

Article

The Magnetosome Protein, Mms6, is a Lipid-Activated Ferric Reductase

Dilini Singappuli-Arachchige ¹, Shuren Feng ², Lijun Wang ³, Pierre E. Palo ⁴, Samuel O. Shobade ⁵, Michelle Thomas ⁶ and Marit Nilsen-Hamilton ^{7,*}

¹ Ames Laboratory, U. S. Department of Energy, and Aptalogic Inc., Ames, Iowa; dilini@iastate.edu

² Ames Laboratory, U. S. Department of Energy, and Roy J. Carver Department of Biochemistry, Biophysics and Molecular Biology, Iowa State University, Ames, Iowa; current Affiliation: Disease Screening Center, Tianjin Women's and Children's Health Center, Tianjin, P.R. China; rustyfengsr@hotmail.com

³ Ames Laboratory, U. S. Department of Energy, Ames, Iowa, and Roy J. Carver Department of Biochemistry, Biophysics and Molecular Biology, Iowa State University, Ames, Iowa; imwanglijun19760830@gmail.com

⁴ Ames Laboratory, U. S. Department of Energy, Ames, Iowa; ppalo@iastate.edu

⁵ Ames Laboratory, U. S. Department of Energy, and Roy J. Carver Department of Biochemistry, Biophysics and Molecular Biology, Iowa State University, Ames, Iowa; shobades@iastate.edu

⁶ Ames Laboratory, U. S. Department of Energy, Ames, Iowa; mjthomas14@gmail.com

⁷ Ames Laboratory, U. S. Department of Energy, Roy J. Carver Department of Biochemistry, Biophysics and Molecular Biology, Iowa State University, and Aptalogic Inc., Ames, Iowa; marit@iastate.edu

* Correspondence: marit@iastate.edu;

Abstract: Magnetosomes of magnetotactic bacteria consist of magnetic nanocrystals with defined morphologies enclosed in vesicles originated from cytoplasmic membrane invaginations. Although many proteins are involved in creating magnetosomes, a single magnetosome protein, Mms6, can direct the crystallization of magnetite nanoparticles *in vitro*. The *in vivo* role of Mms6 in magnetosome formation is debated and the observation that Mms6 binds ferric and not ferrous iron raises the question of how Mms6 could promote the crystallization of magnetite, which contains both ferric and ferrous iron. Here we show that Mms6 is a ferric reductase that reduces ferric to ferrous iron using NADH and FAD as electron donor and cofactor, respectively. Reductase activity is elevated when Mms6 is integrated into either liposomes or bicelles. Analysis of Mms6 mutants suggests that the C-terminal domain binds iron and the N-terminal domain contains the catalytic site. Although Mms6 forms multimers that involve C-terminal and N-terminal domain interactions, a fusion protein with Mms6, which remains a monomer, displays reductase activity, which suggests that the catalytic site is fully in the monomer. These results are consistent with a hypothesis that Mms6, a membrane protein, promotes the formation of magnetite by a mechanism that involves reducing iron.

Keywords: Mms6; ferric reductase; lipid; bicelles

1. Introduction

Since first reported in 1975 [1] magnetotactic bacteria (MTB) have attracted interest because of their abilities to synthesize magnetite crystals in specialized organelles called “magnetosomes” [2-4]. Gene regulation and genomic analysis related to magnetosome formation has been extensively studied [5-7]. Superparamagnetic magnetite crystals of similar size and shape to the bacterial magnetites are formed *in vitro* due to the presence of recombinant Mms6, a magnetosome-associated protein [8-10]. Although Mms6 alone is not responsible for the formation of magnetic nanoparticles *in vivo* [11], its *in vitro* activity provides us an opportunity to better understand the mechanism by which this biomineralization protein functions. Such knowledge helps us to understand how magnetotactic bacteria can synthesize the magnetic crystals in magnetosomes and informs the design of bio inspired routes to synthesize iron oxides and other studies related to magnetic nanoparticles [12-18].

We have previously demonstrated that Mms6 forms a micellar quaternary structure *in vitro* that may provide a surface for magnetite nanoparticle formation [9]. Mms6 consists of two subdomains, with the N-terminal domain responsible for anchoring the C-terminal domain in the micelle from which the C-terminus binds iron and forms magnetic nanoparticles. Analysis of Mms6 and its synthetic C-terminal domain by fluorescence, and CD spectroscopy provided evidence that the protein undergoes a structural change upon binding iron and exhibits two modes of interaction with iron [9, 19].

The magnetite crystal lattice in the magnetosome contains $\text{Fe}^{2+}(\text{Fe}^{3+})_2\text{O}_4$. However, Mms6 binds Fe^{3+} and not Fe^{2+} [20]. Although this deficiency can be circumvented *in vitro* by making available a high concentration of Fe^{2+} , the ratio of $\text{Fe}^{3+}/\text{Fe}^{2+}$ *in vivo* is unlikely to be 2:1. Rather, Fe^{3+} is proposed as the predominant form of iron in magnetosomes [21-25]. Thus, if Mms6 were to be involved in initiating or promoting the growth of magnetite crystals *in vivo*, it would need to cooperate with a protein that could reduce the available Fe^{3+} or itself be a reductase.

Here we show that Mms6 is a ferric reductase, capable of producing the Fe^{2+} required for placement in the magnetite crystal lattice. Mms6 shows structural homology with the ferric reductase superfamily but it does not require the presence of a heme group to reduce iron. Mutational analysis suggests that the reductase catalytic site is in the N-terminal domain. Consistent with its association with magnetosome membranes when isolated from cells and other evidence that it is membrane-localized *in vivo* [8, 11, 26], we show that the reductase activity of Mms6 is enhanced when the protein is integrated in a lipid bicelle membrane. Thus, we propose that the function of Mms6 *in vivo* contributes to both essential elements of magnetic crystal formation: the reduction of iron and the assembly of Fe^{2+} and Fe^{3+} into the crystal structure.

2. Results

2.1. Mms6 is a ferric reductase

The results of our previous studies suggested that Mms6 binds Fe^{3+} cooperatively in groups of 3 [9] and does not bind Fe^{2+} [20]. These binding characteristics are not compatible with an independent role of Mms6 in building the crystal lattice of magnetite, which contains $\text{Fe}^{3+}:\text{Fe}^{2+}$ at a ratio of 2:1. We reasoned that, if Mms6 has a direct role in building magnetite *in vivo* then it should be capable of reducing Fe^{3+} to create the Fe^{2+} necessary for building the crystal lattice. Reduction of Fe^{3+} to Fe^{2+} by Mms6 was monitored by the increase of A_{562} from the Fe^{2+} -ferrozine complex. With this assay we found that Mms6 can reduce Fe^{3+} to Fe^{2+} under aerobic (Fig. 1) and anaerobic (Fig. 1B) conditions. To establish that the observed activity is likely due to catalysis by Mms6, we tested five sequence variants of the protein, which were m1Mms6 (a shuffle of the charged residues in the C-terminal 21 amino acid residues), m2Mms6 (a shuffle of the 9 -OH and -COOH containing amino acids in the C-terminal 21 residues), m3Mms6 (shuffle of the C-terminal 21 amino acid residues), Mms6(W119A), and Mms6(L108A, L110A, L112A, L114A, L116A), all of which demonstrated decreased reductase activity (Fig. 1A, E). A key to the numbering of positions in Mms6 is found in Fig. S1. These mutations cover the length of the protein, which consists of two domains characterized by their hydrophobicity, with the N-terminal being hydrophobic and the C-terminal hydrophilic (Fig. S2).

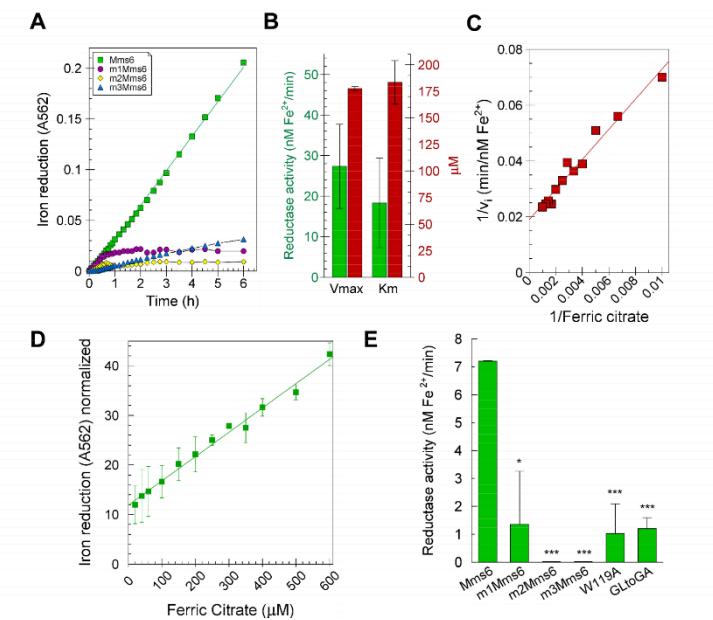


Figure 1. Reductase activity of Mms6. Ferric reductase activity was measured as described in Materials and Methods. **A)** The reduction of 70 μM Fe^{3+} -citrate by 20 μM Mms6, m1Mms6, and m2Mms6 Mms6 in air in the presence of 100 μM each of NADH and FAD. **B)** Comparison of the V_{max} and K_m for Mms6 under aerobic (green) and anaerobic (red) conditions. **(C)** A double reciprocal plot of the initial velocity vs. the ferric citrate concentration in air. **D)** The initial velocity of Mms6 (20 μM) reductase activity as a function of Fe^{3+} -citrate concentration under anaerobic conditions. **E)** The reduction of 70 μM Fe^{3+} -citrate by 20 μM Mms6, and various mutants. The asterisks denote p values of <0.05 (*) and <0.001 (***). in paired comparisons with Mms6.

We have previously demonstrated that Mms6 forms micelles *in vitro*. A trivial reason for the mutant and scrambled forms of Mms6 lacking reductase activity might be that they are aggregated or otherwise structurally impaired at the macromolecular level. To avoid this problem, all mutations, and scrambled versions of Mms6 were chosen to maintain a similar hydrophobicity plot to avoid variations in gross quaternary organization of the protein (Fig. S2). This expectation was confirmed by previous analysis of these mutations that demonstrated their abilities to form micelles [19] and TEM images of Mms6 and its mutants, which show similar morphologies (Fig. S3). Thus, it is unlikely that the mutant versions of Mms6 lack reductase activity due to reorganized quaternary structure. The lack of reductase activity with mutation of Mms6 supports the notion that the reductase activity displayed by Mms6 requires a defined catalytic site.

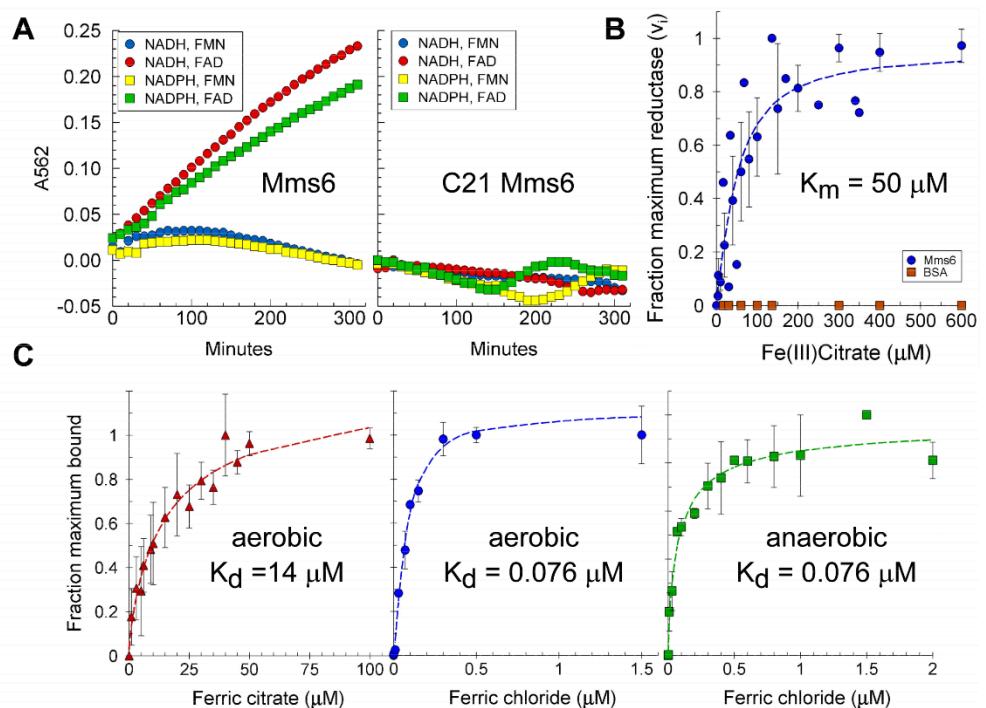


Figure 2. Mms6 ferric reductase specificity for cofactor and electron donor. Panel A) Mms6 (left) and C21Mms6 (right) were tested with 100 μM NADH or NADPH as electron donors combined with either 100 μM FAD or FMN as cofactors in 20 mM Tris, 100 mM KCl, 1 mM ferrozine, 100 mM ferric citrate, pH7.5. Reaction mixes run in parallel without protein were used as the background to subtract from the results of incubation with 20 μM Mms6 with the combinations of cofactor and electron donor shown. B) Reductase activity (initial velocity) was determined at a range of Fe³⁺-citrate concentrations to determine the Km for the reductase. Shown is the compiled data from 9 independent experiments involving six different protein preparations ($r^2 = 0.8$, Hill n set to 1), C) The Kd of Mms6 was determined under aerobic conditions at pH 7.5 and pH 7 for ferric citrate and ferric chloride, respectively and anaerobic conditions at pH 7 for ferric chloride. The ferric citrate binding isotherm was created from the averaged results of 11 independent experiments (duplicates per experiment) with two assessed by intrinsic fluorescence and the remainder by ⁵⁵Fe binding with filter capture. In total, 5 different protein preparations were tested. Both the aerobic and anaerobic ferric chloride binding isotherms are each the average of two independent experiments with one (anaerobic) or two (aerobic) protein preparations and assessed by ⁵⁵Fe binding with filter capture. Goodness of fit: Fe-citrate, aerobic ($r^2 = 0.97$, Hill n=0.84), FeCl₃, aerobic ($r^2 = 0.99$, Hill n=1.3), FeCl₃, anaerobic ($r^2 = 0.98$, Hill n=0.78).

2.2. Electron donor and cofactor requirement

To determine its requirements for electron donor and co-factor, Mms6 was tested for reductase activity in the presence of combinations of electron donor and cofactors. The results show that Mms6 prefers NADH and can also use NADPH as electron donor, but exclusively uses FAD over FMN as cofactor (Fig. 2 panel A left). The synthetic C terminal domain C21Mms6 was also tested under same conditions and no significant activity was observed (Fig. 2 panel A right). The K_m for FAD and NADH were determined as ~25 and ~15 μM by their abilities to stimulate reductase activity and similar activity isotherms were obtained for FAD under aerobic and anaerobic conditions (Fig. S4). The addition of heme had no effect on the activity (Fig. S5).

The K_m of Mms6 for Fe³⁺-citrate is 50 μM, whereas BSA present in the same assay gave no activity (Fig. 2B). In the presence of 100 μM each of FAD and NADH and saturating Fe³⁺, the specific activity was estimated as 0.24 ± 0.20 nmole/min/mg (N=20) and the k_{cat} as $2.0 \times 10^{-5} \pm 2.6 \times 10^{-5}$ sec⁻¹. The affinity for ferric iron was investigated under aerobic and anaerobic conditions for FeCl₃ and under aerobic conditions for ferric citrate. The K_d for FeCl₃ under aerobic and anaerobic conditions were the same at 76 nM, whereas the K_d for ferric citrate was significantly higher at 14 uM (Fig. 2 panel C). This difference is to be

expected as citrate has a high affinity for Fe^{3+} and reduces the concentration of free Fe^{3+} available for Mms6 binding.

2.3. Iron binding residues in Mms6

For reductase activity, Mms6 must bind and reduce Fe^{3+} . To identify potential iron chelating residues, we made a series of alanine exchange mutants for each -OH or -COOH containing side-chain (S,D or E) in the C-terminal domain. We have previously demonstrated that the C terminal mutant m2Mms6 (Table S1) binds Fe^{3+} with a very low affinity that appears as nonspecific [9, 19]. This and other Mms6 mutants were tested for iron binding and ferric reductase activities (Fig 3). For both assays, the activities of the mutants were normalized to that of the wild-type protein. The effect of mutations on reductase and iron binding activity were highly correlated with mutations at positions S138, S143, and S146 and their combinations showing large decreases in iron binding and reductase activities (Fig. 3B, S6). Although E148A had wild-type activity, the Mms6(S146A,E148A) double mutant showed complete loss of both iron binding and reductase activities. The concordance in the results from these two assays strongly suggests that S138, S143, and S146 might be important for iron binding, which is required for reductase activity of Mms6. However, this observation does not rule out the possibility that one or more of these residues plays a structural role for one or both activities.

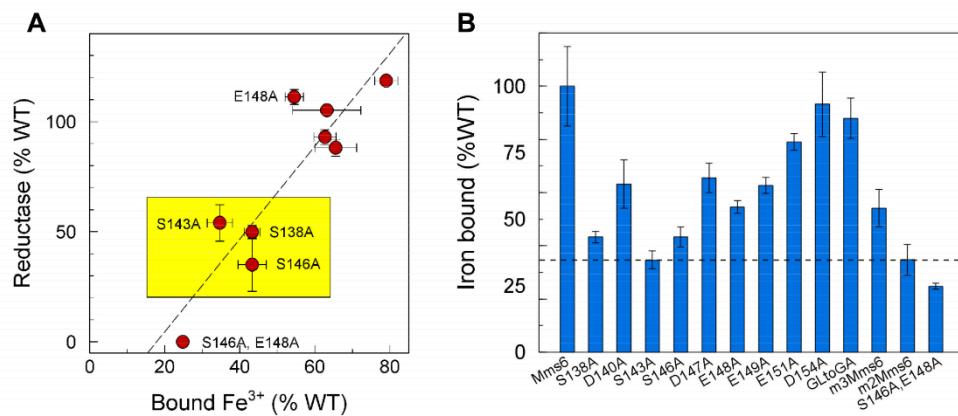


Figure 3. Iron binding and reductase activity correlated in multimer. **A)** The ferric reductase activity of each recombinant protein was determined as described in Materials and Methods with 20 μM protein and 100 μM ferric citrate. **B)** The iron binding activity of each recombinant protein was determined as described in Materials and Methods with 1 μM protein and 20 μM ferric citrate. Reductase and iron binding activities were normalized to that of the wild-type protein. The dashed line is at the level of m3Mms6, which has been previously shown to bind Fe^{3+} nonspecifically [9].

2.4. Mms6 primary sequence and tertiary structure predictions

To better understand the relation between Mms6 and other prokaryotic proteins the primary sequence of Mms6 from *Magnetospirillum magneticum*, AMB-1, used in this study, was compared with sequences of Mms6 from related organisms and with other proteins (Mms7 and Mms13 that were also found tightly associated with magnetosomes [8]. The region of highest identify for was the segment of GL repeats and W119 (Fig. 4A).

We previously predicted a structure for Mms6 [19] using I-TASSER [27] and TM-align [28] to identify related proteins and protein families. This comparison identified homology with reductases (Table S3). Here we developed a 3-dimensional model of Mms6 using SWISS-MODEL [29], which identified the Photosystem II reaction center protein, W, as the most likely structural equivalent for Mms6. Based on the 3D SWISS-MODEL, the catalytically active sites in Mms6 were predicted using CASTp [30]. This analysis identified residues in the N terminal (W119 and GL repeat) as the most likely sites to define the catalytic domain of the protein (Fig. 4B). These findings are consistent with our observations that mutations in these positions of the N-terminal domain result in loss of catalytic activity (Fig. 1E).

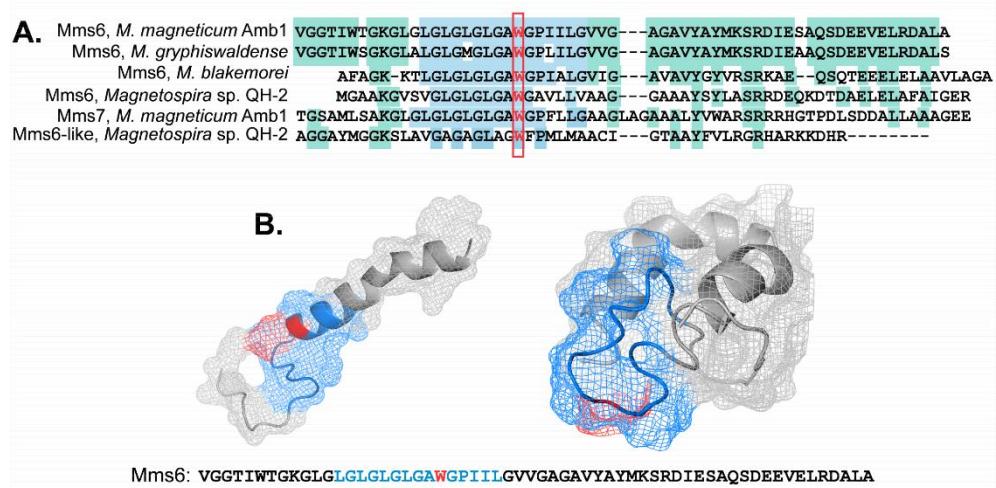


Figure 4. Primary and tertiary structural analysis of Mms6. A) Alignment of Mms6 and proteins with related sequence. Primary sequences were obtained from NIH protein database (<http://www.ncbi.nlm.nih.gov/protein>) and aligned using the clustalw2 online server. The colored regions indicate identity of sequence with the blue region identifying the GL domain and the green rectangle the conserved tryptophane. B) The Swiss model and I-Tasser predicted structures for Mms6 with the conserved Trp and GL domain identified.

2.5. Mms6 integrates into lipid membranes

The N-terminal domain of mature Mms6, an amphipathic protein, is largely hydrophobic and might integrate into a membrane. Numerous observations also suggest that Mms6 is a membrane protein [8, 9, 20, 26]. Membrane integration was tested using DLS and intrinsic fluorescence. Mms6 exists in solution as micelles with a hydrodynamic diameter of ~13 nm (Fig.5A, green). Incubation with 0.5% of Triton-X100 at 24 °C followed by detergent removal with hydrophobic beads increased the Mms6 micellar hydrodynamic diameter to an average of ~45 nm, which we suspect is due to fusion of micelles induced by the detergent (Fig.5A, cyan). DMPC/DHPC bicelles (25 mM) had hydrodynamic diameters of ~10 nm (Fig.5A, red), which is consistent with their expected size [31]. The incorporation of Mms6 into the bicelle was not evidenced by a change in size of the bicelles as they had the hydrodynamic diameter of ~10 nm in the presence or absence of Mms6. However, incorporation of Mms6 into the bicelles is supported by the lack of particles the size of the Mms6 micelles (d=12-15nm) in the Mms6-bicelle sample. This lack of apparent size difference between the bicelle alone or with Mms6 could occur if the incorporation of Mms6 into the bicelles resulted in a change in shape from discoidal to spherical, which might not be observed as a change in hydrodynamic diameter as the analysis of DLS data assumes a spherical shape for all particles. A small change in hydrodynamic diameter was observed with liposomes with the loss of the Mms6-Triton X-100 peak, again suggesting integration of Mms6 into these membranes (Fig. 5A).

To further evaluate the incorporation of Mms6 into the bicelles, we measured intrinsic fluorescence. When excited at 290 nm, Mms6 shows a fluorescence spectrum with a λ_{max} at 355 nm, which shifts to 346 nm when the protein is integrated into bicelles. This blue shift is consistent with the interpretation that tryptophan is experiencing a more hydrophobic environment, which is expected after its integration into lipid membranes. To identify the Trp residue responsible for the blue fluorescence shift, we tested two Mms6 mutants (W103F and W119F). The blue shift was observed with the W103F mutant but not with the W119F mutant (Figs 5B, S7). These results suggest that W119, but not W103, adopts a new, more hydrophobic environment when Mms6 is integrated with bicelles.

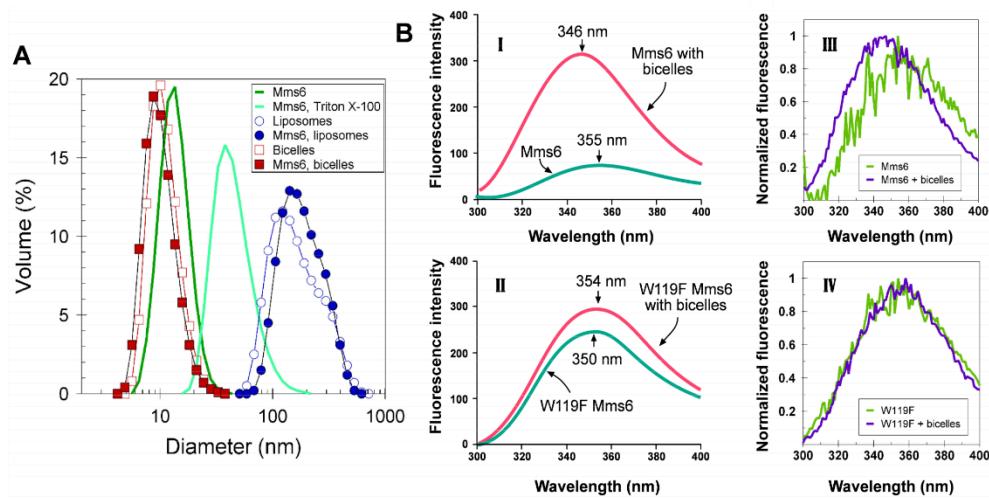


Figure 5. Mms6 interaction with lipids. (A) Dynamic light scattering analysis of samples (500 μ L in 20 mM Tris, 100 mM KCl, pH 7.5) of 25 mM bicelles, 2 mM liposomes or 0.5% Triton X100 with or without 20 μ M Mms6 as stated in the legend. Size distributions for each sample are shown as a volume percentage. (B) Fluorescence spectra were collected ($(\lambda_{\text{ex}}=290 \text{ nm})$) and analyzed as described in Materials and Methods. Smoothed curves for fluorescence intensity of Mms6 (I) or W119F Mms6 (II). The same scans of Mms6 (III) or W119F Mms6 (IV) but normalized to the maximum value and not smoothed.

2.6. Lipids promote higher ferric reductase activity of Mms6

When incorporated into bicelles or liposomes the ferric reductase activity of Mms6 was higher by an average of 2-fold in liposomes (N=2) and 5-fold in bicelles (N=2) (Fig. 6 A,B). The C and N terminal mutants of Mms6, which were inactive in the absence of bicelles (Fig. 1), were also inactive when incorporated in bicelles (Fig 6C). Thus, these mutations identify critical amino acid residues in Mms6 that are required for reductase catalysis regardless of its environment.

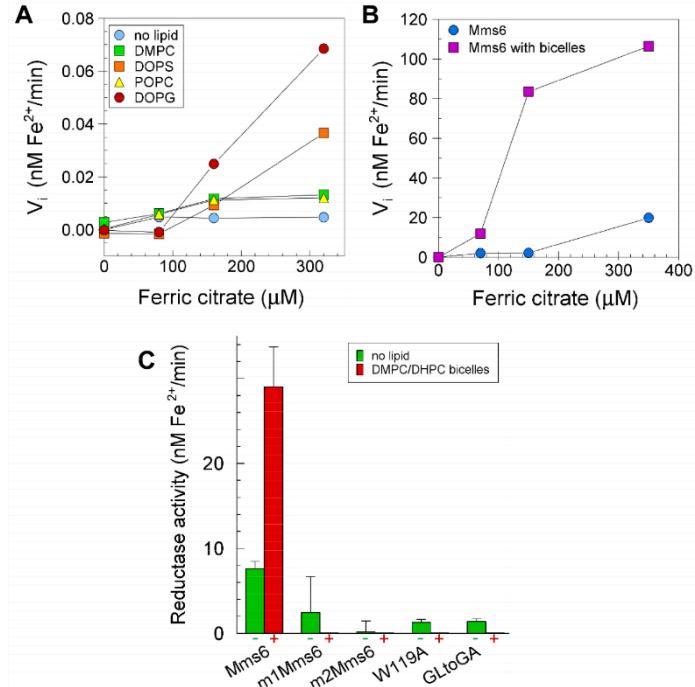


Figure 6. Mms6 has higher ferric reductase activity in lipid environments. A) Mms6 ferric reductase activity in liposomes of various lipid compositions as shown in the legend. DMPC: 1,2-Dimyristoyl-*sn*-glycero-3-phosphocholine, DOPS: 1,2-dioleoyl-*sn*-glycero-3-phospho-L-serine, POPC: 2-Oleoyl-1-palmitoyl-*sn*-glycero-3-phosphocholine, DOPG: 1,2-Dioleoyl-*sn*-glycero-3-phos-

pho-rac-(1-glycerol). **B**) Twenty uM Mms6 alone or incorporated into 25 mM q=1 DMPC/DHPC bicelles were tested as described in Materials and Methods. respectively. **C**) The Vmax of Mms6 and identified mutants while free in solution or associated with bicelles.

2.7. Reductase Activity of the Mms6 Ubiquitin monomer

To evaluate the role of its multimeric state on Mms6 activity, we reasoned that the protein could be retained as a monomer if fused at the N-terminus to a monomeric protein that does not form multimers. Consequently, we created fusion proteins of Mms6 and Mms6(S146A,E148A) with ubiquitin. The monomeric structures of these fusion proteins were validated by size exclusion chromatography (Fig. 7A). Whereas the reductase activity was the same in the monomeric as the multimeric state for the wild-type and mutant forms, the iron binding activity of Mms6(S146A,E148A) was significantly lower than for Mms6 but similar for the monomeric fusion proteins, Ubi-Mms6 and Ubi-Mms6(S146A,E148A) (Fig. 7 B,C).

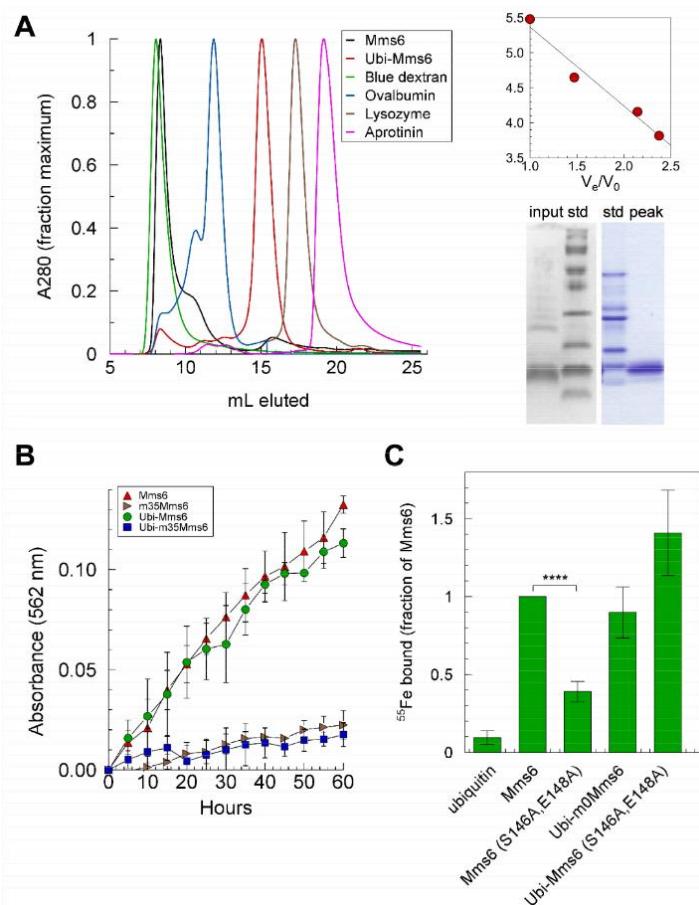


Figure 7. Monomeric Mms6 is a functional reductase but with altered C-terminal domain specificity for Fe³⁺. **A)** Left, eluted peaks of standards, Mms6 and Ubi-Mms6, right top: standard curve, right: gel electrophoretic profile of Ubi-Mms6 prior to loading and in the peak eluted at ~15 mL. **B)** Time courses of reductase activity in the presence of 350 μM ferric citrate. For each isotherm, the data from three independently performed experiments, each normalized to a midpoint value, were averaged, and similarly treated data in the presence of each protein but no ferric citrate was subtracted. The errors include the errors of the average values with and without ferric citrate. **C)** The amount of iron bound per μmole of protein in 20 μM ⁵⁵Ferric citrate expressed as a proportion of the amount bound by Mms6.

3. Discussion

Mms6 *in vivo* function: Identified from the isolated magnetosome membrane of AMB-1 as a magnetite-associated protein [8], Mms6 promotes the formation of magnetic nanoparticles *in vitro* when included in co-precipitation synthesis reactions [8, 32]. Genetic

evidence suggests that Mms6 regulates the morphology of magnetites in the later stage of crystallization *in vivo* and alternatively that it is an accessory protein, unnecessary for magnetite formation [33]. It has been proposed to alternatively hold certain protein such as Mms5, Mms7, and Mms13 to the magnetite and function with them to impart the cubo-octahedral shape of magnetite crystals in AMB-1 [11] and it has been specifically assigned the role of promoting crystal growth on the 110 face of magnetite [26].

Mms6 *in vitro* formation of magnetite: Recombinant Mms6 binds iron with high affinity and high capacity and self-assembles into multimeric micelles that appear to be important for its *in vitro* function of promoting magnetite formation [9, 19]. Mms6 binds Fe³⁺ but not Fe²⁺ [20], which bring up the question of how Mms6 might interact with both Fe²⁺ and Fe³⁺ in the magnetosome to accomplish its assigned role of controlling growth at the 110 crystal surface.

Mms6 reductase activity: Here we have, for the first time, demonstrated that Mms6 is an iron reductase that uses FAD and NADPH as cofactors. Several protein structural prediction algorithms identified reductases for homology modeling and predicted active site residues in Mms6 (W119 or the GL string of residues) for which alanine substitution Mms6 mutants had minimal to no reductase activity. Thus, the combination of modeling and experimental analysis identified the N-terminal half of the protein as the likely reductase catalytic site. The GLGL segment and Trp 119 is conserved in a group of proteins related to Mms6 that are found attached to magnetosomes [8] and also links Mms6 to ferric reductase superfamily [FRD, 34]. The presence of these features in other related proteins suggests that other members of the magnetosome protein family may also possess iron reductase activity.

C-terminal Mms6 amino acids for iron binding: We have previously demonstrated that the C-terminal domain of Mms6 binds iron and when provided with Fe²⁺ and Fe³⁺ it can direct the crystallization of magnetite as can the full-length protein [9]. Consistent with our findings here that alanine substitutions of several C-terminal domain residues result in lower Fe³⁺ binding capability of Mms6, Rawlings and coworkers reported E142, E148 and R153 are key C terminal residues involved in Mms6 ferric binding. In addition, from their simulation studies they suggested that disruption of these residues hinders a C terminal sequence specific motif to ferric iron binding [35].

Iron binding correlates with reductase activity for Mms6 C-terminal mutants, but not N-terminal mutants: We found a correlated drop in reductase activity and iron binding activity with alanine substitutions at positions 138, 143, 146, and 148. The Mms6 reductase activity is also decreased by alanine substitutions in the C-terminal domain. As the rate of catalysis depends on its ability to obtain Fe³⁺, which is mediated by the C-terminal domain, it is reasonable to consider that a decrease in reductase activity might parallel the loss of iron binding capability. However, the observed loss of reductase activity with the GL mutant, which retained iron-binding capability, identified the catalytic site as likely in the N-terminal domain.

The C and N-terminal domains interact structurally as well as functionally: We have previously shown that the binding of iron by the C-terminal domain in the multimer results in a changes in the CD spectrum, intrinsic fluorescence and SANS intensity profile, which suggests that a structural change in the C-terminal domain upon binding iron is transmitted to the N-terminal domain [19].

The kinetic parameters Mms6 are consistent with related enzymes: At 6,191 Daltons, Mms6 is the smallest of reported ferric reductases. However, if it exists *in vivo* as a complex with the remainder of its precursor protein, which is between 12,531 and 14,691 Daltons, depending on the translational start site, then Mms6 would reach the lower end of molecular weight range of 13,000 Daltons reported for the *B. subtilis* ferric reductase [36, Table S**]. A review of kinetic data for 18 reports of ferric reductases from various prokaryotes identified a range of specific activity values for the purified proteins (ferric citrate or ferric EDTA as acceptors) from 5x10⁻³ to 13100 with a median value of 14 nmol/min/mg (Table S4).

Our best estimates for the specific activity and k_{cat} of Mms6 are lower than reported for most reductases (Table S4). We suspect that this activity is lower than what would be observed *in vivo*, in part because Mms6 was refolded from an inclusion body during its preparation. Indeed, in two separate comparisons of the refolded protein with the ubiquitin fusion protein, which is isolated as a soluble monomer, the k_{cat} of the latter was 5 ± 0.2 times that of the refolded protein assayed at the same time. Another likely reason for the low activity of Mms6 *in vitro* is that the protein is out of its natural lipid environment (discussed next) and it may be only part of multisubunit holoenzyme that functions *in vivo* [37-39].

Except for its low activity, the functional characteristics of Mms6 are otherwise consistent with those of other reported reductases [40]. The pH optimum of 7 for the ferric reductase activity of Mms6 is the same as for most other reported ferric reductases (Table S4). Mms6 prefers NADH over NADPH as electron donor and is specific for FAD as co-factor. Our estimated K_m for Mms6 of $50 \mu\text{M}$ for the iron-citrate is within the range of 6 to $213 \mu\text{M}$ reported for other reductases for which the average was $34 \pm 64 \mu\text{M}$ ($N=10$), median = $12 \mu\text{M}$. However, our determined K_d for binding ferric citrate is $14 \mu\text{M}$ suggests that the K_m for enzyme activity is a more complex parameter than just depending on the binding of Mms6 with Fe^{3+} -citrate and might be related to the affinities of FAD and NADH, which we estimated as ~ 25 and $\sim 15 \mu\text{M}$ respectively.

Integration into bicelles increases Mms6 reductase activity: Mms6 spontaneously integrates into liposomes [9] and orients at the surface of a Langmuir trough [20]. These data and those of others [8, 9, 20, 26, 41] supported the expectation that Mms6 would have an affinity for membranes. Mms6, which exists as micelles in the absence of lipid, was incorporated into bicelles and the hydrodynamic diameters of the resulting particles were determined by dynamic light scattering. At concentrations as low as 0.2 mg/mL Mms6 exists as micelles in aqueous solution at pH 3 or pH 7.5. At the latter pH, these micelles are characterized with diameters of $\sim 12\text{-}15 \text{ nm}$ [19]. We observed a specific change in the intrinsic fluorescence of W119 with no change in the fluorescence of W103 when Mms6 is integrated into bicelles (Fig. S7). The shift is consistent with W119 becoming buried in a more hydrophobic location. This shift accompanies an increase in the reductase activity of Mms6 and suggests that the preferred environment of the reductase catalytic site is hydrophobic. However, with their simple and synthetic lipid composition, the bicelle environment is still far from the lipid composition of the magnetosomes.

N-terminal reductase functions as a monomer while C-terminal iron binding varies with structural context: While in the form of a multimer, the roles of individual Mms6 monomers cannot be distinguished and it cannot be known if the Mms6 iron-binding and reductase activities depend on a multimeric structure or can be performed by a monomer. This ability to multimerize also adds complication to the interpretation of data for the effects of some of the mutations on reductase activity as the GL to GA mutation and the W119 mutation destabilize the micelles resulting in a mixture of heteromeric and monomeric Mms6 [19]. To address this question, we created monomeric Mms6 by fusing the sequence to ubiquitin. This monomeric fusion protein possessed the same reductase activity as the multimer but mutations in the C-terminal domain that drastically decreased Fe^{3+} binding in multimeric Mms6, did not influence iron binding by the monomer. This data suggests that C-terminal domain but not the N-terminal domain structure and function is affected by the multimeric state of Mms6 and that the iron binding capability of the Ubi-Mms6(S146,E148) monomer does not support catalysis.

The C-terminal domains interact with each other while associated in the micelle [9]. The results shown here support the hypothesis that the C-terminal domain adopts a “multimeric” structure in the micellar configuration that is different from its structure in the monomer. These alternative structures have similar capabilities to bind iron but may use different residues for capturing iron. Alternatively, although the identified S146 and E148 might be directly chelate iron only in the multimer, they may also be responsible for maintaining the C-terminal domain structure required for high affinity iron binding and the

integration of Fe^{3+} capture with catalysis in the multimer and not directly interact with iron.

Summary: We have demonstrated that Mms6 is an iron reductase with kinetic properties like other prokaryotic iron reductases and with a requirement for FAD and preference for NADH as cofactor and electron donor respectively. The N-terminal domain of Mms6 contains the catalytic site that acts as a monomer and the C-terminal domain binds iron. Reductase activity is enhanced by the incorporation of Mms6 into membranes. This finding that Mms6 is an iron reductase explains how it can be involved in magnetosome formation *in vivo* when it binds only Fe^{3+} but requires Fe^{3+} and Fe^{2+} to create magnetite.

4. Materials and Methods

4.1. Reagents, proteins, and preparation of mutants and sequence variants

Phospholipids used to make bicelles and liposomes were purchased as stocks dissolved in 100% chloroform from Avanti Polar Lipids. Bio-BeadsTM SM-2 Resin used for removing detergents was purchased from Bio-Rad. Other chemical reagents were of analytical grade or higher purity and were obtained from Sigma-Aldrich. Single site-directed mutagenesis of Mms6 was conducted using the QuikChange II mutagenesis kit from Agilent Technologies following the manufacturer's instructions. C-terminal domain variants containing shuffled residues were created by replacing the appropriate segment of the cDNA with oligonucleotides encoding the desired amino acid sequences. All protein sequences used in this study can be found in Table S1. Mms6 is a cleavage product of a longer primary sequence. The residues cited in this work are numbered relative to the proposed start site of the original translated protein based on the gene sequence (Fig. S1). Position 1 in Mms6 is position 98 in the proposed primary sequence. The mature forms of Mms6 and its mutants were expressed and purified as described previously [9, 10, 19]. The C-terminal domain of Mms6 (C21Mms6: KSRDIESAQSQEEVELRDALA) was chemically synthesized by Genscript (Genscript Corp., www.genscript.com).

4.2. Bicelle preparation and integration of Mms6 into bicelles

Bicelle stocks consisting of 1,2-dimyristoyl-sn-glycerol-3-phosphocholine (DMPC) and 1,2-dihexyl-sn-glycero-3-phosphocholine (DHPC) ($M_{\text{DMPC}}:M_{\text{DHPC}}=1:1$, $q=1$) with a total lipid concentration of 250 mM in buffer A (20 mM Tris, 100 mM KCl, pH 7.5 at 24 °C) were prepared as described in [31] with minor modifications. Chloroform was removed from an equimolar mixture of DMPC and DHPC in 100% chloroform in a glass vial on ice under a mild stream of argon in a ventilated hood. This lipid mixture was desiccated overnight under constant vacuum at 4 °C. The desiccated lipids were re-suspended in the appropriate volume of buffer A to achieve a total lipid concentration of 250 mM. The re-suspended bicelle lipids were subjected to repeated warm (45 °C)/cool (ice) cycles until the solution became non-viscous and transparent. The bicelle (DMPC/DHPC) stocks were used immediately or aliquoted and stored at -20 °C until used.

The Mms6-bicelle complex was prepared as described in [42]. Eighty micromolar Mms6 was mixed with 100 mM or 50 mM bicelle ($q=1$) in buffer A. The test tubes were sealed with screw caps and the protein-micelle mixtures were treated with four cycles of freeze (liquid nitrogen) and thaw (24 °C). The protein-bicelle mixtures were stored at 4 °C for up to three weeks or maintained at -20 °C before use. The Mms6-bicelles were incubated at room temperature for one hour before use for experiments.

4.3. Liposome preparation and integration of Mms6 into liposomes

Five individual liposome stocks of 100 mM DMPC, 1-palmitoyl-2-oleoyl-sn-glycero-3-phosphocholine (POPC), 1,2-dioleoyl-sn-glycero-3-phosphocholine (DOPC), 1,2-dioleoyl-sn-glycero-3-phospho-(1'-rac-glycerol) (DOPG), or 1,2-dioleoyl-sn-glycero-3-phospho-L-serine (DOPS) in buffer A were prepared by extrusion through polycarbonate filters [43]. The Mms6-liposome stocks were prepared by mixing Mms6 and liposomes at final concentrations of 80 μM Mms6 and 8 mM liposome in buffer A with 0.5% Triton X-100 and incubating at 24 °C for 2 h with constant inversion. The Triton X-100 was removed

by incubating with Bio-Beads™ SM-2 Resin pre-hydrated in buffer A at 24 °C with constant inversion for 3 h using a ratio of 35 µg Triton X-100 per mg of resin. The Mms6-liposomes were harvested by removing the supernatant after the beads were allowed to settle by gravity.

4.4. Ferric reductase activity

Ferric reductase activity was monitored by the spectral change in ferrozine (3-(2-Pyridyl)-5,6-diphenyl-1,2,4-triazine-p,p-disulfonic acid), which binds Fe²⁺ to form a complex with maximum absorbance at 562nm and molar extinction coefficient of 27,900 M⁻¹cm⁻¹ [44, 45]. The assay mixture for Mms6 (or Mms6 mutant) contained 0.1 mM NADH, 0.1 mM flavin adenine dinucleotide (FAD), 1 mM ferrozine and 20 µM Mms6 in 20 mM Tris-HCl, 100 mM KCl, pH 7.5 in a total volume of 100 or 200 µL. Addition of ferric citrate initiated the reaction

The assay mixture for the full-length protein contained 20 µM Mms6, 100 µM NADH or NADPH, 800 µM ferrozine, 100 µM FAD or FMN in buffer A. The same conditions were used to assay Mms6 integrated in bicelles (12.5 or 25 mM lipid) or liposomes (2 mM lipid). The reaction was started by the addition of ferric citrate and monitored by reading A₅₆₂ each 1 or 2 minute over 6 h. The background absorptions from samples lacking Mms6 read over the same time periods were subtracted from samples with Mms6 before calculating the rate of reductase activity. The V_{max}, K_m, and K_{cat} of Mms6 as a ferric reductase were fitted for K_m and V_{max} (A_{max}) using the formula A=A_{min}+(A_{max}*S)/(S+K_m) using the solver function in Microsoft excel, where A=the change in absorption at 562 nm per min with A_{min} and A_{max} being the minimum and maximum values respectively, and S= substrate concentration. All spectrophotometric measurements were performed at room temperature in 96-well plates (Falcon, Catalog# 351172 or 353948) and read with a Synergy II plate reader (full-length protein) or a Biotek, Model: Ceres 900 plate reader (C21Mms6). All determinations were performed independently at least twice in duplicate. For each incubation time, mixtures containing the same components (excluding peptide or protein) as the assay mixture provided the blank values (averages of duplicates) that were subtracted from the average value obtained in the presence of protein or peptide. The concentration of ferrous iron was determined by A₅₆₂ using extinction coefficient of 27,900 M⁻¹cm⁻¹. The activity is expressed as nmol Fe²⁺/min/mmol protein. In some experiments, particularly those involving Mms6 integrated into bicelles, there was a significant lag before enzymatic conversion was observed. Biphasic kinetics has been observed for iron reductases by others [36, 46-48] and variably attributed to the initial use of oxygen as the electron acceptor [47], lower protein concentrations [48] or functioning the absence of the larger protein complex with which it associates [36]. Consequently, for these instances, the initial velocities for Mms6 activity were taken after the initial lag period.

4.5. Iron binding by filter capture

The binding isotherms of Mms6 for iron citrate and iron chloride binding were determined using ⁵⁵Fe with either sodium citrate or sodium chloride to provide the appropriate anion. Binding was performed in 20 mM Tris-HCl, 145 mM NaCl, 0.01% Ac-BSA, 10 mM KCl, pH 7.0 and incubated for 1 h at 24 °C before filter capture (Fig. 2) or in 20 mM Tris, 100 mM KCl, pH 7.5 and the reaction mixture incubated at 24 °C for 2 h before filter capture (Figs. 3 and 7). The reaction mixture was 100 µL (Fig 2) or 20 µL (Figs. 3 and 7) and the protein-⁵⁵Fe complex was captured by filtration through 0.45µM (Millipore, Cat#: HAWP 02500) followed by two 5 mL washes with the incubation buffer at 25 °C. The capture protein-iron complex was evaluated for ⁵⁵Fe by scintillation spectroscopy in the range 0.0-18.6 meV.

4.6. Intrinsic fluorescence spectroscopy

Twenty micromolar Mms6 or a mutant Mms6 with or without 25 mM bicelles or 2 mM liposomes in buffer A were incubated at 24 °C for 2 h before collecting fluorescence spectra. Spectral analyses of the Trp-containing Mms6 and mutants were performed after

subtracting the fluorescence readings of equimolar samples of the Trp-less Mms6 (W103A, W119A). The fluorescence spectrum of 40 μ M tryptophan was obtained with and without bicelles in buffer A for correction of Mms6 spectra during decomposition of Mms6 fluorescence spectrum by the Protein Fluorescence and the Structure Toolkit [49].

4.7. Dynamic Light Scattering (DLS)

Twenty micromolar Mms6 and its mutants with or without bicelles or liposomes in buffer A were analyzed at 24 °C with a Zetasizer Nanoparticle analyzer (Model: ZEN3690, Malvern Instrument Ltd., Southborough, MA). All samples were centrifuged at 14,000 $\times g$ at 24 °C for one hour to remove particulates prior to taking DLS measurements.

4.8. Size Exclusion Chromatography

Size exclusion chromatography was performed in an AKTA FPLC system (GE healthcare) with a prepacked Superose 12 10/300GL (separation range: 1 kDa to 300 kDa; GE Healthcare, Cat#17517301) at °C and a flow rate of 0.2 mL/min. The inner dimensions of the column were 10x300-310 mm with a bed volume of 24 mL. Prior to being loaded on the column, samples were dialyzed against the column buffer (20 mM Tris, 100 mM KCl, pH7.9) then centrifuged at 15,900 RCF at 4 °C for 30 min. Blue dextran (300 kDa) was used to determine the void column volume (V_0) and the elution volumes (V_e) of ovalbumin (44.3 kDa), lysozyme (14.3 kDa) and aprotinin (6.5 kDa) were used to create a standard curve by which the apparent molecular weights of Mms6 and the ubiquitin-Mms6 fusion protein were determined.

4.9. Analysis of binding isotherms and statistical evaluations

Where appropriate the Student's T-test was used for comparison of data sets to determine statistical significance. The comparisons are noted in specific figures with the relevant p values identified by asterisks. The binding isotherms were analyzed using Microsoft Solver on Excel and the formula $B=B_{\min}+(B_{\max}^*L^n)/(L^n+K_d^n)$ [50].

Supplementary Materials: The following supporting information can be downloaded at: www.mdpi.com/xxx/s1, Figure S1: title; Table S1: title; Video S1: title.

Author Contributions: D.S., S.F., L.J., P.P., and M.T performed the studies with the following highlights: L.J. initiated the study by discovering and early characterization of the reductase activity, S.F discovered and evaluated the effect of lipids on Mms6 activity, P.P. performed the Fe^{3+} binding experiments with mutant versions of Mms6 and discovered the unexpected Fe^{3+} binding activity of the monomeric Ubi-Mms6 C-terminal mutant, D.S. localized the reductase activity to the N-terminal domain and confirmed many of the kinetic parameters, S.O.S. performed the modeling analysis, M.T. with S.F. demonstrated a monomeric structure for Ubi-Mms6. M.N-H initiated and supervised the work, wrote the final versions of the manuscript, and prepared the figures. At various points in its preparation, L.J, S.F and D.S worked with M.N-H to critically review and appropriately revise the manuscript. All authors have read and agreed to the submitted version of the manuscript.

Funding: This work was supported by the U.S. Department of Energy, Office of Basic Energy Science, Division of Materials Sciences and Engineering (cloning Mms6 and variants, initial discoveries, and determining the K_d for Fe^{3+}) and by Aptalotic Inc. (further analysis of effect of bicelles on Mms6 reductase activity, determining kinetic parameters, and identification of the N-terminal domain as the reductase). The research was performed at the Ames Laboratory and at Aptalotic Inc. Ames Laboratory is operated for the U.S. Department of Energy by Iowa State University under Contract No. DE-AC02-07CH11358.

Data Availability Statement: The data that support the findings of this study are available from the corresponding author upon reasonable request.

Acknowledgments: We thank Samir Mehanovic for help in the developing the reductase assay and Lee Bendickson for designing the scrambled mutants to retain a similar hydropathy plot as Mms6 and cloning with the help of Kate Walling to create some of the Mms6 expression plasmids.

Conflicts of Interest: MN-H is founder and CSO of Aptalotic Inc.

References

1. Blakemore, R. "Magnetotactic bacteria." *Science* 190 (1975): 377-9. <https://www.doi.org/10.1126/science.170679>
2. Blakemore, R. P. "Magnetotactic bacteria." *Annu Rev Microbiol* 36 (1982): 217-38. <https://www.doi.org/10.1146/annurev.mi.36.100182.001245>
3. Schüler, D., C. L. Montel and C. T. Lefevre. "Magnetospirillum gryphiswaldense." *Trends in Microbiology* 28 (2020): 947-48. <https://www.doi.org/10.1016/j.tim.2020.06.001>
4. Müller, F. D., D. Schüler and D. Pfeiffer. "A compass to boost navigation: cell biology of bacterial magnetotaxis." *Journal of bacteriology* 202 (2020): e00398-20. <https://www.doi.org/10.1128/jb.00398-20>
5. Zwiener, T., M. Dziuba, F. Mickoleit, C. Rückert, T. Busche, J. Kalinowski, R. Uebe and D. Schüler. "Towards a 'chassis' for bacterial magnetosome biosynthesis: genome streamlining of Magnetospirillum gryphiswaldense by multiple deletions." *Microb Cell Fact* 20 (2021): 35. <https://www.doi.org/10.1186/s12934-021-01517-2>
6. Niu, W., Y. Zhang, J. Liu, T. Wen, T. Miao, A. Basit and W. Jiang. "OxyR controls magnetosome formation by regulating magnetosome island (MAI) genes, iron metabolism, and redox state." *Free Radical Biology and Medicine* 161 (2020): 272-82. <https://www.doi.org/10.1016/j.freeradbiomed.2020.10.015>
7. Lohsse, A., I. Kolinko, O. Raschdorf, R. Uebe, S. Borg, A. Brachmann, J. M. Plitzko, R. Muller, Y. Zhang and D. Schuler. "Over-production of magnetosomes by genomic amplification of biosynthetic gene clusters in a magnetotactic bacterium." *Appl Environ Microbiol* (2016): <https://www.doi.org/10.1128/AEM.03860-15>
8. Arakaki, A., J. Webb and T. Matsunaga. "A novel protein tightly bound to bacterial magnetic particles in *Magnetospirillum magneticum* strain AMB-1." *The Journal of Biological Chemistry* 278 (2003): 8745-50. <https://www.doi.org/10.1074/jbc.M211729200>
9. Wang, L., T. Prozorov, P. E. Palo, X. Liu, D. Vaknin, R. Prozorov, S. Mallapragada and M. Nilsen-Hamilton. "Self-assembly and biphasic iron-binding characteristics of Mms6, a bacterial protein that promotes the formation of superparamagnetic magnetite nanoparticles of uniform size and shape." *Biomacromolecules* 13 (2012): 98-105. <https://www.doi.org/10.1021/bm201278u>
10. Prozorov, T., S. Mallapragada, B. Narasimhan, L. Wang, P. Palo, M. Nilsen-Hamilton, T. Williams, D. Bazylinski, R. Prozorov and P. Canfield. "Protein-mediated synthesis of uniform superparamagnetic magnetite nanocrystals." *Advanced Functional Materials* 17 (2007): 951-57. <https://www.doi.org/10.1002/adfm.200600448>
11. Tanaka, M., E. Mazuyama, A. Arakaki and T. Matsunaga. "Mms6 protein regulates crystal morphology during nano-sized magnetite biominerilization in vivo." *The Journal of Biological Chemistry* 286 (2011): 6386-92. <https://www.doi.org/10.1074/jbc.M110.183434>
12. Mirabello, G., J. J. Lenders and N. A. Sommerdijk. "Bioinspired synthesis of magnetite nanoparticles." *Chem Soc Rev* (2016): <https://www.doi.org/10.1039/c6cs00432f>
13. Bain, J., C. J. Legge, D. L. Beattie, A. Sahota, C. Dirks, J. R. Lovett and S. S. Staniland. "A biomimetic magnetosome: formation of iron oxide within carboxylic acid terminated polymersomes." *Nanoscale* 11 (2019): 11617-25. <https://www.doi.org/10.1039/c9nr00498j>
14. Bird, S. M., A. E. Rawlings, J. M. Galloway and S. S. Staniland. "Using a biomimetic membrane surface experiment to investigate the activity of the magnetite biominerilisation protein Mms6." *RSC advances* 6 (2016): 7356-63. <https://www.doi.org/10.1039/c5ra16469a>
15. Bird, S. M., J. M. Galloway, A. E. Rawlings, J. P. Bramble and S. S. Staniland. "Taking a hard line with biotemplating: cobalt-doped magnetite magnetic nanoparticle arrays." *Nanoscale* 7 (2015): 7340-51. <https://www.doi.org/10.1039/c5nr00651a>
16. Ali, J., N. Ali, L. Wang, H. Waseem and G. Pan. "Revisiting the mechanistic pathways for bacterial mediated synthesis of noble metal nanoparticles." *Journal of Microbiological Methods* 159 (2019): 18-25. <https://www.doi.org/10.1016/j.mimet.2019.02.010>
17. Chen, H., J. Sun, Z. Wang, Y. Zhou, Z. Lou, B. Chen, P. Wang, Z. Guo, H. Tang, J. Ma, et al. "Magnetic cell-scaffold interface constructed by superparamagnetic IONP enhanced osteogenesis of adipose-derived stem cells." *ACS Applied Materials & Interfaces* 10 (2018): 44279-89. <https://www.doi.org/10.1021/acsami.8b17427>
18. Mori, J. F., S. Lu, M. Händel, K. U. Totsche, T. R. Neu, V. V. Iancu, N. Tarcea, J. Popp and K. Küsel. "Schwertmannite formation at cell junctions by a new filament-forming Fe(II)-oxidizing isolate affiliated with the novel genus Acidithrix." *Microbiology* 162 (2016): 62-71. <https://www.doi.org/10.1099/mic.0.000205>
19. Feng, S., L. Wang, P. Palo, X. Liu, S. K. Mallapragada and M. Nilsen-Hamilton. "Integrated self-assembly of the Mms6 magnetosome protein to form an iron-responsive structure." *International journal of molecular sciences* 14 (2013): 14594-606. <https://www.doi.org/10.3390/ijms140714594>
20. Wang, W., W. Bu, L. Wang, P. E. Palo, S. Mallapragada, M. Nilsen-Hamilton and D. Vaknin. "Interfacial properties and iron binding to bacterial proteins that promote the growth of magnetite nanocrystals: X-ray reflectivity and surface spectroscopy studies." *Langmuir* 28 (2012): 4274-82. <https://www.doi.org/10.1021/la205074n>
21. Baumgartner, J., A. Dey, P. H. Bomans, C. Le Coadou, P. Fratzl, N. A. Sommerdijk and D. Faivre. "Nucleation and growth of magnetite from solution." *Nat Mater* 12 (2013): 310-4. <https://www.doi.org/10.1038/nmat3558>
22. Staniland, S., B. Ward, A. Harrison, G. van der Laan and N. Telling. "Rapid magnetosome formation shown by real-time x-ray magnetic circular dichroism." *Proc Natl Acad Sci U S A* 104 (2007): 19524-8. <https://www.doi.org/10.1073/pnas.0704879104>
23. Frankel, R. B. and R. P. Blakemore. "Precipitation of Fe₃O₄ in Magnetotactic Bacteria." *Philosophical Transactions of the Royal Society of London Series B-Biological Sciences* 304 (1984): 567-74. <https://www.doi.org/10.1098/rstb.1984.0047>
24. Frankel, R. B., G. C. Papaefthymiou, R. P. Blakemore and W. Obrien. "Fe₃O₄ Precipitation in Magnetotactic Bacteria." *Biochimica Et Biophysica Acta* 763 (1983): 147-59. [https://www.doi.org/10.1016/0167-4889\(83\)90038-1](https://www.doi.org/10.1016/0167-4889(83)90038-1)

25. Fdez-Gubieda, M. L., A. Muela, J. Alonso, A. Garcia-Prieto, L. Olivi, R. Fernandez-Pacheco and J. M. Barandiaran. "Magnetite biomineralization in *Magnetospirillum gryphiswaldense*: time-resolved magnetic and structural studies." *ACS Nano* 7 (2013): 3297-305. <https://www.doi.org/10.1021/nn3059983>

26. Arakaki, A., A. Yamagishi, A. Fukuyo, M. Tanaka and T. Matsunaga. "Co-ordinated functions of Mms proteins define the surface structure of cubo-octahedral magnetite crystals in magnetotactic bacteria." *Molecular Microbiology* 93 (2014): 554-67. <https://www.doi.org/10.1111/mmi.12683>

27. Yang, J. and Y. Zhang. "I-TASSER server: new development for protein structure and function predictions." *Nucleic Acids Research* 43 (2015): W174-W81. <https://www.doi.org/10.1093/nar/gkv342>

28. Zhang, Y. and J. Skolnick. "TM-align: a protein structure alignment algorithm based on the TM-score." *Nucleic Acids Research* 33 (2005): 2302-09. <https://www.doi.org/10.1093/nar/gki524>

29. Waterhouse, A., M. Bertoni, S. Bienert, G. Studer, G. Tauriello, R. Gumienny, F. T. Heer, T. A. P. de Beer, C. Rempfer, L. Bordoli, et al. "SWISS-MODEL: homology modelling of protein structures and complexes." *Nucleic Acids Research* 46 (2018): W296-W303. <https://www.doi.org/10.1093/nar/gky427>

30. Tian, W., C. Chen, X. Lei, J. Zhao and J. Liang. "CASTp 3.0: computed atlas of surface topography of proteins." *Nucleic Acids Research* 46 (2018): W363-W67. <https://www.doi.org/10.1093/nar/gky473>

31. Beaugrand, M., A. A. Arnold, J. Henin, D. E. Warschawski, P. T. F. Williamson and I. Marcotte. "Lipid concentration and molar ratio boundaries for the use of isotropic bicelles." *Langmuir* 30 (2014): 6162-70. <https://www.doi.org/10.1021/la5004353>

32. Amemiya, Y., A. Arakaki, S. S. Staniland, T. Tanaka and T. Matsunaga. "Controlled formation of magnetite crystal by partial oxidation of ferrous hydroxide in the presence of recombinant magnetotactic bacterial protein Mms6." *Biomaterials* 28 (2007): 5381-9. <https://www.doi.org/10.1016/j.biomaterials.2007.07.051>

33. Lohsse, A., S. Borg, O. Raschdorf, I. Kolinko, E. Tompa, M. Posfai, D. Faivre, J. Baumgartner and D. Schuler. "Genetic dissection of the *mamAB* and *mms6* operons reveals a gene set essential for magnetosome biogenesis in *Magnetospirillum gryphiswaldense*." *Journal of Bacteriology* 196 (2014): 2658-69. <https://www.doi.org/10.1128/Jb.01716-14>

34. Zhang, X., K.-H. Krause, I. Xenarios, T. Soldati and B. Boeckmann. "Evolution of the ferric reductase domain (FRD) superfamily: Modularity, functional diversification, and signature motifs." *PLoS one* 8 (2013): e58126. <https://www.doi.org/10.1371/journal.pone.0058126>

35. Rawlings, A. E., P. Liravi, S. Corbett, A. S. Holehouse and S. S. Staniland. "Investigating the ferric ion binding site of magnetite biominerallisation protein Mms6." *PLoS one* 15 (2020): e0228708-e08. <https://www.doi.org/10.1371/journal.pone.0228708>

36. Hasan, N. and E. W. Nester. "Purification and characterization of NADPH-dependent flavin reductase. An enzyme required for the activation of chorismate synthase in *Bacillus subtilis*." *The Journal of Biological Chemistry* 253 (1978): 4987-92. [https://www.jbc.org/article/S0021-9258\(17\)34645-8/pdf](https://www.jbc.org/article/S0021-9258(17)34645-8/pdf)

37. Nguyen, H. V., E. Suzuki, Z. Oestreicher, H. Minamide, H. Endoh, Y. Fukumori and A. Taoka. "A protein-protein interaction in magnetosomes: TPR protein MamA interacts with an Mms6 protein." *Biochem Biophys Rep* 7 (2016): 39-44. <https://www.doi.org/10.1016/j.bbrep.2016.05.010>

38. Peigneux, A., Y. Jabalera, M. A. F. Vivas, S. Casares, A. I. Azuaga and C. Jimenez-Lopez. "Tuning properties of biomimetic magnetic nanoparticles by combining magnetosome associated proteins." *Scientific Reports* 9 (2019): 8804. <https://www.doi.org/10.1038/s41598-019-45219-7>

39. Wan, J., P. J. Browne, D. M. Hershey, E. Montabana, A. T. Iavarone, K. H. Downing and A. Komeili. "A protease-mediated switch regulates the growth of magnetosome organelles in *Magnetospirillum magneticum*." *Proc Natl Acad Sci U S A* 119 (2022). <https://www.doi.org/10.1073/pnas.2111745119>

40. Cain, T. J. and A. T. Smith. "Ferric iron reductases and their contribution to unicellular ferrous iron uptake." *J Inorg Biochem* 218 (2021): 111407. <https://www.doi.org/10.1016/j.jinorgbio.2021.111407>

41. Duan, N., S. Wu, X. Chen, Y. Huang and Z. Wang. "Selection and Identification of a DNA Aptamer Targeted to *Vibrio parahaemolyticus*." *Journal of Agricultural and Food Chemistry* 60 (2012): 4034-38. <https://www.doi.org/10.1021/jf300395z>

42. Poget, S. F. and M. E. Girvin. "Solution NMR of membrane proteins in bilayer mimics: small is beautiful, but sometimes bigger is better." *Biochimica et Biophysica Acta (BBA)-Biomembranes* 1768 (2007): 3098-106. <https://www.doi.org/10.1016/j.bbamem.2007.09.006>

43. Lu, X., Y. Zhang and Y.-K. Shin. "Supramolecular SNARE assembly precedes hemifusion in SNARE-mediated membrane fusion." *Nat Struct Mol Biol* 15 (2008): 700-06. <https://www.doi.org/10.1038/nsmb.1433>

44. Riemer, J., H. H. Hoepken, H. Czerwinska, S. R. Robinson and R. Dringen. "Colorimetric ferrozine-based assay for the quantitation of iron in cultured cells." *Analytical Biochemistry* 331 (2004): 370-75. <https://www.doi.org/10.1016/j.ab.2004.03.049>

45. Stookey, L. L. "Ferrozine---a new spectrophotometric reagent for iron." *Analytical Chemistry* 42 (1970): 779-81. <https://www.doi.org/10.1021/ac60289a016>

46. Williams, H. D. and R. K. Poole. "Reduction of iron(III) by *Escherichia coli* K12: Lack of involvement of the respiratory chains." *Current Microbiology* 15 (1987): 319-24. <https://www.doi.org/10.1007/BF01577588>

47. Coves, J. and M. Fontecave. "Reduction and mobilization of iron by a NAD(P)H:flavin oxidoreductase from *Escherichia coli*." *Eur J Biochem* 211 (1993): 635-41. <https://www.doi.org/10.1111/j.1432-1033.1993.tb17591.x>

48. Fischer, E., B. Strehlow, D. Hartz and V. Braun. "Soluble and membrane-bound ferrisiderophore reductases of *Escherichia coli* K-12." *Arch Microbiol* 153 (1990): 329-36. <https://www.doi.org/10.1007/bf00249001>

49. Shen, C., R. Menon, D. Das, N. Bansal, N. Nahar, N. Guduru, S. Jaegle, J. Peckham and Y. K. Reshetnyak. "The protein fluorescence and structural toolkit: Database and programs for the analysis of protein fluorescence and structural data." *Proteins: Structure, Function, and Bioinformatics* 71 (2008): 1744-54. <https://www.doi.org/10.1002/prot.21857>

50. Huang, H., N. B. Suslov, N.-S. Li, S. A. Shelke, M. E. Evans, Y. Koldobskaya, P. A. Rice and J. A. Piccirilli. "A G-quadruplex-containing RNA activates fluorescence in a GFP-like fluorophore." *Nat Chem Biol* 10 (2014): 686-91. <https://www.doi.org/10.1038/nchembio.1561>