

Minimum alveolar concentration of isoflurane in rats chronically treated with the synthetic cannabinoid WIN 55,212-2

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Abstract

Background: The isoflurane minimum alveolar concentration (MAC) was measured in rats chronically treated with WIN 55,212-2.

Methods: The isoflurane MAC was determined in 24 male rats from the end expiratory samples at time of tail clamping under the following conditions: without treatment (MAC_{ISO}), in rats treated for 21 days with WIN 55,212-2 ($MAC_{ISO+WIN55}$) and other group 8 days after stopping treatment for 21 days with WIN 55,212-2 ($MAC_{ISO+WIN55+8D}$).

Results: The MAC_{ISO} was 1.32 ± 0.06 . In the $MAC_{ISO+WIN55}$ group the MAC increase to 1.69 ± 0.09 (28%). After 8 days stopping treatment, MAC did not decrease significantly 1.67 ± 0.07 (26%).

Conclusions: The administration for 21 days of WIN 55,212-2 increases the MAC of isoflurane in rats; this effect does not disappear after 8 days of discontinuing treatment with the synthetic cannabinoid.

Introduction

Cannabis sativa, one of the oldest psychotropic drugs known (1), is also the most consumed drug according to the World Drug Report 2019 published by the United Nations, which estimated that there are over 188 million consumers of cannabis (2). Delta-9-tetrahydrocannabinol (Delta 9-THC) was isolated in the 1960s (3), the cannabinoid type 1 receptor (CB1) was identified at the end of the 1980s (4), and the cannabinoid type 2 receptor (CB2) was discovered in 1993 (5); subsequently, the first (6) and second endogenous cannabinoids were discovered (7). The endocannabinoid system (ECS) is involved in many health and disease processes (8), and different research groups have performed investigations (9) by modulating the activity of the ECS and assessing the possible effects of manipulating this system. This information could have important therapeutic applications (10,11,12) in a variety of diseases like epilepsy, acute and chronic pain in humans.

Therefore, we believe it is necessary to study the effects of chronic consumption of these substances on the requirements of inhalation anesthetics in patients that will be submitted to general anesthesia.

The objective of this study was to evaluate the interaction of chronically administered cannabinoid agonist on the minimum alveolar concentration of isoflurane (MAC_{ISO}); to the knowledge of the authors, there are no studies that evaluate this interaction.

Keywords: MAC; Isoflurane; WIN 55,212-2; Rat

Materials and Methods

In this study, with protocol number 3492/2013CHT, 24 male Wistar rats weighing 310 ± 20 g was used. This experiment was approved by the Animal Research Ethics Committee for Animal Experimentation of the Faculty of Veterinary Medicine of the University of the State of Mexico. Rats were housed in groups of four in Plexiglas cages, with a 12 h light/12 h dark cycle (lights on at 07:00), with a relative humidity of 50%–60% and ambient temperature $23 \pm 2^\circ\text{C}$. The animals had free access to water and rodent food (Prolab1 RMH 2500, USA). Animals were allowed to acclimatize for one week before the experiments took place. These experiments were performed during the morning (09:00–12:00). All animals were handled according to the guidelines in the Guide for the Care and Use of Laboratory Animals.

Anesthetic procedure

Anesthetic induction was performed by placing each rat in the induction chamber providing 5% isoflurane (Forane; Baxter Laboratories, USA) with a continuous oxygen flow of 5 L/min. Once the animal was anesthetized and lost jaw tone, it was removed from the induction chamber and placed in dorsal recumbence for endotracheal intubation. The oral cavity was opened and with the aid of an otoscope, the larynx was visualized and a flexible blunt-tip wire guide was inserted inside and used to direct the endotracheal catheter (16G Teflon catheter: Introcan; B-Braun, Brazil), which was secured to the maxilla by means of adhesive tape.

Correct placement of the catheter was confirmed by CO₂ infrared absorption analysis (BeneView T5, mindray, Multi-gas offers, Nanshan, China). The catheter was connected to a small T-piece breathing system with minimal dead space and fresh gas flow of 1 L/min of oxygen. The isoflurane concentration was adjusted as necessary based on assessment of the palpebral reflex and hemodynamic responses during instrumentation. During the study, rats were breathing spontaneously.

The carotid artery was exposed via surgical cut-down and catheterized using a 24-gauge catheter (Introcan; B-Braun, Brazil); this was connected to a pressure transducer system for direct blood pressure monitoring and the collection of arterial blood to determine blood gases. Systolic, diastolic, and mean arterial blood pressures (SAP, DAP and MAP, respectively) and heart rate (HR) were continuously monitored (BeneView T5, Mindray, Nanshan, China). For blood gas analysis (GEM Premier 3000; Instrumentation Laboratory, UK), 0.3 mL of blood was obtained immediately after determining the MAC to ensure (at that time) that values were within normal physiological parameters. Rectal temperature was maintained between 37°C and 38°C by means of a convective warming system (Equator1, SurgiVet1, Smiths Medical PM Inc., USA). Inspired isoflurane ($F_{I\text{iso}}$) and end-tidal ($F_{E\text{iso}}$) concentrations, end-tidal carbon dioxide tension ($P_{E\text{tCO}_2}$) and respiratory rate (RR) were continuously measured with an infrared gas analyzer previously calibrated (BeneView T5, mindray, Multi-gas offers, Nanshan, China) by endotracheal gas sampling (60 mL/min) obtained by means of a catheter inserted through the endotracheal tube with the tip located at the level of the carina.

MAC determination

Once instrumentation was performed and prior to assessing the isoflurane MAC, Fe_{iso} was adjusted to 1.32%, which is a value close to the isoflurane MAC previously reported by the authors (13). Once this concentration was achieved, it was maintained for 15 minutes in order to achieve the equilibrium of isoflurane partial pressure between alveolar gas, arterial blood, and spinal cord. (14) The isoflurane MAC was determined by the tail clamp method described by Quasha *et al.* (15). A painful noxious stimulus was applied with a hemostat clamped (8-inch Rochester Dean hemostatic forceps) on the tail at a specific end-tidal concentration of each volatile agent. The tail was clamped to the first ratchet lock for 60 seconds or until a positive response was observed. The tail was always stimulated proximally to the previous test site. A positive motor response was considered if jerking or twisting motions of the head or body, or movement of the extremities was observed. Negative responses included a lack of movement of the head and limbs, muscle rigidity, shivering, swallowing, and chewing, movement of the tail should not be considered.

If the response was positive, the delivered volatile anesthetic concentration was increased by 10%, and, if the response was negative, the concentration of the volatile anesthetic was decreased by 10%. After an equilibration period of 15 min, the application of the stimulus was repeated. The person assessing the response was

blinded with respect to the drugs administered to each rat. In each rat, the MAC was evaluated in duplicate.

The isoflurane MAC values were corrected to sea level by use of the formula (barometric pressure of location/760 mmHg) x obtained MAC value. The mean barometric pressure was obtained from the official city meteorological station for the altitude at which the experiment was performed (2680 m above sea level) and was 556 mmHg. At the end of each experiment, animals were euthanized with pentobarbital given intravenously (Anestesal, Pfizer, México) to animals deeply anaesthetized with the inhalant agent.

Experimental design

Using a random number generator (Excel 2007, Microsoft Office), the animals were distributed into three groups (n=8).

The control group (MAC_{ISO}) remained untreated for the measurement.

The MAC_{ISO+WIN55} group was treated intraperitoneally (i.p) with 1 mg/kg of WIN 55,212-2 (mesylate salt, Sigma-Aldrich, St. Louis, MO) every 24 hours (at 09:00 h) for 21 days Lawston *et al.* (16).

WIN 55,212-2 was suspended in a vehicle solution of 0.3% Tween 80 in saline (0.9%) as described by Tanda *et al.* (17). Isoflurane MAC measurements were performed 24 hours after the last treatment of WIN 55,212-2 (day 22).

MAC_{ISO+WIN55+8D} group was treated i.p with 1 mg/kg of WIN 55,212-2 every 24 hours

(at 09:00 h) for 21 days. The measurement of isoflurane MAC was performed 8 days after the last treatment with WIN 55,212-2 (day 29).

Statistical analysis

Statistical analysis was performed using Prism 6 (GraphPad Software, Inc., USA). The Shapiro-Wilk test was used for the assessment of data normality. Data are reported as mean \pm standard deviation (SD). Analysis of variance was performed and post hoc comparison of the groups was performed using the Holm-Sidak test. Values were considered statistically different when $p < 0.05$.

Results

The results are summarized in Table 1. The value of the mean \pm SD obtained in the MAC_{ISO} group was 1.32% \pm 0.06. The MAC_{ISO+WIN55} group showed a 28% increase in the isoflurane MAC; the mean value for this group was 1.69% \pm 0.09 and was significantly different when compared to the MAC_{ISO} group ($p < 0.0001$). The mean value of the MAC_{ISO+WIN55+8D} group was 1.67% \pm 0.07; there was no statistically difference when compared to the MAC_{ISO+WIN55} group ($p = 0.6995$), but a significant difference was found compared with the MAC_{ISO} ($p < 0.0001$).

Table 1. Isoflurane MAC in rats chronically treated with the synthetic cannabinoid WIN 55,212-2.

Group	MAC%	SD	% MAC increase	P-value	95% IC
MAC _{ISO}	1.32	0.06		-	1.27 - 1.37
MAC _{ISO+WIN55}	1.69*	0.09	28%	<0.0001	1.58 - 1.77
MAC _{ISO+WIN+8D}	1.67*	0.07	26%	<0.0001	1.60 - 1.75

*Statistically significant compared to the control group CAM_{ISO} ($p < 0.05$).

Discussion

MAC is defined as the alveolar concentration of an inhalatory anesthetic necessary to prevent movement in 50% of subjects exposed to a supramaximal noxious stimulus and represents an index of potency of anesthetic agents (18). In this work we observed that a synthetic cannabinoid chronically administered increases the MAC of isoflurane.

The use of cannabinoids has increased in a recreational and therapeutic way (2), to the knowledge of the authors, there are no reports of the effect of chronic administration of cannabinoids on the requirements of inhalation anesthetics. In this investigation, we observed that 21 days of administering the synthetic cannabinoid WIN 55,212-2 increased the isoflurane MAC in rats. However, this observation may not necessarily reflect the effect that occasional *Cannabis sativa* consumption could generate in humans. The most abundant substance present in the cannabis plant, Δ^9 -THC is responsible for its psychotropic effects (3), and is a phytocannabinoid

and partial agonist of the CB1 receptor. WIN 55,212-2 is a synthetic total cannabinoid agonist of the CB1/CB2 receptors (19) therefore, the effectiveness of the cannabinoid agonist and the duration of exposure could influence the effect on the requirements of inhalation anesthetics.

WIN 55,212-2 cause an increase in the central levels of norepinephrine, Page and collaborators (20) showed that rats treated with WIN 55,212-2 for 8 days presented an increase in noradrenergic activity, furthermore they demonstrated that repeated administration of WIN 55,212-2 stimulates CB1 cannabinoid receptors in cell bodies of

the *locus coeruleus* and in nerve terminals containing norepinephrine, generating an increase in norepinephrine efflux.

Norepinephrine levels in nervous terminals modulates MAC response of isoflurane as Miller *et al.* (21) demonstrated in a previous work, in experiments following the administration of alpha-methyldopa, reserpine and iproniazid, they demonstrated that the requirements of inhalational anesthetics are related to norepinephrine levels. Thus drugs that decrease the concentration of norepinephrine in the central nervous system decrease the MAC. On the other hand, drugs that increase norepinephrine levels cause an increase in the requirement for inhalation anesthetics. Similarly, it has been reported that acute administration of amphetamine (22) and cocaine (23) increases the MAC of halothane in dogs due to an increase in the catecholamine concentration in the central nervous system.

It's also known, diverse neurotransmitters such as norepinephrine, might manifest some of their actions by strongly inhibiting TWIK-related acid-sensitive K⁺ channels (TASK) and thus influence neuronal excitability. Similarly, Inhalation anesthetics activate and cannabinoid agonists inhibit TASK channels (24,25) Perhaps the collective inhibition between norepinephrine and cannabinoids of TASK channels led to the need for a higher concentration of isoflurane to increase in the anesthetic requirements necessary to prevent movement in response to painful stimulus.

Previous studies have reported that acute and subacute administration of Δ^9 -THC decreases the halothane MAC in dogs (26) and the cyclopropane MAC decreases in

rats (27) treated acutely and chronically (for one week) with $\Delta 9$ -THC. This discrepancy with our results may be explained on the basis of the experimental design, while we administered the cannabinoid receptor agonist for 21 days, Stoelting and Vitez administered the treatment for a less period of time, besides we used WIN55, 212-2, while the cited works used $\Delta 9$ -THC.

Another explanation for the differences observed in the isoflurane MAC can be through the observations made by Mechoulam *et al.* (28) and by Marciano *et al.* (29) which indicate that endocannabinoids have paradoxical effects on the central nervous system of mammals, since in some cases they generate an increase in neuronal excitability and in others they decrease it, depending on the dose administered. Similarly, they reported that cannabinoids cause short-term inhibitory effects on the release of glutamate; in contrast, prolonged stimulation of CB1 receptors by exogenous administration of cannabinoids could block the release of the inhibitory neurotransmitter, GABA (29).

In this sense, an increase in the isoflurane MAC suggests that systemic and sustained administration of WIN 55,212-2 reflected as an increase in the anesthetic requirements necessary to prevent movement in response to painful stimulus.

When measuring MAC after 8 days of stopping cannabinoid administration ($MAC_{ISO+WIN55+8D}$ group), we found no statistically significant differences when compared to the $MAC_{ISO+WIN55}$ group. Therefore, the increase in the isoflurane MAC caused by the administration of WIN 55,212-2 does not decrease after 8 days of

stopping the administration of the cannabinoid. This difference with the studies carried out by Page and collaborators (20) could be a consequence of the longer exposure to the cannabinoid used in our study and possibly it will take more time for the decrease in noradrenergic activity (30). To determine whether the effect of WIN 55,212-2 on isoflurane MAC is transient, further experiments will be necessary.

Conclusions

The administration for 21 days of WIN 55,212-2 increases the MAC of isoflurane in rats; this effect does not disappear after 8 days of discontinuing treatment with the synthetic cannabinoid.

References

1. Mechoulam R. The pharmacology of cannabis sativa. In: Mechoulam R, editor. Cannabinoids as therapeutic agents. Boca Raton, FL: CRC Press; 1986. pp 1-16.
2. United Nations Office on Drugs and Crime, World Drug Report 2019 (United Nations publication, Sales No. E.19.XI.8).
3. Gaoni Y, Mechoulam R: Isolation, structure and partial synthesis of an active constituent of hashish. J Am Chem Soc 1964; 86: 1646-1647.
4. Devane WA, Dysarz FA 3rd, Johnson MR, Melvin LS, Howlett AC. Determination and characterization of a cannabinoid receptor in rat brain. Mol Pharmacol. 1988; 34: 605-613.
5. Munro S, Thomas KL, Abu-Shaar M. Molecular characterization of a peripheral receptor for cannabinoids. Nature. 1993; 365: 61-65.
6. Devane WA, Hanuš L, Breuer A, Pertwee RG, Stevenson LA, Griffin G, et al. Isolation and structure of a brain constituent that binds to the cannabinoid receptor. Science. 1992; 258: 1946-1949.
7. Mechoulam R, Ben-Shabat S, Hanuš L, Ligumsky M, Kaminski NE, Schatz AR, et al. Identification of an endogenous 2-monoglyceride, present in canine gut, that binds to cannabinoid receptors. Biochem Pharmacol 1995; 50: 83-90.
8. Pacher P, Kunos G. Modulating the endocannabinoid system in human health and disease successes and failures. FEBS J 2013; 280: 1918-1943.
9. Mechoulam R. Cannabis: the Israeli perspective. J Basic Clin Physiol Pharmacol 2016; 27: 181-187.

10. Tantimonaco M, Ceci R, Sabatini S, Catani MV, Rossi A, Gasperi V, et al. Physical activity and the endocannabinoid system: an overview. *Cell Mol Life Sci.* 2014; 71: 2681-2698.
11. Mechoulam R, Hanus LO, Pertwee R, Howlett AC. Early phytocannabinoid chemistry to endocannabinoids and beyond. *Nature Rev Neurosci.* 2014; 15: 757-776.
12. Maccarrone M, Guzmán M, Mackie K, Doherty P, Harkany T. Programming of neural cells by (endo) cannabinoids: from physiological rules to emerging therapies. *Nat Rev Neurosci* 2014; 15: 786-801.
13. Chavez JR, Ibancovich JA, Sanchez-Aparicio P, Acevedo-Arcique CM, Moran-Muñoz R, Recillas-Morales S. Effect of acetaminophen alone and in combination with morphine and tramadol on the minimum alveolar concentration of isoflurane in rats. *PLoS One.* 2015; 25: 1-9.
14. Antognini JF, Schwartz K. Exaggerated anesthetic requirements in the preferentially anesthetized brain. *Anesthesiology.* 1993; 79(6): 1244-9.
15. Quasha AL, Eger EI, Tinker JH. Determination and applications of MAC. *Anesthesiology.* 1980; 53: 314-334.
16. Lawston J, Borella A, Robinson JK, Whitaker-Azmitia PM. Changes in hippocampal morphology following chronic treatment with the synthetic cannabinoid WIN 55,212-2. *Brain Research.* 2000; 877: 407-410.
17. Tanda G, Pontieri FE, Chiara GD. Cannabinoid and heroin activation of mesolimbic dopamine transmission by a common μ_1 opioid receptor mechanism. *Science.* 1997; 276: 2048-2050.

18. Eger EI, Saidman LJ, Brandstater B. Minimum alveolar anesthetic concentration: a standard of anesthetic potency. *Anesthesiology*. 1965; 26: 756-763.
19. Pertwee RG, Howlett AC, Abood ME, Alexander SPH, Di Marzo V, Elphick MR, et al. International Union of Basic and Clinical Pharmacology. LXXIX. Cannabinoid receptors and their ligands: beyond CB₁ and CB₂. *Pharmacol Rev*. 2010; 62: 588-631.
20. Page ME, Oropeza VC, Sparks SE, Qian Y, Menko AS, Van Bockstaele EJ. Repeated cannabinoid administration increases indices of noradrenergic activity in rats. *Pharmacol Biochem Behav*. 2007; 86: 162-168.
21. Miller RD, Way WL, Eger EI II. The effects of alpha-methylodopa, reserpine, guanethidine and iproniazid on minimum alveolar anesthesia requirement (MAC). *Anesthesiology*. 1968; 29: 1153-1158.
22. Johnston RR, Way WL, Miller RD. Alteration of anesthetic requirement by amphetamine. *Anesthesiology*. 1972; 36: 357-363.
23. Stoelting RK, Creasser CW, Martz RZ. Effect of cocaine administration on halothane MAC in dogs. *Anesth Analg*. 1975; 54: 422-424.
24. Linden AM, Aller MI, Leppa E, Vekovischeva O, Aitta-aho T, Veale EL, Mathie A, Rosenberg P, Wisden W, Korpi ER. The in vivo contributions of TASK-1 containing channels to the actions of inhalation anesthetics, the α 2 adrenergic sedative dexmedetomidine, and cannabinoid agonists. *JPET*; 317:615-626, 2006

25. Linden AM, Sandu C, Aller MI, Vekovischeva OY, Rosenberg PH, Wisden W, Korpi ER: TASK3 Knockout mice exhibit exaggerated nocturnal activity, impairments in cognitive functions, and reduced sensitivity to inhalation anesthetics. *JPET*; 323:924-934, 2000
26. Stoelting RK, Martz RC, Gartner J, Creasser C, Brown DJ, Forney RB. Effects of delta-9-tetrahydrocannabinol on halothane MAC in dogs. *Anesthesiology*. 1973; 38: 521-524.
27. Vitez TS, Way WL, Miller RD, Eger II EI. Effects of delta-9-tetrahydrocannabinol on cyclopropane MAC in the rat. *Anesthesiology*. 1973; 38: 525-527.
28. Mechoulam R, Lichtman AH. Stout guards of the central nervous system. *Science*. 2003; 2003 Oct 3; 302(5642): 65-67.
29. Marsicano G, Goodenough S, Monory K, Hermann H, Eder M, Cannich A, et al. CB1 cannabinoid receptors and on demand defense against excitotoxicity. *Science*. 2003; 302: 84-88.
30. Kobayashi RM, Palkovits M, Kopin IJ, Jacobowitz DM. Biochemical mapping of noradrenergic nerves arising from the rat locus coeruleus. *Brain Res*. 1974; 77: 269-279.