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Review

# Immunotherapy for Colorectal Cancer: Mechanisms and Predictive Biomarkers

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# Simple Summary (max 150 words)

Late-stage colorectal cancer treatment often involves chemotherapy and radiation that can cause dose-limiting toxicity, therefore there is great interest in developing targeted therapies for this disease. Immunotherapy is a targeted therapy that uses peptides, cells, antibodies, viruses, or small molecules to engage or train the immune system to kill cancer. Here, we discuss the preclinical and clinical development of immunotherapy for colorectal cancer and provide an overview of predictive biomarkers for such treatments. We also consider open questions including optimal combination treatments and sensitization of colorectal cancer patients with proficient mismatch repair enzymes.

# Abstract (max 200 words)

Though early-stage colorectal cancer has a high 5-year survival rate of 65-92% depending on the specific stage, this probability drops to 13% after the cancer metastasizes. Frontline treatments for colorectal cancer such as chemotherapy and radiation often produce dose-limiting toxicities in patients and acquired resistance in cancer cells. Additional targeted treatments are needed to improve patient outcomes and quality of life. Immunotherapy involves treatment with peptides, cells, antibodies, viruses, or small molecules to engage or train the immune system to kill cancer cells. Preclinical and clinical investigations of immunotherapy for treatment of colorectal cancer including immune checkpoint blockade, adoptive cell therapy, monoclonal antibodies, oncolytic viruses, anti-cancer vaccines, and immune system modulators have been promising, but demonstrate limitations for patients with proficient mismatch repair enzymes. In this review, we discuss preclinical and clinical studies investigating immunotherapy for treatment of colorectal cancer and predictive biomarkers for response to these treatments. We also consider open questions including optimal combination treatments to maximize efficacy, minimize toxicity, and prevent acquired resistance and approaches to sensitize mismatch repair proficient patients to immunotherapy.

**Keywords (10 max):** colorectal cancer; immunotherapy; checkpoint blockade; adoptive cell therapy; monoclonal antibodies; oncolytic viruses; anti-cancer vaccines; cytokine; T cell; NK cell

#### 1. Introduction

Colorectal cancer (CRC) is a deadly disease with a 5-year survival rate of 13% when metastatic. Among all types of cancer, CRC is the third deadliest. The incidence of CRC among young people is rising in developing nations [1]. Treatment options for CRC include surgery, chemotherapy, radiation, and immunotherapy (IT). Chemotherapy regimens generally include 5-fluorouracil (5-FU) single agent or combinations with either oxaliplatin, irinotecan, or both. Chemotherapy and radiation can elicit severe toxicities in patients, and IT is a promising strategy that aims to kill cancer cells more selectively compared to these traditional treatment options [2].

CRC is classified based on genetic alterations present in the tumor such as mutations in APC, KRAS, BRAF, p53, and mismatch repair (MMR) enzymes [3]. MMR enzymes repair DNA defects, and mutations in MMR that lead to MMR deficiency (dMMR) cause an accumulation of mutations at microsatellite regions or short tandem repeats (STRs) [4]. Patients with dMMR are classified as microsatellite instable (MSI), while patients with proficient MMR (pMMR) are referred to as microsatellite stable (MSS). The accumulation of mutations in MSI tumors results in the presentation of a large repertoire of neoantigens, commonly referred to as neoantigen burden [5]. Often, this increase in neoantigen burden improves IT response in MSI patients as compared to MSS patients. Some clinical trials have reported benefit of IT exclusively in MSI patients [2]. However, MSI is detected in only about 15% of CRC tumors [6], therefore uncovering ways to sensitize MSS CRC to IT is a major area of interest.

IT includes a wide variety of different therapies that engage or train the immune system to kill cancer cells. IT includes immune checkpoint blockade (ICB), adoptive cell therapies (ACT), monoclonal antibodies (mAbs), oncolytic viruses, anti-cancer vaccines, and immune system modulators. Numerous preclinical studies have demonstrated promising results and have been translated into clinical trials (Table 1). While many of these trials are ongoing, some ITs are currently approved for CRC including the mAbs panitumumab (anti-EGFR), cetuximab (anti-EGFR), and bevacizumab (anti-VEGF) [7]. In 2020, pembrolizumab (anti-PD-1) was approved as a first-line treatment for patients with MSI+ CRC based on the results from the KEYNOTE-177 clinical trial, which reported a median progression-free survival (PFS) of 16.5 months for MSI+ CRC patients treated with pembrolizumab and 8.2 months for patients treated with chemotherapy [8]. Nivolumab (anti-PD-1) and ipilimumab (anti-CTLA-4) are also currently approved for patients with MSI+ metastatic CRC (mCRC) [9, 10].

Additional work has been done to identify groups of CRC patients who are likely to benefit from IT based on specific genetic alterations present in CRC tumors, the CRC tumor microenvironment (TME), and analytes that are detectable within patient serum (i.e. liquid biomarkers). This review will discuss the molecular bases of ITs that have been investigated preclinically and clinically for treatment of CRC, provide an overview of current predictive biomarkers for IT in CRC, will consider future requisite topics of investigation including identification of optimal combination treatments and investigation of methods to sensitize MSS CRC to IT.

#### 2. Colorectal cancer response to immunotherapy

The low 5-year survival rate (13% for mCRC) [1], development of resistance to standard therapies, and dose-limiting side effects of cytotoxic drug treatments has inspired the investigation and development of more targeted therapies for treatment of CRC. IT is a promising candidate for treatment of mCRC, especially for patients with MSI-high (MSI-H) tumors [2]. In this section, we detail the molecular mechanisms of ITs that have been investigated clinically or preclinically for treatment of CRC.

# 2.1. Immune checkpoint blockade

Under normal conditions, the body relies on immunosuppressive mechanisms to maintain immune homeostasis and self-tolerance [11, 12]. These immunosuppressive mechanisms involve expression of checkpoint proteins, activation of cell death programs, and accumulation of various immunosuppressive cells. While keeping harmful immune reactions in check, these mechanisms can also be exploited by cancer cells to escape immune attack

[13]. For example, immunosuppressive checkpoint proteins such as PD-1, CTLA-4, LAG-3, TIM-3, and NKG2A/B and are often upregulated in cancer. These molecules are expressed on the surface of immune cells and when bound by their cognate ligand, activate immunosuppressive pathways that initiate apoptosis, reduce proliferation, and/or inhibit cytokine secretion in immune cells (Figure 1).

#### 2.1.1. Anti-PD-1 and anti-PD-L1

Programmed cell death protein 1 (PD-1) is an inhibitory receptor expressed on CD8+ and CD4+ T cells. When bound by ligand, PD-1 modulates the immune response and promotes self-tolerance by activating apoptosis in antigen-specific T cells and inhibiting apoptosis of regulatory T cells (T regs) [11, 12]. PD-1 is bound by programmed cell death ligand 1 (PD-L1), a transmembrane protein that is often overexpressed on tumor cells [12]. Promising *in vitro* and *in vivo* preclinical studies have resulted in clinical trials involving PD-1/PD-L1 inhibition and the consequent approval of several anti-PD-1 mAbs.

ICB therapies that are currently used in the clinic include the FDA-approved compounds pembrolizumab and nivolumab, which can be administered to patients with MSI-H/dMMR mCRC [14, 15]. Pembrolizumab can be given as a frontline treatment, while nivolumab can be given alone or in combination with the anti-CTLA-4 inhibitor ipilimumab to patients who have progressed after chemotherapy. Recent evidence supports the use of ICB as a frontline treatment. For example, a 2020 phase III trial found that frontline pembrolizumab contributed to significantly longer progression-free survival and fewer adverse events when compared to chemotherapy in MSI-H mCRC patients [16]. Similarly, a phase II trial involving neoadjuvant treatment of MSI-H CRC patients with the anti-PD1 mAb toripalimab caused a high pathological complete response rate and an acceptable safety profile [17].

There are several ongoing clinical trials evaluating the use of ICB in the adjuvant setting. These include evaluation of the PD-L1 inhibitor atezolizumab plus FOLFOX compared to FOLFOX alone in MSI-H CRC patients [18], and evaluation of the anti-PD-1 compound avelumab following 5-FU-based treatment compared to 5-FU-based treatment alone in patients with stage III MSI or POLE-mutant CRC [19]. Other clinical trials are testing combination therapies that might improve ICB efficacy in MSS CRC patients. Interest in the potential of MEK inhibitors to sensitize tumors to IT led to the combination of regorafenib (tyrosine kinase inhibitor) and nivolumab, but this treatment resulted in only modest clinical activity in patients with MSS CRC [20]. A phase II study of pembrolizumab plus capecitabine and bevacizumab has shown this treatment to be safe and have some level of efficacy in MSS CRC patients [21]. Recent *in vivo* investigation points to low levels of tumor-infiltrating dendritic cells (DCs) as a reason for limited response in MSS tumors, suggesting co-administration of ICB with Flt-3 Ligand (FLT3L), a DC-promoting cytokine [22, 23]. Further investigation of this nature is needed to determine optimal drug combinations to enhance efficacy of ICB.

#### 2.1.2. Anti-CTLA-4

Cytotoxic T-lymphocyte-associated protein 4 (CTLA-4) is an inhibitory receptor expressed primarily by CD4+ and CD8+ T cells. Antigen-presenting cells and cancer cells express the CTLA-4 ligands CD80 and CD86. These ligands can bind to either CTLA-4 or CD28, resulting in a co-inhibitory or costimulatory response, respectively. Binding of CTLA-4 to CD80 or CD86 results in either cell-intrinsic (affecting the T cell) or cell-extrinsic (affecting the secondary cell) immunosuppressive signaling, though most evidence suggests that the CTLA-4 pathway functions primarily through cell-extrinsic mechanisms [24]. CLTA-4 is also expressed on regulatory T cells (T regs, FOXP3+). When CTLA-4 on T regs binds to CD80/86 on antigen presenting cells (APCs), priming and activation of naïve CD4+ T cells by APCs is blocked [25]. In CRC, the anti-CTLA-4 mAb ipilimumab combined with nivolumab is approved for metastatic patients [26].

#### 2.1.3. Anti-LAG-3

Lymphocyte activation gene-3 (LAG-3) is an inhibitory receptor that is expressed by CD4+ and CD8+ T cells, T regs, and NK cells. LAG-3 on CD4+ cells likely interacts with MHCII on antigen presenting cells, while LAG-3 on

CD8+ T cells and NK cells likely interacts with L-SECtin on cancer cells. These interactions inhibit immune activity by suppressing T cell proliferation, cytokine production, and Ca2+ flux. LAG-3 on T regs stimulates T reg-mediated immune suppression. The precise mechanisms by which LAG-3 mediates these effects remain to be clarified [27]. Several phase II studies are underway to investigate the anti-PD-1 mAbs nivolumab or dostarlimab in combination with other therapies including an anti-LAG-3 antibody [28-30]. One trial will specifically investigate responses in MSS CRC patients [29].

#### 2.1.4. Anti-TIM-3

T cell immunoglobulin and mucin-domain containing-3 (TIM-3), also known as Hepatitis A virus cellular receptor 2 (HAVCR2), is expressed on IFN-γ-producing T cells, T reg cells, NK cells, and APCs such as macrophages and DCs. In CD8+ cells, TIM-3 marks a dysfunctional or exhausted phenotype and limits effector function.

TIM-3 ligands include cell surface proteins CAECAM1 and phosphatidyl serine (PtdSer), soluble protein high mobility group box 1 protein (HMGB1), and galectin-9 (Gal-9), which can be expressed on the cell surface or can be secreted.

CEACAM1 is expressed constitutively on epithelial cells and in a regulated manner on immune cells depending on activation state. Defective CEACAM1 inhibits TIM-3-mediated inhibitory signaling, suggesting that TIM-3 is dependent on CEACAM1 for proper functioning [27]. Co-inhibition of TIM-3 and CEACAM1 results in improved elimination of CRC tumors in mice [31], however no clinical trials are currently investigating this combination for CRC. PtdSer is released when cells undergo apoptosis and directly regulates TIM-3 function [32]. HMGB1 binds DNA released from dying cells and mediates cytokine production and activation of innate immune that is inhibited upon TIM-3-HMGB1 binding [27]. Gal-9 is expressed on and can be secreted by tumor cells and APCs, and upon its binding to TIM-3 induces cell death in Th1 cells. Most anti-TIM-3 therapies target its interaction with CEACAM1 or PtdSer and result in immune synapse disruption, phosphatase recruitment, and apoptosis [33]. Recent investigation revealed that PD-1 physically interacts with Gal-9 and TIM-3 and attenuates Gal-9/TIM-3-induced apoptosis [34]. There is currently one clinical trial (NCT02817633) recruiting patients with advanced solid tumors for treatment with TSR-022, an anti-TIM-3 antibody [35].

#### 2.1.5. Anti-NKG2

The NKG2 family of receptors is expressed on a majority of NK cells and a subset of CD8+ T cells. Different NKG2 isoforms activate either immunosuppressive or immunostimulatory pathways. There are five isoforms, namely NKG2A, NKG2B, NKG2C, NKG2D, NKG2E, and NKG2F [36]. Inhibitory NKG2 receptors include NKG2A and NKG2B, while stimulatory receptors include NKG2C, NKG2D [37], and NKG2E. NKG2F has an unknown function. NKG2A/B bind to HLA-E, a ligand expressed at low levels with a wide tissue distribution [36, 38]. Binding inhibits myriad things including Ca<sup>2+</sup> influx, degranulation, cytokine production, and NK cell proliferation [37]. NKG2D is the most well-understood NK cell stimulatory receptor [39]. It binds to MHCI chain-related proteins MICA and MICB, which are expressed at low levels on healthy tissue and are upregulated in the presence of DNA damage or cellular transformation. NKG2D cognate ligand binding results in enhancement of NK cell cytotoxic potential. Currently, there are no clinical trials targeting NKG2 inhibitory isoforms [40], however, there are NKG2D CAR-T cell-based therapies under clinical evaluation including CYAD-101, NKR-2, and KD-025. All are in early recruitment phases [41-46].

# 2.2. Adoptive cell therapies

Adoptive cell therapy, which is also known as adoptive cell transfer, cellular adoptive IT, or T cell transfer therapy, is a type of IT in which a patient's T cells are removed from their blood, expanded *ex vivo*, sometimes modified to enhance their anti-tumor activity, and then administered back to the patient [13]. Adoptive cell therapies include tumor-infiltrating lymphocyte (TIL) therapy, engineered T cell receptor (TCR) T cell therapy, chimeric antigen receptor T cell (CAR-T cell) therapy, and NK cell therapy [13, 47].

# 2.2.1. Tumor-infiltrating lymphocyte therapy

TILs are lymphocytes that accumulate at the tumor margins or infiltrate within the tumor. These TILs are a heterogeneous population of cells that recognize tumor-associated antigens (TAAs) that are uniquely expressed or overexpressed by cancer cells. These cells can be isolated from single cell suspensions from tumor samples collected during surgery. These tumor samples are homogenized and cultured using various protocols designed to massively expand the TILs, such as culturing them in the presence of specific cytokines or APCs, and are then reinfused into the patient [13]. It was demonstrated as early as 1986 that the adoptive cell transfer of TILs to mice bearing MC-38 colon adenocarcinoma tumors caused significant regression of liver and lung micrometastases [48]. There are currently no approved TIL therapies for CRC, however numerous therapies are under evaluation in clinical trials [49].

# 2.2.2. Engineered T cell therapy

As T cells play a key role in mediating anti-tumor immunity, efforts have been made to genetically engineer or modify T cells to enhance their anti-tumor effects. Therapies developed from these efforts include TCR T cell therapy and CAR-T cell therapy. The modification of T cells can involve viral vector transfection using a lentivirus or retrovirus to express specific TCRs or CARs on the T cell surface. Both of these therapies are autologous, meaning they involve extraction, *ex vivo* modification, and re-infusion of a patient's own T cells, however, their antigen recognition mechanisms differ. TCRs use heterodimers to recognize MHC class I-presented peptides on tumor cells, while CAR-T cells use antibody fragments to directly bind to antigens on cancer cells (Figure 2). Because the peptides presented by TCRs have gone through processing by APCs, TCRs can recognize intracellular proteins, while the antigens that are recognized by CAR T cells must be expressed on the cancer cell surface [50]. Lack of tumor-specific antigens and adverse events after CAR-T treatment of patients with solid tumors necessitates new approaches including the addition of immune-activating molecules on CAR-T cells, regional administration, bispecific engagers, and optimization of CAR structure [51]. Very few clinical studies on TCR T cells are underway, but one ongoing trial is investigating the use of TCRs that were cloned from neoepitope-targeting CD8+ cells in the patient's blood for treatment of solid tumors [52].

# 2.2.3. Natural killer cell therapy

Natural killer (NK) cells are an essential component of the anti-tumor response. Similar to cytotoxic CD8+ T cells, NK cells recognize and directly kill cancer cells through the release of cytotoxic granules containing perforin and granzyme or through other pathways involving FasL/Fas. NK cells also produce IFN-γ and TNF-α as well as other important anti-cancer cytokines. Due to their evident role in eliminating cancerous cells and enhancing the anti-tumor response, NK cell-based therapies have been a topic of interest in cancer therapy development. These therapies involve autologous transfusion of NK cells that have been expanded, primed, and/or modified to enhance tumor cell killing. Similar to CAR-T cell therapy, CAR-NK cell therapy involves genetic modification of NK cells to enhance their recognition of cancer cell antigens [53].

CAR-NK cells in development for treatment of CRC include those that recognize epithelial cell adhesion molecule (EpCAM/CD326), which is overexpressed in many solid tumors and upregulates oncogene expression and cell proliferation [54]. NK cells have also been modified to recognize carcinoembryonic antigen (CEA) [53], which for many years has been the only widely recommended prognostic biomarker for CRC [55]. CEA levels are expected to fall after surgical resection of the tumor [56] and overexpression may contribute to human cancer progression by inhibiting cell-differentiation, apoptosis, and anoikis [57]. Preclinical evaluations have thus far demonstrated positive anti-tumor activity.

Several NK cell therapies have been translated into clinical trials, one of which is indicated for MUC1-positive solid tumors including CRC [58]. MUC-1 is aberrantly overexpressed in CRC [59] and preliminary results of this trial show promise [53]. Other trials involve combination of NK cell-based therapies with various other treatments including radiotherapy [60], cetuximab [61], bevacizumab [62], and chemotherapy [63].

#### 2.3. Monoclonal Antibodies

mAbs used for cancer therapy are a homogenous pool of antibodies that are specific for antigens which are overexpressed in cancer cells. Such antigens are often referred to as cancer-associated antigens (CAAs). mAbs may also be used for imaging when paired with a marker such as a radionuclide [64]. Early development of mAb therapy involved naked mAbs, which simply bind to a target and prevent its downstream signaling. More sophisticated approaches have been taken to improve efficacy of mAb therapy including the conjugation of mAbs to toxins or therapeutic agents, as well as the generation of bispecific mAbs which recognize distinct epitopes on each end of the molecule. In addition to blocking downstream signaling, IgG mAbs can induce antibody-dependent cell-mediated cytotoxicity (ADCC) via binding of the Fc receptor to immune cells [65].

#### 2.3.1. Naked monoclonal antbodies

Several mAbs are currently in clinical use for treatment of CRC. These include panitumumab (anti-EGFR), cetuximab (anti-EGFR), bevacizumab (anti-VEGF), nivolumab (anti-PD-1), pembrolizumab (anti-PD-1), and ipilimumab (anti-CTLA-4). Cetuximab is a chimeric mAb which contains a human constant region and a murine variable region, and is FDA-approved for mCRC patients who overexpress EGFR who are refractory to or intolerant of irinotecan-based chemotherapy. Panitumumab is a human mAb for mCRC patients who overexpress EGFR and who have failed fluoropyrimidine-, oxaliplatin-, and irinotecan-containing chemotherapy regimens. Panitumumab has a higher binding affinity for EGFR, and due to the absence of murine components, it has a longer half-life and causes fewer infusion reactions compared to cetuximab. Anti-EGFR mAbs are thought to induce ADCC via binding of the Fc receptor to immune cells, though the extent of ADCC induction across EGFR inhibitors varies [65]. GA201 and imgatuzumab are anti-EGFR mAbs that have been glycoengineered to enhance intrinsic ADCC induction [66, 67]. Bevacizumab, which is FDA-approved for the treatment of mCRC [68], binds to soluble VEGF and prevents binding of this growth factor with its receptors VEGFR-1 and VEGFR-2. As discussed above, the ICB mAbs nivolumab and pembrolizumab are approved for MSI-H/dMMR mCRC.

#### 2.3.2. Conjugated monoclonal antibodies

The feasibility and clinical efficacy of conjugated mAbs for treatment of CRC was demonstrated decades ago in a 1991 study of mCRC patients treated with either a CEA- or tumor associated glycoprotein (TAG)-targeting mAb conjugated to iodine-13 [69]. Since then, several other studies have demonstrated that conjugated mAbs can be efficacious for treatment of CRC. Panitumumab has been conjugated to platinum agents and demonstrated excellent *in vivo* efficacy in reducing CRC tumor size [70]. Specific populations of cells such as stem cells, which express cell surface marker CD133, within the TME can be targeted with conjugated mAbs. Suppressed tumor growth and decreased recurrence was observed in an HCT116 xenograft model after treatment with anti-CD133 antibody-conjugated SN-38 nanoparticles [71]. HER3, a protein overexpressed in CRC, has been targeted with U3-1402, a novel HER3-antibody conjugated to a topoisomerase I inhibitor. *In vivo* investigation of U3-1402 demonstrated tumor regression independent of KRAS status [72]. Following the success of the HER2-targeting mAb trastuzumab conjugated with the DNA topoisomerase I inhibitor deruxtecan (together known as ENHERTU) in the phase 2 DESTINY-Breast01 trial, this therapy was evaluated in several other types of solid tumors [73] including CRC (DESTINY-CRC01 [74]). Results of the DESTINY-CRC01 trial demonstrated durable responses in patients with previously treated HER2-positive mCRC [75], further supporting the use of conjugated mAbs in the targeted treatment of CRC in the adjuvant setting.

# 2.3.3. Bispecific antibodies

Bispecific antibodies (bsAbs) are antibodies that recognize 2 distinct epitopes on either end. Bispecific antibodies can be separated into combinatorial or obligate. Combinatorial bsAbs have a function or activity that could be achieved by combining two separate mAbs whereas obligate bsAbs have a function or activity that cannot be achieved with two separate mAbs, with the latter being a more attractive format due to the potential of creating novel functionality. Obligate bsAbs create novel functions by 1) bridging cell types such as cytotoxic CD8+ T cells and cancer cells to induce T cell-mediated killing, 2) crosslinking receptors on cells to either inactivate or

activate them, 3) positioning an enzyme and substrate as a cofactor mimetic, or 4) using one binding end to gain access into a restricted cellular compartment and using the other to exert a therapeutic effect, often referred to as "hijacking" or "piggybacking".

Most bsAbs are T cell-engaging bsAbs (bsTCEs) that engage T cells using CD3, a cell surface protein expressed on both CD8+ and CD4+ T cells. bsTCEs have shown great preclinical efficacy. For example, a bsTCE targeting GPA33, which is expressed on >95% of CRC tumors, was used to treat mice with colon or gastric cancer xenografts. Treatment resulted in tumor regression across MSS status [76]. bsTCEs have been developed to target guanylyl cyclase C (GUCY2C), which is also overexpressed in CRC. Treatment using GUCY2C-targeting bsTCEs demonstrated T cell–mediated killing of CRC cells *in vitro* and *in vivo* in CRC xenograft models across KRAS and BRAF status. Furthermore, combination of GUCY2C-targeting bsTCEs is enhanced when combined anti–PD-1/anti-PD-L1 treatment or anti-angiogenic therapies [77]. Treatment of CEA-expressing human CRC xenografts with a CEA x CD3-binding bsTCE induced an inflammatory response in the TME and caused tumor regression [78]. bsTCEs have also been designed to specifically target CD133+ stem cells in the TME and have shown efficacy in *in vivo* CRC models [79]. Development of resistance to bsTCEs may involve upregulation of checkpoint molecules PD-1/PD-L1, however this may be overcome by combination treatment of bsTCEs with anti-PD-1 or anti-PD-L1 antibodies [80].

Mutation-associated neoantigens (MANAs) are proteins encoded by mutated cancer driver genes that have gone through proteolytic processing within the cell and are presented on the cell surface by human leukocyte antigen (HLA) molecules. TCR-mimicking antibodies directed toward these MANAs (MANAbodies) have been developed into bsTCEs that target mutant KRAS or TP53, two genetic alterations frequently found in CRC. These bsTCEs can recognize and kill cancer cells with very low levels of their cognate antigen *in vitro* and *in vivo* [81-83].

Gamma delta  $(\gamma\delta)$  T cells are unconventional T cells that play a major role in the T cell-mediated anti-tumor response despite the fact that they represent only a small fraction of the total T cell population in peripheral blood [84]. bsTCEs targeting EGFR-positive cancer cells for  $\gamma\delta$  T cell-mediated death have been developed and have demonstrated efficacy in killing CRC cells while sparing EGFR+ keratinocytes, suggesting that this treatment may have a high therapeutic index in patients [80].

Some bsAbs are not T cell-engaging but can suppress tumors by different mechanisms. For example, in a CRC xenograft model, a bsAb targeting VEGF and angiopoietin-2 in combination with chemotherapy was superior compared to the clinical standard of anti-VEGF and chemotherapy combination treatment [85]. Vanucizumab, a bsAb targeting VEGF-A and angiopoietin-2, demonstrated acceptable tolerability and decreased tumor vascularity in a heterogeneous population of patients with solid tumors (13/42 had CRC) [86]. Additionally, bsAbs have been conjugated with toxic particles, inducing both photothermal and immune-mediated killing of cancer cells in a heterotopic colorectal tumor model [87].

A disappointing phase I study investigating LY3164530, a bsAb targeting MET and EGFR, demonstrated the important limitation of bsAb fixed stoichiometry. Final results indicated limited efficacy and significant toxicity due to EGFR inhibition. EGFR inhibition causes more severe toxicity than MET inhibition, but stoichiometry of bsAbs is fixed, making it difficult to adjust relative affinities on either end of the molecule. This indicates that targets with vastly different therapeutic indices are likely not an ideal combination for the bsAb approach [88]. Several other clinical trials are underway to investigate both obligate and combinatorial bsAbs (Table 1), for example XmAb841, a bsAb against CTLA-4 and LAG-3 either alone or in combination with pembrolizumab [89].

#### 2.3.4. Nanobodies

Some limitations to mAbs for the treatment of cancer include inadequate pharmacokinetics and tissue accessibility [90]. The emerging field of nanobodies (Nbs) may solve some limitations of mAbs and their derivatives, as these single variable domains of the camelid antibody are smaller, more stable, and have better tissue penetration compared to traditional mAbs [91]. One clinical trial involved treatment of four patients with solid tumors (1 CRC patient) with TAS266, a nanobody that targets death receptor 5 (DR5), a cell surface receptor that triggers apoptosis. However, the trial was terminated early due to dose-limiting toxicity in three patients [91, 92]. Another interesting early phase I trial is recruiting mesothelin+ CRC and ovarian cancer patients for treatment with mesothelin-targeting CAR-T cells that secrete PD-1 Nbs [93]. Further investigation is required before Nbs can be administered regularly. Besides avoiding toxicity, ongoing work with Nbs strives to optimize the size, drug leakage, and release pattern of conjugated mAbs in an effort to improve efficacy [94].

# 2.4. Oncolytic Virus Therapy

Oncolytic viruses are viruses that selectively infect and kill or incapacitate cancer cells without harming normal cells. This selectivity is achieved through genetic engineering. There has been relatively limited clinical investigation of oncolytic viruses for treatment of CRC and no such therapies are FDA-approved [95]. One phase I/II trial evaluated a genetically engineered oncolytic herpes simplex virus (NV1020) in patients with previously treated mCRC. Researchers observed that NV1020 stabilized liver metastases and patients experienced minimal toxicity, justifying a phase II/III trial, however this has not been initiated yet [96]. JX-594 is a vaccinia poxvirus with engineered loss of thymidine kinase, enabling its replication only in cells high levels of thymidine kinase [97]. JX-594 also contains an added GM-CSF gene, encoding a cytokine that induces DC differentiation, maturation, and function, and increases T cell activity. JX-594 did not cause dose-limiting toxicity and achieved stable disease in 67% (n=10) of pre-treated CRC patients [5, 98, 99]. Combination of JX-594 with the anti-PD-1 antibody durvalumab demonstrated tolerability and potential efficacy in mCRC patients [100]. The Newcastle disease virus (NDV) is an interesting natural RNA oncolytic virus. The NDV envelope protein and other intracellular factors induce an immune response to elicit a potent anti-tumor effect in a variety of cancers including CRC. Genetic modification of NDV could enhance this tumor lysis activity or could allow NDV to act as a vector for delivery of therapeutic genes [101].

#### 2.5. Vaccines

Cancer vaccines, or tumor vaccines, are a type of treatment that helps the body's immune system recognize and destroy cancer cells. Treatment generally involves direct administration of cells or antigens to the patient (whole tumor or peptide vaccines, respectively), indirect antigen delivery (viral vector), or delivery of APCs that were trained *ex vivo* to recognize cancer cells. Following antigen delivery, APCs process and present the peptide to CD8+ T cells, stimulating anti-tumor immunity. Adjuvants, or substances designed to enhance or direct the immune system toward a cytotoxic response, are often given in combination with the vaccine. Cancer vaccines are generally therapeutic, meaning that they are administered after the cancer has been diagnosed, as opposed to a preventative vaccine that is administered pre-diagnosis.

#### 2.5.1. Whole tumor

Whole tumor, or whole tumor cell, vaccines involve direct administration of whole tumor cells to a patient. These tumor cells can either be autologous or allogenic. In theory, whole tumor vaccines train the immune system to recognize the entire repertoire of antigens that are present in the heterogeneous TME. Intriguingly, in preclinical CRC models, whole tumor autologous vaccines exacerbated tumor growth when delivered therapeutically [102]. Initial clinical trials with autologous whole tumor vaccines in CRC patients were similarly unimpressive [103-105]. Other limitations of autologous vaccines include the large number of tumor cells needed, restricting this therapy

to more advanced-stage patients with resectable tumors. In contrast to autologous vaccines, allogenic vaccines use cell lines instead of tumor cells and have demonstrated improved efficacy in patients with colon cancer [102].

# 2.5.2. Peptide

Peptide vaccines involve administration of immunogenic cancer-associated antigens to the patient. Several CRC-associated antigens have been targeted with peptide vaccines preclinically, including ephrin type-A receptor 2 (EphA2), carcinoembryonic antigen (CEA), MUC-1, Survivin, and SART3. Though clinical evaluation has revealed that peptide vaccines can cause strong anti-tumor immune responses in CRC patients, no correlation with improved clinical outcome has been made thus far [95, 106].

#### 2.5.3. Viral vector

Viral vector vaccines use a viral vector system to deliver TAAs. Some studies suggest that vector vaccines tend to generate a higher immune response compared to peptide vaccines, but clinical trials have revealed no difference between treatment groups in terms of immune response or clinical outcomes in CRC patients [95].

# 2.5.4. Dendritic cell therapy

The most promising type of cancer vaccine involves vaccination with dendritic cells (DCs), in which DCs are removed from the patient, expanded and modified ex vivo to recognize TAAs, and then re-infused back into the patient to stimulate the cytotoxic T cell response. The most commonly known DC therapy is Sipuleucel-T for treatment of prostate cancer, which improves survival of patients by about 4 months [107]. For CRC, however, results have not been comparable. A randomized phase II clinical trial utilizing an autologous tumor lysate DC vaccine induced a tumor-specific immune response in patients, but did not impact on survival compared to patients receiving the standard of care [95].

# 2.6. Immune System Modulators

Cytokines and chemokines are fundamental to immune responses including the anti-tumor immune response. On a basic level, they regulate the growth of all blood cells and aid the development of immune responses in the event of infection with bacteria or viruses or cancer development. In recent decades, researchers have begun to uncover complex roles of cytokines that contribute to both pro- and anti-tumor effects that are sometimes disparate across cancer types [108-110]. Context-dependent effects of cytokines should be taken into consideration in the development of immunomodulatory therapies, making their development somewhat challenging. Nonetheless, several immune system modulators have entered clinical phases of investigation (Table 1).

# 2.6.1. Interleukins

Interleukins (ILs) are a class of cytokine that are produced by leukocytes. There are currently 38 recognized interleukins with some containing multiple family members such as IL-1 $\alpha$ , IL-1 $\beta$ , etc. for an overall total of more than 60 [111]. Interleukins can have a wide variety of pro- or anti- tumor effects. and many have been therapeutically targeted in CRC clinical trials including IL-2, IL-12, IL-11, IL-6, IL- $\alpha$ , and IL-1 $\beta$ . IL-2 and IL-12 are potent activators of NK cells and CD8+ T cells. IL-11 and IL-6 are tumor-promoting cytokines that function by various mechanisms. It is well-established that IL-1 $\alpha$  and IL-1 $\beta$  contribute to pro-tumor and anti-tumor mechanisms and have pleiotropic effects on immune cells, angiogenesis, cancer cell proliferation, migration, and metastasis [112, 113], and therefore there is much controversy over whether or not they should be inhibited in cancer. Treatment of mCRC patients via activation IL-2 or IL-12, or inhibition of IL-11, IL-6, IL-1 $\alpha$ , or IL-1 $\alpha$ , is currently under clinical investigation. Most are in combination with several other treatment regiments and evaluate the enhancement of these other therapies upon addition of IL. Other ILs have been investigated preclinically, yielding results that may support clinical translation. For example, IL-33 has been shown to have

potent anti-tumor activity in CRC cells that is dependent on the presence of eosinophils, which are recruited, activated, and degranulated by IL-33 [114].

#### 2.6.2. Interferons

Interferons (IFNs) are another a class of leukocyte-produced cytokines. There are three types of IFN: Type I, II, and III. Type I IFNs include 17 distinct proteins that fall into subtypes IFN $\alpha$ , IFN $\beta$ , IFN $\epsilon$ , IFN $\epsilon$  and IFN $\omega$ , but only IFN $\alpha$  and IFN $\beta$  have been well-studied in the context of cancer. There is only one Type II IFN, IFN- $\gamma$ , and Type III IFNs include IFN $\lambda$ 1, IFN $\lambda$ 2 and IFN $\lambda$ 3. Type I and Type III IFNs are induced by pattern recognition receptor pathways, while Type II IFN- $\gamma$  is mainly induced by certain types of mitogens or cytokines including IL-12 and IL-18 produced by T cells and NK cells [115]. The role of IFNs in cancer has largely been described as antitumor, though evidence suggests that in certain contexts IFNs such a IFN- $\gamma$  can be pro-tumor at certain doses [116]. As mentioned above, context should be carefully considered when developing therapeutic IFN treatments. The role of IFN dysregulation in CRC is evidenced by the fact that single nucleotide polymorphisms (SNPs) in the IFN I and IFN II pathways impact risk of CRC development and survival post-diagnosis [115, 117]. Thus, IFNs have been developed as a target for CRC therapy.

IFN-γ is likely the most well-studied IFN for treatment of CRC, but its role in cancer is not clear [118, 119]. IFN-γ cross-talk with immunostimulatory M1 macrophages can inhibit tumor growth. Stimulation of IFN-γ production in T regs in combination with ICB (anti-PD-1) caused tumor regression in MC-38 tumor-bearing mice that were resistant to single treatment with anti-PD-1. Interestingly, because IFNGR is expressed at higher levels on CRC stem-like cells compared to other CRC cells, IFN-γ treatment can selectively induce apoptosis in these cells [116].

Other preclinical studies have shown less promising results. For example, mice with CT26 colon cancer tumors exposed to IFN- $\gamma$ , experienced downregulation of antigen presentation and resultant immune evasion [119]. Another study demonstrated that the tumor suppressor gene SOCS1 mediates its anti-oncogene function through negative regulation of IFN- $\gamma$  [119, 120]. Not surprisingly, clinical trial results have been disappointing or inconclusive. A 1995 trial suggested inefficacy of surgical adjuvant treatment for patients with high-risk colon cancer [121]. Another study combined IFN- $\gamma$  with 5-FU and anti-VEGF therapy bevacizumab, but the specific contribution of IFN- $\gamma$  to the treatment effect was not investigated [122, 123]. There is an ongoing trial that involves treatment with IFN- $\gamma$  and the anti-PD-1 therapy nivolumab for treatment of solid cancers including MSI-high CRC. Preliminary results from other initial clinical trials indicated that low or moderate doses of IFN- $\gamma$  were more effective than high doses [119], suggesting that dose should be carefully considered when administering treatment and interpreting study results.

#### 2.6.3. cGAS-STING

The cyclic GMP-AMP synthase (cGAS)-stimulator of interferon genes (STING) pathway is the primary sensor of cytosolic double stranded DNA (dsDNA) from either invading pathogens or the cell itself when it becomes cancerous or begins to die. In response to cytosolic DNA, the cGAS-STING pathway induces type I IFNs, other inflammatory genes, and cellular senesence to inhibit the development of cancer [124]. STING expression in CRC has independent prognostic value [125] and a STING-related prognostic score may allow identification of high-risk CRC cases [126]. The STING agonist E7766 has shown promise in the preclinical setting in terms of anti-tumor activity and induction of a tumor-specific memory response in a murine metastatic liver CRC model. E7766 is currently in clinical trials for treatment of advanced solid tumors including CRC [127].

some efficacy [129].

Immunomodulatory drugs (IMiDs) therapeutically modulate the immune response to provide patient benefit by various mechanisms. In the context of cancer, IMiDs usually refer to thalidomide and its analogues lenalidomide and pomalidomide. Thalidomide inhibits angiogenesis, enhances production of TNF, and stimulates NK cells. Following the discovery of its anti-angiogenic properties, thalidomide was administered to drug-resistant multiple myeloma patients who subsequently experienced tumor regression. Subsequent investigation of thalidomide, lenalidomide, and pomalidomide treatment in other cancer types *in vitro* and *in vivo* indicated that IMiDs bias the immune response toward anti-tumor immunity, inhibit metastasis, and alter cancer cell signaling [128]. To date, two clinical trials that thalidomide treatment have been completed. In one of these trials, thalidomide and

capecitabine were administered to mCRC patients and the rate and duration of disease stabilization suggested

# 2.7. Targeting the immunosuppressive TME

Tumors with immunosuppressive TMEs are characterized by presence of certain cell types such as myeloid-derived suppressor cells (MDSCs), M2-like tumor-associated macrophages (TAMs), and T regs, and lack of CD8+ T cells. These tumors are often classified as "cold," whereas tumors with large numbers of infiltrating, active CD8+ T cells are classified as "hot." One strategy to transform a cold tumor into a hot tumor involves inhibition of immunosuppressive cell types. One clinical trial is investigating the CCR5 inhibitor Maraviroc for treatment of mCRC. CCR5 is receptor that is expressed on CRC cells, MDSCs, and T regs, thus its inhibition can both slow the cancer cell growth and inhibit immunosuppression in the TME [130, 131].

# 3. Predictive biomarkers for response to IT in colorectal cancer

Similarly to traditional cytotoxic therapies, IT for treatment of CRC comes with risks of side effects such as autoimmunity or cytokine storm, in which the concentration of cytokines in the blood is high enough to cause harmful inflammation and immune effects [132]. As with any therapy that has side effects, development of predictive biomarkers is a crucial step in preventing toxicity by identifying specific patient populations who are likely or unlikely to benefit from the therapy. Currently in CRC, physicians may predict response to IT based on the genetic alterations that are present in the patient's tumor, specific characteristics of the CRC TME, and specific serum-based (liquid) biomarkers present in patient serum.

# 3.1. Prediction based on genetic alterations

Three genetic alterations that are commonly found in CRC include chromosomal instability, MSI, and CpG island methylation [133]. Presence of each alteration can indicate how likely CRC patients are to respond to IT.

# 3.1.1. Chromosomal instability

The majority of CRC cases exhibit a chromosomal instability (CIN) phenotype characterized by a progression of genetic alterations and their resultant histological changes. Common and predicable genomic alterations include the activation of proto-oncogenes *KRAS* and *BRAF* and the inactivation of the tumor suppressor genes *APC* and *TP53*. It is clear that CIN is associated with poor prognosis in multiple tumor types including CRC, however, it is currently unclear if CIN is a biomarker of response to IT [134, 135]. Mutations in oncogenes such as *KRAS* and *BRAF* have recently been shown to impact on immunomodulation [136]. For example, *KRAS*-mutant patients have more T regs and less activated CD4+ memory T cells as compared to *KRAS* wild-type patients, resulting in a more immunosuppressive TME [137]. Moreover, there are several reports linking *KRAS* mutations with enhanced PD-L1 expression on tumor cells, which results in reduced T cell functionality. However, there is limited data on the role of these mutations in the prediction of response to IT [138].

The inactivation of tumor suppressor genes can also play a role in CRC response to IT. For example, the APC tumor suppressor gene is involved in the cellular transition from G1 to S phase and loss of APC can result

in the sustained activation of the Wnt signaling pathway and, thus, increased nuclear  $\beta$ -catenin [139]. It has been shown that both MSI and MSS CRC patients with APC biallelic mutations have decreased T cell tumor infiltration [140]. Another commonly mutated tumor suppressor gene in CRC is TP53. The encoded protein p53 plays a role in controlling cell cycle and apoptosis. While p53 mutations in CRC classically confer resistance to conventional chemotherapies, they provide an additional benefit in the context of IT, as p53 mutated tumors often harbor an increased number of neoantigens. In contrast, wild-type p53 has been shown to enhance MHC class I peptide transport via upregulation of TAP1, the transporter associated with antigen processing [141]. Therefore, the absence of wild-type p53 in cancer cells can diminish cytotoxic T cell-mediated tumor cell death that may impact on IT outcomes in patients.

# 3.1.2. Microsatellite instability

Approximately 15-20% of cases of CRC exhibit the MSI phenotype which is characterized by mutations in the mismatch repair (MMR) system [4]. Patients with MSI-H tumors have dMMR systems and thus, an accumulation of errors in microsatellite regions. In contrast, those with microsatellite low (MSI-L) or MSS tumors tend to have pMMR systems, and thus a lower number of DNA replication errors. Mismatch repair deficiency and the resulting MSI phenotype have long been considered predictors of efficacy for ICB in multiple tumor types. In CRC, MSI is predictive of long-term responses to ICB [142].

# 3.1.3. CpG island methylation

Another common occurrence in CRC is the aberrant epigenetic regulation of gene expression. Hypermethylation occurs when methyl groups are covalently bound in regions of CG dinucleotides or CpG-rich areas of DNA in promoter regions. In instances of normal gene expression, CpG regions are normally maintained in the unmethylated state. Methylation of these promoter regions can lead to gene silencing in tumor suppressor genes, which is denoted as CpG island methylator phenotype (CIMP) [133]. The presence of TILs is associated with a high-degree of CIMP, along with presence of other mutations [143]. CpG island methylator phenotype-high CRC is also associated with PD-L1 status of primary tumors [144].

#### 3.1.4. Tumor mutational burden and neoantigen load

In 2020, the FDA approved pembrolizumab for high tumor mutational burden (TMB-H) solid tumors based on the results of the KEYNOTE-158 study [145]. TMB is a measurement of the number of mutations per megabase of DNA in tumor cells. Typically, TMB-H CRC tumors have more immunogenic neoantigens and a higher neoantigen load. Neoantigen load is a quantification of mutations that can be targeted by T cells and is a biomarker of response to IT. Neoantigen load is correlated with the presence of TILs in both dMMR and pMMR CRC patients [146].

# 3.1.5. Other specific genetic alterations

Another frequently mutated signaling pathway in CRC is the PI3K/AKT pathway. Activating mutations in PIK3CA occur in 25% of CRC cases [147]. PIK3CA mutations are associated with CD8+ T cell infiltration, PD-L1 expression, and improved IT benefit in patients with MSS CRC [148]. The PI3K/AKT signaling pathway is negatively regulated by PTEN, a phosphatase that is mutated in approximately 18% of MSI CRC cases [149]. CRC studies have shown that a loss of PTEN results in increased PD-L1 expression [150]. Moreover, loss of PTEN has been reported to result in decreased TIL presence and an immunosuppressive TME [151]. Another common genetic alteration in CRC is mutations in TGF $\beta$ , a collection of growth factors that regulate myriad cellular processes through their interaction with TGF $\beta$  receptors [133]. TGF $\beta$  activation is predictive of poor prognosis in CRC and is predictive of ICB resistance in several tumor types [152, 153]. The most common mechanism of activation of TGF $\beta$  signaling in CRC is due to mutations in transforming growth factor  $\beta$  receptor type 2 ( $TGF\beta R2$ ), which occurs in approximately 30% of CRC cases [154]. Mutated in  $TGF\beta R2$  is predictive of resistance to ICB in non-small cell lung cancer (NSCLC), but there are currently limited reports concerning the prognostic significance of  $TGF\beta R2$  mutations in the context of CRC [155].

#### 3.2. Prediction based on the tumor microenvironment

The CRC TME is comprised of tumor cells, tumor-associated cells, endothelial cells, immune cells, and the extracellular matrix, all of which can be immunostimulatory or immunosuppressive and can influence the response to IT.

# 3.2.1. PD-L1 expression

PD-L1 is a tumor cell-expressed surface protein that is a common biomarker of response to IT across many tumor types. Tumor cells commonly overexpress PD-L1, and this overexpression is associated with poor overall survival across multiple tumor types [156]. Paradoxically, a study by Droeser et al. reported that PD-L1 expression is correlated with CD8+ T cell infiltration in pMMR CRC patients [157]. Ultimately, the value of PD-L1 alone as a predictive biomarker has limitations.

# 3.2.2. Tumor-infiltrating lymphocytes

The presence and function of TILs such as T cells, B cells, and NK cells can be predictive of response to IT. A positive correlation between TILs and response to ICB has been shown repeatedly in melanoma [158]. A study by Li et al. reported that a signature of high immune cell infiltration in CRC patients was correlated with increased PD-L1 expression and better prognosis as compared to CRC patients clustered in the low and medium immune infiltration groups [159]. Interestingly, in CRC a correlation between T reg cell tumor-infiltration and response to chemo-IT has also been reported [160].

#### 3.2.3. Immune status of the tumor microenvironment

The immune status of the TME can be broadly categorized as immune-inflamed, immune-excluded, or immune-desert. A study by Tang et al. categorized CRC patients into four groups based on the expression signatures of cancer-associated fibroblasts, MDSCs, M2-like tumor-associated macrophages (TAMs), CD8+ T cells, and PD-L1 expression and found that classification of CRC patients into immune subtypes was a reliable predictor of prognosis. The clustering strategy developed here has great potential to guide personalized immunotherapy based on the TME [161].

# 3.2.4. Diversity of T cell repertoires in TME

T cell receptors recognize specific antigens presented by MHCI (for CD8+ T cells) or MHCII (for CD4+ T cells) molecules. A process called genetic recombination occurs in T cells to rearrange the DNA at three loci (TRBV, TRBD and TRBJ) to develop TCRs that are specific for certain antigens. The T cell repertoire refers to all of the unique TCR genetic rearrangements within the adaptive immune system, thus the TME T cell repertoire refers to all of the unique TCR genetic rearrangements within the TME. Not surprisingly, having a diverse TME T cell repertoire is associated with better outcomes in response to immunotherapy. This holds true when evaluating CRC patient populations specifically. In a TCR repertoire analysis of advanced CRC patients treated with a combination of five cancer peptide vaccines and oxaliplatin-based chemotherapy, high TCR diversity scores were associated with improved response [162]. A recent report suggested a significant difference in the usage of TRBV and TRBJ genes between CRC patients and healthy controls, supporting its use as an additional TCR-based predictive biomarker in CRC [163].

#### 3.2.5. Tumor-associated macrophages

TAMs are a type of macrophage that create an immunosuppressive TME by producing immunosuppressive cytokines and growth factors, and by increasing immune checkpoint expression on immune cells. Macrophages are classified into two types: classical-activated M1 and alternate-activated M2. The conversion between these two states is called polarization. Though immunosuppressive TAMs can exhibit either polarization phenotype, it is believed that they generally are more M2-like [164]. Studies suggest that up to 80% of cancer patients, increased levels of TAMs predict worse outcomes. In CRC, however, there are conflicting findings as far as the

prognostic and predictive value of TAMs [164-166] that may be clarified with investigations that make a clear distinction between macrophages with different polarization states or phenotypes. Our knowledge of the role of TAMs in the response to IT across cancer types is very limited [167]. In CRC, TAMs express PD-1 and treatment with anti-PD-1 compounds increased phagocytosis of PD-1+ TAMs. TAMs can also sequester anti-PD-1 antibodies, preventing their therapeutic binding to their target immune cell. Thus, TAMs play key roles in modulating response to immunotherapy in CRC cells but it is not clear whether their levels actually correlate with better or worse response to IT in CRC patients [168].

# 3.2.6. The gut microbiota

The makeup of the gut microbiome may influence responses to immunotherapy. A recent investigation using a mouse model of CRC identified intratumoral presence of the bacterial species *Bifidobacterium pseudolongum*, *Lactobacillus johnsonii*, and *Olsenella* as enhancers of ICB efficacy. *B. pseudolongum* produced inosine, which translocated systemically after ICB-induced gut barrier impairment to activate antitumor T cells [169]. Another study found a significantly different ratio of *Prevotella/Bacteroides* species between ICB responders versus nonresponders in CRC [170]. Thus, the relative abundance of these species within the tumor could predict response to ICB but this remains to be confirmed.

# 3.3. Liquid biomarkers

Peripheral blood offers a non-invasive source of potential biomarkers via liquid biopsies. Liquid biopsies can provide information on circulating factors such as peripheral blood cells, tumor DNA, and cytokines. Other potential liquid biomarkers that may predict responses in CRC include exosomes.

# 3.3.1. Peripheral blood cells

There are significant differences in the proportions of lymphocytes, white blood cells, neutrophils and MDSCs between healthy controls and patients with CRC [171]. This differential peripheral blood cellular composition has been reported as both diagnostic and prognostic in patients with CRC. Peripheral NK cells are an independent predictor of survival in patients with CRC and their prognostic value increases when combined with B cell count [172]. In a study of CRC patients with liver metastases, patients more likely to respond to CAR T therapy had lower-fold changes in their neutrophil-to-lymphocyte (NLR) ratio [173].

# 3.3.2. Circulating tumor DNA

Circulating tumor DNA (ctDNA) is tumor-derived fragmented DNA found in the bloodstream that is no longer cell-associated. In a review of ninety-two clinical studies, ctDNA was demonstrated to be a reliable measure of tumor burden and response to therapy in CRC [174]. In a study of MSS CRC patients treated with combination therapy of regorafenib and PD-1 inhibitors, the authors found that ctDNA may be predictive of early therapeutic efficacy of IT in the MSS CRC patient population [20].

#### 3.3.3. Exosomes

Exosomes are a membrane-bound type of extracellular vesicle, which can carry cargo such as mRNA, microRNA (miRNA), long non-coding RNA (lncRNA), protein, DNA fragments, or debris [175] and deliver this cargo to other cells, mediating various downstream signaling pathways. Extracellular vesicles contribute to tumorigenesis in many ways [175] including by increasing the migration of fibroblasts in the TME [176]. Exosomes containing biomarkers such as PD-1 or PD-L1 predict better response to IT in melanoma and NSCLC, whereas exosomes containing hsa-miR-320b/c/d or hsa-miR-125b-5 predict worse response to IT in NSCLC. It remains to be seen if circulating exosomes predict response to IT in CRC [177].

# 4. Discussion & open questions

IT development for treatment of CRC has made great strides in the past several decades, particularly for dMMR/MSI patients. ITs targeting cancer- or tumor-associated antigens EGFR, PD-1, and CTLA-4 have been approved and are often used for this subset of CRC patients in combination or with or after failure of conventional chemotherapy regimens. One major challenge in developing IT for CRC is sensitizing pMMR CRC tumors, which are present in a majority (85%) of CRC patients. Further investigation of the transcriptomic, cytokine, and immune cell responses to IT in dMMR versus pMMR CRC are likely to reveal differences that will guide combination treatment for pMMR CRC patients. For example, the extent of immune cell, especially T cell, infiltrate is much higher in dMMR CRC patients in response to IT and correlates with positive responses in both dMMR and pMMR CRC. Enhancing immune cell infiltration in pMMR tumors, for example by inhibiting MDSCs, is a promising strategy to sensitize these tumors to IT. Another major challenge in the development of IT is a gap in knowledge regarding optimal treatment strategies, even for FDA approved ICB treatments. It is largely unknown whether IT is more effective in the neoadjuvant or adjuvant setting, or if IT should be combined with chemotherapy. Further investigation will clarify these questions.

The environment in which a cancer grows has profound effects on efficacy of IT. For example, subcutaneous injection of pMMR CRC cells in mice led to the development of tumors that were responsive to ICB, unlike what is seen in patients. When these cells were injected orthotopically into the liver, the main site of CRC cell metastasis, the tumors that developed were resistant to ICB. Comparing these tumors revealed that the resistant tumors in the liver had a significantly lower number of DCs compared to the sensitive subcutaneous tumors. A similar observation was made in liver metastases of CRC patients. As DCs are required for antigen presentation and activation of CD8+ T cells, their low numbers likely contribute to the resistance of liver mCRC cell tumors. This points to combination treatments with FLT3L and ICB, as FLT3L is cytokine that promotes the differentiation and/or expansion of immature DCs [22, 23].

Other exciting new developments could benefit both pMMR and dMMR CRC patients. Mucosal associated invariant T (MAIT) cells are unconventional T cells that play a key role in the immune response to microbes. These cells have been most extensively studied in the context of CRC as compared to other cancer types, but still there is no consensus regarding the role they play in anti-tumor immunity. As MAIT cells are abundant in mucosal tissue such as the gut and are found in a majority of CRC tumors, elucidating this role may guide novel immunotherapies for CRC. Both agonist and antagonist molecules of the semi-invariant TCR on invariant T cells are well-characterized and would ease the development of these therapies [178]. Another recent investigation determined that mutations in more than one hundred tumor suppressor genes can prevent immune cell recognition of cancer cells in vivo. This novel insight into the role of altered tumor suppressors in immune evasion could guide novel immunotherapies which target or restore altered tumor suppressor genes in combination with other IT [179, 180]. Additionally, unraveling the interplay of gut microbiota and IT may reveal approaches to increase abundance of immunostimulatory microbes and decrease abundance of immunosuppressive species to enhance IT efficacy [169, 170, 181]. Ongoing investigation of immune escape mechanisms will continue to reveal next-generation IT targets. For example, recent work has revealed that cancer cells hijack the mitochondria of immune cells via nanotubes. Inhibiting nanotube formation led to improved outcomes after anti-PD-1 therapy in a breast cancer model, and next steps should include similar investigation in colorectal cancer [182].

It is likely that a majority of cancer patients will receive chemotherapy before or concurrently with immunotherapy in the near future. It is well-documented that conventional chemotherapy and targeted therapies have drastic effects on cytokine production and secretion across cell types including cancer cells and immune cells. These cytokines can have effects that enhance the anti-tumor activity of the drug, but could also lead to acquired resistance of cancer cells or systemic toxicity in the patient. Cytokine profiling in cancer cells treated with these drugs can guide logical combination with IT to enhance their anti-tumor effects. This has been completed in HCT116 CRC cells [183] and MCF7 breast cancer cells [184] treated with conventional chemotherapies and in HCT-116, HT-29, and KM12C CRC cells treated with MEK, TRK, tyrosine kinase, RET,

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BRAF, PARP, PI3K, and GSK-3 inhibitors as well as Imipridones [185]. In HCT116 cells treated with chemotherapies 5-FU, irinotecan, oxaliplatin, cisplatin, or clinically relevant combinations, drug-specific effects and an impact of p53 status on the modulation cytokines IL-8, ferritin, and soluble TRAIL-R2 was observed [183]. Possible tissue-specific effects were noted after treatment of breast cancer cells and CRC cells with cisplatin, oxaliplatin, and 5-FU, including enhanced downregulation IFN-β and TRAIL and upregulation of TRAIL-R2 in the breast cancer cells [184]. This finding demonstrates that drug effects on cancer cells are likely tissue type-dependent, and emphasizes the importance of evaluating cytokine induction in the tissue of interest. Treatment of CRC cells with clinically relevant targeted treatments revealed several effects that were observed across drug treatments despite highly heterogeneous cell line mutational profiles and therapeutic mechanisms of action. These effects included downregulation of VEGF, CXCL9, and IL-8, and included upregulation of CXCL14, CCL5, and CXCL5. Other effects were more drug-class dependent [185]. Together, studies like these could support the logical combination of treatments to enhance or direct the immune system towards anti-tumor immunity in CRC.

Table 1. The number of clinical trials involving various types of immunotherapy for the treatment of colorectal cancer.

Number of clinical trials											
	ICB	ACT	mAb	Conj. Ab	bsAb	Virus	Vaccine	IFN	IL	IMiD	STING
Completed	31	8	67	3	3	3	46	11	7	2	0
Active, not recruiting	46	6	19	-	-	1	4	1	4	-	0
Recruiting	111	15	52	2	8	2	18	4	5	-	1
Not yet recruiting	16	7	9	1	3	-	2	1	-	-	0
Terminated	11	4	29	1	1	1	11	2	5	1	0
Withdrawn	10	2	12	2	2	1	5	1	2	-	0
Suspended	-	1	-	-	-	-	1	-	-	-	0
Unknown	9	14	15	-	2	-	9	5	2	3	0
Total	234	57	203	9	19	8	96	25	25	6	1
				Search ter	ms						
ICB	immune checkpoint blockade OR checkpoint blockade OR immune checkpoint inhibitor OR anti-pd1 OR anti-pdl1 OR anti-ctla4 OR anti-lag3 OR anti-tim3 OR anti-nkg2 OR pd1 OR pdl1 OR ctla4 OR lag3 OR tim3 OR nkg2										
ACT	adoptive cell therapy OR adoptive cell transfer OR cellular adoptive immunotherapy OR t cell transfer therapy OR tumor-infiltrating lymphocyte OR engineered t cell OR t cell receptor therapy OR car t cell OR NK cell										
mAb	monoclonal antibody OR monoclonal antibodies										
Conj. Ab	conjugated antibody OR conjugated antibodies										
bsAb	bispecific antibody OR bispecific antibodies										
Virus	oncolytic virus										
Vaccine	vaccine										
IFN	interferon										
IL	interleukin										
IMiD	thalidomide										
STING	sting										

ClinicalTrials.gov search was conducted as follows: condition or disease = colorectal cancer; intervention/treatment = search terms above; study type=interventional. Search was performed on 10/30/2021. Some trials fall into two groups, for example many ICB therapies are mAb-based. Clinical trials involving combination treatments also may fall into two groups. ICB, immune checkpoint blockade; ACT, adoptive cell therapy; mAb, monoclonal antibody; conj. Ab, conjugated antibody; bsAb, bispecific antibody; IFN, interferon; IL: interleukin; IMiD, immunomodulatory drug.

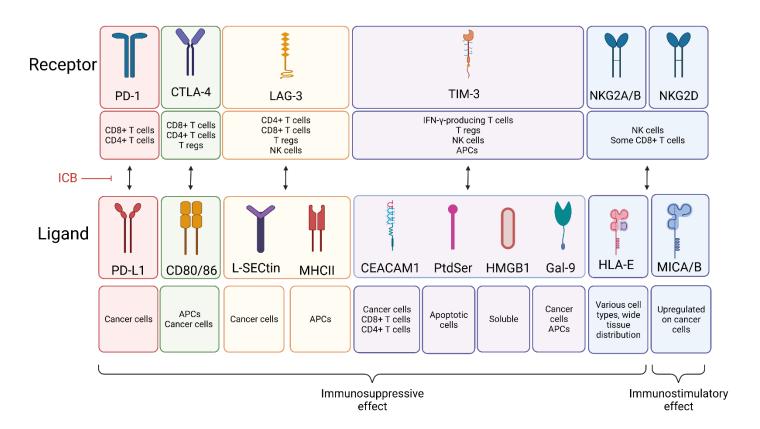
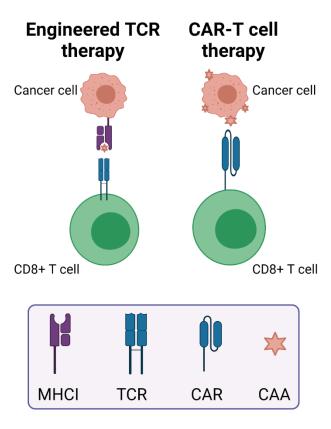


Figure 1. Immune cell and cancer cell interactions relevant to immune checkpoint blockade therapy. CD8+ T cells, CD4+ T cells, NK cells, and T regs express receptors that are susceptible to binding by various cognate ligands present on the surface of colorectal cancer cells or APCs. Ligand binding results in immunosuppressive, or immunostimulatory in the case of NKG2D, signaling. ICB is a therapeutic approach involving inhibition of various receptor-ligand interactions. ICB, immune checkpoint blockade; APC, antigen presenting cell; NK cell, natural killer cell. Created in BioRender.



**Figure 2. Engineered T cell receptor therapy vs. CAR-T cell therapy.** Engineered T cell receptor (TCR) T cell therapy involves engineering T cells with receptors that are specific for MHC class I-presented antigenic peptides. Chimeric antigen receptor T cell (CAR-T cell) therapy involves direct recognition of CAAs on the cancer cell surface. MHCI, major histocompatibility complex class I; TCR, T cell receptor; CAR, chimeric antigen receptor; CAA, cancer-associated antigen. Created in BioRender.

#### References

- 1. Rawla, P., T. Sunkara, and A. Barsouk, *Epidemiology of colorectal cancer: incidence, mortality, survival, and risk factors.* Prz Gastroenterol, 2019. **14**(2): p. 89-103.
- 2. Xie, Y.H., Y.X. Chen, and J.Y. Fang, *Comprehensive review of targeted therapy for colorectal cancer.* Signal Transduct Target Ther, 2020. **5**(1): p. 22.
- 3. Müller, M.F., A.E. Ibrahim, and M.J. Arends, *Molecular pathological classification of colorectal cancer.* Virchows Arch, 2016. **469**(2): p. 125-34.
- 4. Nojadeh, J.N., S. Behrouz Sharif, and E. Sakhinia, *Microsatellite instability in colorectal cancer.* EXCLI Journal, 2018. **17**: p. 159-168.
- 5. Tintelnot, J. and A. Stein, *Immunotherapy in colorectal cancer: Available clinical evidence, challenges and novel approaches.* World J Gastroenterol, 2019. **25**(29): p. 3920-3928.
- 6. Boland, C.R. and A. Goel, *Microsatellite instability in colorectal cancer.* Gastroenterology, 2010. **138**(6): p. 2073-2087.e3.
- 7. Berger, M.D. and H.J. Lenz, *The safety of monoclonal antibodies for treatment of colorectal cancer.* Expert Opin Drug Saf, 2016. **15**(6): p. 799-808.
- 8. Shiu, K.-K., et al., *KEYNOTE-177: Phase III randomized study of pembrolizumab versus chemotherapy for microsatellite instability-high advanced colorectal cancer.* Journal of Clinical Oncology, 2021. **39**(3\_suppl): p. 6-6.
- 9. Marcus, L., et al., FDA Approval Summary: Pembrolizumab for the Treatment of Microsatellite Instability-High Solid Tumors. Clinical Cancer Research, 2019. **25**(13): p. 3753-3758.
- 10. Overman, M.J., et al., *Nivolumab in patients with metastatic DNA mismatch repair-deficient or microsatellite instability-high colorectal cancer (CheckMate 142): an open-label, multicentre, phase 2 study.* Lancet Oncol, 2017. **18**(9): p. 1182-1191.
- 11. Iliopoulos, D., et al., *The negative costimulatory molecule PD-1 modulates the balance between immunity and tolerance via miR-21.* Eur J Immunol, 2011. **41**(6): p. 1754-63.
- 12. Han, Y., D. Liu, and L. Li, *PD-1/PD-L1 pathway: current researches in cancer.* Am J Cancer Res, 2020. **10**(3): p. 727-742.
- 13. Fan, J., et al., *Adoptive Cell Transfer: Is it a Promising Immunotherapy for Colorectal Cancer?* Theranostics, 2018. **8**(20): p. 5784-5800.
- 14. FDA Approves First-Line Immunotherapy for Patients with MSI-H/dMMR Metastatic Colorectal Cancer. Available from: <a href="https://www.fda.gov/news-events/press-announcements/fda-approves-first-line-immunotherapy-patients-msi-hdmmr-metastatic-colorectal-cancer">https://www.fda.gov/news-events/press-announcements/fda-approves-first-line-immunotherapy-patients-msi-hdmmr-metastatic-colorectal-cancer</a>.
- 15. Administration, U.S.F.a.D. *FDA grants nivolumab accelerated approval for MSI-H or dMMR colorectal cancer*. Available from: <a href="https://www.fda.gov/drugs/resources-information-approved-drugs/fda-grants-nivolumab-accelerated-approval-msi-h-or-dmmr-colorectal-cancer">https://www.fda.gov/drugs/resources-information-approved-drugs/fda-grants-nivolumab-accelerated-approval-msi-h-or-dmmr-colorectal-cancer</a>.
- 16. André, T., et al., *Pembrolizumab in Microsatellite-Instability-High Advanced Colorectal Cancer.* N Engl J Med, 2020. **383**(23): p. 2207-2218.
- 17. Hu, H., et al., Neoadjuvant PD-1 blockade with toripalimab, with or without celecoxib, in mismatch repair-deficient or microsatellite instability-high, locally advanced, colorectal cancer (PICC): a single-centre, parallel-group, non-comparative, randomised, phase 2 trial. Lancet Gastroenterol Hepatol, 2021
- 18. Assessing Adjuvant Immunotherapy for MSI-H Colorectal Cancer. 2018; Available from: <a href="https://dailynews.ascopubs.org/do/10.5555/ADN.18.190032/full/">https://dailynews.ascopubs.org/do/10.5555/ADN.18.190032/full/</a>.
- 19. Hirano, H., et al., *Current status and perspectives of immune checkpoint inhibitors for colorectal cancer.* Japanese Journal of Clinical Oncology, 2020. **51**(1): p. 10-19.
- 20. Wang, C., et al., Regorafenib and Nivolumab or Pembrolizumab Combination and Circulating Tumor DNA Response Assessment in Refractory Microsatellite Stable Colorectal Cancer. Oncologist, 2020. **25**(8): p. e1188-e1194.
- 21. Bocobo, A.G., et al., *Phase II study of pembrolizumab plus capecitabine and bevacizumab in microsatellite stable (MSS) metastatic colorectal cancer (mCRC): Interim analysis.* Journal of Clinical Oncology, 2021. **39**(3 suppl): p. 77-77.
- 22. Research, N.C.f.C. *Beating colorectal cancer's immunotherapy resistance*. Available from: <a href="https://news.harvard.edu/gazette/story/2021/10/new-way-to-overcome-colorectal-cancers-resistance-to-immune-response/">https://news.harvard.edu/gazette/story/2021/10/new-way-to-overcome-colorectal-cancers-resistance-to-immune-response/</a>.

- 23. Hospital, M.G. *Research points to a strategy for overcoming colorectal cancers' immunotherapy resistance*. Available from: <a href="https://medicalxpress.com/news/2021-10-strategy-colorectal-cancers-immunotherapy-resistance.html">https://medicalxpress.com/news/2021-10-strategy-colorectal-cancers-immunotherapy-resistance.html</a>.
- 24. Van Coillie, S., B. Wiernicki, and J. Xu, *Molecular and Cellular Functions of CTLA-4.* Adv Exp Med Biol, 2020. **1248**: p. 7-32.
- 25. Sobhani, N., et al., *CTLA-4 in Regulatory T Cells for Cancer Immunotherapy.* Cancers (Basel), 2021. **13**(6).
- 26. ; Available from: <a href="https://www.fda.gov/news-events/press-announcements/fda-approves-first-line-immunotherapy-patients-msi-hdmmr-metastatic-colorectal-cancer">https://www.fda.gov/news-events/press-announcements/fda-approves-first-line-immunotherapy-patients-msi-hdmmr-metastatic-colorectal-cancer</a>.
- 27. Anderson, A.C., N. Joller, and V.K. Kuchroo, *Lag-3, Tim-3, and TIGIT: Co-inhibitory Receptors with Specialized Functions in Immune Regulation*. Immunity, 2016. **44**(5): p. 989-1004.
- 28. An Investigational Immuno-therapy Study of Nivolumab, and Nivolumab in Combination With Other Anti-cancer Drugs, in Colon Cancer That Has Come Back or Has Spread.
- 29. Study of Nivolumab and Relatlimab in Patients With Microsatellite Stable (MSS) Advanced Colorectal Cancer.
- 30. Study of TSR-033 With an Anti-programmed Cell Death-1 Receptor (PD-1) in Participants With Advanced Solid Tumors.
- 31. Huang, Y.H., et al., *CEACAM1 regulates TIM-3-mediated tolerance and exhaustion.* Nature, 2015. **517**(7534): p. 386-90.
- 32. Smith, C.M., et al., *Phosphatidylserine binding directly regulates TIM-3 function.* Biochem J, 2021. **478**(17): p. 3331-3349.
- 33. Wolf, Y., A.C. Anderson, and V.K. Kuchroo, *TIM3 comes of age as an inhibitory receptor.* Nat Rev Immunol, 2020. **20**(3): p. 173-185.
- 34. Yang, R., et al., *Galectin-9 interacts with PD-1 and TIM-3 to regulate T cell death and is a target for cancer immunotherapy.* Nat Commun, 2021. **12**(1): p. 832.
- 35. A Study of TSR-022 in Participants With Advanced Solid Tumors (AMBER).
- 36. Borrego, F., et al., *The CD94/NKG2 family of receptors: from molecules and cells to clinical relevance.* Immunol Res, 2006. **35**(3): p. 263-78.
- 37. Lanier, L.L., *Up on the tightrope: natural killer cell activation and inhibition.* Nat Immunol, 2008. **9**(5): p. 495-502.
- 38. Huang, H., et al., *Up-regulation of NKG2F receptor, a functionally unknown killer receptor, of human natural killer cells by interleukin-2 and interleukin-15.* Oncol Rep, 2010. **24**(4): p. 1043-8.
- 39. Xiao, L., et al., *Adoptive Transfer of NKG2D CAR mRNA-Engineered Natural Killer Cells in Colorectal Cancer Patients*. Mol Ther, 2019. **27**(6): p. 1114-1125.
- 40. *NKG2* | *Colorectal Cancer*. Available from: <a href="https://clinicaltrials.gov/ct2/results?cond=Colorectal+Cancer+&term=NKG2&cntry=&state=&city=&dist="https://clinicaltrials.gov/ct2/results?cond=Colorectal+Cancer+&term=NKG2&cntry=&state=&city=&dist=.</a>
- 41. alloSHRINK Standard cHemotherapy Regimen and Immunotherapy With Allogeneic NKG2D-based CYAD-101 Chimeric Antigen Receptor T-cells. Available from: https://ClinicalTrials.gov/show/NCT03692429.
- 42. Dose Escalation and Dose Expansion Phase I Study to Assess the Safety and Clinical Activity of Multiple Doses of NKR-2 Administered Concurrently With FOLFOX in Colorectal Cancer With Potentially Resectable Liver Metastases. Available from: <a href="https://ClinicalTrials.gov/show/NCT03310008">https://ClinicalTrials.gov/show/NCT03310008</a>.
- 43. Haplo Allogeneic NKG2DL-targeting Chimeric Antigen Receptor-grafted γδ T Cells for Relapsed or Refractory Solid Tumour. Available from: https://ClinicalTrials.gov/show/NCT04107142.
- 44. Hepatic Transarterial Administrations of NKR-2 in Patients With Unresectable Liver Metastases From Colorectal Cancer. Available from: https://ClinicalTrials.gov/show/NCT03370198.
- 45. *NKG2D CAR-T(KD-025) in the Treatment of Relapsed or Refractory NKG2DL+ Tumors.* Available from: https://ClinicalTrials.gov/show/NCT04550663.
- 46. Phase 1b Study to Evaluate the Addition of a Pembrolizumab Treatment After Treatment With CYAD-101 With a FOLFOX Preconditioning in Metastatic Colorectal Cancer Patients. Available from: https://ClinicalTrials.gov/show/NCT04991948.
- 47. *adoptive cell transfer.* NCI Dictionaries; Available from: <a href="https://www.cancer.gov/publications/dictionaries/cancer-terms/def/adoptive-cell-transfer">https://www.cancer.gov/publications/dictionaries/cancer-terms/def/adoptive-cell-transfer</a>.

- 48. Rosenberg, S.A., P. Spiess, and R. Lafreniere, *A new approach to the adoptive immunotherapy of cancer with tumor-infiltrating lymphocytes*. Science, 1986. **233**(4770): p. 1318-21.
- 49. tumor infiltrating lymphocyte | Colorectal Cancer. Available from:

  <a href="https://clinicaltrials.gov/ct2/results?cond=Colorectal+Cancer&term=tumor+infiltrating+lymphocyte&cntry=&state=&city=&dist=.">https://clinicaltrials.gov/ct2/results?cond=Colorectal+Cancer&term=tumor+infiltrating+lymphocyte&cntry=&state=&city=&dist=.</a>
- 50. Zhao, L. and Y.J. Cao, *Engineered T Cell Therapy for Cancer in the Clinic.* Front Immunol, 2019. **10**: p. 2250.
- 51. Li, H., et al., CAR-T cells for Colorectal Cancer: Target-selection and strategies for improved activity and safety. J Cancer, 2021. **12**(6): p. 1804-1814.
- 52. Chmielowski, B., et al., A phase Ia/Ib, open-label first-in-human study of the safety, tolerability, and feasibility of gene-edited autologous NeoTCR-T cells (NeoTCR-P1) administered to patients with locally advanced or metastatic solid tumors. Journal of Clinical Oncology, 2020. **38**(15\_suppl): p. TPS3151-TPS3151.
- 53. Liu, S., et al., *NK cell-based cancer immunotherapy: from basic biology to clinical development.* J Hematol Oncol, 2021. **14**(1): p. 7.
- 54. Zhang, Q., et al., Combination Therapy with EpCAM-CAR-NK-92 Cells and Regorafenib against Human Colorectal Cancer Models. J Immunol Res, 2018. **2018**: p. 4263520.
- 55. Hermunen, K., et al., Postoperative serum CA19-9, YKL-40, CRP and IL-6 in combination with CEA as prognostic markers for recurrence and survival in colorectal cancer. Acta Oncol, 2020. **59**(12): p. 1416-1423.
- 56. Campos-da-Paz, M., et al., Carcinoembryonic Antigen (CEA) and Hepatic Metastasis in Colorectal Cancer: Update on Biomarker for Clinical and Biotechnological Approaches. Recent Pat Biotechnol, 2018. **12**(4): p. 269-279.
- 57. Shiozawa, M., et al., *Pharmacologically upregulated carcinoembryonic antigen-expression enhances the cytolytic activity of genetically-modified chimeric antigen receptor NK-92MI against colorectal cancer cells.* BMC Immunol, 2018. **19**(1): p. 27.
- 58. CAR-pNK Cell Immunotherapy in MUC1 Positive Relapsed or Refractory Solid Tumor. Available from: https://ClinicalTrials.gov/show/NCT02839954.
- 59. Takahashi, H., et al., *MUC1-C activates the TAK1 inflammatory pathway in colon cancer.* Oncogene, 2015. **34**(40): p. 5187-97.
- 60. Radiofrquency Ablation Combined With Cytokine-induced Killer Cells for Colorectal Cancer Liver Metastases. Available from: <a href="https://clinicalTrials.gov/show/NCT02419677">https://clinicalTrials.gov/show/NCT02419677</a>.
- 61. A Phase Ib Study of Immunotherapy with Ex Vivo Pre-Activated and Expanded Cb-Nk Cells in Combination with Cetuximab, in Colorectal Cancer Patients with Minimal Residual Disease (Mrd). Available from: https://ClinicalTrials.gov/show/NCT05040568.
- 62. The Study of CIK Plus S-1 and Bevacizumab as Maintenance Treatment for Patients With Advanced Colorectal Cancer. Available from: https://ClinicalTrials.gov/show/NCT02487992.
- 63. Chemotherapy Combined With CIK Treating Colon Cancer. Available from: https://ClinicalTrials.gov/show/NCT03084809.
- 64. Touchefeu, Y., et al., *Promising clinical performance of pretargeted immuno-PET with anti-CEA bispecific antibody and gallium-68-labelled IMP-288 peptide for imaging colorectal cancer metastases: a pilot study.* Eur J Nucl Med Mol Imaging, 2021. **48**(3): p. 874-882.
- 65. Baysal, H., et al., *The Right Partner in Crime: Unlocking the Potential of the Anti-EGFR Antibody Cetuximab via Combination With Natural Killer Cell Chartering Immunotherapeutic Strategies.* Front Immunol, 2021. **12**: p. 737311.
- 66. Kol, A., et al., ADCC responses and blocking of EGFR-mediated signaling and cell growth by combining the anti-EGFR antibodies imgatuzumab and cetuximab in NSCLC cells. Oncotarget, 2017. **8**(28): p. 45432-45446.
- 67. Oppenheim, D.E., et al., *Glyco-engineered anti-EGFR mAb elicits ADCC by NK cells from colorectal cancer patients irrespective of chemotherapy.* Br J Cancer, 2014. **110**(5): p. 1221-7.
- 68. Goodin, S., *Development of monoclonal antibodies for the treatment of colorectal cancer.* Am J Health Syst Pharm, 2008. **65**(11 Suppl 4): p. S3-7; quiz S22-4.
- 69. Riva, P., et al., *Treatment of metastatic colorectal cancer by means of specific monoclonal antibodies conjugated with iodine-131: a phase II study.* Int J Rad Appl Instrum B, 1991. **18**(1): p. 109-19.

- 70. Tsai, M.H., et al., *Panitumumab-Conjugated Pt-Drug Nanomedicine for Enhanced Efficacy of Combination Targeted Chemotherapy against Colorectal Cancer.* Adv Healthc Mater, 2017. **6**(13).
- 71. Ning, S.T., et al., *Targeting Colorectal Cancer Stem-Like Cells with Anti-CD133 Antibody-Conjugated SN-38 Nanoparticles*. ACS Appl Mater Interfaces, 2016. **8**(28): p. 17793-804.
- 72. Koganemaru, S., et al., *U3-1402, a Novel HER3-Targeting Antibody-Drug Conjugate, for the Treatment of Colorectal Cancer.* Mol Cancer Ther, 2019. **18**(11): p. 2043-2050.
- 73. Keam, S.J., Trastuzumab Deruxtecan: First Approval. Drugs, 2020. 80(5): p. 501-508.
- 74. DS-8201a in Human Epidermal Growth Factor Receptor2 (HER2)-Expressing Colorectal Cancer (DESTINY-CRC01). Available from: <a href="https://ClinicalTrials.gov/show/NCT03384940">https://ClinicalTrials.gov/show/NCT03384940</a>.
- 75. Stenger, M. *Trastuzumab Deruxtecan-nxki in HER2-Positive Metastatic Colorectal Cancer: DESTINY-CRC01*. 2021 [cited 2021 10/7/2021]; Available from: <a href="https://ascopost.com/news/may-2021/trastuzumab-deruxtecan-nxki-in-her2-positive-metastatic-colorectal-cancer-destiny-crc01/">https://ascopost.com/news/may-2021/trastuzumab-deruxtecan-nxki-in-her2-positive-metastatic-colorectal-cancer-destiny-crc01/</a>.
- 76. Wu, Z., et al., *Development of a Tetravalent Anti-GPA33/Anti-CD3 Bispecific Antibody for Colorectal Cancers*. Mol Cancer Ther, 2018. **17**(10): p. 2164-2175.
- 77. Mathur, D., et al., A Novel GUCY2C-CD3 T-Cell Engaging Bispecific Construct (PF-07062119) for the Treatment of Gastrointestinal Cancers. Clin Cancer Res, 2020. **26**(9): p. 2188-2202.
- 78. Bacac, M., et al., A Novel Carcinoembryonic Antigen T-Cell Bispecific Antibody (CEA TCB) for the Treatment of Solid Tumors. Clin Cancer Res, 2016. **22**(13): p. 3286-97.
- 79. Zhao, L., et al., *Targeting CD133high Colorectal Cancer Cells In Vitro and In Vivo With an Asymmetric Bispecific Antibody.* J Immunother, 2015. **38**(6): p. 217-28.
- 80. Labrijn, A.F., et al., *Bispecific antibodies: a mechanistic review of the pipeline.* Nat Rev Drug Discov, 2019. **18**(8): p. 585-608.
- 81. Douglass, J., et al., Bispecific antibodies targeting mutant RAS neoantigens. Sci Immunol, 2021. **6**(57).
- 82. Hsiue, E.H., et al., *Targeting a neoantigen derived from a common TP53 mutation.* Science, 2021. **371**(6533).
- 83. Skora, A.D., et al., Generation of MANAbodies specific to HLA-restricted epitopes encoded by somatically mutated genes. Proc Natl Acad Sci U S A, 2015. **112**(32): p. 9967-72.
- 84. de Bruin, R.C.G., et al., A bispecific nanobody approach to leverage the potent and widely applicable tumor cytolytic capacity of  $V\gamma 9V\delta 2$ -T cells. Oncoimmunology, 2017. **7**(1): p. e1375641.
- 85. Mueller, T., et al., Efficacy of a Bispecific Antibody Co-Targeting VEGFA and Ang-2 in Combination with Chemotherapy in a Chemoresistant Colorectal Carcinoma Xenograft Model. Molecules, 2019. **24**(16).
- 86. Hidalgo, M., et al., First-in-Human Phase I Study of Single-agent Vanucizumab, A First-in-Class Bispecific Anti-Angiopoietin-2/Anti-VEGF-A Antibody, in Adult Patients with Advanced Solid Tumors. Clin Cancer Res, 2018. **24**(7): p. 1536-1545.
- 87. Xu, M., et al., *Hollow mesoporous ruthenium nanoparticles conjugated bispecific antibody for targeted anti-colorectal cancer response of combination therapy.* Nanoscale, 2019. **11**(19): p. 9661-9678.
- 88. Patnaik, A., et al., A phase I study of LY3164530, a bispecific antibody targeting MET and EGFR, in patients with advanced or metastatic cancer. Cancer Chemother Pharmacol, 2018. **82**(3): p. 407-418.
- 89. A Study of XmAb®22841 Monotherapy & in Combination w/ Pembrolizumab in Subjects w/ Selected Advanced Solid Tumors.
- 90. Chames, P., et al., *Therapeutic antibodies: successes, limitations and hopes for the future.* Br J Pharmacol, 2009. **157**(2): p. 220-33.
- 91. Moradi, A., et al., *Nanobody-based therapeutics against colorectal cancer: Precision therapies based on the personal mutanome profile and tumor neoantigens.* Pharmacol Res, 2020. **156**: p. 104790.
- 92. First in Human Trial of TAS266 in Patients With Advanced Solid Tumors. Available from: <a href="https://ClinicalTrials.gov/show/NCT01529307">https://ClinicalTrials.gov/show/NCT01529307</a>, <a href="https://www.novctrd.com/ctrdweb/trialresult/trialresults/pdf?trialResultId=9663">https://www.novctrd.com/ctrdweb/trialresult/trialresults/pdf?trialResultId=9663</a>.
- 93. αPD1-MSLN-CAR T Cells for the Treatment of MSLN-positive Advanced Solid Tumors.
- 94. Dadashi Noshahr, K., et al., *Optimization of post-insertion method to conjugate Doxil with anti-CD133 monoclonal antibodies: Investigating the specific binding and cytotoxicity to colorectal cancer cells in vitro.* Saudi Pharm J, 2020. **28**(11): p. 1392-1401.
- 95. Wrobel, P. and S. Ahmed, *Current status of immunotherapy in metastatic colorectal cancer.* Int J Colorectal Dis, 2019. **34**(1): p. 13-25.

- 96. Geevarghese, S.K., et al., *Phase I/II study of oncolytic herpes simplex virus NV1020 in patients with extensively pretreated refractory colorectal cancer metastatic to the liver.* Hum Gene Ther, 2010. **21**(9): p. 1119-28.
- 97. *pexastimogene devacirepvec*. Available from: <a href="https://www.cancer.gov/publications/dictionaries/cancer-drug/def/pexastimogene-devacirepvec">https://www.cancer.gov/publications/dictionaries/cancer-drug/def/pexastimogene-devacirepvec</a>.
- 98. Samsung Medical, C., A Trial of JX-594 in Refractory Colorectal Carcinoma. 2012.
- 99. Park, S.H., et al., *Phase 1b Trial of Biweekly Intravenous Pexa-Vec (JX-594), an Oncolytic and Immunotherapeutic Vaccinia Virus in Colorectal Cancer.* Mol Ther, 2015. **23**(9): p. 1532-40.
- 100. B., M.C.M., et al., A phase I/II study of Pexa-Vec oncolytic virus in combination with immune checkpoint inhibition in refractory colorectal cancer. Journal of Clinical Oncology, 2020. **38**(4\_suppl): p. 117-117.
- 101. Song, H., et al., *Application of Newcastle disease virus in the treatment of colorectal cancer.* World J Clin Cases, 2019. **7**(16): p. 2143-2154.
- 102. Fifis, T., et al., *Vaccination with in vitro grown whole tumor cells induces strong immune responses and retards tumor growth in a murine model of colorectal liver metastases.* Vaccine, 2008. **26**(2): p. 241-9.
- 103. Schmidt, W., et al., *Transloading of tumor cells with foreign major histocompatibility complex class I peptide ligand: a novel general strategy for the generation of potent cancer vaccines.* Proc Natl Acad Sci U S A, 1996. **93**(18): p. 9759-63.
- 104. de Weger, V.A., et al., Clinical effects of adjuvant active specific immunotherapy differ between patients with microsatellite-stable and microsatellite-instable colon cancer. Clin Cancer Res, 2012. **18**(3): p. 882-9.
- 105. Harris, J.E., et al., Adjuvant active specific immunotherapy for stage II and III colon cancer with an autologous tumor cell vaccine: Eastern Cooperative Oncology Group Study E5283. J Clin Oncol, 2000. **18**(1): p. 148-57.
- 106. Bartnik, A., A.J. Nirmal, and S.Y. Yang, *Peptide Vaccine Therapy in Colorectal Cancer.* Vaccines (Basel), 2012. **1**(1): p. 1-16.
- 107. George, D.J., et al., Survival Outcomes of Sipuleucel-T Phase III Studies: Impact of Control-Arm Cross-Over to Salvage Immunotherapy. Cancer Immunol Res, 2015. **3**(9): p. 1063-9.
- 108. Mirlekar, B. and Y. Pylayeva-Gupta, *IL-12 Family Cytokines in Cancer and Immunotherapy.* Cancers (Basel), 2021. **13**(2).
- 109. *Cytokines and Their Side Effects*. Available from: <a href="https://www.cancer.org/treatment/treatments-and-side-effects/treatment-types/immunotherapy/cytokines.html">https://www.cancer.org/treatment/treatments-and-side-effects/treatment-types/immunotherapy/cytokines.html</a>.
- 110. Chulpanova, D.S., et al., *Molecular Aspects and Future Perspectives of Cytokine-Based Anti-cancer Immunotherapy.* Front Cell Dev Biol, 2020. **8**: p. 402.
- 111. Akdis, M., et al., *Interleukins (from IL-1 to IL-38), interferons, transforming growth factor β, and TNF-α:* Receptors, functions, and roles in diseases. J Allergy Clin Immunol, 2016. **138**(4): p. 984-1010.
- 112. Rébé, C. and F. Ghiringhelli, *Interleukin-1* and Cancer. Cancers (Basel), 2020. **12**(7).
- 113. Baker, K.J., A. Houston, and E. Brint, *IL-1 Family Members in Cancer; Two Sides to Every Story.* Front Immunol, 2019. **10**: p. 1197.
- 114. Kienzl, M., et al., *IL*-33 reduces tumor growth in models of colorectal cancer with the help of eosinophils. Oncoimmunology, 2020. **9**(1): p. 1776059.
- 115. Parker, B.S., J. Rautela, and P.J. Hertzog, *Antitumour actions of interferons: implications for cancer therapy.* Nat Rev Cancer, 2016. **16**(3): p. 131-44.
- 116. Jorgovanovic, D., et al., *Roles of IFN-γ in tumor progression and regression: a review.* Biomark Res, 2020. **8**: p. 49.
- 117. Slattery, M.L., et al., *Interferon-signaling pathway: associations with colon and rectal cancer risk and subsequent survival.* Carcinogenesis, 2011. **32**(11): p. 1660-7.
- 118. Zaidi, M.R. and G. Merlino, *The two faces of interferon-γ in cancer*. Clin Cancer Res, 2011. **17**(19): p. 6118-24.
- 119. Zaidi, M.R., *The Interferon-Gamma Paradox in Cancer.* J Interferon Cytokine Res, 2019. **39**(1): p. 30-38
- 120. Hanada, T., et al., *IFNgamma-dependent, spontaneous development of colorectal carcinomas in SOCS1-deficient mice.* J Exp Med, 2006. **203**(6): p. 1391-7.
- 121. Wiesenfeld, M., et al., Controlled clinical trial of interferon-gamma as postoperative surgical adjuvant therapy for colon cancer. J Clin Oncol, 1995. **13**(9): p. 2324-9.

- 122. Study of Gamma Interfereon in Metastatic Colorectal Carcinoma. Available from: <a href="https://ClinicalTrials.gov/show/NCT00786643">https://ClinicalTrials.gov/show/NCT00786643</a>.
- 123. Qiu, Y., et al., *Clinical Application of Cytokines in Cancer Immunotherapy.* Drug Des Devel Ther, 2021. **15**: p. 2269-2287.
- 124. Jiang, M., et al., *cGAS-STING*, an important pathway in cancer immunotherapy. J Hematol Oncol, 2020. **13**(1): p. 81.
- 125. Chon, H.J., et al., *STING* signaling is a potential immunotherapeutic target in colorectal cancer. J Cancer, 2019. **10**(20): p. 4932-4938.
- 126. Chen, S.Y., et al., A STING-related prognostic score predicts high-risk patients of colorectal cancer and provides insights into immunotherapy. Ann Transl Med, 2021. **9**(1): p. 14.
- 127. Kim, D.S., et al., *E7766, a Macrocycle-Bridged Stimulator of Interferon Genes (STING) Agonist with Potent Pan-Genotypic Activity.* ChemMedChem, 2021. **16**(11): p. 1740-1743.
- 128. Liu, W.M., et al., *Inhibition of metastatic potential in colorectal carcinoma in vivo and in vitro using immunomodulatory drugs (IMiDs).* Br J Cancer, 2009. **101**(5): p. 803-12.
- 129. Goldstein, M.J., et al., *Phase I trial of thalidomide and capecitabine for treatment of metastatic colorectal cancer.* Journal of Clinical Oncology, 2006. **24**(18\_suppl): p. 13552-13552.
- 130. *CCR5-blockade in Metastatic Colorectal Cancer*. Available from: https://ClinicalTrials.gov/show/NCT01736813.
- 131. Mukaida, N., *CCR5 antagonist, an ally to fight against metastatic colorectal cancer.* Translational Cancer Research, 2016: p. S309-S312.
- 132. Kroschinsky, F., et al., New drugs, new toxicities: severe side effects of modern targeted and immunotherapy of cancer and their management. Crit Care, 2017. **21**(1): p. 89.
- 133. Armaghany, T., et al., *Genetic alterations in colorectal cancer.* Gastrointestinal cancer research : GCR, 2012. **5**(1): p. 19-27.
- 134. Wu, C.E., et al., Chromosomal Instability May Not Be a Predictor for Immune Checkpoint Inhibitors from a Comprehensive Bioinformatics Analysis. Life (Basel), 2020. **10**(11).
- 135. Walther, A., R. Houlston, and I. Tomlinson, *Association between chromosomal instability and prognosis in colorectal cancer: a meta-analysis.* Gut, 2008. **57**(7): p. 941-950.
- 136. Hamarsheh, S.a., et al., *Immune modulatory effects of oncogenic KRAS in cancer.* Nature Communications, 2020. **11**(1): p. 5439.
- 137. Liu, J., et al., *Immune landscape and prognostic immune-related genes in KRAS-mutant colorectal cancer patients.* Journal of Translational Medicine, 2021. **19**(1): p. 27.
- 138. Chen, N., et al., *KRAS mutation-induced upregulation of PD-L1 mediates immune escape in human lung adenocarcinoma.* Cancer Immunology, Immunotherapy, 2017. **66**(9): p. 1175-1187.
- 139. Zhang, L. and J.W. Shay, *Multiple Roles of APC and its Therapeutic Implications in Colorectal Cancer.* JNCI: Journal of the National Cancer Institute, 2017. **109**(djw332).
- 140. Grasso, C.S., et al., *Genetic Mechanisms of Immune Evasion in Colorectal Cancer*. Cancer Discov, 2018. **8**(6): p. 730-749.
- 141. Zhu, K., et al., *p53 induces TAP1 and enhances the transport of MHC class I peptides*. Oncogene, 1999. **18**(54): p. 7740-7747.
- 142. Zhao, P., et al., *Mismatch repair deficiency/microsatellite instability-high as a predictor for anti-PD-1/PD-L1 immunotherapy efficacy.* Journal of Hematology & Oncology, 2019. **12**(1): p. 54.
- 143. Hamada, T., et al., *TIME (Tumor Immunity in the MicroEnvironment) classification based on tumor CD274 (PD-L1) expression status and tumor-infiltrating lymphocytes in colorectal carcinomas.*Oncoimmunology, 2018. **7**(7): p. e1442999.
- 144. Kim, J.H., et al., *Characterisation of PD-L1-positive subsets of microsatellite-unstable colorectal cancers.* Br J Cancer, 2016. **115**(4): p. 490-6.
- 145. Marcus, L., et al., FDA Approval Summary: Pembrolizumab for the Treatment of Tumor Mutational Burden-High Solid Tumors. Clin Cancer Res, 2021. **27**(17): p. 4685-4689.
- 146. Overman, M.J., M.S. Ernstoff, and M.A. Morse, *Where We Stand With Immunotherapy in Colorectal Cancer: Deficient Mismatch Repair, Proficient Mismatch Repair, and Toxicity Management.* American Society of Clinical Oncology Educational Book, 2018(38): p. 239-247.
- 147. Samuels, Y., et al., *High Frequency of Mutations of the <i>PIK3CA</i> Gene in Human Cancers.* Science, 2004. **304**(5670): p. 554-554.

- 148. Nusrat, M., et al., Association of PIK3CA mutations (mut) with immune engagement and clinical benefit from immunotherapy in microsatellite stable (MSS) colorectal cancer (CRC) patients (pts). Journal of Clinical Oncology, 2019. **37**(15\_suppl): p. 3604-3604.
- 149. Nassif, N.T., et al., *PTEN mutations are common in sporadic microsatellite stable colorectal cancer.* Oncogene, 2004. **23**(2): p. 617-628.
- 150. Vidotto, T., et al., *Emerging role of PTEN loss in evasion of the immune response to tumours.* British Journal of Cancer, 2020. **122**(12): p. 1732-1743.
- 151. Lin, Z., et al., *PTEN loss correlates with T cell exclusion across human cancers.* BMC Cancer, 2021. **21**(1): p. 429.
- 152. Kim, B.-G., et al., *Novel therapies emerging in oncology to target the TGF-β pathway*. Journal of Hematology & Oncology, 2021. **14**(1): p. 55.
- 153. Bai, X., et al., Blocking TGF-β Signaling To Enhance The Efficacy Of Immune Checkpoint Inhibitor. Onco Targets Ther, 2019. **12**: p. 9527-9538.
- 154. Grady, W.M., et al., *Mutational Inactivation of Transforming Growth Factor β Receptor Type II in Microsatellite Stable Colon Cancers*. Cancer Research, 1999. **59**(2): p. 320-324.
- 155. Li, T., et al., *TGFBR2 mutation predicts resistance to immune checkpoint inhibitors in patients with non-small cell lung cancer.* Therapeutic Advances in Medical Oncology, 2021. **13**: p. 17588359211038477.
- 156. Hu, Y., et al., *Prognostic value of PD-L1 expression in patients with pancreatic cancer: A PRISMA-compliant meta-analysis.* Medicine (Baltimore), 2019. **98**(3): p. e14006.
- 157. Droeser, R.A., et al., *Clinical impact of programmed cell death ligand 1 expression in colorectal cancer.* Eur J Cancer, 2013. **49**(9): p. 2233-42.
- 158. Klein, S., et al., *Tumor infiltrating lymphocyte clusters are associated with response to immune checkpoint inhibition in BRAF V600E/K mutated malignant melanomas.* Scientific Reports, 2021. **11**(1): p. 1834.
- 159. Li, M., et al., An immune landscape based prognostic signature predicts the immune status and immunotherapeutic responses of patients with colorectal cancer. Life Sciences, 2020. **261**: p. 118368.
- 160. Correale, P., et al., Regulatory (FoxP3+) T-cell tumor infiltration is a favorable prognostic factor in advanced colon cancer patients undergoing chemo or chemoimmunotherapy. J Immunother, 2010. **33**(4): p. 435-41.
- 161. Tang, Y.-Q., et al., *The tumor immune microenvironment transcriptomic subtypes of colorectal cancer for prognosis and development of precise immunotherapy.* Gastroenterology Report, 2020. **8**(5): p. 381-389.
- 162. Tamura, K., et al., Characterization of the T cell repertoire by deep T cell receptor sequencing in tissues and blood from patients with advanced colorectal cancer. Oncol Lett, 2016. **11**(6): p. 3643-3649.
- 163. Liu, X., et al., A comprehensive study of immunology repertoires in both preoperative stage and postoperative stage in patients with colorectal cancer. Mol Genet Genomic Med, 2019. **7**(3): p. e504.
- 164. Lin, Y., J. Xu, and H. Lan, *Tumor-associated macrophages in tumor metastasis: biological roles and clinical therapeutic applications.* J Hematol Oncol, 2019. **12**(1): p. 76.
- 165. Zhang, S.Y., et al., *Tumor-associated macrophages: A promising target for a cancer immunotherapeutic strategy.* Pharmacol Res, 2020. **161**: p. 105111.
- 166. Pinto, M.L., et al., *The Two Faces of Tumor-Associated Macrophages and Their Clinical Significance in Colorectal Cancer.* Front Immunol, 2019. **10**: p. 1875.
- 167. Larionova, I., et al., *Tumor-Associated Macrophages in Human Breast, Colorectal, Lung, Ovarian and Prostate Cancers.* Front Oncol, 2020. **10**: p. 566511.
- 168. Ganesh, K., et al., *Immunotherapy in colorectal cancer: rationale, challenges and potential.* Nat Rev Gastroenterol Hepatol, 2019. **16**(6): p. 361-375.
- 169. Mager, L.F., et al., *Microbiome-derived inosine modulates response to checkpoint inhibitor immunotherapy*. Science, 2020. **369**(6510): p. 1481-1489.
- 170. Peng, Z., et al., *The Gut Microbiome Is Associated with Clinical Response to Anti-PD-1/PD-L1 Immunotherapy in Gastrointestinal Cancer.* Cancer Immunol Res, 2020. **8**(10): p. 1251-1261.
- 171. Choi, J., et al., *Diagnostic value of peripheral blood immune profiling in colorectal cancer.* Ann Surg Treat Res, 2018. **94**(6): p. 312-321.
- 172. Tang, Y.-p., et al., *Prognostic value of peripheral blood natural killer cells in colorectal cancer.* BMC Gastroenterology, 2020. **20**(1): p. 31.

- 173. Saied, A., et al., *Neutrophil:lymphocyte ratios and serum cytokine changes after hepatic artery chimeric antigen receptor-modified T-cell infusions for liver metastases.* Cancer Gene Ther, 2014. **21**(11): p. 457-62.
- 174. Reece, M., et al., *The Use of Circulating Tumor DNA to Monitor and Predict Response to Treatment in Colorectal Cancer.* Front Genet, 2019. **10**: p. 1118.
- 175. Chang, Y.C., et al., Exosomal Components and Modulators in Colorectal Cancer: Novel Diagnosis and Prognosis Biomarkers. Biomedicines, 2021. **9**(8).
- 176. Clerici, S.P., et al., Colorectal Cancer Cell-Derived Small Extracellular Vesicles Educate Human Fibroblasts to Stimulate Migratory Capacity. Front Cell Dev Biol, 2021. **9**: p. 696373.
- 177. Zhou, E., et al., Circulating extracellular vesicles are effective biomarkers for predicting response to cancer therapy. EBioMedicine, 2021. **67**: p. 103365.
- 178. Berzins, S.P., et al., A Role for MAIT Cells in Colorectal Cancer. Front Immunol, 2020. 11: p. 949.
- 179. Martin, T.D., et al., *The adaptive immune system is a major driver of selection for tumor suppressor gene inactivation.* Science, 2021. **373**(6561): p. 1327-1335.
- 180. Cancer Cells' Unexpected Genetic Tricks for Evading the Immune System. Available from: https://www.hhmi.org/news/cancer-cells-unexpected-genetic-tricks-evading-immune-system.
- 181. Koulouridi, A., et al., *Immunotherapy in Solid Tumors and Gut Microbiota: The Correlation-A Special Reference to Colorectal Cancer.* Cancers (Basel), 2020. **13**(1).
- 182. Saha, T., et al., *Intercellular nanotubes mediate mitochondrial trafficking between cancer and immune cells.* Nat Nanotechnol, 2021.
- 183. Carlsen, L., et al., *Pan-drug and drug-specific mechanisms of 5-FU, irinotecan (CPT-11), oxaliplatin, and cisplatin identified by comparison of transcriptomic and cytokine responses of colorectal cancer cells.* Oncotarget, 2021. **12**(20): p. 2006-2021.
- 184. Groysman, L., et al., *Chemotherapy-induced cytokines and prognostic gene signatures vary across breast and colorectal cancer*. 2021: American Journal of Cancer Research.
- 185. Huntington, K.E., et al., *A high-throughput customized cytokinome screen of colon cancer cell responses to small-molecule oncology drugs*. Oncotarget, 2021. **12**(20): p. 1980-1991.