

## Identification and pharmacological activities of secondary metabolites of Phytochemicals

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### Abstract

Plant secondary metabolites are organic compounds (alkaloids, flavonoids, phenolics, tannins, anthraquinones, saponins, steroids, lignins, terpenes) produced by plant for self-defense during growth and development in order to protect them from harmful agents. As population is increasing and quality of life is reducing couple with outbreak of some diseases that are posing threat on the economy, a lot of medicinal plants have attracted interest of researchers because of the usefulness of these metabolites in pharmaceuticals, food and cosmetic industries. In this review, the pharmacological activities of some medicinal plants that are rich in secondary metabolites were studied, means of identifying and quantifying using spectrophotometry and chromatography techniques were also discussed. However, this will assist to reduce the uses and dependent on the synthetic drug and the onset of the age related diseases.

**Keywords:** secondary metabolites; pharmacological activities; qualitative and quantitative analysis; techniques.

### 1. Introduction

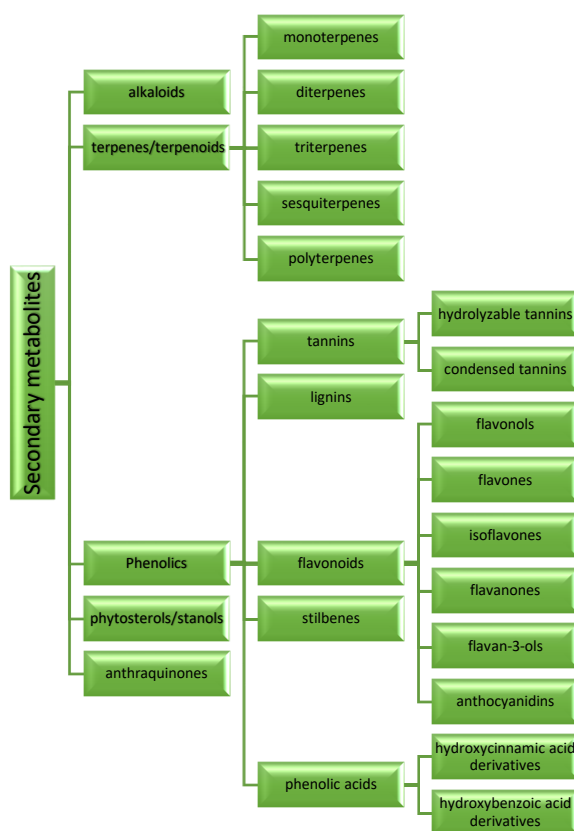
Phytochemicals are name from Greek *φυτόν* (phyton) 'plant'. They are chemical compounds naturally found in plant based and consumed as veggies, beverages, fruits, herbs, and spices contributing to positive or negative health effects. Phytochemicals present interesting biological properties like antioxidant, antibacterial, antiviral, anti-parasitic, anti-inflammatory, antidiarrheal activities. They are biologically classified as primary and secondary metabolites. The primary metabolites are carbohydrates, fats, nucleic acids (purines and pyrimidines), amino acids and chlorophyll, *etc.* They are involved in key functions of living organisms such as metabolism, respiration, growth and development. Secondary metabolites (SMs) are organic compounds that are not part of primary metabolites but are found in different part of the plants, that are used to treat, prevent and cure diseases. SMs are triggered during plants growth and development in order to protect them from harmful agents such as insects and microbes. They are also produced during stressful events such as high ultraviolet (UV) irradiation, flooding, pollution, drought, precipitation, and extreme temperatures [1-2]. SMs are induced and distributed randomly in various parts of the specific plants especially during those biotic and abiotic stressful events. They are unique, colorful with different flavor of pleasant and unpleasant aroma and are produced in low quantity. Secondary metabolites have medicinal, environmental, agricultural and industrial applications. Their applications in pharmaceuticals, food and cosmetic industries have improved the quality of life which boost the economic value, reducing the uses of synthetic drug, increase life expectancy and reduce the

onset of age related diseases such as osteoporosis, susceptible to infection, neurodegeneration, macular degeneration, chronic inflammation, hearing loss (presbycusis), cardiovascular diseases, metabolic syndrome among others.

This review discusses the pharmacological activities of some medicinal plants that are rich in secondary metabolites, the means of identifying and quantifying using spectrophotometry and chromatography techniques.

## 2. Secondary metabolites

SMs are alkaloids, flavonoids, phenolics, tannins, anthraquinones, saponins, steroids, glycosides, terpenes, *etc.* (**Figure 1**), are an alternative to many if not all synthetic drugs because of cost effectiveness and low toxicity.

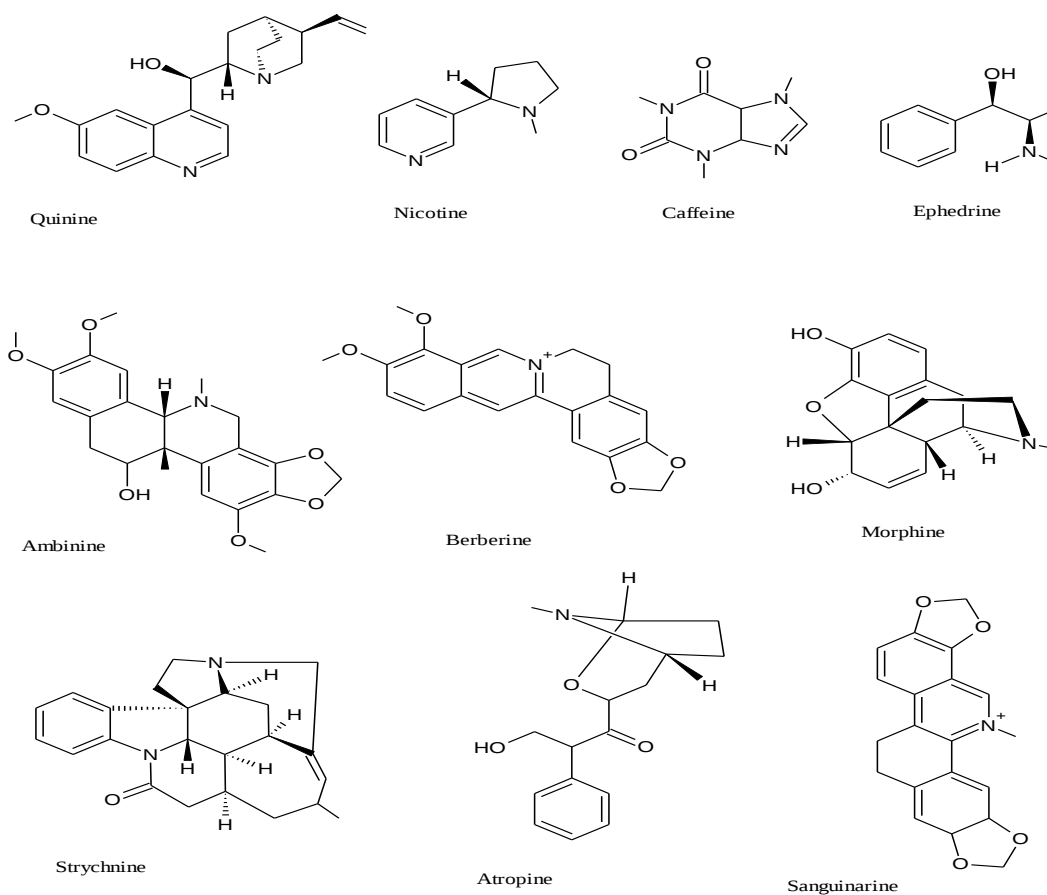


**Figure 1.** Classification of secondary metabolites

### 2.1. Alkaloids

Alkaloids are group of phytochemicals that contain at least one nitrogen in a heterocycle. They have bitter taste and other properties of alkali. Some common alkaloids are morphine, ledecorine, strychnine, berberine, atropine, ambinine, rutaecarpine, colchicine, crinamine, lycorine, scopolamine, rauvoxine, cinchonine, ephedrine, harmaline, schumannificine, salsoline, quinine, aricine, nicotine, tetrahydroalstonine, codeine, coniine, scopolamine, hyoscamine, atropine, caffeine, sangunarine, *etc.* (**Figure 2**). They are found in the roots (*Isatis indigotica*, *Stephenia tetrandra*, *Annona muricata* L, *Strychnos panganensis*, *Strychnos matopensis*, *Stemona aphylla*, *Stephania rotunda*); barks (*Alstonia boonei*, *Guatteria hispida*, *Phellodendron chinense*, *Magnolia officinalis*, *Berberis vulgaris*, *Cinchona*

*calisaya*, *Zanthoxylum leprieurii* Guill. & Perr., *Schumanniohyton magnificum* HARMS.); stems (*Picrasma quassioides*, *Esenbeckia leiocarpa*); leaves (*Vernonia amygdalina*, *Solanum nigrum*, *Nauclea officinalis*, *Pilocarpus microphyllus*, *Rauwolfia vomitoria*); bulbs (*Crinum jagus*, *Crinum glaucum*); seeds (*Hunteria umbrellata*, *Sophora alopecuroides*, *Datura innoxia*, *Datura metel*, *Datura stramonium*, *Cola nitida*, *Cola acuminata*, *Garcinia kola*, *Lupin albus* L., *Centaurea vlachorum*, *Peganum harmala*, *Papaver degenii*, *Erythrina brucei*); and rhizomes (*Sanguinaria canadensis*, *Ligusticum chuanxiong*, *Coptis chinensis*) of many herbaceous and some species of woody plants usually in tropical region. Alkaloids have some biological activity such as antimicrobial, stimulant, analgesic, anthelmintic, anticoagulant, anti-acne, and antioxidant, among others [3-4]. Alkaloids, ambinine and berberine, have been reported to possess antithrombotic and anticoagulant activities [5-6]. Tetramethylpyrazine extracted from *Chuanxiong* rhizome has been used for treatment of Ischemic stroke [7-9]. Quinine extracted from Cinchona tree are used in the treatment of malaria and has served as a source of many other drugs.

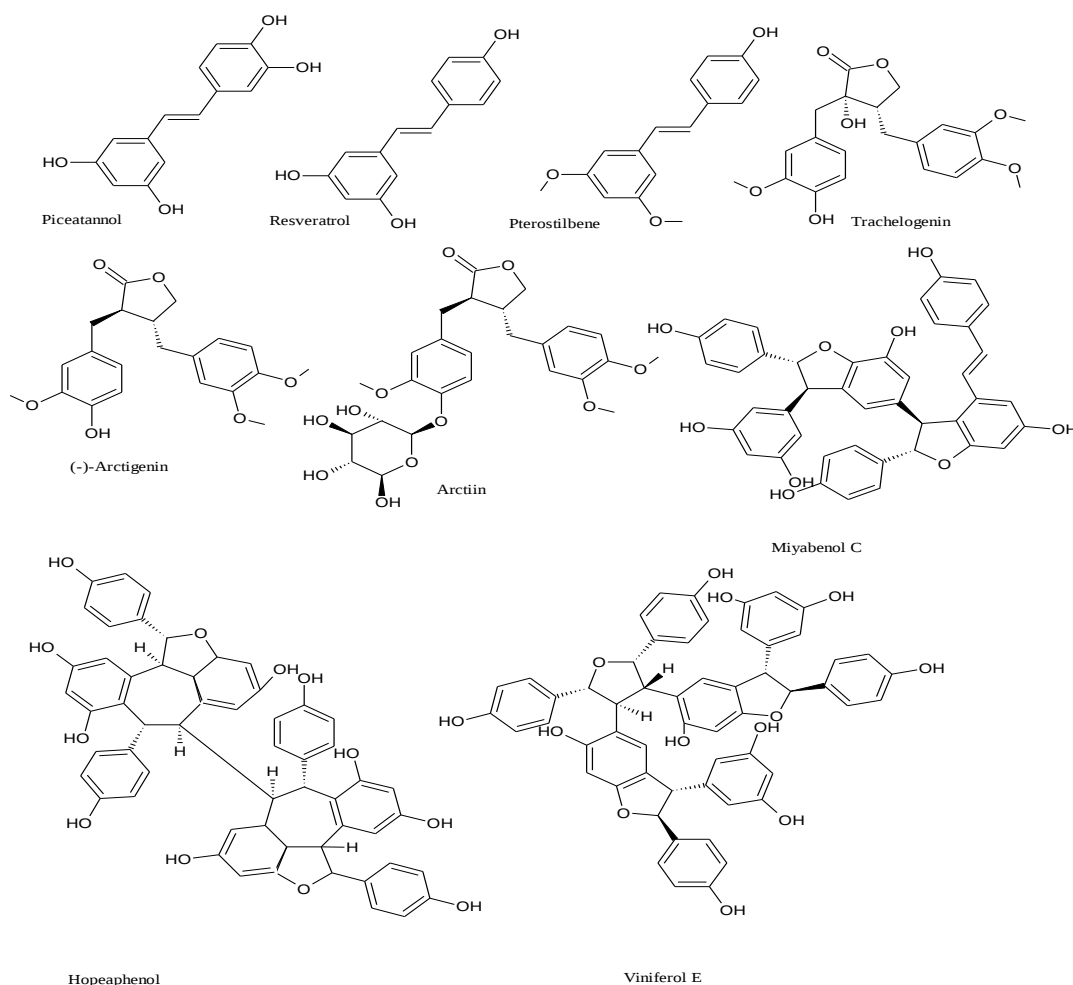


**Figure 2.** Chemical structure of some alkaloids

## 2.2. Phenolics

Phenolic compounds are chemical compounds containing one or more aromatic ring with hydroxyl group and other substituents at different positions on the compounds. They are categorized into classes depending on their structure and subcategorized within each class according to what attaches to the ring. Phenolic compounds are classified as

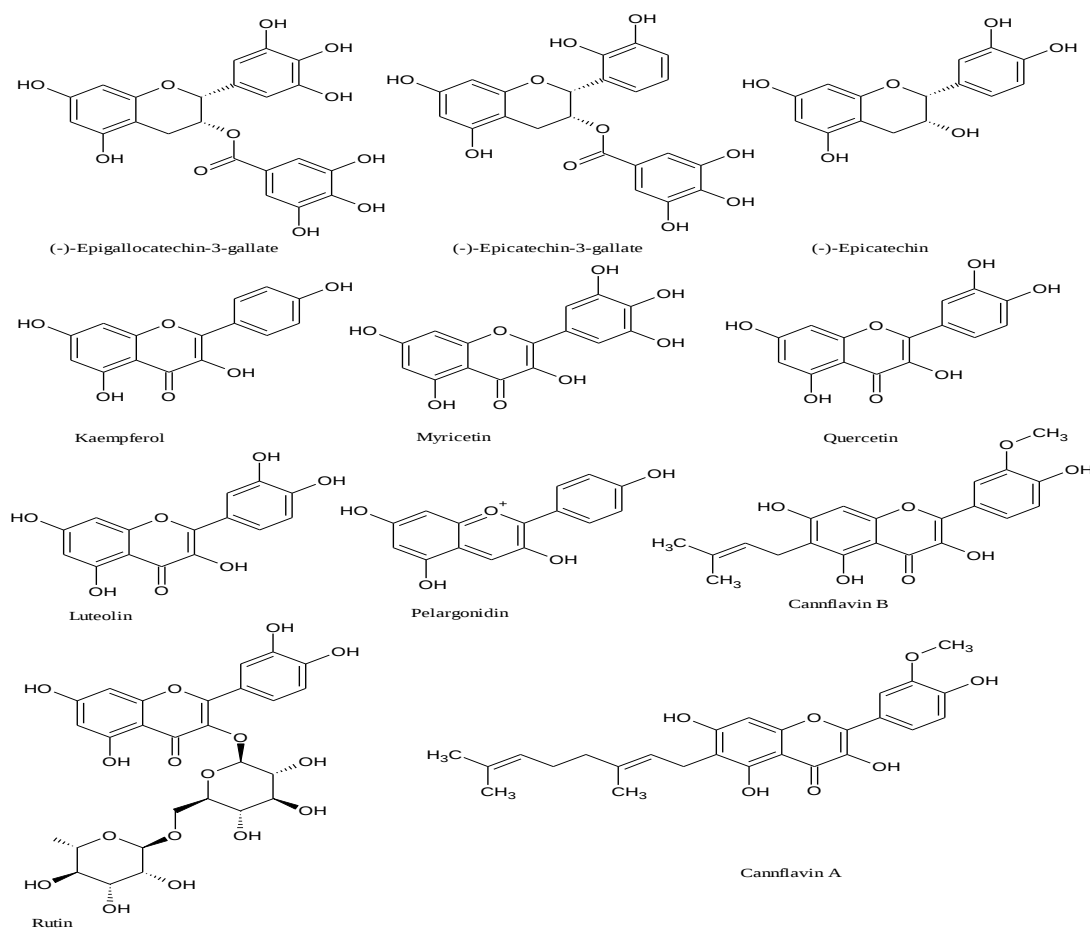
phenolic acids: hydroxybenzoic and hydroxycinnamic acids (*p*-hydroxybenzoic, ferulic, protocatechuic, vanillic, syringic acids, caffeic, ferulic, chlorogenic acid, *o*-, *m*- and *p*-coumaric acids and sinapic acids); flavonoids: flavonols, flavones, isoflavones, flavanones, flavan-3-ols, and anthocyanidins; tannins: condensed or non-hydrolyzable and hydrolyzable; lignins (arctigenin, arctiin, trachelogenin) and stilbenes (piceatannol, miyabenol, viniferol, hopeaphenol, resveratrol) (**Figure 3**). They are present in flowers, fruits, vegetables, cereals, grains, and seeds. For example, berry (blue, straw, choke, black and cran), avocado, plum, pear, cherry (sweet, sour, black), citrus (orange, grapefruit, lemon), apple, peach, potato, lettuce, spinach, coffee beans, and cider. Phenolic compounds have numerous chemicals, biologicals, agricultural, and medical studies. Many phenolic compounds are used in food, pharmaceutical, and cosmetics production because of their organoleptic properties; color, aroma, taste, and astringency [10]. The biological activities of phenolic compounds are exploited in pharmaceutical industries because of their anti-inflammatory, antioxidant, anti-hypertensive, anti-microbial, hepatoprotective properties, antidiabetic, and anticancer, etc. [11]. Arctigenin, arctiin and trachelogenin have been reported as anti-HIV, anticancer [12-13].



**Figure 3.** Chemical structures of some lignins and stilbenes

### 2.3. Flavonoids

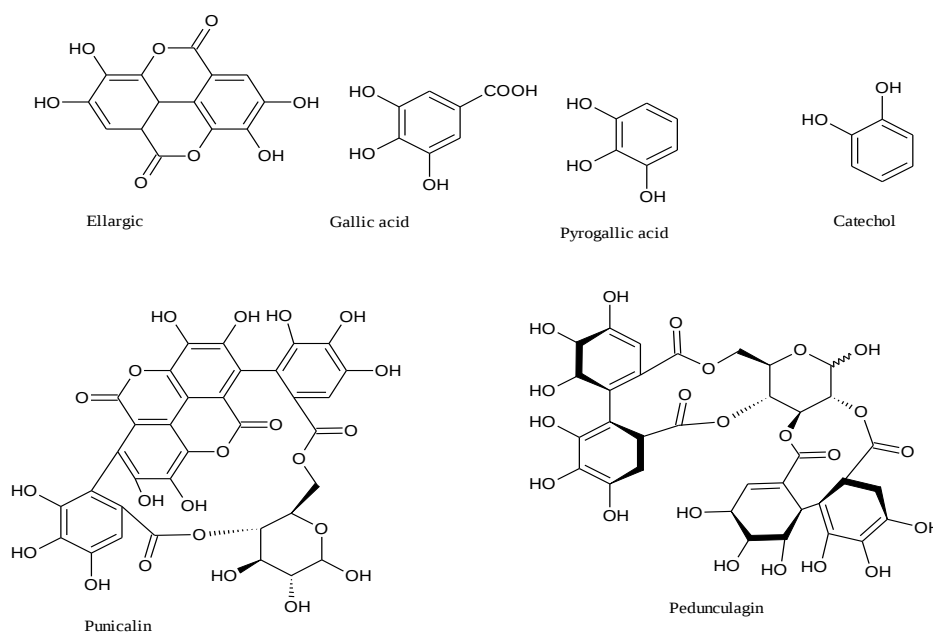
Flavonoids are low molecular weight polyphenols. They are phenolic compounds found in onions, blueberries and other berries, black tea, green tea and oolong tea, bananas, cocoa, all citrus fruits, cannabis, sea-buckthorns, buckwheat, *etc.*, with health benefit. Flavonoids have six major subgroups such as flavonols: quercetin, isorhamnetin, kaempferol, myricetin; flavones: fisetin, luteolin, apigenin; isoflavones: genistein, daidzein, glycitein; flavanones: hesperetin, naringenin, eriodictyol; flavan-3-ols: (+)-catechin, (+)-gallocatechin, (-)-epicatechin, (-)-epigallocatechin, (-)-epicatechin-3-gallate, (-)-epigallocatechin-3-gallate, theaflavin, theaflavin-3-gallate, theaflavin-3'-gallate, theaflavin-3,3'-digallate, thearubigins; anthocyanidins: cyanidin, delphinidin, malvidin, pelargonidin, peonidin, petunidin *etc.* (**Figure 4**). Flavonoids have been reported to reduce the risk of all-causing mortality like cardiovascular diseases (CVD), cancer, Parkinson disease (PD) [14-15]. They have been reported to have antioxidant, anti-inflammatory, anti-apoptotic and neuroprotection [16]. Epigallocatechin-3-gallate, epicatechin gallate and gallocatechin-3-gallate have been reported to have antiviral activities such SARS, SARS-CoV, SAR-CoV and SAR-CoV-2 [17-19]. The flavonoids kaempferol, fisetin, quercetin, *etc.* have been shown to spike protein against SAR-CoV-2 where they have low binding free energy with hACE2-S protein complex [20]. Recently, cannflavin A and B have been reported as potent anticancer, anti-inflammatory and neuroprotector [21- 24].



**Figure 4.** Chemical structure of some flavonoids

#### 2.4. Tannins

Tannins (or tannic acid) are water-soluble phenolic compounds with a molecular weight greater than 500 Da. They have strong affinities for proteins. Tannins are found in different part of the plant like the bark of trees (*Terminalia arjuna*, *Senna auriculata*, *Casuarina equisetifolia*, *Acacia mearnsii*); wood (*Sterculia oblonga*, *Castanea sativa*, *Schinopsis balansae*); fruits (*Sideroxylon obtusifolium* (Roem. & Schult.); seeds (*Doliocarpus multiflorus*, *Tetrapteryx macrocarpa*, *Acacia nilotica*, *Theobroma cacao*, *Platypodium elegans*); and roots (*Potentilla alba*, *Waldsteinia geoides*, *Casuarina equisetifolia*); leaves (*Camellia sinensis*, *Athrixia phylicoides* L., *Psidium Guajava* L., *Carapa procera* DC., *Nephelium lappaceum* L., *Populus tremuloides*, *Pericopsis laxiflora* (Benth. ex Baker), *Camellia formosensis*); flowers (*Punica granatum*, *Hibiscus Sabdariffa*, L.); buds (*Camellia japonica* L.). Many researchers have found tannins to exhibit potent biological properties such as antioxidant, antibacterial, antiviral, anti-parasitic, anti-inflammatory, antidiarrheal activities etc. [25- 27]. Tannins are classified as non-hydrolyzable (condensed tannin) (catechol-type, proanthocyanidins (procyanidins), profisetinidin, polyflavonoid, pyrocatecollic-type) and hydrolysable (ellagic, acid, camelliins A and B, punicatannin C, punicalin, punicalagin, pedunculagin, gallic acid, and pyrogalllic acid) (**Figure 5**). Tannins are used as astringent herbs for hemostatics, gastrointestinal tract, renal, pulmonary, cardiovascular, female reproductive tract and so on. Gallic acid has a wide range of industrial uses: standard for determining phenolic content of analytes in pharmaceutical industry; material for ink, paints and color developer; and in food industry as antioxidants and preservatives.

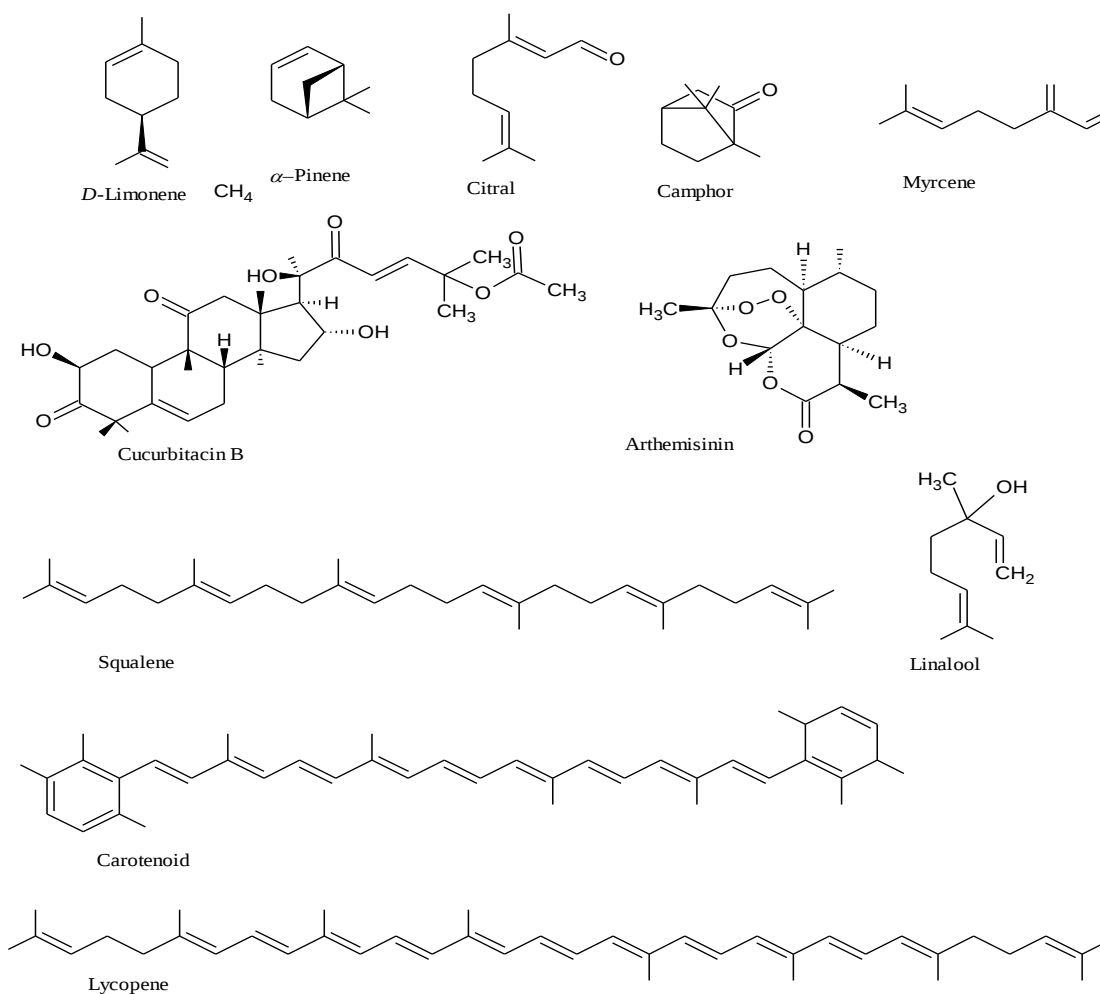


**Figure 5.** Chemical structure of some tannins

## 2.5. Terpenoids

Terpenoids are phytochemicals with chemical units of isoprene ( $C_5H_8$ ) $_n$  where  $n$  represents the number of isoprenes present in the compound. They are found in spices (*Syzygium aromaticum* L., *Zingiber officinale*, *Thymus vulgaris* L., *Curcuma longa*, *Cinnamomum zeylanicum*, *Cinnamomum cassia*, *Salvia miltiorrhiza* Bunge, *Salvia lavandulaefolia*, *Salvia officinalis*), tea (*Camellia sinensis*), cannabis (*Cannabis sativa*), citrus fruits (*Citrus reticulata*, *Citrus sinensis*,

*Citrus aurantium*, *Citrus limon*), *Artemisia annua* L., *Ekebergia capensis*, *Salvia divinorum*, *Thapsia garganica*, *Ricinus communis*, *Ginkgo biloba*, *Crocus sativum* L., etc. Some examples are of terpenes and terpenoids are limonene, citral, squalene, carotenoids artemisinin, lycopene, curcubitacin, camphor, and  $\alpha$ -pinene (**Figure 6**). Different authors have evaluated potential usefulness of terpenes and terpenoids to exhibit anti-inflammatory, antioxidant, antidiabetic, anticancer, geroprotector, and antiatherosclerotic effects [28-29]. *D*-limonene, Linalool,  $\alpha$ -pinene, camphor, myrcene, citral, cucurbitacin B have been reported to be active in inflammatory and oxidative stress diseases [29-31]. Carotenoids, are important natural pigments in carrot, tomatoes etc., pro-vitamin A are rich sources of antioxidant [32]. Artemisinin, a terpenoid from *Artemisia annua*, is used to treat fever, inflammation, malaria and kidney disorder [33-35].

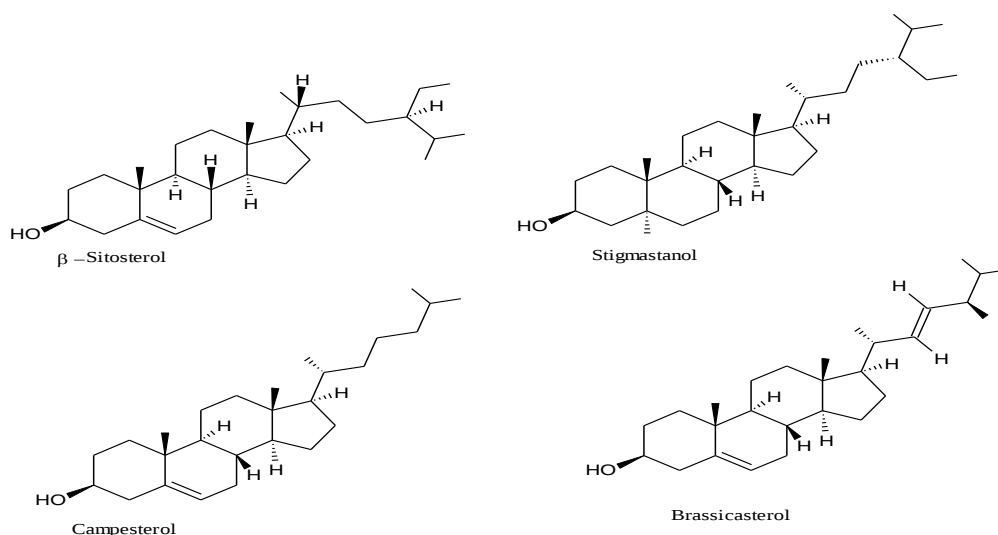


**Figure 6.** Chemical structure of some flavonoids

## 2.6. Phytosterols

Phytosterols are esters found in plants but they are similar to cholesterol in animals. They are characterized by tetracyclic cyclopenta- $\alpha$ -phenanthrene ring with a specific molecular configuration. Phytosterols are  $\beta$ -sitosterol,  $\beta$ -

sitosterol, daucosterol, brassicasterol, campestanol, campesterol, ergosterol, stigmasterol (**Figure 7**). They are found in whole grains, corn oil, wheat germ, rapeseed oil, canola oil, soybeans, green beans, olive oil, sweet potatoes, broccoli, cauliflower, cotton bush (*Asclepias curassavica*), *Grewia tiliaefolia* (GT), etc. The plant sterols and stanols have cholesterol-lowering effects in preventing primary and secondary cardiovascular diseases [36-37]. Several researches have been carried to show interesting pharmacological properties as anti-inflammatory, antioxidant, antidiabetic, chemo-preventive, and anti-atherosclerotic effects [38-39].



**Figure 7.** Chemical structure of some phytosterols

### 3. Pharmacological activities

Phytochemicals (lycopene, lutein, curcumin, resveratrol, perillyl alcohol, *D*-limonene, kaempferol, gingerol, and epigallocatechin-3-gallate etc.), are rich sources of biochemicals, pharmaceuticals and therapeutics agents with little or no toxicity at low doses and of health benefit for treating and preventing most of diseases such as COVID infection [18, 40], coronary heart diseases [41], stroke [6-7], neurodegenerative disorder [24], blood pressure [42-43], osteoporosis [44-46], cancers [47-51], chronic obstructive pulmonary diseases [52-54], diabetes [41, 55-57], diarrheal [58], respiratory problems [59], Human Immunodeficiency Virus (HIV) infection [13, 60-62], mental health problems [24, 63-67] etc.

Despite the health benefit of phytochemicals, some plants are toxic (neurotoxic, genotoxic, gastrotoxic, nephrotoxin, renal toxic, hepatotoxic, cytotoxic) even at the lowest doses. Examples are *Ricinus communis*, *Aesculus hippocastanum*, rhubarb, lupines, sanguinolenta, psorospermin, *Nicotina spp*, *Solanum malacoxylon*, *Phaseolus vulgaris*, *Solanum dulcamara*, *Datura stramonium* [61, 68-71], and some are antinutrients such as glycol-alkaloids and cyanogenic glycosides, lupins, phytohaemagglutinin.

### 4. Qualitative and quantitative analysis of secondary metabolites

Phytochemical analysis is the method of determining organic constituents of the medicinal plants through qualitative and quantitative analysis. The first step in phytochemical analysis is preliminary screening or rather qualitative

analysis before quantitative analysis. Qualitative analysis detects the presence or absence of a phytochemical in the extracts. It is indicated by color changes and formation of precipitate. Quantitative analysis provides information on the amount or the concentration in the plant extracts. This details information assists in drug discovery, safety assessment of herbal drugs, explanation of the medicinal potentials of plants and determination of the toxicity levels in plants [72].

#### 4.1. Techniques for identify and quantifying secondary metabolites

Modern analytical techniques such as spectrophotometry, chromatography have been used to identify, isolate, characterize and elucidate structural formulas of phytochemicals. The conventional method for identifying and quantifying secondary metabolites is spectrophotometry whereas the non-conventional methods is chromatography. Both techniques have been used to separate, identify and quantify series of structurally different SMs in plants include spectrophotometric [57, 73-75], GC-MS [56, 76], LC-MS [77], LC-MS/MS [56, 76, 78], Liquid Chromatography-High Resolution Electrospray Ionization Mass Spectrometry (LC- HRESIMS) [57], Nuclear Magnetic Resonance (NMR) [44, 79], Ultra-Fast Liquid Chromatography (UFLC-Q-TOF-MS/MS) [80-81], UFLC-triple-Q-TOF-MS/MS [82], Ultra-high performance Liquid Chromatography quadrupole- time of flight mass spectrometry [83], UHPLC-LTQ-Orbitrap [84], UHPLC/Q- Orbitrap-MS [85], HPLC-DAD [86], UHPLC-Q-TOF-MS [87], UHPLC-Q-TOF-MS/MS [88], HPLC-DAD-ESI-MS [89], UPLC-QTOF/MSE [90], UHPLC-MS/MS [91], HPLC-ESI-QTOF-MS [92], UPLC-QTOF-MS/MS [93], UHPLC-Q-TOF-MS/MS [94-95], UHPLC-ESI-MS/MS [96], UHPLC-ESI-Orbitrap-MS/MS [97], UPLC-IT-MS [98], UPLC-Q-TOF-MS [98], UPLC-PDA [45], HPLC-ESI-MS and MS<sup>2</sup> [99], reverse phase high performance liquid chromatography-diode array detector (RP-HPLC-DAD) [74], HPLC-HRMS [100].

#### 4.2. Identification of Secondary metabolites

##### 4.2.1. Flavonoids

4.2.1.1. Add FeCl<sub>3</sub> to the plant extract changes from yellow to purple, blackish-red and greenish-blue precipitate [101-103].

4.2.1.2. Shinoda Test: Alcoholic plant extract, magnesium ribbon or powder and conc. HCl turn pink to red color showed the presence of flavonoid [104-106]. The orange to red color indicate flavones, red to crimson indicate flavonoids, crimson to magenta indicate flavonones.

4.2.1.3. The plant extract and conc. HCl turn red color showed the presence of flavonoid [107].

4.2.1.4. Shibata Test: Alcoholic plant extract, metallic magnesium and conc. HCl turn orange to red color. The orange and red color indicate flavones and flavonols respectively [108-109].

4.2.1.5. Pew's Test: Aqueous plant extract, metallic zinc and conc. H<sub>2</sub>SO<sub>4</sub> turn red color showed the presence of flavonols [108].

4.2.1.6. Sodium hydroxide test: Aqueous plant extract and different concentration of sodium hydroxide solution has been used to produce a yellow color that turn colorless on addition of dilute hydrochloric acid confirm flavonoid [102, 104-106, 110-111], and yellow to red color showed the presence of tannin [100-101]. When it produces an emulsion is hydrolyzable tannins.

#### 4.2.2. Test for Tannin

4.2.2.1. Braymer's test: Add  $\text{FeCl}_3$  solution to the plant extract filtrate or crude to give dark green or bluish black precipitate in of tannin catechol or gallic tannin respectively [79, 102, 104-106, 108, 111-113].

4.2.2.2. Crude plant extract and acetic acid give red color showed the presence of tannin.

4.2.2.3. Lead acetate test: Diluted plant extract and lead acetate to give black-green colored precipitate formed revealed the presence of tannins [104]. The yellow precipitate indicates flavonoid [105, 111, 112].

4.2.2.4. Bromine water Test. A filtrate of alcoholic plant extract and bromine water to produce buff color precipitate an indication of condensed tannins meanwhile hydrolyzable tannins decolorize [101, 114].

#### 4.2.3. Test for cardiac glycosides

4.2.3.1. Keller-Killani Test: A crude plant extract filtrate, glacial  $\text{CH}_3\text{COOH}$ ,  $\text{FeCl}_3$  solution mix and conc.  $\text{H}_2\text{SO}_4$  to produce reddish-brown color which turns blue on standing, the acetic layer indicates cardiac glycosides of deoxy sugar moiety [102, 110].

4.2.3.2. Kedee's Test: This method determines cardenolides. The dried plant extract, methanolic KOH and alcoholic 3,5-dinitrobenzoic acid heated to produce purple-violet colour that faded gradually through reddish-brown to brownish-yellow with whitish precipitation indicates the presence of lactone ring [103, 105].

4.2.3.3. Legal's test: The crude plant extract, pyridine, sodium nitroprusside and NaOH solution to produce pink to red color that change to colorless [105].

4.2.3.4. Test for Anthraquinones (Borntrager's test): The test involves alkalization of plant extract with an alkaline usually  $\text{NH}_3$  or KOH to produce the corresponding phenolate ion. crude or filtrate of plant extract with 10% ammonia solution shaken vigorously for 30 s to produce pink [108], or red color [101, 109], in aqueous layer indicates the presence of free anthraquinones. In the case of glycoside, filtrate of hydrolysable plant extract should be used to identify the anthraquinones.

4.2.3.5. Modified Borntrager's Test: Plant extract,  $\text{FeCl}_3$  solution, boil and cool, add benzene, and add  $\text{NH}_3$  to the organic layer. mix, to produce a rose pink to blood red colored solution [115].

#### 4.2.4. Test for Phytosterols

4.2.4.1. Liebermann-Bouchard's or acetic anhydride Test: The aqueous crude extracts,  $(\text{CH}_3\text{CO})_2\text{O}$  heating and cooling and then with conc.  $\text{H}_2\text{SO}_4$  mixed gives a color changes from violet to blue/bluish green confirm steroid core [79, 102, 104-105, 107, 112], but when pink or reddish color appears it indicate triterpene [79, 105, 116].

4.2.4.2. The aqueous crude extracts, chloroform with conc.  $\text{H}_2\text{SO}_4$  mixed with pink ring or red at the bottom chloroform layer [114].

4.2.4.3. Salkowski's Test: The filtrate of aqueous crude extracts, chloroform with conc.  $\text{H}_2\text{SO}_4$  mixed with bluish red to purple color in chloroform layer indicates the presence of steroidal aglycone part of the glycoside and red brown to dark brown at the bottom layer indicates triterpenoids [111].

#### 4.2.4. Test for Terpenoids

4.2.5.1. Add  $\text{CCl}_3\text{COOH}$  to crude plant extract with red precipitate an indicative of the presence of triterpenoids [110].

4.2.5.2. The crude extracts, chloroform, conc.  $\text{H}_2\text{SO}_4$  mixed with reddish brown ring indicates the presence of triterpene [102, 107, 112].

4.2.5.3. Aqueous crude plant extract and copper acetate solution heated to emerald green colour indicate diterpenes [117].

#### 4.2.6. Test for alkaloids

4.2.6.1. Dragendorff reagent, (0.85 g  $\text{Bi}(\text{NO}_3)_3 \cdot 5\text{H}_2\text{O}$  in 40 mL distilled water, 10 mL  $\text{CH}_3\text{COOH}$ , 8 g KI in 20 mL distilled water), is added to the filtrate of acidified extract to form an orange red precipitate [104, 109-110, 118].

4.2.6.2. Mayer's reagent, (1.36 g  $\text{HgCl}_2$  in 60 mL distilled water, 5 g KI in 10 mL distilled water, 30 mL  $\text{H}_2\text{O}_2$ , is added to the filtrate of acidified extract to form a yellowish or white precipitate [105, 110, 112].

4.2.6.3. Wagner's reagent, (1.27 g  $\text{I}_2$ , 2 g KI in 100 mL distilled water), is added to the filtrate of acidified extract to form an orange [79], or reddish-brown precipitate [111, 119].

4.2.6.4 Picric acid test: The filtrate of plant extract and picric acid solution produce yellow color precipitate indicate an alkaloid [102, 110, 120].

4.2.6.5 Bouchardat's test: Acidified filtrate of the extract and Boucharat's reagent (dilute iodine solution) to produce reddish brown colour indicate an alkaloid [119].

4.2.6.6 Tannic acid test: Acidified crude plant extract and tannic acid produce yellow color precipitate indicate an alkaloid [120].

#### 4.2.7. Test for Saponin

4.2.7.1. Foam test: Add or no distilled water to crude plant extract shake vigorously and the soapy appearance [102], foam [79, 105], or honeycomb froth [108, 111-112] showed the presence of saponins glycosides.

4.2.7.2. Frothing test: Add distilled water to aqueous plant extract shake vigorously and heat with the appearance of frothing showed the presence of saponins [107, 115].

4.2.7.3. Olive oil test: Aqueous crude plant extract mixed well with olive oil shake vigorously and the appearance of foam showed the presence of saponins [102, 114].

4.2.7.4.  $\text{Na}_2\text{CO}_3$  test: Add aqueous plant extract to  $\text{Na}_2\text{CO}_3$  solution and distilled water shake vigorously with the appearance of foam show the presence of saponins [110].

### 4.3. Quantification of Secondary metabolites

#### 4.3.1. Test for total phenols

##### 4.3.1.1. Test for total phenols using Folin-Ciocalteu reagent (FCR) method

This method determines the total phenolic compound in plant crude extract where the Folin-Ciocalteu reagent (2.5 g  $\text{Na}_2\text{MoO}_4 \cdot 2\text{H}_2\text{O}$ , 10 g  $\text{Na}_2\text{WO}_4 \cdot 2\text{H}_2\text{O}$  in 70 mL distilled  $\text{H}_2\text{O}$ , then 5 mL 85%  $\text{H}_3\text{PO}_4$  and 10 mL  $\text{HCl}$ ), a mixture of phosphomolybdate and phosphotungstate reagents ( $\text{H}_3\text{PMo}_{12}\text{O}_{40}$  and  $\text{H}_3\text{PW}_{12}\text{O}_{40}$ ) and aluminum chloride ( $\text{AlCl}_3$ ), in the presence of phenolics and carried out in alkaline medium  $\text{Na}_2\text{CO}_3$  to produce a blue color molybdenum tungsten ( $\text{Mo}_8\text{O}_{23}$  and  $\text{W}_8\text{O}_{23}$ ) that can be measured in the range of 720-765 nm in visible spectrophotometer against the standard gallic acid. Also, some other standards including quercetin, ferulic acid and caffeic acids have been reported [57, 74, 121]. In this method different ratio of diluted crude extract and Folin-Ciocalteu reagent have been used. Likewise, the incubation has been carried out at room temperature or heated [57, 73-75].

#### 4.3.2. Test for total flavonoids

##### 4.3.2.1. Test for total flavonoids using aluminum chloride colorimetric assay

Total flavonoid content might involve the nitration of aromatic ring of a catechol with its three or four positions not sterically hinder. The reaction form aluminum complexes with carbonyl and hydroxyl groups of flavones and flavonols respectively. Equivalent amount of aqueous crude extract and diluted with or without  $\text{NaNO}_2$  solution, will be mixed at room temperature, then add to  $\text{AlCl}_3$  solution mix and neutralized with or without  $\text{NaOH}$  solution and read the aluminum complex absorbance in the visible spectrophotometer against either catechin, rutin, or quercetin standard. [57, 74-75, 121-122].

##### 4.3.2.2. Test for total flavonoids by gravimetric

Flavonoid has been quantified when plant extract filtrate is evaporated to dryness to obtain a constant weigh [101-102].

#### 4.3.3. Total tannin content

Plant extract filtrate,  $\text{FeCl}_3$ , and potassium ferricyanide ( $\text{K}_3[\text{Fe}(\text{CN})_6]$ ) in  $\text{HCl}$  are mixed to show a blue color or read the absorbance against gallic acid [101-102].

#### 4.3.4. Total alkaloid

##### 4.3.4.1. Total alkaloid using colorimetric

The plant aqueous extract is dissolved in dimethyl sulphoxide (DMSO), diluted with HCl, the filtrate is mixed with bromocresol green solution and phosphate buffer then measure in the range of visible spectrophotometer against the standard atropine [123].

##### 4.3.4.2. Total alkaloid by gravimetric

Acetic acid in ethanol is added to the crude extract, incubate, concentrate and basified with conc.  $\text{NH}_4\text{OH}$  dropwise to get a precipitate, washed with dilute  $\text{NH}_4\text{OH}$ . The filtrate is dried and weighed [102].

#### 4.3.5. Total saponins

Diethylether is added to concentrated plant extract filtrate, the aqueous layer is washed with alcohol (*n*-butanol) and NaCl solution, then evaporate to a constant weigh [102].

#### 4.3.6. Total terpenoid

Extract the filtrate of plant extract with petroleum ether [102].

#### 4.3.7. Total cardiac glycosides using colorimetric

Baljet reagent (95 mL of picric acid at 1% + 5 mL 10% NaOH) is added to dilute plant extract, incubate to produce a yellow to orange measure in the range of visible spectrophotometer against the standard peruvoside [73, 75].

#### Conclusion

This review has revealed the recent pharmacological activities and has detailed how the plant secondary metabolites can be identified and quantified. However, this will help in the level of new drug discovery and development from the old ones and new secondary metabolites plant based that are safer and effective therapeutic agents in order to improve the quality of life which will boost the economic and reduces the uses and dependent on the synthetic drug. Also, increases life expectancy and reduces the onset of age related diseases.

#### References

1. Huang, Y.; Xie, F.-J.; Cao, X.; Li, M.-Y. Research progress in biosynthesis and regulation of plant terpenoids. *Biotechnol & Biotechnol Equip.* **2021**, 35 (1), 1799-1808. Doi: 10.1080/13102818.2021.2020162.
2. Jan, R.; Asaf, S.; Numan, M.; Lubna; Kim, K.-M. Plant Secondary Metabolite Biosynthesis and Transcriptional Regulation in Response to Biotic and Abiotic Stress Conditions. *Agronomy*, **2021**, 11 (5), 968. Doi: 10.3390/agronomy1105096.

3. Dey, P.; Kundu, A.; Kumar, A.; Gupta, M.; Lee, B.M.; Bhakta, T.; Dash, S.; Kim, H.S. Analysis of alkaloids (indole alkaloids, isoquinoline alkaloids, tropane alkaloids). In: *Recent Advances in Natural Products Analysis*. Elsevier Inc 2020, 15, 505–567. Doi: 10.1016/B978-0-12-816455-6.00015-9.
4. Heinrich, M.; Mah, J.; Amirkia, V. Alkaloids Used as Medicines: Structural Phytochemistry Meets Biodiversity—An Update and Forward Look. *Molecules*. **2021**, 26 (7), 1836. Doi: 10.3390/molecules26071836.
5. Ogugofor, M.O.; Njoku, U.O.; Njoku, O.U.; Batiha, G.E.S. Phytochemical analysis and thrombolytic profiling of *Costus afer* stem fractions. *Futur J Pharm Sci*. **2022**, 8, 4. Doi:10.1186/s43094-021-00392-3.
6. Wang, C.; Cheng, Y.; Zhang, Y.; Jin, H.; Zuo, Z.; Wang, A.; Huang, J.; Jiang, J.; Kong, W. Berberine and Its Main Berberubine Inhibit Platelet Activation Through Suppressing the Class IPI3K $\beta$ /Rasa3/Rap1 Pathway. *Front Pharmacol*. **2021**, 12, 734603. Doi: 10.3389/fphar.2021.734603.
7. Zeng, P.; Yi, Y.; Su, H.-F.; Ye, C.-Y.; Sun, Y.-W.; Zhou, X.-W.; Lu, Y.; Shi, A.; Tian, Q. Key Phytochemicals and Biological Functions of Chuanxiong Rhizoma Against Ischemic Stroke: A Network Pharmacology and Experimental Assessment. *Front Pharmacol*. **2021**, 12, 758049. Doi: 10.3389/fphar.2021.758049.
8. Zhu, T.; Wang, L.; Feng, Y.; Sun, G.; Sun, X. Classical Active Ingredients and Extracts of Chinese Herbal Medicines: Pharmacokinetics, Pharmacodynamics, and Molecular Mechanisms for Ischemic Stroke. *Oxid. Med. Cel. Longev*. **2021**, 2021, 8868941. Doi:10.1155/2021/8868941.
9. Wang, T.M.B.; Jiang, X.M.B.; Ruan, Y.M.B.; Li, M.M.; Chu, L. The mechanism of action of the combination of *Astragalus membranaceus* and *Ligusticum chuanxiong* in the treatment of ischemic stroke based on network pharmacology and molecular docking. *Medicine*. **2022**, 101 (28), e29593. Doi:10.1097/MD.00000000000029593.
10. Ovonramwen, O.B.; Owolabi, B.J.; Falodun, A. Synthesis and Antimicrobial Activities of Some New sulfonyl Phenoxides. *Tanz J Sci*. **2021**, 47 (2), 745-753. Doi: 10.4314/tjs.v47i2.30.
11. Gueboudji, Z.; Addad, D.; Kadi, K.; Nagaz, K.; Secrafi, M.; Yahya, L.B.; Lachehib, B.; Abdelmalek, A. Biological activities and phenolic compounds of olive oil mill wastewater from Abani, endemic Algerian variety. *Sci Rep*. **2022**, 12, 6042. Doi: 10.1038/s41598-022-10052-y.
12. Tanwar, A.K.; Dhiman, N.; Kumar, A.; Jaitak, V. Engagement of phytoestrogens in breast cancer suppression: Structural classification and mechanistic approach. *Eur J Med Chem*. **2021**, 213, 5, Doi: 10.1016/j.ejmech.2020.113037.
13. Wu, D.; Jin, L.; Huang, X.; Deng, H.; Shen, Q.K.; Quan, Z.S.; Zhang, C.; Guo, H.Y. Arctigenin: pharmacology, total synthesis, and progress in structure modification. *J Enzyme Inhib Med Chem*. **2022**, 37 (1), 2452–2477. Doi: 10.1080/14756366.2022.2115035.
14. Hejazi, J.; Ghanavati, M.; Hejazi, E.; Poustchi, H.; Sepanlou, S.G.; Khoshnia, M.; Gharavi, A.; Sohrabpour, A.A.; Sotoudeh, M.; Dawsey, S.M.; Boffetta, P.; Abnet, C.C.; Kamangar, F.; Etemadi, A.; Pourshams, A.; Malekshah, A.F.; Brennan, P.; Malekzadeh, R.; Hekmatdoost, A. Habitual dietary intake

- of flavonoids and all-cause and cause-specific mortality: Golestan cohort study. *Nutr J.* **2020**, 19, 108. Doi: 10.1186/s12937-020-00627-8.
15. Zhang, X.; Molsberry, S.A.; Yeh, T.-S.; Cassidy, A.; Schwarzschild, M. A.; Ascherio, A.; Gao, X. Intake of Flavonoids and Flavonoid-Rich Foods and Mortality Risk Among Individuals With Parkinson Disease: A Prospective Cohort Study. *Neurology.* **2022**, 98 (10), Doi: 10.1212/WNL.00000000000013275.
  16. Jembrek, M.J.; Oršolić, N.; Mandić, L.; Sadžak, A.; Šegota, S. Anti-Oxidative, Anti-Inflammatory and Anti-Apoptotic Effects of Flavonols: Targeting Nrf2, NF-κB and p53 Pathways in Neurodegeneration. *Antioxidants* (Basel)., **2021**, 10 (10), 1628. Doi: 10.3390/antiox10101628.
  17. Ghosh, R.; Chakraborty, A.; Biswas, A.; Chowdhuri, S. Identification of polyphenols from *Broussonetia papyrifera* as SARS CoV-2 main protease inhibitors using in silico docking and molecular dynamics simulation approaches. *J Biomol Struct Dyn.* **2020**, 0, 1–14. Doi: 10.1080/07391102.2020.1802347.
  18. Sharanya, C.S.; Sabu, A.; Haridas, M. Potent phytochemicals against COVID-19 infection from phyto-materials used as antivirals in complementary medicines: a review. *Futur J Pharm Sci.* **2021**, 7 (1), 113. Doi:10.1186/s43094-021-00259-7.
  19. Yang, C.C.; Wu, C.J.; Chien, C.Y.; Chien, C.T. Green Tea Polyphenol Catechins Inhibit Coronavirus Replication and Potentiate the Adaptive Immunity and Autophagy-Dependent Protective Mechanism to Improve Acute Lung Injury in Mice. *Antioxidants.* **2021**, 10 (6), 928; Doi: 10.3390/antiox10060928.
  20. Pandey, P.; Rane, J.S.; Chatterjee, A.; Kumar, A.; Khan, R.; Prakash, A.; Ray, S. Targeting SARS-CoV-2 spike protein of COVID-19 with naturally occurring phytochemicals: an in-silico study for drug development. *J Biomol Struct Dyn.* **2020**, 0, 1–11. Doi:10.1080/07391102.2020.1796811.
  21. Tomko, A.M.; Whynot, E.G.; Dupré, D.J. Anti-cancer properties of cannflavin A and potential synergistic effects with gemcitabine, cisplatin, and cannabinoids in bladder cancer. *J Cannabis Res.* **2022**, 4, 41. Doi: 10.1186/s42238-022-00151-y.
  22. Moreau, M.; Ibeh, U.; Decosmo, K.; Bih, N.; Yasmin-Karim, S.; Toyang, N.; Lowe, H.; Ngwa, W. Flavonoid derivative of cannabis demonstrates therapeutic potential in preclinical models of metastatic pancreatic cancer. *Front Oncol.* **2019**, 9, 660. Doi: 10.3389/fonc.2019.00660.
  23. Bautista, J.L.; Yu, S.; Tian, L. Flavonoids in *Cannabis sativa*: Biosynthesis, Bioactivities, and Biotechnology. *ACS Omega*, **2021**, 6 (8), 5119–5123. Doi.org/10.1021/acsomega.1c00318.
  24. Sahebnaasagh, A.; Eghbali, S.; Saghafi, F.; Sureda, A.; Avan, R. Neurohormetic phytochemicals in the pathogenesis of neurodegenerative diseases. *Immun Ageing.* **2022**, 19, 36. Doi:10.1186/s12979-022-00292-x.
  25. Kaczmarek, B. Tannic acid with Antiviral and antibacterial activity as A Promising component of Biomaterials-A Minireview. *Materials* (Basel). **2020**, 13 (14), 3224. Doi: 10.3390/ma13143224.
  26. Tong, Z.; He, W.; Fan, X.; Guo, A. Biological Function of Plant Tannin and Its Application in Animal Health. *Front Vet Sci.* **2022**, 8, 803657. Doi: 10.3389/fvets.2021.80365732.

27. Zeng, X.; Jiang, W.; Du, Z.; Kokini, J.L. Encapsulation of tannins and tannin rich plant extracts by complex coacervation to improve their physiochemical properties and biological activities: A review. *Crit Rev Food Sci Nutr.* **2022**, *13*, 1-14. Doi:10.1080/10408398.2022.2075313.
28. Proshkina, E.; Plyusnin, S.; Babak, T.; Lashmanova, E.; Maganova, F.; Koval, L.; Platonova, E.; Shaposhnikov, M.; Moskalev, A. Terpenoids as Potential Geroprotectors. *Antioxidants.* **2020**, *9* (6), 529. Doi: 10.3390/antiox9060529.
29. Prado-Audelo, M.L.D.; Cortés, H.; Caballero-Florán, I.H.; González-Torres, M.; Escutia-Guadarrama, L.; Bernal-Chávez, S.A.; Giraldo-Gomez, D.M.; Magaña, J.J.; Leyva-Gómez, G. Therapeutic Applications of Terpenes on Inflammatory Diseases. *Front Pharmacol.* **2021**, *12*, 704197. Doi: 10.3389/fphar.2021.704197.
30. Salehi, B.; Upadhyay, S.; Orhan, I.E.; Jugran, A.K.; Jayaweera, S.L.D.; Dias, D.A.; Sharopov, F.; Taheri, Y.; Martins, N.; Baghalpour, N.; Cho, W.C.; Sharifi-Rad, J. Therapeutic Potential of  $\alpha$ - and  $\beta$ -Pinene: A miracle gift of Nature. *Biomolecules.* **2019**, *9* (11), 738. doi: 10.3390/biom9110738.
31. Muilu-Makela, R.; Aapola, U.; Tienaho, J.; Uusitalo, H.; Sarjala, T. Antibacterial and Oxidative Stress-Protective Effects of Five Monoterpenes from Softwood. *Molecules.* **2022**, *27* (12), 3891. Doi: 10.3390/molecules27123891.
32. Dewanjee, S.; Bhattacharjee, N.; Chakraborty, P.; Bhattacharjee, S. Carotenoids as Antioxidants. In: Zia-Ul-Haq, M., Dewanjee, S., Riaz, M. (eds) *Carotenoids: Structure and Function in the Human Body*. Springer, Cham. 2021. Doi: 10.1007/978-3-030-46459-2\_12.
33. Xia, M.; Liu, D.; Liu, Y.; Liu, H. The Therapeutic Effect of Artemisinin and Its Derivatives in Kidney Disease. *Front Pharmacol.* **2020**, *11*, 380. Doi: 10.3389/fphar.2020.00380.
34. Gusain, P.; Uniyal, D.P.; Joga, R. Conservation and sustainable use of medicinal plants. Preparation of Phytopharmaceuticals for the Management of Disorders. *The Development of Nutraceuticals and Traditional Medicine*. Elsevier. 2021. 23, 409-427.
35. Kellerman, R.D.; Rakel, D. *Malaria*. Conn's Current Therapy. Elsevier Health Sciences. 2021, 1536.
36. Makhmudova, U.; Schulze, P.C.; Lutjohann, D.; Weingartner, O. Phytosterols and Cardiovascular Disease. *Curr Atheroscler Rep.* **2021**, *23*, 68. Doi: 10.1007/s11883-021-00964-x.
37. Poli, A.; Marangoni, F.; Corsini, A.; Manzato, E.; Marrocco, W.; Martini, D.; Medea, G.; Visioli, F. Phytosterols, Cholesterol Control, and Cardiovascular Disease. *Nutrients.* **2021**, *13* (8), 2810. Doi: 10.3390/nu130828810.
38. Salehi, B.; Quispe, C.; Sharifi-Rad, J.; Cruz-Martins, N.; Nigam, M.; Mishra, A.P.; Konovalov, D.A.; Orobinskaya, V.; Abu-Reidah, I.M.; Zam, W.; Sharopov, F.; Venneri, T.; Capasso, R.; Kukula-Koch, W.; Wawruszak, A.; Koch, W. Phytosterols: From Preclinical Evidence to Potential Clinical Applications. *Front Pharmacol.* **2020**, *11*, 599959. Doi: 10.3389/fphar.2020.599959.
39. Ullah, H.; Khan, H. Chapter 16 - Epigenetic drug development for autoimmune and inflammatory diseases. In: *Histone Modifications in Therapy*. Translational Epigenetic. 2020, 20, 395-413. Doi: 10.1016/B978-0-12-816422-8.00017-9.

40. Bhattacharya, S.; Paul, S.M.N. Efficacy of phytochemicals as immunomodulators in managing COVID-19: a comprehensive view. *Virus Dis.* **2021**, *32*, 435–445. Doi: 10.1007/s13337-021-00706-2.
41. Aghdam, S.D.; Siassi, F.; Esfahani, E.N.; Qorbani, M.; Rajab, A.; Sajjadpour, Z.; Bashiri, A.; Aghayan, M.; Sotoudeh, G. Dietary phytochemical index associated with cardiovascular risk factor in patients with type 1 diabetes mellitus. *BMC Cardiovasc Disord.* **2021**, *21*, 293. Doi:10.1186/s12872-021-02106-2.
42. Verma, T.; Sinha, M.; Bansal, N.; Yadav, S.R.; Shah, K.; Chauhan, N.S. Plants Used as Antihypertensive. *Nat Prod Bioprospect.* **2021**, *11* (2), 155–184. Doi:10.1007/s13659-020-00281-x.
43. Jasemi, S.V.; Khazaei, H.; Aneva, I.Y.; Farzaei, M.H.; Echeverría, J. Medicinal Plants and Phytochemicals for the Treatment of Pulmonary Hypertension. *Front Pharmacol.* **2020**, *11*, 145. Doi: 10.3389/fphar.2020.00145.
44. Ibrahim, H.-I.M.; Darrag, H.M.; Alhajhoj, M.R.; Khalil, H.E. Biomolecule from *Trigonella stellata* from Saudi Flora to Suppress Osteoporosis via Osteostromal Regulations. *Plants.* **2020**, *9*, 1610. Doi: 10.3390/plants9111610.
45. Kulkarni, C.; Sharma, S.; Bora, P.S.; Verma, S.; Rajput, S.; Porwal, K.; Rath, S.K.; Gayen, J.R.; Sharma, U.; Chattopadhyay, N. A novel extraction method enhanced the osteogenic and anti-osteoporosis effect of tea extract without any hepatotoxicity in ovariectomized rats. *Front Endocrinol.* **2022**, *13*, 951800. doi: 10.3389/fendo.2022.951800.
46. Rahim, R.B.; Jayusman, P.A.; Lim, V.; Ahmad, N.H.; Hamid, Z.A.A.; Mohamed, S.; Muhammad, N.; Ahmad, F.; Mokhtar, N.; Mohamed, N.; Shuid, A.N.; Mohamed, I.N. Phytochemical Analysis, Antioxidant and Bone Anabolic Effects of *Blainvillea acmella* (L.) Philipson. *Front Pharmacol.* **2022**, *12*, 796509. Doi: 10.3389/fphar.2021.796509.
47. Lai, C.-S.; Ho, C.-T.; Pan, M.-H. The Cancer Chemopreventive and Therapeutic Potential of Tetrahydrocurcumin. *Biomolecules.* **2020**, *10*:831. Doi: 10.3390/biom10060831.
48. Hao, S.; Wang, J.; Li, S.; Shang, F.; Qin, Y.; Wu, T.; Bao, X.; Cao, Q.; Wang, C.; Sun, B. Preparation of Gardenia red pigment and its antineoplastic activity in multiple tumor cells. *Food Biosci.* **2020**, *35*, 100582. Doi: 10.1016/j.fbio.2020.100582.
49. Liang, R.; Wu, J.; Lin, R.; Ran, L.; Shu, B.; Deng, H. Molecular Mechanisms of Gynostemma pentaphyllum in Prevention and Treatment of Non-Small-Cell Lung Cancer. *Evid Based Compl Alternat Med.* **2022**, 9938936. Doi: 10.1155/2022/9938936.
50. Ru, R.; Guo, Y.; Mao, J.; Yu, Z.; Huang, W.; Cao, X.; Hu, H.; Meng, M.; Yuan, L. Cancer Cell Inhibiting Sea Cucumber (*Holothuria leucospilota*) Protein as a Novel Anti-Cancer Drug. *Nutrients.* **2022**, *14* (4), 786. Doi: 10.3390/nu1404786.
51. Prasedya, E.S.; Ardiana, N.; Padmi, H.; Ilhami, B.T.K.; Martyasari, N.W.R.; Sunarwidhi, A.L.; Nikmatullah, A.; Widyastuti, S.; Sunarpi, H.; Frediansyah, A. The Antiproliferative and Apoptosis-Inducing Effects of the Red Macroalgae *Gelidium latifolium* Extract against Melanoma Cells. *Molecules.* **2021**, *26* (21), 6568. Doi: 10.3390/molecules26216568.

52. Devkota, H.P.; Adhikari-Devkota, A.; Paudel K.R.; Panth, N.; Gupta, G.; Chellappan, D.K.; Hansbro, P.M.; Dua, K. Phytochemicals and their Nanoformulations Targeted for Pulmonary Diseases. In: Chellappan, D.K., Pabreja, K., Faiyazuddin, M. (eds) *Advanced Drug Delivery Strategies for Targeting Chronic Inflammatory Lung Diseases*. Springer, Singapore. 2022, 95-106. Doi: 10.1007/978-981-16-4392-7\_5.
53. Jasemi, S.V.; Khazaei, H.; Momtaz, S.; Farzaei, M.H.; Echeverría, J. Natural products in the treatment of pulmonary emphysema: Therapeutic effects and mechanisms of action. *Phytomedicine*. **2022**, 99, 153988. Doi: 10.1016/j.phymed.2022.153988.
54. Mitra, S.; Anand, U.; Ghorai, M.; Vellingiri, B.; Jha, N.K.; Behl, T.; Kumar, M.; Radha, Shekhawat, M.S., Proćków, J.; Dey, A. Unravelling the Therapeutic Potential of Botanicals Against Chronic Obstructive Pulmonary Disease (COPD): Molecular Insights and Future Perspectives. *Front. Pharmacol.* **2022**, 13:824132. Doi: 10.3389/fphar.2022.824132.
55. El Moussaoui, A.; Mechchate, H.; Bourhia, M.; Es-safi, I.; Salamatullah, A.M.; Alkaltham, M.S.; Alyahya, H.K.; Bousta, D.; Bari, A. Glycemic Control Potential of Chemically Characterized Extract from *Withania frutescens* L. Roots in Severe Diabetes-Induced Mice. *Appl Sci*. **2021**, 11, 3998. Doi: 10.3390/app11093998.
56. Saravanakumar, K.; Park, S.J.; Sathiyaseelan, A.; Kim, K.N.; Cho, S.H.; Mariadoss, A.V.A.; Wang, M.H. Metabolite profiling of methanolic extract of *gardenia jaminoides* by LC-MS/MS and GC-MS and its anti-diabetic, and anti-oxidant activities. *Pharmaceuticals*. **2021**, 14 (2), 102. Doi: 10.3390/ph14020102.
57. Elwekeel, A.; Amir, D.E.; Mohamed, E.I.A.; Amin, E.; Hassan, M.H.A.; Zaki, M.A. Characterization of Possible  $\alpha$ -Glucosidase Inhibitors from *Trigonella stellata* Extract Using LC-MS and In Silico Molecular Docking. *Plants* (Basel). **2022**, 11 (2), 208. Doi: 10.3390/plants11020208.
58. Njoku, U.O.; Ogugofor, M.O. Antidiarrheal effect of methanol leaf fraction of *Cola hispida* Brenan and Keay in rats. *Trop J Pharm Res*. **2021**, 20 (7), 1455–1462. Doi:10.4314/tjpr.v20i7.19.
59. Allam, V.S.R.R.; Chellappan, D.K.; Jha, N.K.; Shastri, M.D.; Gupta, G.; Shukla, S.D.; Singh, S.K.; Sunkara, K.; Chitranshi, N.; Gupta, V.; Wich, P.R.; MacLoughlin, R.; Oliver, B.G.G.; Wernersson, S.; Pejler, G.; Dua, K. Treatment of chronic airway diseases using nutraceuticals: Mechanistic insight. *Crit Rev Food Sci Nutr*. **2022**, 62 (27), 7576-7590. Doi: 1080/10408398.2021.1915744.
60. Fonteh, P.; Chingwaru, W.; Meyer, D.; Kapewangolo, P. Invitro Assays for Determining Anti-HIV Potential of Phytochemicals. In: Kumar S, Egbuna C. (eds) *Phytochemistry: An in-silico and in-vitro Update*. Springer, Singapore. 2019, 299-313. Doi: 10.1007/978-981-13-6920-9\_16.
61. Anywar, G.; Kakudidi, E.; Byamukama, R.; Mukonzo, J.; Schubert, A.; Oryem-Origa, H.; Jassoy, C. A Review of the toxicity and Phytochemistry of Medicinal Plant Species Used by Herbalists in Treating People Living With HIV/AIDS in Uganda. *Front. Pharmacol.* **2021**, 12, 615147. Doi: 10.3389/fphar.2021.615147.

62. . Sillapachaiyaporn, C.; Sunita, N.; Ung, A.T.; Chuchawankul, S. Anti-HIV-1 protease activity of the crude extracts and isolated compounds from *Auricularia polytricha*. *BMC Complement Altern Med*. **2019**, 19, 351. Doi: 10.1186/s12906-019-2766-3.
63. Chae, H.K.; Kim, W.; Kim, S.K. Phytochemicals of Cinnamomi Cortex: Cinnamic Acid, but not Cinnamaldehyde, Attenuates Oxaliplatin-Induced Cold and Mechanical Hypersensitivity in Rats. *Nutrients*. **2019**, 11(2), 432; Doi: 10.3390/nu11020432.
64. Phan, H.T.T.; Samarat, K.; Takamura, Y.; Azo-Oussou, A.F.; Nakazono, Y.; Vestergaard, M.C. Polyphenols Modulate Alzheimer's Amyloid Beta Aggregation in a Structure-Dependent Manner. *Nutrients*. **2019**, 11 (4), 756. Doi: 10.3390/nu11040756.
65. Brunetti, G.; Rosa, G.D.; Scuto, M.; Leri, M.; Stefani, M.; Schmitz-Linneweber, C.; Calabrese, V.; Saul, N. Healthspan maintenance and prevention of Parkinson's-like phenotypes with Hydroxytyrosol and Oleuropein Aglycone in *C. elegans*. *Int J Mol Sci*. **2020**, 21 (7), 2588. Doi: 10.3390/ijms21072588.
66. Elmazoglu, Z.; Yar Saglam, A.S.; Sonmez, C.; Karasu, C. Luteolin protects microglia against rotenone-induced toxicity in a hormetic manner through targeting oxidative stress response, genes associated with Parkinson's disease and inflammatory pathways. *Drug Chem Toxicol*. **2020**, 43 (1), 96–103. Doi: 10.1080/01480545.2018.1504961.
67. Esselun, C.; Theyssen, E.; Eckert, G.P. Effects of Urolithin a on mitochondrial parameters in a cellular model of early Alzheimer disease. *Int J Mol Sci*. **2021**, 22 (15), 8333. Doi: 10.3390/ijms22158333.
68. Salem, M.A.; Serag, A.; El-Seedi, H.R.; Hamdan, D.I.; Ezzat, S.M.; Zayed, A. 23-Identification and analysis of toxic phytochemicals. In: *Phytochemistry, the military and health*. Elsevier Inc. 2021, 443-479. Doi:10.1016/B978-0-12-821556-2.00015-3.
69. Pradhan, B.; Kim, H.; Abassi, S.; Ki, J.S. Toxic Effects and Tumor Promotion Activity of Marine Phytoplankton Toxins: A Review. *Toxins (Basel)*. **2022**, 14 (6), 397. Doi: 10.3390/toxins14060397.
70. Rasool, F.; Nizamani, Z.A.; Ahmad, K.S.; Parveen, F.; Khan, S.A.; Sabir, N. Phytoxicological study of selected poisonous plants from Azad Jammu & Kashmir. *PLoS ONE*. **2022**, 17 (5), e0263605. Doi: 10.1371/journal.pone.0263605.
71. Welch, K.D.; Green, B.T.; Gardner, D.R.; Clinton, A.; Cook, D. Toxic plants. In book: *Reproductive and Developmental Toxicology*. Elsevier Inc. 2022, 46, 933-953. Doi: 10.1016/B978-0-323-89773-0.00046-1.
72. Egbuna, C.; Ifemeje, J.C.; Maduako, M.C.; Tijani, H.; Udedi, S.C.; Nwaka, A.C.; Ifemeje, M.O. Phytochemical Test Methods: Qualitative, Quantitative and Proximate Analysis. In: *Phytochemistry, Volume 1: Fundamentals, Modern Techniques, and Applications*. Apple Academic Press, Taylor & Francis, USA. 2019, 15, 381-425.
73. Mendoza, D.; Arias, J.P.; Cuaspu, O.; Arias, M. Phytochemical screening of callus and cell suspensions cultures of *Trevelia peruviana*. *Braz arch boil Technol*. **2020**, 63 (3), 1-14. Doi: 10.1590/1678-4324-2020180735.

74. Yu, M.; Gouvvinhas, I.; Rocha, J.; Barros, A.I.R.N.A. Phytochemical and antioxidant analysis of medicinal and food plants towards bioactive food and pharmaceutical resources. *Sci Rep.* **2021**, *11*, 10041. Doi: 10.1038/s41598-021-89437-4.
75. Galal, T.M.; Al-Yasi, H.M.; Fawzy, M.A.; Abdelkader, T.G.; Hamza, R.Z.; Eid, E.M.; Ali, E.F. Evaluation of the Phytochemical and Pharmacological Potential of Taif's Rose (*Rosa damascena* Mill var. *trigintipetala*) for Possible Recycling of Pruning Wastes. *Life (Basel)*. **2022**, *12* (2), 273. Doi: 10.3390/life12020273.
76. El Sayed, A.M.; Basam, S.M.; El-Naggar, E.-M.B.A.; Marzouk, H.S.; El-Hawary, S. LC–MS/MS and GC–MS profiling as well as the antimicrobial effect of leaves of selected Yucca species introduced to Egypt. *Sci Rep.* **2020**, *10*, 1–15. Doi: 10.1038/s41598-020-74440-y.
77. Ni, Y.; Li, L.; Zhang, W.Y.; Lu, D.; Zang, C.; Zhang, D.; Yu, Y.; Yao, X. Discovery and LC-MS Characterization of new Crocins in *Gardeniae fructus* and their Neuroprotective Potential. *J Agric Food Chem.* **2017**, *65* (14), 2936–2946. Doi: 10.1021/acs.jafc.6b03866.
78. Banerjee, S.; Bhattacharjee, P.; Kar, A.; Mukherjee, P.K. LC–MS/MS analysis and network pharmacology of *Trigonella foenum-graecum*—A plant from Ayurveda against hyperlipidemia and hyperglycemia with combination synergy. *Phytomedicine.* **2019**, *60*, 152944. doi: 10.1016/j.phymed.2019.15294.
79. Nguyen, D.H.; Vo, T.N.N.; Le, N.T.T.; Thi, D.P.N.; Thi, T.T.H. Evaluation of Saponin-rich/poor leaf extract-mediated silver nanoparticles and their antifungal capacity. *Green Process and Syn.* **2020**, *9* (1), 429–439. Doi: 10.1515/gps-2020-0044.
80. Dai, Y.; Dou, Z.; Zhou, R.; Luo, L.; Bian, L.; Chen, Y.; Tao, J.; Chen, Z. Quality Evaluation of *Artemisia capillaris* Thunb. Based on Qualitative Analysis of the HPLC Fingerprint and UFLC-Q-TOF-MS/MS Combined with Quantitative Analysis of Multicomponents. *J Anal Methods Chem.* **2021**, *2021*, 5546446. Doi: 10.1155/2021/5546446.
81. Dou Z.H.; Dai Y.; Zhou, Y.Z.; Wang, S. Quality evaluation of rhubarb based on qualitative analysis of the HPLC fingerprint and UFLC-Q-TOF-MS/MS combined with quantitative analysis of eight anthraquinone glycosides by QAMS. *Biomed Chromatogr.* **2021**, *35* (6), e5074. Doi: 10.1002/bmc.5074.
82. Xu, J.; Zhou, R.; Luo, L.; Dai, Y.; Feng, Y.; Dou, Z. Quality Evaluation of Decoction Pieces of *Gardeniae Fructus* Based on Qualitative Analysis of the HPLC Fingerprint and Triple-Q-TOF-MS/MS Combined with Quantitative Analysis of 12 Representative Components. *J Anal Methods Chem.* **2022**, *2022*, 2219932. Doi: 10.1155/2022/2219932.
83. Li, W.; Ren, C.; Fei, C.; Wang, Y.; Xue, Q.; Li, L.; Yin, F.; Li, W. Analysis of the chemical composition changes of *Gardeniae Fructus* before and after processing based on ultra-high-performance liquid chromatography quadrupole time-of-flight mass spectrometry. *J Sep Sci.* **2021**, *44* (5), 981–991. Doi: 10.1002/jssc.202000957.
84. Ouyang, W.Z.; Shang, Z.P.; Wang, W.J.; Wang, S.X.; Shen, J.X.; Ma, Z.G.; Zhang, J.Y. Rapid characterization of chemical constituents in capillary wormwood extract based on UHPLC-LTQ-

- Orbitrap. *China Journal of Chinese Materia Medica*. **2017**, 42 (3), 523–530. Doi: 10.19540/j.cnki.cjcmm.2017.0010.
85. Jia, L.; Fu, L.L.; Wang, X.Y.; Yang, W.; Wang, H.; Zuo, T.; Zhang, C.; Hu, Y.; Gao, X.; Han, L. Systematic profiling of the multicomponents and authentication of Erzhi Pill by UHPLC/Q-Orbitrap-MS oriented rapid polarity-switching data-dependent acquisition and selective monitoring of the chemical markers deduced from fingerprint analysis. *Molecules*. **2018**, 23 (12), Doi: 10.3390/molecules23123143.
  86. Tian, F.; Ruan, Q.J.; Zhang, Y.; Cao, H.; Ma, Z.G.; Zhou, G.L.; Wu, M.H. Quantitative analysis of six phenolic acids in *Artemisia capillaris* (Yinchen) by HPLC-DAD and their transformation pathways in decoction preparation process. *J Anal Methods Chem*. **2020**, 8950324. Doi: 10.1155/2020/8950324.
  87. Zhu, T.; Liu, X.; Wang, X.; Cao, G.; Qin, K.; Pei, K.; Zhu, H.; Cai, H.; Niu, M.; Ca, B. Profiling and analysis of multiple compounds in rhubarb decoction after processing by wine steaming using UHPLC-Q-TOF-MS coupled with multiple statistical strategies. *J Sep Sci*. **2016**, 39 (15), 3081–3090. Doi: 10.1002/jssc.201600256.
  88. Jia, J.; Liu, M.; Wen, Q.; He, M.; Ouyang, H.; Chen, L.; Li, J.; Feng, Y.; Zhong, G.; Yang, S. Screening of anti-complement active ingredients from *Eucommia ulmoides* Oliv. branches and their metabolism in vivo based on UHPLC-Q-TOF/MS/MS. *J. Chromatogr. B*. **2019**, 1124:26–36. doi: 10.1016/j.jchromb.2019.05.029.
  89. Han, Y.; Wen, J.; Zhou, T.; Fan, G. Chemical fingerprinting of *Gardenia jasminoides* Ellis by HPLC-DAD-ESI-MS combined with chemometrics methods. *Food Chem*. **2015**, 188, 648–657. Doi: 10.1016/j.foodchem.2015.05.039.
  90. Yang, Z.-R.; Wang, Z.-H.; Tang, J.-F.; Yan, Y.; Yue, S.-J.; Feng, W.-W.; Shi, Z.-Y.; Meng, X.-T.; Peng, C.; Wang, C.-Y.; Meng, D.L.; Yan, D. UPLC-QTOF/MS<sup>E</sup> and Bioassay Are Available Approaches for Identifying Quality Fluctuation of Xueshuantong Lyophilized Powder in Clinic. *Front Pharmacol*. **2018**, 9, 633. Doi: 10.3389/fphar.2018.00633.
  91. Wang, X.; Yao, Y.; An, C.; Li, X.; Xiang, F.; Dong, Y.; Li, M. Simultaneous Determination of 20 Bioactive Components in *Chuanxiong Rhizoma* from Different Production Origins in Sichuan Province by Ultra-high-performance Liquid Chromatography Coupled with Triple Quadrupole Mass Spectrometry Combined with Multivariate Statistical Analysis. *Electrophoresis*. **2020**, 41 (18-19), 1606–1616. Doi:10.1002/elps.202000082.
  92. Zhang, Q.; Hu, X.-F.; Xin, M.-M.; Liu, H.-B.; Sun, L.; Morris-Natschke, S.L.; Chen, Y.; Lee, K.-H. Antidiabetic potential of the ethyl acetate extract of *Physalis alkekengi* and chemical constituents identified by HPLC-ESI-QTOF-MS. *J Ethnopharmacol*. **2018**, 225, 202–210. Doi: 10.1016/j.jep.2018.07.007.
  93. Joo, Y.H.; Nam, M.H.; Chung, N.; Lee, Y.K. UPLC-QTOF-MS/MS screening and identification of bioactive compounds in fresh, aged, and browned *Magnolia denudata* flower extracts. *Food Res Int*. **2020**, 133, 109192. Doi: 10.1016/j.foodres.2020.109192.

94. Liu, M.; He, M.; Gao, H.; Guo, S.; Jia, J.; Ouyang, H.; Feng, Y.; Yang, S. Strategy for rapid screening of antioxidant and anti-inflammatory active ingredients in *Gynura procumbens* (Lour.) Merr. based on UHPLC–Q-TOF–MS/MS and characteristic ion filtration. *Biomed Chromatogr.* **2019**, 33:e4635. Doi: 10.1002/bmc.4635.
95. Nadeem, M.; Mumtaz, M.W.; Danish, M.; Rashid, U.; Mukhtar, H.; Irfan, A. Antidiabetic functionality of *Vitex negundo* L. leaves based on UHPLC-QTOF-MS/MS based bioactives profiling and molecular docking insights. *Ind Crops Prod.* **2020**, 152, 112445. Doi: 10.1016/j.indcrop.2020.112445.
96. Stochmal, A.; Moniuszko-Szajwaj, B.; Zuchwski, J.; Pecio, L.; Konttek, B.; Szumacher-strabel, M.; Olas, B.; Cieslak, A. Qualitative and Quantitative Analysis of Secondary Metabolites in Morphological Parts of Paulownia Clon invitro 112® and Their Anticoagulant Properties in Whole Human Blood. *Molecules.* **2022**, 27 (3), 980. Doi:10.3390/molecules27030980.
97. Gampe, N.; Szakács, Z.; Darcsi, A.; Boldizsár, I.; Szőke, É.; Kuzovkina, I.; Kursinszki, L.; Béni, S. Qualitative and Quantitative Phytochemical Analysis of Ononis Hairy Root Cultures. *Front. Plant Sci.* **2021**, 11, 622585. Doi: 10.3389/fpls.2020.622585.
98. Wang, J.; Jiang, W.; Liu, Z.; Wang, J.; Fu, T.; Wang, Y. Analysis and identification of chemical constituents of fenugreek by UPLC-IT-MS n and UPLC-Q-TOF-MS. *Chem Res Chin Univ.* **2017**, 33, 721–730. Doi: 10.1007/s40242-017-7136-4.
99. Pellati, F.; Brighenti, V.; Sperlea, J.; Marchetti, L.; Bertelli, D.; Benvenuti, S. New Methods for the Comprehensive Analysis of Bioactive Compounds in *Cannabis sativa* L. (hemp). *Molecules.* **2018**, 23 (10), 2639, Doi: 10.3390/molecules23102639.
100. Henzelyova, J.; Antalová, M.; Nigutova, K.; Logoida, M.; Schreiberova, A.; Kusari, S.; Cellárová, E. Isolation, Characterization and Targeted Metabolic Evaluation of Endophytic Fungi Harbored in 14 Seed-Derived *Hypericum Species*. *Planta Med.* **2020**, 86 (13/14) 997-1008. Doi. 10.1055/a-1130-4703.
101. Ali, S.; Khan, M.R.; Ullah, I.; Sajid, M.; Zahra, Z. Phytochemical investigation and antimicrobial appraisal of *Parrotiopsis jacquemontiana* (Decne) Rehder. *BMC Complement Altern Med.* **2018**, 18 (1), 43. Doi: 10.1186/s12906-018-2114-z.
102. Batool, R.; Khan, M.R.; Sajid, M.; Ali, S.; Zahra, Z. Estimation of phytochemical constituents and in vitro antioxidant potencies of *Brachychiton populneus* (Schott & Endl.) R.Br.. *BMC Chemistry.* **2019**, 13 (1), 32. Doi: 10.1186/s13065-019-0549-z.
103. Chiu, C.M.; Nootem, J.; Santiwat, T.; Pratumyot, K.; Srisuwannaket, C.; Chantarasriwong, O.; Lin, W.C.; Mingvanish, W.; Niamnont, N. Enhanced stability and bioactivity of *Curcuma comosa* Roxb. extract in electrospun gelatin nanofibers. *Fibers.* **2019**, 7 (9), 76. doi:10.3390/fib7090076.
104. Muhammad, S.L.; Wada, Y.; Mohammed, M.; Ibrahim, S.; Musa, K.Y.; Olonitola, O.S.; Ahmad, M.H.; Mustapha, S.; Rahman, Z.A.; Sha'aban, A. Bioassay-Guided Identification of Bioactive Compounds from *Senna alata* L. against Methicillin-Resistant *Staphylococcus aureus*. *Appl Microbiol.* **2021**, 1 (3), 520-536. Doi: 10.3390/applmicrobiol1030034.

105. Chen, M.; He, X.; Sun, H.; Sun, Y.; Li, L.; Zhu, J.; Xia, G.; Guo, X.; Zang, H. Phytochemical analysis, UPLC-ESI-Orbitrap-MS analysis, biological activity, and toxicity of extracts from *Tripleurospermum limosum* (Maxim.) Pobed. *Arabian J Chem.* **2022**, 15(5), 103797. Doi: 10.1016/j.arabjc.2022.103797.
106. Rahayu, D.U.C.; Hakim, R.A.; Mawarni, S.A.; Satriani, A.R. Indonesian Cinnamon (*Cinnamomum Burmannii*): Extraction, Flavonoid Content, Antioxidant Activity, and Stability in the Presence of Ascorbic Acid. *Cosmetics.* **2022**, 9 (3), 57. Doi: 10.3390/Cosmetics9030057.
107. Ismail, A.M.; Mohamed, E.A.; Marghany, M.R.; Abdel-Motaal, F.F.; Abdel-Farid, I.B.; El-Sayed, M.A. Preliminary phytochemical screening, plant growth inhibition and antimicrobial activity studies of *Faidherbia albida* legume extracts. *J Saudi Soc Agric Sci.* **2016**, 15 (2), 112-117. Doi. 10/1016/j.jssas.2014.06.002.
108. Auwal, M.S.; Saka, S.; Mairiga, I.A.; Sanda, K.A.; Shuaibu, A.; Ibrahim, A. Preliminary phytochemical and elemental analysis of aqueous and fractionated pod extracts of *Acacia nilotica* (Thorn mimosa). *Vet Res Forum.* **2014**, 5 (2) 95-100.
109. Maria, R.; Shirley, M.; Cornejo, X.; Jaime, S.; David, V.; Rosa, S.; Jodie, D. Preliminary phytochemical screening, total phenolic content and antibacterial activity thirteen of native species from Guayas province Ecuador. *J. King Saud Univ sci.* **2018**, 30 (4), 500-505. Doi. 10/1016/j.jksus.2017.03.009.
110. Kancherla, N.; Dhakshinamoorthi, A.; Chitra, K.; Komaram, R. B. Preliminary Analysis of Phytoconstituents and Evaluation of Anthelmintic property of *Cayratia auriculata* (In vitro). *Maedica (Bucur).* **2019**, 14 (2), 350-356. Doi: 10.26574/maedica.2019.14.4.350.
111. Sharif, M.; Ansari, F.; Hassan, N.U.I.; Sultana, K.; Ali, Q. Explore the antiproliferative from ethanolic extracts of *Citrus paradisi* against liver cancer cell line by chemical analysis using TLC and FT-IR spectroscopy. *Braz J Biol.* **2022**, 82. Doi: 10.1590/1519-6984.256856.
112. Labulo, A.H.; David, O.A.; Terna, A.D. Green Synthesis and characterization of silver nanoparticles using *Morinda lucida* leaf extract and evaluation of its antioxidant and antimicrobial activity. *Chem Pap.* **2022**, 1-13, Doi:10.1007/s11696-022-02392-w.
113. Gollo, A.L.; Tanobe, V.O.A.; de Melo Pereira, G.V.; Marin O.; Bonatto S.J.R.; Silva, S.; de Barros, I.R.; Ricardo, C. Phytochemical analysis and biological activities of in vitro cultured *Nidularium procerum*, a bromeliad vulnerable to extinction. *Sci Rep.* **2020**, 10, 7008. Doi: 10.1038/s41598-020-64026-z.
114. Gurgur, E.; Oluyamo, S.S.; Adetuyi, A.O.; Omotunde O.I.; Okoronkwo, A.E. Green synthesis of zinc oxide nanoparticles and zinc oxide–silver, zinc oxide–copper nanocomposites using *Bridelia ferruginea* as biotemplate. *SN Appl Sci.* **2020**, 2, 911. Doi:10.1007/s42452-020-2269-3.
115. Hamza, R.Z.; Al-Motaani, S.E.; Al-Talhi, T. Therapeutic and Ameliorative effects of Active compounds of *Combretum molle* in the Treatment and Relief from Wounds in Diabetes Mellitus Experimental Model. *Coatings.* **2022**, 12 (8), 1188. Doi: 10.3390/coatings12081188.
116. Kontagora, G.F.; Lawal, N.; Adebote, D.A.; Kamba, B.; Nafiu, M.I.; Jumare, A.I. Some preliminary Phytochemical Screening and Assessment of Four Solvents Extracts of Button Weed (*Borreria verticillata*). *J Appl Sci Envirn Manage.* **2020**, 24 (12), 2085-2088. Doi: 10.4314/jasem.v24i12.12.

117. Tamokou, J.D.D.; Mbaveng, A.T.; Kuete, V. Antimicrobial Activities of African Medicinal Spices and Vegetables. In: *Medicinal Spices and Vegetables from Africa*. Elsevier Inc. 2017, 8, 207-237. Doi: 10.1016/B978-0-12-809286.
118. Raal, A.; Meos, A.; Hinrikus, T.; Heinamaki, J.; Romane, E.; Guidene, V.; Jakstas, V.; Koshovyi, O.; Kovaleva, A.; Fursenco, C.; Chiru, T.; Nguyen, H.T. Dragendorff's reagent: Historical perspectives and current status of a versatile reagent introduced over 150 years ago at the University of Dorpat, Tartu, Estonia. *Pharmazie*. **2020**, 75 (7), 299-306. Doi: 10.1691/ph.2020.0438.
119. Habib, N.; Choudhry, S. HPLC Quantification of Thymoquinone Extracted from *Nigella sativa* L. (*Ranunculaceae*) seeds and Antibacterial Activity of its Extracts against *Bacillus* species. *Evid Based Compl and Altern Med*. **2021**, 4, 1-11. Doi: 10.1155/2021/6645680.
120. Dahanayake, J.M.; Perera, P.K.; Galappatty, P.; Perera, H.D.S.M.; Arawwawala, L.D.A.M. Comparative Phytochemical Analysis and Antioxidant Activities of Tamalakyadi Decoction with Its Modified Dosage Forms. *Evid Based Compl and Alternat Med*. **2019**, 6037137. Doi: 10.1155/2019/6037137.
121. Mikolajczak, N.; Tanska, M.; Ogrodowska, D. Phenolic compounds in plant oil: A review of composition, analytical methods, and effect on oxidative stability. *Trends in food sci and technol*. **2021**, 113, 110-138. Doi: 10.1016/j.tifs.2021.04.046.
122. Shraim, A.M.; Ahmed, T.; Rahman, M.; Hijji, Y.M. Determination of total flavonoid content by aluminum chloride assay: A critical evaluation. *LWT*. **2021**, 150 (2), 111932. doi.org/10.1016/j.lwt.2021.111932.
123. Das, K.; Dang, R.; Sivaraman, G.; Ellath, R.P. Phytochemical Screening of Various Secondary Metabolites, Antioxidant, and Anthelmintic Activity of *Coscinium Fenestratum* Fruit Pulp: A New Biosource for Novel Drug Discovery. *Turk J Pharm Sci*. **2018**, 15 (2), 156-165. Doi:10.4274/tjps.54376.