

## Article

# Evaluation of Tumor Grade and Proliferation Indices before and after Short-Course Anti-Inflammatory Prednisone Therapy in Canine Cutaneous Mast Cell Tumors

Shawna Klahn<sup>1\*</sup>, Nikolaos Dervisis<sup>1</sup>, Kevin Lahmers<sup>2</sup> and Marian Benitez<sup>1</sup>

<sup>1</sup> Department of Small Animal Clinical Sciences, Virginia Maryland College of Veterinary Medicine, Virginia Tech, 245 Duck Pond Drive, Blacksburg, VA 24061; dervisis@vt.edu (N.D.); marian.benitez19@gmail.com (M.B.)

<sup>2</sup> Department of Biomedical Sciences and Pathobiology, Virginia Maryland College of Veterinary Medicine, Virginia Tech, 245 Duck Pond Drive, Blacksburg, VA 24061; klahmers@vt.edu (K.L.) ORCID 0000-0002-5290-3426

\* Correspondence: klahn@vt.edu

**Abstract:** Glucocorticoid administration is a common clinical practice in attempt to decrease inflammation associated with, and improve resectability of, canine mast cell tumors (MCTs). However, the impact of neoadjuvant glucocorticoids on histological features and proliferation indices of canine MCTs is unknown. The objective of this study was to evaluate changes in tumor grade, mitotic count, Ki67, AgNOR, and AgNORxKi67 scores following short-course, anti-inflammatory neoadjuvant prednisone in canine patients with MCTs. This was a prospective, single-arm pilot study. Client-owned dogs with treatment-naïve, cytologically-confirmed MCTs were enrolled. Patients underwent an initial incisional biopsy followed by a 10-14 day course of anti-inflammatory prednisone, and surgical resection. All histological samples were randomized, masked, and evaluated by a single pathologist. Unstained paired pre- and post-treatment samples were submitted to a commercial laboratory for Ki67 and AgNOR immunohistochemical analysis. There were 11 dogs enrolled with 11 tumors. There were no statistical differences between the pre- and post-treatment histological parameters of mitotic index, Ki67, AgNOR, or Ki67xAgNOR. There were no clinically-significant alterations between pre-treatment and post-treatment in the assignment of tumor grades. A short-course of anti-inflammatory prednisone does not appear to alter the histological parameters that affect grade-determination, nor significantly alter the proliferation indices in canine MCTs.

**Keywords:** mast cell tumor; dog; canine; proliferation indices; grade; prednisone; Ki67; AgNOR; mitotic count; mitotic index

## 1. Introduction

Mast cell tumors (MCTs) are the most common skin tumor affecting dogs, and tumor grade is the most consistent and clinically-relevant prognostic factor[1-4]. There are two MCT grading systems: the Patnaik system and the Kiupel system[3]. The Patnaik system categorizes tumors as grades 1, 2, or 3, with grade 1 as potentially curable, and grade 3 conferring a poor prognosis[5]. The majority of MCTs, however, are classified as Patnaik grade 2, which demonstrate a wide range of biological behavior[3]. The Kiupel system was proposed to reclassify grade 2 tumors to improve clinical utility, and categorizes all tumors as either low- or high-grade, with low-grade tumors having an excellent prognosis, and high-grade tumors conferring a poor prognosis[6]. The recent consensus document by the Oncology-Pathology Working Group states that the two grading systems are complementary, and recommends that MCT grade is reported using both systems: G1/LG, G2/LG, G2/HG, or G3/HG[1].

Rapid cellular proliferation carries prognostic significance in malignancy and can be quantified by evaluating the mitotic count, defined as the number of mitotic figures in 10 non-overlapping high-powered fields (2.37 mm<sup>2</sup>)[7,8]. Mitotic count is a criterion in MCT grading, but the two grading systems having different cut-offs for grade determination, and interobserver variability and lack of standardization of mitotic count in canine MCTs have been reported [1,7-9]. Tumor proliferation involves not only cells in mitosis, but the number of cells actively engaged in the cell cycle and how quickly cells are progressing through the cell cycle. To that end, mitotic count does not provide a global view of MCT proliferation, and additional markers evaluating the growth fraction and generation time can be employed to improve prognostication[10]. Ki67 is a nuclear protein that is expressed in all phases of the cell cycle except G0 and represents the tumor growth fraction. The relative number of cells expressing Ki67 is an established prognostic factor in canine MCTs and other malignancies in several species[2,11]. There are multiple variants of Ki67, with species, cell-type, and cell-cycle specificity, and it forms the perichromosomal layer during mitosis, preventing chromosomes from sticking together and maintaining chromosomal structural integrity. Following mitosis, Ki67 also functions in aiding in nucleolar organization[11]. Nucleoli are the sites of ribosome biogenesis and form around organizer regions (NORs) which contain tandem arrays of ribosomal gene repeats. The nucleolus is the largest non-membrane-bound subnuclear structure and can be easily visualized in the interphase nucleus[12,13]. A silver-based staining method is used to identify and quantify NORs, termed argyrophilic nucleolar organizing regions (AgNOR), and represent how quickly the cells progress through the cell cycle. Together, AgNOR and Ki67 further refine the anticipated biological behavior, and can provide the clinician with information to guide treatment decisions for patients with tumors with the intermediate tumor grades of G2/LG and G2/HG[2].

Glucocorticoids are foundational in the treatment of canine (MCTs). They are administered orally or intra-lesionally, as sole therapy or in combination with conventional chemotherapy, small molecule targeted therapy, radiation therapy, and/or surgery[14-38]. The majority of responses to glucocorticoids as sole therapy are partial responses, with a response rate of approximately 70%[14,19,22,34,39-41]. Definitive local and local-regional therapy improve outcome and quality of life, regardless of MCT location or grade[1,17,19,22,36,38,42-56]. It is a common clinical practice to attempt cytoreduction of MCTs with glucocorticoid therapy and to reduce morbidity associated with definitive local therapy and/or provide a window of feasibility for curative-intent surgery.

Clinical response to glucocorticoids is attributed to their anti-inflammatory effects and the apoptosis of the mast cells via activation of the glucocorticoid receptor (GCR)[15,57]. As a transcription factor, activation of the cytosolic GCR (cGCR) results in changes in gene expression: anti-inflammatory and regulatory gene expression is transactivated while pro-inflammatory gene expression is transrepressed[57]. Glucocorticoid administration also exerts its clinical effects through non-genomic mechanisms. Activation of cGCR results in apoptosis of cells by targeting pro-survival factors for degradation[58]. Intracellular signaling is altered due to the release of proteins from the cGCR multi-protein complex upon binding of glucocorticoids, resulting in rapid anti-inflammatory effects[57]. Glucocorticoids also act directly by negatively impacting cellular growth through inhibition of arachidonic acid release or via direct interaction with cellular membranes. These direct interactions alter cellular physicochemical properties and the function of membrane-associated proteins, allowing interference of cytokine synthesis, antigen processing, phagocytosis and migration[57]. Glucocorticoid resistance in canine mast cells is reported to be related to inhibition of GCR-mediated gene expression changes, increased cellular efflux, and increase in anti-apoptotic factors[59]. The alteration of gene expression, cellular function, and intra- and intercellular communication by glucocorticoids has potential to affect the number of cells actively in the cell cycle, how rapidly cells are progressing through the cell cycle, cellular and nuclear morphology, and qualities of the tumor and stromal microenvironment.

The clinical implications of potential alterations in the MCT grade or the scoring of proliferation indices are significant. The prognosis and subsequent management of the patient after definitive local therapy is dichotomized by MCT grade. Typically, low-grade tumors require no further treatment, even if incompletely-excised, and patients are expected to have good to excellent outcomes; while patients with high-grade tumors are expected to succumb to their disease and require intensive multi-modal therapeutic strategies[1,2,6,35-39,48,52,55,60-69]. This then raises the question of how pre-operative glucocorticoid treatment may impact histological parameters, criteria for grade determination and immunohistochemical detection of proliferation indices in canine MCTs.

The objective of this study was to evaluate changes in tumor grade, mitotic count, Ki67, AgNOR, and AgNORxKi67 scores following short-course, anti-inflammatory neoadjuvant prednisone in canine patients with MCTs. This pilot study was intended to guide hypothesis generation and future study design, and assist in power analysis calculations regarding the impact of short-term neoadjuvant prednisone administration on the histological and proliferation indices in canine cutaneous mast cell tumors.

## **2. Materials and Methods**

### *2.1. Study Population*

Client-owned dogs presenting to Virginia-Maryland College of Veterinary Medicine Veterinary Teaching Hospital (VMCVM) with naïve or recurrent cutaneous mast cell tumors were recruited. Inclusion criteria included a minimum body weight of 5 kg, cytologic diagnosis of mast cell tumor by board-certified clinical pathologist, tumor size  $\geq 1$  cm and  $< 10$  cm in longest diameter, and expected survival of  $\geq 4$  weeks without therapy. Prior surgery with mast cell tumor recurrence was allowed. Exclusion criteria included creatinine, ALT or AST  $\geq 1.5$  x upper reference limit, albumin  $< 2.0$  g/dL, grade 2 or higher VCOG cytopenia, or concurrent or previous chemotherapy or kinase therapy, steroid administration, or radiation therapy. All clients were informed of the purpose of the study and informed consent was obtained. This study was approved by the Virginia Tech Institutional Animal Care and Use Committee (IACUC) and the Veterinary Hospital Board.

### *2.2. Study Design*

This was a prospective, single-arm, open-label pilot study. All procedures were performed at a single institution. All tumor measurements throughout the study were performed prior to manipulation, taken in three dimensions using digital calipers, and performed by the same investigator throughout the study (S.K.). Baseline evaluation included physical exam, tumor measurements and photographs, CBC, serum biochemistry panel, and urinalysis. Within seven days of screening evaluation, a pre-treatment incisional 4-6mm punch biopsy was performed under sedation using standard sedation protocols selected at clinician discretion. Patients were discharged with oral prednisone at a targeted dose of 1.0 mg/kg administered once daily for 10-14 days. Concurrent treatment with H1 or H2 blocking agents was acceptable. Clients maintained and submitted a daily account of medication administration and observations. Prednisone was discontinued on the day of excisional biopsy. An exam, tumor measurements and photographs, CBC, serum biochemistry, and urinalysis were performed prior to excisional biopsy. Post-treatment tumor measurements are defined as the longest tumor diameter at the end of prednisone therapy but before excisional biopsy. Any adverse events noted were graded according to the VCOG-CTAE [70]. Gross surgical margins were recorded for each tumor. Curative-intent surgical margins were defined as either wide excision (lateral surgical margins  $> 2$ cm) or as lateral surgical margins proportional to the widest tumor diameter[55,71-75]. Surgical margins not meeting the definition of curative-intent was considered marginal excision. Excisional biopsy and post-operative management was performed by or under the supervision of a Diplomate of the American College of Veterinary Surgeons (ACVS) per standard of care at the VMCVM.

### 2.3. Assessment of histologic parameters

Incisional pre-treatment and excisional post-treatment biopsy samples were processed in standard preparation for routine histological evaluation. All samples were interpreted and histologic margins reported by a Diplomate of the American College of Veterinary Pathologists (ACVP) for immediate clinical use. Upon completion of all patient enrollment and participation, all samples were randomized, masked, and digitized by a non-investigator. Images were re-evaluated by a single board-certified (ACVP) pathologist (K.L.). All samples were graded according to the Patnaik and Kiupel grading systems, and assigned to one of four possible categories: Grade 1/Low Grade (G1/LG), Grade 2/Low Grade (G2/LG), Grade 2/High Grade (G2/HG), and Grade 3/High Grade (G3/HG)[1]. The mitotic count reported in number of mitoses per 10 high powered fields. Complete histologic margins were defined in this study as  $\geq 2\text{mm}$ [73].

### 2.4. Assessment of proliferation indices

Unstained histological slides of paired incisional pre-treatment and excisional post-treatment biopsy samples were submitted to a commercially-available reference lab (Michigan State University Diagnostic Center of Population and Animal Health (MSU DCPAH)) for immunohistochemical staining for Ki-67 and AgNOR. Results were reported per standard for all routine samples presented to MSU DCPAH.

### 2.5. Statistical Analyses

Continuous variables were analyzed with paired t-test for normally-distributed data or the Wilcoxon test for data not normally distributed. All p-values were 2-sided and p-values  $< 0.05$  were considered statistically significant. Statistical analyses were performed with standard software (MedCalc Statistical Software version 18.1 (MedCalc Software bvba, Ostend, Belgium; <https://www.medcalc.org>; 2018).

## 3. Results

### 3.1. Study population and tumor details

Thirteen dogs were screened for enrollment. All dogs met the eligibility criteria, were enrolled, underwent incisional biopsy, and initiated prednisone treatment. Two dogs were removed from the study prior to excisional biopsy. One dog was removed from the study due to grade 4 gastrointestinal toxicity (gastrointestinal ulceration), and another dog was excluded due histological diagnosis on pre-treatment biopsy inconsistent with mast cell tumor. Eleven dogs completed the study with 11 paired tumor samples available for evaluation.

Patient and tumor details are listed in **Table 1**. Most of the tumors were novel ( $n=9$ ), and two dogs had recurrent mast cell tumors. The median age was 7.5 years (range, 3 years to 12 years). There were six castrated males and five spayed females. The median weight was 27.3 kg (4.3 kg – 45.3 kg). A variety of breeds were represented, with mixed-breed as the most common ( $n=4$ ), and the remaining dogs each representing one breed. The majority of tumors were located on the trunk, tail, or limbs ( $n=8$ ), with one tumor each in the inguinal region, oral cavity, and ventral to the eye.

The median dose of prednisone was 0.8 mg/kg/day (range: 0.5 – 1.2 mg/kg). The median duration of prednisone administration was 11 days (range: 10-14 days). Pre-treatment, the median tumor volume was 2.89 cm<sup>3</sup> (range: 0.8 cm<sup>3</sup> – 160 cm<sup>3</sup>), and the median longest diameter (LD) was 21 mm (range: 14 – 92 mm). Post-treatment, the median tumor volume was 1.73 cm<sup>3</sup> (range: 0.3 cm<sup>3</sup> – 58.6 cm<sup>3</sup>), and the median LD was 17 mm (range: 10 – 60mm). The overall response rate was 72.7%. Eight tumors decreased in size, one increased in size, and there was no change in size for two tumors. For the tumors that decreased in size, the median decrease in LD was 13.6 mm (range: 3 mm – 32 mm), with a median relative size decrease of 29% (17.6% – 47.4%).

**Table 1.** Patient demographics and tumor details.

Pa-tient #	Breed	Age (y)	Sex	Weig ht (kg)	Recur- rent or novel	Tumor Loca- tion	Tumor vol- ume (mm <sup>3</sup> )	Tumor vol- ume (% change)	Surgical margins <sup>1</sup> and Histologic margins <sup>2</sup>
1	Golden re- triever	7.5	FS	34.5	Novel	proximal lateral left forelimb	Pre: 2205 Post: 1125	-49	Wide Complete
2	Mixed	3.2	FS	45.3	Novel	Tail	Pre: 1425 Post: 500	-64.9	Wide Complete
3	Yorkshire ter- rier	5.3	MC	4.3	Novel	Ventral to left eye	Pre: 1078 Post: 1078	0	Marginal Incomplete
4	Staffordshire terrier	6.7	MC	27.3	Novel	Interdigital	Pre: 3300 Post: 2550	-22.7	Marginal Incomplete
5	Mixed	11.6	MC	30.6	Recurrent	Left abdomen	Pre: 160,080 Post: 39,360	-75.4	Wide Complete
6	Miniature Schnauzer	9.6	MC	10.1	Novel	Right dorsal tar- sus	Pre: 588 Post: 1768	200.7	Proportional Incomplete
7	Mixed	8.3	MC	12.4	Novel	Distal medial left hindlimb	Pre: 765 Post: 300	-60.8	Proportional Incomplete
8	Norwegian elkhound	9.3	FS	24.9	Novel	Oral cavity	Pre: 9996 Post: 1729	-82.7	Marginal Complete
9	German shorthair pointer	3.5	MC	27.4	Novel	Proximal lateral right hindlimb	Pre: 2890 Post: 1260	-56.4	Wide Incomplete
10	Mixed	3.9	MC	38.9	Recurrent	Distal lateral right hindlimb	Pre: 58,608 Post: 58608	0	Wide Complete
11	Beagle	8.2	FS	19.8	Novel	Left inguinal re- gion	Pre: 9620 Post: 4104	-57.3	Wide Complete

<sup>1</sup> Wide excision defined as  $\geq 2$  cm surgical margins, proportional excision defined as lateral surgical margins proportional to the widest tumor diameter. <sup>2</sup> Complete excision is defined as  $\geq 2$  mm histologic margins.

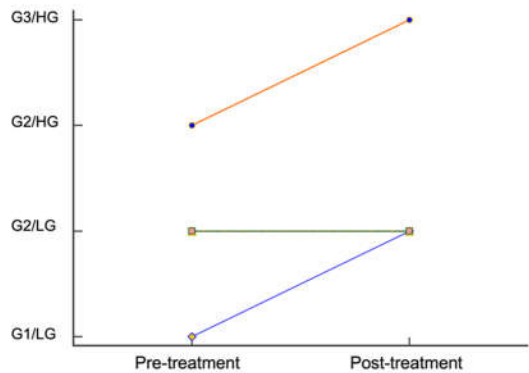
### 3.2. Histological parameters

Most of the tumors were amenable to curative-intent resection (n=8), defined as wide surgical margins  $> 2$  cm (n = 6) or lateral surgical margins proportional to the widest tumor diameter (n = 2) [71,74]. Complete histologic margins were achieved in 54.5% (n=6) of all tumors. All tumors with incomplete histologic margins were reported to have evidence of mast cells present at a surgical margin; i.e., no tumor margins classified as incomplete had "narrow", or clean histologic margins of  $< 2$  mm. Complete histologic margins were achieved in 62.5% (n=5) of tumors resected with curative-intent. All five of these tumors had been resected with wide surgical margins  $> 2$  cm. One tumor increased in size following prednisone treatment (patient #6), and had incomplete histologic margins following curative-intent surgical resection with proportional margins. One tumor (patient #7) with complete histologic margins had been marginally resected. This tumor was located in the oral cavity, was G2/LG, and demonstrated the greatest reduction in tumor volume following prednisone treatment. The patient last presented to the VMCVM with non-MCT-related morbidity 47 months following resection, with no evidence of tumor recurrence.

Individual patient tumor grades and mitotic counts are listed in **Table S1**. In pre-treatment tumor grade classification, G2/LG tumors were the most common (n=9), and there was one tumor classified as G1/LG and one tumor classified as G2/HG. Post-treatment, 10 tumors were classified as G2/LG, and one tumor classified as G3/HG. Two tumors were interpreted to have a different grade following prednisone treatment (**Figure**

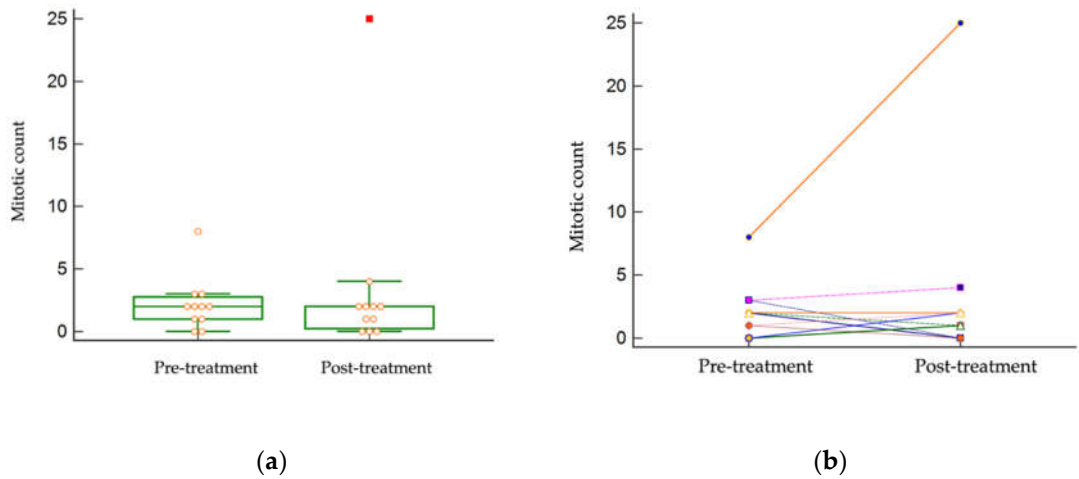


1). In both instances, the Patnaik designation increased, but the Kiupel designation did not change.



**Figure 1.** Pre- and post-treatment tumor grade classifications. The classification for nine G2/LG tumors did not change. The Patnaik classification, but not the Kiupel designation, increased for two tumors following treatment with prednisone.

The median mitotic count pre-treatment was 2 per 10 hpf (range: 0-8), and the median mitotic count post-treatment remained 2 per 10 hpf (range: 0-25). There was no statistically-significant difference between the mitotic counts pre- and post-treatment ( $p=0.4210$ ) (**Figure 2**). The median mitotic count excluding the G2/HG (pre-treatment) was 2 per 10 hpf (range: 0-3), and the median mitotic count excluding the G3/HG (post-treatment) was 1.5 per 10 hpf (range: 0-4). The one Kiupel high-grade tumor (patient #3) was noted to have an increase in the mitotic count post-treatment.



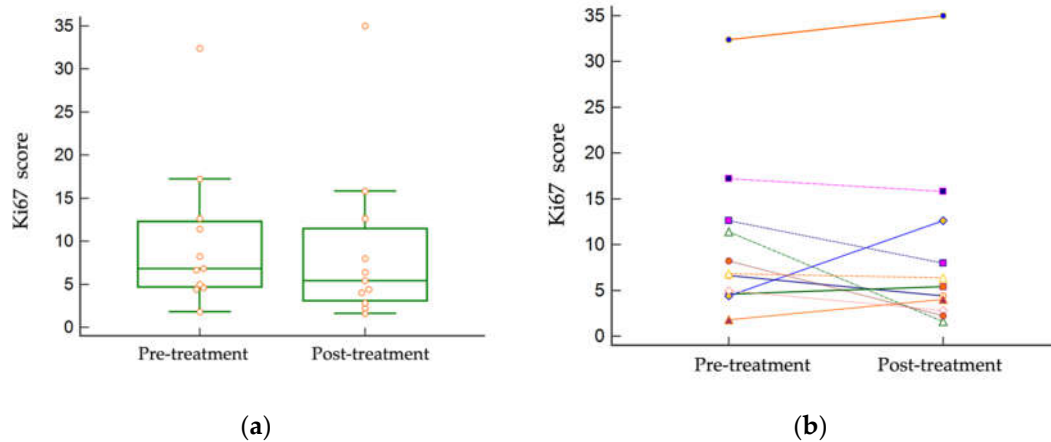
**Figure 2.** Mitotic counts in paired tumor samples. There was no statistically-significant difference between pre- and post-treatment mitotic counts. (a) Distribution of mitotic counts pre- and post-treatment; (b) Individual tumor paired mitotic count.

3.3. Proliferation indices

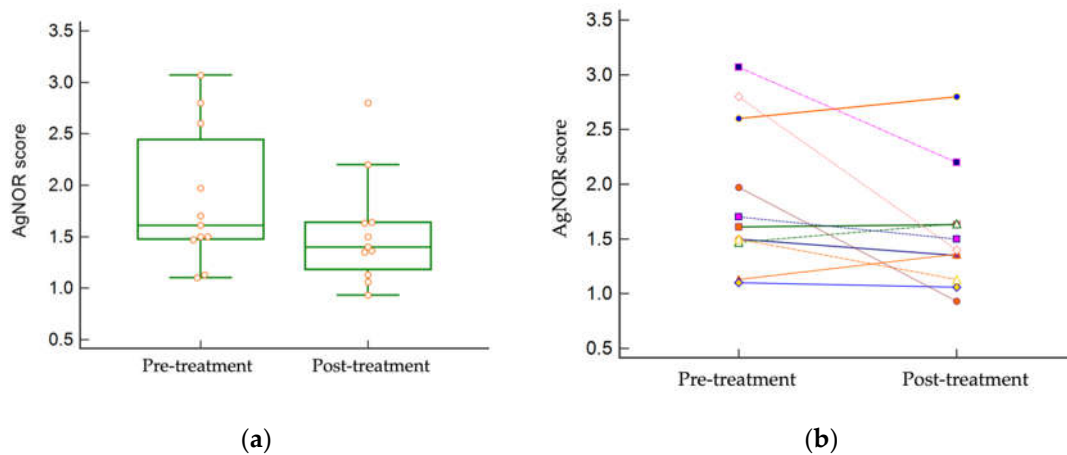
All individual patient tumor proliferation indices pre- and post-treatment are listed in **Table S1**. The median Ki67 score pre-treatment was 6.8 (1.8 – 32.4) and was 5.4 (1.6 – 35) post-treatment. Differences on an individual level varied, with most patients’ scores remaining roughly the same: increasing or decreasing within < 3 points. Other patients’ scores markedly decreased (patients #4 and 10), or markedly increased (patient #7) following treatment. There was no statistically-significant difference between pre- and post-treatment cohort Ki67 scores ( $p = 0.4393$ ) (**Figure 3**).

The median AgNOR score pre-treatment was 1.61 (1.1 – 3.07) and was 1.4 (0.93 – 2.8) post-treatment. Most individual paired tumors had similar AgNOR scores pre- and post-treatment, varying by less than 0.2 points. Three patients' AgNOR scores decreased by > 1 following treatment (patients #5, 8, and 10), although the difference in AgNOR scores between pre- and post-treatment cohorts did not reach significance ( $p = 0.0885$ ) (**Figure 4**).

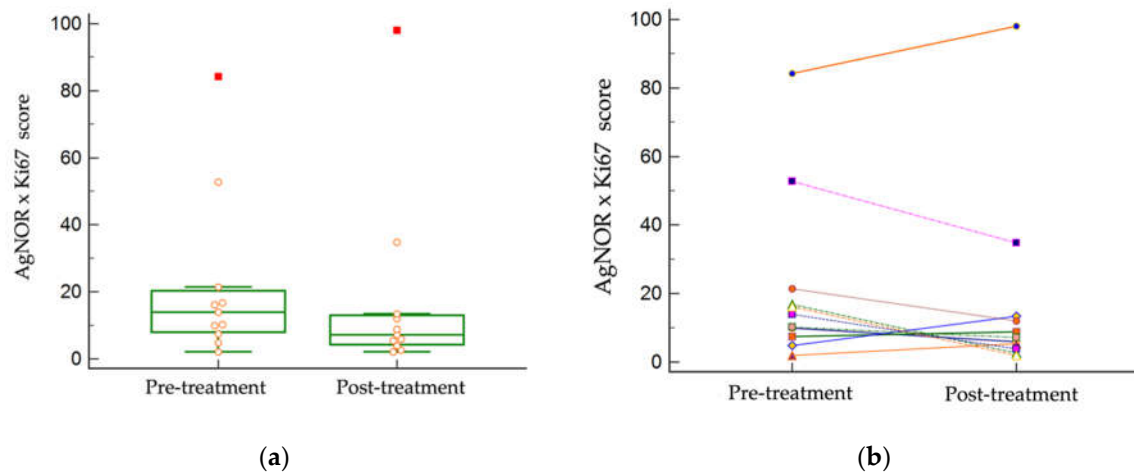
The median AgNORxKi67 product score pre-treatment was 14 (2.0 – 84.2) and the median post-treatment was 7.2 (2.1 – 98). There was no statistically-significant difference in the AgNORxKi67 score between treatment cohorts ( $p = 0.2046$ ) (**Figure 5**). There was only one set of paired tumor samples with AgNORxKi67 scores above 54 (patient #3), whose tumor was classified as G2/HG pre-treatment and G3/HG post-treatment[10].



**Figure 3.** Ki67 scores in paired tumor samples. There was no statistically-significant difference between pre- and post-treatment Ki67 scores. (a) Distribution of Ki67 scores pre- and post-treatment; (b) Individual tumor paired Ki67 scores.



**Figure 4.** AgNOR scores in paired tumor samples. There was no statistically-significant difference between pre- and post-treatment AgNOR scores. (a) The median AgNOR score pre- and post-treatment; (b) Individual tumor paired AgNOR scores. Three patients had a decrease > 1 in the AgNOR score following treatment.



**Figure 5.** AgNORxKi67 scores in paired tumor samples. There was no statistically-significant difference between pre- and post-treatment AgNORxKi67 scores. (a) The distribution of Ag67 scores pre- and post-treatment; (b) Individual tumor paired AgNORxKi67 scores. .

#### 4. Discussion

The objective of this study was to evaluate changes in tumor grade, mitotic count, Ki67, AgNOR, and AgNORxKi67 scores following short-course, anti-inflammatory neoadjuvant prednisone in canine patients with cutaneous mast cell tumors. The impetus for this study was the common clinical practice of neoadjuvant prednisone treatment in attempt to cytoreduce mast cell tumors, but always with the unanswerable question as to the impact on the tumor histopathology and the subsequent prognostication and treatment recommendations.

There is no consensus regarding the utility of pre-treatment biopsy in the initial screening evaluation of canine mast cell tumors[1]. The pre-treatment biopsy in our study had potential to independently affect the histological parameters assessed following treatment with prednisone. Local inflammation and mast cell degranulation associated with a biopsy procedure could lead to alterations in the proliferation indices scores or in the criteria for tumor grading, such as mitotic count, cellular or nuclear morphology, and edema or necrosis. In a recent study by Shaw et al, pre-treatment biopsy samples and subsequent excisional biopsy samples had a very high level of concordance using the Patnaik grading system and high level of concordance using the Kiupel grading system[76]. This was true regardless of the tumor location or the biopsy technique employed: wedge, punch, or needle; specifically, the punch biopsy had a 100% agreement under the Patnaik system and 95% agreement under the Kiupel system. The mean duration between pre-treatment and excisional biopsy in that study was 14 days, and the median was 9 days (2 – 111 days). All pre-treatment biopsies in our study were performed using a 4mm or 6mm punch instrument with a median duration between pre-treatment biopsy and excisional biopsy of 11 days (10 – 14 days). This suggests that the pre-treatment biopsy procedure in our study likely had minimal impact on the histologic parameters of the excisional biopsy.

Tumor grade and mitotic count are the most consistent prognostic factors in canine mast cell tumors[1,9,39,77-80]. Clinically, the prognosis and treatment recommendations can be dichotomized based on tumor grade: G1/LG and G2/LG tumors conferring an excellent prognosis, typically with no additional therapy required; while G2/HG and G3/HG tumors consistently result in a 1-year survival rate of < 50% even with additional local or systemic therapy[1,81]. In our study, there was no statistically-significant difference in the median mitotic count, nor in the tumor grade classification, following treatment with prednisone. The tumor grade classification was altered following treatment for two tumors. In both cases, the Patnaik assignment increased but the Kiupel assignment did not change, and the overall change in tumor grade classification had no clinical impact. Our findings are consistent with those of a recently-published study, in which there



was no statistically-significant difference in the mitotic count following prednisone treatment, and 2/13 paired tumor samples following prednisone treatment had an altered Patnaik grade without change in the Kiupel grade[40]. All but one of the tumors in our study was classified as low-grade. Additional studies restricted to the impact of pre-treatment with prednisone on high-grade tumor mitotic count and grade classification are indicated. It is important to note that tumor grade classification using the Patnaik system is subject to inter-observer variability, reported to have 50-60% discordance, while the Kiupel system reports 96-98% consistency among pathologists[1,6,82]. Furthermore, determination of mitotic count can be subject to individual variation[7,8]. In our study, a single pathologist interpreted all tumor samples after they were digitized, randomized, and masked by a non-investigator, thus controlling for both inter-observer variability and bias due to knowledge of sample origin or treatment status.

The proliferation indices AgNOR and Ki67, and their product AgNORxKi67, have demonstrated utility in refining the prognosis of canine mast cell tumors, especially in intermediate-grade tumors[2,10,39,80,83-87]. Clinically, these indices may also be useful in determining whether adjuvant therapy is warranted following surgical excision, as increasing AgNOR and Ki67 scores have been associated with increased risk of local tumor recurrence and metastasis[10,39,84,86,87]. Most canine mast cell tumors are intermediate-grade tumors (Patnaik grade 2) which are now also classified as either Kiupel grade high or Kiupel grade low[1]. Clinically, there is no standard of care for intermediate-grade tumors and the proliferation indices provide complementary information that are used to guide ancillary treatment decisions. Therefore, understanding how administration of routine peri-operative medications impacts these scores is important during the initial treatment planning. A recent study has evaluated the impact of opioid administration on histologic parameters including the proliferation indices in canine cutaneous mast cell tumors, but the impact of prednisone has not been previously evaluated[40,88]. In our study, there was no trend noted at the individual level, and no statistically-significant difference noted in the paired tumor samples for the Ki67 score. On an individual level, most patients' AgNOR score varied by  $< 0.2$  points between paired samples. However, there were three patients whose AgNOR score decreased by  $> 1$  following prednisone treatment. This did not reach statistical significance ( $p = 0.08$ ), which could be due to type II error. Post hoc power analysis was performed, and 27 paired samples would be required to detect a mean difference of 0.314 in AgNOR score, with  $\alpha = 0.05$ , and  $\beta = 20$ . There are few studies that evaluate AgNOR score as an independent prognostic factor, and unfortunately, AgNOR was not an immunohistochemical marker that was included in a recent systematic review and meta-analysis[2]. All existing studies have demonstrated via multi-variate analyses that increasing AgNOR score is associated with increased risk of local tumor recurrence, distant tumor occurrence, lymph node metastasis, and/or MCT-related mortality[10,83,87]. Based on our findings, larger, randomized, controlled studies evaluating the impact of prednisone on AgNOR score and long-term follow-up would be warranted. It is important to note that in the available literature, there is variability in the methodology and cut-off points for assessing the proliferation indices, particularly Ki67[2]. We evaluated AgNOR and Ki67 in this study via sample submission to an external, commercial laboratory. This provided consistent, unbiased, validated, and reproducible data which have practical and applicable relevance.

Glucocorticoids are used as sole therapy in cutaneous mast cell tumors, administered orally or intra-lesionally. Existing studies classify most responses as partial responses, with all but one study reporting overall response rates between 63-75%[14,19,22,39-41]. All responses in our study were partial responses, with the overall response rate of 72.7%. For the tumors that responded, the median decrease in tumor LD was 1.36 cm and the relative decrease in tumor volume was 29%. Response to glucocorticoid administration has been associated with larger tumor size and low-grade classification[22,39]. Four of the tumors in our study were  $> 5\text{cm}^3$ , and nearly all of them were Kiupel low-grade.

It is consistently reported that complete surgical resection is the treatment of choice for mast cell tumors, and complete histologic margins may be considered curative for low-

grade tumors[1,22,41-44,47,50,51,55,60,62,63,66,67,71-75,84,86,89-102]. Mast cell tumors can be deemed non-resectable or not amenable to curative-intent surgical margins. In a recent study, there was a significant association with increased risk of post-operative complication in patients with MCTs and incomplete histologic margins[103]. It is reasonable to attempt cytoreduction with pre-operative glucocorticoid administration in patients with non-resectable or marginally-resectable MCTs. Concerns regarding glucocorticoid treatment for cytoreduction include whether surgical margins based on post-treatment tumor size would yield complete histologic margins, and whether treatment would be associated with increased risk for post-operative incisional complications. In our study and others, surgical margins based on tumor size following pre-operative treatment with glucocorticoids have yielded complete histologic margins[22,40,103]. Long-term prospective evaluation of these patients is warranted to determine whether local tumor recurrence rate is impacted[55]. Post-operative complications following MCT resection, whether wide or intentionally marginal, is reported to be 13-29%[41,103]. Although not evaluated in our study, others have reported that dogs treated with pre-operative glucocorticoids have not had an increased risk of post-operative complications[22,40,103].

There are several limitations to our study. This study was intended to generate data to guide hypothesis generation and future study design, and assist in power analysis calculations regarding the impact of short-term neoadjuvant prednisone administration on the histological and proliferation indices in canine cutaneous mast cell tumors. As such, there was no control group, the patients did not undergo standardized staging evaluation, and there is no long-term outcome data. Identification of tumor grade was not a study enrollment criteria, resulting in a paucity of high-grade tumors in our study participants. As such, the findings in this study are applicable to low-grade tumors and should not be extrapolated to high-grade tumors. Glucocorticoid resistance mechanisms in canine mast cells is related to inhibition of GCR-mediated gene expression changes and an increase in anti-apoptotic factors[59]. It is possible that as a cohort, the tumors in our study failed to demonstrate significant changes due to variation in gross tumor response to glucocorticoid therapy. Future studies may elect to stratify tumor cohorts or restrict inclusion criteria based on clinical response to glucocorticoids to maximize identification of tumor histological or proliferation alterations following treatment. Our pilot study focused on commercially-accessible parameters of routine tumor histopathology and immunohistochemical detection of AgNOR and Ki67. However, a multi-faceted methodological approach evaluating impact on gross and histologic tumor parameters, differential gene expression, and protein expression may be warranted in future studies to better characterize the role and impact of glucocorticoids in MCT management. Additionally, evaluation of changes in the tumor volumes, the histologic margins, and the relationship between the surgical dose and histologic margins were secondary and exploratory objectives of our study and should be interpreted with caution. Tumor margins were not re-evaluated by a single investigator-pathologist, which introduces the potential for inter-observer error, and the tumor volumes post-treatment must be interpreted carefully in light of the pre-treatment biopsy, in that the decrease in tumor volume and LD may be impacted by the pre-treatment biopsy two weeks prior to final measurement[104]. While our findings corroborate that a pre-treatment biopsy does not seem to impact tumor grade nor mitotic count, the pre-treatment biopsy procedure could have independently affected the proliferation indices, which were not evaluated in a recent study, and larger prospective, randomized placebo-controlled studies are necessary for further investigation[40,76].

## 5. Conclusions

The data from this study provide the catalysts and foundation for the next steps into the investigation of the role glucocorticoids in canine MCT management. The results indicate that there appears to be no clinically-relevant alterations in tumor grade classification, mitotic count, nor the proliferation indices in low-grade mast cell tumors, three criteria consistently relied-upon in the management of canine cutaneous mast cell tumors.

Decreases in the proliferation index of AgNOR warrants further investigation, and randomized, placebo-controlled, appropriately-powered studies are necessary to confirm our results. The findings of our study can guide patient and tumor selection criteria for future studies: stratifying cohorts based on tumor grade, size, and response to glucocorticoid therapy. This study may also provide impetus for multi-tiered molecular investigation to characterize the global impact of neoadjuvant glucocorticoid therapy in canine MCTs. Finally, this study provides support for tangentially-related investigation regarding factors associated with response to prednisone therapy, and the long-term impact of neoadjuvant prednisone administration on surgical dose and the resulting histologic margins.

**Supplementary Materials:** Table S1: Paired tumor grades, mitotic counts, and proliferation indices.

**Author Contributions:** Conceptualization, Shawna Klahn and Nikolaos Dervisis; Data curation, Shawna Klahn; Formal analysis, Shawna Klahn, Nikolaos Dervisis and Kevin Lahmers; Funding acquisition, Shawna Klahn; Investigation, Shawna Klahn, Nikolaos Dervisis and Marian Benitez; Methodology, Shawna Klahn, Nikolaos Dervisis, Kevin Lahmers and Marian Benitez; Project administration, Shawna Klahn; Resources, Shawna Klahn and Kevin Lahmers; Software, Shawna Klahn, Nikolaos Dervisis and Kevin Lahmers; Supervision, Shawna Klahn and Marian Benitez; Validation, Shawna Klahn; Visualization, Shawna Klahn and Nikolaos Dervisis; Writing – original draft, Shawna Klahn; Writing – review & editing, Shawna Klahn, Nikolaos Dervisis, Kevin Lahmers and Marian Benitez. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by the Veterinary Memorial Fund, grant number 441944. This trial was registered in the AVMA Animal Health Studies Database: [https://ebusiness.avma.org/aahsd/study\\_search.aspx](https://ebusiness.avma.org/aahsd/study_search.aspx) #AAHSD000121. The work of Shawna Klahn was conducted with the support of the iTHRIV Scholars Program. The iTHRIV Scholars Program is supported in part by the National Center for Advancing Translational Sciences of the National Institutes of Health under Award Numbers UL1TR003015 and KL2TR003016 as well as by Virginia Tech.

**Institutional Review Board Statement:** Institutional Review Board was not required for this study; The animal study protocol was approved by the equivalent board governing review and oversight in animal research, the Institutional Animal Care and Use Committee of Virginia Tech (#16-048 (CVM), April 28, 2016), and the clinical trial approved by the College of Veterinary Medicine Hospital Board.

**Informed Consent Statement:** Informed consent was obtained from all subjects involved in the study. This work involved non-experimental animals only and followed high medical standards (best practice) for individual patient care. All samples were obtained following standard veterinary diagnostic and surgical procedures.

**Data Availability Statement:** All generated or analyzed during this study are included in this article. The datasets used for analysis are available from the corresponding author upon request.

**Acknowledgments:** We would like to thank the following team members for their contributions to the success of this trial: Mindy Quigley, Clinical Trials Coordinator; Veterinarians involved in screening and case management: Brittanie Partridge, Erin (Fagan) Cletzer, Eric Tempel, Noelle Muro, Sabrina Barry, Otto Lanz; Licensed Veterinary Technicians involved in patient care and client communication: Stefanie Olsen, Lauren Scaletta, Amanda Conrad; ViTALS histotechnicians and Anatomic Pathology laboratory: Kelli Hall-Manning, Jinhua Zhang, and Megan Zalek; Financial fund management: Drema Foster, VTH Business Office Manager, and Tracie Smith, Grant Financial Specialist. This study was presented in part at the Veterinary Cancer Society Annual Conference, Louisville, KY, October 2018.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

- Berlato, D.; Bulman-Fleming, J.; Clifford, C.A.; Garrett, L.; Intile, J.; Jones, P.; Kamstock, D.A.; Liptak, J.M.; Pavuk, A.; Powell, R.; et al. Value, Limitations, and Recommendations for Grading of Canine Cutaneous Mast Cell Tumors: A Consensus of the Oncology-Pathology Working Group. *Veterinary pathology* **2021**, *58*, 858-863, doi:10.1177/03009858211009785.
- Freytag, J.O.; Queiroz, M.R.; Govoni, V.M.; Pereira, I.V.A.; Pulz, L.H.; de Francisco Strefezzi, R.; Queiroga, F.L.; Cogliati, B. Prognostic value of immunohistochemical markers in canine cutaneous mast cell tumours: A systematic review and meta-analysis. *Veterinary and comparative oncology* **2021**, *19*, 529-540, doi:10.1111/vco.12692.
- London, C.A.; Thamm, D.H. 21 - Mast Cell Tumors. In *Withrow and MacEwen's Small Animal Clinical Oncology (Sixth Edition)*, Vail, D.M., Thamm, D.H., Liptak, J.M., Eds.; W.B. Saunders: St. Louis (MO), 2019; pp. 382-403.
- de Nardi, A.B.; Dos Santos Horta, R.; Fonseca-Alves, C.E.; de Paiva, F.N.; Linhares, L.C.M.; Firmo, B.F.; Ruiz Sueiro, F.A.; de Oliveira, K.D.; Lourenço, S.V.; De Francisco Strefezzi, R.; et al. Diagnosis, Prognosis and Treatment of Canine Cutaneous and Subcutaneous Mast Cell Tumors. *Cells* **2022**, *11*, doi:10.3390/cells11040618.
- Patnaik, A.K.; Ehler, W.J.; MacEwen, E.G. Canine cutaneous mast cell tumor: morphologic grading and survival time in 83 dogs. *Veterinary pathology* **1984**, *21*, 469-474, doi:10.1177/030098588402100503.
- Kiupel, M.; Webster, J.D.; Bailey, K.L.; Best, S.; DeLay, J.; Detrisac, C.J.; Fitzgerald, S.D.; Gamble, D.; Ginn, P.E.; Goldschmidt, M.H.; et al. Proposal of a 2-tier histologic grading system for canine cutaneous mast cell tumors to more accurately predict biological behavior. *Veterinary pathology* **2011**, *48*, 147-155, doi:10.1177/0300985810386469.
- Meuten, D.J.; Moore, F.M.; George, J.W. Mitotic Count and the Field of View Area: Time to Standardize. *Veterinary pathology* **2016**, *53*, 7-9, doi:10.1177/0300985815593349.
- Bertram, C.A.; Aubreville, M.; Gurtner, C.; Bartel, A.; Corner, S.M.; Dettwiler, M.; Kershaw, O.; Noland, E.L.; Schmidt, A.; Sledge, D.G.; et al. Computerized Calculation of Mitotic Count Distribution in Canine Cutaneous Mast Cell Tumor Sections: Mitotic Count Is Area Dependent. *Veterinary pathology* **2020**, *57*, 214-226, doi:10.1177/0300985819890686.
- Reynolds, B.D.; Thomson, M.J.; O'Connell, K.; Morgan, E.J.; Gummow, B. Patient and tumour factors influencing canine mast cell tumour histological grade and mitotic index. *Veterinary and comparative oncology* **2019**, *17*, 338-344, doi:10.1111/vco.12477.
- Webster, J.D.; Yuzbasiyan-Gurkan, V.; Miller, R.A.; Kaneene, J.B.; Kiupel, M. Cellular proliferation in canine cutaneous mast cell tumors: associations with c-KIT and its role in prognostication. *Veterinary pathology* **2007**, *44*, 298-308, doi:10.1354/vp.44-3-298.
- Sun, X.; Kaufman, P.D. Ki-67: more than a proliferation marker. *Chromosoma* **2018**, *127*, 175-186, doi:10.1007/s00412-018-0659-8.
- McStay, B. Nucleolar organizer regions: genomic 'dark matter' requiring illumination. *Genes & development* **2016**, *30*, 1598-1610, doi:10.1101/gad.283838.116.
- Gall, J.G. The human nucleolus organizer regions. *Genes & development* **2019**, *33*, 1617-1618, doi:10.1101/gad.334748.119.
- McCaw, D.L.; Miller, M.A.; Ogilvie, G.K.; Withrow, S.J.; Brewer, W.G., Jr.; Klein, M.K.; Bell, F.W.; Anderson, S.K. Response of canine mast cell tumors to treatment with oral prednisone. *Journal of veterinary internal medicine / American College of Veterinary Internal Medicine* **1994**, *8*, 406-408.
- Takahashi, T.; Kadosawa, T.; Nagase, M.; Mochizuki, M.; Matsunaga, S.; Nishimura, R.; Sasaki, N. Inhibitory effects of glucocorticoids on proliferation of canine mast cell tumor. *The Journal of veterinary medical science / the Japanese Society of Veterinary Science* **1997**, *59*, 995-1001.
- Thamm, D.H.; Mauldin, E.A.; Vail, D.M. Prednisone and vinblastine chemotherapy for canine mast cell tumor--41 cases (1992-1997). *Journal of veterinary internal medicine / American College of Veterinary Internal Medicine* **1999**, *13*, 491-497, doi:10.1892/0891-6640(1999)013<0491:pavcf>2.3.co;2.
- Cahalane, A.K.; Payne, S.; Barber, L.G.; Duda, L.E.; Henry, C.J.; Mauldin, G.E.; Frimberger, A.E.; Cotter, S.M.; Moore, A.S. Prognostic factors for survival of dogs with inguinal and perineal mast cell tumors treated surgically with or without adjunctive treatment: 68 cases (1994-2002). *Journal of the American Veterinary Medical Association* **2004**, *225*, 401-408, doi:10.2460/javma.2004.225.401.
- Davies, D.R.; Wyatt, K.M.; Jardine, J.E.; Robertson, I.D.; Irwin, P.J. Vinblastine and prednisolone as adjunctive therapy for canine cutaneous mast cell tumors. *Journal of the American Animal Hospital Association* **2004**, *40*, 124-130.
- Dobson, J.; Cohen, S.; Gould, S. Treatment of canine mast cell tumours with prednisolone and radiotherapy. *Veterinary and comparative oncology* **2004**, *2*, 132-141, doi:10.1111/j.1476-5810.2004.00048.x.
- Camps-Palau, M.A.; Leibman, N.F.; Elmslie, R.; Lana, S.E.; Plaza, S.; McKnight, J.A.; Risbon, R.; Bergman, P.J. Treatment of canine mast cell tumours with vinblastine, cyclophosphamide and prednisone: 35 cases (1997-2004). *Veterinary and comparative oncology* **2007**, *5*, 156-167, doi:10.1111/j.1476-5829.2006.00125.x.
- Hayes, A.; Adams, V.; Smith, K.; Maglennon, G.; Murphy, S. Vinblastine and prednisolone chemotherapy for surgically excised grade III canine cutaneous mast cell tumours. *Veterinary and comparative oncology* **2007**, *5*, 168-176, doi:10.1111/j.1476-5829.2007.00135.x.
- Stanclift, R.M.; Gilson, S.D. Evaluation of neoadjuvant prednisone administration and surgical excision in treatment of cutaneous mast cell tumors in dogs. *Journal of the American Veterinary Medical Association* **2008**, *232*, 53-62, doi:10.2460/javma.232.1.53.
- Vickery, K.R.; Wilson, H.; Vail, D.M.; Thamm, D.H. Dose-escalating vinblastine for the treatment of canine mast cell tumour. *Veterinary and comparative oncology* **2008**, *6*, 111-119, doi:10.1111/j.1476-5829.2007.00147.x.



24. Cooper, M.; Tsai, X.; Bennett, P. Combination CCNU and vinblastine chemotherapy for canine mast cell tumours: 57 cases. *Veterinary and comparative oncology* **2009**, *7*, 196-206, doi:10.1111/j.1476-5829.2009.00190.x.
25. Hosoya, K.; Kisseberth, W.C.; Alvarez, F.J.; Lara-Garcia, A.; Beamer, G.; Stromberg, P.C.; Couto, C.G. Adjuvant CCNU (lomustine) and prednisone chemotherapy for dogs with incompletely excised grade 2 mast cell tumors. *Journal of the American Animal Hospital Association* **2009**, *45*, 14-18, doi:10.5326/0450014.
26. Taylor, F.; Gear, R.; Hoather, T.; Dobson, J. Chlorambucil and prednisolone chemotherapy for dogs with inoperable mast cell tumours: 21 cases. *The Journal of small animal practice* **2009**, *50*, 284-289, doi:10.1111/j.1748-5827.2009.00732.x.
27. Hahn, K.A.; Legendre, A.M.; Shaw, N.G.; Phillips, B.; Ogilvie, G.K.; Prescott, D.M.; Atwater, S.W.; Carreras, J.K.; Lana, S.E.; Ladue, T.; et al. Evaluation of 12- and 24-month survival rates after treatment with masitinib in dogs with nonresectable mast cell tumors. *American journal of veterinary research* **2010**, *71*, 1354-1361, doi:10.2460/ajvr.71.11.1354.
28. Rassnick, K.M.; Bailey, D.B.; Russell, D.S.; Flory, A.B.; Kiselow, M.A.; Intile, J.L.; Malone, E.K.; Balkman, C.E.; Barnard, S.M. A phase II study to evaluate the toxicity and efficacy of alternating CCNU and high-dose vinblastine and prednisone (CVP) for treatment of dogs with high-grade, metastatic or nonresectable mast cell tumours. *Veterinary and comparative oncology* **2010**, *8*, 138-152, doi:10.1111/j.1476-5829.2010.00217.x.
29. Carlsten, K.S.; London, C.A.; Haney, S.; Burnett, R.; Avery, A.C.; Thamm, D.H. Multicenter prospective trial of hypofractionated radiation treatment, toceranib, and prednisone for measurable canine mast cell tumors. *Journal of veterinary internal medicine / American College of Veterinary Internal Medicine* **2012**, *26*, 135-141, doi:10.1111/j.1939-1676.2011.00851.x.
30. Burton, J.H.; Venable, R.O.; Vail, D.M.; Williams, L.E.; Clifford, C.A.; Axiak-Bechtel, S.M.; Avery, A.C.; Thamm, D.H. Pulse-Administered Toceranib Phosphate Plus Lomustine for Treatment of Unresectable Mast Cell Tumors in Dogs. *Journal of veterinary internal medicine / American College of Veterinary Internal Medicine* **2015**, *29*, 1098-1104, doi:10.1111/jvim.13573.
31. Smrkovski, O.A.; Essick, L.; Rohrbach, B.W.; Legendre, A.M. Masitinib mesylate for metastatic and non-resectable canine cutaneous mast cell tumours. *Veterinary and comparative oncology* **2015**, *13*, 314-321, doi:10.1111/vco.12053.
32. Serra Varela, J.C.; Pecceu, E.; Handel, I.; Lawrence, J. Tolerability of a rapid-escalation vinblastine-prednisolone protocol in dogs with mast cell tumours. *Vet Med Sci* **2016**, *2*, 266-280, doi:10.1002/vms3.42.
33. Bavcar, S.; de Vos, J.; Kessler, M.; de Fornel, P.; Buracco, P.; Murphy, S.; Hirschberger, J.; Argyle, D.J. Combination toceranib and lomustine shows frequent high grade toxicities when used for treatment of non-resectable or recurrent mast cell tumours in dogs: A European multicentre study. *Veterinary journal (London, England : 1997)* **2017**, *224*, 1-6, doi:10.1016/j.tvjl.2017.04.010.
34. Case, A.; Burgess, K. Safety and efficacy of intralesional triamcinolone administration for treatment of mast cell tumors in dogs: 23 cases (2005-2011). *Journal of the American Veterinary Medical Association* **2018**, *252*, 84-91, doi:10.2460/javma.252.1.84.
35. Hay, J.K.; Larson, V.S. Lomustine (CCNU) and prednisone chemotherapy for high-grade completely excised canine mast cell tumors. *The Canadian veterinary journal. La revue veterinaire canadienne* **2019**, *60*, 1326-1330.
36. Mendez, S.E.; Drobatz, K.J.; Duda, L.E.; White, P.; Kubicek, L.; Sorenmo, K.U. Treating the locoregional lymph nodes with radiation and/or surgery significantly improves outcome in dogs with high-grade mast cell tumours. *Veterinary and comparative oncology* **2019**, doi:10.1111/vco.12541.
37. Todd, J.E.; Nguyen, S.M.; White, J.; Langova, V.; Thomas, P.M.; Tzannes, S. Combination vinblastine and palladia for high-grade and metastatic mast cell tumors in dogs. *The Canadian veterinary journal. La revue veterinaire canadienne* **2021**, *62*, 1335-1340.
38. Marconato, L.; Stefanello, D.; Kiupel, M.; Finotello, R.; Polton, G.; Massari, F.; Ferrari, R.; Agnoli, C.; Capitani, O.; Giudice, C.; et al. Adjuvant medical therapy provides no therapeutic benefit in the treatment of dogs with low-grade mast cell tumours and early nodal metastasis undergoing surgery. *Veterinary and comparative oncology* **2020**, *18*, 409-415, doi:10.1111/vco.12566.
39. Horta, R.S.; Lavalley, G.E.; Monteiro, L.N.; Souza, M.C.C.; Cassali, G.D.; Araújo, R.B. Assessment of Canine Mast Cell Tumor Mortality Risk Based on Clinical, Histologic, Immunohistochemical, and Molecular Features. *Veterinary pathology* **2018**, *55*, 212-223, doi:10.1177/0300985817747325.
40. Linde, K.J.; Stockdale, S.L.; Mison, M.B.; Perry, J.A. The effect of prednisone on histologic and gross characteristics in canine mast cell tumors. *The Canadian veterinary journal. La revue veterinaire canadienne* **2021**, *62*, 45-50.
41. Cockburn, E.; Janovec, J.; Solano, M.A.; L'Eplattenier, H. Marginal excision of cutaneous mast cell tumors in dogs was not associated with a higher rate of complications or prolonged wound healing than marginal excision of soft tissue sarcomas. *Journal of the American Veterinary Medical Association* **2022**, 1-6, doi:10.2460/javma.21.05.0235.
42. Michels, G.M.; Knapp, D.W.; DeNicola, D.B.; Glickman, N.; Bonney, P. Prognosis following surgical excision of canine cutaneous mast cell tumors with histopathologically tumor-free versus nontumor-free margins: a retrospective study of 31 cases. *Journal of the American Animal Hospital Association* **2002**, *38*, 458-466, doi:10.5326/0380458.
43. Weisse, C.; Shofer, F.S.; Sorenmo, K. Recurrence rates and sites for grade II canine cutaneous mast cell tumors following complete surgical excision. *Journal of the American Animal Hospital Association* **2002**, *38*, 71-73.
44. Gieger, T.L.; Theon, A.P.; Werner, J.A.; McEntee, M.C.; Rassnick, K.M.; DeCock, H.E. Biologic behavior and prognostic factors for mast cell tumors of the canine muzzle: 24 cases (1990-2001). *Journal of veterinary internal medicine / American College of Veterinary Internal Medicine* **2003**, *17*, 687-692.
45. Hahn, K.A.; King, G.K.; Carreras, J.K. Efficacy of radiation therapy for incompletely resected grade-III mast cell tumors in dogs: 31 cases (1987-1998). *Journal of the American Veterinary Medical Association* **2004**, *224*, 79-82.
46. Sfiligoi, G.; Rassnick, K.M.; Scarlett, J.M.; Northrup, N.C.; Gieger, T.L. Outcome of dogs with mast cell tumors in the inguinal or perineal region versus other cutaneous locations: 124 cases (1990-2001). *Journal of the American Veterinary Medical Association* **2005**, *226*, 1368-1374, doi:10.2460/javma.2005.226.1368.



47. Mullins, M.N.; Dernell, W.S.; Withrow, S.J.; Ehrhart, E.J.; Thamm, D.H.; Lana, S.E. Evaluation of prognostic factors associated with outcome in dogs with multiple cutaneous mast cell tumors treated with surgery with and without adjuvant treatment: 54 cases (1998-2004). *Journal of the American Veterinary Medical Association* **2006**, *228*, 91-95, doi:10.2460/javma.228.1.91.
48. Hillman, L.A.; Garrett, L.D.; de Lorimier, L.P.; Charney, S.C.; Borst, L.B.; Fan, T.M. Biological behavior of oral and perioral mast cell tumors in dogs: 44 cases (1996-2006). *Journal of the American Veterinary Medical Association* **2010**, *237*, 936-942, doi:10.2460/javma.237.8.936.
49. Elliott, J.W.; Cripps, P.; Blackwood, L.; Berlato, D.; Murphy, S.; Grant, I.A. Canine oral mucosal mast cell tumours. *Veterinary and comparative oncology* **2013**, doi:10.1111/vco.12071.
50. Schwab, T.M.; Popovitch, C.; DeBiasio, J.; Goldschmidt, M. Clinical outcome for MCTs of canine pinnae treated with surgical excision (2004-2008). *Journal of the American Animal Hospital Association* **2014**, *50*, 187-191, doi:10.5326/JAAHA-MS-6039.
51. Trappler, M.C.; Popovitch, C.A.; Goldschmidt, M.H.; Goldschmidt, K.H.; Risbon, R.E. Scrotal tumors in dogs: a retrospective study of 676 cases (1986-2010). *The Canadian veterinary journal. La revue vétérinaire canadienne* **2014**, *55*, 1229-1233.
52. Lejeune, A.; Skorupski, K.; Frazier, S.; Vanhaezebrouck, I.; Rebhun, R.B.; Reilly, C.M.; Rodriguez, C.O., Jr. Aggressive local therapy combined with systemic chemotherapy provides long-term control in grade II stage 2 canine mast cell tumour: 21 cases (1999-2012). *Veterinary and comparative oncology* **2015**, *13*, 267-280, doi:10.1111/vco.12042.
53. Elliott, J.W.; Cripps, P.; Blackwood, L.; Berlato, D.; Murphy, S.; Grant, I.A. Canine oral mucosal mast cell tumours. *Veterinary and comparative oncology* **2016**, *14*, 101-111, doi:10.1111/vco.12071.
54. Miller, R.L.; Van Lelyveld, S.; Warland, J.; Dobson, J.M.; Foale, R.D. A retrospective review of treatment and response of high-risk mast cell tumours in dogs. *Veterinary and comparative oncology* **2016**, *14*, 361-370, doi:10.1111/vco.12116.
55. Milovancev, M.; Townsend, K.L.; Tuohy, J.L.; Gorman, E.; Bracha, S.; Curran, K.M.; Russell, D.S. Long-term outcomes of dogs undergoing surgical resection of mast cell tumors and soft tissue sarcomas: A prospective 2-year-long study. *Veterinary surgery : VS* **2020**, *49*, 96-105, doi:10.1111/vsu.13225.
56. Sabattini, S.; Kiupel, M.; Finotello, R.; Stefanello, D.; Faroni, E.; Bertazzolo, W.; Bonfanti, U.; Rigillo, A.; Del Magno, S.; Foglia, A.; et al. A retrospective study on prophylactic regional lymphadenectomy versus nodal observation only in the management of dogs with stage I, completely resected, low-grade cutaneous mast cell tumors. *BMC veterinary research* **2021**, *17*, 331, doi:10.1186/s12917-021-03043-0.
57. Stahn, C.; Buttgerit, F. Genomic and nongenomic effects of glucocorticoids. *Nat Clin Pract Rheumatol* **2008**, *4*, 525-533, doi:10.1038/ncprheum0898.
58. C.W., D. Recent insights into the mechanisms of glucocorticosteroid-induced apoptosis. *Cell death and differentiation* **2002**, *9*, 17.
59. Matsuda, A. Long-term in-vitro glucocorticoid treatment induces glucocorticoid resistance in canine mast cell tumors. *Canadian journal of veterinary research = Revue canadienne de recherche vétérinaire* **2021**, *85*, 302-308.
60. Hume, C.T.; Kiupel, M.; Rigatti, L.; Shofer, F.S.; Skorupski, K.A.; Sorenmo, K.U. Outcomes of dogs with grade 3 mast cell tumors: 43 cases (1997-2007). *Journal of the American Animal Hospital Association* **2011**, *47*, 37-44, doi:10.5326/jaaha-ms-5557.
61. A retrospective review of treatment and response of high-risk mast cell tumours in dogs. *Veterinary and comparative oncology* **2016**, *14*, 361-370, doi:10.1111/vco.12116.
62. Smith, J.; Kiupel, M.; Farrelly, J.; Cohen, R.; Olmsted, G.; Kirpensteijn, J.; Brocks, B.; Post, G. Recurrence rates and clinical outcome for dogs with grade II mast cell tumours with a low AgNOR count and Ki67 index treated with surgery alone. *Veterinary and comparative oncology* **2017**, *15*, 36-45, doi:10.1111/vco.12140.
63. Vincenti, S.; Findji, F. Influence of treatment on the outcome of dogs with incompletely excised grade-2 mast cell tumors. *Schweizer Archiv für Tierheilkunde* **2017**, *159*, 171-177, doi:10.17236/sat00109.
64. Olsen, J.A.; Thomson, M.; O'Connell, K.; Wyatt, K. Combination vinblastine, prednisolone and toceranib phosphate for treatment of grade II and III mast cell tumours in dogs. *Vet Med Sci* **2018**, doi:10.1002/vms3.106.
65. Kiupel, M.; Camus, M. Diagnosis and Prognosis of Canine Cutaneous Mast Cell Tumors. *The Veterinary clinics of North America. Small animal practice* **2019**, *49*, 819-836, doi:10.1016/j.cvsm.2019.04.002.
66. Moore, A.S.; Frimberger, A.E.; Taylor, D.; Sullivan, N. Retrospective outcome evaluation for dogs with surgically excised, solitary Kiupel high-grade, cutaneous mast cell tumours. *Veterinary and comparative oncology* **2020**, doi:10.1111/vco.12565.
67. Karbe, G.T.; Davis, E.; Runge, J.J.; Brown, D.C.; Holt, D.E. Evaluation of scar revision after inadequate primary excision of cutaneous mast cell tumors in 85 dogs (2000-2013). *Veterinary surgery : VS* **2021**, *50*, 807-815, doi:10.1111/vsu.13619.
68. Mason, S.L.; Pittaway, C.; Gil, B.P.; Russak, O.M.; Westlake, K.; Berlato, D.; Benoit, J.; Morris, J.; Dobson, J.M. Outcomes of adjunctive radiation therapy for the treatment of mast cell tumors in dogs and assessment of toxicity: A multicenter observational study of 300 dogs. *Journal of veterinary internal medicine / American College of Veterinary Internal Medicine* **2021**, *35*, 2853-2864, doi:10.1111/jvim.16264.
69. Guerra, D.; Faroni, E.; Sabattini, S.; Agnoli, C.; Chalfon, C.; Stefanello, D.; Del Magno, S.; Cola, V.; Grieco, V.; Marconato, L. Histologic grade has a higher-weighted value than nodal status as predictor of outcome in dogs with cutaneous mast cell tumors and overtly metastatic sentinel lymph nodes. *Veterinary and comparative oncology* **2022**, doi:10.1111/vco.12806.
70. Veterinary Co-operative Oncology Group - Common Terminology Criteria for Adverse Events (VCOG-CTCAE) following chemotherapy or biological antineoplastic therapy in dogs and cats v1.0. *Veterinary and comparative oncology* **2004**, *2*, 195-213, doi:10.1111/j.1476-5810.2004.0053b.x.
71. Pratschke, K.M.; Atherton, M.J.; Sillito, J.A.; Lamm, C.G. Evaluation of a modified proportional margins approach for surgical resection of mast cell tumors in dogs: 40 cases (2008-2012). *Journal of the American Veterinary Medical Association* **2013**, *243*, 1436-1441, doi:10.2460/javma.243.10.1436.

72. Chu, M.L.; Hayes, G.M.; Henry, J.G.; Oblak, M.L. Comparison of lateral surgical margins of up to two centimeters with margins of three centimeters for achieving tumor-free histologic margins following excision of grade I or II cutaneous mast cell tumors in dogs. *Journal of the American Veterinary Medical Association* **2020**, *256*, 567-572, doi:10.2460/javma.256.5.567.
73. Selmic, L.E.; Ruple, A. A systematic review of surgical margins utilized for removal of cutaneous mast cell tumors in dogs. *BMC veterinary research* **2020**, *16*, 5, doi:10.1186/s12917-019-2227-8.
74. Itoh, T.; Kojimoto, A.; Uchida, K.; Chambers, J.; Shii, H. Long-term postsurgical outcomes of mast cell tumors resected with a margin proportional to the tumor diameter in 23 dogs. *The Journal of veterinary medical science / the Japanese Society of Veterinary Science* **2021**, *83*, 230-233, doi:10.1292/jvms.20-0281.
75. Saunders, H.; Thomson, M.J.; O'Connell, K.; Bridges, J.P.; Chau, L. Evaluation of a modified proportional margin approach for complete surgical excision of canine cutaneous mast cell tumours and its association with clinical outcome. *Veterinary and comparative oncology* **2021**, *19*, 604-615, doi:10.1111/vco.12630.
76. Shaw, T.; Kudnig, S.T.; Firestone, S.M. Diagnostic accuracy of pre-treatment biopsy for grading cutaneous mast cell tumours in dogs. *Veterinary and comparative oncology* **2018**, *16*, 6, doi:10.1111/vco.12346.
77. Romansik, E.M.; Reilly, C.M.; Kass, P.H.; Moore, P.F.; London, C.A. Mitotic index is predictive for survival for canine cutaneous mast cell tumors. *Veterinary pathology* **2007**, *44*, 335-341, doi:10.1354/vp.44-3-335.
78. Elston, L.B.; Sueiro, F.A.; Cavalcanti, J.N.; Metze, K. The importance of the mitotic index as a prognostic factor for survival of canine cutaneous mast cell tumors: a validation study. *Veterinary pathology* **2009**, *46*, 362-364, author reply 364-365, doi:10.1354/vp.46-2-362.
79. van Lelyveld, S.; Warland, J.; Miller, R.; Maw, H.; Foale, R.; Goodfellow, M.; Dobson, J. Comparison between Ki-67 index and mitotic index for predicting outcome in canine mast cell tumours. *The Journal of small animal practice* **2015**, *56*, 312-319, doi:10.1111/jsap.12320.
80. Vascellari, M.; Giantin, M.; Capello, K.; Carminato, A.; Morello, E.M.; Vercelli, A.; Granato, A.; Buracco, P.; Dacasto, M.; Mutinelli, F. Expression of Ki67, BCL-2, and COX-2 in canine cutaneous mast cell tumors: association with grading and prognosis. *Veterinary pathology* **2013**, *50*, 110-121, doi:10.1177/0300985812447829.
81. Sabattini, S.; Scarpa, F.; Berlato, D.; Bettini, G. Histologic grading of canine mast cell tumor: is 2 better than 3? *Veterinary pathology* **2015**, *52*, 70-73, doi:10.1177/0300985814521638.
82. Northrup, N.C.; Howerth, E.W.; Harmon, B.G.; Brown, C.A.; Carmicheal, K.P.; Garcia, A.P.; Latimer, K.S.; Munday, J.S.; Rakich, P.M.; Richey, L.J.; et al. Variation among Pathologists in the Histologic Grading of Canine Cutaneous Mast Cell Tumors with Uniform Use of a Single Grading Reference. *Journal of Veterinary Diagnostic Investigation* **2005**, *17*, 561-564, doi:10.1177/104063870501700606.
83. Scase, T.J.; Edwards, D.; Miller, J.; Henley, W.; Smith, K.; Blunden, A.; Murphy, S. Canine mast cell tumors: correlation of apoptosis and proliferation markers with prognosis. *Journal of veterinary internal medicine / American College of Veterinary Internal Medicine* **2006**, *20*, 151-158.
84. Seguin, B.; Besancon, M.F.; McCallan, J.L.; Dewe, L.L.; Tenwolde, M.C.; Wong, E.K.; Kent, M.S. Recurrence rate, clinical outcome, and cellular proliferation indices as prognostic indicators after incomplete surgical excision of cutaneous grade II mast cell tumors: 28 dogs (1994-2002). *Journal of veterinary internal medicine / American College of Veterinary Internal Medicine* **2006**, *20*, 933-940.
85. Maglennon, G.A.; Murphy, S.; Adams, V.; Miller, J.; Smith, K.; Blunden, A.; Scase, T.J. Association of Ki67 index with prognosis for intermediate-grade canine cutaneous mast cell tumours. *Veterinary and comparative oncology* **2008**, *6*, 268-274, doi:10.1111/j.1476-5829.2008.00168.x.
86. Smith, J.; Kiupel, M.; Farrelly, J.; Cohen, R.; Olmsted, G.; Kirpensteijn, J.; Post, G. Recurrence rates and clinical outcome for dogs with grade II mast cell tumours with a low AgNOR count and Ki67 index treated with surgery alone. *Veterinary and comparative oncology* **2015**, doi:10.1111/vco.12140.
87. Krick, E.L.; Kiupel, M.; Durham, A.C.; Thaiwong, T.; Brown, D.C.; Sorenmo, K.U. Investigating Associations Between Proliferation Indices, C-kit, and Lymph Node Stage in Canine Mast Cell Tumors. *Journal of the American Animal Hospital Association* **2017**, *53*, 258-264, doi:10.5326/jaaha-ms-6265.
88. Marouda, C.; Anagnostou, T.; Savvas, I.; Papazoglou, L.G.; Psalla, D. The Effect of Opioid Administration on Cytologic and Histopathologic Diagnosis of Canine Cutaneous Mast Cell Tumors Treated by Surgical Excision. *Veterinary Sciences* **2022**, *9*, doi:10.3390/vetsci9050202.
89. Murphy, S.; Sparkes, A.H.; Smith, K.C.; Blunden, A.S.; Brearley, M.J. Relationships between the histological grade of cutaneous mast cell tumours in dogs, their survival and the efficacy of surgical resection. *The Veterinary record* **2004**, *154*, 743-746.
90. Fulcher, R.P.; Ludwig, L.L.; Bergman, P.J.; Newman, S.J.; Simpson, A.M.; Patnaik, A.K. Evaluation of a two-centimeter lateral surgical margin for excision of grade I and grade II cutaneous mast cell tumors in dogs. *Journal of the American Veterinary Medical Association* **2006**, *228*, 210-215, doi:10.2460/javma.228.2.210.
91. Ozaki, K.; Yamagami, T.; Nomura, K.; Narama, I. Prognostic significance of surgical margin, Ki-67 and cyclin D1 protein expression in grade II canine cutaneous mast cell tumor. *The Journal of veterinary medical science / the Japanese Society of Veterinary Science* **2007**, *69*, 1117-1121.
92. Monteiro, B.; Boston, S.; Monteith, G. Factors influencing complete tumor excision of mast cell tumors and soft tissue sarcomas: a retrospective study in 100 dogs. *The Canadian veterinary journal. La revue veterinaire canadienne* **2011**, *52*, 1209-1214.

93. Kry, K.L.; Boston, S.E. Additional local therapy with primary re-excision or radiation therapy improves survival and local control after incomplete or close surgical excision of mast cell tumors in dogs. *Veterinary surgery : VS* **2014**, *43*, 182-189, doi:10.1111/j.1532-950X.2014.12099.x.
94. Miller, R.L.; Van Lelyveld, S.; Warland, J.; Dobson, J.M.; Foale, R.D. A retrospective review of treatment and response of high-risk mast cell tumours in dogs. *Veterinary and comparative oncology* **2014**, doi:10.1111/vco.12116.
95. Donnelly, L.; Mullin, C.; Balko, J.; Goldschmidt, M.; Krick, E.; Hume, C.; Brown, D.C.; Sorenmo, K. Evaluation of histological grade and histologically tumour-free margins as predictors of local recurrence in completely excised canine mast cell tumours. *Veterinary and comparative oncology* **2015**, *13*, 70-76, doi:10.1111/vco.12021.
96. Risselada, M.; Mathews, K.G.; Griffith, E. Surgically planned versus histologically measured lateral tumor margins for resection of cutaneous and subcutaneous mast cell tumors in dogs: 46 cases (2010-2013). *Journal of the American Veterinary Medical Association* **2015**, *247*, 184-189, doi:10.2460/javma.247.2.184.
97. Lowe, R.; Gavazza, A.; Impellizeri, J.A.; Soden, D.M.; Lubas, G. The treatment of canine mast cell tumours with electrochemotherapy with or without surgical excision. *Veterinary and comparative oncology* **2017**, *15*, 775-784, doi:10.1111/vco.12217.
98. Robinson, W.P.; Elliott, J.; Baines, S.J.; Owen, L.; Shales, C.J. Intramuscular mast cell tumors in 7 dogs. *The Canadian veterinary journal. La revue veterinaire canadienne* **2017**, *58*, 931-935.
99. Does, C.B.; Milovancev, M.; Russell, D.S. Comparison of histologic margin status in low-grade cutaneous and subcutaneous canine mast cell tumours examined by radial and tangential sections. *Veterinary and comparative oncology* **2018**, *16*, 125-130, doi:10.1111/vco.12321.
100. Ferrari, R.; Marconato, L.; Buracco, P.; Boracchi, P.; Giudice, C.; Iussich, S.; Grieco, V.; Chiti, L.E.; Favretto, E.; Stefanello, D. The impact of extirpation of non-palpable/normal-sized regional lymph nodes on staging of canine cutaneous mast cell tumours: A multicentric retrospective study. *Veterinary and comparative oncology* **2018**, *16*, 505-510, doi:10.1111/vco.12408.
101. Bae, S.; Milovancev, M.; Bartels, C.; Irvin, V.L.; Tuohy, J.L.; Townsend, K.L.; Leeper, H. Histologically low-grade, yet biologically high-grade, canine cutaneous mast cell tumours: A systematic review and meta-analysis of individual participant data. *Veterinary and comparative oncology* **2020**, doi:10.1111/vco.12581.
102. Marconato, L.; Stefanello, D.; Kiupel, M.; Finotello, R.; Polton, G.; Massari, F.; Ferrari, R.; Agnoli, C.; Capitani, O.; Giudice, C.; et al. Adjuvant medical therapy provides no therapeutic benefit in the treatment of dogs with low-grade mast cell tumours and early nodal metastasis undergoing surgery. *Veterinary and comparative oncology* **2020**, doi:10.1111/vco.12566.
103. Iodence, A.E.; Wallace, M.L.; Grimes, J.A.; Schmiedt, C.W. Dogs undergoing surgical excision of mast cell tumors are not at increased risk of incisional complications. *Journal of the American Veterinary Medical Association* **2021**, *260*, S88-s95, doi:10.2460/javma.20.09.0488.
104. Kiser, P.K.; Löhr, C.V.; Meritet, D.; Spagnoli, S.T.; Milovancev, M.; Russell, D.S. Histologic processing artifacts and inter-pathologist variation in measurement of inked margins of canine mast cell tumors. *Journal of veterinary diagnostic investigation : official publication of the American Association of Veterinary Laboratory Diagnosticians, Inc* **2018**, *30*, 377-385, doi:10.1177/1040638718757582.