Synaptic plasticity is altered by treatment with pharmacological levels of retinoic acid acting nongenomically however endogenous retinoic acid has not been shown to control synaptic plasticity

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Abstract

A paper recently published by eLife on forebrain cortical synaptic plasticity reports that retinoic acid (RA) alters synaptopodin-dependent metaplasticity in mouse dentate granule cells (Lenz et al., 2021). RA is the active form of vitamin A that functions as a ligand for nuclear RA receptors that directly bind genomic control regions to regulate gene expression (Chambon, 1996; Ghyselinck and Duester, 2019). However, some studies have suggested that RA may have nongenomic effects outside of the nucleus, particularly with regard to synaptic plasticity (Aoto et al., 2008; Zhang et al., 2018). The current results reported by Lenz et al. demonstrate that treatment with pharmacological levels of RA can alter synaptic plasticity which may be useful to treat neurological diseases (Lenz et al., 2021). However, the results reported here and those reported by others have not shown that endogenous RA is normally required for synaptic plasticity (or any other nongenomic effect) as there are no reports of genetic loss-of-function studies that remove endogenous RA in adult brain. The implication is that pharmacological levels of RA result in nongenomic effects, some of which may be helpful to treat certain diseases but in other cases this may cause unwanted side-effects.

Keywords

synaptic plasticity; retinoic acid; nongenomic action

Pharmacological Levels of RA Alter Synaptic Plasticity

Lenz et al. report that RA treatment affects forebrain cortical synaptic plasticity which modulates synaptic transmission to effectively respond to specific stimuli; specifically, they report that this effect occurs in the dorsal hippocampus but not ventral hippocampus and requires synaptopodin (Lenz et al., 2021). The method Lenz et al. used to determine RA function was to treat mice with 10 mg/kg RA. This dose of RA results in micromolar concentrations of RA in mouse tissues which is a teratogenic dose (Nau, 1995); RA is normally present in the nanomolar range in brain and other tissues (Kurlandsky et al., 1995). Such high levels of RA cannot be used to determine the normal function of RA due to unwanted side-effects. In fact, one cannot determine the normal in vivo functions of RA by

adding RA. Instead, one must take away RA preferably using RA genetic loss-of-function studies in vivo to be certain that RA is required for any proposed function (Cunningham and Duester, 2015).

The article by Lenz et al. follows up on a series of other articles suggesting that RA controls synaptic plasticity in a nongenomic fashion using a mechanism that does not involve regulation of gene expression by nuclear RA receptors, but instead involves cytoplasmic RA receptors that control mRNA translation or perhaps other nongenomic processes (Aoto et al., 2008; Arendt et al., 2015; Zhang et al., 2018; Hsu et al., 2019). However, although these previous articles suggest that endogenous RA normally controls synaptic plasticity, none report a genetic loss-of-function study to remove endogenous RA to see if it is required for synaptic plasticity via any mechanism. In order to do this in brain is difficult as several enzymes may participate in RA synthesis. However, it has been possible to eliminate RA synthesis in mouse fetal brain using knockouts of the RA-generating enzymes encoded by *Aldh1a2* or *Aldh1a3* (Chatzi et al., 2011); RA production has also been successfully eliminated in adult mouse cornea using a triple conditional knockout of *Aldh1a1*, *Aldh1a2*, and *Aldh1a3* (Kumar et al., 2017). Although sometimes very difficult, genetic loss-of-function studies are needed to determine the function of any gene, protein, or in this case a molecule such as RA.

Summary

As it stands now, Lenz et al. have provided convincing evidence that pharmacological levels of RA can alter synaptic plasticity in a synaptopodin-dependent fashion; they did not address whether endogenous RA normally controls synaptic plasticity (Lenz et al., 2021). Such information may be useful to explore treatment options for neurological disease. However, in order for the reader to be able to fully understand how these studies relate to normal RA signaling, it is useful to point out that future studies are needed to determine whether endogenous RA controls synaptic plasticity. In order to accomplish this goal, one would need to determine whether endogenous RA is present in the relevant tissues, determine where in the adult brain and under what conditions RA is generated, and then genetically knockout RA-generating enzymes to remove endogenous RA and determine if this

has an effect on synaptic plasticity. If so, then it would be relevant to determine whether the mechanism proceeds through nuclear RA receptors or in a nongenomic manner that involves cytoplasmic RA receptors or some other process. This is important as there currently are no RA genetic loss-of-function studies that support any nongenomic mechanism for RA.

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References

Aoto, J., Nam, C.I., Poon, M.M., Ting, P., and Chen, L. (2008). Synaptic signaling by all-trans retinoic acid in homeostatic synaptic plasticity. Neuron *60*, 308-320.

Arendt, K.L., Zhang, Z., Ganesan, S., Hintze, M., Shin, M.M., Tang, Y., Cho, A., Graef, I.A., and Chen, L. (2015). Calcineurin mediates homeostatic synaptic plasticity by regulating retinoic acid synthesis. Proc Natl Acad Sci USA *112*, E5744-5752.

Chambon, P. (1996). A decade of molecular biology of retinoic acid receptors. FASEB J *10*, 940-954.

Chatzi, C., Brade, T., and Duester, G. (2011). Retinoic acid functions as a key GABAergic differentiation signal in the basal ganglia. PLoS Biol *9*, e1000609.

Cunningham, T.J., and Duester, G. (2015). Mechanisms of retinoic acid signalling and its roles in organ and limb development. Nature Rev Mol Cell Biol *16*, 110-123.

Ghyselinck, N.B., and Duester, G. (2019). Retinoic acid signaling pathways. Development *146*, dev167502.

Hsu, Y.T., Li, J., Wu, D., Sudhof, T.C., and Chen, L. (2019). Synaptic retinoic acid receptor signaling mediates mTOR-dependent metaplasticity that controls hippocampal learning. Proc Natl Acad Sci USA *116*, 7113-7122.

Kumar, S., Dolle, P., Ghyselinck, N., and Duester, G. (2017). Endogenous retinoic acid signaling is required for maintenance and regeneration of cornea. Exp Eye Res *154*, 190-195.

Kurlandsky, S.B., Gamble, M.V., Ramakrishnan, R., and Blaner, W.S. (1995). Plasma delivery of retinoic acid to tissues in the rat. J Biol Chem *270*, 17850-17857.

Lenz, M., Eichler, A., Kruse, P., Muellerleile, J., Deller, T., Jedlicka, P., and Vlachos, A. (2021). All-trans retinoic acid induces synaptopodin-dependent metaplasticity in mouse dentate granule cells. eLife *10*, 01.

Nau, H. (1995). Chemical structure - Teratogenicity relationships, toxicokinetics and metabolism in risk assessment of retinoids. Toxicol Lett *82-83*, 975-979.

Zhang, Z., Marro, S.G., Zhang, Y., Arendt, K.L., Patzke, C., Zhou, B., Fair, T., Yang, N., Sudhof, T.C., Wernig, M.*, et al.* (2018). The fragile X mutation impairs homeostatic plasticity in human neurons by blocking synaptic retinoic acid signaling. Science Transl Med *10*, 01.