

Portable FRET-based Biosensor Device for On-site Lead Detec-tions

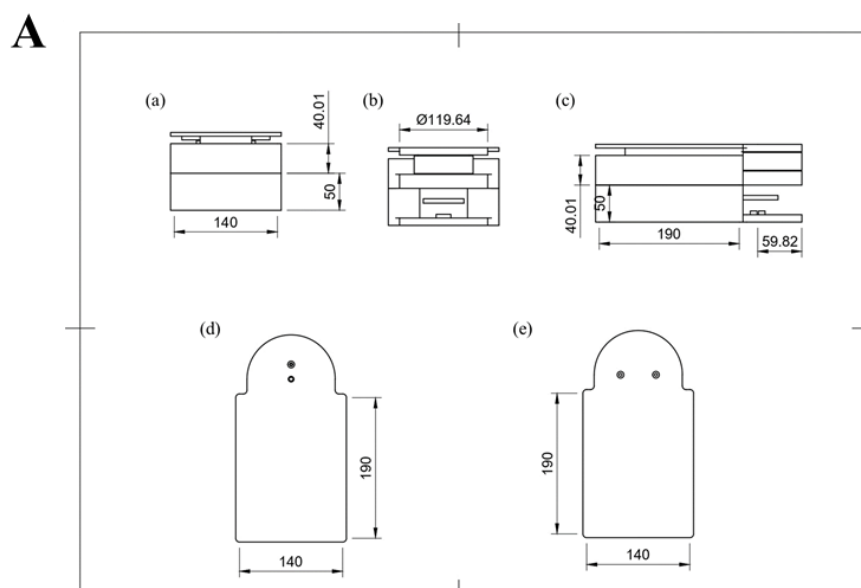
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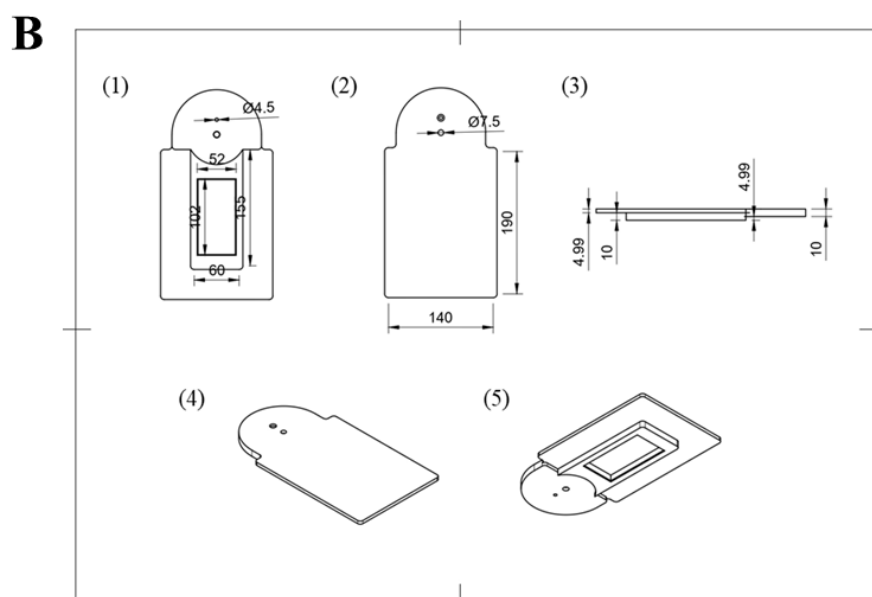
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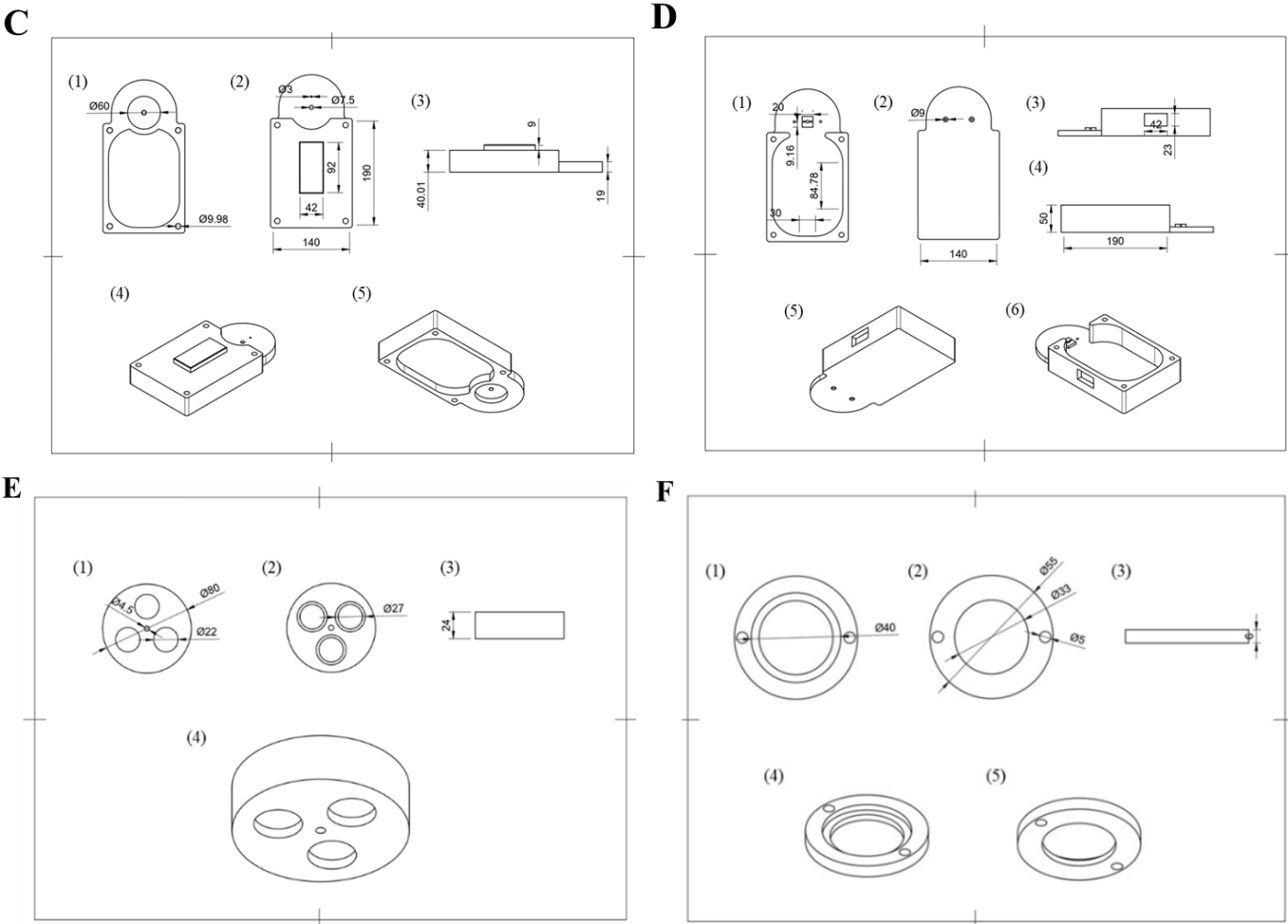
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Additional Figures.





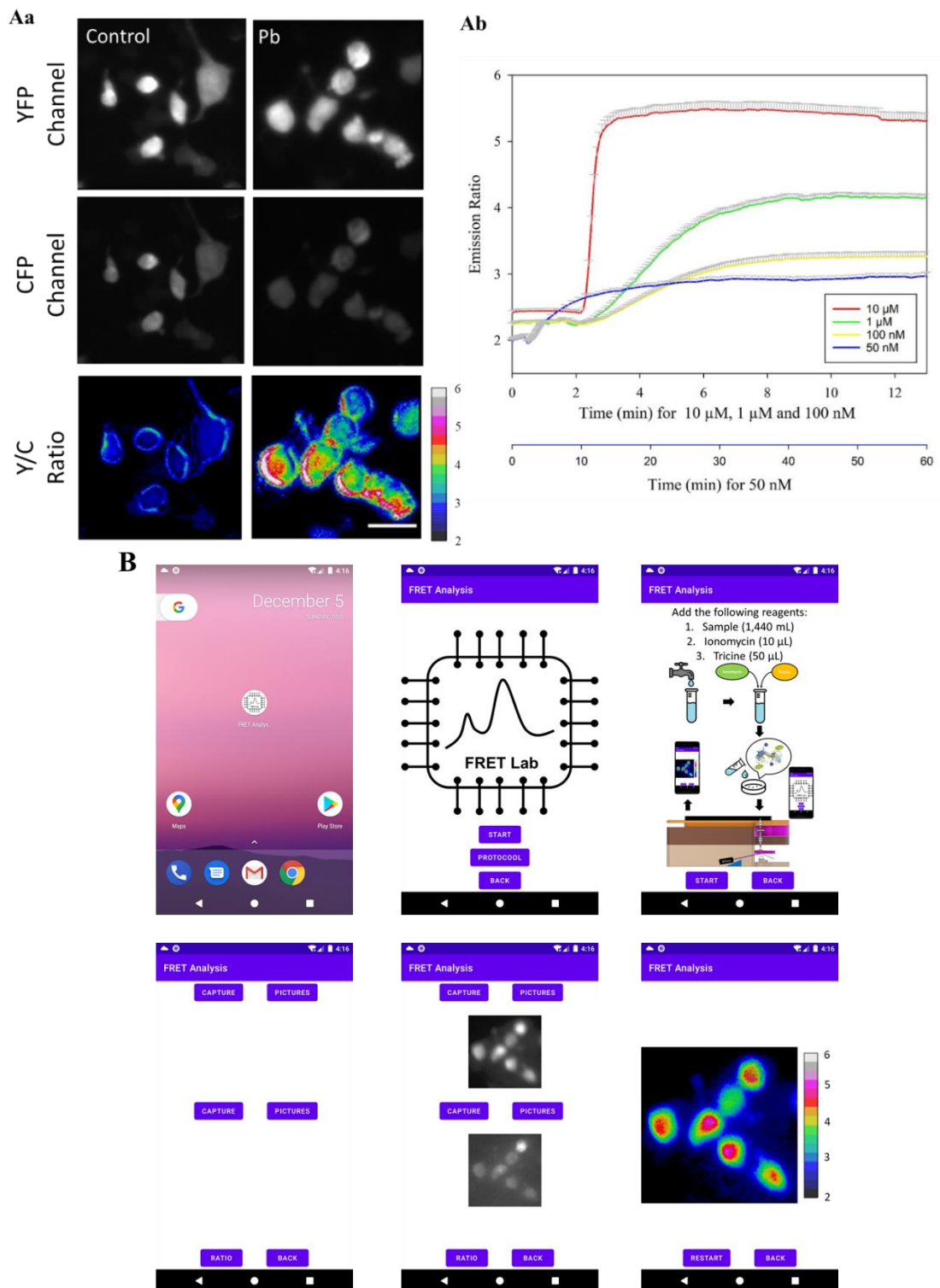
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Figure S1. Design of the smartphone-based Pb sensing device in detail. (A) Outline structure design. (B) Top part design. (C) Middle part design. (D) Bottom part design. (E) Filter turret design. (F) Sample stage design.



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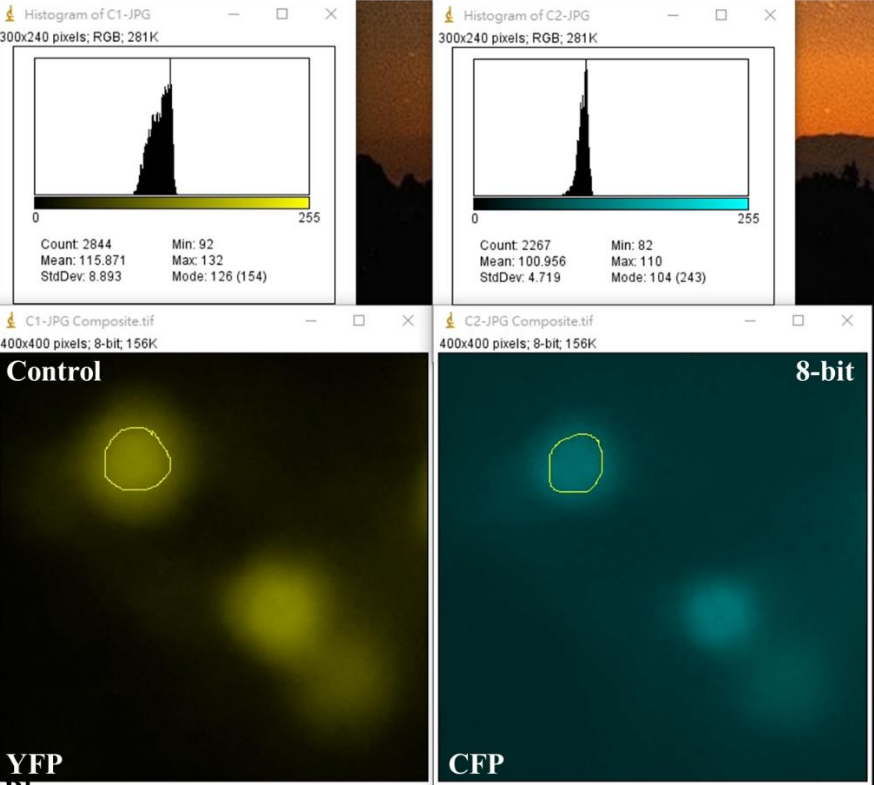
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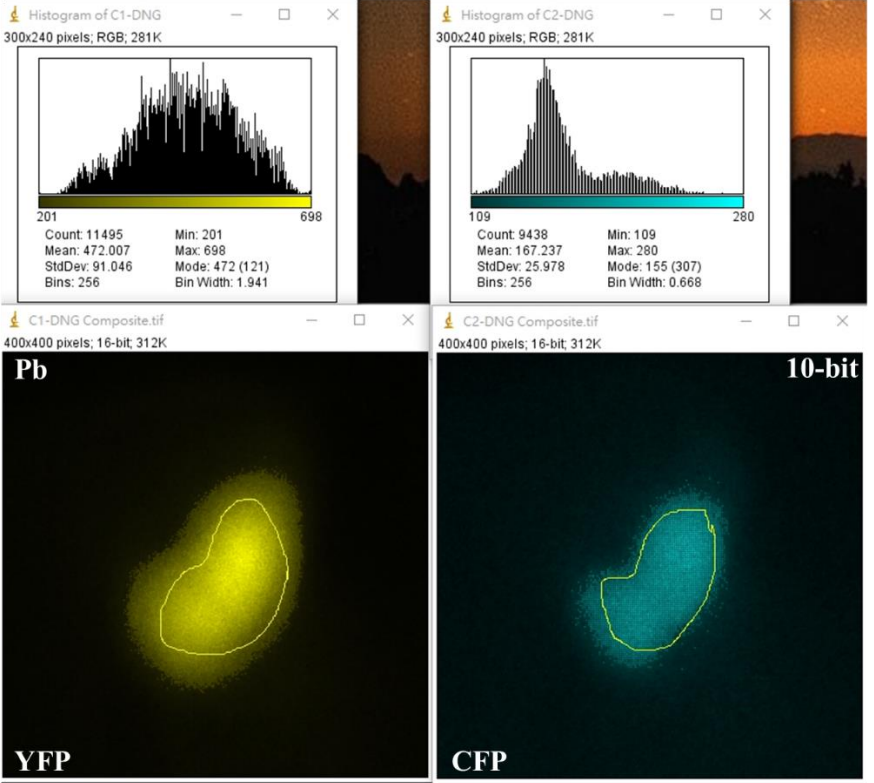
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Figure S2. Sensing function check of Met-lead biochip through conventional FRET microscope and the software for pMet-lead. (A) Functional checking of Met-lead 1.44 M1 under a general FRET ratio microscope with 10 x objectives, representative images are shown in Aa (Control without Pb and Pb with 100 nM or Pb). Various concentrations of Pb (from 50, 100 nM, to 1, 10 μ M) were used under a conventional FRET imaging and shown in Ab. (B) The flowchart of the iMet-lead app for smartphones integrated with the device pMet-lead. **Android Studio** was used through the Java language as an integrated development environment to construct the Android App, iMet-lead. To help users easily proceed Pb sensing with pMet-lead, iMet-lead can preview the operation procedure before sensing analysis. At the stage of preview and the following data acquisition, images of YFP and CFP will be seen on the screen of the smartphone under the interface of iMet-lead. The Ratio bottom can proceed both image acquisition (DNG and JPG format) and image ratio (YFP/CFP, only in DNG format) through smartphone camera and display in color map (the rainbow color plate is the predestined setting). Such kind of visualization can help users to easily judge whether the sample is contaminated with Pb.

Aa

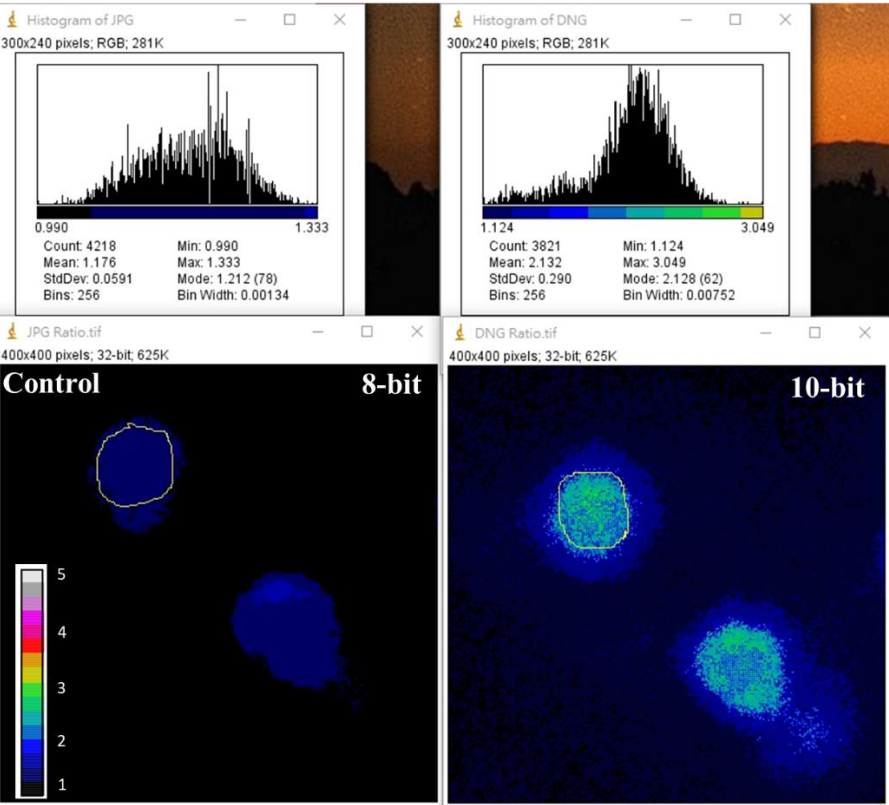


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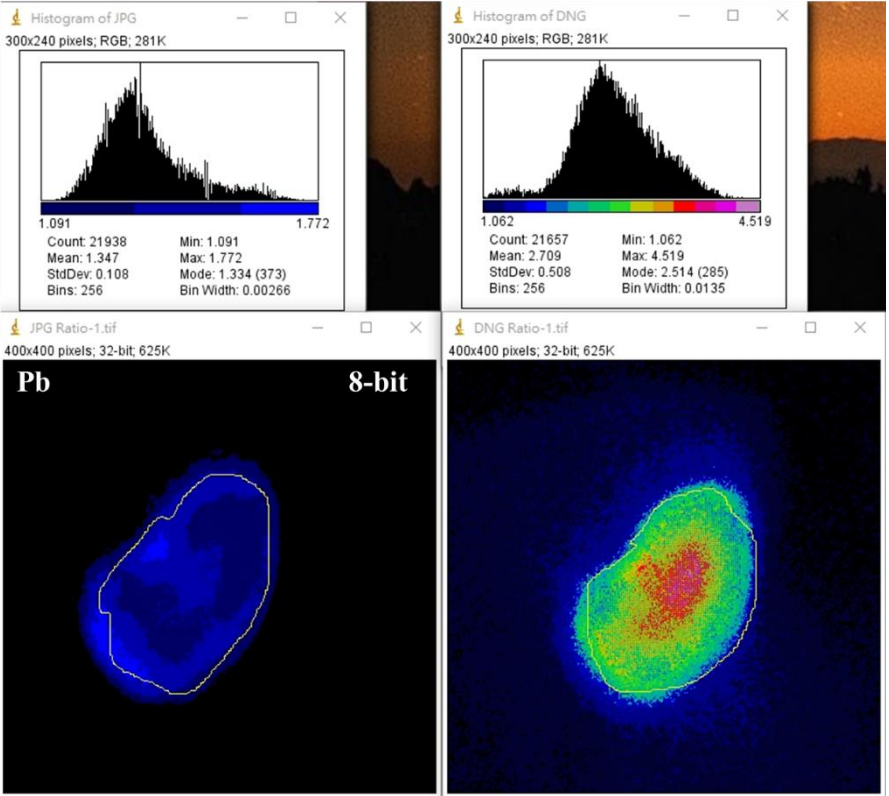
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Ca



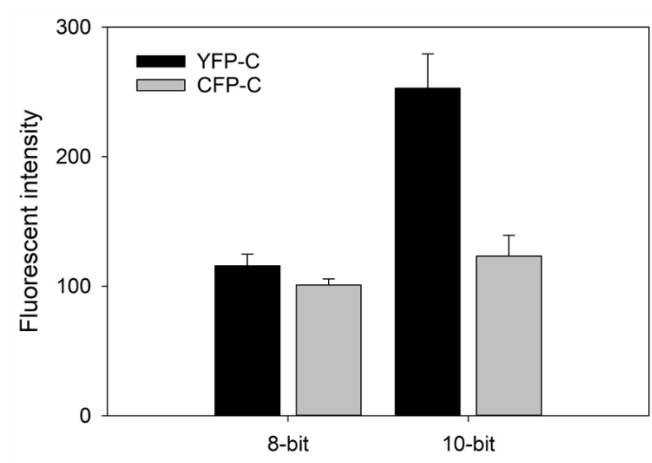
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Cb



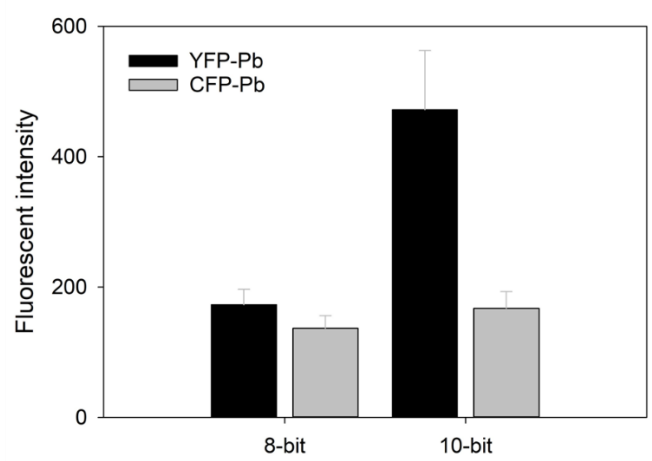
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Da



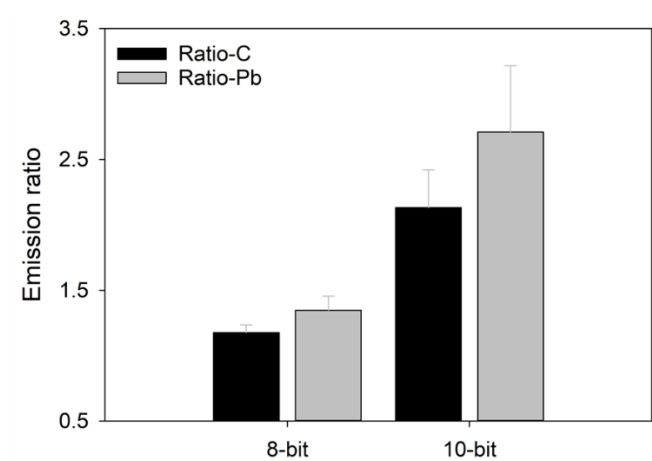
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Db



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Dc



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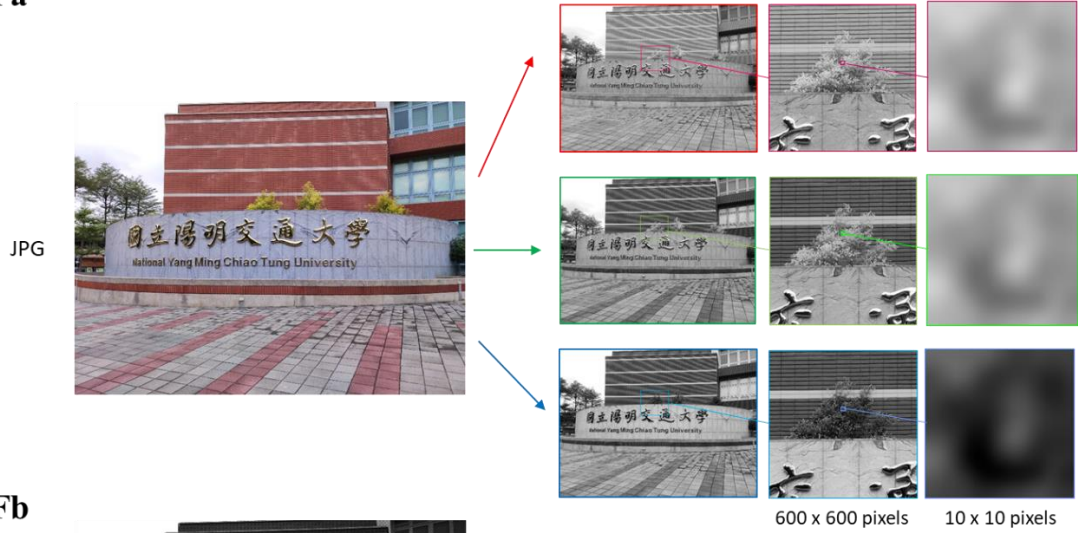
Ea



Eb



Fa



Fb

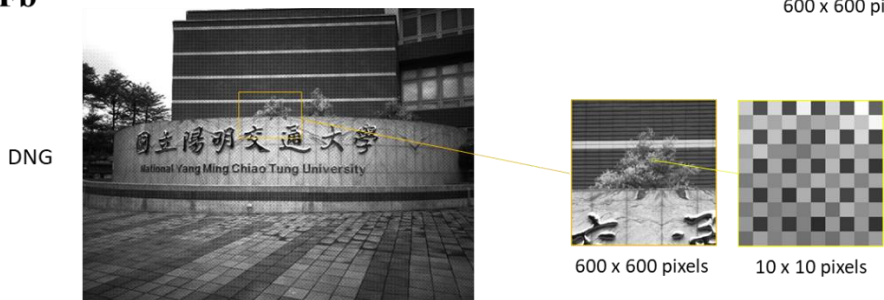


Figure S3. Image depth and image properties taken from smartphones. (A) Image depth at 8 bits in jpg format. (B) Image depth in 10 bits (DNG) format. (C) Ratio images original from 8 bits (A) or 10 bits (B) files. (D) Bar graphs from (A) to (C). (E), (F)- The black spots observed as shown in Figure 3 are not due to pMet-lead.

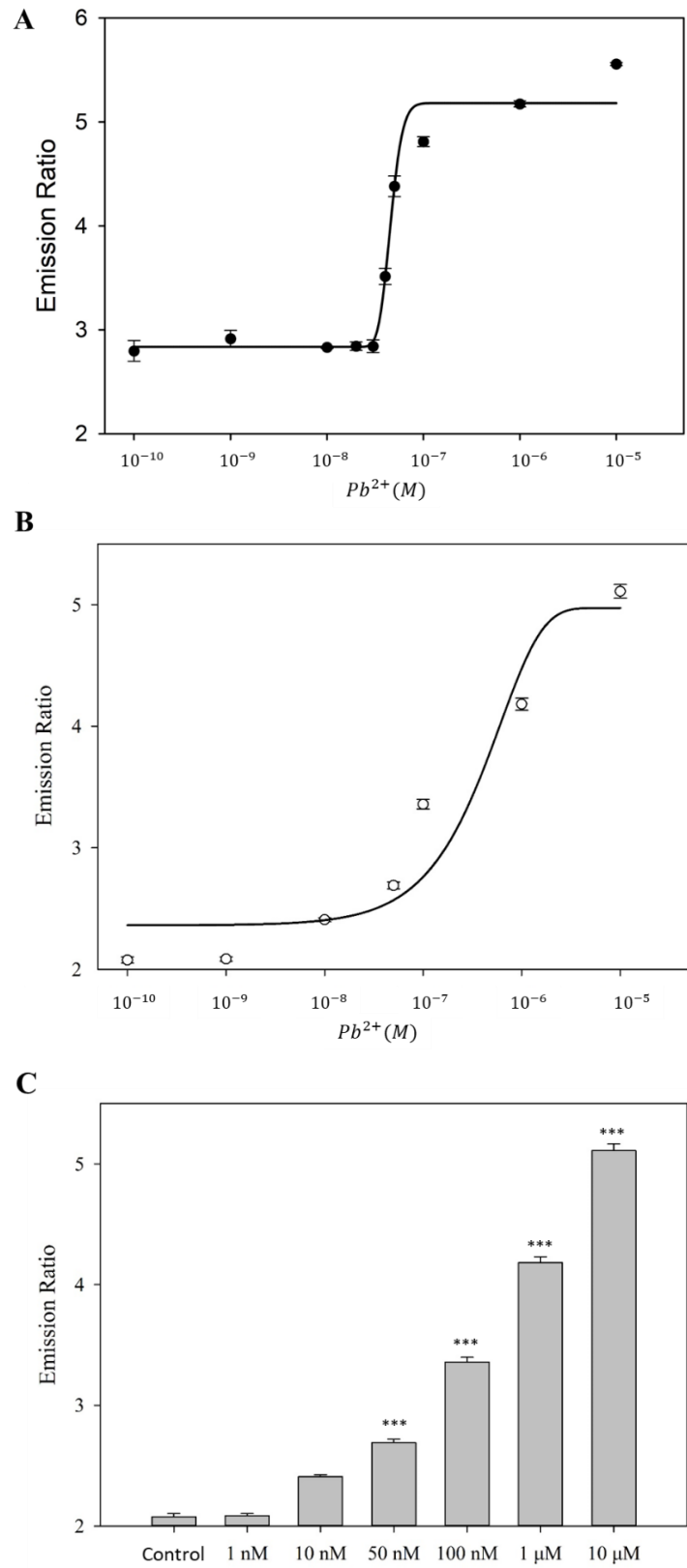
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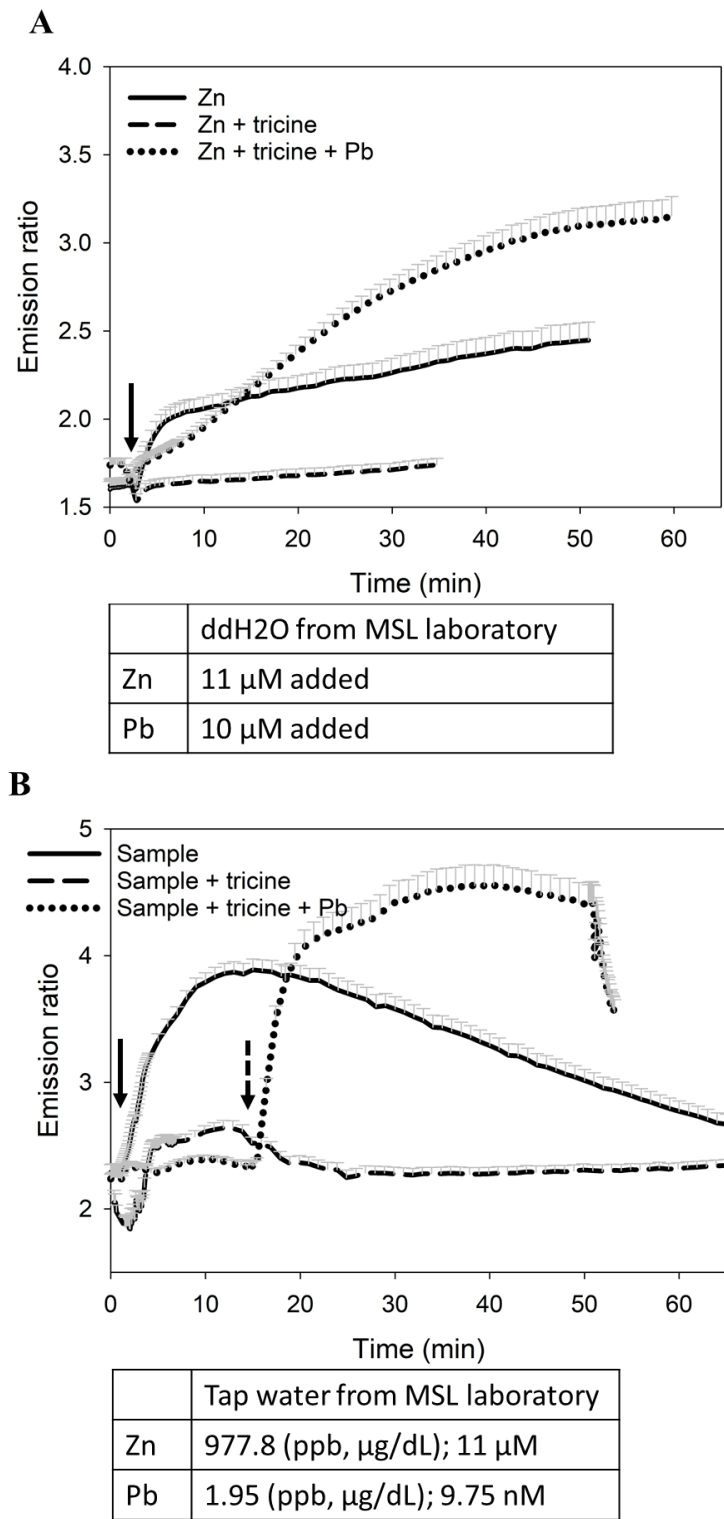
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Figure S4. Sensing ability of pMet-lead and Met-lead 1.44 M1. (A), (B) Titration experiments of Met-lead 1.44 M1 under pMet-lead (A) or under FRET ratio microscope (B). (C) Bar graphs of FRET ratio Pb sensing under various concentrations of Pb through FRET ratio microscope by Met-lead 1.44 M1.



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Figure S5. Full time scale of experiments shown in Figure 4. (A), Time-lapse changes in the Y/C ratio value under the recording of the Met-lead biosensing system. The double-distilled water from the laboratory was pre-mixed with certain concentrations of zinc alone (11 μ M, Zn, in solid line), with additional tricine (10 mM, Zn + tricine, in dash line), or with additional tricine and Pb (10 μ M, Zn + tricine + Pb, in dotted line) introduced at the time point as the arrows indicated. (B) Tap water from the faucet of the laboratory as the sample without (Sample, in solid line) or with tricine (10 mM) pre-mixed (Sample + tricine, in dash line) or with additional Pb (10 μ M, Zn + tricine + Pb, in dotted line) to be added into the Met-lead biosensing system as the arrow indicates. All the experimental sample tests were with ionomycin (5 μ M).

Additional Movies.

Movie S1. Time-lapse FRET ratio color data of Met-lead 1.44 M1 under FRET ratio microscope on pure water when adding certain amounts of zinc (11 μM). The ratio color bar is from 1.1 to 3.5.	57 58 59
Movie S2. Time-lapse FRET ratio color data of Met-lead 1.44 M1 under FRET ratio microscope on pure water when adding certain amounts of zinc (11 μM) pre-incubated with zinc chelator tricine (10 mM). The ratio color bar is from 1.1 to 4.0.	60 61
Movie S3. Time-lapse FRET ratio color data of Met-lead 1.44 M1 under FRET ratio microscope on pure water when adding certain amounts of zinc (11 μM) pre-incubated with zinc chelator tricine (10 mM) and additional treatment with Pb (10 μM). The ratio color bar is from 1.1 to 3.5.	62 63 64
Movie S4. Time-lapse FRET ratio color data of Met-lead 1.44 M1 under FRET ratio microscope on real water sample (tap from laboratory). The ratio color bar is from 1.5 to 3.5.	65 66
Movie S5. Time-lapse FRET ratio color data of Met-lead 1.44 M1 under FRET ratio microscope on real water sample (tap from laboratory) pre-incubated with zinc chelator tricine (10 mM). The ratio color bar is from 1.5 to 3.5.	67 68
Movie S6. Time-lapse FRET ratio color data of Met-lead 1.44 M1 under FRET ratio microscope on real water sample (tap from laboratory) pre-incubated with zinc chelator tricine (10 mM) and additional treatment with Pb (10 μM). The ratio color bar is from 1.5 to 6.0.	69 70
Movie S7. The animated flowchart of the iMet-lead app for smartphones integrated with the device pMet-lead.	71