

**High atmospheric CO<sub>2</sub> concentration causes increased respiration by the oxidative pentose phosphate pathway in chloroplasts – Supporting information**

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**Notes S1. Recalculation of previously reported estimates of flux through the plastidial anaplerotic pathway at low C<sub>a</sub>**

In Wieloch *et al.* (2022), we expressed fractionation signals discussed here as

$$\delta D_i = \frac{D_i}{\sum D_{ME}/6} - 1 \quad \text{Eqn S1}$$

where D<sub>i</sub> and D<sub>ME</sub> denote relative deuterium abundances at specific carbon-bound hydrogens of glucose and the six methyl-group hydrogens of the glucose derivative used for NMR measurements (3,6-anhydro-1,2-O-isopropylidene- $\alpha$ -D-glucofuranose), respectively. Here, I express these signals as

$$\delta D_1 = \frac{D_1}{D_{6S}} - 1 \quad \text{Eqn S2}$$

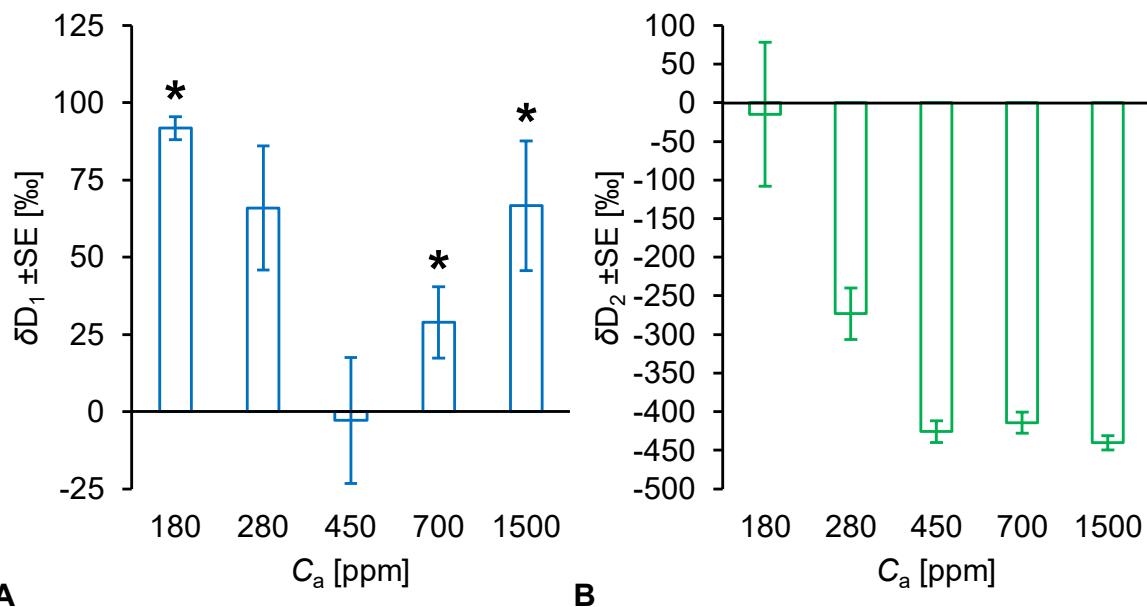
and

$$\delta D_2 = \frac{D_2}{D_{6R}} - 1 \quad \text{Eqn S3}$$

Based on equation S2,  $\delta D_1$  is 66‰ at C<sub>a</sub> = 280 ppm and 92‰ at C<sub>a</sub> = 180 ppm (Figure S1A). Furthermore,  $\delta D_1$  and thus flux through the plastidial anaplerotic pathway is significantly greater than zero at C<sub>a</sub> = 180 ppm ( $p < 0.05$ ,  $n = 2$ ) while it comes close to being significantly greater than zero at C<sub>a</sub> = 280 ppm ( $p < 0.13$ ,  $n = 2$ ). A previously published model describing deuterium fractionation by G6PD can be used to estimate the plastidial anaplerotic flux, and associated respiration (Wieloch *et al.*, 2022). Based on this model,  $\approx 9.3\%$  and  $12.7\%$  of the G6P entering the starch biosynthesis pathway is diverted into the anaplerotic pathway at C<sub>a</sub> = 280 and 180 ppm, respectively. Assuming 50% of all net assimilated carbon becomes starch (Sharkey *et al.*, 1985), anaplerotic flux and associated respiration proceeds at  $\approx 5\%$  and  $\approx 7\%$  relative to the rate of net carbon assimilation at C<sub>a</sub> = 280 and 180 ppm. These rates are probably

strongly underestimated since, at low  $C_a$ , much of the fractionation signal introduced by G6PD can be expected to not arrive in starch (see biochemical explanation in Wieloch *et al.*, 2022).

Based on equation S3,  $\delta D_2$  is  $\approx -427\text{‰}$  at  $C_a \geq 450$  ppm,  $-273\text{‰}$  at  $C_a = 280$  ppm, and  $-15\text{‰}$  at  $C_a = 180$  ppm (Figure S1B). This indicates that the PGI reaction is on the side of F6P at  $C_a \geq 450$  ppm and shifts towards equilibrium with decreasing  $C_a$  below 450 ppm (Wieloch *et al.*, 2022).



**Figure S1** Deuterium abundance at glucose H<sup>1</sup> (A, blue bars), and H<sup>2</sup> (B, green bars) of sunflower leaf starch. Asterisks denote deuterium abundances that are significantly greater than zero (one-tailed one-sample t-test:  $p < 0.05$ ,  $n \geq 2$ ). At 280 ppm,  $\delta D_1$  is close to being significantly greater than zero ( $p < 0.13$ ,  $n = 2$ ). The plants were raised in chambers over 7 to 8 weeks at  $C_a = 450$  ppm. After a day in darkness to drain the starch reserves, the plants were grown for two days at different levels of  $C_a$  (180, 280, 450, 700, 1500 ppm) corresponding to different levels of  $C_i$  (140, 206, 328, 531, 1365 ppm). Data expressed as  $\delta D_1 = D_1/D_{6S}-1$  and  $\delta D_2 = D_2/D_{6R}-1$  where  $D_i$  denotes relative deuterium abundances at specific carbon-bound hydrogens of glucose. Deuterium abundances at glucose H<sup>6S</sup> and H<sup>6R</sup> are used as references because glucose H<sup>1</sup> and H<sup>6S</sup> and H<sup>2</sup> and H<sup>6R</sup> have the same precursors at the chloroplast triose-phosphate level, and H<sup>6S</sup> and H<sup>6R</sup> are not modified in the starch biosynthesis pathway (Wieloch *et al.*, 2022).

## References

**Sharkey TD, Berry JA, Raschke K. 1985.** Starch and sucrose synthesis in *Phaseolus vulgaris* as affected by light, CO<sub>2</sub>, and abscisic acid. *Plant Physiology* **77**: 617–620.

**Wieloch T, Augusti A, Schleucher J. 2022.** Anaplerotic flux into the Calvin-Benson cycle. Hydrogen isotope evidence for *in vivo* occurrence in C<sub>3</sub> metabolism. *New Phytologist* **234**: 405–411.