Article

Altered cortical palmitoylation induces widespread molecular disturbances in Parkinson´s disease

Juan F. Cervilla-Martinez[1]*, Juan J. Rodríguez-Gotor[1,2]*, Krzysztof J. Wypijewski[3,4], Ángela Fontán-Lozano[1,5], Tao Wang[6], Enrique Santamaría[7], William Fuller[3], Rebeca Mejías[1,5#]

1   Department of Physiology, School of Biology, University of Seville, Avenida de la Reina Mercedes, 6, 41012 Sevilla, Spain (J.F.C.M.: juacermar1@alum.us.es; J.J.R.G.: juanjogotor@gmail.com; A.F.L.: afontan@us.es; R.M: rmejias@us.es).

2   Instituto de Neurociencias CSIC-UMH, Avenida Santiago Ramón y Cajal s/n, San Juan de Alicante, 03550 Alicante, Spain.

3   Institute of Cardiovascular and Medical Sciences, University of Glasgow, VPF4+76 Glasgow, UK (W.F.: Will.Fuller@glasgow.ac.uk).

4   School of Life Sciences, University of Dundee, Invergowrie, Dundee, DD2 5DA, Scotland, UK (K.J.W.: KWypijewski001@dundee.ac.uk).

5   Instituto de Biomedicina de Sevilla, Campus Hospital Universitario Virgen del Rocío, Avda. Manuel Siurot, s/n, 41013 Sevilla, Spain.

6   McKusick-Nathans Institute of Genetic Medicine and Department of Pediatrics, Johns Hopkins University School of Medicine, Baltimore, MD, 21205, USA (twang9@jhmi.edu).

7   Clinical Neuroproteomics Unit, Navarrabiomed, Hospital Universitario de Navarra (HUN), Universidad Pública de Navarra (UPNA), IDISNA, Irunlarrea Street, 3, 31008, Pamplona, Navarra, Spain (E.S: esantamma@navarra.es).

* These authors contributed equally to this work.

\# Correspondence: Rebeca Mejías (rmejias@us.es); Department of Physiology, School of Biology, University of Seville, Avenida de la Reina Mercedes, 6, 41012 Sevilla, Spain; Phone: +34-954-559-549.

Abstract: The relationship between Parkinson's disease (PD), the second most common neurodegenerative disease after Alzheimer's disease, and palmitoylation, a post-translational lipid modification, is not well understood. In this study, we have analyzed the differential palmitome present in the cerebral cortex of PD patients compared to controls (n=4 per group) to better understand the role of protein palmitoylation in PD and the pathways altered in this disease. Data-mining of the cortical palmitome from PD patients and controls has allowed to: i) detect a set of 150 proteins with altered palmitoylation in PD subjects in comparison with controls, ii) describe the biological pathways and targets predicted to be altered by these palmitoylation changes, and iii) depict the overlap between the differential palmitome identified in our study with protein interactomes of the PD-linked proteins α-synuclein, LRRK2, DJ-1, PINK1, GBA and UCHL1. In summary, we have partially characterized the altered palmitome in the cortex of PD patients which is predicted to impact cytoskeleton, mitochondrial and fibrinogen functions, as well as cell survival. Our study points out that protein palmitoylation could have a role in the pathophysiology of PD, and that comprehensive palmitoyl-proteomics offers a powerful approach for elucidating novel cellular pathways modulated in this neurodegenerative disease.

Keywords: Parkinson´s disease; proteomics; cerebral cortex; palmitoylation; interactome.

1. Introduction

PD is the second most common neurodegenerative disease, and its pathological hallmarks are the loss of dopaminergic neurons in the substantia nigra pars compacta (SNc) and the presence of Lewy bodies (LBs), insoluble intracytoplasmic neuronal inclusions containing misfolded α-synuclein. The loss of dopaminergic neurons in the SNc leads to a depletion of dopamine in the striatum which in turns is responsible for the cardinal motor symptoms of PD [1]. As the disease evolves, neurodegeneration spreads to the rest of the central nervous system, including the cerebral cortex [2]. In addition to motor disturbances, non-motor symptoms that profoundly affect the quality of life of PD patients, such as REM sleep disturbances, mood, constipation, and smell loss, can appear even before the development of motor symptoms [3]. Although the exact causes of PD are unknown, in 5%-15% of cases a mutation in a PARK gene has been identified, and some environmental factors, such as certain pesticides, have also been linked to the risk of PD [1]. The characterization of the protein products encoded by PD-linked genes has provided important insights into the pathophysiology of familial and sporadic PD. Thus, alterations in mitochondrial function, vesicular transport, protein degradation, and oxidative balance, among other cellular functions, have been postulated as pathways involved in the neurodegeneration associated with PD [4]. At present there is no cure for PD and treatments are symptomatic, aimed to improve motor and non-motor symptoms [5].

S-acylation (called palmitoylation for simplicity through the manuscript) is a post-translational modification in which a palmitate molecule attaches reversibly to a cysteine residue of a protein via a thioester bond. Palmitoylation is exquisitely regulated by enzymes which add palmitate (palmitoyl acyltransferases, PATs) or remove it (acyl protein thioesterases) from proteins [6]. Twenty-three PATs have been identified in mammals to date, but the knowledge about depalmitoylating enzymes remains very restricted [7,8]. Protein palmitoylation can regulate trafficking, stability, and enzymatic activity of proteins, and dysregulation of this lipid modification has been implicated in several human diseases including cancers, metabolic syndrome, and infections [9,10]. More specifically, in the nervous system, several studies have linked alterations in palmitoylation to schizophrenia, Alzheimer´s disease (AD), intellectual disability, neuronal ceroid lipofuscinosis, and Huntington's disease [7,11–17]. It is well-known that multiple neural signaling proteins, ion channels, cell adhesion molecules, SNARE proteins, vesicular trafficking proteins, and G-protein receptors are susceptible to be palmitoylated [18,19]. Moreover, this lipid modification has been clearly associated to neuronal development and synaptic plasticity [8,20,21]. However, a possible link between palmitoylation/depalmitoylation defects and PD has not been studied *in vivo*, and our limited understanding of this relationship comes from *in vitro* and *in silico* studies. Thus, palmitoylation of the dopamine transporter (DAT) and DJ-1, encoded by the PARK7 gene, have been described, and palmitoylation of LRRK2, encoded by the PARK8 gene, has been predicted [22–24]. In addition, it has been recently reported that upregulation of cellular palmitoylation alleviates the accumulation and neurotoxicity induced by α-synuclein [25].

During the last years, neuroproteomics has emerged as a powerful approach to characterize multiple proteotypes associated to the neurodegenerative process [26,27]. Due to the lack of information about the impact of palmitoylation during the progression of the neurodegenerative process associated to PD, we studied the cortical palmitoyl-proteome (palmitome) of PD and control subjects. For this purpose, after isolating palmitoylated proteins from cortical human samples using the Acyl-Rac assay, quantitative proteomics was performed to detect, identify, and quantify palmitoylated proteins levels in both groups. In this pilot study, we have characterised a set of 84 proteins that show altered levels of palmitoylation in the cerebral cortex of PD patients *vs.* controls. Interestingly, around 30% of these proteins were not previously known to be palmitoylated. These post-translational alterations in protein palmitoylation might have an impact on the transport associated to microtubules, inflammation, oxidative stress, and tau pathology, among other physiological effects. Moreover, some of the proteins with altered palmitoylation levels in PD patients belong to the interactome of proteins encoded by most relevant PARK genes, suggesting that alterations in palmitoylation could contribute to pathological mechanisms causing PD. Additionally, thanks to public available RNA-seq datasets, we show that the differentially palmitoylated proteins tend to map in different cell types in the cerebral cortex, but mainly in neurons. Our results demonstrate, for the first time in human brain tissue, a global protein palmitoylation deregulation in the cortex of PD patients. We consider that the deployment of brain palmitoyl-proteomics is an ideal approach that allows zooming-in to explore the potential imbalance in palmitoylation-dependent regulatory mechanisms associated to this disease.

2. Results and Discussion

2.1 Altered palmitoylation levels of cortical proteins in PD

To characterize the palmitoyl-proteome (palmitome) signature in the brain of PD patients, we performed label free LC-MS/MS-based comparative proteomics [28][29] on cortical tissue derived from PD subjects and controls with no history of neurological disorders (Supplementary Table 1), after isolating palmitoylated proteins using the Acyl-RAC assay. Among 2284 putatively palmitoylated proteins identified in this proteomics study (Supplementary Table 2), 84 proteins had statistically significant differences in their levels when comparing PD patients and control subjects (4% of the quantified palmitome; Log2(FC): ≥[0.6]; p-value: ≤0.05) (Supplementary Table 3). We will call these 84 proteins differential palmitoylated proteins (DPPs) henceforth. We consecutively compared our results (set of all palmitoylated proteins identified in human cortex and DPPs) with the database on S-palmitoylation SwissPalm [30]. SwissPalm contains information about the palmitoylation of all annotate proteins in *H. sapiens*, more specifically, whether a protein is known to be palmitoylated and the type of experiments that suggest the lipid modification (proteomics studies and/or further validation experiments) or not known to be palmitoylated. In general, around 20% of all proteins annotated in *Homo sapiens* have been identified as palmitoylated according to SwissPalm database (Figure 1A). Focusing on the 2284 proteins identified in our cortical palmitome study (all palmitoylated proteins identified in human cortex), more than 50% have been described to be palmitoylated according to the SwissPalm database (Figure 1B). Almost 70% of the DPPs are palmitoylated according to the same database (Figure 1C). The increase in the percentage of palmitoylated proteins in the pie charts from Figures 1B and 1C regarding 1A, confirms that the isolation of palmitoylated proteins before the proteomics analysis was successful. Besides, these observations also point out that our proteomics experiment might have identified novel palmitoylated proteins that were not known to carry this lipid modification, although additional targeted studies are needed to validate our data. It would be interesting to compare the palmitoylation profile across different brain regions in PD patients in the future to provide a better understanding of the role of this post-translational modification in the origin and progression of this disease as well as its correlation with
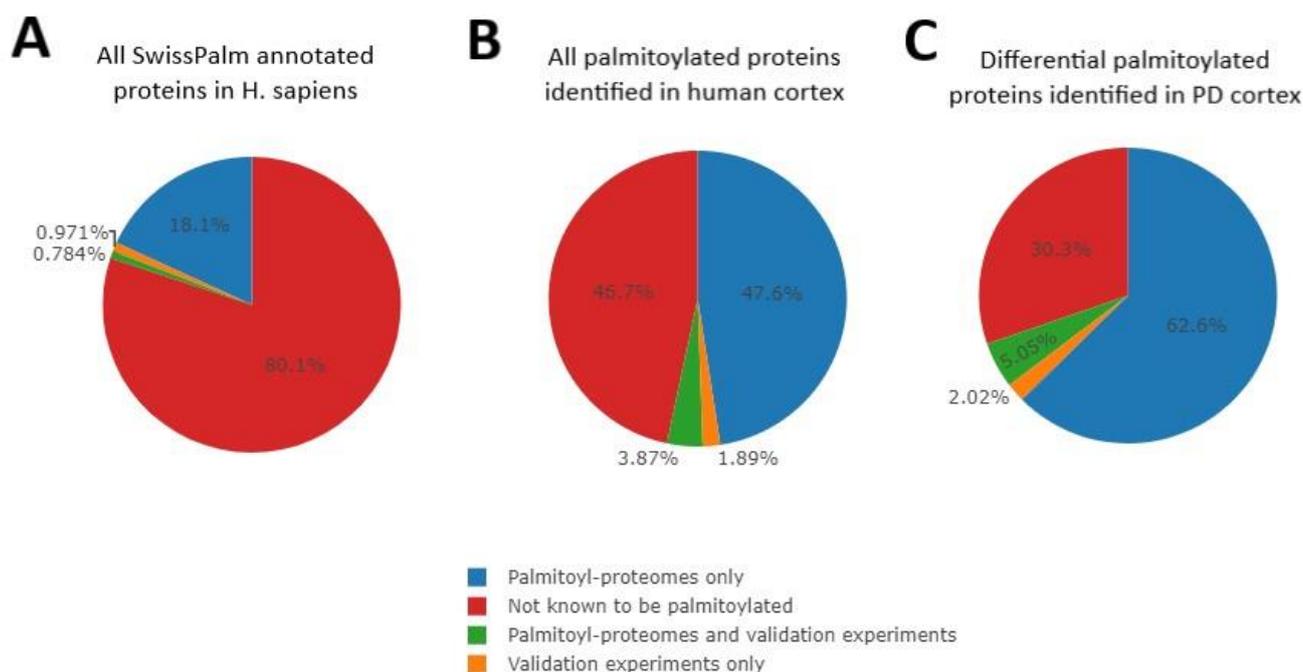
the neuropathological grading. It is worth mentioning that global changes in protein palmitoylation were also found in patients with the neurodegenerative disorder neuronal ceroid lipofuscinosis [31].

Figure 1. Pie charts representing the percentage of proteins per category (palmitoylated with different levels of validation, in blue, green or orange, or non-known to be palmitoylated, in red) included in the SwissPalm database for *Homo sapiens* (A), for all 2284 palmitoylated proteins detected in our workflow study (B), and for the 84 differentially palmitoylated proteins in PD patients identified (C).

From the 84 DPPs identified in this work, 33 and 51 proteins were up-palmitoylated and down-palmitoylated respectively with regards to the control group (Figure 2 and Supplementary table 3).
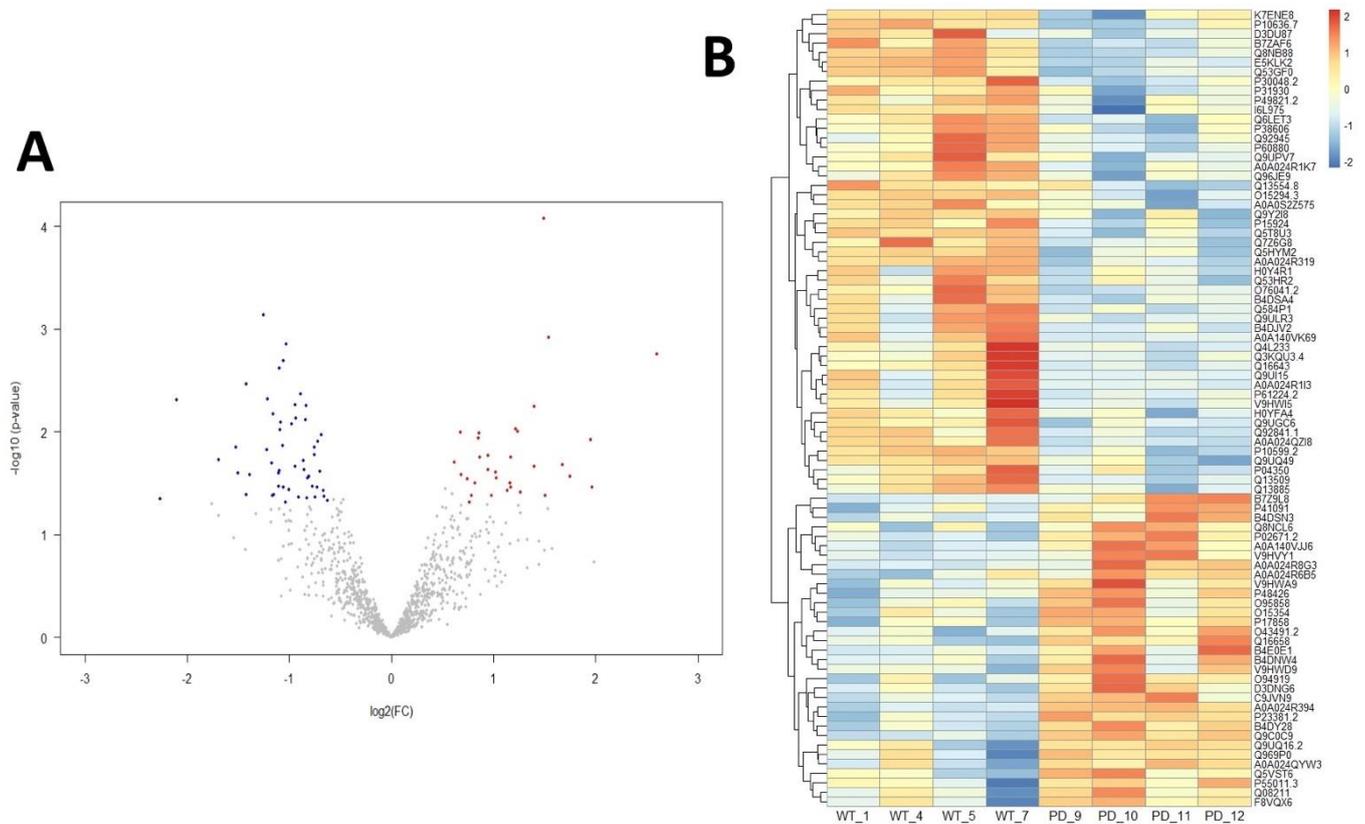

Figure 2. (A) Volcano plot showing up-palmitoylated (red) and down-palmitoylated proteins (blue) detected in cerebral cortex from PD patients. Proteins with no significant changes among groups are represented in grey. (B) Heatmap showing differential palmitoylated proteins arranged by Z-score. The UniProt accession number for each protein is shown.

In order to understand the biological and functional impact of the alterations in cortical palmitoylation levels in PD subjects, we performed GO (Gene Ontology) enrichment analysis [32] (summarized in Supplementary Table 4; adjusted p-value: ≤0.05). Figure 3A shows a heatplot with the 15 most relevant GO terms obtained in this analysis. In Figure 3B we can observe the DPPs (represented by their gene symbols) that are included within these GO terms. Figure 3C displays a cnet plot to visualize the linkages of the DPPs (represented by their gene symbols) shown in Figure 3B, with the most significantly altered biofunctions in PD. In summary, as shown in Figure 3, key palmitoylated proteins are related to coagulation (*FGA*, *FGB*, and *FGG*), dendritic spine, postsynapse, and neuron projection organization (*ABHD17B*, *CAMK2B*, *CFL1*, *DBN1*, *DNM3*, *MAP6*, and *YWHAH*), and cell-cell adhesion (*CSRP1* and *DSP*). Interestingly, ABDH17 proteins have depalmitoylating activity but its palmitoylation has not been reported to date [33]. We observed an increase in the palmitoylation levels of the three polypeptides that constitute the fibrinogen molecule (encoded by genes *FGA*, *FGB*, and *FGG*) in our PD sample cohort. Fibrinogen has essential roles in coagulation, inflammation, extracellular matrix physiology, and tissue repair, having chaperone-like activity [34–37]. Fibrinogen is undetectable in the healthy brain, but it can deposit abundantly after blood-brain barrier (BBB) disruption in a variety of neurological diseases, suggesting a potential pathological role [38–41]. Fibrinogen deposition has not been explored in the brain of PD patients, although disruption of the BBB and inflammation may be involved in disease onset [42,43]. Besides, alterations in normal clotting of blood are not well-described in PD patients, although Sato *et al*. reported a possible relationship between the use of antiparkinsonian drugs and coagulation abnormalities [44]. Studies about fibrinogen levels in plasma and cerebrospinal fluid (CSF) in PD patients have led to conflicting results. Chen *et al*. reported no relationship between

plasma levels of fibrinogen and the risk of developing PD [45], while other reports found high fibrinogen levels in plasma and CSF associated with PD [46–48]. Our data support that PD induces an up-regulation in the levels of palmitoylated fibrinogen polypeptides, at least at cortical level in the brain. Interestingly, palmitoylation of fibrinogen subunits has not been described to date, and consequences of this lipid modification in the functioning of the fibrinogen complex are unknown. Therefore, further experiments are needed to validate fibrinogen palmitoylation and deeply characterize its role in the onset and progression of PD.
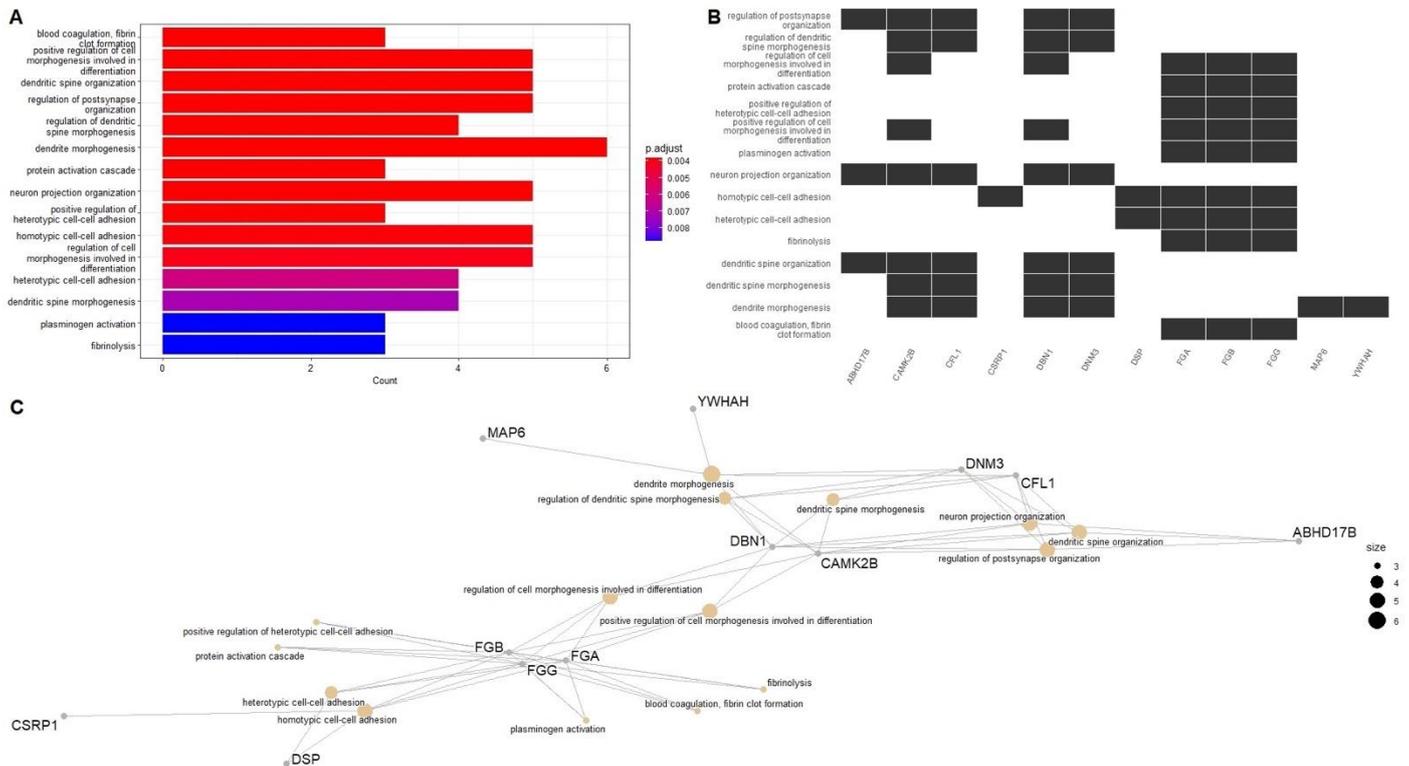

Figure 3. (A) Heatplot showing the 15 most significant terms obtained by performing GO enrichment analysis. (B) Barplot showing the DPPs (represented by their gene symbols) and their relationship with the 15 most relevant GO terms. (A) Cnet plot of significantly enriched GO terms.

To further study pathways affected by the alterations in the DPPs in PD patients and complementing the results obtained by the GO enrichment analysis, we performed Metascape analysis (results summarized in Supplementary Table 5; p-values are provided in logarithmic based ten: "LogP"; p-value: ≤0.05). Metascape is a resource that incorporates a core set of default ontologies for enrichment analysis, including GO processes (GO), KEGG pathways (hsa), Reactome gene sets (R-HSA), and WikiPathways (WP) [49]. In our case, we did not select GO analysis, as it was done previously in our analysis. Interestingly, as shown in Figure 4, Parkinson´s disease pathway (hsa05012 term) was the most affected route according to KEGG Pathway Database. More specifically, nine of the DPPs are involved in PD-related molecular pathways related to PD according to KEGG: *CAMK2B*- Calcium/Calmodulin Dependent Protein Kinase II Beta, *GPR37*- G Protein-Coupled Receptor 37, *MAPT*- Microtubule Associated Protein Tau, *NDUFV1*- NADH: Ubiquinone Oxidoreductase Core Subunit V1 (in mitochondrial Complex I), *TUBB2A*- Tubulin Beta 2A Class IIa, *TXN*- Thioredoxin, *UQCRC1*- Ubiquinol-Cytochrome C Reductase Core Protein 1 (in mitochondrial complex III), *TUBB3*- Tubulin Beta 3 Class III, and *TUBB4A*- Tubulin Beta 4A Class IVa (Gene Symbols- Protein coding). A visualization of the PD pathway can be seen at https://www.kegg.jp/kegg-bin/show_pathway?hsa05012/816/2861/4137/4723/7280/7295/7384/10381/10382 with altered DPPs proteins (in a red rectangle and in red font colour) and their interactions with proteins encoded by PD-associated genes DJ-1, PaelR, Parkin, PINK1, SNCA, and UCHL1 (in a black rectangle and in red font colour). It is worth to point out that disfunction of mitochondrial complex I has been recently linked to the loss of dopaminergic neurons and metabolic changes in the substantia nigra of mice, causing a human-like parkinsonism [50], and that low activity of mitochondrial complexes I , II, and III has been reported in lymphocytes and platelets of PD patients [51,52]. Thioredoxin has antioxidant activity that protects

dopaminergic neurons in animal models of PD [53], and its palmitoylation has been shown to modulate its activity [54,55]. The role of Tubulin Beta in PD and the effects of palmitoylation in its function are not well defined, although its degradation seems to be regulated by Parkin [56] and its palmitoylation has been suggested [57]. Metascape analysis also predicted changes in MAPK signaling for integrins (R-HSA-354194; *FGA, FGB, FGG, RAP1B*) and cellular responses to stimuli (R-HSA-8953897; *ATP6V1A, CAMK2B, CSRP1, EIF2S3, HSPA2, RPL7A, TKT, TUBB2A, TXN, KHSRP, TUBB3, TUBB4A, PRDX3*), among other pathways. Taken together, the analyses of our palmitome study in PD cortex highlights both well-known and not-well characterized molecular pathways that could be affected in this disease.
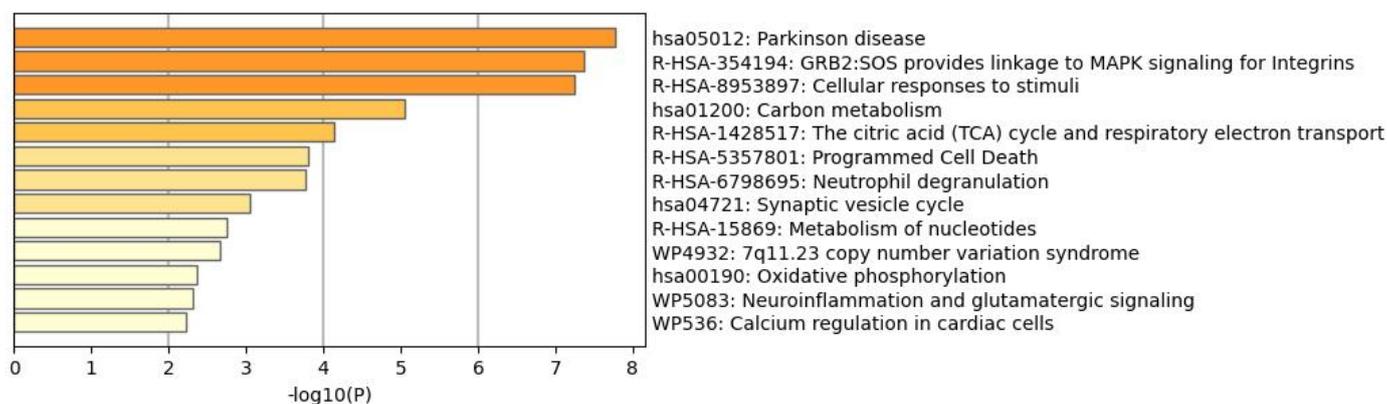

Figure 4. Barplot from Metascape analysis showing the 13 most relevant and non-redundant biofunctions affected by the palmitoylation changes in the DPPs detected at cortical level in PD.

2.2 Network-driven palmitoyl-proteomics reveals an imbalance in cytoskeletal architecture and mitochondrial homeostasis in PD

To enhance the biological outcome of our palmitome approach, proteome-scale interaction networks were constructed using STRING (p≤0.05) [58] and QIAGEN Ingenuity Pathway Analysis (IPA). The outcome obtained by the STRING analysis of DPPs can be seen at https://version-11-5.string-db.org/cgi/network?taskId=bx7tBdv4gtLt&sessionId=bAXrCBiw5ZOI. Figure 5A shows functional and physical protein-protein interaction (PPI) network of DPPs (represented by their gene symbols) and Figure 5B displays just the physical PPI network of DPPs (represented by their gene symbols) obtained using the STRING tool. In this analysis, one of the affected molecular functions involved constituents of the cytoskeleton. Tubulin beta polypeptides (encoded by *TUBB2A, TUBB3* and *TUBB4A*), Microtubule Associated Protein Tau (encoded by *MAPT*), and Calcium/Calmodulin Dependent Protein Kinase II Beta (encoded by *CAMK2B*) are found in the same PPI network with lower levels of palmitoylation in PD patients compared to the controls (Figure 5A). Tubulin alpha and beta heterodimers are structural components of the microtubules and have key roles on multiple cellular functions such as the intracellular molecular trafficking [59]. Microtubule Associated Protein Tau, usually named Tau for simplicity, is responsible for stabilizing the microtubules and its pathological deposition is known as tauopathy. In tauopathies, the soluble tau protein separates from microtubules and creates abnormal, fibrillar structures of aggregated, hyperphosphorylated, and acetylated tau [60,61] Some tauopathies are characterized by parkinsonism, but PD was not initially considered to be a typical tauopathy. However, recent studies have demonstrated increasing evidence of tau pathology in PD. Interestingly, hyperphosphorylated tau proteins have been shown to interact with α-synuclein promoting aggregation and fibrilization to each other, accelerating the formation of Lewy bodies and axonal transport dysfunction [62–65]. Our studies support the existence of tau pathology in the cortex of PD patients and that tau could be susceptible to be palmitoylated *in vivo*. Calcium/Calmodulin Dependent Protein Kinase II Beta (CAMKIIβ), whose possible palmitoylation has not been studied, can associate with the actin cytoskeleton (microfilaments) and it has a role in regulating dendritic arborization and synapse density [66,67]. Interestingly, cytoskeleton disfunctions in PD have been suggested in the literature [68–70], and it would be noteworthy to unravel the consequences of altered palmitoylation in the mentioned proteins on the functioning of this crucial cellular structure.

In accordance with the data obtained from Metascape (MAPK signaling for integrins, R-HSA-354194; Figure 4 and Supplementary Table 5), STRING analysis also predicted a functional and physical PPI network including the fibrinogen components and Ras-Related Protein Rap-1b (encoded by *RAP1B* gene) (Figures 5A and 5B). Furthermore, as shown previously, GO enrichment analysis predicted a biological impact of the alterations in palmitoylation observed in PD patients on the fibrinogen molecule (Figure 3). Modulation of the binding of fibrinogen to integrins by Rap-1b has been

studied in platelets aggregation and megakaryocytes [71,72], but no information is available related to the interactions of these proteins in the brain.
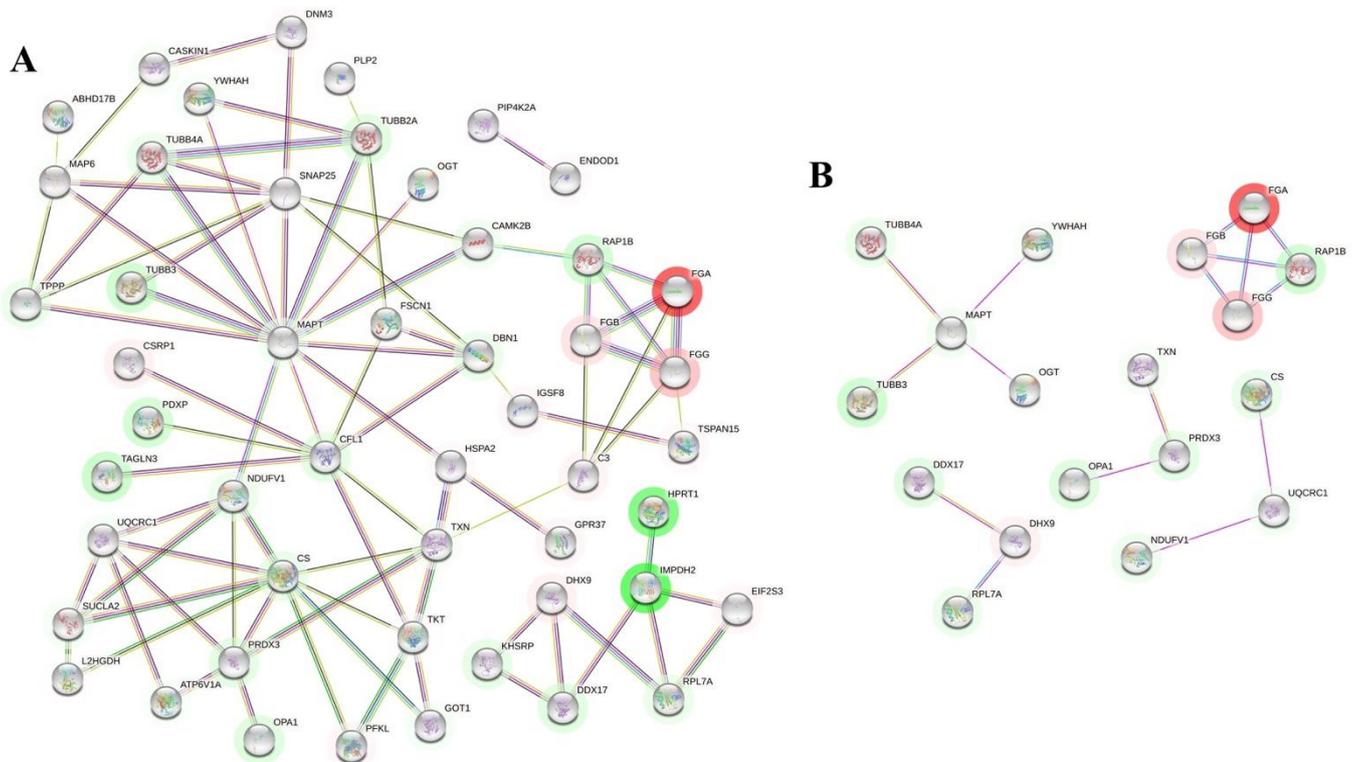

Figure 5. (A) Functional and physical protein-protein interaction (PPI) network of DPPs (represented by their gene symbols) obtained with STRING analysis. (B) Only physical PPI network of DPPs (represented by their gene symbols) obtained with the STRING tool are shown. Green and red halos represent down- or up-regulation of the palmitoylation levels, respectively. Color intensity of the halo is in accordance with the fold-change.

To further study the functional connection of our proteomics data with signaling hubs, we used IPA. To enrich our proteomic dataset, we decided to include the qualitative palmitoylated proteins (QPPs, 66 proteins listed in Supplementary Table 6) to the 84 DPPs in this analysis (total input dataset of 150 proteins). QPPs are proteins whose palmitoylation is almost undetectable in the cortex of one of the groups, either the control or the PD group, but detectable in the other one, and therefore the differences in palmitoylation levels are quite striking but cannot be quantified. As shown in Figure 6, protein network analysis revealed that the deregulated cortical palmitome of PD patients is functionally associated to survival mediators such as Protein Kinase B (Akt) and Protein phosphatase 2 (PP2A). Both, Akt and PP2 have been involved in a variety of neurodegenerative disorders, including PD. Specifically, Akt has been reported to promote neuronal survival [73] and PP2A to modulate mitochondrial function [74,75]. Although palmitome changes were predicted to inhibit AKT (Figure 6), additional research is needed to characterize the potential crosstalk between cellular palmitoylation and survival mediators in the context of PD.
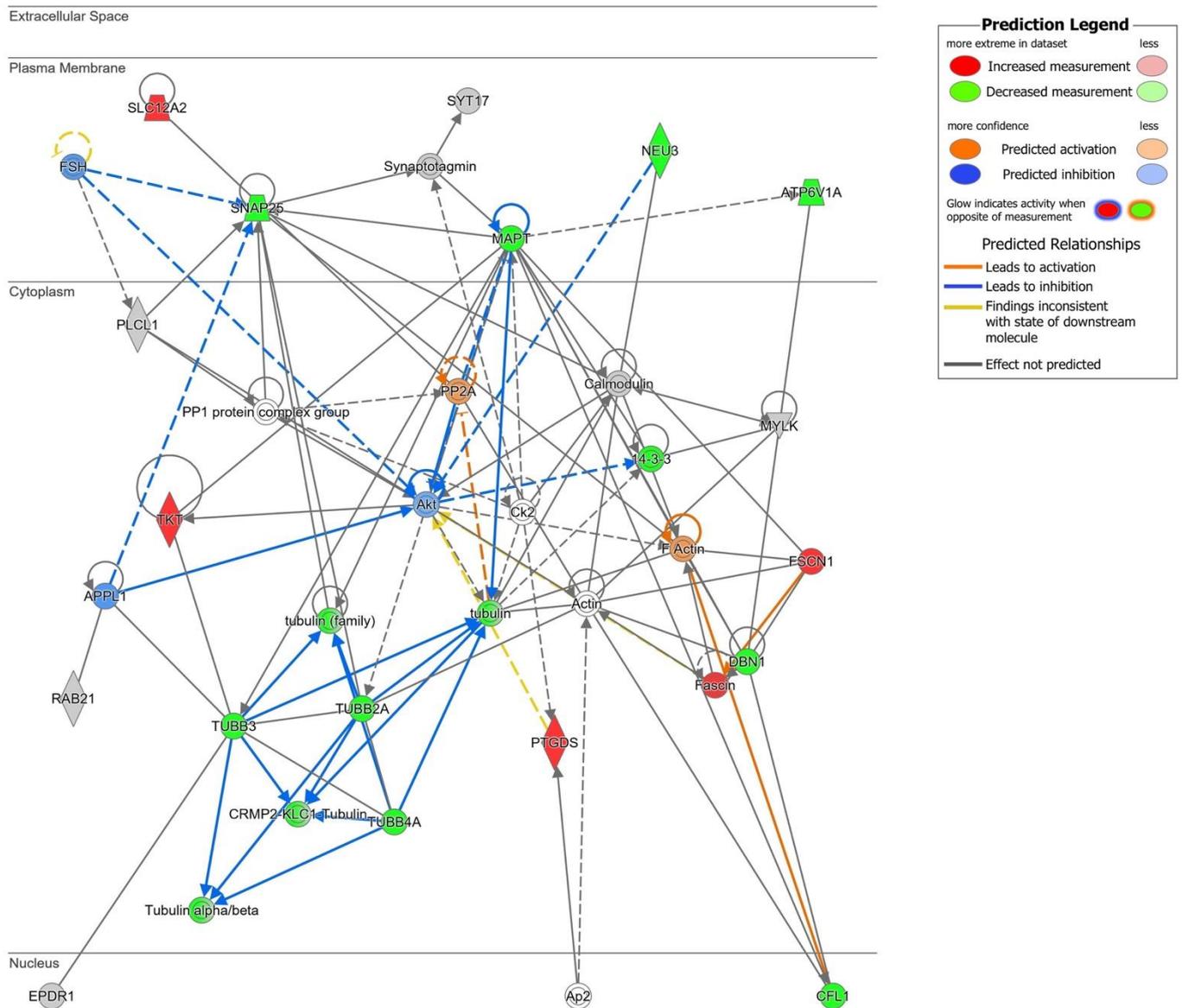
Figure 6. High-scoring protein interactome map focused on cell survival (Score 38). Visual representation of the relationships between DPPs and QPPs with functional interactors. DPPs are highlighted in red when palmitoylation is up-regulated, and in green when palmitoylation is down-regulated in PD cortex. Dataset molecules that did not pass the analyses cutoffs are shown with a grey background. Continuous and discontinuous lines represent direct and indirect interactions respectively. Predicted interactors of DPPs and QPPs, not part of the dataset, are shown in white. See complete QIAGEN IPA legend at https://qiagen.my.salesforce-sites.com/KnowledgeBase/articles/Knowledge/Legend.

Moreover, functional interactome data also recognized a protein set and their PPI interactions that affect mitochondrial homeostasis, specifically the mitochondrial complex I functionality in the cortex of PD subjects (Figure 7). In accordance with this information, Metascape analyses also pointed a mitochondrial disruption linked to dyshomeostatic palmitoylation (Parkinson´s disease pathway, hsa05012 term, Ubiquinone Oxidoreductase Core Subunit V1 and Ubiquinol-Cytochrome C Reductase Core Protein 1). Peroxiredoxin 3 (PRDX3) is associated to the mitochondria, where it exerts its antioxidant role [76]. Both altered mitochondrial complexes activity and oxidative stress has been described as molecular mechanisms underlying the pathophysiology of PD [50–52,77]. On the other hand, O-Linked N-Acetylglucosamine (GlcNAc) Transferase (OGT), catalyzes the post-translational modification of proteins named O-GlcNAcylation, which modulates protein function. Inhibition of OGT might be neuroprotective against AD, and tau protein can be O-GlcNAcylated, at least in animal models [78–80]. GlcNAcylation on α-synuclein has also been established, which has

an inhibitory effect on fibril formation, neurotoxicity, and aggregation [81,82]. Although there is no information about palmitoylation of OGT in the literature, palmitoylation of PRDX3 has been described in mice [83]. According to IPA knowledgebase, PRDX3 and OGT are functionally linked to NF-κβ, a key regulator of inflammation and apoptosis whose dysregulations could be involved in the onset of PD [84,85]. Thus, NF-κβ activation in the brain of PD patients could contribute to chronic neuroinflammation, and its inhibition has been proposed to slow down the progression of this disease [86,87]. Accordingly, IPA predicts a potential activation of NF-κβ in the cortex of PD patients. NF-κβ is not-known to be palmitoylated. In summary, network data from STRING and IPA analyses indicate that cytoskeletal-mito-chondrial axis could be modulated, at least in part, by levels of protein palmitoylation in the cortex of PD patients. Some studies have pointed to a relationship between cytoskeleton alterations and mitochondrial dysfunction in PD [88,89].
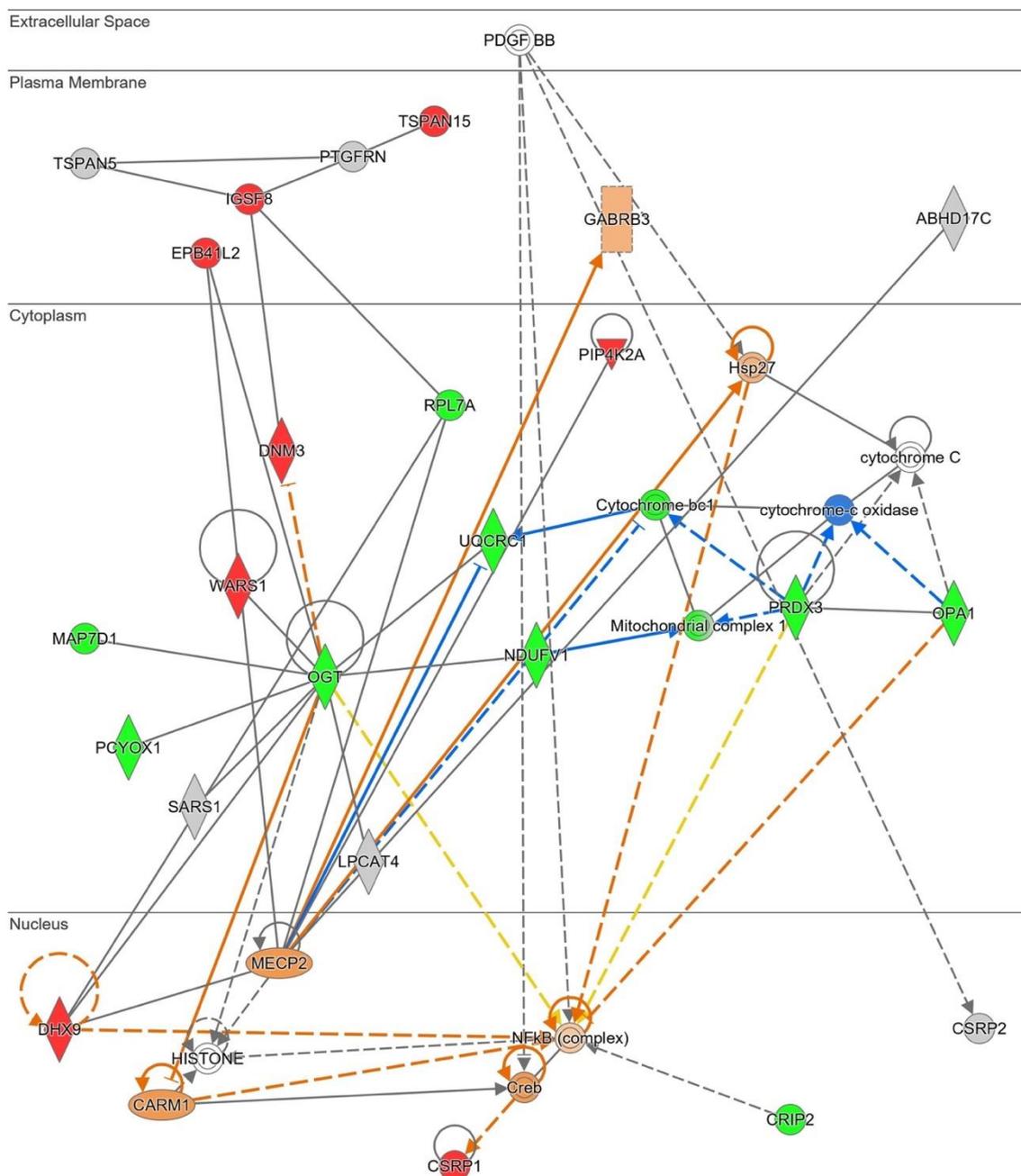

Figure 7. High-scoring protein interactome map unveiling a mitochondrial imbalance (Score 57). Visual representation of the relationships between DPPs and QPPs with functional interactors. DPPs are highlighted in red when pal-mitoylation is up-regulated, and in green when palmitoylation is down-regulated in cortex of PD patients. Dataset

molecules that did not pass the analyses cutoffs are shown with a grey background. Continuous and discontinuous lines represent direct and indirect interactions respectively. Predicted interactors of DPPs and QPPs, not part of the dataset, are shown in white. See complete QIAGEN IPA legend at https://qiagen.my.salesforce-sites.com/Knowledge-Base/articles/Knowledge/Legend.

2.3 Differential palmitoyl-proteins are components of interactomes associated to PD genes and are located in multiple cell types

Discovery of unexpected relationships between apparently unrelated and PD-linked proteins may be a powerful strategy for the characterization of novel PD targets and pathways. We explored whether the interactome of well-established PD-related proteins were indeed potentially interconnected with the DPPs detected at cortical level. As shown in Figure 8 and Supplementary Table 7, we identified several DPPs that are known interactors of proteins encoded by PD-linked genes (SNCA, α-synuclein; LRRK2, leucine-rich repeat kinase 2; GBA, glucocerebrosidase; UCHL1, ubiquitin C-terminal hydrolase L1; PINK1, PTEN-induced kinase 1). Strikingly, we found that PRDX3 (Peroxiredoxin 3) is a common interactor of α-synuclein, LRRK2 and DJ-1 (Figure 8A and Supplementary Table 7). As discussed earlier, PRDX3 has an antioxidant role in the mitochondria and it is known to be palmitoylated in mice [76][83]. PRDX3 is phosphorylated by LRRK2 [90] and its overexpression reduces the neurotoxicity associated to LRRK2 kinase mutant G2019S in flies [91]. We consider of interest to study whether PRDX3 is palmitoylated in human samples, and the influence of this lipid modification on the antioxidant activity of this enzyme.

Secondly, we found that there are 17 DPPs (listed in Supplementary Table 7) which are common interactors of GBA, UCHL1 and PINK1 (Figure 8B). Remarkably, three DPPs: CFL1 (cofilin 1), TKT (transketolase) and DBN1 (Drebrin 1), are shared interactors of most PD-linked proteins studied (LRRK2, DJ-1, PINK1, GBA and UCHL1) (Figure 8C and Supplementary Table 7). Cofilin 1 is an actin-binding protein that can regulate mitochondrial function [92]. Drebrin 1 is also a cytoplasmic actin-binding protein [93], and transketolase is a key enzyme in the non-oxidative branch of the pentose phosphate pathway [94]. However, the precise mechanism by which these enzymes are involved in PD are unknown.
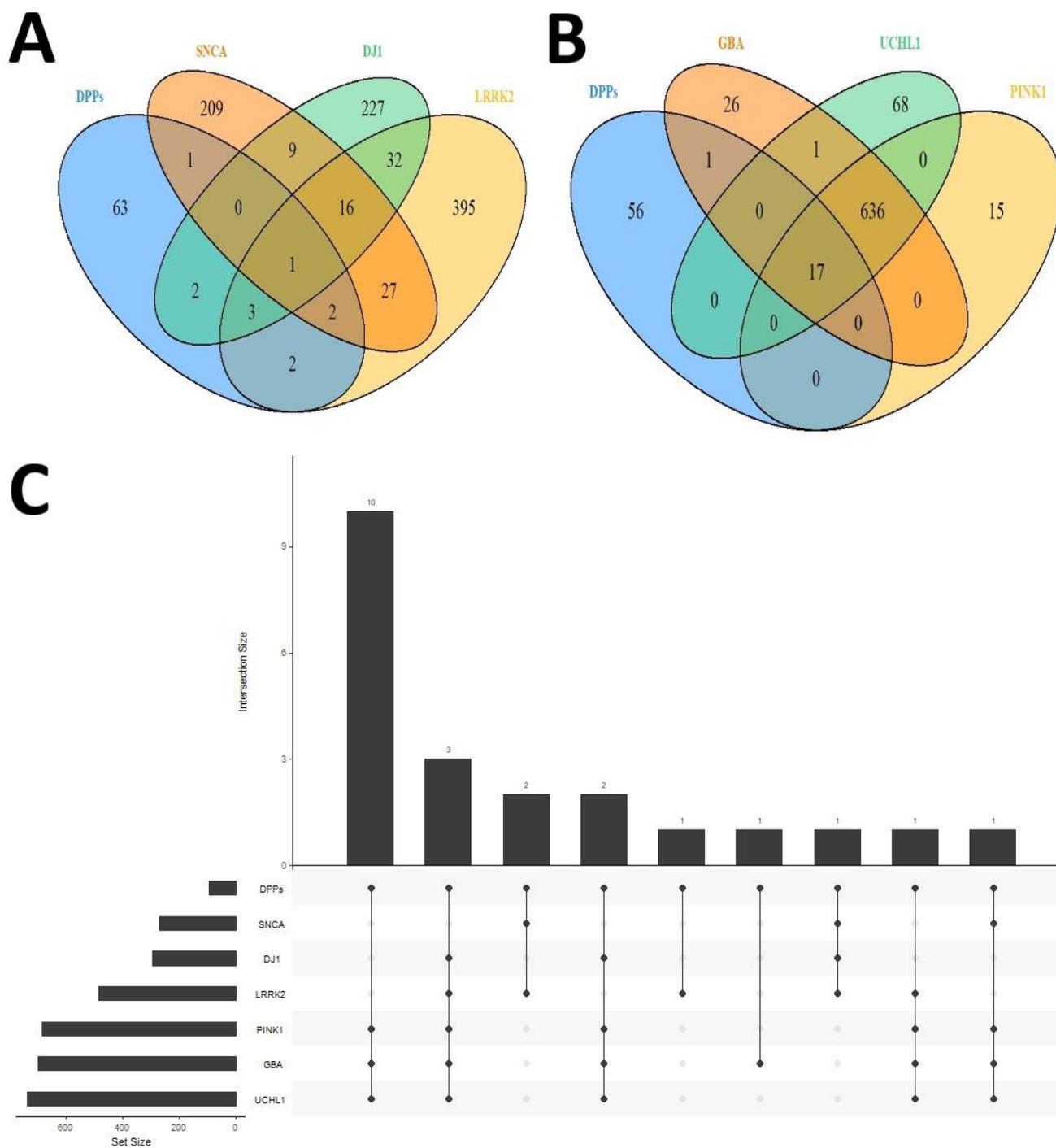
Figure 8. (A) Venn diagram showing the overlap between SNCA (α-synuclein), LRRK2 and DJ-1 interactomes with DPPs. (B) Venn diagram showing the overlap between GBA, UCHL1 and PINK1 interactomes with DPPs. (C) Upset plot showing the sizes of each interactome, and number and overlaps between SNCA, DJ1, LRRK2, PINK1, GBA y UCHL1 interactomes and DPPs.

Lastly, to explore whether alterations in protein palmitoylation occur in specific cell types in the cortex of PD patients, we compared our differential dataset with public available cell-type RNA-seq and proteomics datasets [95,96]. As shown in Supplementary Figure 1 and Supplementary Table 8, a high proportion of DPPs and QPPs tend to be wide-spread distributed across multiple cell-types, being neuronal proteins the most enriched protein set.

3. Materials and Methods

3.1 Subjects and samples

Post-mortem cerebral cortex samples were obtained from the Banner Sun Health Research Institute Brain and Body Donation Program of Sun City, Arizona (https://www.bannerhealth.com/es/services/research/locations/sun-health-institute/programs/body-donation) after approval by the Banner Sun Health Research Institute Tissue Allocation Committee (IRB approval from the wcg IRB of Puyallup, Washington; study number 1132516). Control is a subject without dementia or parkinsonism during life and without a major neuropathological diagnosis. A Parkinson's disease (PD) patient is defined as having two of the three cardinal clinical signs of resting tremor, muscular rigidity, and bradykinesia, along with pigmented neuron loss and Lewy bodies in the substantia nigra. We included in this study four PD and four control cortical samples. There were no differences between control and PD groups regarding postmortem interval to obtain biopsies (around 3 h after death), senile plaque density, or neurofibrillary tangle density. Additional information on subject characteristics is available on Supplementary Table 1.

3.2 Acyl-RAC

The isolation of palmitoylated proteins was based on the Acyl-Rac method [97][98]. Samples were homogenized in blocking buffer (100 mM HEPES, 1.0 mM EDTA, 2.5% SDS, 1% MMTS, pH 7.5) and incubated for 4 hours at 37°C and 1200 rpm. Samples were precipitated in acetone to eliminate the excess of MMTS and then pellets were washed with acetone 70%. Pellets were dried and resolubilized in binding buffer (100 mM HEPES, 1.0 mM EDTA, 1% SDS, pH 7.5) at 40°C and 1400 rpm for 2 hours. Palmitoylated proteins were captured on thiopropyl sepharose beads (T8387, Sigma Aldrich) in the presence of 200 mM hydroxylamine, pH 7.4, by rotating for 2.5 hours at room temperature. The beads were then washed five times in binding buffer and the proteins were eluted in SDS PAGE loading buffer with 100 mM DTT.

3.3 Label free proteomics

To perform label free LC-MS/MS-based comparative proteomics, proteins isolated from Acyl-Rac technique had brief electrophoresis followed by in-gel digest and reductive alkylation. Trypsin-digested peptides were processed for mass spectrometry using a Thermo Scientific LTQ Velos Pro Orbitrap following methodology described before at the FingerPrints Proteomics Service at the University of Dundee (https://proteomics.lifesci.dundee.ac.uk/) [28]. The raw MS/MS spectra searches were processed using the MaxQuant software and default settings (v 1.6.7.0) [99] and searched against the Uniprot proteome reference for Homo sapiens (Proteome ID: UP000005640_9606).

3.4 Data preprocessing

Supplementary Table 2 contains the quantification and identification of proteins from the proteomics experiment. To obtain the list of differential palmitoylated proteins (DPPs), analysis of the LFQ (label-free quantitation) intensities proteins were performed using the software "R" and the integrated development environment "RStudio" [100], through packages found in the repositories CRAN ("The Comprehensive R Archive Network") [101] and Bioconductor [102]. The packages "tidyverse" [103], "cowplot" [104], "gridExtra" [105], and "ggpubr" [106] were used for data transformation and plotting. "Readxl" package [107] was used for opening Excel files in RStudio IDE. Value imputation was performed as follows: data was imputed in a way that those proteins that did not have at least two values for each group (50%) were removed. The average expression value per group was calculated for each protein, filling missing values and completing the data matrix. After imputation, the "NormalyszerDE" package [108] was used to evaluate different data normalization approaches. The dataset with log2(FC) transformation and quantile normalization was selected. The selection of this type of normalization was corroborated by other statistical metrics such as Pearson and Spearman correlation, Q-Q plots, MA plots, etc., generated by the statistical summary of the own package. Looking at the LFQ intensities, certain proteins had an interesting pattern of presence/absence of signal in controls and PD samples. These proteins, named through the manuscript "qualitative palmitoylated proteins" (QPPs), showed values in at least three samples in one group (either control or PD), and missing values (value of 0) in at least three samples in the other group. Since these proteins had no sufficient values for quantitation in both conditions, they could not be used for the study of DPPs. Nevertheless, we considered that these candidates have potential biological interest as the presence or absence of these proteins suggest drastic changes in the levels of palmitoylation between both experimental groups.

3.5 Data analysis

The "limma" package [109] was applied to obtain the DPPs. Proteins with a Log2(FC): ≥[0.6] and a p-value: ≤0.05 were selected. A heatmap of differential palmitoylated proteins was plotted using "pheatmap" package [110]. Z-score normalization [111] was calculated to rearrange DPPs used in the heatmap. SwissPalm database gathers information about the palmitoylation of proteins in the UniprotKB database [30]. We used this database to examine the previous knowledge about palmitoylation of the proteins identified in our cortical palmitome study.

For functional enrichment analysis, "clusterProfiler" package [112,113] and "org.Hs.eg.db" annotation package [114] were used. "enrichGO" function from "cluster Profiler" was applied to search terms from Gene Ontology (GO) database [115][32] using DPPs. In this case, the method for determining the adjusted p-value was the false discovery rate (FDR) [116], with a threshold of 0.05. To further characterize the alteration in metabolic pathways, Metascape web tool was used [49] to simultaneously query the databases KEGG [117–119], a set of databases that gather information about diverse species, as well as mapping tools for network visualization and molecular pathways; Reactome [120], a database which provides curated information about a wide range of biological processes in *Homo sapiens*; and WikiPathways [121], a database that includes molecular pathways linked to rare diseases and lipid metabolism, among other biological pathways. For this purpose, a list of DPPs translated into gene symbols was generated using the web tool BioDBnet [122]. Ten of the DPPs and 9 of the QDPs (see Supplementary Tables 3 and 6) could not be translated into gene symbols, probably due to poor annotation. Within Metascape options, the customized analysis was selected, where only the databases previously mentioned and a p-value threshold of 0.05 were selected. All other enrichment options were set as default. The study of protein-protein interactions (PPI) within DPPs was performed using the STRING database and software [58]. To obtain the network, the list of DPPs, applying a p-value of 0.05 as a threshold, was used in combination with the color and intensity of the aureoles based on log2(FC) depending on their up (red) or down (green) palmitoylated levels. To complement the functional outputs, the Ingenuity Pathway Analysis software (IPA) from QIAGEN was applied [123], using exclusively the database information of experimental and predictive origin regarding central nervous system in order to be confident about the potential affected signaling routes.

To interlock our DPPs dataset with interactomes from relevant PD-associated genes, experimentally demonstrated interactors of these PARK genes were downloaded from the BIOGRID database [124]. The packages "Venndetail" [125] and "UpSetR" [126] were used to visualize the interactomes through Venn diagrams and upset plots. To map DPPs signature in specific brain cell-types, public RNA-seq and proteomics datasets were used [96][95].

4. Conclusions

To the best of our knowledge, these results demonstrate, for the first time, alterations in protein palmitoylation levels in human PD tissue, suggesting a possible role of this posttranslational modification in the pathophysiology of PD. Future palmitoyl-proteomic approaches and palmitoylation assays in human samples could generate new knowledge to better understand the involvement of palmitoylation in PD. Our results confirm the role of well-stablished molecular pathways previously linked to PD, such as mitochondrial dysfunction, oxidative stress, and inflammation. In addition, our proteomics approach pointed out to fibrinogen and cytoskeleton proteins as novel targets linked to neurodegeneration in PD.

References

1.   Balestrino, R.; Schapira, A.H.V. Parkinson Disease. *Eur. J. Neurol.* **2020**, *27*, 27–42, doi:10.1111/ENE.14108.

2.   Foffani, G.; Obeso, J.A. A Cortical Pathogenic Theory of Parkinson's Disease. *Neuron* **2018**, *99*, 1116–1128, doi:10.1016/J.NEURON.2018.07.028.

3.   Schapira, A.H.V.; Chaudhuri, K.R.; Jenner, P. Non-Motor Features of Parkinson Disease. *Nat. Rev. Neurosci.* **2017**, *18*, 435–450, doi:10.1038/NRN.2017.62.

4.   Panicker, N.; Ge, P.; Dawson, V.L.; Dawson, T.M. The Cell Biology of Parkinson's Disease. *J. Cell Biol.* **2021**, *220*, doi:10.1083/JCB.202012095.

5.   Armstrong, M.J.; Okun, M.S. Diagnosis and Treatment of Parkinson Disease: A Review. *JAMA* **2020**, *323*, 548–560, doi:10.1001/JAMA.2019.22360.

6.   Hornemann, T. Palmitoylation and Depalmitoylation Defects. *J. Inherit. Metab. Dis.* **2015**, *38*, 179–186, doi:10.1007/S10545-014-9753-0.

7.   Zaręba-Kozioł, M.; Figiel, I.; Bartkowiak-Kaczmarek, A.; Włodarczyk, J. Insights Into Protein S-Palmitoylation in Synaptic Plasticity and Neurological Disorders: Potential and Limitations of Methods for Detection and Analysis. *Front. Mol. Neurosci.* **2018**, *11*, doi:10.3389/FNMOL.2018.00175.

8.   Fukata, Y.; Fukata, M. Protein Palmitoylation in Neuronal Development and Synaptic Plasticity. *Nat. Rev. Neurosci.* **2010**, *11*, 161–175, doi:10.1038/NRN2788.

9.   Qu, M.; Zhou, X.; Wang, X.; Li, H. Lipid-Induced S-Palmitoylation as a Vital Regulator of Cell Signaling and Disease Development. *Int. J. Biol. Sci.* **2021**, *17*, 4223–4237, doi:10.7150/IJBS.64046.

10.   Jin, J.; Zhi, X.; Wang, X.; Meng, D. Protein Palmitoylation and Its Pathophysiological Relevance. *J. Cell. Physiol.* **2021**, *236*, 3220–3233, doi:10.1002/JCP.30122.

11.   Cho, E.; Park, M. Palmitoylation in Alzheimer's Disease and Other Neurodegenerative Diseases. *Pharmacol. Res.* **2016**, *111*, 133–151, doi:10.1016/J.PHRS.2016.06.008.

12.   Young, F.B.; Butland, S.L.; Sanders, S.S.; Sutton, L.M.; Hayden, M.R. Putting Proteins in Their Place: Palmitoylation in Huntington Disease and Other Neuropsychiatric Diseases. *Prog. Neurobiol.* **2012**, *97*, 220–238, doi:10.1016/j.pneurobio.2011.11.002.

13.   Hornemann, T. Palmitoylation and Depalmitoylation Defects. *J. Inherit. Metab. Dis.* **2015**, *38*, 179–186, doi:10.1007/S10545-014-9753-0.

14.   Pinner, A.L.; Tucholski, J.; Haroutunian, V.; McCullumsmith, R.E.; Meador-Woodruff, J.H. Decreased Protein S-Palmitoylation in Dorsolateral Prefrontal Cortex in Schizophrenia. *Schizophr. Res.* **2016**, *177*, 78–87, doi:10.1016/J.SCHRES.2016.01.054.

15.   Sanders, S.S.; Hayden, M.R. Aberrant Palmitoylation in Huntington Disease. *Biochem. Soc. Trans.* **2015**, *43*, 205–210, doi:10.1042/BST20140242.

16.   Virlogeux, A.; Scaramuzzino, C.; Lenoir, S.; Carpentier, R.; Louessard, M.; Genoux, A.; Lino, P.; Hinckelmann, M.V.;

Perrier, A.L.; Humbert, S.; et al. Increasing Brain Palmitoylation Rescues Behavior and Neuropathology in Huntington Disease Mice. *Sci. Adv.* **2021**, *7*, doi:10.1126/SCIADV.ABB0799.

17. Henderson, M.X.; Wirak, G.S.; Zhang, Y. quan; Dai, F.; Ginsberg, S.D.; Dolzhanskaya, N.; Staropoli, J.F.; Nijssen, P.C.G.; Lam, T.K.T.; Roth, A.F.; et al. Neuronal Ceroid Lipofuscinosis with DNAJC5/CSPα Mutation Has PPT1 Pathology and Exhibit Aberrant Protein Palmitoylation. *Acta Neuropathol.* **2016**, *131*, 621–637, doi:10.1007/S00401-015-1512-2.

18. Kang, R.; Wan, J.; Arstikaitis, P.; Takahashi, H.; Huang, K.; Bailey, A.O.; Thompson, J.X.; Roth, A.F.; Drisdel, R.C.; Mastro, R.; et al. Neural Palmitoyl-Proteomics Reveals Dynamic Synaptic Palmitoylation. *Nature* **2008**, *456*, 904–909, doi:10.1038/NATURE07605.

19. Prescott, G.R.; Gorleku, O.A.; Greaves, J.; Chamberlain, L.H. Palmitoylation of the Synaptic Vesicle Fusion Machinery. *J. Neurochem.* **2009**, *110*, 1135–1149, doi:10.1111/J.1471-4159.2009.06205.X.

20. Ji, B.; Skup, M. Roles of Palmitoylation in Structural Long-Term Synaptic Plasticity. *Mol. Brain* **2021**, *14*, doi:10.1186/S13041-020-00717-Y.

21. Globa, A.K.; Bamji, S.X. Protein Palmitoylation in the Development and Plasticity of Neuronal Connections. *Curr. Opin. Neurobiol.* **2017**, *45*, 210–220, doi:10.1016/J.CONB.2017.02.016.

22. Rastedt, D.E.; Vaughan, R.A.; Foster, J.D. Palmitoylation Mechanisms in Dopamine Transporter Regulation. *J. Chem. Neuroanat.* **2017**, *83–84*, 3–9, doi:10.1016/J.JCHEMNEU.2017.01.002.

23. Kim, K.S.; Kim, J.S.; Park, J.Y.; Suh, Y.H.; Jou, I.; Joe, E.H.; Park, S.M. DJ-1 Associates with Lipid Rafts by Palmitoylation and Regulates Lipid Rafts-Dependent Endocytosis in Astrocytes. *Hum. Mol. Genet.* **2013**, *22*, 4805–4817, doi:10.1093/HMG/DDT332.

24. Schapansky, J.; Nardozzi, J.D.; Felizia, F.; LaVoie, M.J. Membrane Recruitment of Endogenous LRRK2 Precedes Its Potent Regulation of Autophagy. *Hum. Mol. Genet.* **2014**, *23*, 4201, doi:10.1093/HMG/DDU138.

25. Ho, G.P.H.; Ramalingam, N.; Imberdis, T.; Wilkie, E.C.; Dettmer, U.; Selkoe, D.J. Upregulation of Cellular Palmitoylation Mitigates α-Synuclein Accumulation and Neurotoxicity. *Mov. Disord.* **2021**, *36*, 348–359, doi:10.1002/MDS.28346.

26. Hosp, F.; Mann, M. A Primer on Concepts and Applications of Proteomics in Neuroscience. *Neuron* **2017**, *96*, 558–571, doi:10.1016/J.NEURON.2017.09.025.

27. Sanders, S.S.; Martin, D.D.O.; Butland, S.L.; Lavallée-Adam, M.; Calzolari, D.; Kay, C.; Yates, J.R.; Hayden, M.R. Curation of the Mammalian Palmitoylome Indicates a Pivotal Role for Palmitoylation in Diseases and Disorders of the Nervous System and Cancers. *PLoS Comput. Biol.* **2015**, *11*, doi:10.1371/JOURNAL.PCBI.1004405.

28. Aghamaleky Sarvestany, A.; Hunter, G.; Tavendale, A.; Lamont, D.J.; Llavero Hurtado, M.; Graham, L.C.; Wishart, T.M.; Gillingwater, T.H. Label-Free Quantitative Proteomic Profiling Identifies Disruption of Ubiquitin Homeostasis as a Key Driver of Schwann Cell Defects in Spinal Muscular Atrophy. *J. Proteome Res.* **2014**, *13*, 4546–4557, doi:10.1021/PR500492J/SUPPL_FILE/PR500492J_SI_003.PDF.

29. Cox, J.; Hein, M.Y.; Luber, C.A.; Paron, I.; Nagaraj, N.; Mann, M. Accurate Proteome-Wide Label-Free Quantification by Delayed Normalization and Maximal Peptide Ratio Extraction, Termed MaxLFQ. *Mol. Cell. Proteomics* **2014**, *13*, 2513–2526, doi:10.1074/MCP.M113.031591.

30. Blanc, M.; David, F.P.A.; Abrami, L.; Migliozzi, D.; Armand, F.; Jérôme, B.; van der Goot, F.G. SwissPalm: Protein Palmitoylation Database Available online: https://swisspalm.org/reference (accessed on 9 October 2022).

31. Henderson, M.X.; Wirak, G.S.; Zhang, Y. quan; Dai, F.; Ginsberg, S.D.; Dolzhanskaya, N.; Staropoli, J.F.; Nijssen, P.C.G.; Lam, T.K.T.; Roth, A.F.; et al. Neuronal Ceroid Lipofuscinosis with DNAJC5/CSPα Mutation Has PPT1 Pathology and Exhibit Aberrant Protein Palmitoylation. *Acta Neuropathol.* **2016**, *131*, 621–637, doi:10.1007/S00401-015-1512-2.

32. Carbon, S.; Douglass, E.; Good, B.M.; Unni, D.R.; Harris, N.L.; Mungall, C.J.; Basu, S.; Chisholm, R.L.; Dodson, R.J.; Hartline, E.; et al. The Gene Ontology Resource: Enriching a GOld Mine. *Nucleic Acids Res.* **2021**, *49*, D325–D334, doi:10.1093/NAR/GKAA1113.

33. Lin, D.T.S.; Conibear, E. ABHD17 Proteins Are Novel Protein Depalmitoylases That Regulate N-Ras Palmitate Turnover

and Subcellular Localization. *Elife* **2015**, *4*, doi:10.7554/ELIFE.11306.

34. Davalos, D.; Akassoglou, K. Fibrinogen as a Key Regulator of Inflammation in Disease. *Semin. Immunopathol.* **2012**, *34*, 43–62, doi:10.1007/S00281-011-0290-8.

35. Kattula, S.; Byrnes, J.R.; Wolberg, A.S. Fibrinogen and Fibrin in Hemostasis and Thrombosis. *Arterioscler. Thromb. Vasc. Biol.* **2017**, *37*, e13–e21, doi:10.1161/ATVBAHA.117.308564.

36. Halper, J.; Kjaer, M. Basic Components of Connective Tissues and Extracellular Matrix: Elastin, Fibrillin, Fibulins, Fibrinogen, Fibronectin, Laminin, Tenascins and Thrombospondins. *Adv. Exp. Med. Biol.* **2014**, *802*, 31–47, doi:10.1007/978-94-007-7893-1_3.

37. Tang, H.; Fu, Y.; Cui, Y.; He, Y.; Zeng, X.; Ploplis, V.A.; Castellino, F.J.; Luo, Y. Fibrinogen Has Chaperone-like Activity. *Biochem. Biophys. Res. Commun.* **2009**, *378*, 662–667, doi:10.1016/J.BBRC.2008.11.112.

38. Petersen, M.A.; Ryu, J.K.; Akassoglou, K. Fibrinogen in Neurological Diseases: Mechanisms, Imaging and Therapeutics. *Nat. Rev. Neurosci.* **2018**, *19*, 283–301, doi:10.1038/NRN.2018.13.

39. Sulimai, N.; Lominadze, D. Fibrinogen and Neuroinflammation During Traumatic Brain Injury. *Mol. Neurobiol.* **2020**, *57*, 4692–4703, doi:10.1007/S12035-020-02012-2.

40. Merlini, M.; Rafalski, V.A.; Rios Coronado, P.E.; Gill, T.M.; Ellisman, M.; Muthukumar, G.; Subramanian, K.S.; Ryu, J.K.; Syme, C.A.; Davalos, D.; et al. Fibrinogen Induces Microglia-Mediated Spine Elimination and Cognitive Impairment in an Alzheimer's Disease Model. *Neuron* **2019**, *101*, 1099-1108.e6, doi:10.1016/J.NEURON.2019.01.014.

41. Davalos, D.; Mahajan, K.R.; Trapp, B.D. Brain Fibrinogen Deposition Plays a Key Role in MS Pathophysiology - Yes. *Mult. Scler.* **2019**, *25*, 1434–1435, doi:10.1177/1352458519852723.

42. Pajares, M.; I Rojo, A.; Manda, G.; Boscá, L.; Cuadrado, A. Inflammation in Parkinson's Disease: Mechanisms and Therapeutic Implications. *Cells* **2020**, *9*, doi:10.3390/CELLS9071687.

43. Sweeney, M.D.; Sagare, A.P.; Zlokovic, B. V. Blood-Brain Barrier Breakdown in Alzheimer Disease and Other Neurodegenerative Disorders. *Nat. Rev. Neurol.* **2018**, *14*, 133–150, doi:10.1038/NRNEUROL.2017.188.

44. Sato, Y.; Kaji, M.; Metoki, N.; Yoshida, H.; Satoh, K. Coagulation-Fibrinolysis Abnormalities in Patients Receiving Antiparkinsonian Agents. *J. Neurol. Sci.* **2003**, *212*, 55–58, doi:10.1016/S0022-510X(03)00101-1.

45. Chen, H.; O'Reilly, E.J.; Schwarzschild, M.A.; Ascherio, A. Peripheral Inflammatory Biomarkers and Risk of Parkinson's Disease. *Am. J. Epidemiol.* **2008**, *167*, 90–95, doi:10.1093/AJE/KWM260.

46. Wong, K.T.; Grove, J.S.; Grandinetti, A.; Curb, J.D.; Yee, M.; Blanchette, P.; Ross, G.W.; Rodriguez, B.I. Association of Fibrinogen with Parkinson Disease in Elderly Japanese-American Men: A Prospective Study. *Neuroepidemiology* **2010**, *34*, 50–54, doi:10.1159/000260070.

47. Song, I.U.; Kim, Y. Do; Cho, H.J.; Chung, S.W. Is Neuroinflammation Involved in the Development of Dementia in Patients with Parkinson's Disease? *Intern. Med.* **2013**, *52*, 1787–1792, doi:10.2169/INTERNALMEDICINE.52.0474.

48. Maarouf, C.L.; Beach, T.G.; Adler, C.H.; Shill, H.A.; Sabbagh, M.N.; Wu, T.; Walker, D.G.; Kokjohn, T.A.; Roher, A.E. Cerebrospinal Fluid Biomarkers of Neuropathologically Diagnosed Parkinson's Disease Subjects. *Neurol. Res.* **2012**, *34*, 669–676, doi:10.1179/1743132812Y.0000000063.

49. Zhou, Y.; Zhou, B.; Pache, L.; Chang, M.; Khodabakhshi, A.H.; Tanaseichuk, O.; Benner, C.; Chanda, S.K. Metascape Provides a Biologist-Oriented Resource for the Analysis of Systems-Level Datasets. *Nat. Commun.* **2019**, *10*, doi:10.1038/S41467-019-09234-6.

50. González-Rodríguez, P.; Zampese, E.; Stout, K.A.; Guzman, J.N.; Ilijic, E.; Yang, B.; Tkatch, T.; Stavarache, M.A.; Wokosin, D.L.; Gao, L.; et al. Disruption of Mitochondrial Complex I Induces Progressive Parkinsonism. *Nat. 2021 5997886* **2021**, *599*, 650–656, doi:10.1038/s41586-021-04059-0.

51. Haas, R.H.; Nasirian, F.; Nakano, K.; Ward, D.; Pay, M.; Hill, R.; Shults, C.W. Low Platelet Mitochondrial Complex I and Complex II/III Activity in Early Untreated Parkinson's Disease. *Ann. Neurol.* **1995**, *37*, 714–722, doi:10.1002/ANA.410370604.

52. Yoshino, H.; Nakagawa-Hattori, Y.; Kondo, T.; Mizuno, Y. Mitochondrial Complex I and II Activities of Lymphocytes and

Platelets in Parkinson's Disease. *J. Neural Transm. Park. Dis. Dement. Sect.* **1992**, *4*, 27–34, doi:10.1007/BF02257619.

53.   Liu, Z.; ye, Q.; Wang, F.; Guo, Y.; Cui, R.; Wang, J.; Wang, D. Protective Effect of Thioredoxin Reductase 1 in Parkinson's Disease. *Neurosci. Lett.* **2021**, *741*, doi:10.1016/J.NEULET.2020.135457.

54.   Xu, Z.; Zhong, L. New Insights into the Posttranslational Regulation of Human Cytosolic Thioredoxin by S-Palmitoylation. *Biochem. Biophys. Res. Commun.* **2015**, *460*, 949–956, doi:10.1016/J.BBRC.2015.03.132.

55.   Qin, H.; Liang, W.; Xu, Z.; Ye, F.; Li, X.; Zhong, L. Mechanistic Insights into the Inhibitory Effects of Palmitoylation on Cytosolic Thioredoxin Reductase and Thioredoxin. *Biochimie* **2015**, *110*, 25–35, doi:10.1016/J.BIOCHI.2014.12.018.

56.   Ren, Y.; Zhao, J.; Feng, J. Parkin Binds to Alpha/Beta Tubulin and Increases Their Ubiquitination and Degradation. *J. Neurosci.* **2003**, *23*, 3316–3324, doi:10.1523/JNEUROSCI.23-08-03316.2003.

57.   Zambito, A.M.; Wolff, J. Palmitoylation of Tubulin. *Biochem. Biophys. Res. Commun.* **1997**, *239*, 650–654, doi:10.1006/BBRC.1997.7525.

58.   Szklarczyk, D.; Gable, A.L.; Nastou, K.C.; Lyon, D.; Kirsch, R.; Pyysalo, S.; Doncheva, N.T.; Legeay, M.; Fang, T.; Bork, P.; et al. The STRING Database in 2021: Customizable Protein-Protein Networks, and Functional Characterization of User-Uploaded Gene/Measurement Sets. *Nucleic Acids Res.* **2021**, *49*, D605–D612, doi:10.1093/NAR/GKAA1074.

59.   Binarová, P.; Tuszynski, J. Tubulin: Structure, Functions and Roles in Disease. *Cells* **2019**, *8*, doi:10.3390/CELLS8101294.

60.   Saha, P.; Sen, N. Tauopathy: A Common Mechanism for Neurodegeneration and Brain Aging. *Mech. Ageing Dev.* **2019**, *178*, 72–79, doi:10.1016/J.MAD.2019.01.007.

61.   Alquezar, C.; Arya, S.; Kao, A.W. Tau Post-Translational Modifications: Dynamic Transformers of Tau Function, Degradation, and Aggregation. *Front. Neurol.* **2021**, *11*, 1826, doi:10.3389/FNEUR.2020.595532/BIBTEX.

62.   Strang, K.H.; Golde, T.E.; Giasson, B.I. MAPT Mutations, Tauopathy, and Mechanisms of Neurodegeneration. *Lab. Invest.* **2019**, *99*, 912–928, doi:10.1038/S41374-019-0197-X.

63.   Leveille, E.; Ross, O.A.; Gan-Or, Z. Tau and MAPT Genetics in Tauopathies and Synucleinopathies. *Parkinsonism Relat. Disord.* **2021**, *90*, 142–154, doi:10.1016/J.PARKRELDIS.2021.09.008.

64.   Zhang, X.; Gao, F.; Wang, D.; Li, C.; Fu, Y.; He, W.; Zhang, J. Tau Pathology in Parkinson's Disease. *Front. Neurol.* **2018**, *9*, doi:10.3389/FNEUR.2018.00809.

65.   Li, X.; James, S.; Lei, P. Interactions Between α-Synuclein and Tau Protein: Implications to Neurodegenerative Disorders. *J. Mol. Neurosci.* **2016**, *60*, 298–304, doi:10.1007/S12031-016-0829-1.

66.   O'Leary, H.; Lasda, E.; Bayer, K.U. CaMKIIβ Association with the Actin Cytoskeleton Is Regulated by Alternative Splicing. *Mol. Biol. Cell* **2006**, *17*, 4656, doi:10.1091/MBC.E06-03-0252.

67.   Fink, C.C.; Bayer, K.U.; Myers, J.W.; Ferrell, J.E.; Schulman, H.; Meyer, T. Selective Regulation of Neurite Extension and Synapse Formation by the β but Not the α Isoform of CaMKII. *Neuron* **2003**, *39*, 283–297, doi:10.1016/S0896-6273(03)00428-8.

68.   Pellegrini, L.; Wetzel, A.; Grannó, S.; Heaton, G.; Harvey, K. Back to the Tubule: Microtubule Dynamics in Parkinson's Disease. *Cell. Mol. Life Sci.* **2017**, *74*, 409, doi:10.1007/S00018-016-2351-6.

69.   Calogero, A.M.; Mazzetti, S.; Pezzoli, G.; Cappelletti, G. Neuronal Microtubules and Proteins Linked to Parkinson's Disease: A Relevant Interaction? *Biol. Chem.* **2019**, *400*, 1099–1112, doi:10.1515/HSZ-2019-0142.

70.   Oliveira da Silva, M.I.; Liz, M.A. Linking Alpha-Synuclein to the Actin Cytoskeleton: Consequences to Neuronal Function. *Front. cell Dev. Biol.* **2020**, *8*, doi:10.3389/FCELL.2020.00787.

71.   Bertoni, A.; Tadokoro, S.; Eto, K.; Pampori, N.; Parise, L. V.; White, G.C.; Shattil, S.J. Relationships between Rap1b, Affinity Modulation of Integrin Alpha IIbbeta 3, and the Actin Cytoskeleton. *J. Biol. Chem.* **2002**, *277*, 25715–25721, doi:10.1074/JBC.M202791200.

72.   Zhang, G.; Xiang, B.; Ye, S.; Chrzanowska-Wodnicka, M.; Morris, A.J.; Gartner, T.K.; Whiteheart, S.W.; White, G.C.; Smyth, S.S.; Li, Z. Distinct Roles for Rap1b Protein in Platelet Secretion and Integrin AIIbβ3 Outside-in Signaling. *J. Biol. Chem.* **2011**, *286*, 39466–39477, doi:10.1074/JBC.M111.239608.

73.   Rai, S.N.; Dilnashin, H.; Birla, H.; Singh, S. Sen; Zahra, W.; Rathore, A.S.; Singh, B.K.; Singh, S.P. The Role of PI3K/Akt and

ERK in Neurodegenerative Disorders. *Neurotox. Res.* **2019**, *35*, 775–795, doi:10.1007/S12640-019-0003-Y.

74. Su, J.; Zhang, J.; Bao, R.; Xia, C.; Zhang, Y.; Zhu, Z.; Lv, Q.; Qi, Y.; Xue, J. Mitochondrial Dysfunction and Apoptosis Are Attenuated through Activation of AMPK/GSK-3β/PP2A Pathway in Parkinson's Disease. *Eur. J. Pharmacol.* **2021**, *907*, doi:10.1016/J.EJPHAR.2021.174202.

75. Heo, K.; Basu, H.; Gutnick, A.; Wei, W.; Shlevkov, E.; Schwarz, T.L. Serine/Threonine Protein Phosphatase 2A Regulates the Transport of Axonal Mitochondria. *Front. Cell. Neurosci.* **2022**, *16*, 88, doi:10.3389/FNCEL.2022.852245/BIBTEX.

76. Jiang, M.Y.; Han, Z.D.; Li, W.; Yue, F.; Ye, J.; Li, B.; Cai, Z.; Lu, J.M.; Dong, W.; Jiang, X.; et al. Mitochondrion-Associated Protein Peroxiredoxin 3 Promotes Benign Prostatic Hyperplasia through Autophagy Suppression and Pyroptosis Activation. *Oncotarget* **2017**, *8*, 80295–80302, doi:10.18632/ONCOTARGET.17927.

77. Trist, B.G.; Hare, D.J.; Double, K.L. Oxidative Stress in the Aging Substantia Nigra and the Etiology of Parkinson's Disease. *Aging Cell* **2019**, *18*, doi:10.1111/ACEL.13031.

78. Ma, X.; Li, H.; He, Y.; Hao, J. The Emerging Link between O-GlcNAcylation and Neurological Disorders. *Cell. Mol. Life Sci.* **2017**, *74*, 3667–3686, doi:10.1007/S00018-017-2542-9.

79. Yuzwa, S.A.; Shan, X.; MacAuley, M.S.; Clark, T.; Skorobogatko, Y.; Vosseller, K.; Vocadlo, D.J. Increasing O-GlcNAc Slows Neurodegeneration and Stabilizes Tau against Aggregation. *Nat. Chem. Biol.* **2012**, *8*, 393–399, doi:10.1038/NCHEMBIO.797.

80. Liu, F.; Iqbal, K.; Grundke-Iqbal, I.; Hart, G.W.; Gong, C.X. O-GlcNAcylation Regulates Phosphorylation of Tau: A Mechanism Involved in Alzheimer's Disease. *Proc. Natl. Acad. Sci. U. S. A.* **2004**, *101*, 10804–10809, doi:10.1073/PNAS.0400348101.

81. Marotta, N.P.; Lin, Y.H.; Lewis, Y.E.; Ambroso, M.R.; Zaro, B.W.; Roth, M.T.; Arnold, D.B.; Langen, R.; Pratt, M.R. O-GlcNAc Modification Blocks the Aggregation and Toxicity of the Protein α-Synuclein Associated with Parkinson's Disease. *Nat. Chem.* **2015**, *7*, 913–920, doi:10.1038/NCHEM.2361.

82. Lewis, Y.E.; Galesic, A.; Levine, P.M.; De Leon, C.A.; Lamiri, N.; Brennan, C.K.; Pratt, M.R. O-GlcNAcylation of α-Synuclein at Serine 87 Reduces Aggregation without Affecting Membrane Binding. *ACS Publ.* **2017**, *12*, 1020–1027, doi:10.1021/acschembio.7b00113.

83. Cao, Y.; Qiu, T.; Kathayat, R.S.; Azizi, S.A.; Thorne, A.K.; Ahn, D.; Fukata, Y.; Fukata, M.; Rice, P.A.; Dickinson, B.C. ABHD10 Is an S-Depalmitoylase Affecting Redox Homeostasis through Peroxiredoxin-5. *Nat. Chem. Biol.* **2019**, *15*, 1232, doi:10.1038/S41589-019-0399-Y.

84. Lanzillotta, A.; Porrini, V.; Bellucci, A.; Benarese, M.; Branca, C.; Parrella, E.; Spano, P.F.; Pizzi, M. NF-KB in Innate Neuroprotection and Age-Related Neurodegenerative Diseases. *Front. Neurol.* **2015**, *6*, doi:10.3389/FNEUR.2015.00098.

85. Bellucci, A.; Bubacco, L.; Longhena, F.; Parrella, E.; Faustini, G.; Porrini, V.; Bono, F.; Missale, C.; Pizzi, M. Nuclear Factor-KB Dysregulation and α-Synuclein Pathology: Critical Interplay in the Pathogenesis of Parkinson's Disease. *Front. Aging Neurosci.* **2020**, *12*, 68, doi:10.3389/FNAGI.2020.00068/BIBTEX.

86. Singh, S. Sen; Rai, S.N.; Birla, H.; Zahra, W.; Rathore, A.S.; Singh, S.P. NF-KB-Mediated Neuroinflammation in Parkinson's Disease and Potential Therapeutic Effect of Polyphenols. *Neurotox. Res.* **2020**, *37*, 491–507, doi:10.1007/S12640-019-00147-2.

87. Ghosh, A.; Roy, A.; Liu, X.; Kordower, J.H.; Mufson, E.J.; Hartley, D.M.; Ghosh, S.; Mosley, R.L.; Gendelman, H.E.; Pahan, K. Selective Inhibition of NF-KB Activation Prevents Dopaminergic Neuronal Loss in a Mouse Model of Parkinson's Disease. *Proc. Natl. Acad. Sci. U. S. A.* **2007**, *104*, 18754–18759, doi:10.1073/PNAS.0704908104/SUPPL_FILE/04908FIG8.PDF.

88. Ordonez, D.G.; Lee, M.K.; Feany, M.B. α-Synuclein Induces Mitochondrial Dysfunction through Spectrin and the Actin Cytoskeleton. *Neuron* **2018**, *97*, 108, doi:10.1016/J.NEURON.2017.11.036.

89. Singh, A.; Zhi, L.; Zhang, H. LRRK2 and Mitochondria: Recent Advances and Current Views. *Brain Res.* **2019**, *1702*, 96–104, doi:10.1016/J.BRAINRES.2018.06.010.

90. Angeles, D.C.; Gan, B.H.; Onstead, L.; Zhao, Y.; Lim, K.L.; Dachsel, J.; Melrose, H.; Farrer, M.; Wszolek, Z.K.; Dickson, D.W.; et al. Mutations in LRRK2 Increase Phosphorylation of Peroxiredoxin 3 Exacerbating Oxidative Stress-Induced Neuronal Death. *Hum. Mutat.* **2011**, *32*, 1390–1397, doi:10.1002/HUMU.21582.

91.   Angeles, D.C.; Ho, P.; Chua, L.L.; Wang, C.; Yap, Y.W.; Ng, C.; Zhou, Z. dong; Lim, K.L.; Wszolek, Z.K.; Wang, H.Y.; et al. Thiol Peroxidases Ameliorate LRRK2 Mutant-Induced Mitochondrial and Dopaminergic Neuronal Degeneration in Drosophila. *Hum. Mol. Genet.* **2014**, *23*, 3157–3165, doi:10.1093/HMG/DDU026.

92.   Hoffmann, L.; Rust, M.B.; Culmsee, C. Actin(g) on Mitochondria - a Role for Cofilin1 in Neuronal Cell Death Pathways. *Biol. Chem.* **2019**, *400*, 1089–1097, doi:10.1515/HSZ-2019-0120.

93.   Sekino, Y.; Kojima, N.; Shirao, T. Role of Actin Cytoskeleton in Dendritic Spine Morphogenesis. *Neurochem. Int.* **2007**, *51*, 92–104, doi:10.1016/J.NEUINT.2007.04.029.

94.   Kim, H.; Kang, H.; Lee, Y.; Park, C.H.; Jo, A.; Khang, R.; Shin, J.H. Identification of Transketolase as a Target of PARIS in Substantia Nigra. *Biochem. Biophys. Res. Commun.* **2017**, *493*, 1050–1056, doi:10.1016/J.BBRC.2017.09.090.

95.   Sharma, K.; Schmitt, S.; Bergner, C.G.; Tyanova, S.; Kannaiyan, N.; Manrique-Hoyos, N.; Kongi, K.; Cantuti, L.; Hanisch, U.K.; Philips, M.A.; et al. Cell Type- and Brain Region-Resolved Mouse Brain Proteome. *Nat. Neurosci.* **2015**, *18*, 1819–1831, doi:10.1038/NN.4160.

96.   Zhang, Y.; Chen, K.; Sloan, S.A.; Bennett, M.L.; Scholze, A.R.; O'Keeffe, S.; Phatnani, H.P.; Guarnieri, P.; Caneda, C.; Ruderisch, N.; et al. An RNA-Sequencing Transcriptome and Splicing Database of Glia, Neurons, and Vascular Cells of the Cerebral Cortex. *J. Neurosci.* **2014**, *34*, 11929–11947, doi:10.1523/JNEUROSCI.1860-14.2014.

97.   Tewari, R.; West, S.J.; Shayahati, B.; Akimzhanov, A.M. Detection of Protein S-Acylation Using Acyl-Resin Assisted Capture. *J. Vis. Exp.* **2020**, *2020*, doi:10.3791/61016.

98.   Gök, C.; Main, A.; Gao, X.; Kerekes, Z.; Plain, F.; Kuo, C.W.; Robertson, A.D.; Fraser, N.J.; Fuller, W. Insights into the Molecular Basis of the Palmitoylation and Depalmitoylation of NCX1. *Cell Calcium* **2021**, *97*, 102408, doi:10.1016/J.CECA.2021.102408.

99.   Tyanova, S.; Temu, T.; Cox, J. The MaxQuant Computational Platform for Mass Spectrometry-Based Shotgun Proteomics. *Nat. Protoc. 2016 1112* **2016**, *11*, 2301–2319, doi:10.1038/nprot.2016.136.

100.   R Core Team (2020) R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna, Austria. Available online: http://www.r-project.org/index.html (accessed on 9 October 2022).

101.   Hornik, K. The Comprehensive R Archive Network. *Wiley Interdiscip. Rev. Comput. Stat.* **2012**, *4*, 394–398, doi:10.1002/WICS.1212.

102.   Gentleman, R.C.; Carey, V.J.; Bates, D.M.; Bolstad, B.; Dettling, M.; Dudoit, S.; Ellis, B.; Gautier, L.; Ge, Y.; Gentry, J.; et al. Bioconductor: Open Software Development for Computational Biology and Bioinformatics. *Genome Biol. 2004 510* **2004**, *5*, 1–16, doi:10.1186/GB-2004-5-10-R80.

103.   Wickham, H.; Averick, M.; Bryan, J.; Chang, W.; D', L.; Mcgowan, A.; François, R.; Grolemund, G.; Hayes, A.; Henry, L.; et al. Welcome to the Tidyverse. *J. Open Source Softw.* **2019**, *4*, 1686, doi:10.21105/JOSS.01686.

104.   Wilke, C.O. Streamlined Plot Theme and Plot Annotations for "ggplot2" [R Package Cowplot Version 1.1.1] Available online: https://cran.r-project.org/package=cowplot (accessed on 9 October 2022).

105.   Auguie, B. Miscellaneous Functions for "Grid" Graphics [R Package GridExtra Version 2.3]. **2017**.

106.   Kassambara, A. Ggplot2 Based Publication Ready Plots • Ggpubr Available online: https://rpkgs.datanovia.com/ggpubr/ (accessed on 9 October 2022).

107.   Wickham, H.; Bryan, J. Read Excel Files [R Package Readxl Version 1.4.1] Available online: https://cran.r-project.org/package=readxl (accessed on 9 October 2022).

108.   Willforss, J.; Chawade, A.; Levander, F. NormalyzerDE: Online Tool for Improved Normalization of Omics Expression Data and High-Sensitivity Differential Expression Analysis. *J. Proteome Res.* **2019**, *18*, 732–740, doi:10.1021/ACS.JPROTEOME.8B00523.

109.   Ritchie, M.E.; Phipson, B.; Wu, D.; Hu, Y.; Law, C.W.; Shi, W.; Smyth, G.K. Limma Powers Differential Expression Analyses for RNA-Sequencing and Microarray Studies. *Nucleic Acids Res.* **2015**, *43*, e47, doi:10.1093/NAR/GKV007.

110.   Kolde, R. Pheatmap: Pretty Heatmaps Available online: https://cran.r-project.org/web/packages/pheatmap/index.html

(accessed on 9 October 2022).

111. Jain, A.; Nandakumar, K.; Ross, A. Score Normalization in Multimodal Biometric Systems. *Pattern Recognit.* **2005**, *38*, 2270–2285, doi:10.1016/J.PATCOG.2005.01.012.

112. Wu, T.; Hu, E.; Xu, S.; Chen, M.; Guo, P.; Dai, Z.; Feng, T.; Zhou, L.; Tang, W.; Zhan, L.; et al. ClusterProfiler 4.0: A Universal Enrichment Tool for Interpreting Omics Data. *Innov. (Cambridge* **2021**, *2*, doi:10.1016/J.XINN.2021.100141.

113. Yu, G.; Wang, L.G.; Han, Y.; He, Q.Y. ClusterProfiler: An R Package for Comparing Biological Themes among Gene Clusters. *OMICS* **2012**, *16*, 284–287, doi:10.1089/OMI.2011.0118.

114. Carlson, M. Org.Hs.Eg.Db: Genome Wide Annotation for Human. R Package Version 3.8.2. Available online: https://bioconductor.org/packages/release/data/annotation/html/org.Hs.eg.db.html (accessed on 10 October 2022).

115. Ashburner, M.; Ball, C.A.; Blake, J.A.; Botstein, D.; Butler, H.; Cherry, J.M.; Davis, A.P.; Dolinski, K.; Dwight, S.S.; Eppig, J.T.; et al. Gene Ontology: Tool for the Unification of Biology. *Nat. Genet. 2000 251* **2000**, *25*, 25–29, doi:10.1038/75556.

116. Benjaminit, Y.; Hochberg, Y. Controlling the False Discovery Rate: A Practical and Powerful Approach to Multiple Testing. *J. R. Stat. Soc. B* **1995**, *57*, 289–300.

117. Kanehisa, M. Toward Understanding the Origin and Evolution of Cellular Organisms. *Protein Sci.* **2019**, *28*, 1947–1951, doi:10.1002/PRO.3715.

118. Kanehisa, M.; Furumichi, M.; Sato, Y.; Ishiguro-Watanabe, M.; Tanabe, M. KEGG: Integrating Viruses and Cellular Organisms. *Nucleic Acids Res.* **2021**, *49*, D545–D551, doi:10.1093/NAR/GKAA970.

119. Kanehisa, M.; Goto, S. KEGG: Kyoto Encyclopedia of Genes and Genomes. *Nucleic Acids Res.* **2000**, *28*, 27–30, doi:10.1093/NAR/28.1.27.

120. Gillespie, M.; Jassal, B.; Stephan, R.; Milacic, M.; Rothfels, K.; Senff-Ribeiro, A.; Griss, J.; Sevilla, C.; Matthews, L.; Gong, C.; et al. The Reactome Pathway Knowledgebase 2022. *Nucleic Acids Res.* **2022**, *50*, D687–D692, doi:10.1093/NAR/GKAB1028.

121. Martens, M.; Ammar, A.; Riutta, A.; Waagmeester, A.; Slenter, D.N.; Hanspers, K.; Miller, R.A.; Digles, D.; Lopes, E.N.; Ehrhart, F.; et al. WikiPathways: Connecting Communities. *Nucleic Acids Res.* **2021**, *49*, D613–D621, doi:10.1093/NAR/GKAA1024.

122. Mudunuri, U.; Che, A.; Yi, M.; Stephens, R.M. BioDBnet: The Biological Database Network. *Bioinformatics* **2009**, *25*, 555–556, doi:10.1093/BIOINFORMATICS/BTN654.

123. Krämer, A.; Green, J.; Pollard, J.; Tugendreich, S. Causal Analysis Approaches in Ingenuity Pathway Analysis. *Bioinformatics* **2014**, *30*, 523–530, doi:10.1093/BIOINFORMATICS/BTT703.

124. Oughtred, R.; Rust, J.; Chang, C.; Breitkreutz, B.J.; Stark, C.; Willems, A.; Boucher, L.; Leung, G.; Kolas, N.; Zhang, F.; et al. The BioGRID Database: A Comprehensive Biomedical Resource of Curated Protein, Genetic, and Chemical Interactions. *Protein Sci.* **2021**, *30*, 187, doi:10.1002/PRO.3978.

125. Guo, K.; Maintainer, B.M. Package "VennDetail" Type Package Title A Package for Visualization and Extract Details Version 1.12.0. **2022**.

126. Conway, J.R.; Lex, A.; Gehlenborg, N. UpSetR: An R Package for the Visualization of Intersecting Sets and Their Properties. *Bioinformatics* **2017**, *33*, 2938–2940, doi:10.1093/BIOINFORMATICS/BTX364.

127. Okuda, S.; Watanabe, Y.; Moriya, Y.; Kawano, S.; Yamamoto, T.; Matsumoto, M.; Takami, T.; Kobayashi, D.; Araki, N.; Yoshizawa, A.C.; et al. JPOSTrepo: An International Standard Data Repository for Proteomes. *Nucleic Acids Res.* **2017**, *45*, D1107–D1111, doi:10.1093/NAR/GKW1080.