Case Report

Orientia tsutsugamushi and Epstein-Barr virus in a young adult with acalculous cholecystitis: a case report and epidemiology of scrub typhus in the Maldives


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Abstract: Scrub typhus is a neglected tropical disease predominantly occurring in Asia. The causative agent is a bacterium transmitted by the larval stage of mites found in rural vegetation in endemic regions. Cases of scrub typhus frequently present as acute undifferentiated febrile illness, and without early diagnosis and treatment, the disease can develop fatal complications. We retrospectively reviewed de-identified data from a 23-year-old woman who presented to an emergency department with complaints of worsening abdominal pain. On presentation, she appeared jaundiced and toxic-looking. Other positive findings on abdominal examination were a positive Murphy’s sign, abdominal guarding and hepatosplenomegaly. Magnetic resonance cholangiopancreatography demonstrated acalculous cholecystitis. Additional findings included eschar on the medial aspect of the left thigh with inguinal regional lymphadenopathy. Whether that represented acute infection or re-activation of the Epstein-Barr virus could not be determined. As other reports have described acute acalculous cholecystitis in adult scrub typhus patients, we recommend doxycycline to treat acute acalculous cholecystitis in endemic regions while awaiting serological confirmation.

Keywords: acute acalculous cholecystitis; Orientia tsutsugamushi; scrub typhus; eschar; Epstein-Barr virus; re-activation; clinical manifestation; Maldives
1. Introduction

Infectious diseases are well-known as a scourge of wars. Some infamous accounts include endemic typhus during the Franko-Russo war, trench fever in World War I, and scrub typhus during World War II [1]. The latter of these, scrub typhus, is caused by the rickettsial species *Orientia tsutsugamushi*, an intra-cytosolic coccobacillus with three main prototype strains (Karp, Gilliam and Kato) that can inflict disease of varying severities in humans [2].

The earliest description of scrub typhus appears to be an account of “shashitsu” in Chinese literature from 313 AD [3]. Centuries later, the word “tsutsugamushi” was used in Japan to describe a febrile illness following insect exposures. Tsutsugamushi was a feverish delirium associated with regional lymphadenopathy, rash and a pathognomonic cutaneous finding of tache noire or black spot, commonly referred to now as eschar [4].

The skin lesion of eschar is characterized by central necrosis and surrounding erythema with a halo sign [5]. The necrosis results from acidic enzymes channeled through a stylosome, a funnel created by the vector, gaining access via a skin pore or hair follicle [6]. This triggers a host response comprising infiltration of white blood cells into the dermis, causing the peripheral crusting of the scab.

In the Asia-Pacific theater of World War II, stringent vector control measures were employed to protect deployed troops from the parasitic larval stage of a chigger-mite found amongst the vegetation in endemic regions [7, 8]. Nevertheless, this task proved challenging during the pre-antibiotic era and case fatality reached up to 40%. In 1947, the first cases of successful treatment with chloromycetin were reported from the Malay Peninsula [9, 10].

However, despite modern availability of surefire antibiotics with bacteriostatic properties that stalled the bacterium *O. tsutsugamushi* and allowed phagocytes to clear up the infection, the disease still occurred after exposure to the vector. Escalating this problem is the geographic expansion of previously restricted endemic regions, and cases clinically compatible with scrub typhus or showing serological evidence have been reported from Chile, the United Arab Emirates and parts of Africa [11-18].

The Maldives is one endemic region for scrub typhus in Asia [5]. The archipelago has a peculiar historical account comprising a six-decade period with absence of the disease ended by re-emergence with 168 cases, including 10 fatalities [19]. Since that re-emergence in the Maldives, sentinel reporting of scrub typhus has shown occurrences throughout the year, with two periodic peaks observed during holiday seasons [20].

In tropical regions, coinfections with other tropical diseases are often observed, obfuscating the clinical characteristics of diseases [21]. This provides a major impediment to correct clinical judgment and often results in a poor clinical trajectory with missed diagnoses [22]. It has been previously reported that 15% of scrub typhus cases had coinfection with other diseases [23]. Complications often involve multiple systems and fatalities are often due to multi-organ dysfunction syndrome [24]. Commonly involved systems include the respiratory, central nervous, cardiovascular, renal, and gastrointestinal systems. However, reports of complications such as pancreatitis, acute cholecystitis, or acaulcus cholecystitis in scrub typhus are rare.

First described in 1844 by a surgeon, acute acaulcus cholecystitis (AAC) occurs in mainly critically ill geriatric patients. AAC also occurs postoperatively, and in patients requiring medical interventions such as mechanical ventilation or prolonged parenteral nutrition and fasting [25, 26]. This pathology accounts for 10% of all cases of acute cholecystitis and is associated with high morbidity and mortality rates [27, 28]. The pathophysiology involves inflammation of the gallbladder causing bile stasis and disruption of circulation causing ischemia and necrosis or rupture of the gallbladder, which then involves the peritoneum. In such cases, AAC is associated with infectious etiologies, mainly involving Gram-negative Enterobacteriaceae or Gram-positive spore-forming bacilli [29]. Interestingly, some other pathogens like Epstein-Barr virus and *O. tsutsugamushi* have been described to be associated with AAC in otherwise healthy
2. Materials and Methods

The case we described here presented to Indira Gandhi Memorial Hospital (IGMH) in Malé, Republic of Maldives, in July 2021. De-identified clinical and laboratory data during the hospitalization were reviewed using in-patient medical charts. Diagnostics for *O. tsutsugamushi* included the “Scrub Typhus Detect™” immunoglobulin (Ig)M/IgG enzyme-linked immunosorbent assay (InBios International, Seattle, WA, USA). Diagnostics for Epstein-Barr virus included VIDAS®EBV which detected Epstein-Barr viral capsid antigen (EBV-VCA) IgM/IgG, early antigen (EBV-EA) IgG, and nuclear antigen (EBV-NA) IgG by chemiluminescence assays (bioMérieux Italia, Florence, Italy). Hemoculture was performed using an automated culture system (bioMérieux, Durham, NC, USA). We also obtained epidemiological surveillance data for the preceding nine years (2012-2021) on scrub typhus in the Maldives from the Health Protection Agency at the Ministry of Health.

3. Case report

We present the case of a previously healthy 23-year-old woman who presented to the emergency department of IGMH with worsening abdominal pain and fatigue (Figure S1). The presenting complaint was associated with a 9-day history of high-grade fever accompanied by chills, and displaying a diurnal pattern. Four days before presentation, the patient had consulted a primary-care physician and had been prescribed amoxicillin and clavulanic acid for tonsillopharyngitis. Additional history included several episodes of vomiting with the onset of colicky, non-radiating abdominal pain of severe intensity, without any apparent aggravating or relieving factors. The patient had traveled from her home island of Gadhdhoo, in a southern atoll in the Maldives, to the capital city of Malé to visit friends and relatives a couple of days before falling ill, as illustrated in Figure S2. The annual descriptive epidemiology of scrub typhus in the Maldives is presented in Figure S3.

On examination, the patient was conscious and coherent, but appeared dehydrated, jaundiced, and toxic-looking. Oral temperature was 39°C, and heart rate was 110 beats/min with a mean arterial blood pressure of 73 mmHg. Both tonsils were enlarged with evidence of pharyngitis and palpable cervical lymphadenopathy. Evaluations of the cardiovascular, respiratory, and central nervous systems found no abnormalities. Other abdominal examinations revealed a positive Murphy’s sign and guarding on palpation of the abdomen. The results of laboratory investigations on presentation are shown in Table 1.
Table 1. Laboratory results at presentation and before discharge.

<table>
<thead>
<tr>
<th>Day of illness</th>
<th>9</th>
<th>14</th>
</tr>
</thead>
<tbody>
<tr>
<td>Leukocyte/µL</td>
<td>6,650</td>
<td>8,030</td>
</tr>
<tr>
<td>Neutrophils/µL</td>
<td>4,788</td>
<td>4,496</td>
</tr>
<tr>
<td>Lymphocytes</td>
<td>14,616</td>
<td>16,240</td>
</tr>
<tr>
<td>Monocytes/µL</td>
<td>1,340</td>
<td>3,265</td>
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<tr>
<td>Eosinophils/µL</td>
<td>0</td>
<td>37.5</td>
</tr>
<tr>
<td>Basophils/µL</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Platelets/µL</td>
<td>269,000</td>
<td>545,000</td>
</tr>
<tr>
<td>Hemoglobin (g/dL)</td>
<td>10.6</td>
<td>7.8</td>
</tr>
<tr>
<td>Hematocrit (%)</td>
<td>32.9</td>
<td>24.6</td>
</tr>
<tr>
<td>Total Bilirubin (mg/dL)</td>
<td>4.7</td>
<td>3.5</td>
</tr>
<tr>
<td>Direct Bilirubin (mg/dL)</td>
<td>3.5</td>
<td>2.6</td>
</tr>
<tr>
<td>Total protein (g/dL)</td>
<td>6.9</td>
<td>6.8</td>
</tr>
<tr>
<td>Albumin (g/dL)</td>
<td>3.4</td>
<td>2.8</td>
</tr>
<tr>
<td>Alkaline phosphatase (IU/L)</td>
<td>288</td>
<td>250</td>
</tr>
<tr>
<td>Aspartate aminotransferase (IU/L)</td>
<td>140</td>
<td>76</td>
</tr>
<tr>
<td>Alanine aminotransferase (IU/L)</td>
<td>157</td>
<td>93</td>
</tr>
<tr>
<td>Creatinine (mg/dL)</td>
<td>0.8</td>
<td>0.6</td>
</tr>
<tr>
<td>Urea (mg/dL)</td>
<td>10</td>
<td>6.4</td>
</tr>
<tr>
<td>CRP (mg/dL)</td>
<td>7.7</td>
<td>2.8</td>
</tr>
<tr>
<td>Sodium (mmol/L)</td>
<td>131</td>
<td>134</td>
</tr>
<tr>
<td>Potassium (mmol/L)</td>
<td>4.5</td>
<td>4.3</td>
</tr>
<tr>
<td>Ferritin (ng/mL)</td>
<td>1,299.3</td>
<td></td>
</tr>
<tr>
<td>LDH (IU/L)</td>
<td>726</td>
<td></td>
</tr>
<tr>
<td>EBV-VCA IgM (IU/mL)</td>
<td>63.6</td>
<td></td>
</tr>
<tr>
<td>EBV-VCA IgG (IU/mL)</td>
<td>89.9</td>
<td></td>
</tr>
<tr>
<td>EBV-EA IgG (IU/mL)</td>
<td>16.1</td>
<td></td>
</tr>
<tr>
<td>EBV-NA IgG (IU/mL)</td>
<td>574.0</td>
<td></td>
</tr>
<tr>
<td>Hemoculture</td>
<td>no growth</td>
<td></td>
</tr>
<tr>
<td>Orientia tsutsugamushi IgM</td>
<td>positive</td>
<td></td>
</tr>
<tr>
<td>Orientia tsutsugamushi IgG</td>
<td>positive</td>
<td></td>
</tr>
</tbody>
</table>

CRP: C-reactive protein, LDH: lactate dehydrogenase, EBV-VCA: Epstein-Barr virus viral capsid antigen, EBV-EA; Epstein-Barr virus early antigen, EBV-NA: Epstein-Barr virus nuclear antigen, IgM: immunoglobulin M, IgG: immunoglobulin G.

The initial hematological profile on presentation showed lymphocytosis and increased monocytes, although leukocytes, neutrophils, and platelets remained within normal ranges. Hyperbilirubinemia and elevated transaminases were identified from liver function tests. Further, hyperferritinemia and elevated levels of C-reactive protein were identified. Ultrasonography confirmed Murphey’s sign from the probe test and demonstrated edema of the gallbladder wall. Computed tomography (CT) of the abdomen revealed hepatosplenomegaly. Magnetic resonance cholangiopancreatography subsequently confirmed acalculous cholecystitis, as shown in Figure 1.
Figure 1. Imaging of the abdomen during hospitalization. a) Coronal-view CT of the abdomen shows hepatosplenomegaly. b) Axial-view CT of the abdomen demonstrates cholecystitis with enhancement of the gallbladder wall. c) Longitudinal ultrasonography depicts an edematous gallbladder wall (wall thickness > 4 mm) without lithiasis. d) Magnetic resonance cholangiopancreatography reveals a ballooned gallbladder without any ductal obstruction, but with signs of inflammation consistent with acalculous cholecystitis.

The patient was admitted with parenteral fluids to correct dehydration, broad-spectrum antibiotics (cefotaxime at 2 g/day, metronidazole at 1500 mg/day), and parenteral analgesics. During nursing care, eschar was noticed on the medial aspect of the left thigh (Figure S5). Left regional inguinal lymphadenopathy was also detected. Doxycycline was started at a loading stat dose of 200 mg followed by 200 mg/day for seven days. Fever cleared within 52 h of beginning doxycycline (Figure S6). Symptoms of abdominal pain gradually improved, and at the time of discharge from hospital, she had residual signs of fatigue.

4. Discussion

The first description of scrub typhus from the Maldives was reported during World War II [8]. By mid-1944, when the momentum in the war had shifted away from the battlefront following the historic invasion at Normandy, several hundred deployed British troops in the southern atolls of the Maldives fell victim to scrub typhus [42]. This prompted several entomological expeditions to the hotspot atolls identified in the Mal-
dives, such as Gaaf atoll, and the vector and reservoir local to the islands were identified [43, 44]. Reservoir eradication proved effective, with no subsequent cases reported. Scrub typhus was then simply forgotten until its sudden re-emergence in 2002 [5, 19, 45]. Since decades of sentinel data reflect the existence of *O. tsutsugamushi* throughout the archipelago, reports of scrub typhus from Faafu, Vaavu, and Meemu atolls are recent. We loosely associate this peculiar finding with urbanization, which could have exposed individuals to areas of vegetation where the vector is found [5].

The incubation of scrub typhus is 6–21 days [24]. Consequently, when obtaining a travel history, a minimum of three weeks is necessary to consider scrub typhus among the differential diagnoses. The patient in the present case was from an island within a hotspot for scrub typhus in the Maldives. However, she did not recall any particular activities that might have exposed her to the vector, such as lying on grassy vegetation or exploring tropical forest without proper preventive measures (such as use of insecticides, or covering exposed skin with long-sleeved clothes to avoid contact with insects). Nevertheless, the patient did confirm walking down a trail in a forest to get to the beach. The particular trails are narrow, with fringes of vegetation along both sides to the sandy beach. Interestingly, the patient also acknowledged knowing about the disease on the island through folklore of individuals developing fever after returning from the woods, and the eschar was locally referred to as “Gadhho bihi”, translating as “skin lesion from the island of Gadhho”. This reflects historical accounts of the endemicity of scrub typhus in the region.

Clinical manifestations of scrub typhus commonly appear first as undifferentiated febrile illness. During the early phase of disease, systemic symptoms are predominantly present without localizing foci of infection. Nevertheless, our patient presented with intermittent fever and she had complaints of sore throat and documented findings of tonsillopharyngitis from her initial visit to the primary-care physician. Similar manifestations had previously been described in scrub typhus patients, where misdiagnosis unfortunately led to complications from scrub typhus [46]. The exact cause of the intermittent fever observed in scrub typhus is unknown, but is likely to involve a pyrogen and countereffective cryogen production. Here, we assumed the exogenous pyrogen to be the peptidoglycan-like structure of *O. tsutsugamushi* and the cryogen to be interleukin 10 (IL-10) expressed by host cells [47-50]. Expression of IL-10 is one of the mechanisms utilized in immune evasion by the bacterium [51, 52].

Gastrointestinal (GI) symptoms occur in scrub typhus, but less frequently compared to systemic symptoms [53]. GI symptoms were reported in 20–76% of scrub typhus cases, with varying severity [54, 55]. As described in one cohort, the most common GI symptom was nausea, followed by abdominal pain, vomiting, and diarrhea [53]. The patient in the present case developed nausea during the third day after symptoms onset and experienced several episodes of vomiting with bile contents, up until the abdominal pain started. A correlation between endoscopic findings suggestive of gastrointestinal vasculitis and disease severity has previously been demonstrated [55]. *O. tsutsugamushi* shows tropism to several host cells, including endothelial cells with the pathogenicity of endo-theliitis [56]. Dissemination occurs within macrophages, and the bacterium has previously been detected in several organs, including in the human gallbladder [57].

Other studies have previously described hepatosplenomegaly, gallbladder or periportal edema and abdominal lymphadenopathy as GI findings visualized in patients with scrub typhus [58]. In the same cohort, splenomegaly was detected in 15 of 19 cases and 16% developed splenic infarcts due to acute enlargement of the spleen. The patient in our case presented also exhibited massive splenomegaly, but fortunately no splenic infarcts developed. In addition, the liver was enlarged. Animal studies have demonstrated hepatomegaly resulting from perivascular inflammation [59]. Similarly, thickening of the gallbladder wall can result from perivasculitis or vasculitis [35]. This thickening of the gallbladder wall was detected in 9 of 19 cases previously [58]. Presentation with acute abdomen in cases of scrub typhus is rare, but does occur as in this case, and has on occasion resulted in explorative surgeries [60].
The patient in this case showed the triad of fever, right upper quadrant pain, and clinical jaundice, suggestive of acute cholecystitis. Nevertheless, some hematological features like leukocytosis were absent from the hematological profile. In addition, no risk factors for cholecystitis were identified, such as alcohol consumption or evidence of cholelithiasis. With the identification of the eschar during nursing care, scrub typhus was suspected and doxycycline was started, and the fever cleared within two days. The patient was discharged from hospital five days later, once the abdominal pain had completely subsided. We were unable to obtain follow-up images showing resolution of gallbladder wall edema or a clinical specimen for molecular confirmation of *O. tsutsugamushi*, due to the retrospective nature of this report.

Interestingly, EBV was also detected from serology. The patients showed several features consistent with primary mononucleosis, including tonsillitis, adenopathy, hepatosplenomegaly, and lymphocytosis. In addition, no eruption of rash was identified in this case, as previously described [61] We could not determine the percentage increase in atypical lymphocytes, which would have helped distinguish primary mononucleosis from reactivation [62]. Nevertheless, others have also reported mononucleosis-like features in scrub typhus [63]. As mentioned in the Introduction, EBV infection can also complicate and manifest as acute cholecystitis without lithiasis. During the lytic phase, the virus can replicate in tonsillar tissues and hepatocytes and manifest with splenomegaly.

Scrub typhus is a neglected tropical disease and a public health concern. Increasing public awareness about prevention and educating clinicians beginning work in endemic regions about scrub typhus, among other common tropical diseases, could help detect cases, start treatment in a timely manner, and prevent complications and death[65].

In conclusion, scrub typhus can complicate and manifest as acute acalculous cholecystitis in young adults. We therefore recommend using doxycycline empirically in cases presenting from regions endemic for scrub typhus, providing no contraindications are present.

**Supplementary Materials:** The following are available online at www.mdpi.com/xxx/s1: Figure S1. Timeline depicting the clinical course prior to hospitalization; Figure S2. Past and recent epidemiology of scrub typhus in atolls of the Maldives; Figure S3. Two decades of sentinel reporting of scrub typhus in the Maldives; Figure S4. Eschar lesion on the medial aspect of the thigh; and Figure S5. Clinical course during hospitalization.


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**Institutional Review Board Statement:** The reporting of this case was conducted in accordance with the guidelines of the Declaration of Helsinki. An exemption to ethical approval was granted on September 29, 2021 (Ref: 137-B(NHA)/MISC/2021/045) by the National Healthcare Academy at Indira Gandhi Memorial Hospital, Malé, Republic of Maldives.
**Informed Consent Statement:** Informed consent was obtained from the patient involved in the study.

**Data Availability Statement:** The data presented in this study are available on request from the corresponding author. The data are not publicly available to ensure the privacy of the study participant.

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**Conflicts of Interest:** The authors declare no conflicts of interest.

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