

Review

Boosting the Immune Response – Combining the Local and Immune Therapy for Prostate Cancer Treatment

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Abstract: Due to slow progression and susceptibility to radical forms of treatment low-grade PC is associated with high overall survival (OS). With the clinical progression of PC the therapy is getting more complex. The immunosuppressive tumor microenvironment (TME) makes PC a difficult target for most immunotherapeutics. Its general immune resistance is established by i.e. immune evasion through Treg cells, synthesis of immunosuppressive mediators, and defective expression of surface neoantigens. The success of sipuleucel-T in clinical trials initiated several other clinical studies that specifically target the immune escape of the tumor and eliminate the immunosuppressive properties of TME. In the settings of PC treatment, this can be commonly achieved with radiation therapy (RT). Also, focal therapies usually applied for localized PC, such as high-intensity focused ultrasound (HIFU) therapy, cryotherapy, photodynamic therapy (PDT), or irreversible electroporation (IRE) were shown to boost anti-cancer response. Nevertheless, the present guidelines restrict their application to localized and low-grade PC. This review explains how RT and focal therapies enhance the immune response. We also provide data supporting the combination of RT and focal treatments with immune therapies.

Keywords: metastatic castration-resistant prostate cancer; cancer vaccines; immunotherapy; focal therapy; combination immunotherapy; tumor immune microenvironment; in vivo vaccination

1. Introduction

In 2020, prostate cancer (PC) was the second most frequent cancer and the fifth cause of cancer-related death among men. In more than half of the countries of the world it was the most frequently diagnosed cancer in men [1]. While mortality rates are relatively low in comparison to other malignancies, metastatic castration-resistant prostate cancer (mCRPC) remains an incurable condition, with few treatment strategies providing any clinical benefit [2].

Focal therapies are minimally invasive treatment strategies used in the management of PC to provide a local control of the disease, minimizing the risk of possible complications. Despite their limitations, these strategies show some promising oncological results, especially from a short-term perspective [3]. Immunological impact of focal therapies, as well as immunotherapy of PC itself, have been addressed by academic research for years now. Thus, it was a substantial breakthrough when sipuleucel-T became the first therapeutic vaccine for patients with mCRPC approved by the United States Food and Drug Administration (FDA), and the first autologous cellular therapeutic

vaccine in oncology [4,5]. Nevertheless, the clinical benefit of immunotherapy alone remains limited due to low-grade inflammation in the tumor microenvironment (TME). However, both radiotherapy (RT), and various focal therapies have the potential for activating the anti-tumor immune response, and, therefore, enhance efficacy of immunotherapy [6–8].

The purpose of this review is to identify the immune properties of RT, and focal therapies, including high-intensity focused ultrasound (HIFU), cryotherapy, photodynamic therapy (PDT), and irreversible electroporation (IRE). Furthermore, we attempted to compile available knowledge on different combinational therapies including both a focal and an immunotherapeutic component.

2. Immunological Background of Prostate Cancer

2.1. PC Microenvironment

Microenvironment of PC consists of numerous elements, including both neoplastic cells, and diverse host cells. The host component comprises stromal cells and extracellular matrix, endothelial and vascular cells, immune cells, and various soluble factors [9]. The tumor microenvironment (TME) in PC plays an ambiguous role in carcinogenesis. Particularly, the impact of the immune system is highly complex, as both innate and adaptive immune response mechanisms can provide anti-neoplastic activity, as well as propagate carcinogenesis [10]. For example, cytotoxic T-lymphocytes (CTLs), which are one of the most important cancer cell killers, are also able to secrete transforming growth factor-beta (TGF- β), which both supports tumor growth, and induces immune suppression [11].

There is a multitude of mechanisms affecting the TME in PC, including inhibition of neoantigens expression and instability of rapid cell division, DNA damage response (DDR) genes defects, decreased human leucocyte antigens (HLA) expression, phosphatase and tensin homolog (PTEN) protein loss, and dysfunction of in interferon (IFN) type I signalling [12].

2.2. T-cell Infiltration

Many immune cell types play a role in TME functionality, although one considered most vital, is the T-cell population, especially CTLs [13]. They are the key elements of the physiological cancer immunity cycle, which is briefly summarized in Figure 1 [14]. T-cells are recruited from peripheral blood after antigen-presenting cells (APCs), specifically dendritic cells (DCs) and macrophages, capture neoantigens released by the tumor. Presenting the abovementioned antigens to CTLs using a major histocompatibility complex (MHC) is called priming, and takes place in the local lymph nodes. This results in recruiting and stimulating more T-cells, including CD4 $^{+}$ cells. CTLs infiltrate the tumor, recognize cancer cells and kill them. Neoantigens are then released and the process comes full circle [14]. Localization and density of tumor infiltrating lymphocytes (TILs) and memory T-cells within the center of the tumor and its margins were the foundation for creating the “immunoscore”. It divides tumors into two groups: T-cell inflamed (“hot”) and non-T-cell inflamed (“cold”) [15]. This immune contexture is significant in efficacy of the therapy in the variety of cancers. Many publications indicate that a high level of TILs shows a positive prognostic value [16–21]. PC is primarily described as a “cold” tumor, with a low inflammation burden and immune activation [22]. However, the impact of TME on PC oncological outcomes is unclear [12]. Some studies show that the high intratumoral density of CTLs is associated with improved cancer-specific survival (CSS) in PC patients undergoing RP [23,24]. Others show that the higher the level of CTLs infiltration in PC, the greater the risk of distant metastases and biochemical recurrence [10,25]. Although the connection between inflammation and tumorigenesis remains unclear, one of the main goals of various local pre-immunotherapy technics is to propagate inflammation of TME, converting it to inflamed and susceptible to immunotherapy [26].

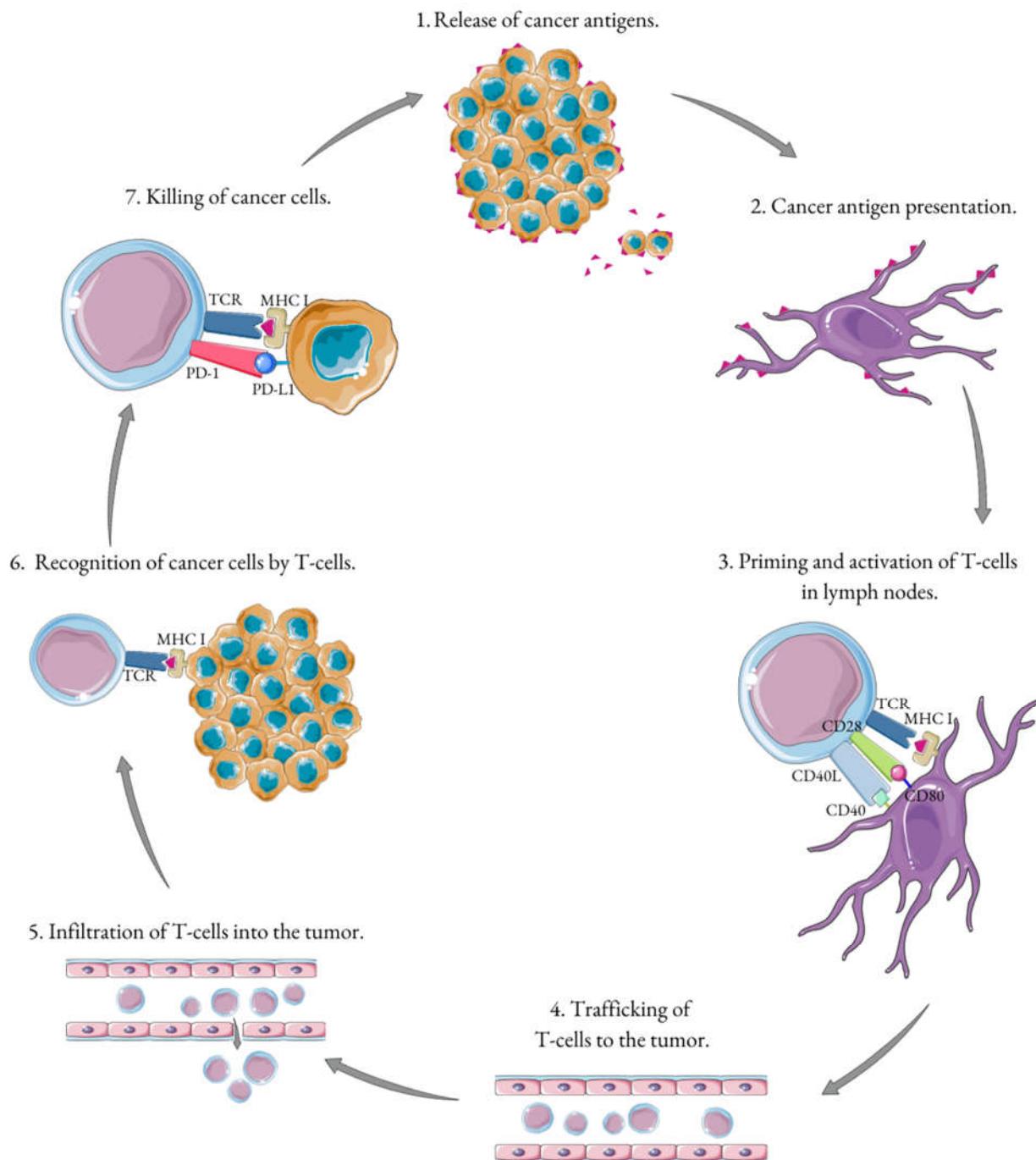


Figure 1. Schematic overview of cancer immunity cycle.

2.3. Regulation of the T-cell Response

After a T-cell is initially activated during priming, the second step of activation takes place: binding of costimulatory molecules, CD80 (B7-1) or CD86 (B7-2), which serve as ligands on APCs, and CD28, a receptor expressed on T-cells [27–29]. Cytotoxic T-lymphocyte antigen 4 (CTLA-4, or CD152) is a coinhibitory glycoprotein receptor expressed on the surface of the T-cell, competing with CD28 for B7 ligands. CTLA-4 is induced after T-cell activation (except for regulatory T cells [Tregs], which express it continuously), and because of its higher affinity for B7 molecules, it successfully outcompetes CD28 receptor [30–33]. The B7:CTLA4 interaction leads to inhibition of cell cycle progression through IL-2 accumulation [34,35]. Programmed death receptor 1 (PD-1) is another co-inhibitory receptor on the surface of T and B-cells. PD-1 ligand 1 (PD-L1)

or CD274) and PD-1 ligand 2 (PD-L2 or CD273) are two known ligands for PD-1 receptor, expressed on macrophages, DCs and other immune cells [36]. Although the interaction of PD-L2 and PD-1 has an immunosuppressive outcome, it is the PD-L1:PD-1 binding, that induces the conversion of naïve T-cells into Tregs [37–40]. CTLA-4, PD-1 and its ligands are parts of B7 superfamily molecules and are the most vital immune checkpoints (ICPs) [41].

2.4. Immune Evasion Mechanisms

Cancer cells have developed several immune evasion mechanisms associated with TME components. Immune evasion may be described as the entirety of biochemical interactions leading to the suppression of the natural immune response to tumor cells. The spectrum of possible “back doors” can be generally divided into a few mechanisms. These include 1) immune evasion through immune cells (most notably Tregs), 2) synthesis of immune-suppressive mediators, and 3) defective expression of surface neoantigens [42].

2.4.1. The Role of Specific Immune Cells

One of the cancer immune evasion mechanisms is CD4+ CD25+ FOXP3+ Tregs activity, as their physiological role is to modulate effector T-cells to support immunological tolerance to self-antigens (self-Ags) [43–45]. Tregs drawn by the tumor have higher suppressive properties compared to circulating Tregs, and are able to inhibit anti-tumor activity of other immune cells directly by cell-cell interactions or indirectly through synthesis and secretion of mediators, e.g. TGF- β , interleukin 10 (IL-10) [46,47]. Many tumor-associated Ags are expressed by host cells and can therefore act like self-Ags, which further emphasizes the Tregs role in immune evasion [48,49].

MDSCs are another heterogeneous group comprising immature DCs, granulocytes, and macrophages. Overproduction and concentration of these cell types in an inflammatory environment are correlated with the immunosuppressive qualities of TME [50,51]. Their functions include the inhibition of CLTs through various mechanisms (e.g. producing reactive oxygen species (ROS) or interactions with T-cell receptor [TCR]), suppressing natural killer (NK) cells, and Tregs induction [52–56]. MDSCs level correlate with the stage of PC, applied treatment, as well as with serum levels of crucial inflammatory mediators – IL-6 and IL-8 [57–59].

DCs are the most professional and efficient APCs, but their functionality is impaired due to tumor's modulatory activity. Impaired DCs have lower levels of CD80, CD86, and CD40, thus they cannot present antigens and activate T-cells effectively enough [42,60]. The role of CD40 is highly complex, as it connects the T and B-cell responses. Namely, when DCs remain active and secrete IL-12, they may interact with CD40L on both T-cells and B-cells [61–64]. The first interaction induces the Th1 and IFN- γ secretion by the T-cells and the latter induces the class switching between IgG and IgA in B cells [65–67]. Also, the reciprocal expression of CD40 and CD40L on DCs, T cells and B cells links the humoral and cellular immune response, thus the reduced level of CD40 might lead to the impairments in both responses [67].

Tumor-associated macrophages (TAMs) are another important group contributing to PC TME. TAMs, especially M2 type, can stimulate tumor growth through the secretion of various mediators such as TGF- β , IL-10 and vascular endothelial growth factor (VEGF) [42,68–70]. Overexpression of TAMs in PC is correlated with unfavorable oncological outcomes in patients with PC, including biochemical recurrence (BCR), or worse distant metastasis-free survival [71–73].

The role of B-cells in immune evasion is not well understood in the case of PC. However, B-cells infiltration has prognostic significance in different cancers like breast cancer and melanoma [74]. B-cell TILs secrete a significant member of the TNF family, the lymphotoxin (LT), which promotes survival and proliferation of androgen-deprived cells, therefore encouraging castration-resistant PC (CRPC) development [75].

2.4.2. Immunosuppressive Mediators

There are many immunosuppressive cytokines, which aid tumor in the immune evasion through the promotion of tumor proliferation, chemoresistance, angiogenesis, or migration, and these are most notably TGF- β , VEGF, IL-6, RANKL, or CXCL family [76]. TGF- β is one of the most vital mediators, acting both as a direct growth-promoting factor, as well as a stimulator of CD4+ T-cells-Tregs transformation [77–79]. Its other roles include promoting angiogenesis, and downregulating HLA-1 expression, thus inducing epithelial-mesenchymal transition (EMT) [76,80–83]. Another important cytokine is VEGF, which also contributes to tumor growth, as well as inhibits DCs differentiation. A similar role is performed by cancer-associated ganglioside antigens, which conduct an immunosuppressive activity through impairing CTLs and DCs [42,84,85].

2.4.3. Dysfunctional Expression of Surface Neoantigens

MHC Class I proteins are found on nucleated cells and platelet surfaces and their role is to be recognized by CD8+ T cells, which trigger the immune response against certain antigens by activating T cells and leading to target cell destruction [5,86,87]. Decreased MHC I presentation of tumor-associated antigens is one of the immune evasion mechanisms of PC [88,89].

2.5. Immune Check-Point Inhibitors

Immune checkpoint inhibitors (ICIs) are novel treatment options gaining more and more interest, as they already appeared to be successful strategies in cancers such as melanoma or lung cancer [90–96]. Among CTLA-4 inhibitors there are ipilimumab and tremelimumab, while the most pivotal PD-1 inhibitors comprise nivolumab and pembrolizumab; atezolizumab belongs to PD-L1 inhibitors [97].

Immune checkpoint blockade in PC remains a poor monotherapeutic tool [98,99]. Among the reasons for this state there are the low level of T-cell infiltration, “cold” immunogenic profile of the tumor, mutational burden, and immune evasion mechanisms [100–102].

3. Cancer Vaccines

3.1. Dendritic Cell Vaccines – Sipuleucel-T, DCvac/PCa, and others

As mentioned above, DCs are one of the most important features of the immune system; they are the most efficient APCs, not only able to activate T-cells (both Tregs and CTLs), but also NK cells. DC vaccines require blood-derived DCs, pulsing them ex vivo with the tumor-associated antigen and activating them by the specific adjuvant, and then reinjecting them to the patient [103]. The first DC vaccine approved by the U.S. Food and Drug Administration (FDA) was sipuleucel-T (Provenge®), and so far it remains the only DC vaccine for mCRPC [104,105]. Sipuleucel-T promotes the immune response against tumor cells using prostatic acid phosphatase (PAP) antigen-activated DCs [41]. A double-blind, placebo-controlled, multicenter phase III trial compared this DC vaccine to the placebo group, with the results of a 22% reduction in the risk of death and more than 4 months of improvement in overall survival (OS) [4]. Another trial showed even greater improvement in OS (up to 8.1 months), if sipuleucel-T therapy is extended by the APC8015F, a variant of the DC vaccine prepared from cryopreserved cells, which were frozen for future use [106].

DC vaccines are a very promising therapeutic tool, although requiring further clinical trials, and more attempts of combining them with different approaches [107]. There is only one ongoing trial assessing the combination of sipuleucel-T and other therapies: sipuleucel-T plus stereotactic ablative body radiation (SABR) (NCT01818986, phase II). Different phase III trial evaluates the efficiency of sipuleucel-T in reducing the progression of CRPC. The study includes active surveillance patients (the ProVent Study; NCT03686683).

DCvac/PCa is an autologous DC-based vaccine, in which case DCs are pulsed with killed lymph node carcinoma of the prostate (LNCaP) cells. Several clinical trials have investigated its efficacy in PC. Podrazil et al. researched the combination of DCvac/PCa and docetaxel in phase I/II clinical trial in mCRPC, concluding this strategy is characterized by longer OS [108]. A similar study was conducted by Kongsted et al., which compared the same combination with docetaxel alone. PFS and disease-specific survival were comparable in both arms [109]. Fucikova et al. assessed the DCvac/PCa impact on PSA in patients with rising PSA after RP or salvage RT. PSA doubling time was elongated significantly in this variant [110]. Although DCvac/PCa immunological impact is quite well documented by now, translation to clinical benefits is needed and further clinical trials are required, especially concerning different combinations of therapies. A recent clinical phase III trial (the Viable) by Vogelzang et al. investigated DCvac/PCa combination with docetaxel and prednisone. The therapy failed to improve OS in patients with mCRPC [111].

Other DC-based vaccines that have been tested in the last decade in PC patients are prostate-specific membrane antigen (PSMA) and survivin loaded DC vaccine, mucin 1 (MUC1) vaccine, or T-cell receptor γ alternate reading frame protein (TARP) vaccine [112–115].

3.2. PROSTVAC – a PSA-Based Viral Vector Vaccine

One of the trailblazing PC vaccines is PROSTVAC (PSA-TRICOM), which comprises two recombinant poxvirus vectors containing transgenes for PSA and three costimulatory molecules: B7.1, ICAM-1, and LFA-3 [116,117]. A phase II trial analyzing neoadjuvant PROSTVAC in patients awaiting RP showed an increase in CD4+ and CD8+ T-cell infiltration of the tumor, as well as the peripheral immune response to neoantigens in 13 of 25 patients [118]. This promising immune response doesn't yet translate into a clinical advantage, however. In a phase III trial Gulley et al. concluded that despite the therapy was well-tolerated and safe for patients, treatment had no impact on median OS and alive without events (AWE) in patients with mCRPC, disappointingly [119]. Parsons et al. evaluated the preventive value of PROSTVAC in patients with localized PC which is managed by an active surveillance strategy. Although some initial data on the immunological effect of the vaccine is already available, we are looking forward to the summary of this phase II trial in the future (NCT02326805) [120]. Madan et al. revealed that the addition of PROSTVAC to enzalutamide doesn't affect PSA levels. The authors concluded that in this particular combination PROSTVAC effect may get lost and remain unseen due to patients' response to enzalutamide [121].

Several ongoing clinical trials are investigating different combinational management strategies including PROSTVAC. These are evaluating, among others, combination with nivolumab (NCT02933255, phase I/II) or nivolumab and ipilimumab (NCT03532217, phase I), with CV301 (a poxviral vaccine) and M7824 (a protein targeting PD-L1 and TGF- β) (NCT03315871, phase II), docetaxel (NCT02649855, phase II), or enzalutamide (NCT01867333, phase II).

TroVax is another viral vector, 5T4 (oncofoetal glycoprotein) targeting vaccination. It's characterized by the good immune response in mCRPC and the potential to efficiently combine with docetaxel [122,123].

3.3. Peptide-Based Vaccines

Among peptide-based vaccines, one of the most interesting is GX301, consisting of four telomerase peptides and two adjuvants – Montanide ISA-51 and Imiquimod. Fenoglio et al. assessed its potential in phase I/II clinical trial, revealing its immunological response in PC and renal cell cancer (RCC). An increase in PFS and OS were observed as well [124]. Filaci et al. evaluated GX301 efficiency and immunological impact in mCRPC. The therapy didn't increase OS, though they observed that higher numbers of drug administration were correlated with increased immunological response [125].

Cell division associated 1 (CDCA1) peptide vaccination was a topic of research in phase I clinical trial by Obara et al. CDCA1 is a peptide overexpressed in a few malignancies, including PC. Authors indicated that the vaccine is well-tolerated, and it boosts immunological response in patients with CRPC. Additionally, they pointed out that CDCA1 vaccine therapy might increase survival rates and aid to maintain the quality of life of CRPC patients, but further clinical trials are required to prove that [126,127].

Other peptide-based vaccinations include personalized peptide vaccination (PPV), which includes administration of different HLA-matched peptides, multi-peptide vaccines, and a vaccine targeting Ras homolog gene family member C (RhoC) [128–130]. Their clinical use requires further phase II and III trials in the future.

3.4. Whole-Tumor-Cell Vaccines

GVAX is a vaccine consisting of genetically modified PC cells, which undergone radiation. Studies suggest that this vaccination induces the immune response by activation of DCs and MDSCs [131]. A combinational therapy with ipilimumab has been investigated in a phase I trial by van den Eertwegh et al., which showed that GVAX is well-tolerated and safe for patients with mCRPC [132]. Once again, further clinical trials are required [133].

4. Focal Ablation and Immune Therapy Combination

4.1. High-Intensity Focused Ultrasound

Lately, HIFU appeared as a potential neoadjuvant-like therapy, serving as the first step of immunotherapeutic treatment. HIFU itself has already made an appearance in guidelines, concerning PC treatment options, although only as an investigational therapeutic tool, or as salvage therapy [134]. The most important benefit of HIFU is that it is minimally invasive when compared to surgical treatment, and it is devoid of systemic toxicity in comparison with androgen deprivation therapy (ADT) or chemotherapy, nevertheless possible adverse effects may occur quite frequently, and they include erectile dysfunction, urinary tract infections, rectal injuries, and more [135,136]. Properties of HIFU can be divided into a few groups – ablative and non-ablative (mechanical), immune, and biological effects; induced activity depends on a multitude of factors including frequency, pressure, duty cycle, and treatment time, achieved temperature, tissue susceptibility, and more. This allows to distinguish several possible technique variants, such as thermal ablation, thermal stress and hyperthermia, mechanical perturbation, or histotripsy [137]. However, first and foremost effect of HIFU is thermal ablation (by heating tumor tissue above approximately 55°C), resulting in coagulative necrosis, combined with additional cavitation formation, the most captivating secondary effect is anti-tumor immunity induction [138,139].

HIFU immunotherapeutic effect has lately been investigated in many kinds of malignancies. Hu et al. confirmed HIFU promotes DCs infiltration and activation in mice bearing colon adenocarcinoma and indicated that the mechanical components of this procedure may be successfully combined with other types of therapy [140]. Ran et al. showed that HIFU increases peripheral blood CD3+, CD4+ levels and CD4+/CD8+ ratio, enhances CTLs cytotoxicity against murine hepatocarcinoma, and inhibits tumor growth and progression in mice [141]. The impact on the CD4+/CD8+ ratio has been observed in the past by Rosberger et al. [142]. Activation of anti-tumor immunity promoted by HIFU can be partially explained by tumor debris “left-over” antigens immunogenicity, which was demonstrated by Zhang et al. in the murine hepatocellular carcinoma model [143]. Similar investigations have been conducted with other malignancies, such as melanoma, neuroblastoma, or pancreatic cancer [144–149]. Wu et al. researched tumor debris immunogenic properties in 23 patients with breast cancer. Using HIFU, they ablated primary tumors, and evaluated the expression of tumor antigens and heat-shock protein 70 (HSP-70), also pointing out the immunogenic potential of neoplastic debris [150].

Sonodynamic therapy (SDT) is another promising strategy concerning the usage of ultrasound. It is based on the application of sonosensitizers, which is followed by their activation with the ultrasound. Activated particles then transfer the energy to oxygen accumulated in TME, creating ROS, which kill or damage tumor cells [151,152]. HIFU, and the spectrum of ultrasound-based therapies in general, is still a very modern approach used for enhancing the immune response. Further investigation is required, especially concerning PC.

4.2. Cryotherapy

Cryoablation or cryotherapy performed either as a focal therapy, or as the whole-gland procedure, is an ablation technique using extremely low temperatures to induce both necrosis and apoptosis of tumor cells. With the use of special cryoprobes, liquid nitrogen or argon, passing from high pressure to an atmospheric pressure revealing its cooling effect, is implemented inside a prostate gland. Although it may be used as monotherapy, for this review we will only focus on its immunomodulatory activity and its synergy with immunotherapy.

Cryotherapy has a great enhancing potential to enhance the immune response, due to its significant preservation of tumor antigens and cytokines, compared to other ablation techniques based on high temperatures rather than hypothermia [153]. It is believed to leave tumor's intracellular molecules intact and, through attracting the immune system by these factors, stimulate tumor-specific immunity. However, cryotherapy can prompt both immunostimulatory and immunosuppressive response, which is strongly dependent on the type of induced cell death; studies suggest that necrosis, occurring mainly in the inner zone of the tissue, causes tumor cells to release danger-associated molecular patterns, which boost the immune response through the maturation of DCs, and consequently T-cells activation. However, apoptosis occurring primarily in the peripheral margin of the ablated organ leads to a lack of secretion of danger signals, therefore caring immunosuppressive impact [154]. The cryoimmunological effect is further described by the term "abscopal effect". This rare phenomenon refers to the systemic immunological impact a focal therapy has, and primarily refers to the reduction of a metastasis preceded by a localized treatment in a different location [155]. This process was proved to be mediated by CD8+ T-cells and correlated with a low level of CD4+CD5+ Tregs, as well as an increased level of IFN- γ [156,157].

Various investigations have been conducted on the theme of cryoimmunological synergy, both in murine models and in clinical trials. For instance, Gaitanis and Bassukas researched the impact of immunocryosurgery on basal cell carcinomas (BCC). Their study indicated that cryoablation combined with TLR7 agonist, imiquimod, can be a very effective substitute for surgical treatment for BCC under 20 mm in diameter [158]. In another study, Lin et al. prospectively evaluated allogeneic NK cell immunotherapy combined with cryosurgery in renal cell carcinoma (RCC). They once again proved a synergistic effect of the two therapies [159]. The same group of researchers conducted similar investigations in patients with lung and hepatocellular cancers, with similarly favorable outcome results [160,161].

So far clinical trials including synergy of cryosurgery and immunotherapy in patients with PC are rarely conducted. One of them is a therapy using granulocyte-macrophage colony-stimulating factor (GM-CSF), a cytokine regulating functions of granulocytes and macrophages, as well as promoting survival of DCs [162,163]. These investigations revealed that GM-CSF administration enhances INF- γ secretion by T-cells on the base of prior cryoablation procedures, as well as the fact that GM-CSF increases levels of prostate-specific and nonspecific antigens. Ross et al. examined cryosurgery combined with short term ADT and pembrolizumab, a PD-1 inhibitor, proving local disease control, but questioning its potential for management of systemic disease [164].

4.3. Photodynamic Therapy

PDT is an example of another targeted treatment option, that has already been used as an alternative to radical therapies, with intention of reducing levels of side effects, while maintaining favorable oncological outcomes [165]. This focal therapy is based on the usage of a laser of a specific wavelength, which activates the photosensitizer (PS), administered systemically or locally, and therefore generates ROS resulting in necrosis of the tumor cells. Depending on the qualities of photoagents, different effects can be achieved. Photothermal therapy (PTT) is a subtype of phototherapy different from PDT, as it engages PS properties not to produce ROS, but to execute a thermal effect through the conversion of absorbed laser light into heat [166].

As for PDT in PC, researchers point out high efficacy and low level of adverse effects of vascular-targeted photodynamic therapy (VTP) in comparison to other therapies, while addressing the great need for long-term benefit evaluation in randomized clinical trials (RCT) [167]. Rastinehad et al. introduced the results of a clinical trial in which they used gold-silica nanoshells (AuroShells) to conduct PTT in 15 patients with PC. The study revealed high-profile feasibility of the procedure, and once again pointed out its low-rate adverse effects burden [168]. Another study by Azzouzi et al. compared padeliporfin VTP with active surveillance strategy in a phase III RCT. They evaluated VTP as a safe and effective treatment for low-risk, localized PC, with a longer time to progression and a higher proportion of negative biopsy results in comparison to active surveillance [169]. On the other hand, a review of this investigation, aroused by the Oncologic Drugs Advisory Committee within the FDA, resulted in voting against approval of this therapeutic strategy in the United States, which emphasizes that the topic requires more RCTs proving its safety and efficacy [170]. Besides, a lot more clinical trials have been conducted, evaluating different doses of various PS, varying laser wavelengths, and manipulating other parameters [170].

Nevertheless, more and more papers these days have been turning its attention to immunological aspects of PDT, as it propagates inflammatory response, induces necrosis, and promotes recruitment of neutrophils, and other immune cells. Furthermore, PDT can promote immune cells and engage them to eradicate distant metastases [171]. Therefore, the term photoimmunotherapy (PIT) has been forged, and it may be described as a combination of immunogenic properties of PDT and immunotherapy treatment [172]. The immunological effect obtained by PDT is complex and multi-level. First of all, it affects immune cells directly through the recruitment of neutrophils, DCs maturation, and macrophage activation, as well as accumulation of CTLs and affecting them through regulation of NK cells migration [173,174]. Secretion of IL-1 α/β , IL-6, IL-8, IL-10 and IL-12 is boosted, as is the release of a few secondary inflammatory mediators, including thromboxane and prostaglandins [175]. Furthermore, a few strategies concerning the combination of PDT with different immunotherapeutic strategies have been conducted and their results are promising. Li et al. evaluated the synergistic effect of CTLA-4 antibodies and single-walled carbon nanotube-glycated chitosan complex (SWNT-GC) in metastatic mammary tumors in mice. Local administration was then followed by PTT. The results showed that this strategy prolonged survival time, suppressed primary tumors, and inhibited metastases [176]. Huang et al. introduced a drug conjugate consisting of protoporphyrin IX and NLG919, a potent indoleamine-2,3-dioxygenase (IDO) inhibitor, which is applied to the cells through liposomal delivery (PpIX-NLG@Lipo). They showed its strong ability to generate ROS after phototherapeutic procedure, as well as its potential of increasing CD8+ T-cells infiltration [177]. Kim et al. investigated the impact of PDT with Ce6-embedded nanophotosensitizer (FIC-PDT) with rapasudil, a rho-kinase (ROCK) inhibitor on the immune response in mice with uveal melanoma. Their research indicated that this combination demonstrates vaccine-like function, leads to CD8+ T-cells accumulation in the primary tumor and, in further synergy with anti-PD-L1 antibody, to metastasis inhibition [178].

Nagaya et al. presented the effects of near-infrared photoimmunotherapy (NIR-PIT) with prostate-specific membrane antigen (PSMA) antibody in the PC cell line. The anti-PSMA antibody was conjugated to the light-absorbing agent, IR700DX. This antibody-PS conglomerate was observed to bind cell-specifically and to effectively kill PC cells after activation using NIR-PIT, with over two-thirds of the investigated tumors cured [179]. Research on the same topic was conducted by Watanabe et al. and it pointed to the possibility of using only fragments of anti-PSMA antibodies instead of the full antibodies, which may clinically translate to a more thorough penetration of the tumor milieu. Using smaller parts of antibodies should also shorten the time gap between injection of the PS and NIR-PIT [180].

4.4. Irreversible Electroporation

IRE is the permeabilization of cell membranes with electrical pulses, which affect membranous electrochemical potentials, creating pores in a lipid bilayer [181]. IRE has been already used in PC management, both as a focal therapy, and as the whole gland ablation. The procedure is based on needle electrodes, which are placed inside or nearby the targeted tissue. Then short electrical pulses are delivered, which induces apoptosis through a non-thermal mechanism [182]. Despite its role in the immune response is still unexplored, IRE seems to have immunomodulatory properties. The most pivotal immunological effect of IRE is a decrease of Tregs in TME; additionally, a decrease of MDSCs occurs as well [183].

The field of IRE-immunotherapy combinations in treating malignancies is still uninvestigated, though there are a few articles, especially on pancreatic cancer. Yang et al. for example revealed a connection between IRE and tumor-associated immune evasion in a mice model of pancreatic ductal adenocarcinoma (PDAC). They indicated that IRE combined with DC cancer vaccination increases the level of tumor-infiltrating cells including CD8+ T-cells and granzyme B+ cells in PDAC [184]. Similar investigations have been conducted by Zhao et al. and by He et al. Both studies showed promising results of the combination of IRE and PD-1 inhibitors in mice with PDAC [185,186].

A study by Burbach et al. examined the combination of IRE and ICI in mice with PC. Focal treatment using IRE combined with ICI led to the expansion of tumor-specific CD8+ T-cells in blood and TME [187].

5. Radiation and Immune Therapy Combination

RT has been used as a management strategy both in PC and in many other malignancies for years now. Its primary property exploited for the tumor treatment purposes was the effect on double-strand DNA, leading to its breakdown, and thus resulting in cell death, majorly through senescence, slightly less frequently through mitotic catastrophe, apoptosis, and necrosis [188]. Traditionally RT was considered to be a therapy of immunosuppressive qualities, therefore its combination with immunotherapy appeared to be irrational at first [189]. However, rapidly growing interest in TME affected the way RT is perceived, as its game with the immune system is far more complex and ambiguous [190,191].

Immune-stimulating effect of RT is generally achieved through induced cell death and modulating the composition of TME. One of the initial steps following tumor cell damage is enhanced release of damage-associated molecules, such as calreticulin, adenosine triphosphate (ATP), GM-CSF, high-mobility group box 1 (HMGB1), or heat shock proteins (HSPs) [192,193]. Afterwards, these damage signals activate DCs and APCs, which takes place in lymph nodes and leads to priming naïve T-cells in consequence [193]. Additionally, one of the radiation effects is the release of other inflammatory molecules, such as chemokines (e.g. CXCL10 or CXCL16) and other cytokines, including IL-1 β , TNF- α , and type 1 and 2 interferons, which further contribute to increase inflammation in TME [194]. Finally, RT triggers upregulation of MHC I, NKG2D ligand, Fas/CD95, and other co-stimulatory molecules, resulting in cell death and further antigen exposure [193,194].

Eckert et al. investigated the impact of RT on the immune system in 18 patients with localized PC. The study revealed the ambiguous effect of ionizing radiation, as RT resulted in a decrease in absolute leukocyte and lymphocyte counts, and an increase in Tregs and NK cells counts after over eight weeks since radiation. However, during RT an increase was observed in all immune cells counts excluding Tregs. Importantly, the percentage of CD8+ T-cells had its peak early during RT [195]. Nevertheless, Harris et al. researched a combination of RT and immunotherapy in a transgenic murine model and observed that the anti-tumor immune response occurred when immune therapy was administered 3 to 5 weeks after RT [19]. This further suggests the existence of a certain type of therapeutic time window, in which immunostimulatory properties of RT are emphasized, and the immunosuppressive component is partially inhibited. Nickols et al. researched the impact of stereotactic body radiotherapy (SBRT) on immunological homeostasis in a clinical trial evaluating resected prostate specimens of 16 patients. While prostates without SBRT were mainly lymphoid-diverse, specimens after SBRT were immunologically dominated by myeloid cells [196]. Keam et al. proved in their 24 patient clinical trial that high dose-rate brachytherapy (HDRBT) has a substantial potential in enhancing inflammation in prostate. In response to HDRBT an increase in CD4+ T-cells, macrophages and DCs counts was observed. Moreover, they evaluated tumor inflammation signature (TIS) and concluded that 80% of immunologically “cold” tumors were converted to “intermediate” or “hot” types [26].

Interestingly, RT is another management strategy with proven abscopal effect, hence resulting in regression of metastases, probably due to the outburst of tumor-associated antigens. This extremely rare effect is observed more often when RT is combined with immune therapy, particularly with checkpoint inhibitors [192]. Dudzinski et al. studied the combination of anti-PD-1 or anti-PD-L1 and radiation in mice, and they not only observed an increase in median survival in comparison to the drug alone (70% longer for anti-PD-1 and 130% for anti-PD-L1), but also detected the abscopal effect – a regression of unirradiated distant metastases [197].

In the research concerning the effects of RT and immunotherapy combination in mice, there have been a few distinguishing articles, including the paper from Wada et al. They assessed the efficacy of this therapy (immunotherapeutic component being GM-CSF) using an autochthonous model of PC. Improved OS and increase of the effector-to-regulatory TILs ratio, as well as treatment effect in both primary tumor and metastases, were observed [198]. Another investigation by Philippou et al. assayed the combination of anti-PD-L1 and RT and its impact on TME in PC. They observed macrophages and DCs counts increase, as well as upregulation of PD-1/PD-L1 in both arms of the study 7 days after RT. Radiation was observed to delay tumor growth and affect TME immunological composition. However, PD-L1 inhibition administered in one of the arms didn't affect tumor growth delay when compared to monotherapy [199]. Table 1 presents ongoing trials evaluating different combinations of RT and immunotherapy in PC management.

Table 1. Ongoing trials assessing combination of radiotherapy and immunotherapy.

NCT Number	Phase	n	Setting	Immunotherapeutics	Radiotherapy
NCT03835533	I	45	mCRPC	NKTR-214, Nibolumab, CDX-301, Poly-ICLC, INO-5151	SBRT
NCT03795207	II	96	mPC	Durvalumab	SBRT
NCT03543189	I/II	44	PC	Nivolumab	Brachytherapy, EBRT
NCT03217747	I/II	173	mCRPC	Anti-OX40, Avelumab, Utomilumab	RT*
NCT03007732	II	42	PC	Pembrolizumab, SD-101	SBRT
NCT01818986	II	20	mCRPC	Sipuleucel-T	SBRT
NCT01436968	III	711	PC	Aglatimagine Besadenovec	EBRT

NCT: The National Clinical Trial; n: number of patients enrolled; PC: prostate cancer; mPC: metastatic prostate cancer; mCRPC: metastatic castration-resistant prostate cancer; RT: radiotherapy; SBRT: stereotactic body radiation therapy; EBRT: external beam radiation therapy.

* The specific variant of radiation therapy wasn't specified.

The efficiency of radioimmunotherapy in patients with PC has been explored willingly in clinical trials for the last 10 years. Slovin et al. assessed the combination of anti-CTLA-4 antibody, ipilimumab, with external-beam radiotherapy (EBRT) in comparison to the drug alone. This phase I/II study on 50 patients evaluated adverse effects, defining them as manageable, and indicated anti-tumor activity [200]. In another investigation, a phase III trial concerning ipilimumab versus placebo after radiotherapy in patients with mCRPC that progressed after docetaxel chemotherapy has been conducted by Kwon et al. No notable difference in OS was found, though ipilimumab use was associated with a decrease in PSA levels and an increase in progression-free survival. Additionally, OS increase was observed in the ipilimumab subgroup without visceral metastases, with non-raised or mildly raised alkaline phosphatase, and without anaemia. Accordingly, the authors suggested that a specific constellation of prognostic features could potentially enhance clinical outcomes of radioimmunotherapy [201]. The final analysis of this phase III trial revealed that OS was two to three times higher at 3 years and beyond in favor of radiotherapy and ipilimumab combination [201]. Different clinical trials assessing nivolumab and brachytherapy or EBRT, as well as sipuleucel-T and EBRT combinations indicated that these therapies are safe and well-tolerated, though immunogenic effect and anti-tumor activity of radiation with nivolumab were observed, while radiation with sipuleucel-T showed no particular increase in the immune response [202,203]. Another phase II trial assessed the combination of sipuleucel-T and a radioisotope, radium-223, in patients with mCRPC. Despite paradoxically decreased the immune response in the combination arm, PSA levels were decreased and PFS and OS longer [204]. A case report by Han et al. presents a significant clinical response to pembrolizumab and radiation combination in patient heavily treated mCRPC with rectal involvement. After radiation and six cycles of the drug PSA was undetectable, prostate mass was decreased and rectal invasion was imperceptible in imaging studies [205].

Table 2 presents a comparison of the immunomodulatory impact of different local therapies on TME.

Table 2. Immunomodulatory impact of local treatment strategies on TME.

Local Therapy	Immunomodulatory Effects	References
HIFU	<ul style="list-style-type: none"> Promotion of DCs infiltration and activation. Increase of CD3+ and CD4+ levels cells, and CD4+/CD8+ ratio. Enhancement of CTLs cytotoxicity. 	[140,141,150–152,142–149]
Cryotherapy	<ul style="list-style-type: none"> Activation of T-cells. DCs maturation. The abscopal effect. 	[154,155,164,156–163]
PDT	<ul style="list-style-type: none"> Promotion of neutrophils recruitment. DCs maturation. Activation of macrophages. Regulation of CTLs and NK cells migration, increase of CD8+ T-cells infiltration. Secretion of IL-1, IL-6, IL-8, IL-10, IL-12, thromboxane and prostaglandins. 	[171–180]
IRE	<ul style="list-style-type: none"> Decrease of Tregs and MDSCs levels. 	[183–187]
RT	<ul style="list-style-type: none"> Enhancement of damage-associated molecules release. Activation of DCs and other APCs. Release of various cytokines (e.g. CXCL10, CXCL16, IL-1, TNF-α, interferons). Upregulation of MHC I, NKG2D ligand and Fas/CD95. The abscopal effect. 	[188,192,201–205,193–200]

HIFU: high-intensity focused ultrasound; PDT: photodynamic therapy; IRE: irreversible electroporation; RT: radiotherapy; DC: dendritic cell; CTL: cytotoxic T-lymphocyte; NK: natural killer; IL: interleukin; Treg: regulatory T cell; MDSC: myeloid-derived suppressor cell; APC: antigen-presenting cell; TNF: tumor necrosis factor; MHC: major histocompatibility complex.

6. Conclusions

The immunotherapy for PC remains an unexplored field, despite the initial success of sipuleucel-T. Further phase I/II clinical trials investigating combinations of focal and immune therapies are highly desirable. The RT and immunotherapy combo is an approach of the greatest potential to increase anti-tumor qualities of TME. Thus, it may be the most effective strategy stimulating the cancer-related immune response in PC.

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References

1. Sung, H.; Ferlay, J.; Siegel, R.L.; Laversanne, M.; Soerjomataram, I.; Jemal, A.; Bray, F. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA. Cancer J. Clin.* **2021**, *71*, 209–249, doi:10.3322/caac.21660.
2. He, L.; Fang, H.; Chen, C.; Wu, Y.; Wang, Y.; Ge, H.; Wang, L.; Wan, Y.; He, H. Metastatic Castration-Resistant Prostate Cancer: Academic Insights and Perspectives through Bibliometric Analysis. *Med. (United States)* **2020**, *99*, doi:10.1097/MD.00000000000019760.
3. Ahdoot, M.; Lebastchi, A.H.; Turkbey, B.; Wood, B.; Pinto, P.A. Contemporary Treatments in Prostate Cancer Focal Therapy. *Curr. Opin. Oncol.* **2019**, *31*, 200–206, doi:10.1097/CCO.0000000000000515.
4. May, K.F.; Gulley, J.L.; Drake, C.G.; Dranoff, G.; Kantoff, P.W. Prostate Cancer Immunotherapy. *Clin. Cancer Res.* **2011**, *17*, 5233–5238, doi:10.1158/1078-0432.CCR-10-3402.
5. Bansal, D.; Reimers, M.A.; Knoche, E.M.; Pachynski, R.K. Immunotherapy and Immunotherapy Combinations in Metastatic Castration-Resistant Prostate Cancer. *Cancers (Basel)* **2021**, *13*, 1–22, doi:10.3390/cancers13020334.
6. Carvalho, H. de A.; Villar, R.C. Radiotherapy and Immune Response: The Systemic Effects of a Local Treatment. *Clinics* **2018**, *73*, doi:10.6061/CLINICS/2018/E557S.
7. Borges, R.C.; Tourinho-Barbosa, R.R.; de la Rosette, J. Tumour Microenvironment and Focal Therapy for Prostate Cancer. *Curr. Opin. Urol.* **2022**, *32*, 248–253, doi:10.1097/MOU.0000000000000987.
8. Chavez, M.; Silvestrini, M.T.; Ingham, E.S.; Fite, B.Z.; Mahakian, L.M.; Tam, S.M.; Illovitsh, A.; Monjazeb, A.M.; Murphy, W.J.; Hubbard, N.E.; et al. Distinct Immune Signatures in Directly Treated and Distant Tumors Result from TLR Adjuvants and Focal Ablation. *Theranostics* **2018**, *8*, 3611–3628, doi:10.7150/thno.25613.
9. Dai, J.; Lu, Y.; Roca, H.; Keller, J.M.; Zhang, J.; McCauley, L.K.; Keller, E.T. Immune Mediators in the Tumor Microenvironment of Prostate Cancer. *Chin. J. Cancer* **2017**, *36*, 1–8, doi:10.1186/s40880-017-0198-3.
10. Giraldo, N.A.; Sanchez-Salas, R.; Peske, J.D.; Vano, Y.; Becht, E.; Petitprez, F.; Validire, P.; Ingels, A.; Cathelineau, X.; Fridman, W.H.; et al. The Clinical Role of the TME in Solid Cancer. *Br. J. Cancer* **2019**, *120*, 45–53, doi:10.1038/s41416-018-0327-z.
11. Donkor, M.K.; Sarkar, A.; Savage, P.A.; Franklin, R.A.; Johnson, L.K.; Jungbluth, A.A.; Allison, J.P.; Li, M.O. T Cell Surveillance of Oncogene-Induced Prostate Cancer Is Impeded by T Cell-Derived TGF-B1 Cytokine. *Immunity* **2011**, *35*, 123–134, doi:10.1016/j.jimmuni.2011.04.019.
12. Vitkin, N.; Nersesian, S.; Siemens, D.R.; Koti, M. The Tumor Immune Contexture of Prostate Cancer. *Front. Immunol.* **2019**, *10*, 1–10, doi:10.3389/fimmu.2019.00603.
13. Kwon, J.T.W.; Bryant, R.J.; Parkes, E.E. The Tumor Microenvironment and Immune Responses in Prostate Cancer Patients. *Endocr. Relat. Cancer* **2021**, *28*, T95–T107, doi:10.1530/ERC-21-0149.
14. Chen, D.S.; Mellman, I. Oncology Meets Immunology: The Cancer-Immunity Cycle. *Immunity* **2013**, *39*, 1–10, doi:10.1016/j.jimmuni.2013.07.012.
15. Angell, H.; Galon, J. From the Immune Contexture to the Immunoscore: The Role of Prognostic and Predictive Immune Markers in Cancer. *Curr. Opin. Immunol.* **2013**, *25*, 261–267, doi:10.1016/j.coi.2013.03.004.
16. Gao, G.; Wang, Z.; Qu, X.; Zhang, Z. Prognostic Value of Tumor-Infiltrating Lymphocytes in Patients with Triple-Negative Breast Cancer: A Systematic Review and Meta-Analysis. *BMC Cancer* **2020**, *20*, doi:10.1186/s12885-020-6668-z.
17. Ropponen, K.M.; Eskelinen, M.J.; Lipponen, P.K.; Alhava, E.; Kosma, V.M. Prognostic Value of Tumour-Infiltrating Lymphocytes (TILs) in Colorectal Cancer. *J. Pathol.* **1997**, *182*, 318–324, doi:10.1002/(SICI)1096-9896(199707)182:3<318::AID-PATH862>3.0.CO;2-6.
18. Fu, Q.; Chen, N.; Ge, C.; Li, R.; Li, Z.; Zeng, B.; Li, C.; Wang, Y.; Xue, Y.; Song, X.; et al. Prognostic Value of Tumor-Infiltrating Lymphocytes in Melanoma: A Systematic Review and Meta-Analysis. *Oncotarget* **2019**, *8*, doi:10.1080/2162402X.2019.1593806.
19. Harris, T.J.; Hipkiss, E.L.; Borzillary, S.; Wada, S.; Grosso, J.F.; Yen, H.R.; Getnet, D.; Bruno, T.C.; Goldberg, M. V.; Pardoll, D.M.; et al. Radiotherapy Augments the Immune Response to Prostate Cancer in a Time-Dependent Manner. *Prostate* **2008**, *68*, 1319–1329, doi:10.1002/pros.20794.
20. Li, J.; Wang, J.; Chen, R.; Bai, Y.; Lu, X. The Prognostic Value of Tumor-Infiltrating T Lymphocytes in Ovarian Cancer. *Oncotarget* **2017**, *8*, 15621–15631, doi:10.18632/oncotarget.14919.
21. Ding, W.; Xu, X.; Qian, Y.; Xue, W.; Wang, Y.; Du, J.; Jin, L.; Tan, Y. Prognostic Value of Tumor-Infiltrating Lymphocytes in Hepatocellular Carcinoma A Meta-Analysis. *Med. (United States)* **2018**, *97*, doi:10.1097/MD.00000000000013301.
22. de Bono, J.S.; Guo, C.; Gurel, B.; De Marzo, A.M.; Sfanos, K.S.; Mani, R.S.; Gil, J.; Drake, C.G.; Alimonti, A. Prostate Carcinogenesis: Inflammatory Storms. *Nat. Rev. Cancer* **2020**, *20*, 455–469, doi:10.1038/s41568-020-0267-9.
23. Yang, Y.; Attwood, K.; Bshara, W.; Mohler, J.L.; Guru, K.; Xu, B.; Kalinski, P.; Chatta, G. High Intratumoral CD8+ T-Cell Infiltration Is Associated with Improved Survival in Prostate Cancer Patients Undergoing Radical Prostatectomy. *Prostate* **2021**, *81*, 20–28, doi:10.1002/pros.24068.
24. Vicier, C.; Werner, L.; Huang, Y.; Hamid, A.; Evan, C.; Loda, M.; Sweeney, C. Immune Infiltrate with CD8 Low or PDL1 High Associated with Metastatic Prostate Cancer after Radical Prostatectomy (RP). *J. Clin. Oncol.* **2019**, *37*, 86–86, doi:10.1200/jco.2019.37.7_suppl.86.

25. Ness, N.; Andersen, S.; Valkov, A.; Nordby, Y.; Donnem, T.; Al-Saad, S.; Busund, L.T.; Bremnes, R.M.; Richardsen, E. Infiltration of CD8+ Lymphocytes Is an Independent Prognostic Factor of Biochemical Failure-Free Survival in Prostate Cancer. *Prostate* **2014**, *74*, 1452–1461, doi:10.1002/pros.22862.
26. Keam, S.P.; Halse, H.; Nguyen, T.; Wang, M.; Van Kooten Losio, N.; Mitchell, C.; Caramia, F.; Byrne, D.J.; Haupt, S.; Ryland, G.; et al. High Dose-Rate Brachytherapy of Localized Prostate Cancer Converts Tumors from Cold to Hot. *J. Immunother. cancer* **2020**, *8*, doi:10.1136/jitc-2020-000792.
27. Hammerstrom, A.E.; Cauley, D.H.; Atkinson, B.J.; Sharma, P. Cancer Immunotherapy: Sipuleucel-T and Beyond. *Pharmacotherapy* **2011**, *31*, 813–828, doi:10.1592/phco.31.8.813.
28. Melichar, B.; Nash, M.A.; Lenzi, R.; Platsoucas, C.D.; Freedman, R.S. Expression of Costimulatory Molecules CD80 and CD86 and Their Receptors CD28, CTLA-4 on Malignant Ascites CD3+ Tumour-Infiltrating Lymphocytes (TIL) from Patients with Ovarian and Other Types of Peritoneal Carcinomatosis. *Clin. Exp. Immunol.* **2000**, *119*, 19–27, doi:10.1046/j.1365-2249.2000.01105.x.
29. Nakajima, A.; Watanabe, N.; Yoshino, S.; Yagita, H.; Okumura, K.; Azuma, M. Requirement of CD28-CD86 Co-Stimulation in the Interaction between Antigen-Primed T Helper Type 2 and B Cells. *Int. Immunopharmacol.* **1997**, *9*, 637–644, doi:10.1093/intimm/9.5.637.
30. Takahashi, B.T.; Tagami, T.; Yamazaki, S.; Uede, T.; Shimizu, J.; Sakaguchi, N.; Mak, T.W.; Sakaguchi, S. Immunologic Self-Tolerance Maintained by CD25. *J. Exp. Med.* **2000**, *192*.
31. Alegre, M.L.; Frauwirth, K.A.; Thompson, C.B. T-Cell Regulation by CD28 and CTLA-4. *Nat. Rev. Immunol.* **2001**, *1*, 220–228, doi:10.1038/35105024.
32. Rudd, C.E.; Taylor, A.; Schneider, H. CD28 and CTLA-4 Coreceptor Expression and Signal Transduction. *Immunol. Rev.* **2009**, *229*, 12–26, doi:10.1111/j.1600-065X.2009.00770.x.
33. Sansom, D.M. CD28, CTLA-4 and Their Ligands: Who Does What and to Whom? *Immunology* **2000**, *101*, 169–177, doi:10.1046/j.1365-2567.2000.00121.x.
34. Krummel, M.F.; Allison, J.P. CTLA-4 Engagement Inhibits IL-2 Accumulation and Cell Cycle Progression upon Activation of Resting T Cells. *J. Exp. Med.* **1996**, *183*, 2533–2540, doi:10.1084/jem.183.6.2533.
35. Belani, R.; Weiner, G.J. Expression of Both B7-1 and CD28 Contributes to the IL-2 Responsiveness of CTLL-2 Cells. *Immunology* **1996**, *87*, 271–274, doi:10.1046/j.1365-2567.1996.461532.x.
36. Loke, P.; Allison, J.P. PD-L1 and PD-L2 Are Differentially Regulated by Th1 and Th2 Cells. *Proc. Natl. Acad. Sci. U. S. A.* **2003**, *100*, 5336–5341, doi:10.1073/pnas.0931259100.
37. Latchman, Y.; Wood, C.R.; Chernova, T.; Chaudhary, D.; Borde, M.; Chernova, I.; Iwai, Y.; Long, A.J.; Brown, J.A.; Nunes, R.; et al. PD-L2 Is a Second Ligand for PD-1 and Inhibits T Cell Activation. *Nat. Immunol.* **2001**, *2*, 261–268, doi:10.1038/85330.
38. Wang, L.; Pino-lagos, K.; Vries, V.C. De; Guleria, I.; Sayegh, M.H.; Noelle, R.J. PNAS July 8, 2008 Vol. 105 No. 27 9331–9336.Pdf. **2008**, *105*, 9331–9336.
39. Brown, J.A.; Dorfman, D.M.; Ma, F.-R.; Sullivan, E.L.; Munoz, O.; Wood, C.R.; Greenfield, E.A.; Freeman, G.J. Blockade of Programmed Death-1 Ligands on Dendritic Cells Enhances T Cell Activation and Cytokine Production. *J. Immunol.* **2003**, *170*, 1257–1266, doi:10.4049/jimmunol.170.3.1257.
40. Blank, C.; Gajewski, T.F.; Mackensen, A. Interaction of PD-L1 on Tumor Cells with PD-1 on Tumor-Specific T Cells as a Mechanism of Immune Evasion: Implications for Tumor Immunotherapy. *Cancer Immunol. Immunother.* **2005**, *54*, 307–314, doi:10.1007/s00262-004-0593-x.
41. Jafari, S.; Molavi, O.; Kahroba, H.; Hejazi, M.S.; Maleki-Dizaji, N.; Barghi, S.; Kiaie, S.H.; Jadidi-Niaragh, F. Clinical Application of Immune Checkpoints in Targeted Immunotherapy of Prostate Cancer. *Cell. Mol. Life Sci.* **2020**, *77*, 3693–3710, doi:10.1007/s00018-020-03459-1.
42. Vinay, D.S.; Ryan, E.P.; Pawelec, G.; Talib, W.H.; Stagg, J.; Elkord, E.; Lichtor, T.; Decker, W.K.; Whelan, R.L.; Kumara, H.M.C.S.; et al. Immune Evasion in Cancer: Mechanistic Basis and Therapeutic Strategies. *Semin. Cancer Biol.* **2015**, *35*, S185–S198, doi:10.1016/j.semcan.2015.03.004.
43. Sasidharan Nair, V.; Elkord, E. Immune Checkpoint Inhibitors in Cancer Therapy: A Focus on T-Regulatory Cells: A. *Immunol. Cell Biol.* **2018**, *96*, 21–33, doi:10.1111/imcb.1003.
44. Wolf, A.M.; Wolf, D.; Steurer, M.; Gastl, G.; Gunsilius, E.; Grubeck-Loebenstein, B. Increase of Regulatory T Cells in the Peripheral Blood of Cancer Patients. *Clin. Cancer Res.* **2003**, *9*, 606–612.
45. Dannull, J.; Su, Z.; Rizzieri, D.; Yang, B.K.; Coleman, D.; Yancey, D.; Zhang, A.; Dahm, P.; Chao, N.; Gilboa, E.; et al. Enhancement of Vaccine-Mediated Antitumor Immunity in Cancer Patients after Depletion of Regulatory T Cells. *J. Clin. Invest.* **2005**, *115*, 3623–3633, doi:10.1172/JCI25947.
46. Miller, A.M.; Lundberg, K.; Özenci, V.; Banham, A.H.; Hellström, M.; Egevad, L.; Pisa, P. CD4 + CD25 High T Cells Are Enriched in the Tumor and Peripheral Blood of Prostate Cancer Patients . *J. Immunol.* **2006**, *177*, 7398–7405, doi:10.4049/jimmunol.177.10.7398.
47. Karpisheh, V.; Mousavi, S.M.; Naghavi Sheykholeslami, P.; Fathi, M.; Mohammadpour Saray, M.; Aghebati-Maleki, L.; Jafari, R.; Majidi Zolbanin, N.; Jadidi-Niaragh, F. The Role of Regulatory T Cells in the Pathogenesis and Treatment of Prostate Cancer. *Life Sci.* **2021**, *284*, 119132, doi:10.1016/j.lfs.2021.119132.
48. Houghton, A.N.; Guevara-Patiño, J.A. Immune Recognition of Self in Immunity against Cancer. *J. Clin. Invest.* **2004**, *114*, 468–471, doi:10.1172/JCI22685.

49. Touloukian, C.E.; Leitner, W.W.; Robbins, P.F.; Li, Y.F.; Kang, X.; Lapointe, R.; Hwu, P.; Rosenberg, S.A.; Restifo, N.P. Expression of a "Self-" Antigen by Human Tumor Cells Enhances Tumor Antigen-Specific CD4+ T-Cell Function. *Cancer Res.* **2002**, *62*, 5144–5147.
50. Bunt, S.K.; Yang, L.; Sinha, P.; Clements, V.K.; Leips, J.; Ostrand-Rosenberg, S. Reduced Inflammation in the Tumor Microenvironment Delays the Accumulation of Myeloid-Derived Suppressor Cells and Limits Tumor Progression. *Cancer Res.* **2007**, *67*, 10019–10026, doi:10.1158/0008-5472.CAN-07-2354.
51. Choi, J.-N.; Sun, E.G.; Cho, S.-H. IL-12 Enhances Immune Response by Modulation of Myeloid Derived Suppressor Cells in Tumor Microenvironment. *Chonnam Med. J.* **2019**, *55*, 31, doi:10.4068/cmj.2019.55.1.31.
52. Li, H.; Han, Y.; Guo, Q.; Zhang, M.; Cao, X. Cancer-Expanded Myeloid-Derived Suppressor Cells Induce Anergy of NK Cells through Membrane-Bound TGF-B1. *J. Immunol.* **2009**, *182*, 240–249, doi:10.4049/jimmunol.182.1.240.
53. Serafini, P.; Mgebroff, S.; Noonan, K.; Borrello, I. Myeloid-Derived Suppressor Cells Promote Cross-Tolerance in B-Cell Lymphoma by Expanding Regulatory T Cells. *Cancer Res.* **2008**, *68*, 5439–5449, doi:10.1158/0008-5472.CAN-07-6621.
54. Hoechst, B.; Ormandy, L.A.; Ballmaier, M.; Lehner, F.; Krüger, C.; Manns, M.P.; Greten, T.F.; Korangy, F. A New Population of Myeloid-Derived Suppressor Cells in Hepatocellular Carcinoma Patients Induces CD4+CD25+Foxp3+ T Cells. *Gastroenterology* **2008**, *135*, 234–243, doi:10.1053/j.gastro.2008.03.020.
55. Srivastava, M.K.; Sinha, P.; Clements, V.K.; Rodriguez, P.; Ostrand-Rosenberg, S. Myeloid-Derived Suppressor Cells Inhibit T-Cell Activation by Depleting Cystine and Cysteine. *Cancer Res.* **2010**, *70*, 68–77, doi:10.1158/0008-5472.CAN-09-2587.
56. Sanaei, M.J.; Salimzadeh, L.; Bagheri, N. Crosstalk between Myeloid-Derived Suppressor Cells and the Immune System in Prostate Cancer: MDSCs and Immune System in Prostate Cancer. *J. Leukoc. Biol.* **2020**, *107*, 43–56, doi:10.1002/JLB.4RU0819-150RR.
57. Chi, N.; Tan, Z.; Ma, K.; Bao, L.; Yun, Z. Increased Circulating Myeloid-Derived Suppressor Cells Correlate with Cancer Stages, Interleukin-8 and -6 in Prostate Cancer. *Int. J. Clin. Exp. Med.* **2014**, *7*, 3181–3192.
58. Bosas, P.; Zaleskis, G.; Dabkevičiene, D.; Dobrovolskiene, N.; Mlynška, A.; Tikuišis, R.; Ulys, A.; Pašukoniene, V.; Jarmalaitė, S.; Jankevičius, F. Immunophenotype Rearrangement in Response to Tumor Excision May Be Related to the Risk of Biochemical Recurrence in Prostate Cancer Patients. *J. Clin. Med.* **2021**, *10*, doi:10.3390/jcm10163709.
59. Siemińska, I.; Rychlicka-Buniowska, E.; Jaszczyński, J.; Palaczynski, M.; Bukowska-Strakova, K.; Rys, J.; Dumański, J.; Siedlar, M.; Baran, J. The Level of Myeloid Derived-Suppressor Cells in Peripheral Blood of Patients with Prostate Cancer after Various Types of Therapy. *Polish J. Pathol.* **2020**, *71*, 46–54, doi:10.5114/pjp.2020.95415.
60. Murugaiyan, G.; Martin, S.; Saha, B. Levels of CD40 Expression on Dendritic Cells Dictate Tumour Growth or Regression. *Clin. Exp. Immunol.* **2007**, *149*, 194–202, doi:10.1111/j.1365-2249.2007.03407.x.
61. Wykes, M.; Macpherson, G. Dendritic Cell-B-Cell Interaction: Dendritic Cells Provide B Cells with CD40-Independent Proliferation Signals and CD40-Dependent Survival Signals. *Immunology* **2000**, *100*, 1–3, doi:10.1046/j.1365-2567.2000.00044.x.
62. Wesa, A.; Galy, A. Increased Production of Pro-Inflammatory Cytokines and Enhanced T Cell Responses after Activation of Human Dendritic Cells with IL-1 and CD40 Ligand. *BMC Immunol.* **2002**, *3*, doi:10.1186/1471-2172-3-14.
63. Hernandez, M.G.H.; Shen, L.; Rock, K.L. CD40-CD40 Ligand Interaction between Dendritic Cells and CD8+ T Cells Is Needed to Stimulate Maximal T Cell Responses in the Absence of CD4+ T Cell Help. *J. Immunol.* **2007**, *178*, 2844–2852, doi:10.4049/jimmunol.178.5.2844.
64. Hill, K.S.; Errington, F.; Steele, L.P.; Merrick, A.; Morgan, R.; Selby, P.J.; Georgopoulos, N.T.; O'Donnell, D.M.; Melcher, A.A. OK432-Activated Human Dendritic Cells Kill Tumor Cells via CD40/CD40 Ligand Interactions. *J. Immunol.* **2008**, *181*, 3108–3115, doi:10.4049/jimmunol.181.5.3108.
65. Wykes, M.; Pombo, A.; Jenkins, C.; MacPherson, G.G. Dendritic Cells Interact Directly with Naive B Lymphocytes to Transfer Antigen and Initiate Class Switching in a Primary T-Dependent Response. *J. Immunol.* **1998**, *161*, 1313–1319.
66. Kelsall, B.L.; Stüber, E.; Neurath, M.; Strober, W. Interleukin-12 Production by Dendritic Cells. The Role of CD40-CD40L Interactions in Th1 T-Cell Responses. *Ann. N. Y. Acad. Sci.* **1996**, *795*, 116–126, doi:10.1111/j.1749-6632.1996.tb52660.x.
67. Ma, D.Y.; Clark, E.A. The Role of CD40 and CD154/CD40L in Dendritic Cells. *Semin. Immunol.* **2009**, *21*, 265–272, doi:10.1016/j.smim.2009.05.010.
68. de Waal Malefyt, R.; Hans, Y.; Roncarolo, M.G.; Spits, H.; de Vries, J.E. Interleukin-10. *Curr. Opin. Immunol.* **1992**, *4*, 314–320, doi:10.1016/0952-7915(92)90082-P.
69. Solinas, G.; Germano, G.; Mantovani, A.; Allavena, P. Tumor-Associated Macrophages (TAM) as Major Players of the Cancer-Related Inflammation. *J. Leukoc. Biol.* **2009**, *86*, 1065–1073, doi:10.1189/jlb.0609385.
70. Schoppmann, S.F.; Birner, P.; Stöckl, J.; Kalt, R.; Ullrich, R.; Caucig, C.; Nagy, K.; Alitalo, K.; Kerjaschki, D. Tumor-Associated Macrophages Express Lymphatic Endothelial Growth Factors and Are Related to Peritumoral Lymphangiogenesis. *Am. J. Pathol.* **2002**, *161*, 947–956, doi:10.1016/S0002-9440(10)64255-1.
71. Larionova, I.; Tuguzbaeva, G.; Ponomaryova, A.; Stakheyeva, M.; Cherdynseva, N.; Pavlov, V.; Choinzonov, E.; Kzhyshkowska, J. Tumor-Associated Macrophages in Human Breast, Colorectal, Lung, Ovarian and Prostate Cancers. *Front. Oncol.* **2020**, *10*, 1–34, doi:10.3389/fonc.2020.566511.
72. Yuri, P.; Shigemura, K.; Kitagawa, K.; Hadibrata, E.; Risan, M.; Zulfiqar, A.; Soeroharjo, I.; Hendri, A.Z.; Danarto, R.; Ishii, A.; et al. Increased Tumor-Associated Macrophages in the Prostate Cancer Microenvironment Predicted Patients' Survival and Responses to Androgen Deprivation Therapies in Indonesian Patients Cohort. *Prostate Int.* **2020**, *8*, 62–69, doi:10.1016/j.prnil.2019.12.001.

73. Gollapudi, K.; Galet, C.; Grogan, T.; Zhang, H.; Said, J.W.; Huang, J.; Elashoff, D.; Freedland, S.J.; Rettig, M.; Aronson, W.J. Association between Tumor-Associated Macrophage Infiltration, High Grade Prostate Cancer, and Biochemical Recurrence after Radical Prostatectomy. *Am. J. Cancer Res.* **2013**, *3*, 523–529.
74. Woo, J.R.; Liss, M.A.; Muldong, M.T.; Palazzi, K.; Strasner, A.; Ammirante, M.; Varki, N.; Shabaik, A.; Howell, S.; Kane, C.J.; et al. Tumor Infiltrating B-Cells Are Increased in Prostate Cancer Tissue. *J. Transl. Med.* **2014**, *12*, 1–9, doi:10.1186/1479-5876-12-30.
75. Ammirante, M.; Luo, J.L.; Grivennikov, S.; Nedospasov, S.; Karin, M. B-Cell-Derived Lymphotoxin Promotes Castration-Resistant Prostate Cancer. *Nature* **2010**, *464*, 302–305, doi:10.1038/nature08782.
76. Adekoya, T.O.; Richardson, R.M. Cytokines and Chemokines as Mediators of Prostate Cancer Metastasis. *Int. J. Mol. Sci.* **2020**, *21*, 1–29, doi:10.3390/ijms21124449.
77. Konkel, J.E.; Zhang, D.; Zanvit, P.; Chia, C.; Zangarle-Murray, T.; Jin, W.; Wang, S.; Chen, W.J. Transforming Growth Factor- β Signaling in Regulatory T Cells Controls T Helper-17 Cells and Tissue-Specific Immune Responses. *Immunity* **2017**, *46*, 660–674, doi:10.1016/j.jimmuni.2017.03.015.
78. Massagué, J. TGF β in Cancer. *Cell* **2008**, *134*, 215–230, doi:10.1016/j.cell.2008.07.001.
79. Batlle, E.; Massagué, J. Transforming Growth Factor- β Signaling in Immunity and Cancer. *Immunity* **2019**, *50*, 924–940, doi:10.1016/j.jimmuni.2019.03.024.
80. Ferrari, G.; Cook, B.D.; Terushkin, V.; Pintucci, G.; Mignatti, P. Transforming Growth Factor-Beta 1 (TGF-B1) Induces Angiogenesis through Vascular Endothelial Growth Factor (VEGF)-Mediated Apoptosis. *J. Cell. Physiol.* **2009**, *219*, 449–458, doi:10.1002/jcp.21706.
81. Chen, X.H.; Liu, Z.C.; Zhang, G.; Wei, W.; Wang, X.X.; Wang, H.; Ke, H.P.; Zhang, F.; Wang, H.S.; Cai, S.H.; et al. TGF- β and EGF Induced HLA-I Downregulation Is Associated with Epithelial-Mesenchymal Transition (EMT) through Upregulation of Snail in Prostate Cancer Cells. *Mol. Immunol.* **2015**, *65*, 34–42, doi:10.1016/j.molimm.2014.12.017.
82. Zuber, P.; Kuppner, M.C.; Tribolet, N. De Transforming Growth Factor- β 2 Down-regulates HLA-DR Antigen Expression on Human Malignant Glioma Cells. *Eur. J. Immunol.* **1988**, *18*, 1623–1626, doi:10.1002/eji.1830181023.
83. Lebrin, F.; Deckers, M.; Bertolino, P.; Ten Dijke, P. TGF- β Receptor Function in the Endothelium. *Cardiovasc. Res.* **2005**, *65*, 599–608, doi:10.1016/j.cardiores.2004.10.036.
84. McKallip, R.; Li, R.; Ladisch, S. Tumor Gangliosides Inhibit the Tumor-Specific Immune Response. *J. Immunol.* **1999**, *163*, 3718–3726.
85. Péguel-Navarro, J.; Sportouch, M.; Popa, I.; Berthier, O.; Schmitt, D.; Portoukalian, J. Gangliosides from Human Melanoma Tumors Impair Dendritic Cell Differentiation from Monocytes and Induce Their Apoptosis. *J. Immunol.* **2003**, *170*, 3488–3494, doi:10.4049/jimmunol.170.7.3488.
86. Norment, A.M.; Salter, R.D.; Parham, P.; Engelhard, V.H.; Littman, D.R. Cell-Cell Adhesion Mediated by CD8 and MHC Class I Molecules. *Nature* **1988**, *336*, 79–81, doi:10.1038/336079a0.
87. Natarajan, K.; Li, H.; Mariuzza, R.A.; Margulies, D.H. MHC Class I Molecules, Structure and Function. *Rev. Immunogenet.* **1999**, *1*, 32–46.
88. Movassaghi, M.; Chung, R.; Anderson, C.B.; Stein, M.; Saenger, Y.; Faiena, I. Overcoming Immune Resistance in Prostate Cancer: Challenges and Advances. *Cancers (Basel.)* **2021**, *13*, 1–16, doi:10.3390/cancers13194757.
89. Bander, N.H.; Yao, D.; Liu, H.; Chen, Y.T.; Steiner, M.; Zuccaro, W.; Moy, P. MHC Class I and II Expression in Prostate Carcinoma and Modulation by Interferon-Alpha and -Gamma. *Prostate* **1997**, *33*, 233–239, doi:10.1002/(SICI)1097-0045(19971201)33:4<233::AID-PROS2>3.0.CO;2-I.
90. Wolchok, J.D.; Neyns, B.; Linette, G.; Negrier, S.; Lutzky, J.; Thomas, L.; Waterfield, W.; Schadendorf, D.; Smylie, M.; Guthrie, T.; et al. Ipilimumab Monotherapy in Patients with Pretreated Advanced Melanoma: A Randomised, Double-Blind, Multicentre, Phase 2, Dose-Ranging Study. *Lancet Oncol.* **2010**, *11*, 155–164, doi:10.1016/S1470-2045(09)70334-1.
91. Hodi, F.S.; O'Day, S.J.; McDermott, D.F.; Weber, R.W.; Sosman, J.A.; Haanen, J.B.; Gonzalez, R.; Robert, C.; Schadendorf, D.; Hassel, J.C.; et al. Improved Survival with Ipilimumab in Patients with Metastatic Melanoma. *N. Engl. J. Med.* **2010**, *363*, 711–723, doi:10.1056/nejmoa1003466.
92. Robert, C.; Schachter, J.; Long, G. V.; Arance, A.; Grob, J.J.; Mortier, L.; Daud, A.; Carlino, M.S.; McNeil, C.; Lotem, M.; et al. Pembrolizumab versus Ipilimumab in Advanced Melanoma. *N. Engl. J. Med.* **2015**, *372*, 2521–2532, doi:10.1056/nejmoa1503093.
93. Huang, A.C.; Zappasodi, R. A Decade of Checkpoint Blockade Immunotherapy in Melanoma: Understanding the Molecular Basis for Immune Sensitivity and Resistance. *Nat. Immunol.* **2022**, doi:10.1038/s41590-022-01141-1.
94. Herbst, R.S.; Baas, P.; Kim, D.W.; Felip, E.; Pérez-Gracia, J.L.; Han, J.Y.; Molina, J.; Kim, J.H.; Arvis, C.D.; Ahn, M.J.; et al. Pembrolizumab versus Docetaxel for Previously Treated, PD-L1-Positive, Advanced Non-Small-Cell Lung Cancer (KEYNOTE-010): A Randomised Controlled Trial. *Lancet* **2016**, *387*, 1540–1550, doi:10.1016/S0140-6736(15)01281-7.
95. Brahmer, J.; Reckamp, K.L.; Baas, P.; Crino, L.; Eberhardt, W.E.E.; Poddubskaya, E.; Antonia, S.; Pluzanski, A.; Vokes, E.E.; Holgado, E.; et al. Nivolumab versus Docetaxel in Advanced Squamous-Cell Non-Small-Cell Lung Cancer. *N. Engl. J. Med.* **2015**, *373*, 123–135, doi:10.1056/nejmoa1504627.
96. Carlino, M.S.; Larkin, J.; Long, G. V. Immune Checkpoint Inhibitors in Melanoma. *Lancet* **2021**, *398*, 1002–1014, doi:10.1016/S0140-6736(21)01206-X.
97. Seidel, J.A.; Otsuka, A.; Kabashima, K. Anti-PD-1 and Anti-CTLA-4 Therapies in Cancer: Mechanisms of Action, Efficacy, and Limitations. *Front. Oncol.* **2018**, *8*, 1–14, doi:10.3389/fonc.2018.00086.
98. Venkatachalam, S.; McFarland, T.R.; Agarwal, N.; Swami, U. Immune Checkpoint Inhibitors in Prostate Cancer. *Cancers (Basel.)* **2021**, *13*, doi:10.3390/cancers13092187.

99. H, D.; TF, H.; AH, Z. The Current Role of Immunotherapy in MCRPC: A Systematic Review. *Austin J. Clin. Case Reports* **2021**, *8*, doi:10.26420/austinjclincaserep.2021.1222.
100. Karzai, F.; Vanderweele, D.; Madan, R.A.; Owens, H.; Cordes, L.M.; Hankin, A.; Couvillon, A.; Nichols, E.; Bilusic, M.; Beshiri, M.L.; et al. Activity of Durvalumab plus Olaparib in Metastatic Castration-Resistant Prostate Cancer in Men with and without DNA Damage Repair Mutations 1112 Medical and Health Sciences 1112 Oncology and Carcinogenesis. *J. Immunother. Cancer* **2018**, *6*, doi:10.1186/s40425-018-0463-2.
101. Antonarakis, E.S.; Isaacsson Velho, P.; Fu, W.; Wang, H.; Agarwal, N.; Santos, V.S.; Maughan, B.L.; Pili, R.; Adra, N.; Sternberg, C.N.; et al. CDK12 -Altered Prostate Cancer: Clinical Features and Therapeutic Outcomes to Standard Systemic Therapies, Poly (ADP-Ribose) Polymerase Inhibitors, and PD-1 Inhibitors. *JCO Precis. Oncol.* **2020**, 370–381, doi:10.1200/po.19.00399.
102. Graf, R.P.; Fisher, V.; Weerpals, J.; Gjoerup, O.; Tierno, M.B.; Huang, R.S.P.; Sayegh, N.; Lin, D.I.; Raskina, K.; Schrock, A.B.; et al. Comparative Effectiveness of Immune Checkpoint Inhibitors vs Chemotherapy by Tumor Mutational Burden in Metastatic Castration-Resistant Prostate Cancer. *JAMA Netw. Open* **2022**, *5*, doi:10.1001/jamanetworkopen.2022.5394.
103. Sadeghzadeh, M.; Bornehdeli, S.; Mohahammadrezakhani, H.; Abolghasemi, M.; Poursaei, E.; Asadi, M.; Zafari, V.; Aghebati-Maleki, L.; Shanehbandi, D. Dendritic Cell Therapy in Cancer Treatment; the State-of-the-Art. *Life Sci.* **2020**, *254*, 117580, doi:10.1016/j.lfs.2020.117580.
104. Cheever, M.A.; Higano, C.S. PROVENGE (Sipuleucel-T) in Prostate Cancer: The First FDA-Approved Therapeutic Cancer Vaccine. *Clin. Cancer Res.* **2011**, *17*, 3520–3526, doi:10.1158/1078-0432.CCR-10-3126.
105. Polo, S.H.; Muñoz, D.M.; Rodríguez, A.C.R.; Ruiz, J.S.; Rodríguez, D.I.R.; Couñago, F. Changing the History of Prostate Cancer with New Targeted Therapies. *Biomedicines* **2021**, *9*, 1–20, doi:10.3390/biomedicines9040392.
106. George, D.J.; Nabhan, C.; Devries, T.; Whitmore, J.B.; Gomella, L.G. Survival Outcomes of Sipuleucel-T Phase III Studies: Impact of Control-Arm Cross-over to Salvage Immunotherapy. *Cancer Immunol. Res.* **2015**, *3*, 1063–1069, doi:10.1158/2326-6066.CIR-15-0006.
107. Draube, A.; Klein-González, N.; Mattheus, S.; Brillant, C.; Hellmich, M.; Engert, A.; von Bergwelt-Bailedon, M. Dendritic Cell Based Tumor Vaccination in Prostate and Renal Cell Cancer: A Systematic Review and Meta-Analysis. *PLoS One* **2011**, *6*, doi:10.1371/journal.pone.0018801.
108. Podrazil, M.; Horvath, R.; Becht, E.; Rozkova, D.; Bilkova, P.; Sochorova, K.; Hromadkova, H.; Kayserova, J.; Vavrova, K.; Lastovicka, J.; et al. Phase I/II Clinical Trial of Dendritic-Cell Based Immunotherapy (DCVAC/PCa) Combined with Chemotherapy in Patients with Metastatic, Castration-Resistant Prostate Cancer. *Oncotarget* **2015**, *6*, 18192–18205, doi:10.18632/oncotarget.4145.
109. Kongsted, P.; Borch, T.H.; Ellebaek, E.; Iversen, T.Z.; Andersen, R.; Met, Ö.; Hansen, M.; Lindberg, H.; Sengelov, L.; Svane, I.M. Dendritic Cell Vaccination in Combination with Docetaxel for Patients with Metastatic Castration-Resistant Prostate Cancer: A Randomized Phase II Study. *Cytotherapy* **2017**, *19*, 500–513, doi:10.1016/j.jcyt.2017.01.007.
110. Fucikova, J.; Podrazil, M.; Jarolim, L.; Bilkova, P.; Hensler, M.; Becht, E.; Gasova, Z.; Klouckova, J.; Kayserova, J.; Horvath, R.; et al. Phase I/II Trial of Dendritic Cell-Based Active Cellular Immunotherapy with DCVAC/PCa in Patients with Rising PSA after Primary Prostatectomy or Salvage Radiotherapy for the Treatment of Prostate Cancer. *Cancer Immunol. Immunother.* **2018**, *67*, 89–100, doi:10.1007/s00262-017-2068-x.
111. Vogelzang, N.J.; Beer, T.M.; Gerritsen, W.; Oudard, S.; Wiechno, P.; Kukielka-Budny, B.; Samal, V.; Hajek, J.; Feyerabend, S.; Khoo, V.; et al. Efficacy and Safety of Autologous Dendritic Cell-Based Immunotherapy, Docetaxel, and Prednisone vs Placebo in Patients with Metastatic Castration-Resistant Prostate Cancer: The VIABLE Phase 3 Randomized Clinical Trial. *JAMA Oncol.* **2022**, doi:10.1001/jamaoncol.2021.7298.
112. Xi, H.B.; Wang, G.X.; Fu, B.; Liu, W.P.; Li, Y. Survivin and PSMA Loaded Dendritic Cell Vaccine for the Treatment of Prostate Cancer. *Biol. Pharm. Bull.* **2015**, *38*, 827–835, doi:10.1248/bpb.b14-00518.
113. Scheid, E.; Major, P.; Bergeron, A.; Finn, O.J.; Salter, R.D.; Eady, R.; Yassine-Diab, B.; Favre, D.; Peretz, Y.; Landry, C.; et al. Tn-MUC1 DC Vaccination of Rhesus Macaques and a Phase I/II Trial in Patients with Nonmetastatic Castrate-Resistant Prostate Cancer. *Cancer Immunol. Res.* **2016**, *4*, 881–892, doi:10.1158/2326-6066.CIR-15-0189.
114. Castiello, L.; Sabatino, M.; Ren, J.; Terabe, M.; Khuu, H.; Wood, L. V.; Berzofsky, J.A.; Stroncek, D.F. Expression of CD14, IL10, and Tolerogenic Signature in Dendritic Cells Inversely Correlate with Clinical and Immunologic Response to TARP Vaccination in Prostate Cancer Patients. *Clin. Cancer Res.* **2017**, *23*, 3352–3364, doi:10.1158/1078-0432.CCR-16-2199.
115. Wood, L. V.; Fojo, A.; Roberson, B.D.; Hughes, M.S.B.; Dahut, W.; Gulley, J.L.; Madan, R.A.; Arlen, P.M.; Sabatino, M.; Stroncek, D.F.; et al. TARP Vaccination Is Associated with Slowing in PSA Velocity and Decreasing Tumor Growth Rates in Patients with Stage D0 Prostate Cancer. *Oncoimmunology* **2016**, *5*, 1–13, doi:10.1080/2162402X.2016.1197459.
116. Sanda, M.G.; Smith, D.C.; Charles, L.G.; Hwang, C.; Pienta, K.J.; Schlimo, J.; Milenic, D.; Panicali, D.; Montie, J.E. Recombinant Vaccinia-PSA (PROSTVAC) Can Induce a Prostate-Specific Immune Response in Androgen-Modulated Human Prostate Cancer. *Urology* **1999**, *53*, 260–266, doi:10.1016/S0090-4295(98)00539-1.
117. Lasek, W.; Zapała, Ł. Therapeutic Metastatic Prostate Cancer Vaccines: Lessons Learnt from Urologic Oncology. *Cent. Eur. J. Urol.* **2021**, *74*, 300–307, doi:10.5173/ceju.2021.0094.
118. Abdul Sater, H.; Marté, J.L.; Donahue, R.N.; Walter-Rodriguez, B.; Heery, C.R.; Steinberg, S.M.; Cordes, L.M.; Chun, G.; Karzai, F.; Bilusic, M.; et al. Neoadjuvant PROSTVAC Prior to Radical Prostatectomy Enhances T-Cell Infiltration into the Tumor Immune Microenvironment in Men with Prostate Cancer. *J. Immunother. Cancer* **2020**, *8*, 1–9, doi:10.1136/jitc-2020-000655.

119. Gulley, J.L.; Borre, M.; Vogelzang, N.J.; Ng, S.; Agarwal, N.; Parker, C.C.; Pook, D.W.; Rathenborg, P.; Flraig, T.W.; Carles, J.; et al. Phase III Trial of PROSTVAC in Asymptomatic or Minimally Symptomatic Metastatic Castration-Resistant Prostate Cancer. *J. Clin. Oncol.* **2019**, *37*, 1051–1061, doi:10.1200/JCO.18.02031.
120. Parsons, J.K.; Pinto, P.A.; Pavlovich, C.P.; Uchio, E.; Kim, H.L.; Nguyen, M.N.; Gulley, J.L.; Jamieson, C.; Hsu, P.; Wojtowicz, M.; et al. A Randomized, Double-Blind, Phase II Trial of PSA-TRICOM (PROSTVAC) in Patients with Localized Prostate Cancer: The Immunotherapy to Prevent Progression on Active Surveillance Study. *Eur. Urol. Focus* **2018**, *4*, 636–638, doi:10.1016/j.euf.2018.08.016.
121. Madan, R.A.; Karzai, F.; Donahue, R.N.; Al-Harthi, M.; Bilusic, M.; Rosner, I.I.; Singh, H.; Arlen, P.M.; Theoret, M.R.; Marté, J.L.; et al. Clinical and Immunologic Impact of Short-Course Enzalutamide Alone and with Immunotherapy in Non-Metastatic Castration Sensitive Prostate Cancer. *J. Immunother. Cancer* **2021**, *9*, doi:10.1136/jitc-2020-001556.
122. Maiorano, B.A.; Schinzari, G.; Ciardiello, D.; Rodriquenz, M.G.; Cisternino, A.; Tortora, G.; Maiello, E. Cancer Vaccines for Genitourinary Tumors: Recent Progresses and Future Possibilities. *Vaccines* **2021**, *9*, doi:10.3390/vaccines9060623.
123. Cappuccini, F.; Pollock, E.; Stribbling, S.; Hill, A.V.S.; Redchenko, I. 5T4 Oncofoetal Glycoprotein: An Old Target for a Novel Prostate Cancer Immunotherapy. *Oncotarget* **2017**, *8*, 47474–47489, doi:10.18632/oncotarget.17666.
124. Fenoglio, D.; Traverso, P.; Parodi, A.; Tomasello, L.; Negrini, S.; Kalli, F.; Battaglia, F.; Ferrera, F.; Sciallero, S.; Murdaca, G.; et al. A Multi-Peptide, Dual-Adjuvant Telomerase Vaccine (GX301) Is Highly Immunogenic in Patients with Prostate and Renal Cancer. *Cancer Immunol. Immunother.* **2013**, *62*, 1041–1052, doi:10.1007/s00262-013-1415-9.
125. Filaci, G.; Fenoglio, D.; Nolè, F.; Zanardi, E.; Tomasello, L.; Aglietta, M.; Del Conte, G.; Carles, J.; Morales-Barrera, R.; Guglielmini, P.; et al. Telomerase-Based GX301 Cancer Vaccine in Patients with Metastatic Castration-Resistant Prostate Cancer: A Randomized Phase II Trial. *Cancer Immunol. Immunother.* **2021**, *70*, 3679–3692, doi:10.1007/s00262-021-03024-0.
126. Obara, W.; Sato, F.; Takeda, K.; Kato, R.; Kato, Y.; Kanehira, M.; Takata, R.; Mimata, H.; Sugai, T.; Nakamura, Y.; et al. Phase I Clinical Trial of Cell Division Associated 1 (CDCA1) Peptide Vaccination for Castration Resistant Prostate Cancer. *Cancer Sci.* **2017**, *108*, 1452–1457, doi:10.1111/cas.13278.
127. Obara, W.; Kanehira, M.; Katagiri, T.; Kato, R.; Kato, Y.; Takata, R. Present Status and Future Perspective of Peptide-Based Vaccine Therapy for Urological Cancer. *Cancer Sci.* **2018**, *109*, 550–559, doi:10.1111/cas.13506.
128. Uemura, H.; Fujimoto, K.; Mine, T.; Uejima, S.; de Velasco, M.A.; Hirao, Y.; Komatsu, N.; Yamada, A.; Itoh, K. Immunological Evaluation of Personalized Peptide Vaccination Monotherapy in Patients with Castration-Resistant Prostate Cancer. *Cancer Sci.* **2010**, *101*, 601–608, doi:10.1111/j.1349-7006.2009.01459.x.
129. Noguchi, M.; Fujimoto, K.; Arai, G.; Uemura, H.; Hashine, K.; Matsumoto, H.; Fukasawa, S.; Kohjimoto, Y.; Nakatsu, H.; Takenaka, A.; et al. A Randomized Phase III Trial of Personalized Peptide Vaccination for Castration-Resistant Prostate Cancer Progressing after Docetaxel. *Oncol. Rep.* **2021**, *45*, 159–168, doi:10.3892/or.2020.7847.
130. Schuhmacher, J.; Heidu, S.; Balchen, T.; Richardson, J.R.; Schmeltz, C.; Sonne, J.; Schweiker, J.; Rammensee, H.G.; Thor Straten, P.; Røder, M.A.; et al. Vaccination against RhoC Induces Long-Lasting Immune Responses in Patients with Prostate Cancer: Results from a Phase I/II Clinical Trial. *J. Immunother. Cancer* **2020**, *8*, 1–12, doi:10.1136/jitc-2020-001157.
131. Santegoets, S.J.A.M.; Stam, A.G.M.; Lougheed, S.M.; Gall, H.; Jooss, K.; Sacks, N.; Hege, K.; Lowy, I.; Scheper, R.J.; Gerritsen, W.R.; et al. Myeloid Derived Suppressor and Dendritic Cell Subsets Are Related to Clinical Outcome in Prostate Cancer Patients Treated with Prostate Gvax and Ipilimumab. *J. Immunother. Cancer* **2014**, *2*, 1, doi:10.1186/s40425-014-0031-3.
132. Van den Eertwegh, A.J.M.; Versluis, J.; Van den Berg, H.P.; Santegoets, S.J.A.M.; Van Moorselaar, R.J.A.; Van der Sluis, T.M.; Gall, H.E.; Harding, T.C.; Jooss, K.; Lowy, I.; et al. Combined Immunotherapy with Granulocyte-Macrophage Colony-Stimulating Factor-Transduced Allogeneic Prostate Cancer Cells and Ipilimumab in Patients with Metastatic Castration-Resistant Prostate Cancer: A Phase 1 Dose-Escalation Trial. *Lancet Oncol.* **2012**, *13*, 509–517, doi:10.1016/S1470-2045(12)70007-4.
133. Karan, D.; Van Veldhuizen, P. Combination Immunotherapy with Prostate GVAX and Ipilimumab: Safety and Toxicity. *Immunotherapy* **2012**, *4*, 577–580, doi:10.2217/imt.12.53.
134. Mottet, N.; Bastian, P.; Bellmunt, J.; van den Bergh, R.; Bolla, M.; van Casteren, N.; Cornford, P.; Joniau, S.; Matveev, V.; van der Kwast, T.; et al. EAU - EANM - ESTRO - SIOG: Guidelines on Prostate Cancer. *Eur. Assoc. Urol.* **2020**, 1–182.
135. Ramsay, C.R.; Adewuyi, T.E.; Gray, J.; Hislop, J.; Shirley, M.D.; Jayakody, S.; MacLennan, G.; Fraser, C.; MacLennan, S.; Brazzelli, M.; et al. Ablative Therapy for People with Localised Prostate Cancer: A Systematic Review and Economic Evaluation. *Health Technol. Assess. (Rockv.)* **2015**, *19*, 1–8, doi:10.3310/hta19490.
136. Schmid, F.A.; Schindele, D.; Mortezaei, A.; Spitznagel, T.; Sulser, T.; Schostak, M.; Eberli, D. Prospective Multicentre Study Using High Intensity Focused Ultrasound (HIFU) for the Focal Treatment of Prostate Cancer: Safety Outcomes and Complications. *Urol. Oncol. Semin. Orig. Investig.* **2020**, *38*, 225–230, doi:10.1016/j.urolonc.2019.09.001.
137. Joiner, J.B.; Pylayeva-Gupta, Y.; Dayton, P.A. Focused Ultrasound for Immunomodulation of the Tumor Microenvironment. *J. Immunol.* **2020**, *205*, 2327–2341, doi:10.4049/jimmunol.1901430.
138. Finley, D.S.; Pouliot, F.; Shuch, B.; Chin, A.; Pantuck, A.; Dekernion, J.B.; Belldegrun, A.S. Ultrasound-Based Combination Therapy: Potential in Urologic Cancer. *Expert Rev. Anticancer Ther.* **2011**, *11*, 107–113, doi:10.1586/era.10.174.
139. ter Haar, G.; Coussios, C. High Intensity Focused Ultrasound: Physical Principles and Devices. *Int. J. Hyperth.* **2007**, *23*, 89–104, doi:10.1080/02656730601186138.
140. Hu, Z.; Yang, X.Y.; Liu, Y.; Sankin, G.N.; Pua, E.C.; Morse, M.A.; Lyerly, H.K.; Clay, T.M.; Zhong, P. Investigation of HIFU-Induced Anti-Tumor Immunity in a Murine Tumor Model. *J. Transl. Med.* **2007**, *5*, 1–11, doi:10.1186/1479-5876-5-34.

141. Ran, L.F.; Xie, X.P.; Xia, J.Z.; Xie, F.L.; Fan, Y.M.; Wu, F. Specific Antitumour Immunity of HIFU-Activated Cytotoxic T Lymphocytes after Adoptive Transfusion in Tumour-Bearing Mice. *Int. J. Hyperth.* **2016**, *32*, 204–210, doi:10.3109/02656736.2015.1112438.
142. Rosberger, D.F.; Coleman, D.J.; Silverman, R.; Woods, S.; Rondeau, M.; Cunningham-Rundles, S. Immunomodulation in Choroidal Melanoma: Reversal of Inverted CD4/CD8 Ratios Following Treatment with Ultrasonic Hyperthermia. *Biotechnol. Ther.* **1994**, *5*, 59–68.
143. Zhang, Y.; Deng, J.; Feng, J.; Wu, F. Enhancement of Antitumor Vaccine in Ablated Hepatocellular Carcinoma by High-Intensity Focused Ultrasound. *World J. Gastroenterol.* **2010**, *16*, 3584–3591, doi:10.3748/wjg.v16.i28.3584.
144. Xing, Y.; Lu, X.; Pua, E.C.; Zhong, P. The Effect of High Intensity Focused Ultrasound Treatment on Metastases in a Murine Melanoma Model. *Biochem. Biophys. Res. Commun.* **2008**, *375*, 645–650, doi:10.1016/j.bbrc.2008.08.072.
145. Singh, M.P.; Sethuraman, S.N.; Ritchey, J.; Fiering, S.; Guha, C.; Malayer, J.; Ranjan, A. In-Situ Vaccination Using Focused Ultrasound Heating and Anti-CD-40 Agonistic Antibody Enhances T-Cell Mediated Local and Abscopal Effects in Murine Melanoma. *Int. J. Hyperth.* **2019**, *36*, 64–73, doi:10.1080/02656736.2019.1663280.
146. Yang, R.; Reilly, C.R.; Rescorla, F.J.; Sanghvi, N.T.; Fry, F.J.; Franklin, T.D.; Grosfeld, J.L. Effects of High-Intensity Focused Ultrasound in the Treatment of Experimental Neuroblastoma. *J. Pediatr. Surg.* **1992**, *27*, 246–251, doi:10.1016/0022-3468(92)90321-W.
147. Erranki, A.; Srinivasan, P.; Ries, M.; Kim, A.R.; Lazarski, C.A.; Rossi, C.T.; Khokhlova, T.D.; Wilson, E.; Knoblauch, S.M.; Sharma, K. V.; et al. High-Intensity Focused Ultrasound (HIFU) Triggers Immune Sensitization of Refractory Murine Neuroblastoma to Checkpoint Inhibitor Therapy. *Clin. Cancer Res.* **2020**, *26*, 1152–1161, doi:10.1158/1078-0432.CCR-19-1604.
148. Wang, X.; Sun, J. High-Intensity Focused Ultrasound in Patients with Late-Stage Pancreatic Carcinoma. *Chin. Med. J. (Engl.)* **2002**, *115*, 1332–1335.
149. Mouratidis, P.X.E.; Costa, M.; Rivens, I.; Repasky, E.E.; Ter Haar, G. Pulsed Focused Ultrasound Can Improve the Anti-Cancer Effects of Immune Checkpoint Inhibitors in Murine Pancreatic Cancer. *J. R. Soc. Interface* **2021**, *18*, doi:10.1098/rsif.2021.0266.
150. Wu, F.; Wang, Z.B.; Cao, Y. De; Zhou, Q.; Zhang, Y.; Xu, Z.L.; Zhu, X.Q. Expression of Tumor Antigens and Heat-Shock Protein 70 in Breast Cancer Cells after High-Intensity Focused Ultrasound Ablation. *Ann. Surg. Oncol.* **2007**, *14*, 1237–1242, doi:10.1245/s10434-006-9275-6.
151. Ji, C.; Si, J.; Xu, Y.; Zhang, W.; Yang, Y.; He, X.; Xu, H.; Mou, X.; Ren, H.; Guo, H. Mitochondria-Targeted and Ultrasound-Responsive Nanoparticles for Oxygen and Nitric Oxide Codelivery to Reverse Immunosuppression and Enhance Sonodynamic Therapy for Immune Activation. *Theranostics* **2021**, *11*, 8587–8604, doi:10.7150/THNO.62572.
152. Zhang, D.; Lin, Z.; Zheng, Y.; Song, J.; Li, J.; Zeng, Y.; Liu, X. Ultrasound-Driven Biomimetic Nanosystem Suppresses Tumor Growth and Metastasis through Sonodynamic Therapy, CO Therapy, and Indoleamine 2,3-Dioxygenase Inhibition. *ACS Nano* **2020**, *14*, 8985–8999, doi:10.1021/acsnano.0c03833.
153. Yakkala, C.; Denys, A.; Kandalaft, L.; Duran, R. Cryoablation and Immunotherapy of Cancer. *Curr. Opin. Biotechnol.* **2020**, *65*, 60–64, doi:10.1016/j.copbio.2020.01.006.
154. Aarts, B.M.; Klompenhouwer, E.G.; Rice, S.L.; Imani, F.; Baetens, T.; Bex, A.; Horenblas, S.; Kok, M.; Haanen, J.B.A.G.; Beets-Tan, R.G.H.; et al. Cryoablation and Immunotherapy: An Overview of Evidence on Its Synergy. *Insights Imaging* **2019**, *10*, doi:10.1186/s13244-019-0727-5.
155. Ablin, R.J.; Soanes, W.A.; Gonder, M.J. Immunologic Studies of the Prostate. A Review. *Int. Surg.* **1969**, *52*, 8–21.
156. Abdo, J.; Cornell, D.L.; Mittal, S.K.; Agrawal, D.K. Immunotherapy plus Cryotherapy: Potential Augmented Abscopal Effect for Advanced Cancers. *Front. Oncol.* **2018**, *8*, 1–16, doi:10.3389/fonc.2018.00085.
157. Stamell, E.F.; Wolchok, J.D.; Gnjatic, S.; Lee, N.Y.; Brownell, I. The Abscopal Effect Associated with a Systemic Anti-Melanoma Immune Response. *Int. J. Radiat. Oncol. Biol. Phys.* **2013**, *85*, 293–295, doi:10.1016/j.ijrobp.2012.03.017.
158. Gaitanis, G.; Bassukas, I.D. Immunocryosurgery for Non-Superficial Basal Cell Carcinomas ≤ 20 mm in Maximal Diameter: Five-Year Follow-Up. *J. Geriatr. Oncol.* **2019**, *10*, 475–478, doi:10.1016/j.jgo.2018.08.012.
159. Lin, M.; Xu, K.; Liang, S.; Wang, X.; Liang, Y.; Zhang, M.; Chen, J.; Niu, L.Z. Prospective Study of Percutaneous Cryoablation Combined with Allogenic NK Cell Immunotherapy for Advanced Renal Cell Cancer. *Immunol. Lett.* **2017**, *184*, 98–104, doi:10.1016/j.imlet.2017.03.004.
160. Lin, M.; Liang, S.; Wang, X.; Liang, Y.; Zhang, M.; Chen, J.; Niu, L.; Xu, K. Cryoablation Combined with Allogenic Natural Killer Cell Immunotherapy Improves the Curative Effect in Patients with Advanced Hepatocellular Cancer. *Oncotarget* **2017**, *8*, 81967–81977, doi:10.18632/oncotarget.17804.
161. Lin, M.; Liang, S.Z.; Wang, X.H.; Liang, Y.Q.; Zhang, M.J.; Niu, L.Z.; Chen, J.B.; Li, H.B.; Xu, K.C. Clinical Efficacy of Percutaneous Cryoablation Combined with Allogenic NK Cell Immunotherapy for Advanced Non-Small Cell Lung Cancer. *Immunol. Res.* **2017**, *65*, 880–887, doi:10.1007/s12026-017-8927-x.
162. Si, T.; Guo, Z.; Hao, X. Combined Cryoablation and GM-CSF Treatment for Metastatic Hormone Refractory Prostate Cancer. *J. Immunother.* **2009**, *32*, 86–91, doi:10.1097/CJI.0b013e31818df785.
163. Barqawi, A.B.; Rodrigues Pessoa, R.; Crawford, E.D.; Al-Musawi, M.; MacDermott, T.; O'Donell, C.; Kendl, R.M. Boosting Immune Response with GM-CSF Optimizes Primary Cryotherapy Outcomes in the Treatment of Prostate Cancer: A Prospective Randomized Clinical Trial. *Prostate Cancer Prostatic Dis.* **2021**, *24*, 750–757, doi:10.1038/s41391-021-00321-8.
164. Ross, A.E.; Hurley, P.J.; Tran, P.T.; Rowe, S.P.; Benzon, B.; Neal, T.O.; Chapman, C.; Harb, R.; Milman, Y.; Trock, B.J.; et al. A Pilot Trial of Pembrolizumab plus Prostatic Cryotherapy for Men with Newly Diagnosed Oligometastatic Hormone-Sensitive Prostate Cancer. *Prostate Cancer Prostatic Dis.* **2020**, *23*, 184–193, doi:10.1038/s41391-019-0176-8.

165. Wang, G.; Zhao, D.; Spring, D.J.; Depinho, R.A. Genetics and Biology of Prostate Cancer. *Genes Dev.* **2018**, *32*, 1105–1140, doi:10.1101/gad.315739.118.
166. Wang, M.; Rao, J.; Wang, M.; Li, X.; Liu, K.; Naylor, M.F.; Nordquist, R.E.; Chen, W.R.; Zhou, F. Cancer Photo-Immunotherapy: From Bench to Bedside. *Theranostics* **2021**, *11*, 2218–2231, doi:10.7150/thno.53056.
167. Algorri, J.F.; Ochoa, M.; Roldán-Varona, P.; Rodríguez-Cobo, L.; López-Higuera, J.M. Photodynamic Therapy: A Compendium of Latest Reviews. *Cancers (Basel.)* **2021**, *13*, doi:10.3390/cancers13174447.
168. Rastinehad, A.R.; Anastos, H.; Wajswol, E.; Winoker, J.S.; Sfakianos, J.P.; Doppalapudi, S.K.; Carrick, M.R.; Knauer, C.J.; Taouli, B.; Lewis, S.C.; et al. Gold Nanoshell-Localized Photothermal Ablation of Prostate Tumors in a Clinical Pilot Device Study. *Proc. Natl. Acad. Sci. U. S. A.* **2019**, *116*, 18590–18596, doi:10.1073/pnas.1906929116.
169. Azzouzi, A.R.; Vincendeau, S.; Barret, E.; Cicco, A.; Kleinlauss, F.; van der Poel, H.G.; Stief, C.G.; Rassweiler, J.; Salomon, G.; Solsona, E.; et al. Padeliporfin Vascular-Targeted Photodynamic Therapy versus Active Surveillance in Men with Low-Risk Prostate Cancer (CLIN1001 PCM301): An Open-Label, Phase 3, Randomised Controlled Trial. *Lancet Oncol.* **2017**, *18*, 181–191, doi:10.1016/S1470-2045(16)30661-1.
170. Osuchowski, M.; Bartusik-Aebisher, D.; Osuchowski, F.; Aebisher, D. Photodynamic Therapy for Prostate Cancer – A Narrative Review. *Photodiagnosis Photodyn. Ther.* **2021**, *33*, doi:10.1016/j.pdpt.2020.102158.
171. Hwang, H.S.; Shin, H.; Han, J.; Na, K. Combination of Photodynamic Therapy (PDT) and Anti-Tumor Immunity in Cancer Therapy. *J. Pharm. Investig.* **2018**, *48*, 143–151, doi:10.1007/s40005-017-0377-x.
172. Zou, J.; Li, L.; Yang, Z.; Chen, X. Phototherapy Meets Immunotherapy: A Win-Win Strategy to Fight against Cancer. *Nanophotonics* **2021**, *10*, 3229–3245, doi:10.1515/nanoph-2021-0209.
173. Castano, A.P.; Mroz, P.; Hamblin, M.R. Photodynamic Therapy and Anti-Tumour Immunity. *Nat. Rev. Cancer* **2006**, *6*, 535–545, doi:10.1038/nrc1894.
174. Kabingu, E.; Vaughan, L.; Owczarczak, B.; Ramsey, K.D.; Gollnick, S.O. CD8+ T Cell-Mediated Control of Distant Tumours Following Local Photodynamic Therapy Is Independent of CD4+ T Cells and Dependent on Natural Killer Cells. *Br. J. Cancer* **2007**, *96*, 1839–1848, doi:10.1038/sj.bjc.6603792.
175. Mroz, P.; Hashmi, J.T.; Huang, Y.Y.; Lange, N.; Hamblin, M.R. Stimulation of Anti-Tumor Immunity by Photodynamic Therapy. *Expert Rev. Clin. Immunol.* **2011**, *7*, 75–91, doi:10.1586/eci.10.81.
176. Li, Y.; Li, X.; Doughty, A.; West, C.; Wang, L.; Zhou, F.; Nordquist, R.E.; Chen, W.R. Phototherapy Using Immunologically Modified Carbon Nanotubes to Potentiate Checkpoint Blockade for Metastatic Breast Cancer. *Nanomedicine Nanotechnology, Biol. Med.* **2019**, *18*, 44–53, doi:10.1016/j.nano.2019.02.009.
177. Huang, Z.; Wei, G.; Zeng, Z.; Huang, Y.; Huang, L.; Shen, Y.; Sun, X.; Xu, C.; Zhao, C. Enhanced Cancer Therapy through Synergetic Photodynamic/Immune Checkpoint Blockade Mediated by a Liposomal Conjugate Comprised of Porphyrin and IDO Inhibitor. *Theranostics* **2019**, *9*, 5542–5557, doi:10.7150/thno.35343.
178. Kim, S.; Kim, S.A.; Nam, G.H.; Hong, Y.; Kim, G.B.; Choi, Y.; Lee, S.; Cho, Y.; Kwon, M.; Jeong, C.; et al. In Situ Immunogenic Clearance Induced by a Combination of Photodynamic Therapy and Rho-Kinase Inhibition Sensitizes Immune Checkpoint Blockade Response to Elicit Systemic Antitumor Immunity against Intraocular Melanoma and Its Metastasis. *J. Immunother. Cancer* **2021**, *9*, 1–15, doi:10.1136/jitc-2020-001481.
179. Nagaya, T.; Nakamura, Y.; Okuyama, S.; Ogata, F.; Maruoka, Y.; Choyke, P.L.; Kobayashi, H. Near-Infrared Photoimmunotherapy Targeting Prostate Cancer with Prostate-Specific Membrane Antigen (PSMA) Antibody. *Mol. Cancer Res.* **2017**, *15*, 1153–1162, doi:10.1158/1541-7786.MCR-17-0164.
180. Watanabe, R.; Hanaoka, H.; Sato, K.; Nagaya, T.; Harada, T.; Mitsunaga, M.; Kim, I.; Paik, C.H.; Wu, A.M.; Choyke, P.L.; et al. Photoimmunotherapy Targeting Prostate-Specific Membrane Antigen: Are Antibody Fragments as Effective as Antibodies? *J. Nucl. Med.* **2015**, *56*, 140–144, doi:10.2967/jnumed.114.149526.
181. Miller, L.; Leor, J.; Rubinsky, B. Cancer Cells Ablation with Irreversible Electroporation. *Technol. Cancer Res. Treat.* **2005**, *4*, 699–705, doi:10.1177/153303460500400615.
182. Ting, F.; Tran, M.; Böhm, M.; Siriwardana, A.; Van Leeuwen, P.J.; Haynes, A.M.; Delprado, W.; Shnier, R.; Stricker, P.D. Focal Irreversible Electroporation for Prostate Cancer: Functional Outcomes and Short-Term Oncological Control. *Prostate Cancer Prostatic Dis.* **2016**, *19*, 46–52, doi:10.1038/pcan.2015.47.
183. Kielbik, A.; Szlasi, W.; Saczko, J.; Kulbacka, J. Electroporation-Based Treatments in Urology. *Cancers (Basel.)* **2020**, *12*, 1–26, doi:10.3390/cancers12082208.
184. Yang, J.; Eresen, A.; Shangguan, J.; Ma, Q.; Yaghmai, V.; Zhang, Z. Irreversible Electroporation Ablation Overcomes Tumor-Associated Immunosuppression to Improve the Efficacy of DC Vaccination in a Mice Model of Pancreatic Cancer. *Oncoimmunology* **2021**, *10*, 1–9, doi:10.1080/2162402X.2021.1875638.
185. Zhao, J.; Wen, X.; Tian, L.; Li, T.; Xu, C.; Wen, X.; Melancon, M.P.; Gupta, S.; Shen, B.; Peng, W.; et al. Irreversible Electroporation Reverses Resistance to Immune Checkpoint Blockade in Pancreatic Cancer. *Nat. Commun.* **2019**, *10*, 1–14, doi:10.1038/s41467-019-08782-1.
186. He, C.; Sun, S.; Zhang, Y.; Li, S. Irreversible Electroporation plus Anti-Pd-1 Antibody versus Irreversible Electroporation Alone for Patients with Locally Advanced Pancreatic Cancer. *J. Inflamm. Res.* **2021**, *14*, 4795–4807, doi:10.2147/JIR.S331023.
187. Burbach, B.J.; O'Flanagan, S.D.; Shao, Q.; Young, K.M.; Slaughter, J.R.; Rollins, M.R.; Street, T.J.L.; Granger, V.E.; Beura, L.K.; Azarin, S.M.; et al. Irreversible Electroporation Augments Checkpoint Immunotherapy in Prostate Cancer and Promotes Tumor Antigen-Specific Tissue-Resident Memory CD8+ T Cells. *Nat. Commun.* **2021**, *12*, 1–16, doi:10.1038/s41467-021-24132-6.

188. Eriksson, D.; Stigbrand, T. Radiation-Induced Cell Death Mechanisms. *Tumor Biol.* **2010**, *31*, 363–372, doi:10.1007/s13277-010-0042-8.
189. Finkelstein, S.E.; Salenius, S.; Mantz, C.A.; Shore, N.D.; Fernandez, E.B.; Shulman, J.; Myslicki, F.A.; Agassi, A.M.; Rotterman, Y.; Devries, T.; et al. Combining Immunotherapy and Radiation for Prostate Cancer. *Clin. Genitourin. Cancer* **2015**, *13*, 1–9, doi:10.1016/j.clgc.2014.09.001.
190. Nesslinger, N.J.; Sahota, R.A.; Stone, B.; Johnson, K.; Chima, N.; King, C.; Rasmussen, D.; Bishop, D.; Rennie, P.S.; Gleave, M.; et al. Standard Treatments Induce Antigen-Specific Immune Responses in Prostate Cancer. *Clin. Cancer Res.* **2007**, *13*, 1493–1502, doi:10.1158/1078-0432.CCR-06-1772.
191. Lin, L.; Kane, N.; Kobayashi, N.; Kono, E.A.; Yamashiro, J.M.; Nickols, N.G.; Reiter, R.E. High-Dose per Fraction Radiotherapy Induces Both Antitumor Immunity and Immunosuppressive Responses in Prostate Tumors. *Clin. Cancer Res.* **2021**, *27*, 1505–1515, doi:10.1158/1078-0432.CCR-20-2293.
192. Donlon, N.E.; Power, R.; Hayes, C.; Reynolds, J. V.; Lysaght, J. Radiotherapy, Immunotherapy, and the Tumour Microenvironment: Turning an Immunosuppressive Milieu into a Therapeutic Opportunity. *Cancer Lett.* **2021**, *502*, 84–96, doi:10.1016/j.canlet.2020.12.045.
193. Solanki, A.A.; Bossi, A.; Efsthathiou, J.A.; Lock, D.; Mondini, M.; Ramapriyan, R.; Welsh, J.; Kang, J. Combining Immunotherapy with Radiotherapy for the Treatment of Genitourinary Malignancies. *Eur. Urol. Oncol.* **2019**, *2*, 79–87, doi:10.1016/j.euo.2018.09.013.
194. Demaria, S.; Bhardwaj, N.; McBride, W.H.; Formenti, S.C. Combining Radiotherapy and Immunotherapy: A Revived Partnership. *Int. J. Radiat. Oncol. Biol. Phys.* **2005**, *63*, 655–666, doi:10.1016/j.ijrobp.2005.06.032.
195. Eckert, F.; Schaedle, P.; Zips, D.; Schmid-Horch, B.; Rammensee, H.G.; Gani, C.; Gouttefangeas, C. Impact of Curative Radiotherapy on the Immune Status of Patients with Localized Prostate Cancer. *Oncoimmunology* **2018**, *7*, 1–11, doi:10.1080/2162402X.2018.1496881.
196. Nickols, N.G.; Ganapathy, E.; Nguyen, C.; Kane, N.; Lin, L.; Diaz-Perez, S.; Nazarian, R.; Mathis, C.; Felix, C.; Basehart, V.; et al. The Intraprostatic Immune Environment after Stereotactic Body Radiotherapy Is Dominated by Myeloid Cells. *Prostate Cancer Prostatic Dis.* **2021**, *24*, 135–139, doi:10.1038/s41391-020-0249-8.
197. Dudzinski, S.O.; Cameron, B.D.; Wang, J.; Rathmell, J.C.; Giorgio, T.D.; Kirschner, A.N. Combination Immunotherapy and Radiotherapy Causes an Abscopal Treatment Response in a Mouse Model of Castration Resistant Prostate Cancer. *J. Immunother. Cancer* **2019**, *7*, 1–8, doi:10.1186/s40425-019-0704-z.
198. Wada, S.; Harris, T.J.; Tryggestad, E.; Yoshimura, K.; Zeng, J.; Yen, H.R.; Getnet, D.; Gross, J.F.; Bruno, T.C.; De Marzo, A.M.; et al. Combined Treatment Effects of Radiation and Immunotherapy: Studies in an Autochthonous Prostate Cancer Model. *Int. J. Radiat. Oncol. Biol. Phys.* **2013**, *87*, 769–776, doi:10.1016/j.ijrobp.2013.07.015.
199. Philippou, Y.; Sjoberg, H.T.; Murphy, E.; Alyacoubi, S.; Jones, K.I.; Gordon-Weeks, A.N.; Phyu, S.; Parkes, E.E.; Gillies McKenna, W.; Lamb, A.D.; et al. Impacts of Combining Anti-PD-L1 Immunotherapy and Radiotherapy on the Tumour Immune Microenvironment in a Murine Prostate Cancer Model. *Br. J. Cancer* **2020**, *123*, 1089–1100, doi:10.1038/s41416-020-0956-x.
200. Slovin, S.F.; Higano, C.S.; Hamid, O.; Tejwani, S.; Harzstark, A.; Alumkal, J.J.; Scher, H.I.; Chin, K.; Gagnier, P.; McHenry, M.B.; et al. Ipilimumab Alone or in Combination with Radiotherapy in Metastatic Castration-Resistant Prostate Cancer: Results from an Open-Label, Multicenter Phase I/I Study. *Ann. Oncol.* **2013**, *24*, 1813–1821, doi:10.1093/annonc/mdt107.
201. Kwon, E.D.; Drake, C.G.; Scher, H.I.; Fizazi, K.; Bossi, A.; Van den Eertwegh, A.J.M.; Krainer, M.; Houede, N.; Santos, R.; Mahammedi, H.; et al. Ipilimumab versus Placebo after Radiotherapy in Patients with Metastatic Castration-Resistant Prostate Cancer That Had Progressed after Docetaxel Chemotherapy (CA184-043): A Multicentre, Randomised, Double-Blind, Phase 3 Trial. *Lancet Oncol.* **2014**, *15*, 700–712, doi:10.1016/S1470-2045(14)70189-5.
202. Twardowski, P.; Wong, J.Y.C.; Pal, S.K.; Maughan, B.L.; Frankel, P.H.; Franklin, K.; Junqueira, M.; Prajapati, M.R.; Nachaegari, G.; Harwood, D.; et al. Randomized Phase II Trial of Sipuleucel-T Immunotherapy Preceded by Sensitizing Radiation Therapy and Sipuleucel-T Alone in Patients with Metastatic Castrate Resistant Prostate Cancer. *Cancer Treat. Res. Commun.* **2019**, *19*, 100116, doi:10.1016/j.ctarc.2018.100116.
203. Yuan, Z.; Fernandez, D.; Dhillon, J.; Abraham-Miranda, J.; Awasthi, S.; Kim, Y.; Zhang, J.; Jain, R.; Serna, A.; Pow-Sang, J.M.; et al. Proof-of-Principle Phase I Results of Combining Nivolumab with Brachytherapy and External Beam Radiation Therapy for Grade Group 5 Prostate Cancer: Safety, Feasibility, and Exploratory Analysis. *Prostate Cancer Prostatic Dis.* **2021**, *24*, 140–149, doi:10.1038/s41391-020-0254-y.
204. Marshall, C.H.; Fu, W.; Wang, H.; Park, J.C.; DeWeese, T.L.; Tran, P.T.; Song, D.Y.; King, S.; Afful, M.; Hurrelbrink, J.; et al. Randomized Phase II Trial of Sipuleucel-T with or without Radium-223 in Men with Bone-Metastatic Castration-Resistant Prostate Cancer. *Clin. Cancer Res.* **2021**, *27*, 1623–1630, doi:10.1158/1078-0432.CCR-20-4476.
205. Han, H.J.; Li, Y.R.; Roach, M.; Aggarwal, R. Dramatic Response to Combination Pembrolizumab and Radiation in Metastatic Castration Resistant Prostate Cancer. *Ther. Adv. Med. Oncol.* **2020**, *12*, 1–8, doi:10.1177/1758835920936084.