

## The tumor microenvironment in tumorigenesis and therapy resistance revisited

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## Abstract

Tumorigenesis is a complex and dynamic process involving cell-cell and cell-extracellular matrix (ECM) interactions that allow tumor cell growth, drug resistance and metastasis. This review provides an updated summary of the role played by the tumor microenvironment (TME) components and hypoxia in tumorigenesis and highlight various ways through which tumor cells reprogram normal cells including into phenotypes that are pro-tumorigenic including cancer associated- fibroblasts, -macrophages and -endothelial cells. Tumor cells secrete numerous factors leading to transformation of a previously anti-tumorigenic environment into a pro-tumorigenic environment. Once formed, solid tumors continue to interact with various stromal cells including local and infiltrating fibroblasts, macrophages, mesenchymal stem cells, endothelial cells, pericytes, and secreted factors and the ECM within the tumor microenvironment (TME). The TME is key to tumorigenesis, drug response and treatment outcome. Importantly, stromal cells and secreted factors can initially be anti-tumorigenic but over time promote tumorigenesis and induce therapy resistance. To counter hypoxia, increased angiogenesis leads to formation of new vascular networks in order to actively promote and sustain tumor growth via supply of oxygen and nutrients whilst removing metabolic waste. Angiogenic vascular network formation aid in tumor cell metastatic dissemination. Successful tumor treatment and novel drug development require the identification and therapeutic targeting of pro-tumorigenic components of the TME including cancer-associated- fibroblasts (CAFs) and -macrophages (CAMs), hypoxia, blocking ECM-receptor interactions, in addition to targeting of tumor cells. Re-programming of stromal cells and the immune response to be anti-tumorigenic is key to therapeutic success. Lastly, this review highlights potential TME- and hypoxia-centred therapies under investigations.

**Keywords:** Tumor microenvironment; stromal cells; immune cells; ECM; cancer hallmarks; hypoxia; exosomes; drug resistance; targeted therapy.

Methodology

We retrieved relevant published manuscripts via an electronic search on Embase, Scopus, PubMed and Web of Science using keywords including tumor microenvironment; stromal cells; immune cells; ECM; cancer hallmarks; hypoxia; chemotherapy; multi-drug resistance and targeted therapy. This search yielded a rich source of data on the role of tumor microenvironment in tumorigenesis and therapy resistance (Figure 1). We removed duplicate articles and only full articles were included in compiling this review.

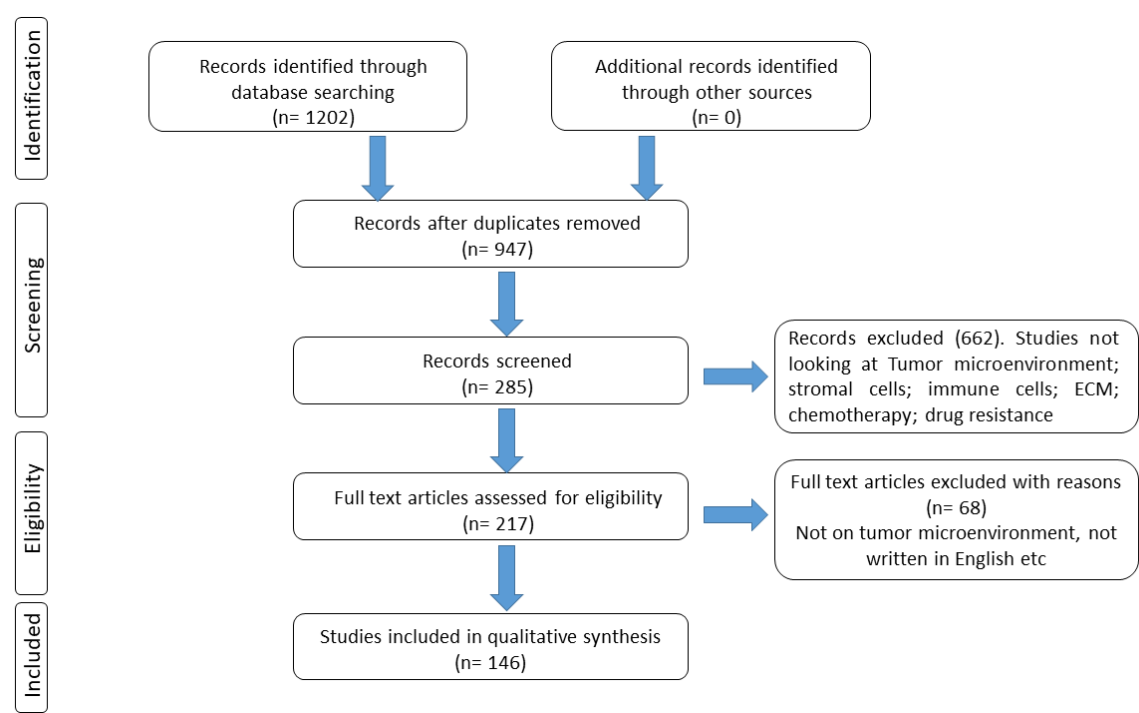


Figure 1. Selection of manuscripts used in the production of this review manuscript.

The tumor microenvironment in brief

It is universally accepted that cancer has major hallmarks including the presence of genomic instability and mutations, unrestricted growth, evasion of growth suppressors, resisting cell death, enhanced inflammation, enhanced metabolism, the ability to promote angiogenesis, invasion and metastasis [1, 2]. It is also scientifically accepted that tumors are more than just tumor cells and

include recruited stromal cells and the non-cellular component, the extracellular matrix (ECM) (Figure 2) [3-6]. Stromal cells and the ECM are active participants during tumorigenesis, starting as anti-tumorigenic during the initial stages to being pro-tumorigenic over time and contributing to the attainment of specific cancer hallmarks [3-6]. Thus, the study and understanding of cancer and tumorigenesis now extends beyond tumor cells to include the stromal cells and the ECM, which make up the tumor microenvironment (TME) [3, 5-18]. Stromal cells include normal fibroblasts, cancer associated fibroblasts (CAFs), cancer associated macrophages (CAMs), mesenchymal stem cells (MSCs), inflammatory cells and endothelial cells [3, 7, 11, 13, 17, 19, 20]. Beside the contribution of the TME during tumorigenesis and metastasis, the TME and common features including hypoxia also play a critical role in therapy resistance [4, 6, 8, 9, 14, 16, 18]. Cell-cell and cell-ECM interactions involve a myriad of biomolecular factors such as growth factors, cytokines, enzymes and chemokines. In addition, exosomes and apoptotic bodies are shown to play roles in promoting tumorigenesis and drug resistance [21]. This review provides a comprehensive description of how the TME, characterized by hypoxia, contribute to tumorigenesis and therapy resistance, and presents ways to reprogramme cells and factors to increase therapy efficacy.

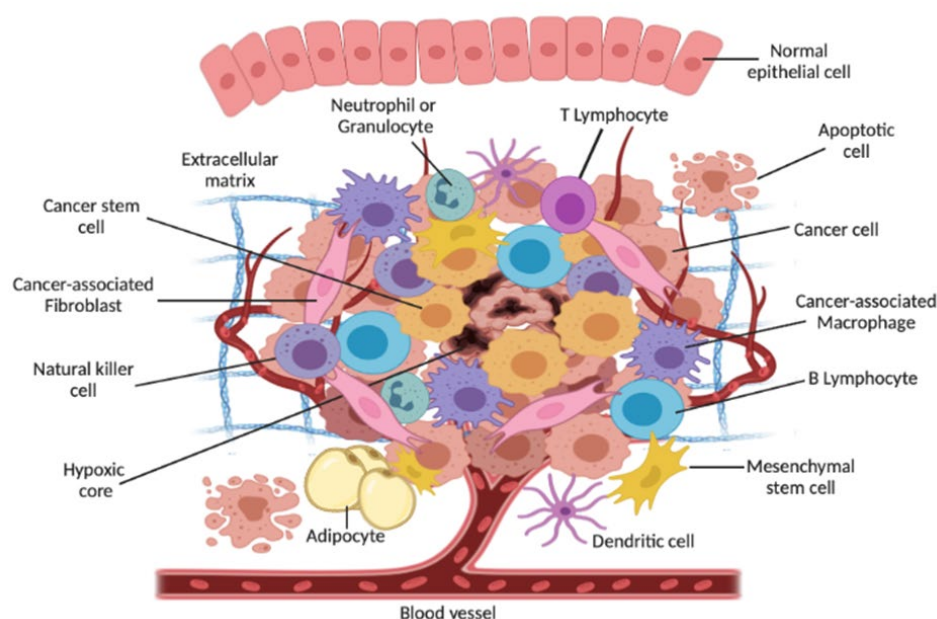


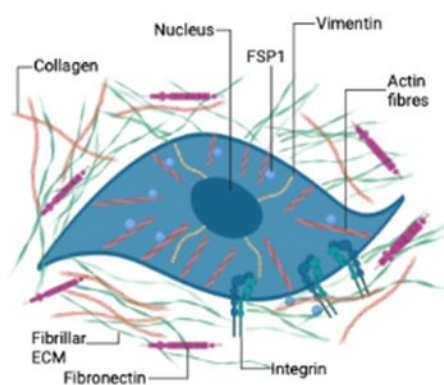
Figure 2. The tumor microenvironment components and their contribution during tumorigenesis.

## Stromal and Immune Cells within the tumor microenvironment

### *Cancer Associated Fibroblasts*

Reports have shown that resident and recruited fibroblasts are part of the TME, where they contribute during tumorigenesis and in drug resistance [3-5, 7, 12, 17, 20, 22]. Initially, fibroblasts are anti-tumorigenic as they are involved in the synthesis of the ECM which surrounds and isolate tumor cells from normal tissue during the early stages of tumorigenesis [5, 6, 23]. Over time, a subpopulation of activated fibroblasts, referred to as cancer associated fibroblasts, obtain a myofibroblastic phenotype characterized by increased synthesis of ECM and release of pro-tumorigenic factors (Figure 3) [6, 24]. Similar to myofibroblasts linked to fibrosis, the CAFs are perpetually activated and promote tumorigenesis via release of factors, activation of pro-tumorigenic signaling, angiogenesis, microRNA, and cytokines [25-29]. At each stage of tumorigenesis, CAFs continue to produce and interact with various TME components including the ECM, cytokines, and growth factors

#### a. Fibroblast



#### b. Cancer-associated fibroblast

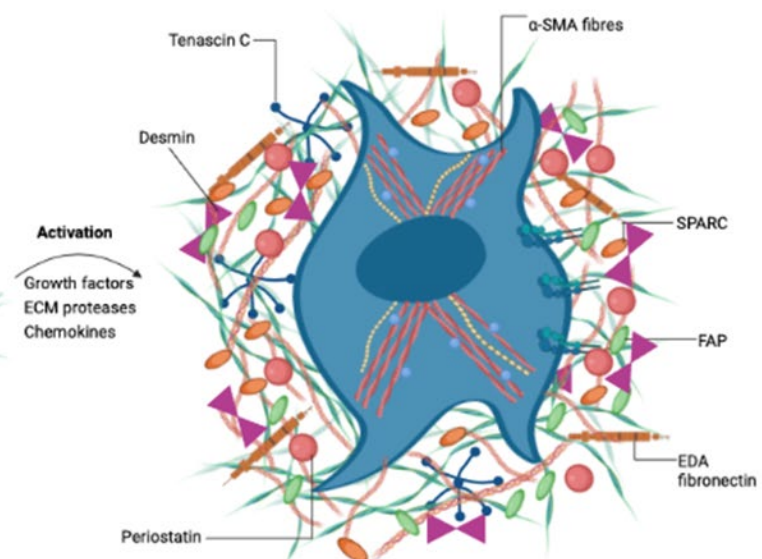
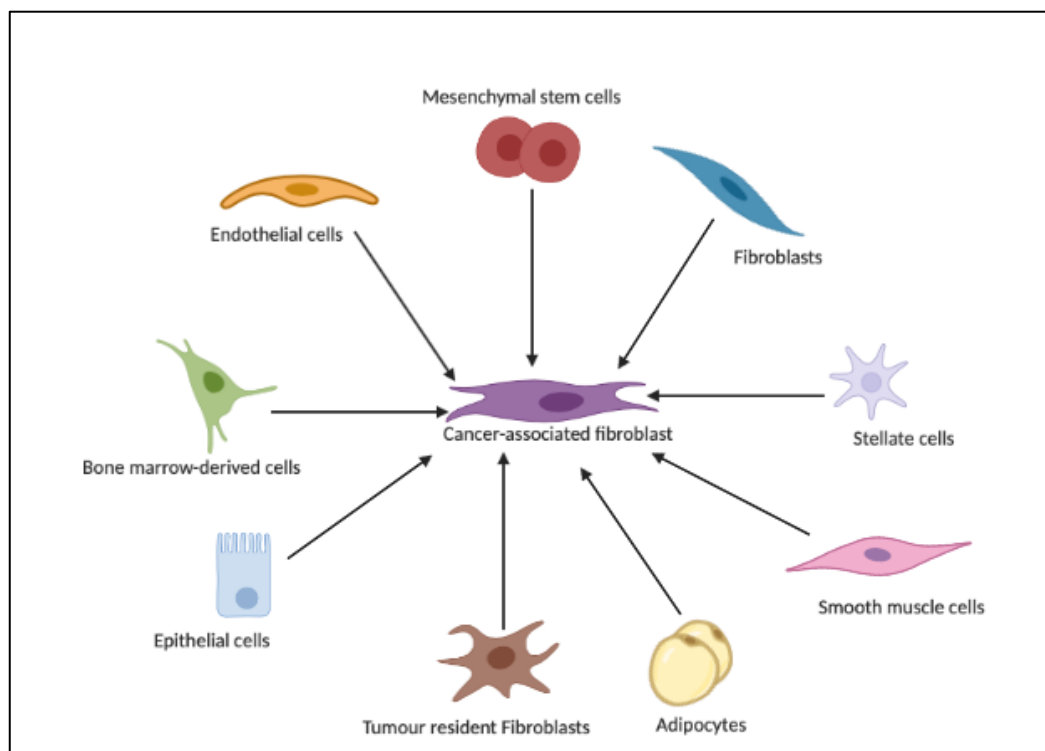


Fig 3. Normal fibroblasts are initially anti-tumorigenic within the TME but over time become activated into CAFs and contribute to tumorigenesis through increased synthesis of factors and the ECM.

Together with several other stromal cells including CAMs and MSCs, CAFs release factors such as TGF- $\beta$  and cytokines involved in ECM remodelling, promotion of tumor cell proliferation, suppression of immune response, recruitment of MSCs as well as induction of angiogenesis [3, 5, 6, 16, 17, 30, 31]. For example, TGF- $\beta$ , from both tumor cells and CAFs, has been shown to promote tumor cell proliferation and to induce EMT transition [32-37]. TGF- $\beta$  overexpression is correlated with poor prognosis in prostate cancer, colorectal cancer, and hepatocellular carcinoma [38-40]. In addition to the expression of TGF- $\beta$ , CAFs also express vascular endothelial growth factor (VEGF) and platelet derived growth factor (PDGF) and this allow their involvement in tumor metastasis [41, 42]. CAF-derived interleukin-6 (IL-6) activation of the Janus kinase (JAK)-signal transducer and activator of transcription (STAT) (JAK-STAT) signaling pathway leads to increased TGF- $\beta$  signaling, promoting tumor growth and metastasis [43-45]. In addition, increased CAFs within the TME and synthesized CXC chemokines correlated with low patient survival in various cancers including colorectal cancer and esophageal cancer [46-49]. Several investigations also show that CAF-derived matrix metalloproteases participate in tumor cell migration and invasion through creation of 'matrix highways' after ECM molecules degradation [50-54]. Using a cell-derived ECM, Senthebane and colleagues demonstrated that fibroblast-derived MMPs contribute towards cancer cell migratory and invasive behaviour [5].

Mounting reports indicate that CAFs originate from different cells and therefore display complex heterogeneity (Figure 4) [16]. Tissue-resident fibroblasts contribute to most CAFs within the TME in addition to other stromal cells such as stellate cells, bone marrow derived- and tissue adult derived-mesenchymal stem cells, pericytes and endothelial cells (Figure 2) [55]. In case of an injury,

the activation of tissue-resident fibroblasts and stellate cells in the liver, for example, reversibly transform these cells into a myofibroblast phenotype characterised by the elevated expression of  $\alpha$ -SMA [55]. Various studies have shown the involvement of growth factors including fibroblast growth factor 2 and TGF- $\beta$  signaling in the transformation of stromal cells into myofibroblastic cells or CAFs [5, 25, 32, 43]. These myofibroblastic cells are the activated fibroblasts responsible for enhanced ECM synthesis in liver cancers. Several studies have also shown that fibrocytes are present in blood [56]. Barth and colleagues demonstrated the presence and the role of CD34+ fibrocytes in invasive ductal carcinoma [57]. Besides breast cancer, the same authors also demonstrated a role for fibrocytes in pancreatic and cervical cancer [58]. Overall, increased levels of CAFs within the TME is associated with tumor relapse and poor prognosis in various cancers.



**Figure 4.** Cancer associated fibroblasts are diverse in origin. The origin of CAFs range from tissue resident fibroblasts, pericytes, endothelial cells to mesenchymal stem cells.



Another potential origin of CAFs is epithelial cells. Epithelial cells near cancer cells can undergo epithelial-to-mesenchymal transition (EMT) and end up as CAFs [59]. Epithelial cancers may display elevated levels of CAFs that drive tumorigenesis [60, 61]. Epithelial cells lose the normal cell-cell adhesive abilities and gain migratory abilities. Endothelial cells can undergo endothelial-to-mesenchymal transition (EMT), transforming these cells into CAFs [62]. CAFs originating from epithelial and endothelial cells produce CAF markers such as S100A4 [59, 62]. Both adipocytes and pericytes can undergo trans-differentiation into CAFs [63]. It is important to note that while CAFs are pro-tumorigenic, studies also indicate that CAFs can act in an anti-tumorigenic manner [64, 65]. Only recently, is a clear and well-defined picture of CAFs and their role in tumorigenesis emerging.

CAFs heterogeneity means numerous subgroups exist with contrasting phenotypes and functions within the TME [66, 67]. Reports also show that CAFs heterogeneity is linked to stage of tumour development [68]. ECM remodelling and stromal cell transformations during different stages of tumorigenesis can lead to CAFs being genetically unstable [69, 70]. Thus, CAFs co-evolve with tumour cells during tumorigenesis. The initial anti-tumour activity of stromal cells become ‘tumour-promoting’ activity over time [5, 6]. Various signalling cascades modulate CAFs activation and activity and these include the lysophosphatidic acid and TGF- $\beta$  family ligands which influence serum response factor (SRF) and SMAD transcription factors activities, respectively, to promote the expression of the activated fibroblast marker  $\alpha$ -SMA [71]. Co-culture of cancer cells and fibroblasts demonstrate the promotion of CAF activation in breast cancer via the Notch signalling [72]. Furthermore, inflammatory modulators including interleukin-1 $\beta$  (IL-1 $\beta$ ) can induce NF- $\kappa$ B activation in CAFs [73]. CAF markers include FAP, PDGFR $\alpha/\beta$ , tenascin C, vimentin, desmin, CD90 and podoplanin (PDPN) (Table 1) [74]. CAFs heterogeneity means that there is no universal marker and early studies utilized  $\alpha$ -SMA and FAP-alpha [74]. A combination of these markers is



the ideal means to identify CAFs. Other markers include  $\alpha$ -SMA, vimentin and CD10. The expression of  $\alpha$ -SMA is not exclusive to CAFs as other cells such as smooth muscle cells and pericytes express the same marker [66]. CAFs found in several cancers, such as breast and pancreatic cancers, express high levels of  $\alpha$ -SMA and vimentin [55, 75].

**Table 1.** Markers for cancer-associated fibroblasts

CAFs Marker	Description and function of protein	Effect within TME
$\alpha$ -SMA	Actin isoform: cellular contraction and maintenance of structure	Promote tumor cell proliferation; involved in immunosuppression
Tenascin-C	Extracellular matrix glycoprotein: cell migration; wound healing	Impeding drug delivery; protect tumor cells
Vimentin	Type III intermediate filament protein: cell migration; cell structure maintenance	Tumor cell migration and invasion
PDGFR $\alpha/\beta$	Protein tyrosine kinase receptor: cellular signaling	Macrophage polarization; angiogenesis
FAP	Membrane-bound gelatinase: protease activity; ECM remodelling	Angiogenesis, macrophage polarization; immunosuppression; metastasis
GPR77	Complement component 5a receptor 2: Activation of complement; promote inflammation	Maintains tumor cell stemness; Drug resistance
Vimentin	Intermediate filament protein: Maintains cell structure and integrity; cell migration	Tumor migration, invasion, and metastasis
Caveolin-1	Scaffolding protein within caveolar membranes: maintains cellular structure and signaling	Low caveolin-1 linked to poor prognosis

Targeting CAFs, with their significant heterogeneity, involves reversal of the transformation from normal fibroblasts into CAFs. Reports indicate that the use of microRNA can achieve such de-

activation or reprogramming of CAFs into normal fibroblasts [76-78]. De-differentiation of CAFs into quiescent cells is another strategy under consideration [79].

#### *Cancer associated endothelial cells*

New blood vessel formation during tumorigenesis is initiated by endothelial cells and these cells constitute the innermost layer of blood vessels [80]. The usually thin vascular endothelium separates blood from tissues in addition to delivering important nutrients, ions, and water [81]. The vascular endothelium is also important in carrying away all toxic metabolic waste products. Immune cells are also carried to tumors via the blood stream. Whilst diffusion is responsible for oxygen supply and carbon dioxide removal during the initial stages of tumorigenesis, increase in size of tumor will require increased supply of oxygen as well as removal of metabolic waste [82]. As the tumor increase in size, a hypoxic core is formed, activating the tumor to form new blood vessels to supply much-needed nutrients and oxygen [83, 84]. Vascular networks are formed as a result of the action of various transcription factors induced by hypoxia. The transcription factors induced by hypoxia act on endothelial cells which release growth factors such as epidermal growth factor (EGF), PDGF to form new blood vessels [85, 86]. Old blood vessels can also sprout and form new branching vessels. Beside growth factors, endothelial cells also release proteins required for the formation of basement membranes. Due to the unregulated release of cytokines and growth factors, blood vessel formation is not proper within a tumor. This results in 'makeshift' blood vessels that are leaky [87]. Being responsible for new blood vessel formation makes endothelial cells important for cancer cell migration and metastasis. As the blood vessels within tumors are leaky, cancer cells can easily invade new tissues and intravasate into blood vessels to be transported to new sites [88]. Endothelial cells can also undergo 'endothelial to mesenchymal transition' to become cancer associated fibroblasts as they are very plastic [89, 90]. Various growth factors including TGF- $\beta$  are known to be involved in this transition [91]. Cancer associated endothelial cells promotes tumorigenesis by being immunosuppressive, growth factor synthesis and enhanced

migratory behaviour of tumor cells [92, 93]. Cancer associated endothelial cell also aid immunosuppressing myeloid cells infiltration into tumors.

#### *Cancer-associated macrophages*

In the human body, macrophages, mostly originating from circulating monocytes, participate in various processes from clearing infections, wound healing as well as repair of tissues [94]. As part of the innate immune system macrophages respond to the presence of pathogens by presenting antigens and carrying out phagocytosis [95]. M1 macrophages are the predominant type of macrophages during the initial stages of tumorigenesis, as they participate in phagocytosis of pathogens and antigen presentation [96]. A tumor is sometimes referred to as a 'wound' that does not heal. Thus, within the tumor microenvironment the M2 macrophages are present and actively participate in suppressing the immune system and wound healing [97]. Deep inside the tumor, lack of oxygen and various cytokines are known to promote the M2 type of macrophages [97, 98]. Infiltration of tumors with macrophages occur throughout the process of tumorigenesis and macrophages can account up to a third of the mass of the tumor at some stages. Reports indicate that an elevated levels of macrophages within tumors are associated with low survival rates in various cancers [99, 100]. This is attributed to macrophages' promotion of angiogenesis via release of various cytokines and thus enhance formation of new blood vessels. Recent data also show that CAMs play key roles in chemoresistance to drugs such as paclitaxel and 5-fluorouracil [101-106]. Furthermore, CAMs have been shown to promote CSCs tumorigenic capacity as well as their therapeutic resistance via increased enzyme synthesis (cytidine deaminase) involved in drug metabolism [102-104].

*Cancer-associated neutrophils*

When an infection occurs, circulating leukocytes, and specifically neutrophils provide the first line of defence against pathogens [107]. Within the tumor microenvironment, neutrophils can have both pro- and anti-tumorigenic properties [108]. During the initial stages of tumorigenesis, recruited neutrophils release various cytokines including IL6 thereby inducing inflammation [109, 110]. This causes tumor cells to undergo apoptosis. Neutrophils also release reactive oxygen species that induce apoptosis in tumor cells [94]. In later stages of tumorigenesis, neutrophils release various growth factors such as VEGF involved in angiogenesis, and therefore promotes tumorigenesis through new blood vessel formation [111, 112]. Neutrophils are also involved in ECM remodelling via production of matrix metalloproteases (MMPs) [113]. MMPs are also actively involved in promoting tumor cell invasion and eventual metastasis via degradation of ECM molecules [114].

*T cells*

Various populations of T cells have been identified within the tumor microenvironment at various stages of tumor development [115]. Specific T cell populations have specific receptors used in antigen identification. For example, cytotoxic T cells with specific receptors identify abnormal antigens expressed on tumor cells and their attachment to tumor cells leads to the destruction of the cells [115, 116]. Cytotoxic T cells also play a key role in preventing formation of new blood vessels via the release of the pleiotropic cytokine interferon-gamma [117]. Thus, cytotoxic T cells demonstrate anti-tumorigenic behaviour within the tumor microenvironment [118]. Another population of T cells found within the tumor microenvironment are the CD4<sup>+</sup> T cells. CD4<sup>+</sup> T cells are mainly involved in immune responses within the tumor microenvironment and over time differentiate into several cells [119]. For example, CD4<sup>+</sup> T cells can become T-helper 1 cells, which participate in inflammation induction and their presence within various tumors is linked to increased patient survival [120-122]. Another T cell type found within tumors is the regulatory T

cells. Regulatory T cells participate in suppressing inflammation and anti-tumor immune responses [123-126]. Regulatory T cells releases interleukin-2 which controls the function of natural killer cells [127-129]. Furthermore, regulatory T cells secrete various growth factors and cytokines and advertently supports tumorigenesis

#### *B cells*

B cells are responsible for antibody production in the body as well as secretion of various cytokines [130-133]. B cells are mostly localized at the periphery of tumors and within lymph nodes near the tumor site [131, 133]. Thus, few B cells are found within tumors [131, 133]. The main function of B cells during tumorigenesis is their close relationship with T cells, allowing T cells to act against tumor cells. B cells act as antigen presenting cells to T cells [134-137]. B cells are also involved in secretion of anti-tumorigenic cytokines such as IFN- $\gamma$  [134-137]. However, several studies also show that B cells are pro-tumorigenic in some tumors [138-140]. It has been shown that regulatory B cells produce various cytokines including IL-10 and TGF- $\beta$  that promote immune suppression via their effects on macrophages and T cells [141-143].

#### *Natural Killer cells*

Cells found in the blood and infected with viruses are destroyed through the action of natural killer cells [8, 144, 145]. Natural killers do not distinguish between virally infected-tumor cells and -host cells. A sub-category of natural killer cells participates directly in killing tumor cells whilst another sub-category produces inflammatory cytokines [8, 122, 144]. Inflammation will lead to accumulation of various immune cells involved in tumor cell killing. By seeking and destroying tumor cells within the bloodstream, natural killer cells are important in preventing metastasis and formation of secondary tumors [146-148]. Within tumors, natural killer cells are less efficient at killing tumor cells.

### *Dendritic cells*

The function of dendritic cells is mostly to recognize and capture antigens as well as present them to T cells [149-151]. Dendritic cells are mostly found within lymph nodes where they participate in T cell response to specific pathogen infection [152, 153]. Depending on the prevailing environment within tumors, dendritic cells can be both anti- and pro-tumorigenic [151]. Overproduction of pro-tumorigenic growth factors and cytokines can lead to dendritic cells tolerating the presence of tumor cells and act to prevent an immune reaction [151].

### *Stellate Cells*

Found in the liver and the pancreas, stellate cells originated from mesenchymal tissue and are mostly involved in promoting tumorigenesis via differentiation into myofibroblasts [154-156]. Injury to the liver and pancreas induce stellate cell differentiation into myofibroblasts after which they synthesize enormous quantities of ECM molecules and growth factors including VEGF [157-159]. The development of a tumor, akin to 'wound healing' induce stellate cells differentiation into myofibroblasts. One major function of stellate cells is the accumulation of vitamin A in lipids droplets [160-162]. The lipid droplets are then utilized during ECM synthesis and production of MMPs. Tumor cell-derived TGF- $\beta$  is known to be involved in the activation of hepatic stellate cells into myofibroblast in liver cancer. Both liver cancer and pancreatic cancer tend to be associated with fibrosis. Quiescent pancreatic stellate cells are involved in remodelling of the ECM via MMPs synthesis and ECM protein synthesis [163]. Once activated, pancreatic stellate cells secrete various biomolecules leading to their increased migratory behaviour and proliferation.

### *Adipocytes*

Two cell types, adipocytes, and white adipose tissue, constitute the adipose tissue [164]. Energy storage as well as maintenance of energy balance in the body is the function of adipocytes or fat cells. Given the high energy required by tumor cells during tumor initiation and progression, it is not surprising therefore that adipocytes play a key role in this process [164]. Adipocytes have been shown to secrete various biomolecules from growth factors, enzymes to cytokines [165, 166]. The secretion of enzymes including MMPs leads to ECM remodelling, allowing tumor cells to migrate and metastasize. Obesity is considered a high-risk factor in many cancers with close to half cancer patients being obese for example in breast and ovarian cancers [167]. Reports show that white adipose tissue is linked to increased risk of cancers and the formation of secondary tumors in lungs for example [168]. Organs with a high number of adipocytes include the breast and these cells have been shown to be pro-tumorigenic [169]. As tumor cells require a lot of energy, adipocytes can be induced to undergo lipolysis which converts lipids into fatty acids that can be used by tumor cells during tumorigenesis [164, 170]. Furthermore, adipocytes secrete various hormones including leptin that promotes tumor cell proliferation and migration as well as the recruitment of immune cells to the TME [171]. Adipose-derived adult stem cells, which can differentiate into different cell lineages, also come from adipose tissue. These stem cells have the ability to enhance inflammation within the TME and thus are pro-tumorigenic [172, 173]. It is possible that adipose-derived stem cells can differentiate into cancer-associated stromal cells such as CAFs.

### *Mesenchymal stem cells*

Important for maintenance of health tissue and repair of tissue in case of injury, mesenchymal stem cells or mesenchymal stromal cells are able to differentiate into cell types such as osteoblasts, and chondrocytes [174, 175]. This differentiation ability is the reason why MSCs recruited to tumors can transform into various tumor associated cells. Reports indicate that beside resident



fibroblasts differentiation into CAFs, recruited MSCs can also be transformed into CAFs [4, 16, 17, 20, 25]. Whilst resident fibroblasts may initially have an anti-tumorigenic phenotype, it is reported that over time all fibroblasts are pro-tumorigenic [5]. During the initial stages of tumorigenesis, fibroblasts synthesise large quantities of ECM proteins, in what appear to be an attempt at isolating the tumor from the rest of the tissue [5]. Increase in ECM synthesis also causes stiffening of tissue. Increase in tissue stiffness has been associated with tumorigenesis [176]. In later stages of tumorigenesis MSCs demonstrate immunoregulatory effects by contributing to the dampening of the anti-tumor immunity [177, 178].

### *Pericytes*

Pericytes have multiple roles within the tumor microenvironment including covering endothelial cells along the surface of the endothelium, in the remodelling of the basement membrane during tumorigenesis and formation of new blood vessels [87, 179]. Pericytes have also been involved in immunoregulatory process through activation of immune cells such as lymphocytes and in phagocytosis [180, 181]. Although clinical trials targeting pericytes involvement in angiogenesis has been done, results so far are not promising. Some reports even show that targeting pericytes leads to more tumor cells metastasizing [182, 183]. For example, targeting pericytes in animal models of breast cancer resulted in aggressive pulmonary tumor process [184]. It has been postulated that pericytes may display heterogeneity and there is need to target the correct pericyte subpopulation with a specific phenotype to stop tumorigenesis [185-187].

### **The Extracellular Matrix**

One key component of the TME is the ECM. Forming the structural part of the TME, the ECM is located under the epithelial layer surrounding the connective tissue cells [188, 189]. CAFs are

the main source of ECM components. It is made up of many macromolecules including vitronectin, collagens, proteoglycans, and glycoproteins (e.g., fibronectin, laminin) (Figure 5) [188]. Its composition is always changing depending on the stage of tumorigenesis [190], and this is facilitated by enzymes such as cathepsins, lysyl oxidase (LOX), MMPs, and their inhibitors [191]. In solid tumors, the ECM can constitute about half of the tumor mass (desmoplastic tumors) and has been linked to poor patient survival [192].

The elasticity and rigidity of the ECM promote tumorigenesis via integrin signalling [193]. Changes in ECM composition and elasticity influence many aspects of tumorigenesis varying from cancer cell growth, survival and therapy resistance [193]. Collagen, the most abundant ECM molecule in tumors [193], provides structural support to tumor cells and regulating other processes such as tumor cell adhesion, supporting chemotaxis and migration. Enhanced levels of type I collagen also increase ECM stiffness and promote tumorigenesis in the process [193]. Enzyme-linked changes in ECM composition and levels facilitate tumor cell migration via the creation of 'pores' allowing tumor cells to invade surrounding tissues and travel to distant tissues and organs [194]. Increased collagen production and the resulting stiffness influence integrin signaling and tumor cell survival [194].

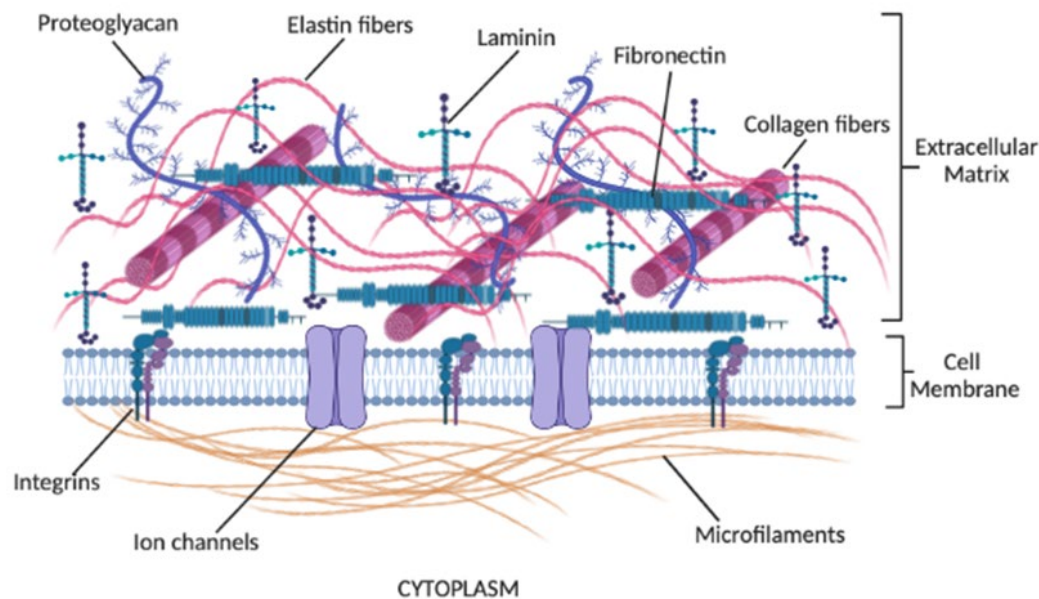


Figure 5. Components of the ECM include glycoproteins, collagens, proteoglycans, and polysaccharides. Collagens and glycoproteins are ligand for integrins and play key roles in tumor cell signaling necessary for survival.

Importantly, the ECM presents a physical hindrance to drug distribution within tumors [195]. In most cases, this physical hindrance as well as sequestration of drugs through direct binding to ECM molecules contributes to development of drug resistance in many solid tumors [196]. Furthermore, various reports show that the ECM is key to tumour vascularisation [197]. New blood vessel formation is important to tumorigenesis. As production of ECM molecules such as collagen increases, the resulting increased ECM density causes a decrease in vascularisation. A stiff ECM compresses blood vessels, limiting the flow of drugs and oxygen within the TME [197, 198]. The lack of enough oxygen within tumors influence vascularisation via the activation of HIF-1 $\alpha$ . HIF-1 $\alpha$  promotes chemoresistance via activation of MDR1 expression in hypoxic colon cancer, for example [199, 200]. Lastly, the ECM can sequester various growth factors and cytokines that can promote tumorigenesis such as TGF- $\beta$ , VEGF and PDGF.

## Vascular Networks

Tumor cells require supplies of oxygen and nutrients to maintain their uncontrolled growth [201]. This is achieved through the vascular and lymphatic networks that allows gaseous exchange and removal of toxic waste from the tumor (Figure 6) [202, 203]. A major hallmark of cancer is the process of angiogenesis. The tumor microenvironment becomes hypoxic as the tumor continues to grow as the vasculature cannot supply oxygen to all cells within the TME [203]. New blood vessels formed from pre-existing ones are 'leaky' and convoluted [204, 205]. Similar to growth of tumor cells which is uncontrolled, blood vessel formation continues unabated with no proper control, resulting in a complex structure. Leaky vessels also help tumor cells to migrate to other tissues to form secondary tumors as well as contribute to ineffective distribution of drugs within the TME. Lymphatic vessels also provide a 'throughfare' through which tumor cells can migrate to other sites [206].

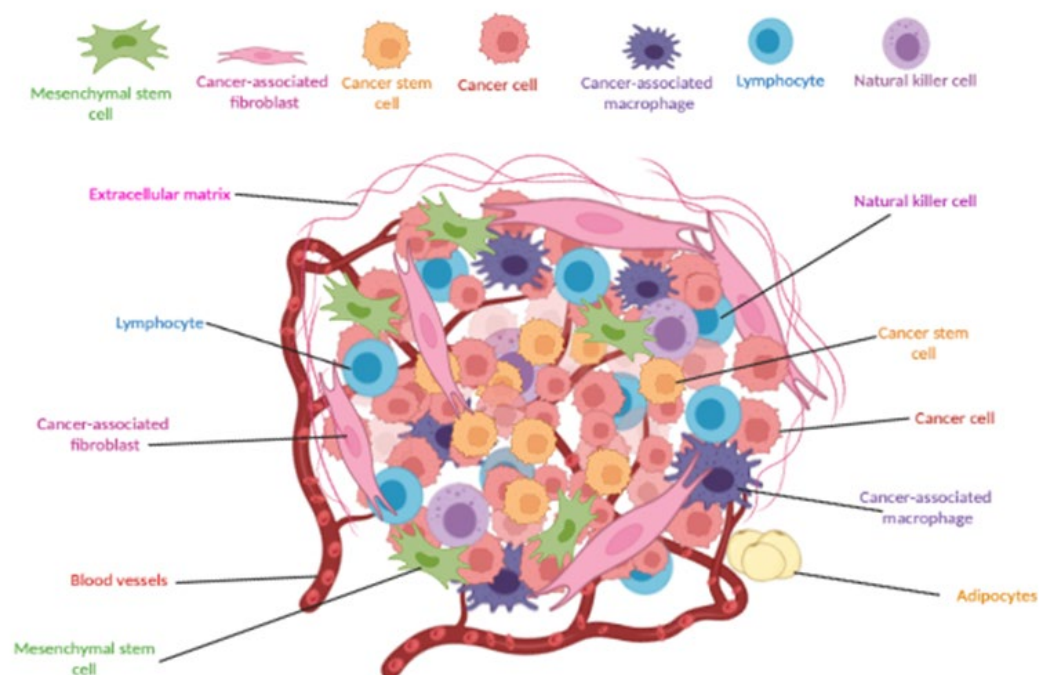


Figure 6. The blood and lymphatic vascular networks are important during tumorigenesis. Supply of oxygen and removal of carbon dioxide and other metabolic waste products is achieved by the blood vessels. Leaky blood and lymphatic vessels also allow tumor cells to migrate to other tissues.

## Hypoxia within the TME

A hallmark of the unregulated proliferation of tumor cells is the unavailability of oxygen or hypoxia and nutrients in some parts of a growing solid tumor [207, 208]. Synthesis of new blood vessels via angiogenesis does not occur fast enough to provide oxygen to rapidly growing tumor cells. The result is tumors with some regions having less than 2 % oxygen levels, thus are hypoxic [208, 209]. Importantly, angiogenesis within a growing tumor leads to dysregulated vasculature and oxygenated blood is not supplied to all regions. Tumor cells within hypoxic regions obtain a different phenotype to those in regions properly supplied with oxygenated blood, are more aggressive and become resistant to commonly used drugs [210]. Indeed, oxygen gradients within solid tumors is a common feature. Tumor cells within hypoxic regions also express elevated levels of hypoxia-inducible factor alpha (HIF-1 $\alpha$ ), with three isoforms having been found in mammals [211]. HIFs play central roles in tumorigenesis in which they influence hypoxia-induced gene expression and metabolism [212]. For example, HIF-1 is especially important in tumor cell response to therapy [213]. HIF-1 $\alpha$  also enhances the activities of transcriptional factors including Twist and Snail, leading to increased endothelial-to-mesenchymal transition (EMT) [214, 215]. By modulating collagen synthesis and collagen fibre alignment as well as integrin-ECM interactions within the TME, HIF-1 $\alpha$  also aid tumor cell migration and metastasis [216, 217]. In addition, due to lack of oxygen, tumor cells within hypoxic regions of TME divide slowly, thus can circumvent common drugs targeting rapidly dividing tumor cells.

As a tumor grows, de novo angiogenesis leads to formation of leaky blood vessels leading to increase in interstitial fluid pressure [88, 218]. Furthermore, leaky blood vessels aid tumor cell metastasis as tumor cells can easily escape the blood vessels with discontinuous endothelium. Various reports documented that cells within hypoxic TME region also promote immunosuppression. For example, cancer-associated macrophages of the M2 type have been

found in hypoxic regions [219, 220]. The immunosuppressive properties of CAMs are well documented. HIF-1 $\alpha$  can modulate the behaviour of myeloid-derived suppressor cells within hypoxic regions of TME [221]. Hypoxia also cause the TME to be acidic and under these conditions T cells are not able to perform their cytotoxic functions [222]. Further data show that hypoxia can induce the over-expression of various proteins involved in drug efflux [223]. Reports show that blocking of HIF-1 $\alpha$  expression can reverse drug resistance in cancers [224, 225]. Drug resistance can also emanate from tumor cells altering their metabolism and avoiding apoptosis. Hypoxia can also induce autophagy, which can lead to multi-drug resistance [226]. Overall, hypoxia within the TME can be used as an independent prognostic factor in cancers and predicts poor outcomes [227, 228]. Thus, novel strategies must target tumor hypoxia together with various components of the TME.

### **Exosomes and Exosomal miRNAs in tumor microenvironment**

Ranging in size from 30 to 200 nm, exosomes play key roles in cellular communication between tumor cells and stromal cells and are secreted into the extracellular space by cells regularly [229]. The contents of exosomes depend on their origin, with stromal cell-derived exosomes containing various growth factors, cytokines and other signaling molecules that can impact tumor cell behaviour as well as cell-cell interactions [230]. In most cases, the contents of exosomes promote tumorigenesis via impacting processes such as angiogenesis, migration, and metastasis [231]. Reports indicate that tumor cells under conditions of low oxygen and nutrients produce increased levels of exosomes and leads to alterations of stromal cells into pro-tumorigenic cells including CAFs and CAMs [21, 232]. Tumor cell-derived exosomes also have the ability to prepare some tissue-specific cells for colonization by tumor cells [233, 234].

Importantly, exosomes are key to transporting microRNAs [21]. Stromal cells can alter microRNAs (miRNAs) expression in both tumor cells and stromal cells [232]. Alteration of miRNA expression can be induced by tumor and stromal cell interactions through release of auto- and paracrine factors [79, 231]. For example, microRNA-122 from breast cancer cells has been shown to reprogram normal cell metabolism by reducing uptake of glucose by lung cells in preparation for lung colonization [235-237]. This will make sure there is enough nutrients for metastatic breast tumor cells upon lung colonization. Delineating miRNAs functions within the TME can lead to new therapeutic targets identification. Exosomes are useful as diagnostic biomarkers as well as therapeutic targets [238]. Exosomes are stable within the circulatory system and their contents can be used for diagnosis purposes and can predict tumor metastasis accurately [239, 240]. Given their many functions during tumorigenesis, reports indicate that abrogation of exosome production can inhibit tumorigenesis [21, 241]. The suppression of tumor-derived exosomes uptake through the use of heparin resulted in decreased metastatic ability of oral squamous cell carcinoma [242].

In terms of cancer treatment, exosomes can be used to deliver drugs as they non-toxic and biodegradable [243]. Ligands specific for certain tumors can be expressed on the surface of the exosomes so as to direct them to specific tumor cells [244, 245]. Such tumor cell-specific exosomes can then deliver therapeutic siRNA or drugs for example to kill cancer cells [246].

### **Advances in Therapeutic targeting of TME**

Great improvements have been brought to cancer treatment through combinations of various drugs and immunotherapy in the past few years. Chemotherapy, used mostly as the first line of cancer treatment, target rapidly growing cancer cells, and tends to be broad in their focus [247,



248]. By specifically targeting sub-populations of cancer cells within the TME including CSCs, improvements have been made in cancer treatment [3, 18]. In addition, the introduction of immunotherapy and specifically immune checkpoint blockade such as PD1 that targets several immune cells within the TME brought remarkable success in cancer treatment [249]. Immune checkpoint inhibitors are antibodies or drugs that block proteins called checkpoints from immune system cells including T cells as well as some cancer cells [250, 251]. Normally these checkpoints prevent the immune responses from being too strong and this impact T cells' ability to kill cancer cells [252, 253]. Importantly, the identification of biomarkers can lead to the grouping of patients that can benefit from specific drugs and therapies.

New therapies also include the prevention of new blood vessel formation. Tumorigenesis is a process that depends on the constant supply of oxygen and nutrients to growing tumor cells [87]. Furthermore, metabolic waste must be removed, without which the microenvironment becomes toxic even for tumor cells. Thus, the prevention of angiogenesis through the use of anti-angiogenic drugs including those neutralizing growth factors such as VEGF, decoy receptors for growth factors is an appealing strategy under intense investigation. Small molecule inhibitors of several factors released within the TME including AMD3465 can prevent stromal cell-derived factors from being pro-tumorigenic [254, 255]. Antagonists of integrins can prevent cell-cell and cell-ECM interactions within the TME, increasing cancer cell response to drugs in the process [256]. Inactivation of HIF-1 $\alpha$  has been shown to enhance the effect of carboplatin on tumor cell proliferation and thus can be used as a hypoxia-centred therapy [257, 258]. The acidification of the TME by hypoxia also reduces some drug effectiveness as this depends on the surrounding pH. Adjustment of TME pH can therefore be used to enhance or decrease efficacy of drugs [259]. Drugs that can be activated in hypoxic regions of tumors have been suggested. These hypoxic pro-drugs can be activated into cytotoxic drugs by enzymes found within hypoxic regions of tumors.

For example, TH-302 is a hypoxic pro-drug utilized together with gemcitabine in the treatment of pancreatic cancer, which is highly hypoxic with oxygen levels averaging around 0.7% [260, 261]. Various signaling cascades important in hypoxia including the unfolded protein response are appealing targets to treat solid tumors characterized by hypoxia [262, 263]. Other strategies to avoid hypoxia-induced changes to drug effectiveness make use of nanoparticles to deliver drugs directly to tumor cells. Other strategies to inhibit hypoxia-mediated HIF response is to use small-interfering RNA.

As expected, therapy resistance is a major problem when these strategies are used. Combination therapy involving the use of two or more anti-tumor strategies results in better responses. More research is needed including evaluating the efficiency of these strategies before these strategies are commonplace in clinics.

## **Conclusion**

Treatment of cancer ranging from the use of surgery, chemotherapy, radiotherapy, and recently introduced immunotherapy have all had limited success when used alone. In most cases, combination therapy is the best strategy to use for successful treatment. However, therapy resistance develops as tumor cells are heterogeneous and plastic in nature and tumor cells can convert a non-supporting 'anti-tumorigenic' environment into a 'pro-tumorigenic' environment. The contribution of tumor microenvironment to tumorigenesis, metastasis, and development of therapy resistance is of note. Thus, it is important to delineate the role played by various TME components in tumorigenesis, metastasis and therapy development. This review discusses the identification of predictive, prognostic biomarkers via the analysis of TME components and how this reveals the complexity of tumor biology as well as lead to development of targeted therapies

for specific cancers and patients. Importantly, the recruitment of non-tumorigenic cells and non-cellular components by tumor cells for their benefit, allows tumorigenesis to proceed without hindrances. Stromal cells and immune cells are reprogrammed by tumor cells to release various factors that favor tumor cell growth and survival. Hypoxic microenvironment has been noted to play key roles in tumorigenesis and drug resistance. Understanding the processes involved in regulating hypoxia can lead to new therapeutic targets. In this regard, exosomes have been identified as useful as diagnostic and therapeutic tools by revealing tumor-derived secretome and can deliver drugs to tumor cells, respectively. Currently, combination therapy targeting various components of the TME can lead to best results during treatment.

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## References

1. Hanahan, D. and R.A. Weinberg, *Hallmarks of cancer: the next generation*. cell, 2011. **144**(5): p. 646-674.
2. Fouad, Y.A. and C. Aanei, *Revisiting the hallmarks of cancer*. American journal of cancer research, 2017. **7**(5): p. 1016.
3. Dzobo, K., *Taking a Full Snapshot of Cancer Biology: Deciphering the Tumor Microenvironment for Effective Cancer Therapy in the Oncology Clinic*. Omics, 2020. **24**(4): p. 175-179.
4. Erin, N., et al., *Tumor microenvironment and epithelial mesenchymal transition as targets to overcome tumor multidrug resistance*. Drug Resist Updat, 2020. **53**: p. 100715.
5. Senthebane, D.A., et al., *The Role of Tumor Microenvironment in Chemoresistance: 3D Extracellular Matrices as Accomplices*. Int J Mol Sci, 2018. **19**(10).
6. Senthebane, D.A., et al., *The Role of Tumor Microenvironment in Chemoresistance: To Survive, Keep Your Enemies Closer*. Int J Mol Sci, 2017. **18**(7).
7. Arneth, B., *Tumor Microenvironment*. Medicina (Kaunas), 2019. **56**(1).
8. Hinshaw, D.C. and L.A. Shevde, *The Tumor Microenvironment Innately Modulates Cancer Progression*. Cancer Res, 2019. **79**(18): p. 4557-4566.
9. Hui, L. and Y. Chen, *Tumor microenvironment: Sanctuary of the devil*. Cancer Lett, 2015. **368**(1): p. 7-13.
10. Jarosz-Biej, M., et al., *Tumor Microenvironment as A "Game Changer" in Cancer Radiotherapy*. Int J Mol Sci, 2019. **20**(13).
11. Kim, J. and J.S. Bae, *Tumor-Associated Macrophages and Neutrophils in Tumor Microenvironment*. Mediators Inflamm, 2016. **2016**: p. 6058147.
12. Soysal, S.D., A. Tzankov, and S.E. Muenst, *Role of the Tumor Microenvironment in Breast Cancer*. Pathobiology, 2015. **82**(3-4): p. 142-52.
13. Vitale, I., et al., *Macrophages and Metabolism in the Tumor Microenvironment*. Cell Metab, 2019. **30**(1): p. 36-50.
14. Wu, T. and Y. Dai, *Tumor microenvironment and therapeutic response*. Cancer Lett, 2017. **387**: p. 61-68.
15. Dzobo, K., *Integrins Within the Tumor Microenvironment: Biological Functions, Importance for Molecular Targeting, and Cancer Therapeutics Innovation*. Omics, 2021. **25**(7): p. 417-430.
16. Dzobo, K. and C. Dandara, *Broadening Drug Design and Targets to Tumor Microenvironment? Cancer-Associated Fibroblast Marker Expression in Cancers and Relevance for Survival Outcomes*. Omics, 2020. **24**(6): p. 340-351.
17. Dzobo, K. and C. Dandara, *Architecture of Cancer-Associated Fibroblasts in Tumor Microenvironment: Mapping Their Origins, Heterogeneity, and Role in Cancer Therapy Resistance*. Omics, 2020. **24**(6): p. 314-339.
18. Dzobo, K., et al., *Advances in Therapeutic Targeting of Cancer Stem Cells within the Tumor Microenvironment: An Updated Review*. Cells, 2020. **9**(8).
19. Bussard, K.M., et al., *Tumor-associated stromal cells as key contributors to the tumor microenvironment*. Breast Cancer Research, 2016. **18**(1): p. 84.
20. Denton, A.E., E.W. Roberts, and D.T. Fearon, *Stromal Cells in the Tumor Microenvironment*. Adv Exp Med Biol, 2018. **1060**: p. 99-114.
21. Zhao, L., et al., *The role of exosomes and "exosomal shuttle microRNA" in tumorigenesis and drug resistance*. Cancer letters, 2015. **356**(2): p. 339-346.
22. Bussard, K.M., et al., *Tumor-associated stromal cells as key contributors to the tumor microenvironment*. Breast Cancer Res, 2016. **18**(1): p. 84.
23. Hanley, C.J., et al., *Targeting the Myofibroblastic Cancer-Associated Fibroblast Phenotype Through Inhibition of NOX4*. J Natl Cancer Inst, 2018. **110**(1): p. 109-20.

24. Augsten, M., *Cancer-associated fibroblasts as another polarized cell type of the tumor microenvironment*. *Frontiers in oncology*, 2014. **4**: p. 62.
25. Orimo, A., et al., *Stromal fibroblasts present in invasive human breast carcinomas promote tumor growth and angiogenesis through elevated SDF-1/CXCL12 secretion*. *Cell*, 2005. **121**(3): p. 335-48.
26. Olumi, A.F., et al., *Carcinoma-associated fibroblasts direct tumor progression of initiated human prostatic epithelium*. *Cancer Res*, 1999. **59**(19): p. 5002-11.
27. Busch, S., et al., *TGF-beta receptor type-2 expression in cancer-associated fibroblasts regulates breast cancer cell growth and survival and is a prognostic marker in pre-menopausal breast cancer*. *Oncogene*, 2015. **34**(1): p. 27-38.
28. Tanaka, K., et al., *miR-27 is associated with chemoresistance in esophageal cancer through transformation of normal fibroblasts to cancer-associated fibroblasts*. *Carcinogenesis*, 2015. **36**(8): p. 894-903.
29. Min, A., et al., *Downregulation of Microrna-148a in Cancer-Associated Fibroblasts from Oral Cancer Promotes Cancer Cell Migration and Invasion by Targeting Wnt10b*. *J Biochem Mol Toxicol*, 2016. **30**(4): p. 186-91.
30. Wiseman, B.S. and Z. Werb, *Stromal effects on mammary gland development and breast cancer*. *Science*, 2002. **296**(5570): p. 1046-9.
31. Xue, X., et al., *Galectin-1 secreted by activated stellate cells in pancreatic ductal adenocarcinoma stroma promotes proliferation and invasion of pancreatic cancer cells: an in vitro study on the microenvironment of pancreatic ductal adenocarcinoma*. *Pancreas*, 2011. **40**(6): p. 832-9.
32. Sun, D.-Y., et al., *Cancer-associated fibroblast regulate proliferation and migration of prostate cancer cells through TGF- $\beta$  signaling pathway*. *Life sciences*, 2019. **235**: p. 116791.
33. Maluccio, M., et al., *Tacrolimus enhances transforming growth factor- $\beta$ 1 expression and promotes tumor progression*. *Transplantation*, 2003. **76**(3): p. 597-602.
34. Moses, H. and M.H. Barcellos-Hoff, *TGF- $\beta$  biology in mammary development and breast cancer*. *Cold Spring Harbor perspectives in biology*, 2011. **3**(1): p. a003277.
35. Wang, L., et al., *Cancer-associated fibroblasts enhance metastatic potential of lung cancer cells through IL-6/STAT3 signaling pathway*. *Oncotarget*, 2017. **8**(44): p. 76116.
36. Yu, Y., et al., *Cancer-associated fibroblasts induce epithelial-mesenchymal transition of breast cancer cells through paracrine TGF- $\beta$  signalling*. *British journal of cancer*, 2014. **110**(3): p. 724-732.
37. Hao, Y., D. Baker, and P. Ten Dijke, *TGF- $\beta$ -mediated epithelial-mesenchymal transition and cancer metastasis*. *International journal of molecular sciences*, 2019. **20**(11): p. 2767.
38. Reis, S.T.d., et al., *Tgf- $\beta$ 1 expression as a biomarker of poor prognosis in prostate cancer*. *Clinics*, 2011. **66**: p. 1143-1147.
39. Robson, H., et al., *Transforming growth factor  $\beta$  1 expression in human colorectal tumours: an independent prognostic marker in a subgroup of poor prognosis patients*. *British journal of cancer*, 1996. **74**(5): p. 753-758.
40. Huang, C.-y., et al., *Transforming growth factor  $\beta$  is a poor prognostic factor and inhibits the favorable prognostic value of CD8+ CTL in human hepatocellular carcinoma*. *Journal of Immunotherapy*, 2017. **40**(5): p. 175-186.
41. Desmouliere, A., C. Guyot, and G. Gabbiani, *The stroma reaction myofibroblast: a key player in the control of tumor cell behavior*. *International Journal of Developmental Biology*, 2004. **48**(5-6): p. 509-517.
42. Huang, J., et al., *Different roles of myofibroblasts in the tumorigenesis of nonsmall cell lung cancer*. *Tumor Biology*, 2016. **37**(12): p. 15525-15534.
43. Shi, J., et al., *Targeted blockade of TGF- $\beta$  and IL-6/JAK2/STAT3 pathways inhibits lung cancer growth promoted by bone marrow-derived myofibroblasts*. *Scientific Reports*, 2017. **7**(1): p. 8660.

44. Yamamoto, T., et al., *Cross-talk between IL-6 and TGF- $\beta$  signaling in hepatoma cells*. FEBS Letters, 2001. **492**(3): p. 247-253.
45. O'Reilly, S., et al., *Interleukin-6 (IL-6) Trans Signaling Drives a STAT3-dependent Pathway That Leads to Hyperactive Transforming Growth Factor- $\beta$  (TGF- $\beta$ ) Signaling Promoting SMAD3 Activation and Fibrosis via Gremlin Protein*. Journal of Biological Chemistry, 2014. **289**(14): p. 9952-9960.
46. Verbeke, H., et al., *The expression and role of CXC chemokines in colorectal cancer*. Cytokine & growth factor reviews, 2011. **22**(5-6): p. 345-358.
47. Yang, F., et al., *CXCR1 correlates to poor outcomes of EGFR-TKI against advanced non-small cell lung cancer by activating chemokine and JAK/STAT pathway*. Pulmonary Pharmacology & Therapeutics, 2021. **67**: p. 102001.
48. Goto, M. and M. Liu, *Chemokines and their receptors as biomarkers in esophageal cancer*. Esophagus, 2020. **17**(2): p. 113-121.
49. Do, H.T.T., C.H. Lee, and J. Cho, *Chemokines and their receptors: multifaceted roles in cancer progression and potential value as cancer prognostic markers*. Cancers, 2020. **12**(2): p. 287.
50. Cui, N., M. Hu, and R.A. Khalil, *Biochemical and Biological Attributes of Matrix Metalloproteinases*. Prog Mol Biol Transl Sci, 2017. **147**: p. 1-73.
51. Huang, H., *Matrix Metalloproteinase-9 (MMP-9) as a Cancer Biomarker and MMP-9 Biosensors: Recent Advances*. Sensors (Basel), 2018. **18**(10).
52. Karamanou, K., et al., *Epithelial-to-mesenchymal transition and invadopodia markers in breast cancer: Lumican a key regulator*. Semin Cancer Biol, 2020. **62**: p. 125-133.
53. Najafi, M., B. Farhood, and K. Mortezaee, *Extracellular matrix (ECM) stiffness and degradation as cancer drivers*. J Cell Biochem, 2019. **120**(3): p. 2782-2790.
54. Pittayapruerk, P., et al., *Role of Matrix Metalloproteinases in Photoaging and Photocarcinogenesis*. Int J Mol Sci, 2016. **17**(6).
55. Yin, C., et al., *Hepatic stellate cells in liver development, regeneration, and cancer*. The Journal of clinical investigation, 2013. **123**(5): p. 1902-1910.
56. Gomperts, B.N. and R.M. Strieter, *Fibrocytes in lung disease*. Journal of leukocyte biology, 2007. **82**(3): p. 449-456.
57. Barth, P.J., et al., *CD34+ fibrocytes in invasive ductal carcinoma, ductal carcinoma in situ, and benign breast lesions*. Virchows Archiv, 2002. **440**(3): p. 298-303.
58. Barth, P.J., et al., *CD34+ fibrocytes in neoplastic and inflammatory pancreatic lesions*. Virchows Archiv, 2002. **440**(2): p. 128-133.
59. Iwano, M., et al., *Evidence that fibroblasts derive from epithelium during tissue fibrosis*. The Journal of clinical investigation, 2002. **110**(3): p. 341-350.
60. Orimo, A. and R.A. Weinberg, *Stromal fibroblasts in cancer: a novel tumor-promoting cell type*. Cell cycle, 2006. **5**(15): p. 1597-1601.
61. Dzobo, K. and C. Dandara, *Architecture of cancer-associated fibroblasts in tumor microenvironment: Mapping their origins, heterogeneity, and role in cancer therapy resistance*. Omics: a journal of integrative biology, 2020. **24**(6): p. 314-339.
62. Zeisberg, E.M., et al., *Discovery of endothelial to mesenchymal transition as a source for carcinoma-associated fibroblasts*. Cancer research, 2007. **67**(21): p. 10123-10128.
63. Jotzu, C., et al., *Adipose tissue derived stem cells differentiate into carcinoma-associated fibroblast-like cells under the influence of tumor derived factors*. Cellular oncology, 2011. **34**(1): p. 55-67.
64. Gorchs, L., et al., *The vitamin D analogue calcipotriol promotes an anti-tumorigenic phenotype of human pancreatic CAFs but reduces T cell mediated immunity*. Scientific reports, 2020. **10**(1): p. 1-15.
65. Dzobo, K. and C. Dandara, *Broadening drug design and targets to tumor microenvironment? Cancer-associated fibroblast marker expression in cancers and relevance for survival outcomes*. Omics: a journal of integrative biology, 2020. **24**(6): p. 340-351.



66. Öhlund, D., E. Elyada, and D. Tuveson, *Fibroblast heterogeneity in the cancer wound*. Journal of Experimental Medicine, 2014. **211**(8): p. 1503-1523.
67. Öhlund, D., et al., *Distinct populations of inflammatory fibroblasts and myofibroblasts in pancreatic cancer*. Journal of Experimental Medicine, 2017. **214**(3): p. 579-596.
68. Huelken, J. and D. Hanahan, *A subset of cancer-associated fibroblasts determines therapy resistance*. Cell, 2018. **172**(4): p. 643-644.
69. Campbell, I., W. Qiu, and I. Haviv, *Genetic changes in tumour microenvironments*. The Journal of pathology, 2011. **223**(4): p. 450-458.
70. Lim, K.P., et al., *Fibroblast gene expression profile reflects the stage of tumour progression in oral squamous cell carcinoma*. The Journal of pathology, 2011. **223**(4): p. 459-469.
71. Sahai, E., et al., *A framework for advancing our understanding of cancer-associated fibroblasts*. Nature Reviews Cancer, 2020. **20**(3): p. 174-186.
72. Strell, C., et al., *Impact of Epithelial-Stromal Interactions on Peritumoral Fibroblasts in Ductal Carcinoma in Situ*. J Natl Cancer Inst, 2019. **111**(9): p. 983-995.
73. Erez, N., et al., *Cancer-Associated Fibroblasts Are Activated in Incipient Neoplasia to Orchestrate Tumor-Promoting Inflammation in an NF-kappaB-Dependent Manner*. Cancer Cell, 2010. **17**(2): p. 135-47.
74. Kalluri, R., *The biology and function of fibroblasts in cancer*. Nature Reviews Cancer, 2016. **16**(9): p. 582.
75. Ayala, G., et al., *Reactive stroma as a predictor of biochemical-free recurrence in prostate cancer*. Clinical Cancer Research, 2003. **9**(13): p. 4792-4801.
76. Musumeci, M., et al., *Control of tumor and microenvironment cross-talk by miR-15a and miR-16 in prostate cancer*. Oncogene, 2011. **30**(41): p. 4231-42.
77. Bronisz, A., et al., *Reprogramming of the tumour microenvironment by stromal PTEN-regulated miR-320*. Nat Cell Biol, 2011. **14**(2): p. 159-67.
78. Aprelikova, O., et al., *Silencing of miR-148a in cancer-associated fibroblasts results in WNT10B-mediated stimulation of tumor cell motility*. Oncogene, 2013. **32**(27): p. 3246-53.
79. Yang, X., et al., *Role of exosomes in crosstalk between cancer-associated fibroblasts and cancer cells*. Frontiers in Oncology, 2019. **9**: p. 356.
80. Davies, G., et al., *Levels of expression of endothelial markers specific to tumour-associated endothelial cells and their correlation with prognosis in patients with breast cancer*. Clinical & experimental metastasis, 2004. **21**(1): p. 31-37.
81. De Sanctis, F., et al. *The dark side of tumor-associated endothelial cells*. in *Seminars in Immunology*. 2018. Elsevier.
82. Ronca, R., J.A. Van Ginderachter, and A. Turtoi, *Paracrine interactions of cancer-associated fibroblasts, macrophages and endothelial cells: tumor allies and foes*. Current Opinion in Oncology, 2018. **30**(1): p. 45-53.
83. Rankin, E.á. and A. Giaccia, *The role of hypoxia-inducible factors in tumorigenesis*. Cell Death & Differentiation, 2008. **15**(4): p. 678-685.
84. Jensen, R.L., *Hypoxia in the tumorigenesis of gliomas and as a potential target for therapeutic measures*. Neurosurgical focus, 2006. **20**(4): p. E24.
85. Ye, J. and C. Koumenis, *ATF4, an ER stress and hypoxia-inducible transcription factor and its potential role in hypoxia tolerance and tumorigenesis*. Current molecular medicine, 2009. **9**(4): p. 411-416.
86. Jensen, R.L., *Brain tumor hypoxia: tumorigenesis, angiogenesis, imaging, pseudoprogression, and as a therapeutic target*. Journal of neuro-oncology, 2009. **92**(3): p. 317-335.
87. Bergers, G. and L.E. Benjamin, *Tumorigenesis and the angiogenic switch*. Nature reviews cancer, 2003. **3**(6): p. 401-410.
88. Yehya, A.H.S., et al., *Angiogenesis: managing the culprits behind tumorigenesis and metastasis*. Medicina, 2018. **54**(1): p. 8.



89. Kovacic, J.C., et al., *Epithelial-to-mesenchymal and endothelial-to-mesenchymal transition: from cardiovascular development to disease*. Circulation, 2012. **125**(14): p. 1795-1808.
90. Potenta, S., E. Zeisberg, and R. Kalluri, *The role of endothelial-to-mesenchymal transition in cancer progression*. British journal of cancer, 2008. **99**(9): p. 1375-1379.
91. Wesseling, M., et al., *The morphological and molecular mechanisms of epithelial/endothelial-to-mesenchymal transition and its involvement in atherosclerosis*. Vascular pharmacology, 2018. **106**: p. 1-8.
92. Motz, G.T. and G. Coukos, *The parallel lives of angiogenesis and immunosuppression: cancer and other tales*. Nature Reviews Immunology, 2011. **11**(10): p. 702-711.
93. Frumento, G., et al., *Targeting tumor-related immunosuppression for cancer immunotherapy*. Endocrine, Metabolic & Immune Disorders-Drug Targets (Formerly Current Drug Targets-Immune, Endocrine & Metabolic Disorders), 2006. **6**(3): p. 223-237.
94. Galdiero, M.R., et al., *Tumor associated macrophages and neutrophils in cancer*. Immunobiology, 2013. **218**(11): p. 1402-1410.
95. Solinas, G., et al., *Tumor-associated macrophages (TAM) as major players of the cancer-related inflammation*. Journal of leukocyte biology, 2009. **86**(5): p. 1065-1073.
96. Chen, J.J., et al., *Tumor-associated macrophages: the double-edged sword in cancer progression*. Journal of clinical oncology, 2005. **23**(5): p. 953-964.
97. Jayasingam, S.D., et al., *Evaluating the polarization of tumor-associated macrophages into M1 and M2 phenotypes in human cancer tissue: technicalities and challenges in routine clinical practice*. Frontiers in Oncology, 2020. **9**: p. 1512.
98. Hu, W., et al., *Tumor-associated macrophages in cancers*. Clinical and Translational Oncology, 2016. **18**(3): p. 251-258.
99. Almatroodi, S.A., et al., *Characterization of M1/M2 tumour-associated macrophages (TAMs) and Th1/Th2 cytokine profiles in patients with NSCLC*. Cancer Microenvironment, 2016. **9**(1): p. 1-11.
100. Heusinkveld, M. and S.H. van Der Burg, *Identification and manipulation of tumor associated macrophages in human cancers*. Journal of translational medicine, 2011. **9**(1): p. 1-14.
101. Paulus, P., et al., *Colony-stimulating factor-1 antibody reverses chemoresistance in human MCF-7 breast cancer xenografts*. Cancer research, 2006. **66**(8): p. 4349-4356.
102. DeNardo, D.G., et al., *Leukocyte complexity predicts breast cancer survival and functionally regulates response to chemotherapy*. Cancer discovery, 2011. **1**(1): p. 54-67.
103. Dijkgraaf, E.M., et al., *Chemotherapy Alters Monocyte Differentiation to Favor Generation of Cancer-Supporting M2 Macrophages in the Tumor Microenvironment*. Effect of Chemotherapy on Tumor Microenvironment. Cancer research, 2013. **73**(8): p. 2480-2492.
104. Mantovani, A. and P. Allavena, *The interaction of anticancer therapies with tumor-associated macrophages*. Journal of Experimental Medicine, 2015. **212**(4): p. 435-445.
105. Jinushi, M. and Y. Komohara, *Tumor-associated macrophages as an emerging target against tumors: Creating a new path from bench to bedside*. Biochimica et Biophysica Acta (BBA)-Reviews on Cancer, 2015. **1855**(2): p. 123-130.
106. Weizman, N., et al., *Macrophages mediate gemcitabine resistance of pancreatic adenocarcinoma by upregulating cytidine deaminase*. Oncogene, 2014. **33**(29): p. 3812-3819.
107. Shaul, M.E. and Z.G. Fridlender, *Tumour-associated neutrophils in patients with cancer*. Nature reviews Clinical oncology, 2019. **16**(10): p. 601-620.
108. Coffelt, S.B., M.D. Wellenstein, and K.E. de Visser, *Neutrophils in cancer: neutral no more*. Nature Reviews Cancer, 2016. **16**(7): p. 431-446.
109. Wu, L., et al., *Tumor-associated neutrophils in cancer: going pro*. Cancers, 2019. **11**(4): p. 564.
110. Gregory, A.D. and A. McGarry Houghton, *Tumor-associated neutrophils: new targets for cancer therapy*. Cancer research, 2011. **71**(7): p. 2411-2416.
111. Masucci, M.T., M. Minopoli, and M.V. Carriero, *Tumor associated neutrophils. Their role in tumorigenesis, metastasis, prognosis and therapy*. Frontiers in oncology, 2019. **9**: p. 1146.

112. Tolle, F., et al., *Neutrophils in Tumorigenesis: Missing Targets for Successful Next Generation Cancer Therapies?* International Journal of Molecular Sciences, 2021. **22**(13): p. 6744.
113. Houghton, A.M., *The paradox of tumor-associated neutrophils: fueling tumor growth with cytotoxic substances.* Cell cycle, 2010. **9**(9): p. 1732-1737.
114. Jabłońska-Trypuć, A., M. Matejczyk, and S. Rosochacki, *Matrix metalloproteinases (MMPs), the main extracellular matrix (ECM) enzymes in collagen degradation, as a target for anticancer drugs.* Journal of enzyme inhibition and medicinal chemistry, 2016. **31**(sup1): p. 177-183.
115. Maimela, N.R., S. Liu, and Y. Zhang, *Fates of CD8+ T cells in tumor microenvironment.* Computational and structural biotechnology journal, 2019. **17**: p. 1-13.
116. Vlachonikola, E., K. Stamatopoulos, and A. Chatzidimitriou, *T cells in chronic lymphocytic leukemia: a two-edged sword.* Frontiers in Immunology, 2021. **11**: p. 612244.
117. Andersen, M.H., et al., *Cytotoxic T cells.* Journal of Investigative Dermatology, 2006. **126**(1): p. 32-41.
118. Finlay, D. and D.A. Cantrell, *Metabolism, migration and memory in cytotoxic T cells.* Nature Reviews Immunology, 2011. **11**(2): p. 109-117.
119. Lindau, D., et al., *The immunosuppressive tumour network: myeloid-derived suppressor cells, regulatory T cells and natural killer T cells.* Immunology, 2013. **138**(2): p. 105-115.
120. Navasardyan, I. and B. Bonavida, *Regulation of T Cells in Cancer by Nitric Oxide.* Cells, 2021. **10**(10).
121. Tang, X.X., H. Shimada, and N. Ikegaki, *Clinical Relevance of CD4 Cytotoxic T Cells in High-Risk Neuroblastoma.* Front Immunol, 2021. **12**: p. 650427.
122. Zheng, Y., et al., *Immune suppressive landscape in the human esophageal squamous cell carcinoma microenvironment.* Nat Commun, 2020. **11**(1): p. 6268.
123. Dees, S., et al., *Regulatory T cell targeting in cancer: Emerging strategies in immunotherapy.* Eur J Immunol, 2021. **51**(2): p. 280-291.
124. Elkord, E. and V. Sasidharan Nair, *T-Regulatory Cells in Health and Disease.* J Immunol Res, 2018. **2018**: p. 5025238.
125. Sasidharan Nair, V., et al., *Metabolic reprogramming of T regulatory cells in the hypoxic tumor microenvironment.* Cancer Immunol Immunother, 2021. **70**(8): p. 2103-2121.
126. Wang, H., et al., *Regulatory T-cell and neutrophil extracellular trap interaction contributes to carcinogenesis in non-alcoholic steatohepatitis.* J Hepatol, 2021. **75**(6): p. 1271-1283.
127. Kordasti, S.Y., et al., *CD4+CD25<sup>high</sup> Foxp3<sup>+</sup> regulatory T cells in myelodysplastic syndrome (MDS).* Blood, 2007. **110**(3): p. 847-50.
128. Lim, K.P., et al., *CD4+CD25<sup>hi</sup>CD127<sup>low</sup> regulatory T cells are increased in oral squamous cell carcinoma patients.* PLoS One, 2014. **9**(8): p. e103975.
129. Mitchell, D.A., et al., *Monoclonal antibody blockade of IL-2 receptor  $\alpha$  during lymphopenia selectively depletes regulatory T cells in mice and humans.* Blood, 2011. **118**(11): p. 3003-12.
130. Chen, V.E., et al., *The Underappreciated Role of the Humoral Immune System and B Cells in Tumorigenesis and Cancer Therapeutics: A Review.* Int J Radiat Oncol Biol Phys, 2020. **108**(1): p. 38-45.
131. Corsiero, E., et al., *B cells in the formation of tertiary lymphoid organs in autoimmunity, transplantation and tumorigenesis.* Curr Opin Immunol, 2019. **57**: p. 46-52.
132. Mintz, M.A. and J.G. Cyster, *T follicular helper cells in germinal center B cell selection and lymphomagenesis.* Immunol Rev, 2020. **296**(1): p. 48-61.
133. Roghanian, A., et al., *B Cells Promote Pancreatic Tumorigenesis.* Cancer Discov, 2016. **6**(3): p. 230-2.
134. Bruno, T.C., et al., *Antigen-Presenting Intratumoral B Cells Affect CD4(+) TIL Phenotypes in Non-Small Cell Lung Cancer Patients.* Cancer Immunol Res, 2017. **5**(10): p. 898-907.
135. Chen, X. and P.E. Jensen, *The role of B lymphocytes as antigen-presenting cells.* Arch Immunol Ther Exp (Warsz), 2008. **56**(2): p. 77-83.

136. Ghosh, D., et al., *New insights into B cells as antigen presenting cells*. Curr Opin Immunol, 2021. **70**: p. 129-137.
137. Rastogi, I., et al., *Role of B cells as antigen presenting cells*. Front Immunol, 2022. **13**: p. 954936.
138. Edechi, C.A., et al., *Regulation of Immunity in Breast Cancer*. Cancers (Basel), 2019. **11**(8).
139. Gupta, S.L., et al., *B-Cell-Based Immunotherapy: A Promising New Alternative*. Vaccines (Basel), 2022. **10**(6).
140. Kuroda, H., et al., *Tumor microenvironment in triple-negative breast cancer: the correlation of tumor-associated macrophages and tumor-infiltrating lymphocytes*. Clin Transl Oncol, 2021. **23**(12): p. 2513-2525.
141. Catalán, D., et al., *Immunosuppressive Mechanisms of Regulatory B Cells*. Front Immunol, 2021. **12**: p. 611795.
142. Rosser, E.C. and C. Mauri, *Regulatory B cells: origin, phenotype, and function*. Immunity, 2015. **42**(4): p. 607-12.
143. Wang, L., Y. Fu, and Y. Chu, *Regulatory B Cells*. Adv Exp Med Biol, 2020. **1254**: p. 87-103.
144. Barry, K.C., et al., *A natural killer-dendritic cell axis defines checkpoint therapy-responsive tumor microenvironments*. Nat Med, 2018. **24**(8): p. 1178-1191.
145. Terrén, I., et al., *NK Cell Metabolism and Tumor Microenvironment*. Front Immunol, 2019. **10**: p. 2278.
146. Chan, I.S., et al., *Cancer cells educate natural killer cells to a metastasis-promoting cell state*. J Cell Biol, 2020. **219**(9).
147. Guillerey, C., N.D. Huntington, and M.J. Smyth, *Targeting natural killer cells in cancer immunotherapy*. Nat Immunol, 2016. **17**(9): p. 1025-36.
148. López-Soto, A., et al., *Control of Metastasis by NK Cells*. Cancer Cell, 2017. **32**(2): p. 135-154.
149. Gajewski, T.F., H. Schreiber, and Y.X. Fu, *Innate and adaptive immune cells in the tumor microenvironment*. Nat Immunol, 2013. **14**(10): p. 1014-22.
150. Gardner, A. and B. Ruffell, *Dendritic Cells and Cancer Immunity*. Trends Immunol, 2016. **37**(12): p. 855-865.
151. Lee, Y.S. and K.J. Radford, *The role of dendritic cells in cancer*. Int Rev Cell Mol Biol, 2019. **348**: p. 123-178.
152. Volovitz, I., et al., *Dendritic Cells in the Context of Human Tumors: Biology and Experimental Tools*. Int Rev Immunol, 2016. **35**(2): p. 116-35.
153. Zhou, B., T. Lawrence, and Y. Liang, *The Role of Plasmacytoid Dendritic Cells in Cancers*. Front Immunol, 2021. **12**: p. 749190.
154. Barry, A.E., et al., *Hepatic Stellate Cells and Hepatocarcinogenesis*. Front Cell Dev Biol, 2020. **8**: p. 709.
155. Wu, Y., et al., *The Role of Stellate Cells in Pancreatic Ductal Adenocarcinoma: Targeting Perspectives*. Front Oncol, 2020. **10**: p. 621937.
156. Zhao, S., et al., *Highly-metastatic colorectal cancer cell released miR-181a-5p-rich extracellular vesicles promote liver metastasis by activating hepatic stellate cells and remodelling the tumour microenvironment*. J Extracell Vesicles, 2022. **11**(1): p. e12186.
157. Baglieri, J., D.A. Brenner, and T. Kisseleva, *The Role of Fibrosis and Liver-Associated Fibroblasts in the Pathogenesis of Hepatocellular Carcinoma*. Int J Mol Sci, 2019. **20**(7).
158. Elechalawar, C.K., et al., *Targeting Pancreatic Cancer Cells and Stellate Cells Using Designer Nanotherapeutics in vitro*. Int J Nanomedicine, 2020. **15**: p. 991-1003.
159. Tan, H.X., et al., *CXCR4/TGF- $\beta$ 1 mediated hepatic stellate cells differentiation into carcinoma-associated fibroblasts and promoted liver metastasis of colon cancer*. Cancer Biol Ther, 2020. **21**(3): p. 258-268.
160. Lua, I., et al., *Characterization of hepatic stellate cells, portal fibroblasts, and mesothelial cells in normal and fibrotic livers*. J Hepatol, 2016. **64**(5): p. 1137-1146.

161. Senoo, H., Y. Mezaki, and M. Fujiwara, *The stellate cell system (vitamin A-storing cell system)*. Anat Sci Int, 2017. **92**(4): p. 387-455.
162. Senoo, H., et al., *Hepatic stellate cell (vitamin A-storing cell) and its relative--past, present and future*. Cell Biol Int, 2010. **34**(12): p. 1247-72.
163. Ferdek, P.E. and M.A. Jakubowska, *Biology of pancreatic stellate cells—more than just pancreatic cancer*. Pflügers Archiv-European Journal of Physiology, 2017. **469**(9): p. 1039-1050.
164. Nieman, K.M., et al., *Adipose tissue and adipocytes support tumorigenesis and metastasis*. Biochimica et Biophysica Acta (BBA)-Molecular and Cell Biology of Lipids, 2013. **1831**(10): p. 1533-1541.
165. Hefetz-Sela, S. and P.E. Scherer, *Adipocytes: impact on tumor growth and potential sites for therapeutic intervention*. Pharmacology & therapeutics, 2013. **138**(2): p. 197-210.
166. Gao, Y., et al., *Adipocytes promote breast tumorigenesis through TAZ-dependent secretion of Resistin*. Proceedings of the National Academy of Sciences, 2020. **117**(52): p. 33295-33304.
167. De Pergola, G. and F. Silvestris, *Obesity as a major risk factor for cancer*. Journal of obesity, 2013. **2013**.
168. Ntikoudi, E., et al., *Hormones of adipose tissue and their biologic role in lung cancer*. Cancer treatment reviews, 2014. **40**(1): p. 22-30.
169. Wang, Y.-Y., et al., *Adipose tissue and breast epithelial cells: a dangerous dynamic duo in breast cancer*. Cancer letters, 2012. **324**(2): p. 142-151.
170. Camarda, R., et al., *Tumor cell-adipocyte gap junctions activate lipolysis and are essential for breast tumorigenesis*. bioRxiv, 2018: p. 277939.
171. Strong, A.L., et al., *Obesity associated alterations in the biology of adipose stem cells mediate enhanced tumorigenesis by estrogen dependent pathways*. Breast Cancer Research, 2013. **15**(5): p. 1-15.
172. El Atat, O., et al., *An evaluation of the stemness, paracrine, and tumorigenic characteristics of highly expanded, minimally passaged adipose-derived stem cells*. PLoS One, 2016. **11**(9): p. e0162332.
173. MacIsaac, Z.M., et al., *Long-term in-vivo tumorigenic assessment of human culture-expanded adipose stromal/stem cells*. Experimental cell research, 2012. **318**(4): p. 416-423.
174. Dzobo, K., *Recent trends in multipotent human mesenchymal stem/stromal cells: Learning from history and advancing clinical applications*. OMICS: A Journal of Integrative Biology, 2021. **25**(6): p. 342-357.
175. Dzobo, K., et al., *Fibroblast-derived extracellular matrix induces chondrogenic differentiation in human adipose-derived mesenchymal stromal/stem cells in vitro*. International journal of molecular sciences, 2016. **17**(8): p. 1259.
176. Kass, L., et al., *Mammary epithelial cell: influence of extracellular matrix composition and organization during development and tumorigenesis*. The international journal of biochemistry & cell biology, 2007. **39**(11): p. 1987-1994.
177. Vieweg, J., et al., *Reversal of tumor-mediated immunosuppression*. Clinical Cancer Research, 2007. **13**(2): p. 727s-732s.
178. Papait, A., et al., *The multifaceted roles of MSCs in the tumor microenvironment: interactions with immune cells and exploitation for therapy*. Frontiers in Cell and Developmental Biology, 2020. **8**: p. 447.
179. Baker, S.G., *The detached pericyte hypothesis: A novel explanation for many puzzling aspects of tumorigenesis*. Organisms. Journal of Biological Sciences, 2018. **2**(1): p. 25-42.
180. Xian, X., et al., *Pericytes limit tumor cell metastasis*. The Journal of clinical investigation, 2006. **116**(3): p. 642-651.
181. Greenberg, J.I., et al., *A role for VEGF as a negative regulator of pericyte function and vessel maturation*. Nature, 2008. **456**(7223): p. 809-813.



182. Viski, C., et al., *Endosialin-expressing pericytes promote metastatic dissemination*. Cancer research, 2016. **76**(18): p. 5313-5325.
183. Raza, A., M.J. Franklin, and A.Z. Dudek, *Pericytes and vessel maturation during tumor angiogenesis and metastasis*. American journal of hematology, 2010. **85**(8): p. 593-598.
184. Cooke, V.G., et al., *Pericyte depletion results in hypoxia-associated epithelial-to-mesenchymal transition and metastasis mediated by met signaling pathway*. Cancer cell, 2012. **21**(1): p. 66-81.
185. Pieterse, Z., D. Sinha, and P. Kaur, *Pericytes in metastasis*. Pericyte Biology in Disease, 2019: p. 125-135.
186. Gerhardt, H. and H. Semb, *Pericytes: gatekeepers in tumour cell metastasis?* Journal of molecular medicine, 2008. **86**(2): p. 135-144.
187. Prazeres, P.H.D.M., et al., *Pericytes are heterogeneous in their origin within the same tissue*. Developmental biology, 2017. **427**(1): p. 6-11.
188. Dzobo, K., V.D. Leaner, and M.I. Parker, *Feedback regulation of the alpha2(1) collagen gene via the Mek-Erk signaling pathway*. IUBMB Life, 2012. **64**(1): p. 87-98.
189. Dzobo, K., V.D. Leaner, and M.I. Parker, *Absence of feedback regulation in the synthesis of COL1A1*. Life Sci, 2014. **103**(1): p. 25-33.
190. Stupack, D.G. and D.A. Cheresh, *ECM remodeling regulates angiogenesis: endothelial integrins look for new ligands*. Science's STKE, 2002. **2002**(119): p. pe7-pe7.
191. Cox, T.R. and J.T. Erler, *Remodeling and homeostasis of the extracellular matrix: implications for fibrotic diseases and cancer*. Dis Model Mech, 2011. **4**(2): p. 165-78.
192. Sirica, A.E. and G.J. Gores, *Desmoplastic stroma and cholangiocarcinoma: clinical implications and therapeutic targeting*. Hepatology (Baltimore, Md.), 2014. **59**(6): p. 2397.
193. Lu, P., V.M. Weaver, and Z. Werb, *The extracellular matrix: a dynamic niche in cancer progression*. J Cell Biol, 2012. **196**(4): p. 395-406.
194. Page-McCaw, A., A.J. Ewald, and Z. Werb, *Matrix metalloproteinases and the regulation of tissue remodelling*. Nat Rev Mol Cell Biol, 2007. **8**(3): p. 221-33.
195. Holle, A.W., J.L. Young, and J.P. Spatz, *In vitro cancer cell-ECM interactions inform in vivo cancer treatment*. Adv Drug Deliv Rev, 2016. **97**: p. 270-9.
196. Morin, P.J., *Drug resistance and the microenvironment: nature and nurture*. Drug Resist Updat, 2003. **6**(4): p. 169-72.
197. Campbell, N.E., et al., *Extracellular Matrix Proteins and Tumor Angiogenesis*. Journal of Oncology, 2010. **2010**: p. 586905.
198. Cox, T.R., *The matrix in cancer*. Nat Rev Cancer, 2021. **21**(4): p. 217-238.
199. Rohwer, N. and T. Cramer, *Hypoxia-mediated drug resistance: novel insights on the functional interaction of HIFs and cell death pathways*. Drug Resist Updat, 2011. **14**(3): p. 191-201.
200. Lv, Y., et al., *Hypoxia-inducible factor-1 $\alpha$  induces multidrug resistance protein in colon cancer*. Onco Targets Ther, 2015. **8**: p. 1941-8.
201. Wang, M., et al., *Role of tumor microenvironment in tumorigenesis*. Journal of Cancer, 2017. **8**(5): p. 761.
202. Franco, P.I.R., et al., *Tumor microenvironment components: Allies of cancer progression*. Pathology-Research and Practice, 2020. **216**(1): p. 152729.
203. Hinshaw, D.C. and L.A. Shevde, *The tumor microenvironment innately modulates cancer progression*. Cancer research, 2019. **79**(18): p. 4557-4566.
204. Arneth, B., *Tumor microenvironment*. Medicina, 2019. **56**(1): p. 15.
205. Roma-Rodrigues, C., et al., *Targeting tumor microenvironment for cancer therapy*. International journal of molecular sciences, 2019. **20**(4): p. 840.
206. Vaahtomeri, K. and K. Alitalo, *Lymphatic Vessels in Tumor Dissemination versus Immunotherapy* Lymphatics in Metastasis vs. Immunotherapy. Cancer research, 2020. **80**(17): p. 3463-3465.

207. Harris, A.L., *Hypoxia—a key regulatory factor in tumour growth*. Nature reviews cancer, 2002. **2**(1): p. 38-47.
208. Brown, J.M. and A.J. Giaccia, *The unique physiology of solid tumors: opportunities (and problems) for cancer therapy*. Cancer research, 1998. **58**(7): p. 1408-1416.
209. Jing, X., et al., *Role of hypoxia in cancer therapy by regulating the tumor microenvironment*. Molecular Cancer, 2019. **18**(1): p. 157.
210. Finger, E.C. and A.J. Giaccia, *Hypoxia, inflammation, and the tumor microenvironment in metastatic disease*. Cancer and Metastasis Reviews, 2010. **29**(2): p. 285-293.
211. Denko, N.C., *Hypoxia, HIF1 and glucose metabolism in the solid tumour*. Nature Reviews Cancer, 2008. **8**(9): p. 705-713.
212. Ke, Q. and M. Costa, *Hypoxia-inducible factor-1 (HIF-1)*. Molecular pharmacology, 2006. **70**(5): p. 1469-1480.
213. Ziello, J.E., I.S. Jovin, and Y. Huang, *Hypoxia-Inducible Factor (HIF)-1 regulatory pathway and its potential for therapeutic intervention in malignancy and ischemia*. The Yale journal of biology and medicine, 2007. **80**(2): p. 51.
214. Hung, J.-J., et al., *Prognostic significance of hypoxia-inducible factor-1 $\alpha$ , TWIST1 and Snail expression in resectable non-small cell lung cancer*. Thorax, 2009. **64**(12): p. 1082-1089.
215. Liu, K., et al., *Hypoxia promotes vasculogenic mimicry formation by the Twist1-Bmi1 connection in hepatocellular carcinoma*. International journal of molecular medicine, 2015. **36**(3): p. 783-791.
216. Gilkes, D.M., et al., *Hypoxia-inducible factor 1 (HIF-1) promotes extracellular matrix remodeling under hypoxic conditions by inducing P4HA1, P4HA2, and PLOD2 expression in fibroblasts*. Journal of Biological Chemistry, 2013. **288**(15): p. 10819-10829.
217. Gilkes, D.M., G.L. Semenza, and D. Wirtz, *Hypoxia and the extracellular matrix: drivers of tumour metastasis*. Nature Reviews Cancer, 2014. **14**(6): p. 430-439.
218. Shieh, A.C., *Biomechanical forces shape the tumor microenvironment*. Annals of biomedical engineering, 2011. **39**(5): p. 1379-1389.
219. Comito, G., et al., *Cancer-associated fibroblasts and M2-polarized macrophages synergize during prostate carcinoma progression*. Oncogene, 2014. **33**(19): p. 2423-2431.
220. Tripathi, C., et al., *Macrophages are recruited to hypoxic tumor areas and acquire a pro-angiogenic M2-polarized phenotype via hypoxic cancer cell derived cytokines Oncostatin M and Eotaxin*. Oncotarget, 2014. **5**(14): p. 5350.
221. Chiu, D.K.-C., et al., *Hypoxia inducible factor HIF-1 promotes myeloid-derived suppressor cells accumulation through ENTPD2/CD39L1 in hepatocellular carcinoma*. Nature communications, 2017. **8**(1): p. 1-12.
222. Švastová, E., et al., *Hypoxia activates the capacity of tumor-associated carbonic anhydrase IX to acidify extracellular pH*. FEBS letters, 2004. **577**(3): p. 439-445.
223. Jing, X., et al., *Role of hypoxia in cancer therapy by regulating the tumor microenvironment*. Molecular cancer, 2019. **18**(1): p. 1-15.
224. du Souich, P. and C. Fradette, *The effect and clinical consequences of hypoxia on cytochrome P450, membrane carrier proteins activity and expression*. Expert opinion on drug metabolism & toxicology, 2011. **7**(9): p. 1083-1100.
225. Park, T.-E., et al., *Hypoxia-enhanced Blood-Brain Barrier Chip recapitulates human barrier function and shuttling of drugs and antibodies*. Nature communications, 2019. **10**(1): p. 1-12.
226. Mazure, N.M. and J. Pouyssegur, *Hypoxia-induced autophagy: cell death or cell survival?* Current opinion in cell biology, 2010. **22**(2): p. 177-180.
227. Brouqui, P., et al., *Asymptomatic hypoxia in COVID-19 is associated with poor outcome*. International Journal of Infectious Diseases, 2021. **102**: p. 233-238.
228. Kashani, K.B. *Hypoxia in COVID-19: sign of severity or cause for poor outcomes*. in Mayo Clinic Proceedings. 2020. Elsevier.
229. Pegtel, D.M. and S.J. Gould, *Exosomes*. Annual review of biochemistry, 2019. **88**: p. 487-514.

230. Kalluri, R., *The biology and function of exosomes in cancer*. The Journal of clinical investigation, 2016. **126**(4): p. 1208-1215.
231. Milman, N., L. Ginini, and Z. Gil, *Exosomes and their role in tumorigenesis and anticancer drug resistance*. Drug Resistance Updates, 2019. **45**: p. 1-12.
232. Melo, S.A., et al., *Cancer exosomes perform cell-independent microRNA biogenesis and promote tumorigenesis*. Cancer cell, 2014. **26**(5): p. 707-721.
233. Su, L.-L., et al., *Exosomes in esophageal cancer: A review on tumorigenesis, diagnosis and therapeutic potential*. World Journal of Clinical Cases, 2019. **7**(8): p. 908.
234. Yu, D.d., et al., *Exosomes in development, metastasis and drug resistance of breast cancer*. Cancer science, 2015. **106**(8): p. 959-964.
235. Najminejad, H., et al., *Emerging roles of exosomal miRNAs in breast cancer drug resistance*. IUBMB life, 2019. **71**(11): p. 1672-1684.
236. Uen, Y., et al., *Mining of potential microRNAs with clinical correlation-regulation of syndecan-1 expression by miR-122-5p altered mobility of breast cancer cells and possible correlation with liver injury*. Oncotarget, 2018. **9**(46): p. 28165.
237. Fong, M.Y., et al., *Breast-cancer-secreted miR-122 reprograms glucose metabolism in premetastatic niche to promote metastasis*. Nature cell biology, 2015. **17**(2): p. 183-194.
238. Jiang, L., et al., *Exosomes: diagnostic biomarkers and therapeutic delivery vehicles for cancer*. Molecular pharmaceutics, 2019. **16**(8): p. 3333-3349.
239. Kim, J.-H., E. Kim, and M.Y. Lee, *Exosomes as diagnostic biomarkers in cancer*. Molecular & Cellular Toxicology, 2018. **14**(2): p. 113-122.
240. An, T., et al., *Exosomes serve as tumour markers for personalized diagnostics owing to their important role in cancer metastasis*. Journal of extracellular vesicles, 2015. **4**(1): p. 27522.
241. Jafari, R., et al., *Hypoxic exosomes orchestrate tumorigenesis: molecular mechanisms and therapeutic implications*. Journal of Translational Medicine, 2020. **18**(1): p. 1-14.
242. Ohnishi, Y., et al., *Heparin-binding epidermal growth factor-like growth factor is a potent regulator of invasion activity in oral squamous cell carcinoma*. Oncology reports, 2012. **27**(4): p. 954-958.
243. Wang, J., Y. Zheng, and M. Zhao, *Exosome-based cancer therapy: implication for targeting cancer stem cells*. Frontiers in pharmacology, 2017. **7**: p. 533.
244. Chitadze, G., et al., *Generation of Soluble NKG 2 D Ligands: Proteolytic Cleavage, Exosome Secretion and Functional Implications*. Scandinavian journal of immunology, 2013. **78**(2): p. 120-129.
245. Lundholm, M., et al., *Prostate tumor-derived exosomes down-regulate NKG2D expression on natural killer cells and CD8+ T cells: mechanism of immune evasion*. PloS one, 2014. **9**(9): p. e108925.
246. Andaloussi, S.E., et al., *Exosomes for targeted siRNA delivery across biological barriers*. Advanced drug delivery reviews, 2013. **65**(3): p. 391-397.
247. Ruzzo, A., et al., *Pharmacogenetic profiling in patients with advanced colorectal cancer treated with first-line FOLFIRI chemotherapy*. The pharmacogenomics journal, 2008. **8**(4): p. 278-288.
248. Harries, M. and M. Gore, *Part I: Chemotherapy for epithelial ovarian cancer—treatment at first diagnosis*. The lancet oncology, 2002. **3**(9): p. 529-536.
249. Flemming, A., *PD1 makes waves in anticancer immunotherapy*. Nature reviews Drug discovery, 2012. **11**(8): p. 601-601.
250. Haanen, J.B. and C. Robert, *Immune checkpoint inhibitors*. Immuno-Oncology, 2015. **42**: p. 55-66.
251. Jenkins, R.W., D.A. Barbie, and K.T. Flaherty, *Mechanisms of resistance to immune checkpoint inhibitors*. British journal of cancer, 2018. **118**(1): p. 9-16.
252. Grywalska, E., et al., *Immune-checkpoint inhibitors for combating T-cell dysfunction in cancer*. OncoTargets and therapy, 2018. **11**: p. 6505.



253. Sharma, P. and J.P. Allison, *The future of immune checkpoint therapy*. Science, 2015. **348**(6230): p. 56-61.
254. Xue, L.J., et al., *Inhibition of CXCL12/CXCR4 axis as a potential targeted therapy of advanced gastric carcinoma*. Cancer medicine, 2017. **6**(6): p. 1424-1436.
255. Bule, P., et al., *Chemokine-directed tumor microenvironment modulation in cancer immunotherapy*. International Journal of Molecular Sciences, 2021. **22**(18): p. 9804.
256. Perdih, A. and M. Sollner Dolenc, *Small molecule antagonists of integrin receptors*. Current medicinal chemistry, 2010. **17**(22): p. 2371-2392.
257. Unruh, A., et al., *The hypoxia-inducible factor-1 $\alpha$  is a negative factor for tumor therapy*. Oncogene, 2003. **22**(21): p. 3213-3220.
258. Rohwer, N., et al., *Hypoxia-inducible factor 1 $\alpha$  determines gastric cancer chemosensitivity via modulation of p53 and NF- $\kappa$ B*. PloS one, 2010. **5**(8): p. e12038.
259. Chen, Q., et al., *Intelligent albumin-MnO<sub>2</sub> nanoparticles as pH-/H<sub>2</sub>O<sub>2</sub>-responsive dissociable nanocarriers to modulate tumor hypoxia for effective combination therapy*. Advanced materials, 2016. **28**(33): p. 7129-7136.
260. Saggar, J.K. and I.F. Tannock, *Activity of the hypoxia-activated pro-drug TH-302 in hypoxic and perivascular regions of solid tumors and its potential to enhance therapeutic effects of chemotherapy*. International journal of cancer, 2014. **134**(11): p. 2726-2734.
261. Hajj, C., et al., *A combination of radiation and the hypoxia-activated prodrug evofosfamide (TH-302) is efficacious against a human orthotopic pancreatic tumor model*. Translational oncology, 2017. **10**(5): p. 760-765.
262. Wouters, B.G. and M. Koritzinsky, *Hypoxia signalling through mTOR and the unfolded protein response in cancer*. Nature Reviews Cancer, 2008. **8**(11): p. 851-864.
263. Feldman, D.E., V. Chauhan, and A.C. Koong, *The unfolded protein response: a novel component of the hypoxic stress response in tumors*. Molecular Cancer Research, 2005. **3**(11): p. 597-605.