

Figure S1: Generation of Calu6 ACE2 cells and SARS CoV-2 pseudotyping. (A) The human lung-derived epithelial cell line Calu6 was transduced with lentiviral particles encoding for human ACE2 (hACE2) and a puromycin resistance cassette. After sufficient selection with Puromycin, a stable sub-cell line was obtained, henceforward termed Calu6 ACE2. (B) Phase contrast (left) and Immunofluorescence staining of Calu6 and Calu6 ACE2 cells with fluorescently tagged ACE2 antibodies (right). Micrographs were acquired using a Keyence BZ-X700 fluorescence microscope, using a 10x air (10x/0.30NA Ph1 DL) objective. Scale bar 100 μ m. (C) Testing of VSV* SARS CoV-2 virus particles. Both cell lines were infected for 24 h with increasing concentrations of VSV* SARS mock (no pseudotype), VSV* SARS CoV-2 and VSV* VSVg, respectively. Bar charts show frequencies of GFP positive, live cells assessed by flow cytometry.

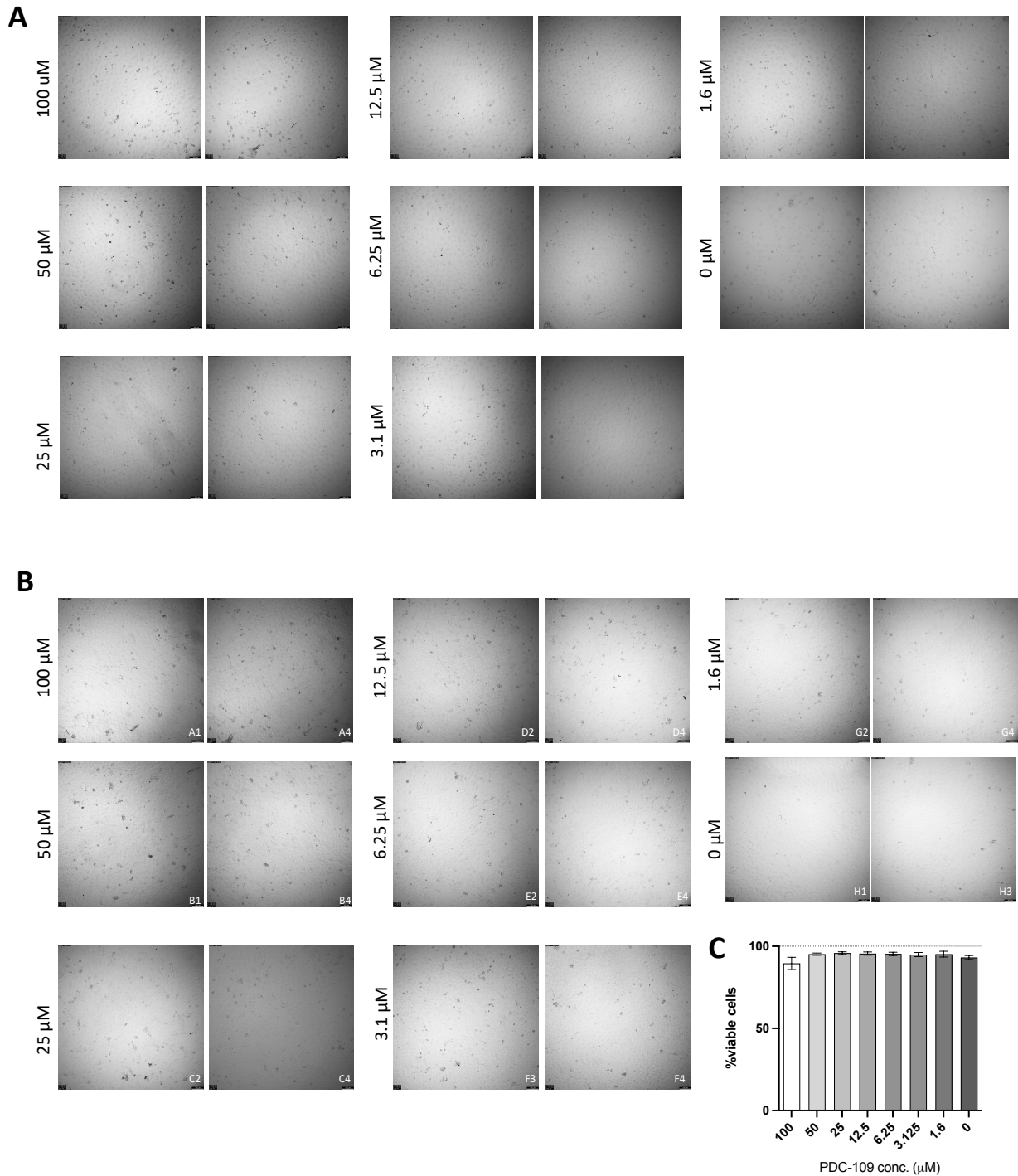


Figure S2: Cytotoxicity of PDC-109 in VeroE6 cells. Cells were plated in flat-bottom 96 well plates and at the next day treated with increasing concentrations PDC-109 for 2 hours. Subsequently, supernatants were replaced with fresh media and cells were cultured for another **(A)** 24 or **(B)** 72 hours. Then, cells were imaged using a Thunder Imaging System (Leica, Wetzlar, Germany) in order to assess cell morphology and overall confluence of the monolayer. **(C)** Cells treated as described in **(B)** were subjected to cell viability staining using Zombie-Aqua (BioLegend, San Diego, USA), followed by flow cytometry at a LSRII instrument. Cells were pre-gated for singlets. Bars show mean with SEM of three repeats.