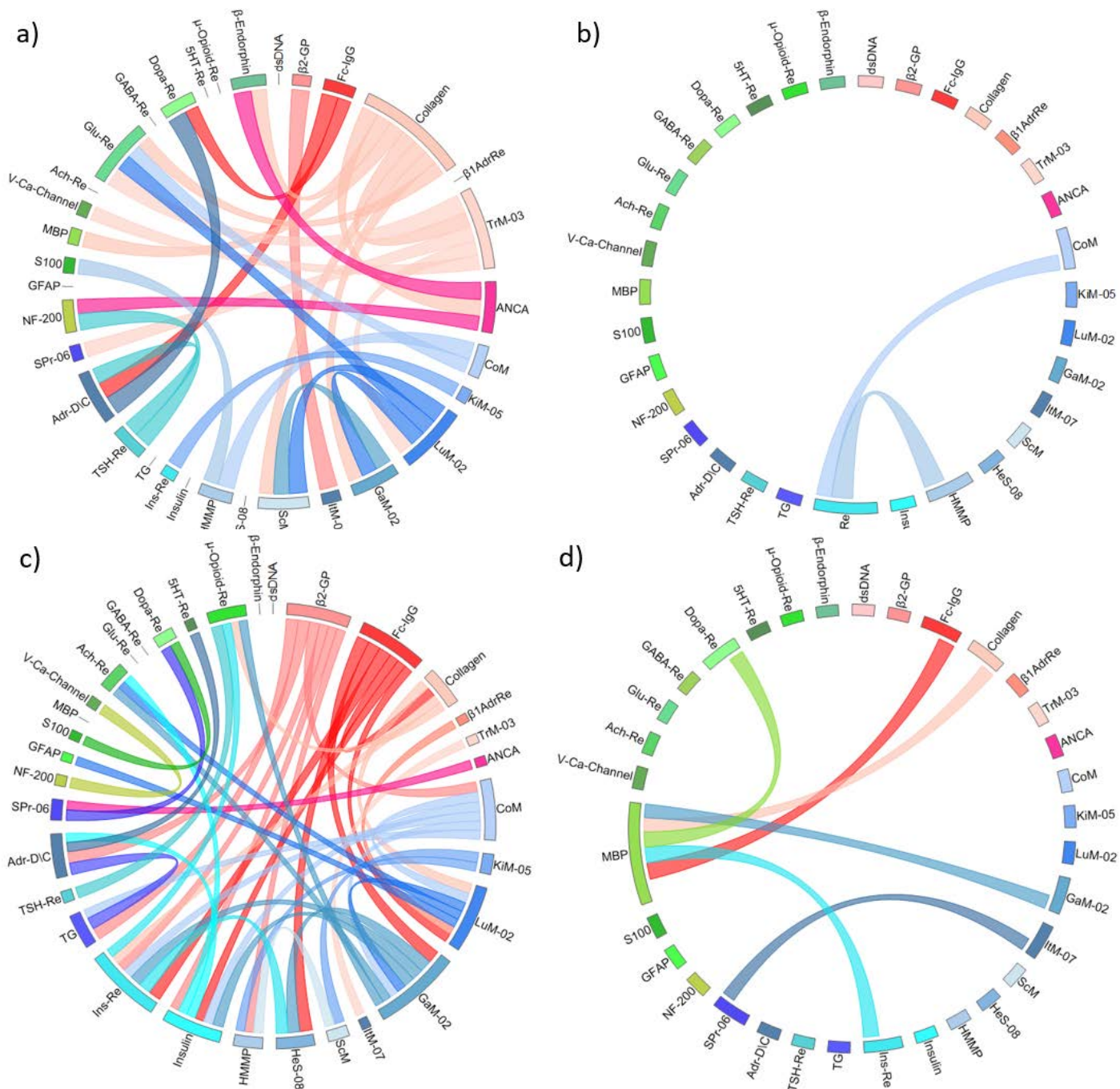


Figure 1 Correlation matrices of AAb targeting organ-specific, neural and non-organ-specific antigens (denoted by abbreviations as per legend in Table 1) for a) ME/CFS(-)FM (n = 11), b) ME/CFS(+)FM (n = 11), c) healthy controls (n = 11). The color scale bar represents the range of Spearman's rank correlation coefficient. Significant correlations are marked with *

The aforementioned data on the correlation between R(dev)agNs and some symptoms score even in HC suggested the presence of physiological relationships among natural AAb. Based on these findings, we expanded our study to include the analysis of HC

sera for correlations between the studied AAb (Figure 1c). As previously shown for a few types of AAb (especially those targeting GPCR)[9], we observed that a number of AAb targeting various organ-specific, neural and non-organ-specific antigens correlated with each other in HC.

To gain better insights into the antibodyome, we presented the significant AAb correlations for each study cohort in chord diagrams. The relationship between the AAb from three antigen groups (organ-specific, neural and non-organ-specific), the number of AAb correlations in patients and HC, and the loss of normal correlation signatures in the disease are more descriptive in these plots (Figure 2 a-f)



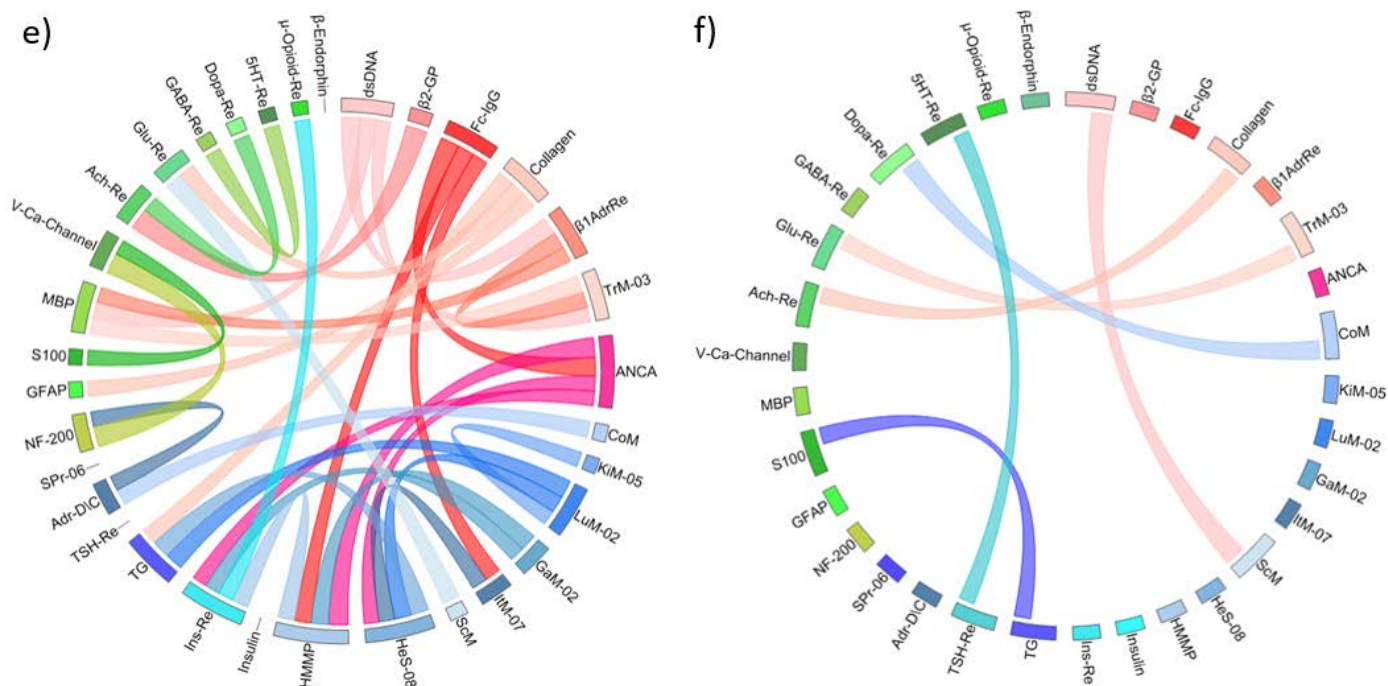


Figure 2 Chord diagrams show the correlation matrix of AAb comparing the study cohorts. Segments in circles indicate studied AAb (see Table 1 for abbreviations employed), which are grouped according to the target antigen (red/pink = organ-specific, green = neural, blue = non-organ-specific antigens). Chords linking AAb indicate significant correlations (at least $p < 0.05$) according to Spearman rank correlations, while chord thickness is directly proportional to correlation coefficient. For clarity, positive and negative correlations are shown separately. A) Positive correlations, ME/CFS(+)/FM; b) Negative correlations, ME/CFS(+)/FM; c) Positive correlations, ME/CFS(-)/FM; d) Negative correlations, ME/CFS(-)/FM; e) Positive correlations, HC; f) Negative correlations, HC.

4. Discussion

The role of autoimmunity in ME/CFS and FM is widely discussed now, largely due to the emerging data on functional anti-GPCR AAb in ME/CFS[8] and successful passive transfer of FM pain from patients to mice[1]. At the same time, it should be kept in mind that the presence of AAb does not imply of the presence of autoimmune condition, as AAb are also recognized in non-autoimmune diseases[18,19]. According to the modern interpretation of the phenomenon of physiological autoimmunity, AAb act as adaptive bioregulators of cell functions and growth such as neurotransmitters or hormones both in health and disease[20]. Moreover, there is a concept of “Immunculus” or “Immune Homoculus” based on the assumption that the network of physiological autoreactive antiidiotypic autoantibodies may dynamically reflect the whole individual antigenome as a totality of internal im-munological images of the autoantigens[21–23].

The results, obtained in our study suggest that ME/CFS and FM are rather not autoimmune diseases, but the conditions with dysregulated natural autoimmunity. In particular, while 54,5% and 45,5% of patients in ME/CFS(+)/FM and ME/CFS(-)/FM groups presented with abnormal absolute deviations of the patients’ serum reactivity against GABA Re compared to none in the control group, but these changes were not very pronounced (since no significant differences have been observed in median R(dev)agNs between the three groups). Classical autoimmune diseases with pathogenic AAb are characterized with a significant increase in the level of the AAb.

According to the concept of physiological autoimmunity, quantitative changes in the content of natural AAb are related to variations of expression and secretion of the relative antigen, reflecting functional state of the corresponding cell type[17,23]. GABA Re is an element of endogenous stress-regulating mechanisms, preventing distress[24]. The last factor has long been considered increasing a risk of ME/CFS[25]. An imbalance

between excitatory and inhibitory neurotransmission has been linked to ME/CFS and FM[26]. Interestingly, that pregabalin, which is one of the three FDA-approved drugs for the treatment of FM, is a lipophilic analogue of GABA. At the same time, it neither acts like GABA nor binds to GABA receptors, but binds strongly to the auxiliary α -2 delta subunit of the presynaptic voltage-gated calcium channel receptor to reduce the activation of postsynaptic neurotransmitter release[26]. Regarding treatment implications of our findings, it should be mentioned that a number of complementary dietary supplements have been reported to rebalance of glutamate:GABA, namely Omega-3 PUFAs, CoQ10, Withania Somnifera (Ashwagandha, Indian Ginseng), N-acetylcysteine, vitamin B12, curcumin (contained in turmeric), zinc, magnesium, 2-aminoethanesulfonic acid (L-Taurine), and carnitine (L-Carnitine)[27].

The majority of Aab, for which significant correlations with the symptom scores were identified in HC, target neural antigens, which is expected, as the analyzed symptoms are neuropsychological. This association was disturbed in both ME/CFS(+)/FM and ME/CFS(-)/FM groups – a number AAb targeting internal organ-specific and non-tissue specific antigens correlated with the symptoms scores in these groups. Thus, our findings imply that ME/CFS and FM are organic multisystem diseases, rather than psychological disorder.

The pattern of correlations found in ME/CFS(-)/FM (according to the manufacturer of the ELI-tests) suggests the role of the gut microbiom, disturbed detoxification mechanisms (namely, liver and kidney functioning) and an inflammatory process in the pelvic organs in the symptoms development. At the same time, the mechanisms, underlying the observed correlations, remains largely unclear and can differ between patients and healthy controls. For example, in HC anti-GABA-Re AAb were associated with lower mental component score, i.e. worse self-perceived mental health, while in patients from ME/CFS(-)/FM group these AAb were inversely correlated with depression.

Changes in immunoreactivity to β Endorphin were associated with more pronounced fatigue in both ME/CFS(-)/FM and ME/CFS(+)/FM groups, but not in HC. These findings suggest the dysfunction of endogenous opioid system in ME/CFS and FM. Interestingly, that a significant factor that differentiates β -endorphin from other endogenous opiates is its high affinity for and lasting effect on μ -opioid receptors[28]. When comparing patient groups and HC with regard to the associations of bodily pain with the studied AAb, it can be noted that patients from both groups abrogated the normal interconnection between anti- μ -Opioid-Re AAb and pain intensity. It was shown by Schrepf et al. in the study employed PET and fMRI together that dysregulation of the endogenous opiate system in FM could lead to less excitation in antinociceptive brain regions by incoming noxious stimulation, resulting in the hyperalgesia and allodynia commonly observed in this population[26].

Increasing evidence suggests that GFAP might be a biomarker for a number of neurological conditions, which is characterized by strong brain-specificity and high expression levels to the brain. Brain injury causes the release of GFAP and its breakdown products from injured astrocytes to the extracellular space, where these proteins equilibrate into the subarachnoid CSF compartment, then release to the circulating blood by glymphatic pathway or by diffusing pass the (possibly compromised) brain blood barrier[29]. It has been also reported that GFAP can serve as autoantigen, triggering AAb response in a subset of patients[29]. In our study changes in anti-GFAP AAb level were positively associated with worse mental component score in ME/CFS(+)/FM group and with illness duration in ME/CFS(-)/FM group.

The network-based analyses has been recently implemented in the study of physiological autoimmunity and its disturbances. In particular, distinct signatures of anti-GPCR AAb in HC, which were influenced by age, gender, and various diseases have been revealed[9].

Here, we determined the correlation signatures of some AAb targeting neural, internal organ-specific and non-organ-specific autoantigens in health and disease based on enzyme-linked immunosorbent assay (ELISA). AAb to β 2-GP, Fc-IgG, CoM, LuM-02, GaM-02 had obviously more associations with other AAb in ME/CFS(-)FM group than in HC. Danilenko et al[30] showed with the same method as in our study, that anti- β 2-GP AAb were increased in ME/CFS patients, but not in healthy participants, that alluded the link between ME/CFS and antiphospholipid syndrome earlier suspected by Berg et al. in 1999[31].

ME/CFS(+)FM group was characterized by an increase of associations of anti-collagen AAb and anti-TrM-03 AAb with other AAb. It was been recently reported that 81% of patients with ME/CFS and/or FM met Brighton criteria for hypermobility syndrome and 18% met 2017 hypermobile Ehlers–Danlos syndrome criteria. Hypermobility scores significantly predicted symptom levels in these patients[32]. Notably, that a high titer of AAb to type I collagen has been recently found in patients with undifferentiated connective tissue dysplasia and joint hypermobility[33]. Earlier such individuals also were demonstrated to be predisposed to anti-thyroid autoimmunity[34]. Another feature of AAb correlation pattern in ME/CFS(+)FM group was the abolished intragroup correlations between anti-neural AAb.

Based on our observations, we assume that AAb are natural components of the immune system and may become dysregulated not only in classical autoimmune conditions, but also in FM and ME/CFS. This assumption is in accordance with the perception of the role of the immune system in homeostatic regulation beyond host defense.

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Institutional Review Board Statement: The study was conducted in accordance with the Declaration of Helsinki, and approved by the Ethics Committee of Saint-Petersburg State University (protocol code 115-02-5 of 25-06-2020).

Informed Consent Statement: Informed consent was obtained from all subjects involved in the study.

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References

- [1] Goebel A, Krock E, Gentry C, Israel MR, Jurczak A, Urbina CM, et al. Passive transfer of fibromyalgia symptoms from patients to mice. *J Clin Invest* 2021;131. doi:10.1172/JCI144201.
- [2] Krock E, Morado-Urbina Msc CE, Menezes Msc J, Hunt Phd MA, Sandström Phd A, Kadetoff D, et al. Fibromyalgia patients with high levels of anti-satellite glia cell IgG antibodies present with more severe symptoms. *BioRxiv* 2022:2022.07.06.498940. doi:10.1101/2022.07.06.498940.
- [3] Rivera MC, Mastronardi C, Silva-Aldana CT, Arcos-Burgos M, Lidbury BA. Myalgic encephalomyelitis/chronic fatigue syndrome: A comprehensive review. *Diagnostics* 2019;9:91. doi:10.3390/diagnostics9030091.

-
- [4] Natelson BH. Myalgic Encephalomyelitis/Chronic Fatigue Syndrome and Fibromyalgia: Definitions, Similarities, and Differences. *Clin Ther* 2019;41:612–8. doi:10.1016/j.clinthera.2018.12.016.
 - [5] Ramírez-Morales R, Bermúdez-Benítez E, Martínez-Martínez LA, Martínez-Lavín M. Clinical overlap between fibromyalgia and myalgic encephalomyelitis. A systematic review and meta-analysis. *Autoimmun Rev* 2022;21. doi:10.1016/J.AUTREV.2022.103129.
 - [6] Komaroff AL. Advances in Understanding the Pathophysiology of Chronic Fatigue Syndrome. *JAMA* 2019;322:499–500. doi:10.1001/jama.2019.8312.
 - [7] Missailidis D, Annesley SJ, Fisher PR. Pathological Mechanisms Underlying Myalgic Encephalomyelitis/Chronic Fatigue Syndrome. *Diagnostics* 2019;9. doi:10.3390/DIAGNOSTICS9030080.
 - [8] Freitag H, Szklarski M, Lorenz S, Sotzny F, Bauer S, Philippe A, et al. Autoantibodies to vasoregulative g-protein-coupled receptors correlate with symptom severity, autonomic dysfunction and disability in myalgic encephalomyelitis/chronic fatigue syndrome. *J Clin Med* 2021;10. doi:10.3390/JCM10163675/S1.
 - [9] Cabral-Marques O, Marques A, Giil LM, De Vito R, Rademacher J, Günther J, et al. GPCR-specific autoantibody signatures are associated with physiological and pathological immune homeostasis. *Nat Commun* 2018;9:5224. doi:10.1038/s41467-018-07598-9.
 - [10] Fukuda K, Straus SE, Hickie I, Sharpe MC, Dobbins JG, Komaroff A. The Chronic Fatigue Syndrome: A Comprehensive Approach to Its Definition and Study. *Ann Intern Med* 1994;121:953. doi:10.7326/0003-4819-121-12-199412150-00009.
 - [11] Carruthers BM, Jain AK, De Meirleir KL, Peterson DL, Klimas NG, Lerner AM, et al. Myalgic Encephalomyelitis/Chronic Fatigue Syndrome. *J Chronic Fatigue Syndr* 2003;11:7–115. doi:10.1300/J092v11n01_02.
 - [12] Institute of Medicine. *Beyond Myalgic Encephalomyelitis/Chronic Fatigue Syndrome*. Washington, D.C.: National Academies Press; 2015. doi:10.17226/19012.
 - [13] Wolfe F, Clauw DJ, Fitzcharles MA, Goldenberg DL, Häuser W, Katz RL, et al. 2016 Revisions to the 2010/2011 fibromyalgia diagnostic criteria. *Semin Arthritis Rheum* 2016;46:319–29. doi:10.1016/j.semarthrit.2016.08.012.
 - [14] Ware JE, Sherbourne CD. The MOS 36-item short-form health survey (SF-36). I. Conceptual framework and item selection. *Med Care* 1992;30:473–83. doi:10.1097/00005650-199206000-00002.
 - [15] Smets EMA, Garssen B, Bonke B, De Haes JCJM. The multidimensional Fatigue Inventory (MFI) psychometric qualities of an instrument to assess fatigue. *J Psychosom Res* 1995;39:315–25. doi:10.1016/0022-3999(94)00125-O.
 - [16] Bjelland I, Dahl AA, Haug TT, Neckelmann D. The validity of the Hospital Anxiety and Depression Scale: An updated literature review. *J Psychosom Res* 2002;52:69–77. doi:10.1016/S0022-3999(01)00296-3.
 - [17] Poletaev A, Boura P. The immune system, natural autoantibodies and general homeostasis in health and disease. *Hippokratia* 2011;15:295.
 - [18] Meier LA, Binstadt BA. The Contribution of Autoantibodies to Inflammatory Cardiovascular Pathology. *Front Immunol* 2018;9:911. doi:10.3389/FIMMU.2018.00911.
 - [19] Fukushima K, Tsujino K, Futami S, Kida H. Natural Autoantibodies in Chronic Pulmonary Diseases. *Int J Mol Sci* 2020, Vol 21, Page 1138 2020;21:1138. doi:10.3390/IJMS21031138.
 - [20] Pashnina IA, Krivolapova IM, Fedotkina T V., Ryabkova VA, Cheresheva M V., Churilov LP, et al. Antinuclear Autoantibodies in Health: Autoimmunity Is Not a Synonym of Autoimmune Disease. *Antibodies* 2021;10:9. doi:10.3390/antib10010009.
 - [21] Cohen IR. Biomarkers, self-antigens and the immunological homunculus. *J Autoimmun* 2007;29:246–9. doi:10.1016/J.JAUT.2007.07.016.
 - [22] Poletaev A, Osipenko L. General network of natural autoantibodies as immunological homunculus (Immunculus). *Autoimmun Rev* 2003;2:264–71. doi:10.1016/S1568-9972(03)00033-8.

-
- [23] Poletaev AB, Churilov LP, Stroev YI, Agapov MM. Immunophysiology versus immunopathology: Natural autoimmunity in human health and disease. *Pathophysiology* 2012;19:221–31. doi:10.1016/j.pathophys.2012.07.003.
 - [24] Laborit H. The major mechanisms of stress. *Methods Achiev Exp Pathol* 1991;15:1–26.
 - [25] Cope H, Mann A, Pelosi A, David A. Psychosocial risk factors for chronic fatigue and chronic fatigue syndrome following presumed viral illness: a case-control study. *Psychol Med* 1996;26:1197–209. doi:10.1017/S0033291700035923.
 - [26] Baraniuk JN. Review of the Midbrain Ascending Arousal Network Nuclei and Implications for Myalgic Encephalomyelitis/Chronic Fatigue Syndrome (ME/CFS), Gulf War Illness (GWI) and Postexertional Malaise (PEM). *Brain Sci* 2022, Vol 12, Page 132 2022;12:132. doi:10.3390/BRAINSKI12020132.
 - [27] Glassford JAG. The Neuroinflammatory Etiopathology of Myalgic Encephalomyelitis/Chronic Fatigue Syndrome (ME/CFS). *Front Physiol* 2017;8:88. doi:10.3389/fphys.2017.00088.
 - [28] Clark A, Lowry P, Smyth DG. 60 YEARS OF POMC: Lipotropin and beta-endorphin: a perspective. *J Mol Endocrinol* 2016;56:T13–25. doi:10.1530/JME-16-0033.
 - [29] Yang Z, Wang KKW. Glial Fibrillary acidic protein: From intermediate filament assembly and gliosis to neurobiomarker. *Trends Neurosci* 2015;38:364. doi:10.1016/J.TINS.2015.04.003.
 - [30] Danilenko O V., Gavrilova NY, Churilov LP. Chronic Fatigue Exhibits Heterogeneous Autoimmunity Characteristics Which Reflect Etiology. *Pathophysiol* 2022, Vol 29, Pages 187-199 2022;29:187–99. doi:10.3390/PATHOPHYSIOLOGY29020016.
 - [31] Berg D, Berg LH, Couvaras J, Harrison H. Chronic fatigue syndrome and/or fibromyalgia as a variation of antiphospholipid antibody syndrome: An explanatory model and approach to laboratory diagnosis. *Blood Coagul Fibrinolysis* 1999;10:435–8. doi:10.1097/00001721-199910000-00006.
 - [32] Eccles JA, Thompson B, Themelis K, Amato ML, Stocks R, Pound A, et al. Beyond bones: The relevance of variants of connective tissue (hypermobility) to fibromyalgia, ME/CFS and controversies surrounding diagnostic classification: an observational study. *Clin Med* 2021;21:53–8. doi:10.7861/CLINMED.2020-0743.
 - [33] Babamuradova Z, Shodikulova G. Content of type I collagen antibodies and their association with clinical manifestations of undifferentiated connective tissue dysplasia. *Eur Sci* 2020;2–1:82–5.
 - [34] Churilov LP, Stroev YI, Serdyuk IY, Kaminova-Mudzhikova OM, Belyaeva IV, Gvozdetsky AN, et al. Autoimmune thyroiditis: Centennial jubilee of a social disease and its comorbidity. *Pathophysiology* 2014;21:135–45. doi:10.1016/j.pathophys.2013.11.002.