

Article

Similarity-based Virtual Screening to Find Antituberculosis Agents based on Novel Scaffolds

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Abstract: A method is developed to identify molecular scaffolds potentially active against the Mycobacterium tuberculosis complex (MTBC). A structurally heterogeneous set of compounds active against MTBC was used to obtain a structural pattern model based on structural invariants. This model was statistically validated through a Leave-n-Out test. It successfully discriminated between active or inactive compounds over 86% in database sets and was also able to select new active chemical structures in external databases. The selection of new substituted pyrimidines, pyrimidones and triazolo[1,5-*a*]pyrimidines was particularly interesting because these structures could provide new scaffolds in this field. The seven selected candidates were synthesized and six of them showed activity *in vitro*.

Keywords: MTBC; virtual screening; topological indices; linear discriminant analysis; pharmacological activity distribution diagrams; antimicrobial drugs; drug design

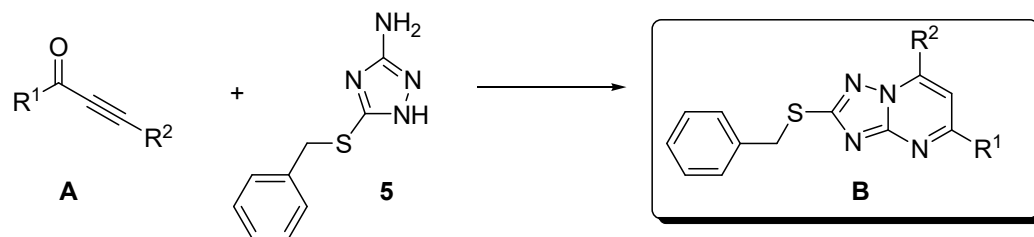
1. Introduction

Tuberculosis is one of the deadliest infections in the world, killing almost 1.45 million people annually [1,2]. Most of these deaths occur in poor countries, and a more rational distribution of wealth could prevent them. However, the increasing incidence of drug-resistant MTBC has caused its resurgence in developed countries and makes therapeutic alternatives necessary [3,4]. Although new compounds potentially active against these bacteria are constantly being sought, very few new treatments have been developed in recent years [5-9]. Therefore, more extensive investigations are needed to find new molecular scaffolds capable of generating new inhibitors of mycobacteria.

The α -acetylenic ketones of type A (Scheme 1) have been shown to be highly versatile building blocks. For instance, these conjugated ynones have proven to be very suitable substrates for the synthesis of (E)-3-acylpropenoic acids [10], and of a wide range of heterocyclic systems [11-21], including the synthesis of the natural product L-lathyrine and related analogues [22,23]. Furthermore, when properly functionalized, compounds of type A have also proven to be valuable substrates for combinatorial and parallel synthesis on solid support of highly molecular diverse 2,4,6-trisubstituted pyrimidines [24-26]. These α -acetylenic ketones are produced readily, mainly by direct palladium-catalyzed coupling of acyl chlorides with 1-alkynes (Sonogashira reaction), [28,29] by reaction of alkynylzinc chlorides with acid halides, [30,31] or by reaction of lithium or magnesium acetylides with aldehydes followed by subsequent oxidation with oxalyl chloride in

DMSO (Swern oxidation) [32], MnO₂ [12,13,15,33], t-butyl hydroperoxide [34] or with IBX [33-35].

In this paper we have further expanded the synthetic usefulness of these conjugated ynones A, and we wish to report the synthesis of novel 2,5,7-substituted triazolo[1,5-*a*]pyrimidines B by cyclocondensation of different α -acetylenic ketones A with 3-amino-5-benzylsulfanyl-1,2,4-triazole 5 (Scheme 1).



Scheme 1

The triazolo[1,5-*a*]pyrimidine nucleus is of considerable chemical and pharmacological interest. Antibiotic, anticholesteremic, antidiabetic, antiallergic, anti-inflammatory, antipyretic, antiphlogistic, analgesic and anticancer activities have been described for these types of compounds. They also serve in the treatment and prevention of circulatory diseases such as hypertension, heart diseases, stroke, hypercholesterol, arteriosclerosis and are an effective coronary vasodilators and bronchodilators [36-49].

On the other hand, several triazolo[1,5-*a*]pyrimidine derivatives have found applications on the agrochemical industry. 1,2,4-triazolo[1,5-*a*]pyrimidinesulfonamides are used as herbicides, and plant growth inhibitors, and they show activity against acetolactate synthase [46,50,51].

Currently, there are many methods available to synthesize triazolo[1,5-*a*]pyrimidines, mainly they are cyclocondensations between 3-amino-1,2,4-triazoles and 1,3-bifunctional synthons such as 1,3-dicarbonyl compounds or their equivalents [52-59], vinylogous iminium salts [47,60,61], ketene dithioacetals [48,62,63], and 3-ketovinyl compounds [46]. We used α -acetylenic Ketones of types A as 1,3-bifunctional synthons.

Although a few pyrimidines [64], pyrimidones [65] and triazolo[1,5-*a*]pyrimidines [66] have already been synthesized and tested for anti-TB activity, their substitution patterns and synthesis methods are different from the compounds described here.

The identification of new targets requires knowledge of the specific biochemical pathways of mycobacteria, but many metabolic processes are still unknown and the structure-based design of new anti-TB agents is a complex task [67].

On the other hand, extra-mechanistic virtual screening methodologies have demonstrated their ability to model the presence of activity within structurally heterogeneous groups of compounds in different therapeutic areas [68-77] as well as in predicting toxicological properties [78,79] and drug-like character [80]. In these models, structural similarity is the key. Molecules are characterized through structural invariants, that is, by descriptors that are independent of molecular conformation. Many of them are topological indices (TI) [81-87], which are capable of characterizing most of the molecular structure [88-95].

The aim of this study was to develop new models based on structural invariants in order to screen structural databases to identify new potentially useful scaffolds against MTBC.

2. Results and Discussion

2.1. Antituberculosis activity modelling

A group of 32 compounds with known activity against MTBC has been compiled from various sources [96-98]. A set of 45 compounds with a different pharmacological activity was also used as inactive group. Each compound was characterized by a set of 90

structural invariants calculated using the DesMol program [99]. The descriptors used with their symbols, definitions and references are shown in table 1. These were used to build a model capable of discriminating between active and inactive antituberculous compounds, as follows.

Linear discriminant analysis functions were calculated with randomly selected subsets of 25 active and 35 inactive compounds by using BMDP New System [103]. Descriptor selection was based on the Fisher-Snedecor F parameter. The variables were introduced step by step in the DF: in each step the variable that added the most to the separation of the groups was entered in the equation, or the variable that contributed the least to improving said separation was eliminated. The classification criterion was the shortest Mahalanobis distance. The discriminant ability of the DF was evaluated by two parameters, Wilks λ , and the percentage of correct classification in each group. The independent variables in this study were the calculated structural invariants, and the discrimination property was the presence of activity against MTBC.

The validation of the selected DF was performed by two methods. One internal leave- n -out test in which the program randomly chose and pulled out a ratio of compounds, and used them to evaluate the DF obtained with the rest; and another external test with a previously unused data set.

To choose the optimal ranges of values for this equation, the corresponding Pharmacological Distribution Diagram, PDD [104], was obtained. These diagrams are useful to determine the intervals of the equation in which the probability of finding new candidates is maximum. This is a graph similar to a histogram in which expectancies appear on the ordinate axis. For an arbitrary range of values of a given function, the activity expectancy is $E_a = a/(i+100)$, where a is the percentage of active compounds in the range and i is the corresponding percentage of inactive within the same range. The expectancy of inactivity is similarly defined as $E_i = i/(a+100)$. This plot provides good visualization of the regions of minimal overlap between the active and inactive compounds and helps to select the optimal interval of the DF.

The 90 descriptors and the selected discriminant function were used as filters to select potential candidates in structural databases. For this, the maximum and minimum values of each descriptor were established as thresholds for the 90 variables, while for DF, the optimal interval was taken. Compounds exhibiting all the 90 values within the thresholds and DF value within the optimal interval were selected as candidates.

The following equation DF was obtained by stepwise LDA using the set shown in table 2:

$$\text{DF} = 0.88 - 11.99 J_1 + 4.86 J_1^v - 11.11 J_3^v + 0.81 {}^1\text{D} - 0.2 {}^4\text{C}_c \quad (1)$$

$$N = 60 \quad \lambda = 0.34 \quad F = 21.12$$

Where $N=60$ represents 25 anti-MTBC drugs and 35 presumably inactive compounds.

Table 1. Descriptors used.

Symbol	Name	Definition	Reference
N	Molecular size	Number of non-hydrogen atoms.	Error! Reference source not found.
V _k k=3,4	Vertices of degree <i>k</i>	Number of atoms having <i>k</i> bonds, σ or π, to non-hydrogen atoms.	Error! Reference source not found.
R	Ramification	Number of single structural branches.	Error! Reference source not found.
W	Wiener path number	Sum of the distances between any two atoms in terms of bonds.	100
L	Length	Maximal distance between atoms in terms of bonds.	Error! Reference source not found.
PR _k k=0-3	Pairs of ramifications at distance <i>k</i>	Number of pairs of single branches at distance <i>k</i> in terms of bonds.	Error! Reference source not found.
${}^k\chi_t$ k=0-4 t=p,c,pc	Randić-like indices of order <i>k</i> and type path (p), cluster (c) and path-cluster (pc)	${}^k\chi_t = \sum_{j=1}^{n_t} \left(\prod_{i \in S_j} \delta_i \right)^{-1/2}$ <p>δ_{<i>i</i>}, number of bonds, σ or π, of the atom <i>i</i> to non-hydrogen atoms. S_{<i>j</i>}, <i>j</i>th sub-structure of order <i>k</i> and type <i>t</i>.</p>	Error! Reference source not found. Error! Reference source not found. Error! Reference source not found.
${}^k\chi_t^v$ k=0-4 t=p,c,pc	Kier-Hall indices of order <i>k</i> and type path (p), cluster (c) and path-cluster (pc)	${}^k\chi_t^v = \sum_{j=1}^{n_t} \left(\prod_{i \in S_j} \delta_i^v \right)^{-1/2}$ <p>δ_{<i>i</i>}^v, Kier-Hall valence of the atom <i>i</i>. S_{<i>j</i>}, <i>j</i>th sub-structure of order <i>k</i> and type <i>t</i>.</p>	Error! Reference source not found. Error! Reference source not found. Error! Reference

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G_k $k=1-5$	Topological charge indices of order k	$G_k = \sum_{i=1}^{N-1} \sum_{j=i+1}^N \mathbf{M}_{ij} - \mathbf{M}_{ji} \delta(k, \mathbf{D}_{ij})$ $\mathbf{M}=\mathbf{A}\mathbf{Q}$, product of the adjacency and inverse squared distance matrices for the hydrogen-depleted molecular graph. \mathbf{D} , distance matrix. δ , Kronecker delta	Error! Reference source not found.,Error! Reference source not found.
G_k^v $k=1-5$	Valence topological charge indices of order k	$G_k^v = \sum_{i=1}^{N-1} \sum_{j=i+1}^N \mathbf{M}_{ij}^v - \mathbf{M}_{ji}^v \delta(k, \mathbf{D}_{ij})$ $\mathbf{M}^v=\mathbf{A}^v\mathbf{Q}$, product of the electronegativity-modified adjacency and inverse squared distance matrices for the hydrogen-depleted molecular graph. \mathbf{D} , distance matrix. δ , Kronecker delta	Error! Reference source not found.,Error! Reference source not found.
J_k $k=1-5$	Pondered topological charge indices of order k	$J_k = \frac{G_k}{N-1}$	Error! Reference source not found.,Error! Reference source not found.
J_k^v $k=1-5$	Pondered valence topological charge indices of order k	$J_k^v = \frac{G_k^v}{N-1}$	Error! Reference source not found.,Error! Reference source not found.
${}^k\mathbf{D}_t$ $k=0-4$ $t=p,c,pc$	Connectivity differences of order k and type path (p), cluster (c) and path-cluster (pc)	${}^k\mathbf{D}_t = {}^k\chi_t - {}^k\chi_t^v$	Error! Reference source not found.
E_k $k=1-5$	Topological charge	$E_k = G_k^v - G_k$	Error! Reference

	differences of order k		source not found.
F_k $k=1-5$	Pondered topological charge differences of order k	$F_k = J_k^v - J_k$	Error! Reference source not found.
${}^k C_t$ $k=0-4$ $t=p,c,pc$	Connectivity quotients of order k and type path (p), cluster (c) and path-cluster (pc)	${}^k C_t = \frac{{}^k \chi_t}{{}^k \chi_t^v}$	Error! Reference source not found.
${}^k Q_t$ $k=0-4$ $t=p,c,pc$	Inverse connectivity quotients of order k and type path (p), cluster (c) and path-cluster (pc)	${}^k Q_t = \frac{{}^k \chi_t^v}{{}^k \chi_t}$	Error! Reference source not found.
CG_k $k=1-5$	Topological charge quotients of order k	$CG_k = \frac{G_k}{G_k^v}$	Error! Reference source not found.
QG_k $k=1-5$	Inverse topological charge quotients of order k	$QG_k = \frac{G_k^v}{G_k}$	Error! Reference source not found.

Table 2. Classification for each compound by DF.

Active Group				Inactive Group			
Compound	DF	Prob. ^a active	Predicted activity ^b	Compound	DF	Prob. ^a inactive	Predicted activity ^b
Streptonicozid	3.14	1.000	+	Butibufen	-3.14	1.000	-
Tobramycin	3.49	1.000	+	Aldicarb	-2.74	1.000	-
Kanamycin	3.88	1.000	+	Antrafenine	-2.55	1.000	-
Amikacin	4.14	1.000	+	Carprofen	-1.89	0.997	-
Dihydrostreptomycin	2.80	0.999	+	Beclobrate	-1.84	0.997	-
Streptomycin	2.32	0.997	+	Benzoctamine	-1.77	0.996	-
Ethambutol	2.23	0.996	+	Carmofur	-1.77	0.996	-
Pyrazinamide	1.97	0.992	+	Aminothiazole	-1.73	0.996	-
Enviomycin	1.92	0.991	+	Acifran	-1.72	0.996	-
Ofloxacin	1.82	0.988	+	Brilliant Blue	-1.72	0.996	-
Moxifloxacin	1.66	0.982	+	Amitraz	-1.68	0.995	-
Gatifloxacin	1.66	0.981	+	Clofibrate	-1.62	0.994	-
Rifampin	1.60	0.978	+	Paraoxon	-1.52	0.992	-

Azithromycin	1.49	0.971	+	Piroxicam	-1.39	0.989	-
Verazide	1.44	0.967	+	Carmustine	-1.11	0.977	-
Isoniazid	1.20	0.938	+	Ornithine	-1.11	0.977	-
Capreomycin	1.16	0.930	+	Alpidem	-1.10	0.976	-
Sparfloxacin	0.98	0.889	+	Alprazolam	-1.04	0.972	-
Clarithromycin	0.93	0.874	+	Bixin	-1.04	0.972	-
Salinazid	0.92	0.871	+	Benzoic Acid	-1.03	0.971	-
Tuberin	0.91	0.869	+	Amsacrine	-0.98	0.967	-
Clofazimine	0.13	0.432	NC	Azaserine	-0.96	0.965	-
Imipenem	0.09	0.404	NC	Theofibrate	-0.88	0.957	-
Ph-Aminosalicylate	-0.200	0.230	-	Azacosterol	-0.83	0.951	-
PAS	-1.000	0.031	-	Allicin	-0.77	0.943	-
				Prazepam	-0.77	0.943	-
				Camazepam	-0.71	0.933	-
				Aminopromazine	-0.57	0.904	-
				Bromazepam	-0.45	0.870	-
				Carnitine	-0.32	0.824	-
				Acipimox	-0.28	0.809	-
				Buspirone	-0.03	0.677	-
				Azapicyl	0.03	0.640	-
				Acronine	0.11	0.588	-
				Captodiamine	0.28	0.470	NC

^a Prob., probability; ^b When probability active or probability inactive >0.60, the compound is classified as active "+" or inactive "-", respectively. In any other case, the compound is considered Non Classified "NC".

The topological descriptors selected in this equation were: the charge indices (J_1 , J_1^v , J_3^v); the difference index ($^1D = ^1\chi - ^1\chi^v$); and the quotient index ($^4C_c = ^4\chi_c / ^4\chi_c^v$), where $^m\chi_t$ and $^m\chi_t^v$ are, respectively, single and valence Randić-Kier-Hall indices of order m and type t . Topological charge-transfer indices, J_1 , J_1^v and J_3^v , are measures of the contribution of molecular topological structure to the charge transfer at topological distance 1 and 3, respectively [68,101]. Difference and quotient indices are related to charge distributions within molecular fragments [68]. Thus, 1D is the net contribution of the heteroatoms to the electronic charge within fragments of order 1 (bonds). The 4C_c index can be related to electron densities of cluster-type fragments of order 4 (three atoms bonded to a central one) in which there is at least one heteroatom.

Table 2 shows the results of the classification for each one of the compounds included in the LDA.

The linear equation gave good results since most compounds were classified with a probability over 86% (table 2). When the probabilities are between 40% and 60%, the compounds were counted as Non Classified (NC), and finally below 40%, they were considered as inactive. Under this framework, the error percentage in the active set was about 9%, whereas in the inactive was 0%.

The results of the internal validation are illustrated in table 4, with the percentage of correct classification within each group. Five runs were performed. A number of compounds ranging from 9 to 16 were randomly extracted from the training to a test set. Wilks values are shown for each equation. Correct classification percentages are shown for

training and test sets, for active and inactive compounds. The number of compounds classified as active (+) or inactive (-) appears in parentheses, where (a/b) = number of (+) compounds / number of (-) compounds. Average values are also shown, as well as the performance of DF function.

The results for the training and test groups are within the same range. The mean percentage of success obtained with the training group for DF (table 3) was 87% for active (+) and 96% for inactive (-). For the test it was 80% and 92%, respectively. The results were similar to those obtained with the DF equation, which points out the validity of the LDA equation.

The results of the external validation test are shown in table 4. As can be seen, for the active set there was a misclassified compound, namely ethionamide. The same is found in the opposite group, where glucosamine was misclassified.

Figure 1 shows the PDD obtained from DF. As can be inferred from Figure 1, the optimal range of DF to find active compounds can be established between 0 and 4.5.

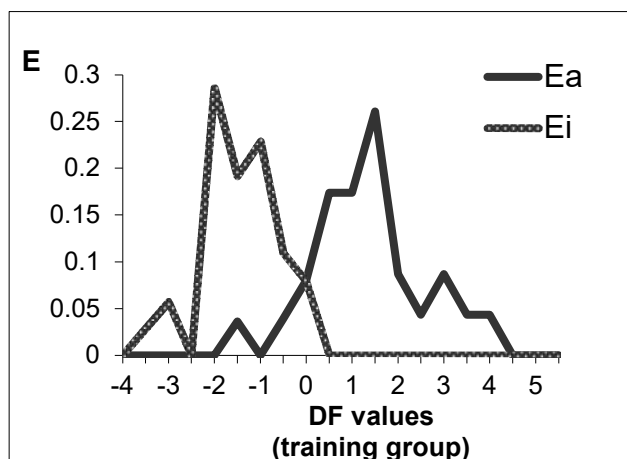
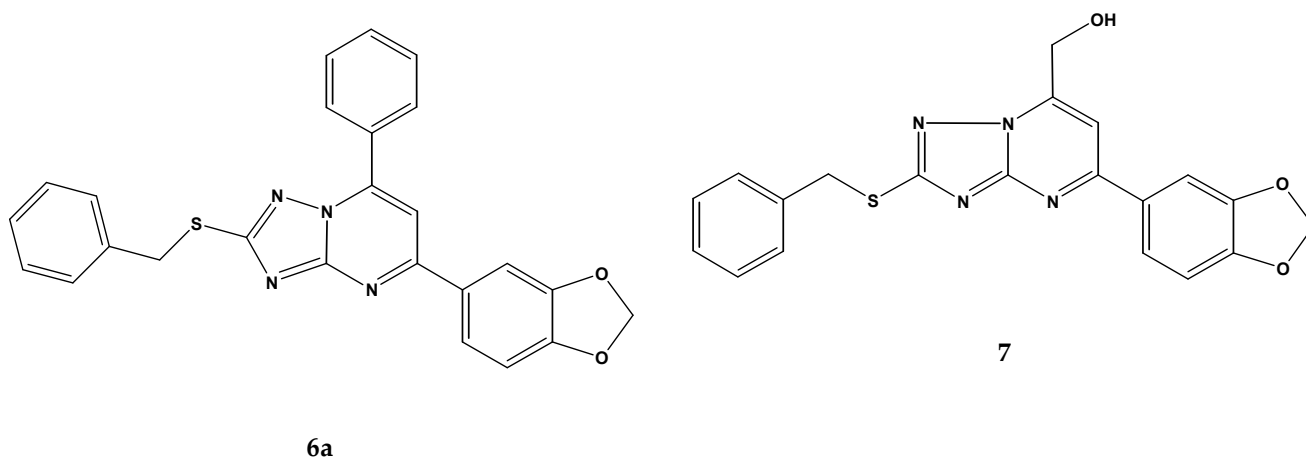


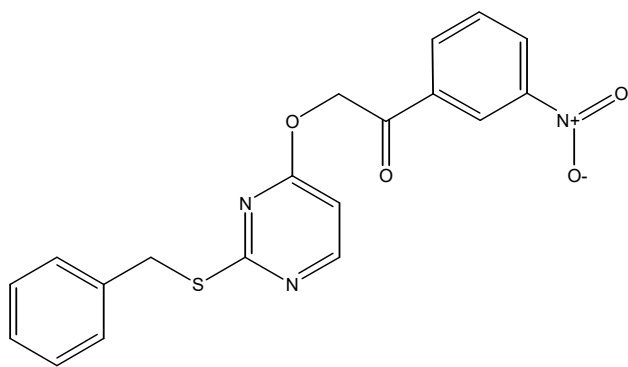
Figure 1. PDD in the training set, obtained by DF equation. E, expectancy of activity or inactivity; black line, active group; grey line, inactive group.

2.2. Similarity-based Virtual screening

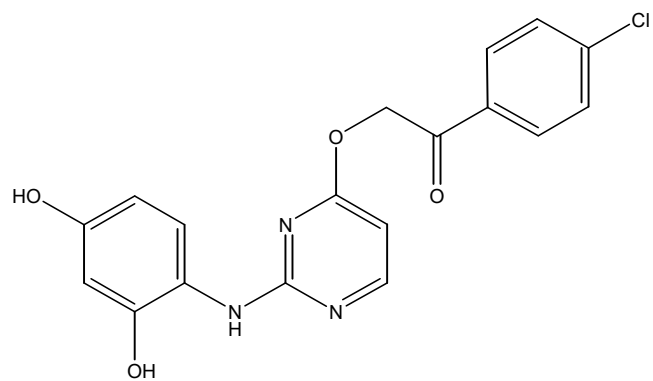
A virtual library containing new pyrimidine derivatives generated by combining the chemical scaffolds and substituents depicted in table 5 was screened by the formalism described above. Compounds selected as possible candidates were synthesized and tested *in vitro*.

After building the database containing the substituted pyrimidines, the virtual screening was performed. Seven potentially active molecules were selected. The structures of the selected compounds are presented in figure 2.

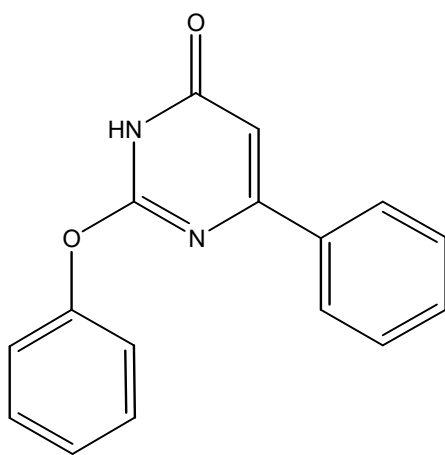




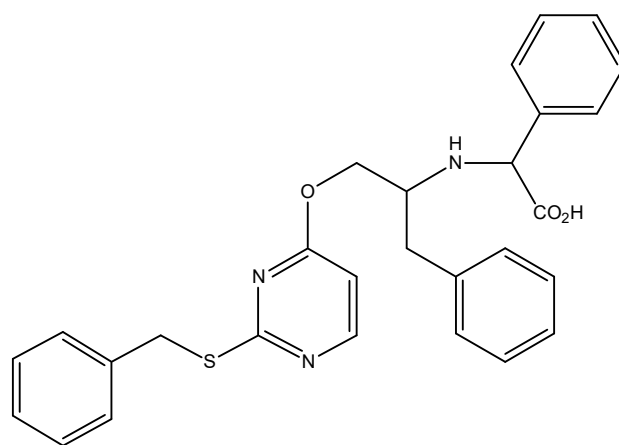
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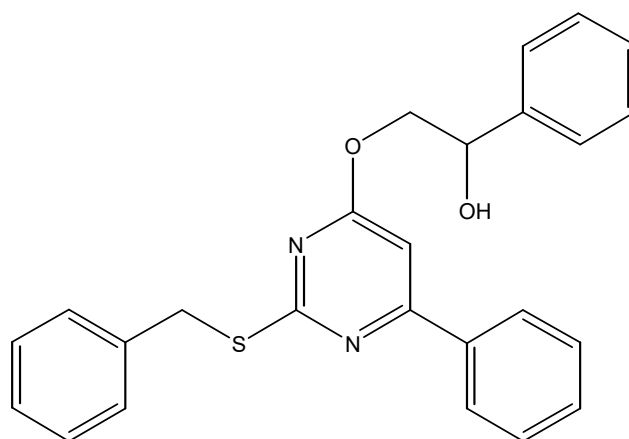
11b



12



13



14

Figure 2. Structures of the selected compounds.

2.3. Chemistry

After building the database containing the substituted pyrimidines, the virtual screening was performed. Seven potentially active molecules were selected. The structures of the selected compounds are presented in figure 2.

Synthesis of 1,2,4-triazolo[1,5-a]pyrimidine derivatives. Reaction of the required magnesium acetylides, generated from alkynes **1a-b** and *i*-PrMgCl in dry THF at 0°C, with piperonal **2**, afforded the expected propargylic alcohols **3a-b**. Oxidation of **3a-b** with MnO₂ afforded α -acetylene ketones **4a** and **4b** in 79% and 37% overall yield, respectively (Scheme 2).

Cyclocondensation of 3-amino-5-benzylsulfanyl-1,2,4-triazole **5** with α -acetylene ketones **4a-b** in dry DMF at 40°C, gave the corresponding triazolo[1,5-a]pyrimidines **6a-b** in moderate yields (73% and 38%, respectively) (Scheme 2).

Finally, deprotection of **6b** in acidic conditions afforded **7** in 77% yield (Scheme 3).

Synthesis of pyrimidine derivatives. Two of us reported on the synthesis of novel 4-alkoxypyrimidines starting from 2-alkylsulfanylpurimidinones of type **8** **Error! Reference source not found.** The method is based on a selective O-alkylation reaction with bulky aliphatic alcohols using the Mitsunobu conditions (method A, Scheme 4) or in basic medium with sterically demanding agents like α -halo-ketones (method B, Scheme 4).

Oxidation of the thioether moiety to the corresponding sulfone **10** using *m*-CPBA and nucleophilic displacement by different nucleophiles produced the corresponding highly molecular diverse pyrimidines of type **11**. In addition, when 4-isopropoxypyrimidine **11a** was treated with a 1:1 mixture of H₂SO₄/AcOH at 90 °C during 15 min. the selective cleavage of the 4-isopropoxy group took place affording to 2-aryloxypyrimidinone **12** (Scheme 5).

On the other hand, the reaction of the deprotected amine **9e** with phenylboronic acid and glyoxylic acid (Petasis reaction) **Error! Reference source not found.** gave the desired α -phenyl glycine derivative **13** in moderated yield (Scheme 6).

Table 3. Results of the *internal validation* for DF.

Run n ^o		Training Group		Test Group	
		(+)	(-)	(+)	(-)
1	0.28	90%(18/2)	100%(0/26)	60%(3/2)	100%(0/9)
2	0.31	87%(20/3)	96%(1/27)	50%(1/1)	100%(0/7)
3	0.35	86%(19/3)	89%(3/24)	100%(3/0)	75%(2/6)
4	0.33	82%(14/3)	100%(0/27)	88%(7/1)	100%(0/8)
5	0.33	91%(20/2)	96%(1/27)	100%(3/0)	86%(1/6)
Average	-	87%	96%	80%	92%
DF	0.34	84%(21/4)	97%(1/34)	No	No

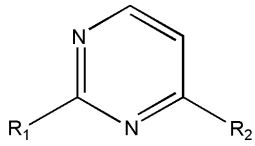
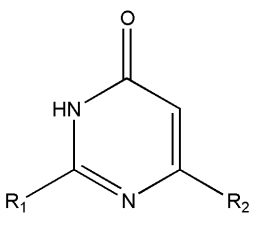
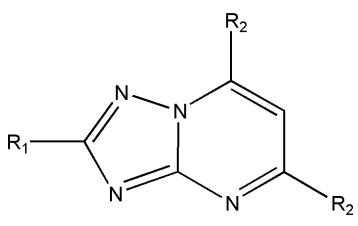

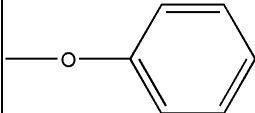
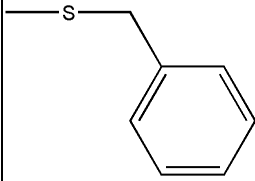
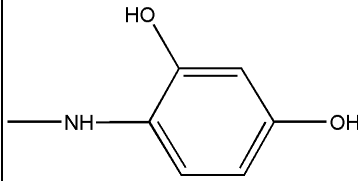

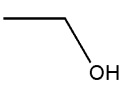

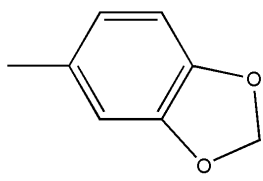
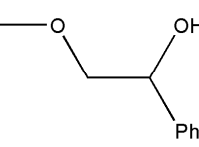
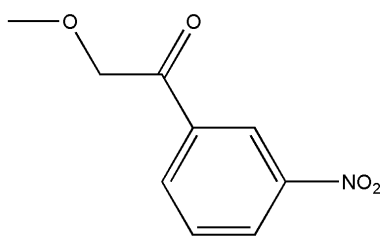
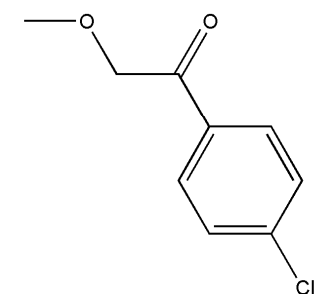
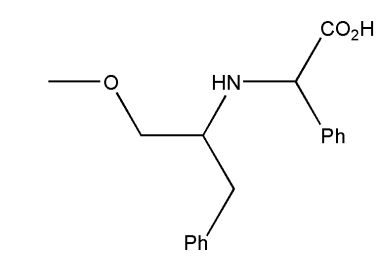
(a/b) = number of (+) compounds / number of (-) compounds.

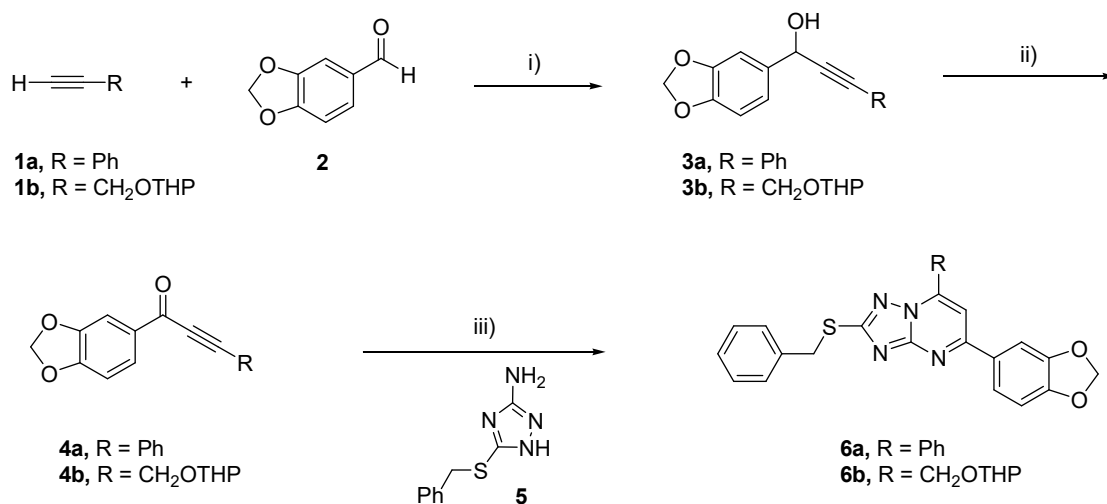
Table 4. Results of the *external validation* for DF.

ACTIVE GROUP				INACTIVE GROUP			
Compound	DF	Prob. ^a active	Predicted activity	Compound	DF	Prob. ^a inactive	Predicted activity
Morphazinamide	4.32	1.000	+	Canthaxanthin	-3.46	1.000	-
Neomycin	5.02	1.000	+	Genite	-2.57	1.000	-
Tubercidin	2.85	0.999	+	Altretamine	-1.91	0.997	-
Ciprofloxacin	2.15	0.995	+	Etifoxin	-1.86	0.997	-
Viomycin	1.75	0.986	+	Dichlone	-1.78	0.996	-
Rifabutin	0.73	0.799	+	Feprazone	-1.47	0.991	-
Ethionamide	-1.30	0.014	-	Antipyrine	-0.44	0.868	-
				Benorylate	-0.25	0.792	-
				Chloropal	0.19	0.526	-
				Glucosamine	1.77	0.014	+

^a Prob., probability; ^b When probability active or probability inactive >0.60, the compound is classified as active “+” or inactive “-”, respectively. In any other case, the compound is considerate Non Classified “NC”.

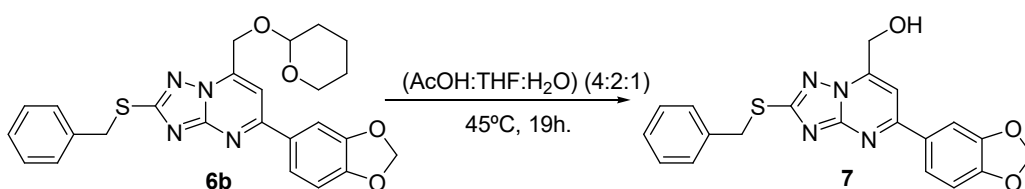
Table 5. Scaffolds and fragments used for the generation of the virtual library.

Chemical scaffolds	R ₁	R ₂
  	   	       

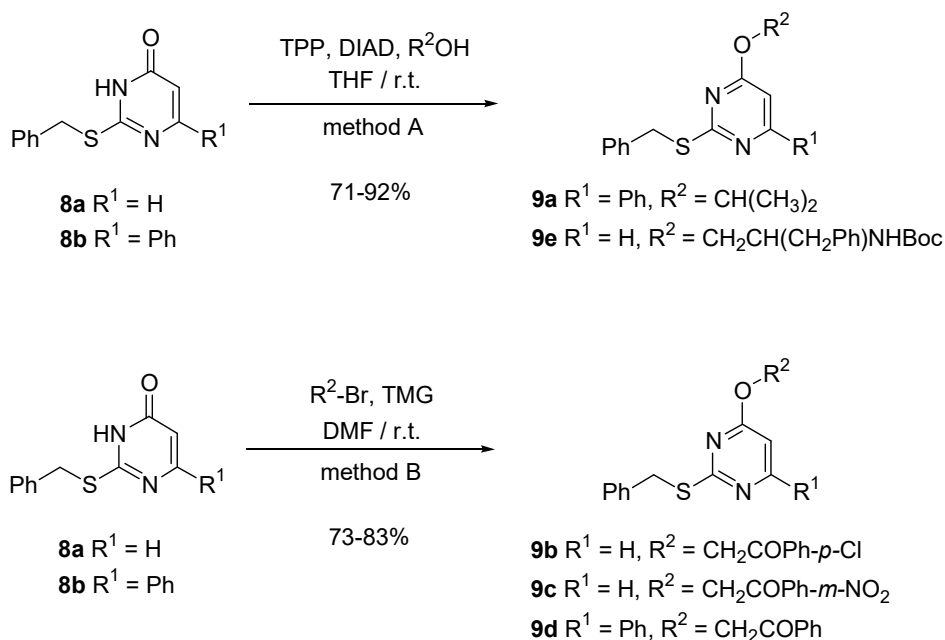


Reagents and conditions: i) *i*-PrMgCl, THF, 0°C, 5-12h., ii) MnO₂, CH₂Cl₂, 0°C-r.t., 3-12h., iii) DBU, DMF, MgSO₄, 40°C, 1-2h.

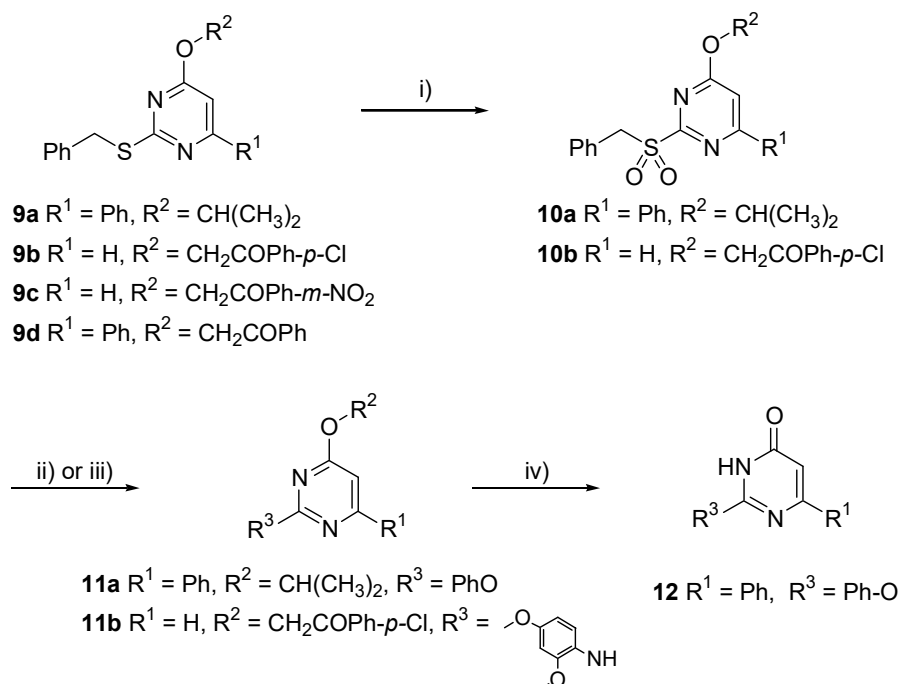
Scheme 2



Scheme 3

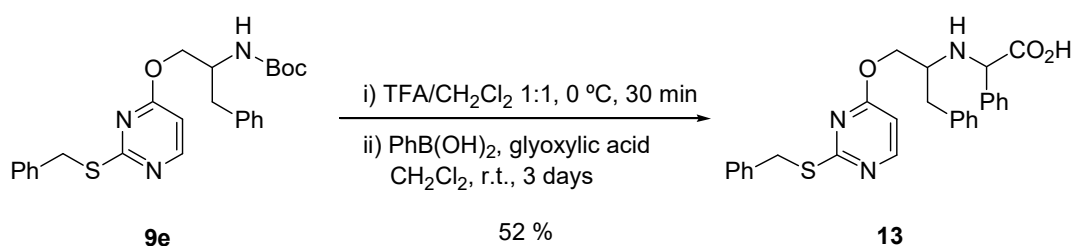


Scheme 4

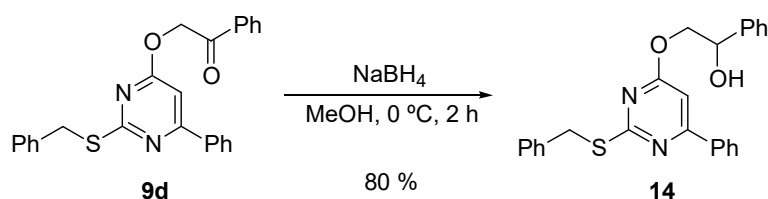


Reagents and conditions: i) *m*-CPBA, CH_2Cl_2 , 0°C to r.t., 2 h; ii) Ar-NH₂, dioxane, reflux, 30 h; iii) PhOH, Cs_2CO_3 , dioxane, 60°C , 3 h; iv) $\text{H}_2\text{SO}_4/\text{AcOH}$ 1:1, 90°C , 15 min

Scheme 5



Scheme 6



Scheme 7

Finally, we prepared the compound **14** by reducing the carbonyl group of compound **9d** with NaBH_4 in MeOH. This reduction afforded the hydroxy derivative **14** in 80% isolated yield (Scheme 7).

Microbiological study. To check the predicted antituberculosis activity of the selected candidates, microbiological tests were performed. The result of *in vitro* susceptibility test of conventional drugs is shown in table 6, and the experimental results of the selected compounds are illustrated in table 7. Two compounds, **7** and **12**, showed

MIC₅₀=MIC₉₀=32 mg/L, corresponding to 81.5 μ M and 121.1 μ M, respectively; four compounds, **6a**, **9c**, **11b** and **13**, showed MIC₉₀=64 mg/L against *M. tuberculosis*, which corresponds to 146, 167.8, 160.1, 131.8 μ M, respectively; and **14** was inactive at the assayed concentrations. The compound **11b** showed the best MIC₅₀ = 80 μ M, but its MIC₉₀ raised to 160.1 μ M. Thus, the results pointed out that **7** was the most active agent, showing molar MIC₉₀ four times lower than Ethambutol but within the same magnitude order.

Table 6. Experimental MIC of conventional drugs against MTBC.

Compound	MIC range / mg/L	MIC ₅₀ / mg/L	MIC ₉₀ / mg/L	Molecular mass / Da	MIC ₅₀ / μ M	MIC ₉₀ / μ M
Ethambutol	1-8	4.0	4.0	204.3	19.6	19.6
Isoniazid	0.05-0.2	0.05	0.05	137.1	0.4	0.4
Rifampin	0.125-1	0.125	0.5	823.0	0.2	0.6
Streptomycin	0.125-0.5	0.25	0.5	581.6	0.4	0.9

MIC range: minimal and maximal inhibitory values found

Table 7. Experimental MIC of the selected compounds against MTBC.

Compound	MIC range / mg/L	MIC ₅₀ / mg/L	MIC ₉₀ / mg/L	Molecular mass / Da	MIC ₅₀ / μ M	MIC ₉₀ / μ M
6a	64	64	64	438.5	146.0	146.0
7	32	32	32	392.4	81.5	81.5
9c	64	64	64	381.4	167.8	167.8
11b	16 - 64	32	64	399.8	80.0	160.1
12	32	32	32	264.3	121.1	121.1
13	32 - 64	64	64	485.6	131.8	131.8
14	>128	>128	>128	414.5	>308.8	>308.8

MIC range: minimal and maximal inhibitory values found

3. Materials and Methods

3.1 Chemical Methods

DMF was dried over activated molecular sieves (4 Å). THF was dried over Na/benzophenone prior use. All the other commercially available chemicals were used as purchased without further purification. Reactions involving magnesium acetylides and synthesis of triazolo[1,5-*a*]pyrimidines were run under a dry Ar atmosphere. Melting points (capillary tube) were measured with an Electrothermal digital melting point apparatus IA 91000 and are uncorrected. IR spectra were recorded on a Mattson-Galaxy Satellite FT-IR. ¹H and ¹³C NMR spectra were recorded at 200 and 50 MHz respectively, on a Bruker DPX200 Advance instrument with TMS as internal standard. MS spectra were recorded on a VG Quattro instrument in the positive ionization FAB mode, using 3-NBA or 1-thioglycerol as the matrix or in a Thermo Quest 2000 series apparatus for the EI (70eV) mode. Analytical TLC was performed on precoated TLC plates, silica gel 60 F₂₅₄ (Merck). Flash-chromatography (FC) purifications were performed on silica gel 60 (230-400 mesh, Merck).

Synthesis of propargylic alcohols 3a-b. General procedure.

To a cooled (0°C) solution of the corresponding alkyne **1a-b** in dry THF (2 mL/mmol), *i*-PrMgCl (2 M solution in THF) was added dropwise under Ar. The mixture was stirred at that temperature for 4 hours. Then, a solution of the piperonal **2** (1.3 equiv) in dry THF (1 mL/mmol) was slowly added dropwise over a period of 15 min. The reaction mixture, under Ar, was stirred from 0°C to r.t. until total consumption of **1a-b** (5-12 h., monitored by TLC). The reaction was quenched with saturated solution of NH₄Cl (3 mL/mmol) at r.t. and the organic solvent was eliminated under reduced pressure. The aqueous layer was extracted with AcOEt (3×3 mL/mmol) and the combined organic layers were dried over

MgSO₄, the solvent was evaporated and the resulting residue purified by flash-chromatography (n-hexane:AcOEt).

1-Benzo[1,3]dioxol-5-yl-3-phenyl-prop-2-yn-1-ol (3a). According to the general procedure described above, reaction between **1a** (4.01 g, 39.26 mmol) and piperonal **2** (7.65 g, 50.9 mmol) afforded 8.02 g (81%) of **3a** as a white solid. M.p.: 60-61°C. IR (KBr): ν 3439 (br., OH). ¹H NMR (CDCl₃): δ 7.5-7.4 (m, 2H_{arom}), 7.3 (m, 3H_{arom}), 7.2-7.1 (m, 2H_{arom}), 6.85 (d, 1H_{arom}, J = 8.0 Hz), 6.00 (s, 2H, OCH₂O), 5.63 (s, 1H, CHC \equiv C), 2.59 (br., 1H, OH). ¹³C NMR (CDCl₃): δ 147.8, 147.6, 134.6 (3s, 3C_{arom}), 131.7, 128.6, 128.3 (3d, 5CH_{arom}), 122.2 (s, C_{arom}), 120.4, 108.1, 107.4 (3d, 3CH_{arom}), 101.2 (t, CH₂), 88.6 (s, C \equiv CPh), 86.5 (s, C \equiv CPh), 64.8 (d, CH). MS (FAB⁺) m/e : 253 ([M+1]⁺, 9). Anal. Calcd for C₁₆H₁₂O₃: C, 76.18; H, 4.79. Found: C, 76.39; H, 4.61%.

1-Benzo[1,3]dioxol-5-yl-3-(tetrahydro-pyran-2-yloxy)-prop-2-yn-1-ol (3b). According to the general procedure described above, reaction between **1b** (0.21 g, 1.48 mmol) and piperonal **2** (0.29 g, 1.92 mmol) afforded the crude of the compound **3b**. The reaction mixture was used in the next step of synthesis without further purification.

Synthesis of α -acetylenic ketones 4a-b. General procedure.

Over a cooled (0°C), mechanically stirred suspension of MnO₂ (5 equiv) in CH₂Cl₂ (3 mL/mmol), a solution of the propargyl alcohol **3a** or of the reaction mixture **3b** in CH₂Cl₂ (3 mL/mmol) was added dropwise. The reaction mixture was stirred from 0°C to r.t. until total consumption of **3a-b** (3-12 h., monitored by TLC) and then filtered through a Celite® pad. The solvents were removed under reduced pressure and the resulting residue purified by flash-chromatography (n-hexane:AcOEt).

1-Benzo[1,3]dioxol-5-yl-3-phenyl-propynone (4a). According to the general procedure described above, reaction between **3a** (1.98 g, 7.87 mmol) and MnO₂ (3.84 g, 39.35 mmol) afforded after crystallization of the residue with MeOH:H₂O instead of the chromatographic purification 1.91 g (97%) of **4a** as a yellow solid. M.p.: 100-101°C. IR (KBr): ν 1625 (s, C=O). ¹H NMR (CDCl₃): δ 7.92 (dd, 1H_{arom}, J = 8.2 Hz, J' = 1.8 Hz), 7.7-7.6 (m, 3H_{arom}), 7.5-7.4 (m, 3H_{arom}), 6.93 (d, 1H_{arom}, J = 8.2 Hz), 6.10 (s, 2H, OCH₂O). ¹³C NMR (CDCl₃): δ 176.0 (s, C=O), 152.8, 148.1 (2s, 2C_{arom}), 132.9 (d, 2CH_{arom}), 131.9 (s, C_{arom}), 130.6, 128.6, 127.2 (3d, 4CH_{arom}), 120 (s, C_{arom}), 108.2, 107.9 (2d, 2CH_{arom}), 102.1 (t, CH₂), 92.3 (s, C \equiv CPh), 86.7 (s, C \equiv CPh). MS (FAB⁺) m/e : 251 ([M+1]⁺, 100). Anal. Calcd for C₁₆H₁₀O₃: C, 76.79; H, 4.03. Found: C, 76.53; H, 4.24%.

1-Benzo[1,3]dioxol-5-yl-4-(tetrahydro-pyran-2-yloxy)-but-2-yn-1-one (4b). According to the general procedure described above, reaction between the reaction mixture of **3b** and MnO₂ (0.72 g, 7.4 mmol) afforded 157 mg (overall yield = 37%) of **4b** as a yellow oil. IR (NaCl): ν 1640 (m, C=O). ¹H NMR (CDCl₃): δ 7.82 (dd, 1H_{arom}, J = 7.4 Hz, J' = 1.8 Hz), 7.55 (d, 1H_{arom}, J = 1.6 Hz), 6.89 (d, 1H_{arom}, J = 8.2 Hz), 6.08 (s, 2H, OCH₂O), 4.9-4.8 (m, 1H, CH), 4.56 (s, 2H, CH₂OThp), 3.9-3.8 (m, 1H, CH₂), 3.6-3.5 (m, 1H, CH₂), 1.9-1.6 (m, 6H, 3CH₂). ¹³C NMR (CDCl₃): δ 175.6 (s, C=O), 152.9, 148.1 (2s, 2C_{arom}), 131.5 (s, C_{arom}), 127.4 (d, CH_{arom}), 108.2, 107.9 (2d, 2CH_{arom}), 102.1 (t, CH₂), 97.3 (d, CH), 89.6 (s, C \equiv CCH₂), 83.3 (s, C \equiv CCH₂), 62.0, 54.0, 30.1, 25.2, 18.8 (5t, 5CH₂). MS (FAB⁺) m/e : 289 ([M+1]⁺, 100). Anal. Calcd for C₁₆H₁₆O₅: C, 66.66; H, 5.59. Found: 66.94; H, 5.76%.

Synthesis of triazolo[1,5-*a*]pyrimidines 6a-b. General procedure.

A mixture of 3-amino-5-benzylsulfanyl-1,2,4-triazole **5**, DBU and anhydrous MgSO₄ (5g/g) in dry DMF (3 mL/mmol) was heated at 40°C under Ar for 30 min. Then, a solution of the corresponding α -acetylenic ketone **4a-b** in dry DMF (3 mL/mmol) was slowly added using a *syringe pump* over a period of 5h. The reaction mixture, under Ar, was stirred at 40°C until total consumption of **5** (1-2 h., monitored by TLC). The reaction was filtered and the organic solvent was eliminated under reduced pressure. The residue was dissolved with CH₂Cl₂ (15 mL/mmol) and was washed with saturated solution of NH₄Cl (3×3 mL/mmol). The organic layer was dried over MgSO₄, the solvent was evaporated and the resulting residue purified by flash-chromatography (n-hexane:AcOEt).

5-Benzo[1,3]dioxol-5-yl-2-benzylsulfanyl-7-phenyl-1,2,4-triazolo[1,5-*a*]pyrimidine (6a). According to the general procedure described above, reaction between **5** (106 mg, 0.52 mmol), **4a** (303 mg, 1.21 mmol, 2.5 equiv), DBU (81 μ l, 0.53 mmol, 1.1 equiv) and

anhydrous MgSO_4 (0.5g) afforded 166 mg (73%) of **6a** as a white solid. M.p.: 143-144°C. ^1H NMR ($\text{DMSO}-d_6$): δ 8.3-7.1 (m, $14H_{\text{arom}}$), 6.21 (s, 2H, OCH_2O), 4.59 (s, 2H, PhCH_2S). ^{13}C NMR ($\text{DMSO}-d_6$): δ 166.4, 159.4, 158.1, 150.1, 148.1, 146.1, 137.7 (7s, $8C_{\text{arom}}$), 131.5 (d, CH_{arom}), 130.1 (s, C_{arom}), 129.7, 128.9, 128.4, 128.3, 127.2, 123.0, 108.5, 107.3, 105.6 (9d, $13\text{CH}_{\text{arom}}$), 101.8, 34.4 (2t, 2CH_2). MS (FAB $^+$) m/e : 439 ($[\text{M}+1]^+$, 100). Anal. Calcd for $\text{C}_{25}\text{H}_{18}\text{N}_4\text{O}_2\text{S}$: C, 68.48; H, 4.14; N, 12.78; S, 7.31. Found: C, 68.29; H, 4.27; N, 13.01; S, 7.02%.

5-Benzo[1,3]dioxol-5-yl-2-benzylsulfanyl-7-(tetrahydro-pyran-2-yloxymethyl)-1,2,4-triazolo[1,5-a]pyrimidine (6b). According to the general procedure described above, reaction between **5** (255 mg, 1.24 mmol), **4b** (698 mg, 2.42 mmol, 2 equiv), DBU (12 μL , 0.082 mmol, 5% mol.) and anhydrous MgSO_4 (1.25g), afforded 225 mg (38%) of **6b** as a yellow solid. M.p.: 62-64°C. ^1H NMR (CDCl_3): δ 7.8-6.9 (m, $9H_{\text{arom}}$), 6.09 (s, 2H, OCH_2O), 5.24 (d, 1H, $J = 17$ Hz, CH_2OThp), 5.01 (d, 1H, $J = 17$ Hz, CH_2OThp), 4.9 (m, 1H, CH), 4.60 (s, 2H, PhCH_2S), 4.0-3.9 (m, 1H, CH_2), 3.7-3.6 (m, 1H, CH_2), 1.9-1.7 (m, 6H, 3CH_2). ^{13}C NMR (CDCl_3): δ 168.3, 160.1, 155.8, 150.4, 148.5, 146.5, 137.2, 130.7 (8s, $8C_{\text{arom}}$), 129.1, 128.5, 127.4, 122.7, 108.4, 107.8, 102.7 (7d, 9CH_{arom}), 101.7 (t, CH_2), 99.14 (d, CH), 62.6, 62.5, 35.6, 30.2, 25.1, 19.3 (6t, 6CH_2). MS (FAB $^+$) m/e : 477 ($[\text{M}+1]^+$, 100). Anal. Calcd for $\text{C}_{25}\text{H}_{24}\text{N}_4\text{O}_4\text{S}$: C, 63.01; H, 5.08; N, 11.76; S, 6.73. Found: C, 63.22; H, 4.84; N, 11.92; S, 6.50%.

Synthesis of (5-benzo[1,3]dioxol-5-yl-2-benzylsulfanyl-1,2,4-triazolo[1,5-a]pyrimidin-7-yl)-methanol (7). A solution of 5-benzo[1,3]dioxol-5-yl-2-benzylsulfanyl-7-(tetrahydro-pyran-2-yloxymethyl)-1,2,4-triazolo[1,5-a]pyrimidine **6b** (99 mg, 0.21 mmol) in ($\text{AcOH}:\text{THF}:\text{H}_2\text{O}$) (4:2:1) (4 mL/mmol, 1 mL) was heated at 45°C for 19 h. The organic solvent was eliminated under reduced pressure, was added saturated solution of NaHCO_3 (3 mL/mmol) and the aqueous layer was extracted with AcOEt (3×3 mL/mmol). The combined organic layers were dried over MgSO_4 , the solvent was evaporated and the resulting residue purified by flash-chromatography (n-hexane: AcOEt) afforded 64 mg (77%) of **7** as a yellow solid. M.p.: 175-177°C. IR (KBr): ν 3199 (br., OH). ^1H NMR ($\text{DMSO}-d_6$): δ 7.9-7.2 (m, $9H_{\text{arom}}$), 6.25 (s, 2H, OCH_2O), 6.17 (t, 1H, $J = 5.5$ Hz, CH_2OH), 5.05 (d, 2H, $J = 5.5$ Hz, CH_2OH), 4.63 (s, 2H, PhCH_2S). ^{13}C NMR ($\text{DMSO}-d_6$): δ 166.6, 159.3, 155.2, 150.5, 150.1, 148.3, 137.7, 130.2 (8s, $8C_{\text{arom}}$), 128.9, 128.4, 127.2, 122.7, 108.7, 107.0, 102.5 (7d, 9CH_{arom}), 101.9, 57.7, 34.4 (3t, 3CH_2). MS (FAB $^+$) m/e : 393 ($[\text{M}+1]^+$, 100). Anal. Calcd for $\text{C}_{20}\text{H}_{16}\text{N}_4\text{O}_3\text{S}$: C, 61.21; H, 4.11; N, 14.28; S, 8.17. Found: C, 60.94; H, 4.36; N, 14.09; S, 8.39%.

Method A for the preparation of the alkoxy pyrimidines 9a and 9e. Mitsunobu reaction.

A solution of DIAD (1.2 equiv) in dry THF (1 mL/mmol) is added dropwise to a solution of Ph_3P (1.2 equiv), the appropriate 2-benzylsulfanylpyrimidinone **8a-b** (1 equiv) and different alcohols in tetrahydrofuran (2 mL/mmol) at room temperature. The reaction mixture was stirred at room temperature until total disappearance of **8a-b** (TLC monitoring). The solvent was evaporated until dryness and the crude product adsorbed over silica purified by flash chromatography (n-hexane: EtOAc).

2-Benzylsulfanyl-4-isopropoxy-6-phenyl-pyrimidine (9a). According to the general procedure described above, reaction between **8b** (500 mg, 1.70 mmol), TPP (675 mg, 2.55 mmol), and DIAD (0.50 mL, 2.55 mmol) in dry THF (6 mL), afforded after 2 h. 526 mg (92%) of **9a** isolated as colourless solid. M.p.: 81-82°C. ^1H NMR (CDCl_3): δ 8.1-8.0 (s, $2H_{\text{arom}}$), 7.7-7.2 (m, $8H_{\text{arom}}$), 6.77 (s, 1H, H_{pyrim}), 5.46 (hept, 1H, $J = 6.2$ Hz, $\text{CH}(\text{CH}_3)_2$), 4.54 (s, 2H, PhCH_2S), 1.38 (d, 6H, $J = 6.2$ Hz, $\text{CH}(\text{CH}_3)_2$). ^{13}C NMR (CDCl_3): δ 170.8, 169.4, 164.6 (3s, $3C_{\text{pyrim}}$), 138.0, 136.8 (2s, $2C_{\text{arom}}$), 130.5, 128.8, 128.7, 128.5, 128.4, 127.0 (6d, $10\text{CH}_{\text{arom}}$), 99.7 (d, CH_{pyrim}), 69.5 (d, CH), 35.4 (t, CH_2), 21.9 (q, 2CH_3). MS (EI) m/e : 336 ($[\text{M}]^+$, 90). Anal. Calcd for $\text{C}_{20}\text{H}_{20}\text{N}_2\text{OS}$: C, 71.40; H, 5.99; N, 8.33; S, 9.53. Found: C, 71.51; H, 6.17; N, 8.14; S, 9.25%.

[1-Benzyl-2-(2-benzylsulfanyl-pyrimidin-4-yloxy)-ethyl]-carbamic acid tert-butyl ester (9e). According to the general procedure described above, reaction between **8a** (1.00 g, 4.59 mmol), TPP (1.58 g, 5.96 mmol), *N*-Boc-phenylalaninol (1.50 g, 5.96 mmol) and DIAD (1.15 mL, 5.96 mmol) in dry THF (15 mL), afforded after 5 h. 1.47 g (71%) of **9e** isolated as colourless solid. M.p.: 108-110°C. IR (KBr): ν 3388 (br., NH), 1685 (s, C=O). ^1H NMR (CDCl_3): δ 8.30 (d, 1H, $J = 4.8$ Hz, H_{pyrim}), 7.5-7.4 (m, $10H_{\text{arom}}$), 6.48 (d, 1H, $J = 4.8$ Hz,

H_{pyrim}), 4.81 (br., 1H, NH), 4.37 (s, 2H, CH_2), 4.30 (s, 2H, CH_2), 4.25 (br., 1H, CH), 2.90 (m, 2H, PhCH_2C), 1.39 (s, 9H, $(\text{CH}_3)_3$). ^{13}C NMR (CDCl_3): δ 171.3, 168.4 (2s, 2C_{pyrim}), 157.5 (d, CH_{pyrim}), 152.2 (s, C=O), 137.8, 137.3 (2s, 2C_{arom}), 129.3, 128.8, 128.5, 127.1, 126.6, 126.4 (6d, $10\text{CH}_{\text{arom}}$), 103.7 (d, CH_{pyrim}), 79.6 (s, C), 64.1 (t, CH_2), 50.8 (d, CH), 37.8, 35.2 (2t, 2CH_2), 28.3 (q, 3CH_3). MS (FAB⁺) m/e : 452 ($[\text{M}+1]^+$, 17). Anal. Calcd for $\text{C}_{25}\text{H}_{29}\text{N}_3\text{O}_3\text{S}$: C, 66.49; H, 6.47; N, 9.31; S, 7.10. Found: C, 66.30; H, 6.68; N, 9.07; S, 6.08%.

Method B for the preparation of the alkoxy pyrimidines 9b-d.

To a solution of the corresponding 2-benzylsulfanylpurimidinone **8a-b** (1 equiv) in DMF (3 mL per mmol), 1.1 equiv of TMG were added. Then, the corresponding phenacyl bromide (1.1 equiv) was added dropwise. The reaction mixture was until total disappearance of **8a-b** (TLC monitoring). The solvent was evaporated until dryness and the crude product adsorbed over silica purified by flash chromatography (n-hexane:EtOAc).

2-(2-Benzylsulfanyl-pyrimidin-4-yloxy)-1-(4-chloro-phenyl)-ethanone (9b). According to the general procedure described above, reaction between **8a** (600 mg, 2.8 mmol), (4-chloro-phenyl)-acetyl bromide (850 mg, 3.6 mmol) and TMG (0.45 mL, 3.6 mmol) in dry DMF (8 mL), afforded after 6 h. 856 mg (83%) of **9b** isolated as colourless solid. M.p.: 111-112°C. IR (KBr): ν 1698 (s, C=O). ^1H NMR (CDCl_3): δ 8.34 (d, 1H, $J = 5.6$ Hz, H_{pyrim}), 7.9-7.3 (m, $9H_{\text{arom}}$), 6.64 (d, 1H, $J = 5.6$ Hz, H_{pyrim}), 5.54 (s, 2H, CH_2O), 4.25 (s, 2H, PhCH_2S). ^{13}C NMR (CDCl_3): δ 191.7 (s, C=O), 171.0, 167.6 (2s, 2C_{pyrim}), 157.8 (d, CH_{pyrim}), 140.3, 137.0, 132.5 (3s, 3C_{arom}), 129.2, 128.5, 128.4, 127.1, (4d, 9CH_{arom}), 103.8 (d, CH_{pyrim}), 67.4, 35.1 (2t, 2CH_2). MS (EI) m/e : 372 ($[\text{M}+2]^+$, 19), 370 ($[\text{M}]^+$, 52). Anal. Calcd for $\text{C}_{19}\text{H}_{15}\text{ClN}_2\text{O}_2\text{S}$: C, 61.53; H, 4.08; N, 7.55; S, 8.65. Found: C, 61.32; H, 3.98; N, 7.76; S, 8.46%.

2-(2-Benzylsulfanyl-pyrimidin-4-yloxy)-1-(3-nitro-phenyl)-ethanone (9c). According to the general procedure described above, reaction between **8a** (1.00 g, 4.6 mmol), (3-nitro-phenyl)-acetyl bromide (1.64 g, 5.7 mmol) and TMG (0.72 mL, 5.7 mmol) in dry DMF (15 mL), afforded after 6 h. 1.31 g (75%) of **9c** isolated as colourless solid. M.p.: 97-98°C. IR (KBr): ν 1705 (s, C=O). ^1H NMR (CDCl_3): δ 8.74 (s, $1H_{\text{arom}}$), 8.46 (d, $1H_{\text{arom}}$, $J = 8.2$ Hz), 8.35 (d, 1H, $J = 5.6$ Hz, H_{pyrim}), 8.24 (d, $1H_{\text{arom}}$, $J = 7.8$ Hz), 7.71 (t, $1H_{\text{arom}}$), 7.3-7.2 (m, $5H_{\text{arom}}$), 6.65 (d, 1H, $J = 5.6$ Hz, H_{pyrim}), 5.60 (s, 2H, CH_2O), 4.26 (s, 2H, PhCH_2S). ^{13}C NMR (CDCl_3): δ 191.1 (s, C=O), 171.1, 167.4 (2s, 2C_{pyrim}), 158.0 (d, CH_{pyrim}), 148.4, 137.0, 135.4 (3s, 3C_{arom}), 133.3, 130.2, 128.5, 128.3, 128.0, 127.1, 122.7 (7d, 9CH_{arom}), 103.8 (d, CH_{pyrim}), 67.5, 35.0 (2t, 2CH_2). MS (EI) m/e : 381 ($[\text{M}]^+$, 18). Anal. Calcd for $\text{C}_{19}\text{H}_{15}\text{N}_3\text{O}_4\text{S}$: C, 59.83; H, 3.96; N, 11.02; S, 8.41. Found: C, 59.57; H, 4.11; N, 11.16; S, 8.14%.

2-(2-Benzylsulfanyl-6-phenyl-pyrimidin-4-yloxy)-1-phenyl-ethanone (9d). According to the general procedure described above, reaction between **8b** (500 mg, 1.7 mmol), phenyl-acetyl bromide (415 mg, 2.0 mmol) and TMG (0.26 mL, 2.0 mmol) in dry DMF (5 mL), afforded after 4 hr. 512 mg (73%) of **9d** isolated as colourless solid. M.p.: 134-135°C. IR (KBr): 1706 (m, C=O). ^1H NMR (CDCl_3): δ 8.1-8.0 (m, $4H_{\text{arom}}$), 7.6-7.5 (m, $6H_{\text{arom}}$), 7.4-7.1 (m, $5H_{\text{arom}}$), 7.05 (s, 1H, H_{pyrim}), 5.67 (s, 2H, OCH_2), 4.39 (s, 2H, PhCH_2S). ^{13}C NMR (CDCl_3): δ 192.9 (s, C=O), 170.7, 168.9, 165.2 (3s, 3C_{pyrim}), 137.5, 136.4, 134.3 (3s, 3C_{arom}), 133.8, 130.7, 128.8, 128.7, 128.6, 128.4, 127.7, 127.1, 127.0 (9d, $15\text{CH}_{\text{arom}}$), 99.1 (d, CH_{pyrim}), 67.7, 35.2 (2t, 2CH_2). MS (EI) m/e : 412 ($[\text{M}]^+$, 25). Anal. Calcd for $\text{C}_{25}\text{H}_{20}\text{N}_2\text{O}_2\text{S}$: C, 72.79; H, 4.89; N, 6.79; S, 7.77. Found: C, 72.57; H, 5.00; N, 6.56; S, 8.09%.

Oxidation of 9a,b to Sulfones 10a,b. General Procedure.

To a cooled (0°C) solution of pyrimidine derivatives **9a,b** (1 equiv) in CH_2Cl_2 (5 mL per mmol), 2.5 equiv of *m*-CPBA (60 % purity) were added in small portions. The mixture was then stirred at 0°C during 2 h., then diluted with CH_2Cl_2 (20 mL per mmol) and washed with aq. satd. NaHCO_3 solution (2×5 mL per mmol) and brine (5 mL per mmol). The separated organic layer was dried (MgSO_4), filtered and evaporated to give a residue which was purified by flash chromatography using n-hexane/EtOAc.

4-Isopropoxy-6-phenyl-2-phenylmethanesulfonyl-pyrimidine (10a). According to the general procedure described above, the reaction of **9a** (450 mg, 1.34 mmol) and *m*-CPBA (1.15 g, 3.34 mmol) in CH_2Cl_2 (7 mL), afforded to **10a** (432 mg, 88%) isolated as colourless solid. M.p.: 124-125°C. ^1H NMR (CDCl_3): δ 8.1-8.0 (m, $2H_{\text{arom}}$), 7.6-7.2 (m, $8H_{\text{arom}}$), 6.99 (s, 1H, H_{pyrim}), 5.65 (hept, 1H, $J = 6.0$ Hz, $\text{CH}(\text{CH}_3)_2$), 4.88 (s, 2H, PhCH_2S), 1.43 (d, 6H,

$J = 6.2$ Hz, $\text{CH}(\text{CH}_3)_2$). ^{13}C NMR (CDCl_3): δ 170.9, 165.5, 164.7 (3s, 3C_{pyrim}), 135.0 (s, C_{arom}), 131.6, 131.3, 128.8, 128.7, 128.2, 127.2 (6d, $10\text{CH}_{\text{arom}}$), 127.1 (s, C_{arom}), 106.0 (d, CH_{pyrim}), 71.6 (d, CH), 57.3 (t, CH_2), 21.7 (q, 2CH_3). MS (EI) m/e : 368 ($[\text{M}]^+$, 67). Anal. Calcd for $\text{C}_{20}\text{H}_{20}\text{N}_2\text{O}_3\text{S}$: C, 65.20; H, 5.47; N, 7.60; S, 8.70. Found: C, 65.29; H, 5.60; N, 7.48; S, 8.88%.

1-(4-Chloro-phenyl)-2-(2-phenylmethanesulfonyl-pyrimidin-4-yloxy)-ethanone (10b). According to the general procedure described above, the reaction of **9b** (675 mg, 1.82 mmol) and *m*-CPBA (1.31 g, 4.55 mmol) in CH_2Cl_2 (9 mL), afforded to **10b** (613 mg, 84%) isolated as colourless solid. M.p.: 141-142°C. IR (KBr): ν 1692 (s, $\text{C}=\text{O}$). ^1H NMR (CDCl_3): δ 8.65 (d, 1H, $J = 5.8$ Hz, H_{pyrim}), 7.92 (d, $2H_{\text{arom}}$, $J = 6.8$ Hz), 7.52 (d, $2H_{\text{arom}}$, $J = 6.8$ Hz), 7.3-7.2 (m, $5H_{\text{arom}}$), 7.12 (d, 1H, $J = 5.6$ Hz, H_{pyrim}), 5.72 (s, 2H, CH_2O), 4.54 (s, 2H, PhCH_2S). ^{13}C NMR (CDCl_3): δ 190.9 (s, $\text{C}=\text{O}$), 169.3, 163.9 (2s, 2C_{pyrim}), 158.3 (d, CH_{pyrim}), 140.8, 132.2 (2s, 2C_{arom}), 131.0, 129.4, 129.2, 128.8, 128.7 (5d, 9CH_{arom}), 126.4 (s, C_{arom}), 111.4 (d, CH_{pyrim}), 68.3, 57.6 (2t, 2CH_2). MS (FAB $^+$) m/e : 405 ($[\text{M}+3]^+$, 24), 403 ($[\text{M}+1]^+$, 62). Anal. Calcd for $\text{C}_{19}\text{H}_{15}\text{ClN}_2\text{O}_4\text{S}$: C, 56.65; H, 3.75; N, 6.95; S, 7.96. Found: C, 56.81; H, 3.54; N, 7.18; S, 8.15%.

Ipso-substitution reaction of pyrimidinyl sulfone derivatives 10a,b.

Synthesis of 4-isopropoxy-2-phenoxy-6-phenyl-pyrimidine (11a). To a solution of phenol (55 mg, 0.57 mmol) in dioxane (2 mL), the Cs_2CO_3 (210 mg, 0.59 mmol) was added. The reaction mixture was stirred at r.t. for 15-20 min. Then the sulfone **10a** (200 mg, 0.54 mmol) was added. After stirring at 60°C for 3 hours the solvent was removed *in vacuo* and the mixture was acidified with 2 *N* hydrochloric acid and extracted with ethyl acetate. The residue was purified using flash chromatography to give **11a** (131 mg, 80 %) as colourless solid. M.p.: 68-69°C. ^1H NMR (CDCl_3): δ 8.0-7.9 (m, $2H_{\text{arom}}$), 7.5-7.3 (m, $8H_{\text{arom}}$), 6.84 (s, 1H, H_{pyrim}), 5.28 (hept, 1H, $J = 6.2$ Hz, $\text{CH}(\text{CH}_3)_2$), 1.34 (d, 6H, $J = 6.2$ Hz, $\text{CH}(\text{CH}_3)_2$). ^{13}C NMR (CDCl_3): δ 171.9, 166.4, 165.0 (3s, 3C_{pyrim}), 153.2, 136.5 (2s, 2C_{arom}), 130.6, 129.1, 128.7, 127.0, 124.8, 121.9 (6d, $10\text{CH}_{\text{arom}}$), 98.6 (d, CH_{pyrim}), 69.9 (d, CH), 21.8 (q, 2CH_3). MS (EI) m/e : 306 ($[\text{M}]^+$, 24). Anal. Calcd for $\text{C}_{19}\text{H}_{18}\text{N}_2\text{O}_2$: C, 74.49; H, 5.92; N, 9.14. Found: C, 74.36; H, 5.75; N, 9.40%.

Synthesis of 1-(4-chloro-phenyl)-2-[2-(2,4-dimethoxy-phenylamino)-pyrimidin-4-yloxy]-ethanone (11b). To a solution of the sulfone **10b** (125 mg, 0.31 mmol) in dioxane (2 mL), the 2,4-dimethoxyaniline (100 mg, 0.62 mmol) was added. The reaction mixture with good stirring was heated at 100°C until total consumption of the starting material (30 h., TLC monitoring). The solvent was removed under reduced pressure and the residue purified by flash-chromatography (n-hexane:EtOAc) to give **11b** (25 mg, 20 %) as an orange crystalline solid. M.p.: 122-123°C. IR (KBr): ν 3244 (br., NH), 1705 (m, $\text{C}=\text{O}$). ^1H NMR (CDCl_3): δ 8.20 (d, 1H, $J = 5.4$ Hz, H_{pyrim}), 7.96 (d, $2H_{\text{arom}}$, $J = 8.2$ Hz), 7.77 (d, $1H_{\text{arom}}$, $J = 8.8$ Hz), 7.54 (d, $2H_{\text{arom}}$, $J = 8.2$ Hz), 7.30 (s, $1H_{\text{arom}}$), 6.44 (d, $1H_{\text{arom}}$, $J = 2.4$ Hz), 6.36 (d, 1H, $J = 5.2$ Hz, H_{pyrim}), 5.90 (d, 1H, $J = 8.2$ Hz, NH), 5.58 (s, 2H, CH_2O), 3.83 (s, 3H, CH_3O), 3.72 (s, 3H, CH_3O). ^{13}C NMR (CDCl_3): δ 192.2 (s, $\text{C}=\text{O}$), 168.5, 159.6 (2s, 2C_{pyrim}), 158.6 (d, CH_{pyrim}), 155.4, 149.7, 140.2, 132.9 (4s, 4C_{arom}), 129.3, 129.2 (2d, 4CH_{arom}), 121.9 (s, C_{arom}), 120.3, 102.7 (2d, 2CH_{arom}), 98.6 (d, CH_{pyrim}), 98.5 (d, CH_{arom}), 67.3 (t, CH_2), 55.6, 55.4 (2q, 2CH_3). MS (EI) m/e : 401 ($[\text{M}+2]^+$, 17), 399 ($[\text{M}]^+$, 52). Anal. Calcd for $\text{C}_{20}\text{H}_{18}\text{ClN}_3\text{O}_4$: C, 60.08; H, 4.54; N, 10.51. Found: C, 60.29; H, 4.48; N, 10.32%.

Removal of the 4-isopropoxy group. Synthesis of 2-phenoxy-6-phenyl-3H-pyrimidin-4-one (12). The 4-isopropoxypyrimidine **11a** (87 mg, 0.28 mmol) was added to a mixture of AcOH (0.6 mL) and con. H_2SO_4 (0.6 mL). The reaction mixture was stirred at 90°C for 15 min. After cooling, the mixture was neutralised with aq. 5 *N* NaOH and extracted with CH_2Cl_2 (3×5 mL). The combined organic layers were washed with brine and the separated organic layer was dried (MgSO_4), filtered and eliminated under reduced pressure to afford the pure pyrimidinone **12** (59 mg, 80 %) as colourless solid. M.p.: 263-264°C. IR (KBr): ν 3060-2750 (br., NH), 1669 (s, $\text{C}=\text{O}$). ^1H NMR ($\text{DMSO}-d_6$): δ 12.75 (br., NH), 7.9-7.8 (m, $2H_{\text{arom}}$), 7.6-7.5 (m, $8H_{\text{arom}}$), 6.74 (s, 1H, H_{pyrim}). ^{13}C NMR ($\text{DMSO}-d_6$): δ 166.0, 161.4, 158.7 (3s, 3C_{pyrim}), 151.8, 135.8 (2s, 2C_{arom}), 130.6, 129.5, 128.8, 126.6, 125.7, 121.7 (6d, $10\text{CH}_{\text{arom}}$), 102.4 (d, CH_{pyrim}). MS (EI) m/e : 264 ($[\text{M}]^+$, 54). Anal. Calcd for $\text{C}_{16}\text{H}_{12}\text{N}_2\text{O}_2$: C, 72.72; H, 4.58; N, 10.70. Found: C, 72.47; H, 4.81; N, 10.86%.

Synthesis of [1-benzyl-2-(2-benzylsulfanyl-pyrimidin-4-yloxy)-ethylamino]-phenyl-acetic acid (**13**): Petasis reaction.

To a stirred solution of glyoxylic acid monohydrate (74 mg, 0.78 mmol) in dichloromethane (5 mL) was added the primary amine **9e**, previous deprotection of Boc using standard conditions, (275 mg, 0.78 mmol), followed by phenylboronic acid (99 mg, 0.78 mmol). After the flask was purged with nitrogen and sealed, the reaction mixture was stirred vigorously at room temperature for 3 days. The resulting precipitate was isolated by filtration and washed with dichloromethane to give the pure pyrimidinone **13** (197 mg, 52 %) as colourless solid. M.p.: 134-135°C. IR (KBr): ν 3217 (br, NH), 1713 (s, C=O). ^1H NMR (DMSO- d_6): δ 8.42 (d, 1H, J = 5.8 Hz, H_{pyrim}), 7.9-7.8 (m, 2H, NH + COOH), 7.5-7.2 (m, 15 H_{arom}), 6.69 (d, 1H, J = 5.8 Hz, H_{pyrim}), 4.63 (s, 1H, CHCOOH), 4.34 (s, 2H, PhCH₂S), 4.31 (dd, 1H, J = 4.0 Hz, J' = 11.0 Hz, OCH₂), 4.17 (dd, 1H, J = 6.0 Hz, J' = 11.0 Hz, OCH₂), 3.15 (br., 1H, PhCH₂CH), 3.02 (dd, 1H, J = 4.8 Hz, J' = 13.6 Hz, PhCH₂CH), 2.85 (dd, 1H, J = 7.5 Hz, J' = 13.6 Hz, PhCH₂CH). ^{13}C NMR (DMSO- d_6): δ 172.6, 170.0, 168.0 (3s, 3C), 158.0 (d, CH_{pyrim}), 137.8, 137.7 (2s, 3C_{arom}), 134.1, 130.0, 129.2, 128.8, 128.4, 127.8, 127.3, 127.0, 126.4 (9d, 15CH_{arom}), 104.1 (d, CH_{pyrim}), 66.9 (t, CH₂), 62.4, 55.5 (2d, 2CH), 36.4, 34.2 (2t, 2CH₂). MS (FAB⁺) m/e : 486 ([M+1]⁺, 28). Anal. Calcd for C₂₈H₂₇N₃O₃S: C, 69.25; H, 5.60; N, 8.65; S, 6.60. Found: C, 69.52; H, 5.84; N, 8.48; S, 6.31%.

Synthesis of 2-(2-benzylsulfanyl-6-phenyl-pyrimidin-4-yloxy)-1-phenyl-ethanol (14**).** To a stirred and cooled (0 °C) solution of pyrimidinone **9d** (500 mg, 1.21 mmols) in MeOH (6 mL) was added NaBH₄ (165 mg, 4.24 mmols) in small portions while stirring (vigorous evolution of gas observed). Stirring was continued for 2 h. at 0°C. The solution was evaporated to dryness, and the crude residue was partitioned between EtOAc (10 mL) and aq satd NH₄Cl solution (15 mL). The organic layer was separated, washed with H₂O (5 mL), dried (MgSO₄), and evaporated to give a residue which was purified by flash chromatography using hexanes/EtOAc to afford pure (**14**) as a colourless solid (403 mg, 80%). M.p.: 140-141°C. IR (KBr): 3400 (br., OH). ^1H NMR (CDCl₃): δ 8.1-8.0 (m, 2 H_{arom}), 7.5-7.3 (m, 13 H_{arom}), 6.90 (s, 1H, H_{pyrim}), 5.17 (dd, 1H, J = 8.4 Hz, J' = 3.2 Hz, PhCH), 4.53 (s, 2H, PhCH₂S), 4.47 (dd, 1H, J = 11.6 Hz, J' = 8.4 Hz, CH₂O), 4.03 (dd, 1H, J = 11.4 Hz, J' = 3.2 Hz, CH₂O), 2.90 (br., 1H, OH). ^{13}C NMR (CDCl₃): δ 170.9, 169.6, 165.1 (3s, 3C_{pyrim}), 139.8, 137.7, 136.4 (3s, 3C_{arom}), 130.8, 128.8, 128.7, 128.6, 128.5, 128.4, 128.2, 127.1, 126.2 (9d, 15CH_{arom}), 99.1 (d, CH_{pyrim}), 72.5 (d, CH), 71.7, 35.4 (2t, 2CH₂). MS (EI) m/e : 414 ([M]⁺, 33). Anal. Calcd for C₂₅H₂₂N₂O₂S: C, 72.44; H, 5.35; N, 6.76; S, 7.74. Found: C, 72.62; H, 5.14; N, 6.52; S, 7.46%.

3.2 Microbiological Methods

Thirty-two isolates of *M. tuberculosis* from respiratory (n=21) and non-respiratory (n=11) clinical samples, identified by conventional methods **Error! Reference source not found.** and by DNA hybridization probe (Accuprobe® Gen Probe Inc., San Diego, California) were selected from the laboratory collection of Dept. Microbiología, Hospital Clínico Universitario. Valencia, Spain. Susceptibility to first-line antituberculosis drugs (ethambutol, isoniazid, rifampin and streptomycin) was tested by a fluorometric method (Bactec® MGIT 960, Becton-Dickinson) and by a microdilution method **Error! Reference source not found.**

The organisms were grown in modified Middlebrook 7H9 broth supplemented with 10% OADC enrichment (Difco Laboratories) for seven days at 37°C. The inoculum size was obtained by dilution of *M. tuberculosis* isolates suspensions in 7H9 broth to yield an absorbance equivalent to that of a MacFarland n° 0.5 standard.

Antimicrobial susceptibility test was performed in 96-well microplates using serial twofold microdilution in 7H9 broth. Initial drug dilutions were prepared in deionized water or, if no soluble, dimethyl sulfoxide. Subsequent twofold dilutions were performed in 150 μL of modified 7H9 broth in the microplates to provide a final test range of 128 to 0.125 mg/L. Ten μL of a suspension of mycobacteria were added to the wells. Plates were covered with Parafilm "M"® (Laboratory Film, American national Can TM), and incubated for 12 days at 37°C. Starting at 13 day of incubation, 20 μL of Resazurina® (Sigma 2127) with a concentration of 250 mg/L were added to the wells, and the microplates were

reincubated at 37°C for an additional period of 48h. MIC₅₀ and MIC₉₀ were determined as the lowest concentrations of the compounds yielding no visible changes from blue to pink
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4. Conclusions

New chemical scaffolds have been identified that could render new lead drugs in this field, by using easy to calculate descriptors, such as structural invariants. The only sub-structure common to all the selected molecules is the pyrimidine ring, and the most frequent substituent is the benzylsulfanyl in position 2. The 1,2,4-triazolo[1,5-*a*]pyrimidine system is present in structures **7** and **6a**, which have the same substituted groups in 2 and 5. Only one pyrimidone was selected (**12**). All these structures are proposed as new base structures in order to design new combinatorial synthesis projects.

Although several action mechanisms have been considered in the training group the validity of the approach is not negligible. One possible explanation to this fact could be that the equation retains the structural features involved in all the mechanisms considered. In opinion of the authors, this approach is not able to find new action mechanisms, but it is possible to obtain new unexpected molecular structures acting through the known mechanisms which combine properties of the known compounds. This feature could be useful in order to avoid problems of resistance.

Supplementary Materials: Table with all the thresholds for the descriptors and DF (Eq. 1), Representative spectra.

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References

1. Tuberculosis <https://www.who.int/westernpacific/health-topics/tuberculosis> (accessed Jun 25, 2022).
2. Dhote, A.; Kawishwar, V. Utility of Concentration Bleach Method and Fluorescent Auramine Rhodamine Staining in Diagnosis of *Mycobacterium tuberculosis* on Formalin Fixed Tissue. *New Horizons in Medicine and Medical Research Vol. 4* **2022**, 67–70.
3. Besalú, E.; Ponec, R.; Julián-Ortiz, J. V. de Virtual generation of agents against *Mycobacterium tuberculosis*. A QSAR study. *Mol. Divers.* **2003**, *6*, 107–120.
4. Acharya, B.; Acharya, A.; Gautam, S.; Ghimire, S. P.; Mishra, G.; Parajuli, N.; Sapkota, B. Advances in Diagnosis of Tuberculosis: An Update into Molecular Diagnosis of *Mycobacterium Tuberculosis*. *Molec. Biol. Rep.* **2020**, *47* (5), 4065–4075.
5. Hoffner, S. E.; Gezelius, L.; Olsson-Liljequist, B. *In vitro* activity of fluorinated quinolones and macrolides against drug-resistant *Mycobacterium tuberculosis*. *J. Antimicrob. Chemother.* **1997**, *40*, 885–888.
6. Tomioka, H. Prospects for development of new antituberculous drug. *Kekkaku.* **2002**, *77*, 573–584.
7. Koketsu, M.; Tanaka, K.; Takenaka, Y.; Kwong, C. D.; Ishihara, H. Synthesis of 1,3-thiazine derivatives and their evaluation as potential antimycobacterial agents. *Eur. J. Pharm. Sci.* **2002**, *15*, 307–310.
8. dos Santos Fernandes, G.; Jornada, Paula, D.; Souza, C.; Chin, C.; Pavan, F.; dos Santos, J. Current Advances in Antitubercular Drug Discovery: Potent Prototypes and New Targets. *Curr. Med. Chem.* **2015**, *22* (27), 3133–3161.
9. Fernandes, G. F.; Thompson, A. M.; Castagnolo, D.; Denny, W. A.; Dos Santos, J. L. Tuberculosis Drug Discovery: Challenges and New Horizons. *J. Med. Chem.* **2022**, *65* (11), 7489–7531.
10. Obrecht, D.; Weiss, B. A new method for the preparation of (*E*)-3-acylprop-2-enoic acids. *Helv. Chim. Acta* **1989**, *72*, 117–122.
11. Utimoto, K.; Miwa, H.; Nozaki, H. Palladium-catalyzed synthesis of pyrroles. *Tetrahedron Lett.* **1981**, *22*, 4277–4278.
12. Obrecht, D. Acid-catalyzed cyclization reactions of substituted acetylenic ketones: a new approach for the synthesis of 3-halofurans, flavones and styrylchromones. *Helv. Chim. Acta* **1989**, *72*, 447–456.
13. Masquelin, T.; Obrecht, D. A facile preparation of 2- and 5-substituted 3-bromothiophenes. *Tetrahedron Lett.* **1994**, *35*, 9387–9390.
14. Garvey, D.S.; Wasicak, J.T.; Elliot, R.L.; Lebold, S.A.; Hettinger, A.-M.; Carrera, G.M.; Lin, N.-H.; He, Y.; Holladay, M.W.; Anderson, D.J.; Cadman, E.D.; Raszkiewicz, J.L.; Sullivan, J.P.; Arneric, S.P. Ligands for brain cholinergic channel receptors: synthesis and in vitro characterization of novel isoxazoles and isothiazoles as bioisosteric replacements for the pyridine ring in nicotine. *J. Med. Chem.* **1994**, *37*, 4455–4463.

15. Masquelin, T.; Obrecht, D. A new approach to the synthesis of N-protected 2- and 5-substituted 3-halopyrroles. *Synthesis* **1995**, 276-384.
16. Degl'Inocenti, A.; Scafato, P.; Capperucci, A.; Bartoletti, L.; Mordini, A.; Reginato, G. Azide cyclocondensation with acetylenic silyl ketone: a general access to functionalized-1,2,3-triazolylacylsilanes and -aldehydes. *Tetrahedron Lett.* **1995**, 36, 9031-9034.
17. Masquelin, T.; Obrecht, D. A novel access to 2,4-substituted quinolines from acetylenic ketones. *Tetrahedron* **1997**, 53, 641-646.
18. Falorni, M.; Giacomelli, G.; Spanedda, A.M. Synthesis of chiral pyrazoles and isoxazoles as constrained amino acids. *Tetrahedron: Asymmetry* **1998**, 9, 3039-3046.
19. Cabarrocas, G.; Rafel, S.; Ventura, M.; Villalgordo, J.M. A new approach toward the stereoselective synthesis of novel quinolyl glycines: Synthesis of the enantiomerically pure quinolyl- β -amino alcohol precursors. *Synlett* **2000**, 5, 595-598.
20. Cabarrocas, G.; Ventura, M.; Maestro, M.; Mahía José; Villalgordo, J. M. Synthesis of Novel Optically Pure Quinolyl- β -Amino Alcohol Derivatives from 2-Amino Thiophenol and Chiral α -Acetylenic Ketones and Their IBX-Mediated Oxidative Cleavage to N-BOC Quinolyl Carboxamides. *Tetrahedron: Asymmetry* **2001**, 12 (13), 1851-1863.
21. Satheesha Rai, N.; Kalluraya, B.; Lingappa, B.; Shenoy, S.; Puranic, V. G. Convenient Access to 1,3,4-Trisubstituted Pyrazoles Carrying 5-Nitrothiophene Moiety via 1,3-Dipolar Cycloaddition of Sydnone with Acetylenic Ketones and Their Antimicrobial Evaluation. *Eur. J. Med. Chem.* **2008**, 43 (8), 1715-1720.
22. Harigae, R.; Moriyama, K.; Togo, H. Preparation of 3,5-Disubstituted Pyrazoles and Isoxazoles from Terminal Alkynes, Aldehydes, Hydrazines, and Hydroxylamine. *J. Org. Chem.* **2014**, 79 (5), 2049-2058.
23. Adlington, R. M.; Baldwin, J. E.; Catterick, D.; Pritchard, G. J. The Synthesis of Pyrimidin-4-Yl Substituted α -Amino Acids. A Versatile Approach from Alkynyl Ketones. *J. Chem. Soc., Perkin Trans. 1* **1999**, No. 8, 855-866.
24. Nájera, C.; Sydnes, L. K.; Yus, M. Conjugated Ynones in Organic Synthesis. *Chem. Rev.* **2019**, 119 (20), 11110-11244.
25. Chucholowski, A.; Masquelin, T.; Obrecht, D.; Stadlwieser, J.; Villalgordo, J. M. Cheminform Abstract: Novel Solution- and Solid-Phase Strategies for the Parallel and Combinatorial Synthesis of Low-Molecular-Weight Compound Libraries. *ChemInform* **2010**, 28 (13).
26. Obrecht, D.; Abrecht, C.; Grieder, A.; Villalgordo, J.M. A novel and efficient approach for the combinatorial synthesis of structurally diverse pyrimidines on solid support. *Helv. Chim. Acta* **1997**, 80, 65-72.
27. Aparna, E. P.; Devaky, K. S. Advances in the Solid-Phase Synthesis of Pyrimidine Derivatives. *ACS Comb. Sci.* **2019**, 21 (2), 35-68.
28. Sonogashira, K.; Tohda, Y.; Hagihara, N. Convenient synthesis of acetylenes. Catalytic substitutions of acetylenic hydrogen with bromo alkenes, iodo arenes and bromopyridines. *Tetrahedron Lett.* **1975**, 50, 4467-4470.
29. Tohda, Y.; Sonogashira, K.; Hagihara, N. A convenient synthesis of 1-alkynyl ketones and 2-alkynamides. *Synthesis* **1977**, 777-778.
30. Corriu, R. J. P.; Bolin, G.; Iqbal, J.; Moreau, J. J. E.; Vernhet, C. Aminosilanes in organic synthesis. Addition of organocopper reagents on γ -bis(trimethylsilyl)amino- α -acetylenic amides, esters and ketones. Stereochemistry and some synthetic uses. *Tetrahedron* **1993**, 49, 4603-4618.
31. Verkruijsse, H.D.; Heus-Kloos, Y.A.; Brandsma, L. Efficient methods for the preparation of acetylenic ketones. *J. Organomet. Chem.* **1988**, 338, 289-294.
32. Larson, D.P.; Heathcock, C.H. Total synthesis of tricolorin A. *J. Org. Chem.* **1997**, 62, 8406-8418.
33. Palombi, L.; Arista, L.; Lattanzi, A.; Bonadies, F.; Scettri, Q. Zeolite-catalyzed oxidation of benzylic and acetylenic alcohols with *t*-butyl hydroperoxide. *Tetrahedron Lett.* **1996**, 37, 7849-7850.
34. Serrat, X.; Cabarrocas, G.; Rafel, S.; Ventura, M.; Linden, A.; Villalgordo, J.M. A highly efficient and straightforward stereoselective synthesis of novel chiral α -acetylenic ketones. *Tetrahedron: Asymmetry* **1999**, 10, 3417-3430.
35. Frigerio, M.; Santagostino, M.; Sputore, S.; Palmisano, G. Oxidation of alcohols with *o*-iodoxybenzoic acid in DMSO: a new insight into an old hypervalent iodine reagent. *J. Org. Chem.* **1995**, 60, 7272-7276.
36. El Ashry, E.S.H.; Rashed, N. 1,2,4-triazolo- and tetrazolo[x,y-z]pyrimidines. *Adv. Heterocycl. Chem.* **1999**, 72, 127-171.
37. El Ashry, E.S.H.; El Kilany, Y.; Rashed, N. Assafir, H.R. Dimroth rearrangement: translocation of heteroatoms in heterocyclic rings and its role in ring transformations of heterocycles. *Adv. Heterocycl. Chem.* **1999**, 75, 79-167.
38. Pugmire, R.J.; Smith, J.C.; Grant, D.M.; Stanovnik, B.; Tisler, M.; Vercek, B. Correlation of ring nitrogen substituents with carbon-13 nuclear magnetic resonance data in azoloazines. *J. Heterocycl. Chem.* **1987**, 24, 805-809.
39. Salas, J.M.; Romero, M.A.; Sánchez, M.P.; Quirós, M. Metal complexes of 1,2,4-triazolo[1,5-*a*]pyrimidine derivatives. *Coord. Chem. Review*, **1999**, 1119-1142.
40. Sato, Y.; Shimoji, Y.; Fujita, H.; Nishino, H.; Mizuno, H.; Kobayashi, S.; Kamakura, S. Studies on cardiovascular agents. 6. Synthesis and coronary vasodilating and antihypertensive activities of 1,2,4-triazolo[1,5-*a*]pyrimidines fused to heterocyclic systems. *J. Med. Chem.* **1980**, 23, 927-937.
41. Tenor, E.; Ludwig, R. Pharmaceutical-chemical research on *s*-triazolo[1,5-*a*]pyrimidines. *Pharmazie* **1971**, 26, 534-539.
42. Navarro, J.A.R.; Salas, J.M.; Romero, M.A.; Faure, R. Influence of anions and crystallization conditions on the solid-state structure of some binuclear silver (I) complexes supported by triazolopyrimidine bridges. *J. Chem. Soc., Dalton Trans.* **1998**, 901-904.
43. Velders, A.H.; Pazderski, L.; Ugozzoli, F.; Biagini-Cingi, M.; Manotti-Lanfredi, A.M.; Haasnot, J.G.; Reedijk, J. Synthesis, characterization and crystal structure of trans-aquatrichlorobis(5,7-dimethyl-1,2,4-triazolo[1,5-*a*]pyrimidine-N3)ruthenium (III) monohydrate. *Inorg. Chim. Acta* **1998**, 273, 259-265.
44. Navarro, J.A.R.; Romero, M.A.; Salas, J.M.; Molina, J.; Tiekink, E.R.T. Ternary copper (II) complexos with the versatile 4,7-dihydro-5-methyl-7-oxo-1,2,4-triazolo[1,5-*a*]pyrimidine ligand. *Inorg. Chim. Acta* **1998**, 274, 53-63.

45. Davies, G.E. Anthibronchoconstrictor activity of two new phosphodiesterase inhibitors, a triazolopyrazine (ICI 58301) and a triazolopyrimidine (ICI 63197). *J. Pharm. Pharmacol.* **1973**, *25*, 681-689.
46. Fischer, G. 1,2,4-Triazolo[1,5-*a*]pyrimidines. *Adv. Heterocycl. Chem.* **1993**, *57*, 81-138.
47. Gupton, J.T.; Petrich, S.A.; Hicks, F.A.; Wilkinson, D.R.; Vargas, M.; Hosein, K.N.; Sikorski, J.A. The preparation of heterocyclic appended vinyllogous iminium salts and their application to the regioselective preparation of biheterocyclic systems. *Heterocycles*, **1998**, *47*, 689-702.
48. Tominaga, Y.; Sakai, S.; Kohra, S.; Tsuka, J.; Matsuda, Y.; Kobayashi, G. Pyrimidine and fused pyrimidine derivatives. III. Synthesis of *s*-triazolo[1,5-*a*]pyrimidines derivatives by using ketene dithioacetals. *Chem. Pharm. Bull.* **1985**, *33*, 962-970.
49. Oukoloff, K.; Lucero, B.; Francisco, K. R.; Brunden, K. R.; Ballatore, C. 1,2,4-Triazolo[1,5-*a*]Pyrimidines in Drug Design. *Eur. J. Med. Chem.* **2019**, *165*, 332-346.
50. Namgoong, S.K.; Lee, H.J.; Kim, Y.S.; Shin, J.-H.; Che, J.-K.; Jang, D.Y.; Kim, G.S.; Yoo, J.K.; Kang, M.-K.; Kil, M.-W.; Choi, J.D.; Chang, S.-I. Synthesis of the quinoline-linked triazolopyrimidine analogues and their interections with the recombinant tobacco acetolactate synthase. *Biochem. Biophys. Res. Commun.* **1999**, *258*, 797-801.
51. Johnson, T. C.; Mann, R. K.; Schmitzer, P. R.; Gast, R. E. Acetohydroxyacid Synthase Inhibiting Triazolopyrimidine Herbicides. *Bioact. Heterocycl. Compd. Cl.* **2012**, 51-60.
52. Monte, W.T.; Kleschick, W.A.; Meikle, R.W.; Snider, S.W.; Bordner, J. Methods for controlling the regioselection in the reaction of 3-amino-5-(benzylthio-1,2,4-triazole with acetylacetaldehyde dimethyl acetal. *J. Heterocycl. Chem.* **1989**, *26*, 1393-1396.
53. Kleschick, W.A.; Bordner, J. Regioselection in the reaction of 3-amino-5-benzylthio-1,2,4-triazole with unsymmetrical 1,3-diketones. *J. Heterocycl. Chem.* **1989**, *26*, 1489-1493.
54. Monte, W.T.; Kleschick, W.A.; Bordner, J. Controlling substitution patterns on the 1,2,4-triazolo[1,5-*a*]pyrimidine ring. Selective removal of chlorine at the 7-position. *J. Heterocycl. Chem.* **1999**, *36*, 183-188.
55. Shankar, R.B.; Pews, R.G. Synthesis of 1,2,4-triazolo[1,5-*a*]pyrimidines-2-sulfonamides. *J. Heterocycl. Chem.* **1993**, *30*, 169-172.
56. Okabe, T.; Bhooshan, B.; Novinson, T.; Hillyard, I.W.; Garner, G.E.; Robins, R.K. Dialkyl bicyclic heterocycles with a bridgehead nitrogen as purine analogs possessing significant cardiac inotropic activity. *J. Heterocycl. Chem.* **1983**, *20*, 735-751.
57. Williams, L.A. Structure of certain polyazaindenes. VI. Structure of some products obtained from 3-amino-1,2,4-triazoles with acetylacetone and ethyl acetoacetate. *J. Chem. Soc.* **1960**, 1829-1832.
58. Novinson, T.; Springer, R.H.; O'Brien, D.E.; Scholten, M.B.; Miller, J.P.; Robins, R.K. 2-(alkylthiol-1,2,4-triazolo[1,5-*a*]pyrimidines as adenosine 3',5'-monophosphate phosphodiesterase inhibitors with potential as new cardiovascular agents. *J. Med. Chem.* **1982**, *25*, 420-426.
59. Kidwai, M.; Chauhan, R. Nafion-H® Catalyzed Efficient One-Pot Synthesis of Triazolo[5,1-*b*]Quinazolinones and Triazolo[1,5-*a*]Pyrimidines: A Green Strategy. *J. Mol. Catal. A: Chem.* **2013**, *377*, 1-6.
60. Petrick, S.A.; Qian, Z.; Santiago, L.M.; Grupton, J.T. The application of unsymmetrical vinyllogous iminium salts and related synthons to the preparation of monosubstituted triazolo[1,5-*a*]pyrimidines. *Tetrahedron* **1994**, *50*, 12113-12124.
61. Petrick, S.A.; Qian, Z.; Santiago, L.M.; Gupton, J.T. The application of symmetrical vinamidinium salts to the preparation of monosubstituted triazolo[1,5-*a*]pyrimidines. *Heterocycles* **1995**, *40*, 729-742.
62. Tominaga, Y.; Kohra, S.; Honkawa, H.; Hosomi, A. Synthesis of pyrimidine derivatives and their related compounds using ketene dithioacetals. *Heterocycles* **1989**, *29*, 1409-1429.
63. Gami, S. P.; Vilapara, K. V.; Khunt, H. R.; Babariya, J. S.; Naliapara, Y. T. Synthesis and Antimicrobial Activities of Some Novel Triazolo[1,5-*a*]Pyrimidine Derivatives. *Int. Lett. Chem., Phys. Astron.* **2014**, *30*, 127-134.
64. Gasse, C.; Douguet, D.; Huteau, V.; Marchal, G.; Munier-Lehmann, H.; Pochet, S. Substituted Benzyl-Pyrimidines Targeting Thymidine Monophosphate Kinase of Mycobacterium Tuberculosis: Synthesis and in Vitro Anti-Mycobacterial Activity. *Bioorg. Med. Chem.* **2008**, *16* (11), 6075-6085.
65. Jadhav, S. B.; Fatema, S.; Bhagat, S. S.; Farooqui, M. Thiazolo[3,2- *a*] Pyrimidones as a Novel Anti-TB Agents. *J. Heterocycl. Chem.* **2018**, *55* (12), 2893-2900.
66. Bhatt, J. D.; Chudasama, C. J.; Patel, K. D. Pyrazole Clubbed Triazolo[1,5-*a*]Pyrimidine Hybrids as an Anti-Tubercular Agents: Synthesis, in Vitro Screening and Molecular Docking Study. *Bioorg. Med. Chem.* **2015**, *23* (24), 7711-7716.
67. Anand, P.; Akhter, Y. A Review on Enzyme Complexes of Electron Transport Chain from *Mycobacterium tuberculosis* as Promising Drug Targets. *Int. J. Biol. Macromol.* **2022**, *212*, 474-494.
68. Gálvez, J.; García-Domenech, R.; Julián-Ortiz, J. V. de; Soler, R. Topological approach to drug design. *J. Chem. Inf. Comp. Sci.* **1995**, *35*, 272-284. Errata: *J. Chem. Inf. Comp. Sci.* **1995**, *35*, 938.
69. García-March, F. J.; García-Domenech, R.; Gálvez, J.; Antón-Fos, G. M.; Julián-Ortiz, J. V. de; Giner-Pons, R.; Recio-Iglesias, M. C. Pharmacological Studies of 1-(*p*-Chlorophenyl)Propanol and 2-(1-Hydroxy-3-Butenyl)Phenol: Two New Non-Narcotic Analgesics Designed by Molecular Connectivity. *J. Pharm. Pharmacol.* **1997**, *49* (1), 10-15.
70. García-Domenech, R.; Julián-Ortiz, J. V. de Antimicrobial activity characterization in a heterogeneous group of compounds. *J. Chem. Inf. Comput. Sci.* **1998**, *38*, 445-449.
71. Julián-Ortiz, J. V. de; Gálvez, J.; Muñoz-Collado, C.; García-Domenech, R.; Gimeno-Cardona, C. Virtual Combinatorial Syntheses and Computational Screening of New Potential Anti-Herpes Compounds. *J. Med. Chem.* **1999**, *42* (17), 3308-3314.
72. Bruno-Blanch, L.; Galvez, J.; Garcia-Domenech, R. Topological virtual screening: a way to find new anticonvulsant drugs from chemical diversity. *Bioorg. Med. Chem. Lett.* **2003**, *13*, 2749-2754.
73. Julián-Ortiz, J. V. de; Besalú, E. Internal Test Sets Studies in a Group of Antimalarials. *Int. J. Molec. Sci.* **2006**, *7* (10), 456-468.

74. García-Domenech, R.; Gálvez, J.; Julián-Ortiz, J. V. de; Pogliani, L. Some New Trends in Chemical Graph Theory. *Chem. Rev.* **2008**, *108* (3), 1127–1169.
75. Garcia-Domenech, R.; Zanni, R.; Galvez-Llompart, M.; Julian-Ortiz, J. V. de Modeling Anti-Allergic Natural Compounds by Molecular Topology. *Comb. Chem. High Throughput Screening* **2013**, *16* (8), 628–635.
76. Zanni, R.; Galvez-Llompart, M.; Morell, C.; Rodríguez-Henche, N.; Díaz-Laviada, I.; Recio-Iglesias, M. C.; Garcia-Domenech, R.; Galvez, J. Novel Cancer Chemotherapy Hits by Molecular Topology: Dual Akt and Beta-Catenin Inhibitors. *PLOS ONE* **2015**, *10* (4).
77. Castillo-Garit, J. A.; Flores-Balmaseda, N.; Álvarez, O.; Pham-The, H.; Pérez-Doñate, V.; Torrens, F.; Pérez-Giménez, F. Computational Identification of Chemical Compounds with Potential Activity against Leishmania Amazonensis Using Nonlinear Machine Learning Techniques. *Curr. Top. Med. Chem.* **2019**, *18* (27), 2347–2354.
78. García-Domenech, R.; Julián-Ortiz, J. V. de; Duart, M. J.; García-Torrecillas, J. M.; Antón-Fos, G. M.; Ríos-Santamarina, I.; Gregorio Alapont, C. de; Gálvez, J. Search of a topological pattern to evaluate toxicity of heterogeneous compounds. *SAR QSAR Environ. Res.* **2001**, *12*, 237–254.
79. Lavado, G. J.; Baderna, D.; Carnesecchi, E.; Toropova, A. P.; Toropov, A. A.; Dorne, J. L.; Benfenati, E. QSAR Models for Soil Ecotoxicity: Development and Validation of Models to Predict Reproductive Toxicity of Organic Chemicals in the Collembola Folsomia Candida. *J. Hazard. Mater.* **2022**, *423*, 127236.
80. Gálvez, J.; Julián-Ortiz, J. V. de; García-Domenech, R. General topological patterns of known drugs. *J. Mol. Graphics Modell.* **2001**, *20*, 84–94.
81. Kier, L. B.; Murray, W. J.; Randić, M.; Hall, L. H.. Molecular Connectivity V: connectivity series concept applied to density. *J. Pharm. Sci.* **1976**, *65*, 1226–1230.
82. Kier, L. B.; Hall, L. H. Molecular Connectivity VII: specific treatment of heteroatoms. *J. Pharm. Sci.* **1976**, *65*, 1806–1809.
83. Randić, M. The connectivity index 25 years after. *J. Mol. Graphics Modell.* **2001**, *20*, 19–35.
84. Estrada, E.; Uriarte, E. Recent advances on the role of topological indices in drug discovery research. *Curr. Med. Chem.* **2001**, *8*, 1573–1588.
85. Selassie, C. D.; Mekapati, S.B.; Verma, R. P. QSAR: then and now. *Curr. Top. Med. Chem.* **2002**, *2*, 1357–1379.
86. Ivanciuc, O. Cheminform Abstract: Chemical Graphs, Molecular Matrices and Topological Indices in Chemoinformatics and Quantitative Structure-Activity Relationships. *ChemInform* **2013**, *44* (49).
87. Havare, Ö. Ç. Quantitative Structure Analysis of Some Molecules in Drugs Used in the Treatment of COVID-19 with Topological Indices. *Polycyclic Aromat. Compd.* **2021**, 1–12.
88. Balaban, A. T. Highly discriminating distance based topological index. *Chem. Phys. Lett.* **1982**, *89*, 399–404.
89. Bonchev, D. Overall connectivity: a next generation molecular connectivity. *J. Mol. Graphics Modell.* **2001**, *20*, 65–75.
90. Estrada, E.; Molina, E. Novel local (fragment-based) topological molecular descriptors for QSpr/QSAR and molecular design. *J. Mol. Graphics Modell.* **2001**, *20*, 54–64.
91. Tropsha, A.; Zheng, W. Identification of the descriptor pharmacophores using variable selection QSAR: applications to database mining. *Curr. Pharm. Des.* **2001**, *7*, 599–612.
92. Gozalbes, R.; Doucet, J. P.; Derouin, F. Application of topological descriptors in QSAR and drug design: history and new trends. *Curr. Drug. Targets Infect. Disord.* **2002**, *2*, 93–102.
93. Agatonovic-Kustrin, S.; Ling, L. H.; Tham, S. Y.; Alany, R. G. Molecular descriptors that influence the amount of drugs transfer into human breast milk. *J. Pharm. Biomed. Anal.* **2002**, *29*, 103–119.
94. Basak, S. C.; Mills, D. R.; Balaban, A. T.; Gute, B. D. Cheminform Abstract: Prediction of Mutagenicity of Aromatic and Heteroaromatic Amines from Structure: A Hierarchical QSAR Approach. *ChemInform* **2010**, 32.
95. Orosz, Á.; Héberger, K.; Rácz, A. Comparison of Descriptor- and Fingerprint Sets in Machine Learning Models for ADME-Tox Targets. *Front. Chem.* **2022**, *10*.
96. Jacobs, M. R. Activity of Quinolones against Mycobacteria. *Drugs* **1995**, *49* (Supplement 2), 67–75.
97. García-Sánchez, J. E.; López, R.; Prieto, J. *Antimicrobianos en Medicina*; Prous Science: Barcelona, 2006.
98. O'Neil, M. J. The Merck Index: An Encyclopedia of Chemicals, Drugs, and Biologicals; Royal Society of Chemistry: Cambridge, UK, 2013.
99. García-Domenech, R. DesMol. Unidad de Investigación de Diseño de Fármacos y Conectividad Molecular, Dep. Química-Física, Facultad de Farmacia, Universitat de València, Spain, 2013.
100. Wiener, H. Structural Determination of Paraffin Boiling Points. *J. Am. Chem. Soc.* **1947**, *69*, 17–20.
101. Gálvez, J.; García-Domenech, R.; Salabert, M. T.; Soler, R. Charge indices. New topological descriptors. *J. Chem. Inf. Comp. Sci.* **1994**, *34*, 520–525.
102. Julián-Ortiz, J.V. de; Besalú, E.; García-Domenech, R. True prediction by consensus for small sets of cyclooxygenase-2 inhibitors. *Indian J. Chem.* **2003**, *42A*, 1392–1404.
103. BMDP New System 2.0. Statistical Solutions Ltd, Ireland.
104. Gálvez, J.; García-Domenech, R.; Gregorio Alapont, C. de; Julián-Ortiz, J. V. de; Popa, L. Pharmacological distribution diagrams: a tool for the new drug design. *J. Mol. Graphics* **1996**, *14*, 272–276.
105. Font, D.; Heras, M.; Villalgordo, J.M. Development of an efficient and straightforward methodology toward the synthesis of molecularly diverse 2,6-disubstituted 3,4-dihydropyrimidin-4(3h)-ones. *Synthesis* **2002**, 1833–1842.
106. Font, D.; Heras, M.; Villalgordo, J.M. Solution- and solid-phase parallel synthesis of 4-alkoxy-substituted pyrimidines with high molecular diversity. *J. Comb. Chem.* **2003**, *5*, 311–321.

-
107. Petasis, N.A.; Goodman, A.; Zavialov, I. A. A new synthesis of α -arylglycines from aryl boronic acids. *Tetrahedron* **1997**, *53*, 16463-16470.
 108. Petasis, N. A.; Zavialov, I. A. A new and practical synthesis of α -amino acids from alkenyl boronic acids *J. Am. Chem. Soc.* **1997**, *119*, 445-446.
 109. Shinnick, T. M.; Good, R. C. Mycobacterial taxonomy. *Eur. J. Clin. Microbiol. Infect. Dis.* **1994**, *13*, 884-901.
 110. Franzblau, S. G.; Witzig, R. S.; McLaughlin, J. C.; Torres, P.; Madico, G.; Hernandez, A.; Degnan, M. T.; Cook, M. B.; Quenzer, V. K.; Ferguson, R. M.; Gilman, R. H. Rapid low technology MIC determination with clinical *Mycobacterium* isolates by using the microplate alamar blue assay. *J. Clin. Microbiol.* **1998**, *36*, 362-366.