

# Potassium at the origins of life: Did biology emerge from biotite in micaceous clay?

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## Abstract

Intracellular potassium concentrations,  $[K^+]$ , are high in all types of living cells, but the origins of this  $K^+$  are unknown. The simplest hypothesis is that life emerged in an environment that was high in  $K^+$ . One such environment is the spaces between the sheets of the clay mineral, mica. The best mica for life's origins is the black mica, biotite, because it has a high content of  $Mg^{++}$  and it has iron in various oxidation states. Life also has many of the characteristics of the environment between mica sheets, giving further support for the possibility that mica was the substrate on and within which life emerged.

**Keywords:** clay, mica, biotite, muscovite, origin of life, abiogenesis, mechanical energy, work, wet-dry cycles

## 1. Introduction

All types of living cells have high intracellular potassium concentrations,  $[K^+]$  [1]. When and how did this high  $[K^+]$  appear? There are two options for when high intracellular  $[K^+]$  might have appeared in living systems: before or after the origins of life. The strongest hypothesis is arguably that life originated in a high- $K^+$  environment, because maintaining the  $K^+$  gradient across the cell membrane is energetically expensive [2-5]. The earliest membrane-bound cells would also have had leaky membranes, causing them to be in equilibrium with the extracellular ionic environment [6].

As Morowitz and others have noted, features that are ubiquitous in biology are likely to have evolved early in life's origins [7-9]. This also argues for the origin of life in a high  $[K^+]$  environment. Why does life have intracellular  $K^+$  when it could seemingly use intracellular  $Na^+$  for the same purpose?, Bracher asks [8].

In living systems,  $K^+$ -dependent enzymes are typically intracellular, and  $Na^+$ -dependent enzymes are typically extracellular [10,11]. Ribosomes require  $K^+$  and are essential for life [12]. Many other key cellular processes also require  $K^+$  [13].

Most research on the origins of life ignores potassium  $K^+$  or mentions it only superficially. The question of  $K^+$  at life's origins is, arguably, an elephant in the room of research on the origin of life. Dubina and colleagues propose that life emerged in an environment high in  $[K^+]$  and Dubina has shown that potassium ions are better than sodium ions for polymerizing glutamic acid [14-16]. Bracher's group also has research on the advantages of  $K^+$  at life's origins [8,17]. For example,  $K^+$  stabilizes linear dipeptides against hydrolysis, while  $Na^+$  stabilizes cyclic dipeptides, consistent with the predominance of linear peptides in living systems.

Where was there a prebiotic environment high in  $[K^+]$ ? The ocean is not high in  $[K^+]$ . Concentrations of  $Na^+$ ,  $[Na^+]$ , are 40 times as high as  $[K^+]$  in the ocean. Similarly, river water is not high in  $[K^+]$  [18].

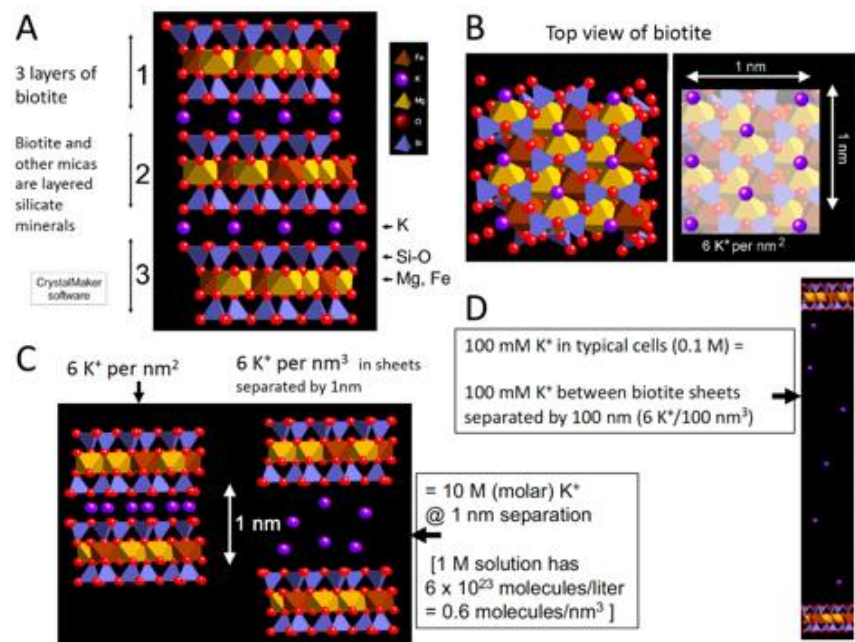
Two main possibilities have been published, for the origins of life in high  $[K^+]$ : in geothermal fields [1] and between the sheets of mica, perhaps in micaceous clay [19,20]. Both of these options might have been true, if micaceous clay was present in geothermal fields. Some advantages of mica are the partial confinement provided by mica sheets and the hexagonal grid of  $K^+$  holding mica's anionic mineral sheets together. This grid has a periodicity of 0.5 nm, which is also the spacing of anionic phosphate groups in extended single-stranded nucleic acids, DNA and RNA.

## 2. $K^+$ between mica sheets.

There are high concentrations of  $K^+$  between mica sheets (Fig. 1). Fig. 1A shows 3 sheets of the black mica, biotite, bridged by  $K^+$  (purple) between adjacent sheets.  $K^+$  are at the sites of partial negative charges from recessed hydroxyl groups on the adjacent sheets.

With an 0.5-nm hexagonal grid of  $K^+$ , there are 6  $K^+$  per  $nm^2$  between pairs of mica sheets (Fig. 1B), giving a concentration of 10 M  $K^+$  when the mica sheets are separated by 1 nm (Fig. 1C). The sheets need to be separated to a distance of 100 nm to give a 100 mM concentration of  $K^+$ , comparable to  $[K^+]$  in living cells (Fig. 1D). 100 mM is

the initial  $[K^+]$  when sheets are separated; the  $[K^+]$  will be decreasing at the edges of the sheets in contact with the external environment, and the  $[K^+]$  will also be increasing on the inside regions of the split mica, where the sheets are separated by less than 100 nm.

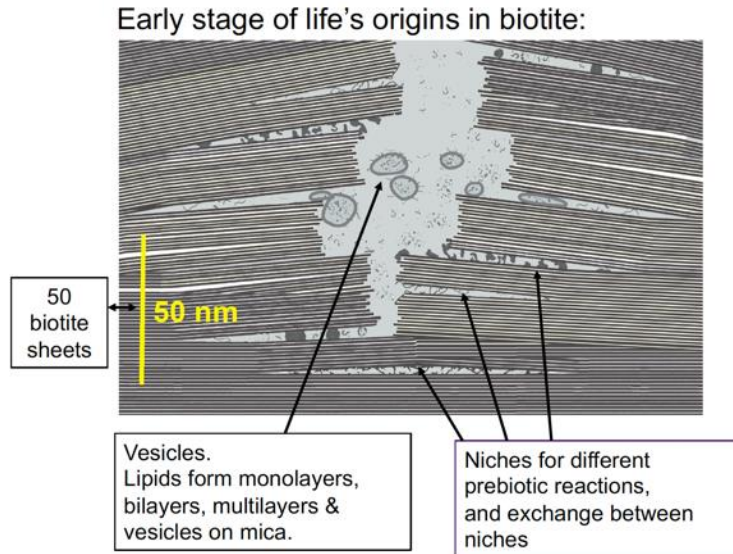


**Figure 1.  $[K^+]$  between mica sheets.** Structure of the black mica, biotite. **(A)** Side view of 3 biotite sheets. **(B)** Top view of 1  $nm^2$  biotite, with  $K^+$  highlighted in the right-hand image, showing that there are 6  $K^+$  per  $nm^2$  between mica sheets. **(C)** Side view of 2 mica sheets, not separated and separated to a distance of 1 nm, where  $[K^+] = 10$  M between the sheets. **(D)** Scale model of biotite sheets at a separation of 100 nm, where  $[K^+] = 100$  mM. (CrystalMaker X software, version 10.6.4, CrystalMaker Software Ltd.)

### 3. A scenario for life's origins between mica sheets.

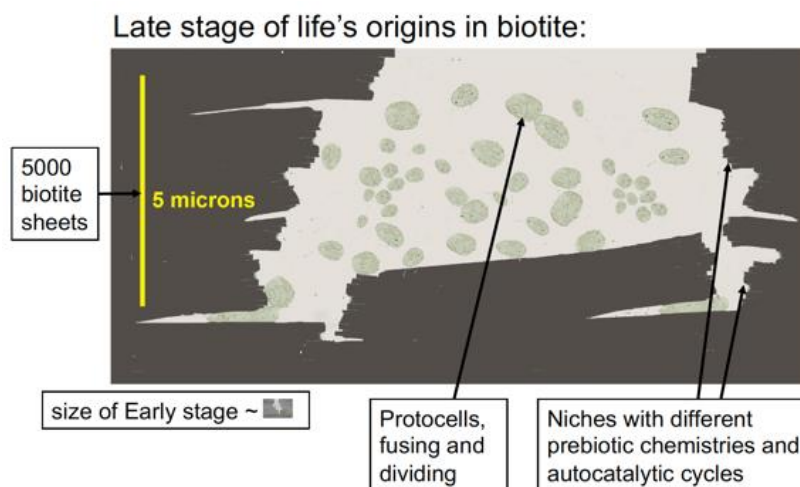
The spaces between mica sheets provide a semi-enclosed environment for life's emergence (Fig. 2). According to de Duve, in the early stages of life's origins, the need for free exchanges would have given an advantage to open systems, due to the constraints of encapsulation [21]. The spaces between mica sheets have the advantages of open exchanges at the outer edges and also the advantages of partial isolation farther within the spaces between the sheets. This might be ideal, for example, for processes such as the evolution of ribozymes, where isolated niches prevent easily replicated ribozymes from dominating the entire population; and interactions such as ligations can occur in other niches and in the space beyond the sheets, to allow ribozymes to change and evolve [19,22,23].

Niches between biotite sheets could also provide spaces where auto-catalytic cycles and proto-metabolic cycles were forming and evolving (e.g., [24-26]). In a beautiful piece of work showing the possibilities of prebiotic syntheses, Muchowska et al. have synthesized 9 of the 11 main components of the TCA cycle, from glyoxylate and pyruvate, with Fe(II), in a test tube at 70°C in only hours [25,27]. Vast numbers of niches exist between mica sheets, providing spaces also for the evolution of genetic coding and ribosomes.



**Figure 2. Nanometer-scale diagram of how the early stages of life might have originated between biotite mica sheets.** Niches within the biotite sheets provide partially enclosed spaces for molecular evolution of different processes essential for life. Vesicles form, encapsulating molecules and molecular complexes from the niches.

Membranes would be forming and encapsulating molecular complexes that were accumulating between the biotite sheets, forming vesicles and protocells. These would tend to aggregate and fuse, bringing together the molecular complexes for metabolism, self-replication, protein synthesis and other necessary processes for life. This would be a slow, gradual, complex process, occurring at many locations in the mica (Fig. 3). After a long long time, membranes would occasionally encapsulate *everything* needed for self-reproducing living cells. Some of these living cells would survive, while others would die after a few generations or more. Life has indeed emerged on Earth, providing conclusive evidence that some living cells survived.



**Figure 3. Micron-scale diagram of how life might have originated between biotite mica sheets.** Protocells in the aqueous environment encapsulate prebiotic molecular aggregates in the niches between mica sheets. Mechanical energy from moving mica sheets can bleb off protocells, as seen in the lower left corner of the figure. Eventually, occasionally, a living cell will be produced, capable of self-reproduction.

Mica is old enough to be a site for the origins of life [28]. Muscovite and biotite are among the major minerals found in zircon grains from the Hadean [29]. Most of the mica would not have been in clays as early as the Hadean, but, as Hazen says, even traces of a mineral could have been sufficient for the mineral to be involved in life's origins [30,31]. Borates, for example, were not present in large quantities at life's origins [28]. Borates, however, are valuable for stabilizing ribose; and even traces of borate on the early Earth might have served this function [32,33]. Similarly, even traces of micaceous clay might have been the site of life's origins on Earth.

Biotite mica has advantages over muscovite mica. Biotite is rich in iron (Fe) and magnesium (Mg). The iron is predominately Fe(II) [34]. Especially in the Hadean reducing environment, Fe(II) predominated over Fe(III). Mg(II) is a major inorganic divalent cation in living systems, where it stabilizes DNA and RNA structures and provides the counterions for ATP, among other things. [Biotite is the most conductive mica, because of its iron content. Electrical conductivity increases exponentially with the iron content of micas [35,36]. Biotite's iron may have been useful for redox reactions [37] at life's origins, in the redox-active and conducting environment of clay [38,39] and the reducing atmosphere of the Hadean [40]. Acid accelerates the dissolution of biotite, acting primarily at step edges of biotite sheets and at etch pits [41]. Biotite is also found on Mars, which may have been the original source of life in the Solar System, seeding life on earth [42].

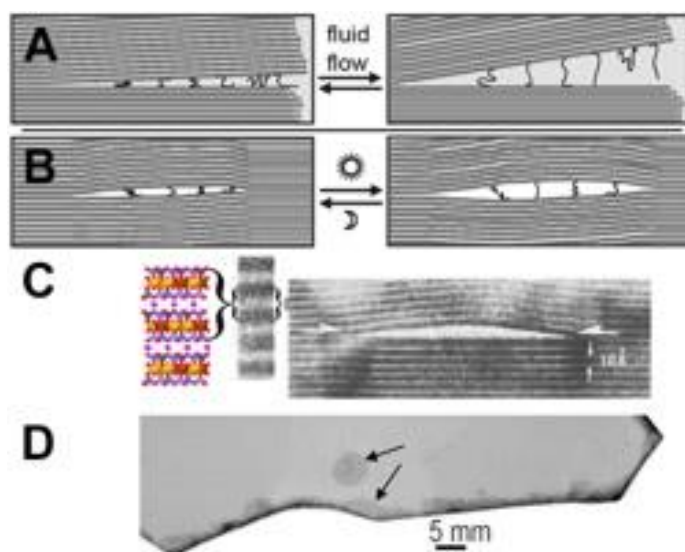
#### 4. Energy from mica

**4.1 Mechanical energy.** (Fig. 4) Moving mica sheets can produce endless energy for the origins of life [43]. This energy of moving mica sheets is mechanical energy that can be used for mechanochemistry, to make and break chemical bonds, when stacks of mica sheets move, open and shut. Mechanochemistry is a growing research field, in which biomolecules are synthesized with mechanical forces [44]. Mechanochemistry has been used in possible prebiotic syntheses [45,46] and nucleobase pairing [47]. Glycine polymerizes by mechanochemistry in mica, by ball milling [48].

Both mica sheets and enzymes have open and shut motions that do work on the molecules between them. As the title of a recent article says, 'Enzymes at work are enzymes in motion' [49]. Experimental results support this statement [50,51]. Further evidence for the importance of mechanical energy in biology is that *Molecular Biology of the Cell* is soliciting submissions for its Sixth Special Issue on Forces on and Within Cells" [52].

How much energy can moving mica sheets provide? If the mica sheets move even 0.1 nanometer (nm) closer together, in air, they can push together 2 molecules to form a covalent bond, if the mica has a spring constant stiff enough to provide 170 piconewtons (pN) of force [19]. The equation for a spring constant,  $F=kx$ , with  $x = 0.1$  nm, shows that a spring constant ( $k$ ) of 1.7 N/m (Newtons/meter) is stiff enough. The spring constant of the mica depends on the number of mica sheets in the layer that is moving open and shut. Each mica sheet is ~1 nm thick. Only about 7 mica sheets are needed to provide this spring constant, in air [53]. In practice, the layers of moving mica sheets will often have thickness of microns, not nanometers, due to the fragility and consequent damage to nanometers-thick layers of mica sheets.

Mechanochemical polymerizations can create oligomers and longer polymers that can bind to the mica surface more strongly than monomers and short oligomers. Monomers and short oligomers can be preferentially washed off the mica sheets, favoring polymerization by mechanochemistry over polymer breakdown.



**Figure 4. Mica and Mechanical energy.** **A.** Diagram of mechanical forces between biotite mica sheets, stretching and compressing polymers, due to water flow at the edges of the biotite sheets. **B.** Diagram of mechanical forces between biotite mica sheets due to heat pumps in a biotite bubble. This mechanical energy can be used to synthesize prebiotic molecules, stretch and compress polymers (as shown in the diagram), or bleb off protocells [19]. Seven mica sheets, as shown in **A**, provides enough force to form a covalent bond in air, when moved a distance of 0.1 nm. **C.** Biotite bubble imaged by HRTEM (high-resolution transmission electron microscopy) [54], with expanded view of HRTEM image and CrystalMaker model of biotite on left. “{“ or “}” = 2 biotite layers. The thickness of a single biotite sheet is 1 nm (10 Angstroms). **D.** Photograph of muscovite mica, showing a bubble (upper arrow) and separation at the edges of the mica sheet (lower arrow). Bubbles are common even in ‘high grade’ micas such as this one.

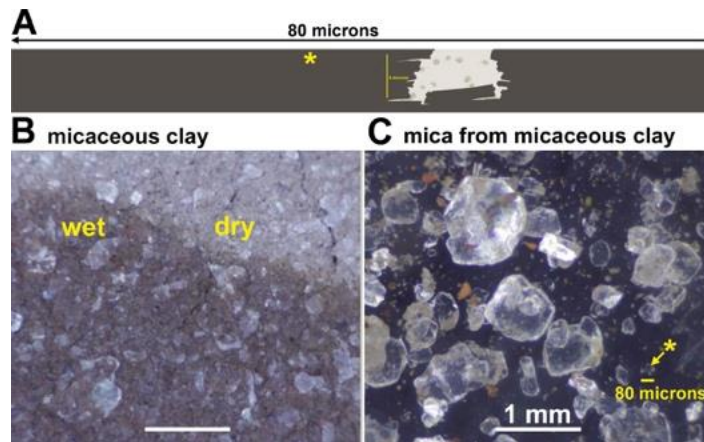
**4.2 Wet-dry cycles.** Entropy drives polymerization during wet-dry cycles [55]. Polymers of amino acids and nucleotides form by dehydration, but polymers hydrolyze in the presence of water [56-59]. In mica, wet-dry cycles occur at split edges of mica sheets, as in Fig. 4A and 4D (lower arrow). The slow wet-dry cycles that can occur at the edges of mica sheets will generate longer polymers during the longer drying cycles, before hydrolysis occurs during the wet phase. These longer polymers will bind to mica better than short polymers and will, consequently, be more likely to remain bound to the surface. This is seen in Atomic Force Microscopy (AFM), where long DNA molecules bind to mica strongly enough for AFM imaging, but short DNA molecules do not bind to mica well enough to be imaged [60]. In contrast, rapid wet-dry cycles in small clay particles will cause polymerization to be followed more quickly by depolymerization, resulting in shorter polymers that will more easily detach from the surface.

How much water was on the Hadean Earth? The question has been reviewed recently, giving evidence that water may have covered up to 80% of Earth in the Hadean and more in the Archaean [61].

## 5. How big do the mica sheets need to be?

One needs only tiny pieces of mica for mechanochemistry. The mica fragments in micaceous clays are large enough. The ‘mica world’ diagram in Fig. 3 is lengthened, in Fig. 5A, to show that even sub-millimeter-sized mica fragments are big enough to generate mechanical energy for life’s emergence. A submillimeter mica fragment of the same length is highlighted in Fig. 5C. Therefore, life may have emerged in micaceous clay, as opposed to larger pieces of mica. The swelling clay particles surrounding the mica fragments would also be advantageous for life’s emergence. For example, polymer syntheses might occur during the drying phase of wet-dry cycles in clay, and solutes in the surrounding fluid would be concentrated during drying. Some of these molecules and other solutes would move between the mica sheets.



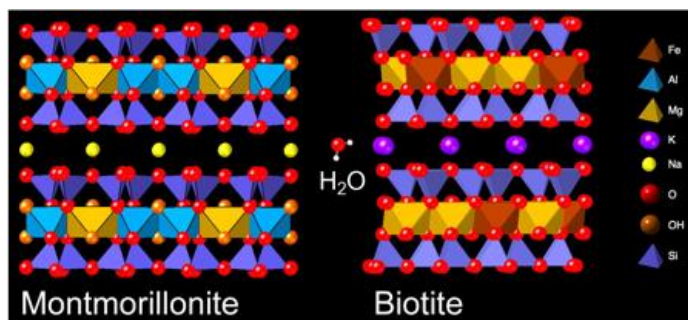


**Figure 5. Micaceous clay and the origin of life between mica sheets.** **A.** Mica origins diagram of Fig. 3, a late stage in the origin of life, extended to a length of 80 microns. **B.** Mica Red Micaceous Clay, from New Mexico Clay Store, containing pale reflecting pieces of mica, in the middle of a wet-dry cycle. **C.** Mica and a few clay particles, washed from the micaceous clay. Yellow asterisk and arrow point to a mica fragment with a diameter of ~80 microns.

## 6. Origin of life in micaceous clay?

Micas are non-swelling clay minerals, unlike smectite clays such as montmorillonite (Fig. 6), which swells and shrinks during wet-dry cycles [62]. Typical clay particles, ~1-2 microns in size, are also much smaller than mica particles. The swelling and shrinking of these clays is as if they were tiny sandwiches whose filling were growing thicker and thinner, and wetter and dryer. Non-swelling mica provides a more stable environment for life's origins than swelling clays. Wet-dry cycles do occur, however, at the split edges of mica sheets, where water seeps slowly in and out (Fig. 4D), leaving dry and nearly dry regions beyond the wet edges. Experimentally, water seeped a few millimeters between the sheets of mica pieces that were cycled daily between 22°C and 4°C for 2 weeks [19].

Why does montmorillonite clay swell, while micas do not swell? Anhydrous  $K^+$  is larger than anhydrous  $Na^+$  (Figure 6 caption). The larger ions of  $K^+$  fill the spaces at the recessed hydroxyls between mica sheets, while the smaller ions of  $Na^+$  are hydrated at the recessed hydroxyls between the sheets of clays such as montmorillonite.



**Figure 6. Swelling clay [montmorillonite] and non-swelling clay [biotite].** Molecular models showing 2 sheets of montmorillonite clay (left) and biotite mica (right); water molecule (center). Surfaces of the sheets are tetrahedral silicon-oxygen (Si-O) layers; see Fig. 1B for top view. A major difference between these clays is that  $Na^+$  [yellow] bridges sheets of montmorillonite and  $K^+$  [purple] bridges sheets of biotite. Ionic radii are 0.095 nm for  $Na^+$  and 0.133 nm for  $K^+$  [63]. The smaller  $Na^+$  are hydrated between montmorillonite sheets, which causes montmorillonite to swell and shrink with wetting and drying. The larger  $K^+$  between biotite sheets are not hydrated; biotite does not swell and shrink with wetting and drying. (CrystalMaker X software, version 10.6.4, CrystalMaker Software Ltd.)

Why clay? Hyman Hartman explains it thus: “The genetic code drives all biological life. But even a mechanism this fundamental rests on still more ancient biochemical processes, as well as the intriguing chemical properties of a seemingly nondescript material—clay. ... Formed through the reaction of silicates with water, clay minerals have layered crystal structures that provide ideal surfaces for molecules to bind to and interact with each other in close proximity. In fact, we have long used these very properties of clay to speed up chemical reactions in oil refineries and in the catalytic converters found in cars.” [64].

Clay mineral surfaces catalyze or support syntheses of amino acids from simple precursors and polymerizations of amino acids and nucleotides into oligopeptides and oligonucleotides, e.g., [57,65-69]. Nucleotides on clay polymerize preferentially in the 3’-5’ orientation, as in life, and not in the non-biological 2’-5’ orientation [70]. A Molecular Dynamics study [71] of montmorillonite clay indicates that polymerization in the 3’-5’ direction occurs fastest in clay sheets that are closer together, compared with the non-biological 2’-5’ direction, which occurs fastest at greater separations of the clay sheets. The mineral sheets in mica are more often close together than the mineral sheets in clay, which swells and shrinks. Polymerizations occur preferentially in the clay interlayer, as opposed to the edges of the sheets. Homochiral polymerizations are favored over achiral polymerizations on clay. Clays form in association with the water needed for life [72], which is another advantage of clay over other rocks and minerals. The mica in micaceous clay might have been the site where life originated.

7. Biology and biotite

Biotite and other micas have similarities with life, as would be expected for places where life might have originated (Table 1) [19,73].

Table 1. Characteristics of life and mica.

Life:	Mica:
Cellular compartments	Compartments between mica sheets
High intracellular potassium, [K <sup>+</sup> ]	High [K <sup>+</sup> ]; potassium ions bridge mica sheets
Nucleotides polymerize to DNA & RNA	Nucleotides polymerize to RNA in wet/dry cycles [74]
0.5 nm spacing of anionic phosphates in ssDNA	0.5 nm anionic crystal lattice on mica surface
Exchangeable inorganic cations bridge anionic sites on molecules such as DNA	Exchangeable inorganic cations bridge anionic sites between mica sheets
Water-rich; aqueous	Hydrophilic
Forms H-bonds	Forms H-bonds [75]
Mechanical energy of enzyme motion <sup>1</sup>	Mechanical energy from moving mica sheets <sup>1</sup>
Synthesis of biomolecules in confined spaces	Supports chemistry of confinement
Filled and covered with lipid membranes	Supports lipid membranes & vesicles

<sup>1</sup>The mechanical energy of enzymes is powered by chemical energy, primarily ATP. The mechanical energy of moving mica sheets is powered primarily by thermal disequilibria.

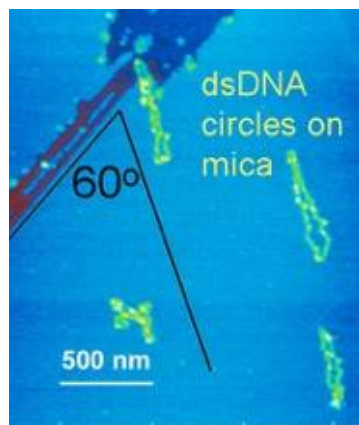
**7.1 RNA and DNA on mica.** RNA polymers form spontaneously on mica [74]. RNA monomers polymerize, non-enzymatically, on a mica surface during wet-dry cycles. Nucleotide monophosphates of Adenine (A), Guanine (G), Cytosine (C) and Uracil (U) on mica were cycled through wet-dry cycles at 80°C and imaged by Atomic Force Microscopy (AFM) [74]. This simple process, with no enzymes or activated nucleotides, produced RNA on bare mica. RNA lengths were ~100-1000 nucleotides, which is about an order of magnitude longer than the RNA lengths obtained when polymerization occurred in the presence of lipids, without mica [76]. It is reasonable that mica’s anionic crystal lattice is a better substrate than lipids, for polymerizing RNA, because RNA has the same periodicity – 0.5 nm – as mica’s crystal lattice.

Mica may have been a template for RNA polymerization at life’s origins. Perhaps nucleic acid linkages are 3’-5’ and not 2’-5’ because mica sheets served as a template that favored 3’-5’ linkages. Perhaps nucleotide templating on mica’s crystal lattice prevented diphosphate linkages, which form a bent polymer, and other irregularities of nucleotide polymerization.

DNA binds reversibly to mica in the presence of various divalent inorganic cations. For example, freshly cleaved mica was soaked in 33 mM magnesium acetate to bind DNA to mica for early AFM imaging (Fig. 7) [77,78]. With AFM in aqueous fluid, stable DNA imaging on mica was observed when Ni<sup>++</sup>, Co<sup>++</sup>, and Zn<sup>++</sup> salts were present; in contrast, DNA binding was not strong enough for AFM imaging when salts of Mn<sup>++</sup>, Cd<sup>++</sup>, Hg<sup>++</sup>, or

$K^+$  were used [79]. DNA transcription by RNA polymerase was observed by AFM when  $Zn^{++}$  was alternately added, to bind the DNA to mica, and removed, to allow polymerase activity [80].

If polymers that have an affinity for a mineral surface, longer polymers will be more firmly bound to the surface than shorter polymers, facilitating the accumulation of long polymers [81]. This has been observed for AFM of DNA [60].



**Figure 7. Atomic Force Microscopy of circular double-stranded DNA (dsDNA) on mica with cracks.** (Cracks are dark streaks at the upper left). Three of the 4 dsDNA circles form a 60-degree angle with the mica crack, consistent with alignment on mica's hexagonal crystal lattice.

**7.2 Sugars.** Sugars, especially ribose, are a major biomolecule in living systems. A plausible prebiotic reaction for forming sugars is the formose reaction, in which formaldehyde reacts to form sugars [82]. In a test tube, the end products become increasingly large polymers of sugars, branched sugar polymers, and eventually a tarry mess. Monosaccharides, especially ribose for RNA, are a desired product, at the origins of life [83]. The spacing of sugars in oligosaccharides is 0.5 nm, like the periodicity of the mica lattice.

If the formose reaction is tightly confined between mica sheets, simpler sugars might predominate. Mica's anionic hexagonal lattice may also favor linear oligosaccharides over branched or bent ones. The formose reaction produces a simple sugar when the reactants are confined in vesicles [84]. This is an example of the advantage that confinement gives for limiting the products of the formose reaction.

**7.3 Peptides.** Peptide amyloids are proposed, in an intriguing new hypothesis for the functional molecules at life's origins [85]. Beta-sheets are the simple linear structure for peptides that can form amyloids, which form into stable ordered structures capable of various catalytic activities.

Peptide bond formation varies in difficulty, depending on whether the reactants are amino acids or peptides. Bond formation is easiest when it forms between 2 polypeptides. Intermediate in difficulty is bond formation between an amino acid and a polypeptide. Bond formation is most difficult between 2 amino acids [86]. This favors the formation of longer polypeptides at life's origins.

**7.4 Membranes and the origins of life.** Membranes on mica have been observed by Atomic Force Microscopy [87-89]. Vesicles on mica fuse to form extended bilayers and multilayers. Even without lipids, however, mica sheets could have provided partially enclosed spaces for emerging life, before the molecules of emerging life were enclosed in membranes. Membranes can be fragile. They leak, acquire and lose molecules, swell, and rupture. Membranes of living cells are highly evolved structures that provide more extensive support, and selective permeabilities, for their contents than primitive vesicles and membranes.

Lipid membranes might not have been essential at the early stages of the origins of life. Root-Bernstein et al. say that the evolution of membranes would be a late development, in their paper about 'prebiotic ecology' [90]. An 'ecosystems first' perspective is proposed by Baum and others, based on their intriguing research involving chemical selection on mineral surfaces [91].



Perhaps, instead of membranes, protolife evolved as an acellular ecosystem, sharing all the necessary enzymes in an open system. Imagine pieces of this ecosystem periodically encapsulating in membranes. Nearly all of these membrane-encapsulated protocells would lack some essential component of life or enzyme. Occasionally, membrane-encapsulated protocells would contain all the essential components of life and became alive. Occasionally some of these living protocells would reproduce successfully and begin seeding Earth with Life.

On the other hand, there is also a school of thought in which membranes are the enclosed spaces where proto-life first evolved, e.g., [92].

**7.5 Coacervates and Membraneless organelles.** There is an increasingly popular alternative to membranes at the origins of life – ‘membraneless organelles’ or ‘membraneless biomolecular condensates’ also known as ‘coacervates’. Peptides/proteins and RNA interact in membraneless organelles in living cells, as nucleoli and other particles [93-95]. Nucleoli, the membraneless organelles in cell nuclei where ribosomes are formed, are now known to contain other membraneless organelles inside them [96,97]. Membraneless organelles form by liquid-in-liquid phase separation (LLPS) [98]. Membraneless organelles are increasingly of interest to origins-of-life researchers [99-106].

Ribosomes are ancient biomolecular condensates, composed of proteins and RNA, and are now necessary for translating nucleic acids into proteins. Ribosomes were present in the Last Universal Common Ancestor of life (LUCA) [107]. When life was coming into being, in the pre-LUCA stages, ribosomes and their precursors may have been early ‘membraneless organelles’, protected within mica sheets [101]. Prokaryotic ribosomes are ~20 nm in diameter, comparable to the thickness of 20 mica sheets (see Figs. 1 and 2) and much smaller than the 100-nm separation of mica sheets at which the  $[K^+]$  is 100 mM (Fig. 1D).

## 8. Dielectric constant at surfaces

The Dielectric constant, or permittivity, of water is 80 for bulk water but only ~2 for the first 2 or 3 water layers above a surface (~2 nm) [108,109]. This means that the charges on charged molecules will become progressively unscreened as the charged molecules approach the mica surface. Electrostatic forces will be stronger, resulting in stronger interactions between charged organic molecules, the anionic mica surfaces, and inorganic cations.

## 9. Crowding at the origins of life

“There is a growing consensus that confinement may have facilitated the transformation of inanimate matter into living organisms” [110]. Confinement exists within compartments of various sizes between the sheets of mica; these compartments may have provided confined spaces for the isolation and stabilization of supramolecular assemblies and protocells.

Molecules in cells are crowded. The space between protein molecules in cells is typically only 10 nm [111]. Crowding speeds up the rates of reactions that are diffusion limited [112]. Crowding may even be the origin of homochirality [113]. Given the molecular crowding in living cells, molecular crowding at life’s origins is a desirable scenario for hypotheses about the origins of life. Molecules in wet-dry cycles become crowded during the drying phase. Molecules that bind to a surface, e.g., the mica surface, will become concentrated and crowded. Molecules in narrow spaces between mica sheets will typically be crowded, by the mica sheets above and below, in addition to crowding by other molecules. Clays will also crowd molecules between their sheets, but clay’s swelling will then dilute molecules. Swelling to even 2 layers of water molecules between Na-montmorillonite clay sheets reduces the interaction energy between sheets to near zero, according to molecular modeling [114]. Thermophoresis is another way to concentrate molecules, in a spatially confined thermal gradient, and even to escalate nucleotide polymerization [115,116].

Confinement chemistry would occur between mica’s sheets during drying and during the compression stage of mechanochemistry. Chemistry in confined spaces produces fewer different molecules and simpler molecules [117,118]. Confined spaces also help proteins fold [119,120]. Enzymes confine their substrates to facilitate the enzymatic reactions. Zeolites mimic enzymes in some respects [121]. Nano-confined liquids have very different properties from bulk liquids [122]. Crowding is also proposed to have enhanced evolutionary capabilities through the networks created by the proximity of components in crowded environments [123]. Confinement chemistry is likely to be a characteristic of any good hypothesis for the origins of life.

## 10. Hierarchy, Complexification, and Error Tolerance

Herbert Simon describes the need for a hierarchy of structures in abiogenesis. [124] He uses the watchmaker as an example. The watchmaker needs stable intermediates in the watchmaking process. If no intermediates were stable, the partially assembled watch would disassemble whenever the watchmaker set it down – to answer the phone, for example, for getting a new order for a watch, to use Herbert Simon's example. Stable intermediates in abiogenesis can assemble on the hexagonal 0.5-nm anionic grids on and between the 'ceilings' and 'floors' of the spaces between mica sheets. As Pross and Paschal say, complexification is also an often-ignored but necessary aspect of abiogenesis [125]. Molecular interactions with mica sheets will stabilize intermediates and enable further complexifications.

Freeman Dyson says error tolerance is essential for life's origins [126]. With the redundancy of the vast areas between biotite mica sheets, in micaceous clay, almost everything can go wrong, and life can still emerge. If not from micaceous clay, life emerged from some other habitat with vast error tolerance.

## 11. Conclusions

Somewhere in the universe there was a hospitable habitat, with everything needed for the origins of life. We know this because life now exists on Earth. The habitat may have been on Earth, ~4 billion years ago [127]. In this habitat, the components and processes of life were evolving, resulting, eventually, in LUCA, a Last Universal Common Ancestor [128,129], which may have been a primitive acellular community of some type [90,91,130,131]. Among these components and processes, RNA-peptide 'worlds' [132-140] were evolving to create and replicate genetic information [141]; proto-metabolic cycles were evolving into early metabolism [142,143], and ribosomes were evolving to synthesize proteins [144,145]. This may have occurred in 'lipid worlds' [76,146,147]; hydrothermal vents have also been proposed [148-151]. Vast numbers of interconnections were needed, for bringing the precursors to Earth or synthesizing them on Earth and bringing them, eventually, to the hospitable habitat from which 'biology' emerged. The habitat was likely clay [152,153], the stuff of life.

Biology may have emerged from the spaces between biotite sheets in micaceous clay. The spaces between the sheets in these mica 'books' might have been ancient pre-cellular habitats where prebiotic molecules were confined, concentrated, and synthesized, before membrane-bound cells emerged (Fig. 2) [19,43,73,101,113,154]. These pre-cellular spaces have an anionic crystal lattice, with 'ceilings' and 'floors' that could have templated molecular syntheses and polymerizations. The mineral sheets of biotite and muscovite micas have a layered silicate structure, like montmorillonite and other clays, which have been used successfully to catalyze reactions non-enzymatically, such as the synthesis of biopolymers. [65-67,155]. Mica sheets move, open and shut, at their edges, as fluids flow and temperature changes; they are thus in a constant state of the thermodynamic non-equilibrium necessary for life.

We will never know for certain whether this, or any origins hypothesis, is completely true. The experimental method traditionally starts with a hypothesis, followed by experimentally testing the hypothesis. Testable hypotheses are presented here. Experimental results will show what is possible today, but these results may, in fact, be false positives or false negatives. The origins of life are partly an ahistorical science in which much will remain hypothetical, both experiments and ideas [125]. Experimental results give us ideas about how life might have originated, but they cannot absolutely prove how life originated, though strong experimental results are seen as convincing evidence [27].

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## References:

1. Mulkidjanian, A.Y.; Bychkov, A.Y.; Dibrova, D.V.; Galperin, M.Y.; Koonin, E.V. Origin of first cells at terrestrial, anoxic geothermal fields. *Proc Natl Acad Sci U S A* **2012**, *109*, E821-830.
2. Mulkidjanian, A.Y.; Dibrov, P.; Galperin, M.Y. The past and present of sodium energetics: may the sodium-motive force be with you. *Biochim Biophys Acta* **2008**, *1777*, 985-992.
3. Faller, L.D. Mechanistic studies of sodium pump. *Archives of Biochemistry and Biophysics* **2008**, *476*, 12-21.
4. Kay, A.R.; Blaustein, M.P. Evolution of our understanding of cell volume regulation by the pump-leak mechanism. *Journal of General Physiology* **2019**, *151*, 407-416.
5. Stautz, J.; Hellmich, Y.; Fuss, M.F.; Silberberg, J.M.; Devlin, J.R.; Stockbridge, R.B.; Hänel, I. Molecular mechanisms for bacterial potassium homeostasis. *Journal of Molecular Biology* **2021**, *433*, 166968.
6. Koonin, E.V.; Mulkidjanian, A.Y. Evolution of cell division: from shear mechanics to complex molecular machineries. *Cell* **2013**, *152*, 942-944.
7. Brunk, C.F.; Marshall, C.R. 'Whole Organism', Systems Biology, and Top-Down Criteria for Evaluating Scenarios for the Origin of Life. *Life* **2021**, *11*, 690.
8. Campbell, T.D.; Hart, C.A.; Febrian, R.; Cheneler, M.L.; Bracher, P.J. The opposite effect of K<sup>+</sup> and Na<sup>+</sup> on the hydrolysis of linear and cyclic dipeptides. *Tetrahedron Letters* **2018**, *59*, 2264-2267.
9. Morowitz, H.J. A theory of biochemical organization, metabolic pathways, and evolution. *Complexity* **1999**, *4*, 39-53.
10. Clausen, M.J.V.; Poulsen, H. Sodium/potassium homeostasis in the cell. In *Metallomics and the Cell*; Springer: 2013; pp. 41-67.
11. Page, M.J.; Di Cera, E. Role of Na<sup>+</sup> and K<sup>+</sup> in enzyme function. *Physiological reviews* **2006**, *86*, 1049-1092.
12. Rozov, A.; Khusainov, I.; El Omari, K.; Duman, R.; Mykhaylyk, V.; Yusupov, M.; Westhof, E.; Wagner, A.; Yusupova, G. Importance of potassium ions for ribosome structure and function revealed by long-wavelength X-ray diffraction. *Nature communications* **2019**, *10*, 1-12.
13. Dibrova, D.; Galperin, M.Y.; Koonin, E.; Mulkidjanian, A. Ancient systems of sodium/potassium homeostasis as predecessors of membrane bioenergetics. *Biochemistry (Moscow)* **2015**, *80*, 495-516.
14. Natchin, Y.V. The physiological evolution of animals: sodium is the clue to resolving contradictions. *Herald of the Russian Academy of Sciences* **2007**, *77*, 581-591.
15. Natchin, Y.V. The origin of membranes. *Paleontological Journal* **2010**, *44*, 860-869.
16. Dubina, M.V.; Vyazmin, S.Y.; Boitsov, V.M.; Nikolaev, E.N.; Popov, I.A.; Kononikhin, A.S.; Eliseev, I.E.; Natchin, Y.V. Potassium ions are more effective than sodium ions in salt induced peptide formation. *Origins of Life and Evolution of Biospheres* **2013**, *43*, 109-117.
17. Campbell, T.D.; Febrian, R.; McCarthy, J.T.; Kleinschmidt, H.E.; Forsythe, J.G.; Bracher, P.J. Prebiotic condensation through wet-dry cycling regulated by deliquescence. *Nature communications* **2019**, *10*, 1-7.
18. Maurer, S.E.; Nguyen, G. Prebiotic vesicle formation and the necessity of salts. *Origins of Life and Evolution of Biospheres* **2016**, *46*, 215-222.
19. Hansma, H.G. Possible origin of life between mica sheets. *Journal of Theoretical Biology* **2010**, *266*, 175-188.
20. Hansma, H.G. Did Biology Emerge from Biotite in Micaceous Clay? *Preprints* **2020**.
21. De Duve, C. *Vital Dust: The Origin and Evolution of life on earth*; Basic Books: 1995.
22. Joyce, G.F. Molecular evolution: booting up life. *Nature* **2002**, *420*, 278-279, doi:10.1038/420278a
23. Szabo, P.; Scheuring, I.; Czaran, T.; Szathmary, E. In silico simulations reveal that replicators with limited dispersal evolve towards higher efficiency and fidelity. *Nature* **2002**, *420*, 340-343.
24. Smith, E.; Morowitz, H.J. *The origin and nature of life on earth: the emergence of the fourth geosphere*; Cambridge University Press: 2016.
25. Muchowska, K.B.; Varma, S.J.; Moran, J. Synthesis and breakdown of universal metabolic precursors promoted by iron. *Nature* **2019**, *569*, 104-107.
26. Camprubi, E.; Jordan, S.F.; Vasiliadou, R.; Lane, N. Iron catalysis at the origin of life. *IUBMB life* **2017**, *69*, 373-381.
27. Pascal, R. A possible non-biological reaction framework for metabolic processes on early Earth. **2019**.
28. Hazen, R.M.; Papineau, D.; Bleeker, W.; Downs, R.T.; Ferry, J.M.; McCoy, T.J.; Sverjensky, D.A.; Yang, H. Mineral Evolution. *American Mineralogist* **2008**, *93*, 1693-1720

29. Papineau, D. Mineral environments on the earliest Earth. *Elements* **2010**, *6*, 25-30.
30. Morrison, S.M.; Runyon, S.E.; Hazen, R.M. The Paleomineralogy of the Hadean Eon Revisited. *Life* **2018**, *8*, 64.
31. Hazen, R.M.; Sverjensky, D.A.; Azzolini, D.; Bish, D.L.; Elmore, S.C.; Hinnov, L.; Milliken, R.E. Clay mineral evolution. *American Mineralogist* **2013**, *98*, 2007-2029.
32. Benner, S.A.; Ricardo, A.; Carrigan, M.A. Is there a common chemical model for life in the universe? *Current Opinion in Chemical Biology* **2004**, *8*.
33. Benner, S.A. Prebiotic plausibility and networks of paradox-resolving independent models. *Nature communications* **2018**, *9*, 1-3.
34. Shane, P.; Smith, V.; Nairn, I. Biotite composition as a tool for the identification of Quaternary tephra beds. *Quaternary Research* **2003**, *59*, 262-270.
35. Crine, J.; Friedmann, A.; Wertheimer, M.; Yelon, A. The relationship between chemical composition and electrical conductivity of some North American micas. *Canadian Journal of Physics* **1977**, *55*, 270-275.
36. Meunier, M.; Currie, J.; Wertheimer, M.; Yelon, A. Electrical conduction in biotite micas. *Journal of applied physics* **1983**, *54*, 898-905.
37. Bouda, S.; Isaac, K. Influence of soil redox conditions on oxidation of biotite. *Clay Minerals* **1986**, *21*, 149-157.
38. Gorski, C.A.; Aeschbacher, M.; Soltermann, D.; Voegelin, A.; Baeyens, B.; Marques Fernandes, M.; Hofstetter, T.B.; Sander, M. Redox properties of structural Fe in clay minerals. 1. Electrochemical quantification of electron-donating and-accepting capacities of smectites. *Environmental science & technology* **2012**, *46*, 9360-9368.
39. Xiang, Y.; Villemure, G. Electrodes modified with synthetic clay minerals: Evidence of direct electron transfer from structural iron sites in the clay lattice. *Journal of Electroanalytical Chemistry* **1995**, *381*, 21-27.
40. Zahnle, K.J.; Lupu, R.; Catling, D.C.; Wogan, N. Creation and Evolution of Impact-generated Reduced Atmospheres of Early Earth. *The Planetary Science Journal* **2020**, *1*, 11.
41. Li, J.; Zhang, W.; Li, S.; Li, X.; Lu, J. Effects of citrate on the dissolution and transformation of biotite, analyzed by chemical and atomic force microscopy. *Applied geochemistry* **2014**, *51*, 101-108.
42. Bridges, J.C.; Warren, P.H. The SNC meteorites: basaltic igneous processes on Mars. *Journal of the Geological Society* **2006**, *163*, 229-251.
43. Hansma, H.G. Mechanical Energy before Chemical Energy at the Origins of Life? *Sci* **2020**, *2*, 19.
44. Wang, G.-W. Mechanochemical organic synthesis. *Chemical Society Reviews* **2013**, *42*, 7668-7700.
45. Lamour, S.; Pallmann, S.; Haas, M.; Trapp, O. Prebiotic Sugar Formation Under Nonaqueous Conditions and Mechanochemical Acceleration. *Life* **2019**, *9*, 52.
46. Bolm, C.; Mocci, R.; Schumacher, C.; Turberg, M.; Puccetti, F.; Hernández, J.G. Mechanochemical Activation of Iron Cyano Complexes: A Prebiotic Impact Scenario for the Synthesis of  $\alpha$ -Amino Acid Derivatives. *Angewandte Chemie* **2018**, *130*, 2447-2450.
47. Stolar, T.; Lukin, S.; Etter, M.; Linarić, M.R.; Užarević, K.; Meštrović, E.; Halasz, I. DNA-specific selectivity in pairing of model nucleobases in the solid state. *Chemical Communications* **2020**.
48. Stolar, T.; Grubešić, S.; Cindro, N.; Meštrović, E.; Užarević, K.; Hernández, J.G. Mechanochemical Prebiotic Peptide Bond Formation. **2020**, doi:DOI:10.26434/chemrxiv.13187852.v1.
49. Saleh, T.; Kalodimos, C.G. Enzymes at work are enzymes in motion. *Science* **2017**, *355*, 247-248.
50. Benkovic, S.J.; Hammes-Schiffer, S. A perspective on enzyme catalysis. *Science* **2003**, *301*, 1196-1202.
51. Kim, T.H.; Mehrabi, P.; Ren, Z.; Sljoka, A.; Ing, C.; Bezginov, A.; Ye, L.; Pomès, R.; Prosser, R.S.; Pai, E.F. The role of dimer asymmetry and protomer dynamics in enzyme catalysis. *Science* **2017**, *355*.
52. MBoC Sixth Special Issue on Forces On and Within Cells. Available online: <https://www.ascb.org/publications-columns/mboc-sixth-special-issue-on-forces-on-and-within-cells/> (accessed on
53. Castellanos-Gomez, A.; Poot, M.; Amor-Amorós, A.; Steele, G.A.; van der Zant, H.S.; Agraït, N.; Rubio-Bollinger, G. Mechanical properties of freely suspended atomically thin dielectric layers of mica. *Nano Research* **2012**, *5*, 550-557.
54. Banos, J.O.; Amouric, M.; De Fouquet, C.; Baronnet, A. Interlayering and interlayer slip in biotite as seen by HRTEM. *American Mineralogist* **1983**, *68*, 754-758.
55. Ross, D.; Deamer, D. Prebiotic Oligomer Assembly: What Was the Energy Source? *Astrobiology* **2019**, *19*, 517-521.

56. Cafferty, B.J.; Hud, N.V. Abiotic synthesis of RNA in water: a common goal of prebiotic chemistry and bottom-up synthetic biology. *Current opinion in chemical biology* **2014**, *22*, 146-157.
57. Lahav, N.; White, D.; Chang, S. Peptide formation in the prebiotic era: thermal condensation of glycine in fluctuating clay environments. *Science* **1978**, *201*, 67-69, doi:10.1126/science.663639.
58. Grover, M.A.; He, C.Y.; Hsieh, M.-C.; Yu, S.-S. A chemical engineering perspective on the origins of life. *Processes* **2015**, *3*, 309-338.
59. Benner, S.A.; Kim, H.-J.; Carrigan, M.A. Asphalt, water, and the prebiotic synthesis of ribose, ribonucleosides, and RNA. *Accounts of chemical research* **2012**, *45*, 2025-2034.
60. Hansma, H.; Revenko, I.; Kim, K.; Laney, D. Atomic force microscopy of long and short double-stranded, single-stranded and triple-stranded nucleic acids. *Nucl. Acids Res.* **1996**, *24*, 713-720, doi:10.1093/nar/24.4.713.
61. Korenaga, J. Was There Land on the Early Earth? *Life* **2021**, *11*, 1142.
62. Stucki, J.W.; Lee, K.; Zhang, L.; Larson, R.A. Effects of iron oxidation states on the surface and structural properties of smectites. *Pure and Applied Chemistry* **2002**, *74*, 2145-2158.
63. Israelachvili, J.N. *Intermolecular and surface forces*; Academic press: 2011.
64. Hartman, H. FROM CLAY TO THE CODE OF LIFE. In *Sydney Brenner's 10-on-10: The Chronicles of Evolution*, Shuzhen Sim, B.S., Ed.; World Scientific Publishing Company: 2018; pp. 33-44.
65. Ferris, J.P.; Aubrey R. Hill, J.; Liu, R.; Orgel, L.E. Synthesis of long prebiotic oligomers on mineral surfaces. *Nature* **1996**, *381*, 59-61.
66. Ferris, J.P.; Ertem, G. Montmorillonite catalysis of RNA oligomer formation in aqueous solution. A model for the prebiotic formation of RNA. *Journal of the American Chemical Society* **1993**, *115*, 12270-12275, doi:10.1021/ja00079a006.
67. Joshi, P.C.; Aldersley, M.F.; Delano, J.W.; Ferris, J.P. Mechanism of Montmorillonite Catalysis in the Formation of RNA Oligomers. *Journal of the American Chemical Society* **2009**, *131*, 13369-13374, doi:10.1021/ja9036516.
68. Bu, H.; Yuan, P.; Liu, H.; Liu, D.; Qin, Z.; Zhong, X.; Song, H.; Li, Y. Formation of macromolecules with peptide bonds via the thermal evolution of amino acids in the presence of montmorillonite: Insight into prebiotic geochemistry on the early Earth. *Chemical Geology* **2019**, *510*, 72-83.
69. Hashizume, H. Role of clay minerals in chemical evolution and the origins of life. *Clay Minerals in Nature—Their characterization, modification and application* **2012**.
70. Huang, W.; Ferris, J.P. One-Step, Regioselective Synthesis of up to 50-mers of RNA Oligomers by Montmorillonite Catalysis. *Journal of the American Chemical Society* **2006**, *128*, 8914-8919, doi:10.1021/ja061782k.
71. Mathew, D.; Luthey-Schulten, Z. Influence of Montmorillonite on Nucleotide Oligomerization Reactions: A Molecular Dynamics Study. *Origins of Life and Evolution of Biospheres* **2010**, *40*, 303-317, doi:10.1007/s11084-010-9207-0.
72. Westall, F. Life on the early Earth: a sedimentary view. *Science* **2005**, *308*, 366-367.
73. Hansma, H.G. Possible Origin of Life between Mica Sheets: Does Life Imitate Mica? *J. Biol. Struct. Dynamics* **2013**, *31*, 888-895.
74. Hassenkam, T.; Damer, B.; Mednick, G.; Deamer, D. Viroid-sized rings self-assemble from mononucleotides through wet-dry cycling: implications for the origin of life. *Life* **2020**, *10*, 321.
75. Yu, J.; Kan, Y.; Rapp, M.; Danner, E.; Wei, W.; Das, S.; Miller, D.R.; Chen, Y.; Waite, J.H.; Israelachvili, J.N. Adaptive hydrophobic and hydrophilic interactions of mussel foot proteins with organic thin films. *Proceedings of the National Academy of Sciences* **2013**, *110*, 15680-15685.
76. Rajamani, S.; Vlassov, A.; Benner, S.; Coombs, A.; Olasagasti, F.; Deamer, D. Lipid-assisted Synthesis of RNA-like Polymers from Mononucleotides. *Origins of Life and Evolution of Biospheres* **2008**, *38*, 57-74.
77. Hansma, H.G.; Sinsheimer, R.L.; Li, M.Q.; Hansma, P.K. Atomic force microscopy of single- and double-stranded DNA. *Nucleic Acids Res* **1992**, *20*, 3585-3590.
78. Hansma, H.G.; Vesenska, J.; Siegerist, C.; Kelderman, G.; Morrett, H.; Sinsheimer, R.L.; Bustamante, C.; Elings, V.; Hansma, P.K. Reproducible Imaging and Dissection of Plasmid DNA under Liquid with the Atomic Force Microscope. *Science* **1992**, *256*, 1180-1184.
79. Hansma, H.G.; Laney, D.E. DNA binding to mica correlates with cationic radius: assay by atomic force microscopy. *Biophys. J.* **1996**, *70*, 1933-1939.
80. Kasas, S.; Thomson, N.H.; Smith, B.L.; Hansma, H.G.; Zhu, X.; Guthold, M.; Bustamante, C.; Kool, E.T.; Kashlev, M.; Hansma, P.K. *E. coli* RNA polymerase activity observed using atomic force microscopy. *Biochemistry* **1997**, *36*, 461-468.



81. Orgel, L.E. Polymerization on the rocks: theoretical introduction. *Origins of Life and Evolution of the Biosphere* **1998**, 28, 227-234.
82. Lambert, J.B.; Gurusamy-Thangavelu, S.A.; Ma, K. The silicate-mediated formose reaction: bottom-up synthesis of sugar silicates. *Science* **2010**, 327, 984-986, doi:327/5968/984 [pii] 10.1126/science.1182669.
83. Delidovich, I.V.; Simonov, A.N.; Taran, O.P.; Parmon, V.N. Catalytic formation of monosaccharides: From the formose reaction towards selective synthesis. *ChemSusChem* **2014**, 7, 1833-1846.
84. Cooper, G.J.; Cronin, L. How to sweet-talk bacteria. *Nature chemistry* **2009**, 1, 342-343.
85. Greenwald, J.; Kwiatkowski, W.; Riek, R. Peptide amyloids in the origin of life. *Journal of molecular biology* **2018**, 430, 3735-3750.
86. Martin, R.B. Free energies and equilibria of peptide bond hydrolysis and formation. *Biopolymers* **1998**, 45, 351-353.
87. Dufrêne, Y.F.; Lee, G.U. Advances in the characterization of supported lipid films with the atomic force microscope. *Biochimica et Biophysica Acta (BBA)-Biomembranes* **2000**, 1509, 14-41.
88. Hansma, H.G.; Hoh, J. Biomolecular imaging with the atomic force microscope. *Annual Review of Biophysics and Biomolecular Structure* **1994**, 23, 115-139.
89. Weisenhorn, A.L.; Egger, M.; Ohnesorge, F.; Gould, S.A.C.; Heyn, S.-P.; Hansma, H.G.; Sinsheimer, R.L.; Gaub, H.E.; Hansma, P.K. Molecular-Resolution Images of Langmuir-Blodgett Films and DNA by Atomic Force Microscopy. *Langmuir* **1991**, 7, 8-12.
90. Hunding, A.; Kepes, F.; Lancet, D.; Minsky, A.; Norris, V.; Raine, D.; Sriram, K.; Root-Bernstein, R. Compositional complementarity and prebiotic ecology in the origin of life. *Bioessays* **2006**, 28, 399-412, doi:10.1002/bies.20389.
91. Vincent, L.; Berg, M.; Krismer, M.; Saghafi, S.T.; Cosby, J.; Sankari, T.; Vetsigian, K.; Cleaves, H.J.; Baum, D.A. Chemical Ecosystem Selection on Mineral Surfaces Reveals Long-Term Dynamics Consistent with the Spontaneous Emergence of Mutual Catalysis. *Life* **2019**, 9, 80.
92. Van Kranendonk, M.J.; Deamer, D.W.; Djokic, T. Life springs. *Scientific American* **2017**, 317, 28-35.
93. Hyman, A.A.; Weber, C.A.; Jülicher, F. Liquid-liquid phase separation in biology. *Annual review of cell and developmental biology* **2014**, 30, 39-58.
94. Marko, J.F. The liquid drop nature of nucleoli. *Nucleus* **2012**, 3, 115-117.
95. Weber, S.C.; Brangwynne, C.P. Getting RNA and protein in phase. *Cell* **2012**, 149, 1188-1191.
96. Lafontaine, D.L.; Riback, J.A.; Bascetin, R.; Brangwynne, C.P. The nucleolus as a multiphase liquid condensate. *Nature Reviews Molecular Cell Biology* **2021**, 22, 165-182.
97. Mountain, G.A.; Keating, C.D. Chapter Five - Practical considerations for generation of multi-compartment complex coacervates. In *Methods in Enzymology*, Keating, C.D., Ed.; Academic Press: 2021; Volume 646, pp. 115-142.
98. Brangwynne, C.P. Phase transitions and size scaling of membrane-less organelles. *The Journal of cell biology* **2013**, 203, 875-881.
99. Jia, T.Z.; Wang, P.-H.; Niwa, T.; Mamajanov, I. Connecting primitive phase separation to biotechnology, synthetic biology, and engineering. *Journal of biosciences* **2021**, 46, 1-28.
100. Hansma, H.G. Liquid-Liquid Phase Separation at the Origins of Life. In *Droplets of Life: Membrane-Less Organelles, Biomolecular Condensates, and Biological Liquid-Liquid Phase Separation*, Uversky, V., Ed.; Elsevier: 2022; p. in press.
101. Hansma, H.G. Better than Membranes at the Origin of Life? *Life* **2017**, 7, 28.
102. Cakmak, F.P.; Keating, C.D. Combining catalytic microparticles with droplets formed by phase coexistence: Adsorption and activity of natural clays at the aqueous/aqueous interface. *Scientific reports* **2017**, 7, 1-14.
103. Tena-Solsona, M.; Wanzke, C.; Riess, B.; Bausch, A.R.; Boekhoven, J. Self-selection of dissipative assemblies driven by primitive chemical reaction networks. *Nature communications* **2018**, 9, 1-8.
104. Poudyal, R.R.; Guth-Metzler, R.M.; Veenis, A.J.; Frankel, E.A.; Keating, C.D.; Bevilacqua, P.C. Template-directed RNA polymerization and enhanced ribozyme catalysis inside membraneless compartments formed by coacervates. *Nature communications* **2019**, 10, 1-13.
105. Drobot, B.; Iglesias-Artola, J.M.; Le Vay, K.; Mayr, V.; Kar, M.; Kreysing, M.; Mutschler, H.; Tang, T.D. Compartmentalised RNA catalysis in membrane-free coacervate protocells. *Nature communications* **2018**, 9, 1-9.

106. Jia, T.Z.; Chandru, K.; Hongo, Y.; Afrin, R.; Usui, T.; Myojo, K.; Cleaves, H.J. Membraneless polyester microdroplets as primordial compartments at the origins of life. *Proceedings of the National Academy of Sciences* **2019**, *116*, 15830-15835.
107. Hsiao, C.; Mohan, S.; Kalahar, B.K.; Williams, L.D. Peeling the onion: ribosomes are ancient molecular fossils. *Molecular Biology and Evolution* **2009**, *26*, 2415-2425.
108. Fumagalli, L.; Esfandiar, A.; Fabregas, R.; Hu, S.; Ares, P.; Janardanan, A.; Yang, Q.; Radha, B.; Taniguchi, T.; Watanabe, K.; et al. Anomalous low dielectric constant of confined water. *Science* **2018**, *360*, 1339-1342, doi:10.1126/science.aat4191.
109. Kalinin, S.V. Feel the dielectric force. *Science* **2018**, *360*, 1302-1302, doi:10.1126/science.aat9875.
110. Grommet, A.B.; Feller, M.; Klajn, R. Chemical reactivity under nanoconfinement. *Nature Nanotechnology* **2020**, 1-16.
111. Phillips, R.; Kondev, J.; Theriot, J. *Physical biology of the cell*; Garland Science: New York, 2008.
112. Ross, D.S.; Deamer, D. Dry/wet cycling and the thermodynamics and kinetics of prebiotic polymer synthesis. *Life* **2016**, *6*, 28.
113. Hansma, H.G. The Power of Crowding for the Origins of Life. *Origins of Life and Evolution of Biospheres* **2014**, *44*, 307-311.
114. Pradhan, S.M.; Katti, K.S.; Katti, D.R. Evolution of molecular interactions in the interlayer of Na-montmorillonite swelling clay with increasing hydration. *International Journal of Geomechanics* **2015**, *15*, 04014073.
115. Duhr, S.; Braun, D. Why molecules move along a temperature gradient. *Proceedings of the National Academy of Sciences* **2006**, *103*, 19678-19682.
116. Mast, C.B.; Schink, S.; Gerland, U.; Braun, D. Escalation of polymerization in a thermal gradient. *Proceedings of the National Academy of Sciences* **2013**, *110*, 8030-8035.
117. Sozzani, P.; Bracco, S.; Comotti, A.; Simonutti, R.; Valsesia, P.; Sakamoto, Y.; Terasaki, O. Complete shape retention in the transformation of silica to polymer micro-objects. *Nature materials* **2006**, *5*, 545-551.
118. Sozzani, P.; Behling, R.W.; Schilling, F.C.; Briickner, S.; Helfand, E.; Bovey, F.A.; Jelinski, L.W. Traveling Defects in 1,4-trans-Polybutadiene as an Inclusion Complex in Perhydrotriphenylene Canals and a Comparison with Molecular Motions in the Crystalline Solid State. *Macromolecules* **1989**, *22*, 3318-3322.
119. Thirumalai, D.; Klimov, D.K.; Lorimer, G.H. Caging helps proteins fold. *Proc Natl Acad Sci U S A* **2003**, *100*, 11195-11197, doi:10.1073/pnas.2035072100
- 2035072100 [pii].
120. Klimov, D.K.; Thirumalai, D. Dissecting the assembly of Abeta16-22 amyloid peptides into antiparallel beta sheets. *Structure* **2003**, *11*, 295-307, doi:10.1016/S0969212603000315 [pii].
121. Turro, N.J. From boiling stones to smart crystals: supramolecular and magnetic isotope control of radical-radical reactions in zeolites. *Accounts of Chemical Research* **2000**, *33*, 637-646.
122. Munoz-Santiburcio, D.; Marx, D. Chemistry in nanoconfined water. *Chemical science* **2017**, *8*, 3444-3452.
123. Saha, R.; Pohorille, A.; Chen, I.A. Molecular crowding and early evolution. *Origins of Life and Evolution of Biospheres* **2014**, *44*, 319-324.
124. Herbert, S. The architecture of complexity. *Proceedings of the American Philosophical Society* **1962**, *106*, 467-482.
125. Pross, A.; Pascal, R. The origin of life: what we know, what we can know and what we will never know. *Open biology* **2013**, *3*, 120190.
126. Dyson, F.J. *Origins of life*, Rev. ed.; Cambridge University Press: Cambridge [England] ; New York, 1999; pp. ix, 100 p.
127. Kipping, D. An objective Bayesian analysis of life's early start and our late arrival. *Proceedings of the National Academy of Sciences* **2020**, *117*, 11995-12003.
128. Weiss, M.C.; Sousa, F.L.; Mrnjavac, N.; Neukirchen, S.; Roettger, M.; Nelson-Sathi, S.; Martin, W.F. The physiology and habitat of the last universal common ancestor. *Nature microbiology* **2016**, *1*, 1-8.
129. Krupovic, M.; Dolja, V.V.; Koonin, E.V. The LUCA and its complex virome. *Nature Reviews Microbiology* **2020**, 1-10.
130. Koonin, E.V.; Martin, W. On the origin of genomes and cells within inorganic compartments. *TRENDS in Genetics* **2005**, *21*, 647-654.
131. Woese, C. The universal ancestor. *Proceedings of the national academy of Sciences* **1998**, *95*, 6854-6859.

132. Joyce, G.F.; Orgel, L.E. Progress toward Understanding the Origin of the RNA World. In *The RNA World : the nature of modern RNA suggests a prebiotic RNA*, Third Edition ed.; Gesteland, R.F., Cech, T.R., Atkins, J.F., Eds.; Cold Spring Harbor Monograph Series; Cold Spring Harbor Laboratory Press: Cold Spring Harbor, New York, 2006; pp. 23-56.
133. Cech, T.R. Crawling out of the RNA world. *Cell* **2009**, *136*, 599-602.
134. Gilbert, W. The RNA World. *Nature* **1986**, *319*, 618.
135. Pressman, A.; Blanco, C.; Chen, I.A. The RNA world as a model system to study the origin of life. *Current Biology* **2015**, *25*, R953-R963.
136. Van der Gulik, P.T.; Speijer, D. How amino acids and peptides shaped the RNA world. *Life* **2015**, *5*, 230-246.
137. Kaddour, H.; Sahai, N. Synergism and mutualism in non-enzymatic RNA polymerization. *Life* **2014**, *4*, 598-620.
138. Chatterjee, S.; Yadav, S. The origin of prebiotic information system in the peptide/RNA world: a simulation model of the evolution of translation and the genetic code. *Life* **2019**, *9*, 25.
139. Orgel, L.E. The origin of life on the earth. *Scientific American* **1994**, *271*, 76-83.
140. Gesteland, R.F.; Cech, T.R.; Atkins, J.F., (Eds.) *The RNA World : the nature of modern RNA suggests a prebiotic RNA*. Third Edition ed.; Cold Spring Harbor Laboratory Press: Cold Spring Harbor, New York, 2006.
141. Koonin, E.V.; Krupovic, M.; Ishino, S.; Ishino, Y. The replication machinery of LUCA: common origin of DNA replication and transcription. *BMC Biology* **2020**, *18*, 1-8.
142. Goldman, A.D.; Baross, J.A.; Samudrala, R. The enzymatic and metabolic capabilities of early life. *PLoS One* **2012**, *7*.
143. Xavier, J.C.; Hordijk, W.; Kauffman, S.; Steel, M.; Martin, W.F. Autocatalytic chemical networks at the origin of metabolism. *Proceedings of the Royal Society B* **2020**, *287*, 20192377.
144. Roberts, E.; Sethi, A.; Montoya, J.; Woese, C.R.; Luthey-Schulten, Z. Molecular signatures of ribosomal evolution. *Proc Natl Acad Sci U S A* **2008**, *105*, 13953-13958, doi:0804861105 [pii] 10.1073/pnas.0804861105.
145. Opron, K.; Burton, Z.F. Ribosome structure, function, and early evolution. *International Journal of Molecular Sciences* **2019**, *20*, 40.
146. Damer, B.; Deamer, D.; Van Kranendonk, M.; Walter, M. An Origin of Life through Three Coupled Phases in Cycling Hydrothermal Pools with Distribution and Adaptive Radiation to Marine Stromatolites. In *Proceedings of the Proceedings of the 2016 Gordon Research Conference on the Origins of Life*, 2016.
147. Lancet, D.; Segrè, D.; Kahana, A. Twenty Years of "Lipid World": A Fertile Partnership with David Deamer. *Life* **2019**, *9*, 77.
148. Branscomb, E.; Russell, M.J. Frankenstein or a submarine alkaline vent: who is responsible for abiogenesis? Part 2: as life is now, so it must have been in the beginning. *BioEssays* **2018**, *40*, 1700182.
149. Martin, W.; Russell, M.J. On the origin of biochemistry at an alkaline hydrothermal vent. *Philosophical Transactions of the Royal Society B: Biological Sciences* **2007**, *362*, 1887-1926.
150. Baross, J.A. The rocky road to biomolecules. **2018**.
151. Ménez, B.; Pisapia, C.; Andreani, M.; Jamme, F.; Vanbellinghen, Q.P.; Brunelle, A.; Richard, L.; Dumas, P.; Réfrégiers, M. Abiotic synthesis of amino acids in the recesses of the oceanic lithosphere. *Nature* **2018**, *564*, 59-63.
152. Hartman, H.; Cairns-Smith, A.G. *Clay Minerals and the Origin of Life*; CUP Archive: 1986.
153. Bernal, J.D. *The Physical Basis of Life*; Routledge & Kegan Paul: London, 1951.
154. Hansma, H.G. Could Life Originate between Mica Sheets?: Mechanochemical Biomolecular Synthesis and the Origins of Life. In *Probing Mechanics at Nanoscale Dimensions*, in *Probing Mechanics at Nanoscale Dimensions*, edited by N. Tamura, A. Minor, C. Murray, L. Friedman (Mater. Res. Soc. Symp. Proc. **Volume 1185**, Warrendale, PA, 2009), 1185-II03-15 ed.; Tamura, N., Minor, A., Murray, C., Friedman, L., Eds.; Materials Research Society: Warrendale, PA, 2009; Volume 1185, pp. II03-15.
155. Fatma Pir Cakmak; Keating, C.D. Combining Catalytic Microparticles with Droplets Formed by Phase Coexistence: Adsorption and Activity of Natural Clays at the Aqueous/Aqueous Interface. *Scientific Reports* **2017**, *7*, 1-14, doi:10.1038/s41598-017-03033-z.