

## **Precision public health through serological biomarkers: An integrated surveillance platform to inform public health interventions**

Kirsten E. Wiens<sup>1</sup>, Barbara Jauregui<sup>2</sup>, Benjamin F. Arnold<sup>3,4</sup>, Kathryn Banke<sup>5</sup>, Djibril Wade<sup>6</sup>, Kyla Hayford<sup>1</sup>, Adriana Costero-Saint Denis<sup>7</sup>, Robert H. Hall<sup>7</sup>, Henrik Salje<sup>8</sup>, Isabel Rodriguez-Barraquer<sup>9</sup>, Andrew S. Azman<sup>1,10,11</sup>, Guy Vernet<sup>2,12</sup>, Daniel T. Leung<sup>13\*</sup>, On behalf of the Collaboration on Integrated Biomarkers Surveillance<sup>^</sup>

<sup>1</sup> Johns Hopkins Bloomberg School of Public Health, Baltimore, United States

<sup>2</sup> Mérieux Foundation USA, Washington, DC, United States

<sup>3</sup> Francis I. Proctor Foundation, University of California, San Francisco, United States

<sup>4</sup> University of California, San Francisco, United States

<sup>5</sup> Bill & Melinda Gates Foundation, Seattle, WA, United States

<sup>6</sup> Institut de Recherche en Santé, de Surveillance Epidémiologique et de Formation (IRESSEF), Dakar, Senegal

<sup>7</sup> National Institute of Allergy and Infectious Diseases (NIAID), Bethesda, United States

<sup>8</sup> University of Cambridge, Cambridge, United Kingdom

<sup>9</sup> University of California, San Francisco, CA, USA

<sup>10</sup> Médecins Sans Frontières, Geneva, Switzerland

<sup>11</sup> University of Geneva, Geneva, Switzerland

<sup>12</sup> Institute Pasteur de Bangui, Bangui, Central African Republic

<sup>13</sup> University of Utah, Salt Lake City, United States

\* Corresponding author

E-mail: daniel.leung@utah.edu

^Membership of the Collaboration on Integrated Biomarkers Surveillance is provided in the Acknowledgements.

**Keywords:** serology, biomarkers, serosurveillance, sero-epidemiology, precision public health

## Abstract

The use of biomarkers to measure immune responses in serum is crucial for understanding population-level exposure and susceptibility to human pathogens. Advances in sample collection, multiplex testing, and computational modeling are transforming serosurveillance into a powerful tool for public health program design and response to infectious threats. In July 2018, 70 scientists from 20 countries met to perform a landscape analysis of approaches that support an integrated serosurveillance platform, including the consideration of issues for successful implementation. Here, we summarize the group's insights and proposed roadmap for implementation, including objectives, technical requirements, ethical issues, logistical considerations, and monitoring and evaluation.

## Introduction

Infectious diseases remain a major cause of morbidity and mortality worldwide. In 2019, 3.68 million deaths were attributable to tuberculosis and other respiratory infections, 1.75 million to enteric diseases, and 747 000 to malaria and neglected tropical diseases (NTDs) (1). The majority of this burden falls on low- and middle-income countries (LMICs) (1). The global spread of SARS-CoV-2 has further shown how all countries are deeply vulnerable to emerging and re-emerging infectious threats. Routine surveillance is a critical component of mitigating spread of these pathogens and depends largely on clinical and microbiological confirmation of infected individuals that seek testing or care. While these tools are valuable for identifying symptomatic cases, they say little about asymptomatic or non-medically attended infections or the population-level immune landscape. Serological surveys using biomarkers that measure

immune responses in serum (i.e. serosurveillance), combined with advances in computational modelling, provide an opportunity to bridge this gap (2,3).

The detection of immune responses in serum has been used for many years, but technological advances are transforming serosurveillance into a powerful tool for epidemiology, mathematical modeling, and public health program design. Sero-epidemiology has guided vaccination strategies for measles and rubella (4), informed vector-control strategies to reduce transmission of malaria (5), and guided tetanus elimination programs (6). Immunological biomarkers have been used to quantify community exposure to a broad range of pathogens, from antigenically-variable viruses such as dengue and chikungunya (7,8) to diarrheal diseases such as cholera (9). Antibodies against vector salivary proteins may also be useful for estimating human exposure to vector bites (10,11) and comparing the efficacy of different vector control strategies (12–14).

Despite the utility of serosurveillance, the costs and logistical challenges involved are prohibitive for comprehensive implementation, particularly in low-resource settings. Integrated serosurveillance systems that measure seroprevalence of multiple pathogens simultaneously could help overcome these barriers (15) and create opportunities to shift from vertical programs to integrative program delivery (16). Integrated platforms would reduce costs of serosurveillance as the cost of adding antigens to a multiplex assay is small compared to the cost of collecting specimen (15). Moreover, integrated platforms could provide a holistic understanding of co-circulating pathogens that contribute to population vulnerability, improving policymakers' ability to decide how to most effectively allocate limited resources.

Given the momentum built by these technological advances, a group of approximately 70 scientists from 20 countries gathered in Annecy, France in July 2018 for an Expert Meeting on sero-epidemiology organized by the Mérieux Foundation USA. This group, the Collaboration on Integrated Biomarkers Surveillance (CIBS), conducted a landscape analysis of existing technologies and approaches that support developing an integrated serosurveillance platform. Based on the results, CIBS established the objectives, technical requirements, ethical issues, logistical considerations, and funding that would be needed for such a platform. Here we summarize CIBS's insights and their proposed roadmap for implementation. Many of the areas for development overlap with recommendations recently released by the Pan American Health Organization (PAHO) for integrative serological surveillance in the Americas (17), as well as a recent review on elimination surveillance for neglected tropical diseases (NTDs) (18).

### Objectives of an integrated platform

The objectives of an integrated platform are twofold (Box 1). First, to identify use-cases for serosurveillance (e.g., identifying recent exposure versus immunity) and support the validation of serological biomarkers markers for each. Second, to develop a serosurveillance system, using these biomarkers, to provide actionable health outcome measures for interventions.

To effectively meet these objectives, the platform should include international, regional, and national components. A generic platform model should be created at the international level that countries could adapt to their national priorities and needs. This would include standard guidelines, operating procedures, training modules, as well as technical support when needed, and would create a global avenue for inter-platform collaboration and exchange of experiences

and practices. Biomarkers for specific pathogens and use-cases would need to be validated at the regional or international level, as is appropriate and feasible. The platform would support these efforts by defining the minimum characteristics of appropriate biomarker validation studies and maintaining biorepositories of gold standard samples for use in these studies. At the national level, the platform would provide feedback on community advocacy, funding sources, setting up immunological assays, designing sampling frames, selecting biomarkers, organizing logistics, and analyzing the data (Box 1).

**Box 1. Objectives of an integrated platform.**

Specific objectives of an integrated platform
<p><b>1) Define use-cases and identify serological biomarkers</b></p> <ul style="list-style-type: none"><li>a) Define use-cases for different pathogens and objectives (e.g., recent exposure, cumulative exposure, or susceptibility of the population)</li><li>b) Identify potential biomarkers for each use-case, including published and experimental biomarkers</li><li>c) Define minimum characteristics of appropriate biomarker validation studies</li><li>d) Maintain biorepositories of and/or access to international standards (reference reagents) that can be used during biomarker development</li></ul> <p><b>2) Develop serosurveillance systems</b></p> <ul style="list-style-type: none"><li>a) Provide support to countries in identifying funding sources and supplies</li><li>b) Provide guidance for setting up immunological assays</li><li>c) Provide feedback and protocols for designing a sampling frame, conducting community advocacy, and demographic and clinical data collection</li><li>d) Provide feedback on organizing logistics and transportation</li><li>e) Support development of analytical frameworks for integrating serological and epidemiologic data to translate test results into actionable information</li></ul>

Ultimately, the platform would serve as a public health resource for sero-epidemiology that informs vaccine campaigns, prophylactic treatments, and other infection control strategies focused on improving the health of the most vulnerable populations. By providing information on a regular basis, it could also enable monitoring the impact of these programs.

## Pathogens

Depending on use-cases, the platform could test biomarkers that measure seroprevalence or recent exposure for a broad range of blood-borne, enteric, respiratory, and vector-borne infections (Table 1). For some pathogens, serological biomarkers may additionally be useful for estimating incidence rates, cumulative infection rates, and correlates of protection, among other applications (Table 1). The performance characteristics (sensitivity and specificity) of relatively few serological markers for serosurveillance have been established to date. Therefore, initial versions of the platform would include validated and experimental markers, with a focus on priority pathogens as defined by the implementing country.

**Table 1. Pathogens to be considered for an integrated platform.** Pathogens that could be considered for an integrated platform are listed and grouped by primary source of infections. Ways in which sero-epidemiology has previously been used in surveillance of each pathogen are indicated and accompanied by published examples, including both reviews and primary research articles.

Primary source of infection	Pathogen for consideration in an integrated platform	Incidence estimates from cross-sectional data	Cumulative infection rate estimates (lasting/saturating Abs)	Vaccine vs. natural infection potentially discernible	Cross-sectional correlates of protection	Used for confirming elimination	Included on Luminex bead assays
Blood and/or other bodily fluids	<i>Chlamydia trachomatis</i>					(19)	(15,17,20,21)
	Ebola virus						(15)
	Hepatitis B virus			(2)	(22)		

	Hepatitis C virus						
	HIV	(23–27)					(15)
	<i>Neisseria meningitidis</i>						
Food, water, and/or soil	<i>Campylobacter jejuni</i>						(28)
	<i>Clostridium tetani</i>		(29)	(2)	(30)	(31)	(6,15,17)
	<i>Cryptosporidium parvum</i>						(15,17,28)
	Enterotoxigenic <i>Escherichia coli</i>						(28)
	<i>Giardia intestinalis</i>						(15,17,28)
	Hepatitis A virus		(32)		(22)		
	Hepatitis E virus		(33)				(34)
	Lassa virus						(15)
	Norovirus						(28)
	Poliovirus				(35)		
	<i>Salmonella enterica</i> serotype enteritidis	(36,37)					(15,28)
	<i>Salmonella enterica</i> serotype typhimurium						(15)
	<i>Schistosoma mansoni</i>					(38)	(15,39)
	<i>Shigella</i>						
	<i>Strongyloides stercoralis</i>					(40)	(15,17,39)
	<i>Taenia solium</i>						(15,17)
	<i>Toxoplasma gondii</i>	(41)					(17)
	<i>Vibrio cholerae</i>	(9,42)			(43,44)		(28)
Respiratory droplets and/or aerosols	<i>Bordetella pertussis</i>	(45)			(22,46)		(15)
	<i>Corynebacterium diphtheriae</i>		(47)		(47)		(15,17)
	<i>Haemophilus influenzae</i> B				(22,30)		
	Measles		(2)		(2)		(15,17)
	Mumps		(48)		(48)		(15,17)
	Respiratory syncytial virus						
	Rhinoviruses						
	Rubella		(2)		(2)		(15,17)
	SARS-CoV-2			(49)			(50,51)
Arthropod vectors	Chikungunya virus	(52)	(7,8)		(53)		(15,54)
	Crimean-Congo hemorrhagic fever virus						(15)
	Dengue virus	(55,56)	(57)				(15)
	Mayaro virus	(58)	(57)				(58)



	<i>Onchocerca volvulus</i>					(38)	(15)
	<i>Plasmodium falciparum</i> & <i>Plasmodium vivax</i>	(5,59,60)					(15,17,39)
	Vector saliva antigens						
	<i>Wuchereria bancrofti</i> & <i>Brugia malayi</i>					(38)	(15,17,39)
	Yellow fever virus						
	Zika virus		(57,61)				

### Study population

The study population will also depend on specific pathogens and use-cases (e.g., estimating force of infection, seroprevalence, or population susceptibility; see Table 1 for examples). This is an area where an integrative platform would be instrumental for providing guidance and sharing expertise. For example, to estimate incidence rates for endemic pathogens that infect individuals from a young age, such as many NTDs and enteric pathogens, measuring serological responses in children may be important to capture differences age-specific seroprevalence that might plateau in older age-groups (15,28). In contrast, teens and adults are more relevant for serosurveillance of pathogens such as HIV, with efforts to sample high-risk groups that may be less likely to be sampled in traditional study designs (15). For integrated surveillance of pathogens that require measurements in different age groups, initial population-based surveys could be conducted across a wide age range, followed by more targeted, adaptive surveys that focus on disease- or program-specific use-cases.

Timing of surveys would also depend on the biomarkers included and specific use-cases. An annual survey would be sufficient for studying long-lasting antibody responses to pathogens such

as measles or rubella, while biannual surveys would be required for antibodies with shorter life such as *Vibrio cholerae* and other enteric pathogens. For vaccine-preventable diseases, immunization schedules or campaigns would also need to be taken into consideration.

### Ethical considerations

Collecting biological specimen and socio-demographic data for research purposes requires careful ethical review and clearance through institutional review boards. Clear information for participants prior to obtaining formal consent is required. Risks including safety issues, even if minimal, need to be considered, as well as any socio-cultural differences. Rules for ownership of data and specimen should be established, with efforts made to process specimen locally within the implementing country when possible and to build this capacity when it is not. Intellectual property on research conducted as part of the program must consider rights of the countries from which the specimens originate and provide guarantees that any analytical and/or laboratory surveillance tools derived from the research will be made available in the country.

### Community and stakeholder engagement

Identifying all relevant community stakeholders and engaging with them is critical for the successful implementation of any new program. Effective community and stakeholder engagement (CSE) requires dedicated funding, sponsor commitments, and moral support. It requires engaging a broad range of individual stakeholders, including women, frontline healthcare workers, and teachers, among many others, who may not be all represented on, for example, community advisory boards. Engagement needs to emphasize listening to stakeholders, identifying opportunities for deliberation, and developing relationships and conditions needed to

integrate the program into existing health infrastructure, where results warrant. It also requires making sure that the research has a direct and timely impact on public health programs in the surveyed community. Finally, any CSE strategy needs to be implemented in a way that allows for meaningful evaluation of its effectiveness.

### Specimen collection and testing

The most appropriate specimen will depend on the scale of the survey and resources available. For cultural and practical reasons, urine, cerebrospinal fluid, and throat and nose swabs are often not easily accessible. While saliva is the most practical specimen for large epidemiological surveys, oral fluid assays have historically had lower sensitivity than comparable blood-based assays. Although, a recent study showed that saliva-based tests have similar performance to plasma-based tests for SARS-CoV-2 (50,62), suggesting that utility of saliva-based tests may be pathogen-specific. Overall, blood remains the most reliable specimen for biomarker detection. However, venipuncture is an invasive procedure that requires specific training, generates substantial biohazardous waste, and requires transporting blood tubes safely in below zero conditions. Collecting capillary blood through dried blood spots (DBS) provide a more scalable alternative. DBS show comparable antibody measurements to serum samples for falciparum malaria, some bacterial and protozoal pathogens, and numerous viral pathogens, including vaccine-preventable diseases (63).

DBS also provide flexibility in testing locations. DBS may be used on site in rapid lateral-flow assays. DBS can be transported to remote sites for testing with more resource-intensive methods such as multiplex immunoassays. They can be kept at cool or ambient temperatures for several

weeks before they are frozen for long-term storage (63), as long as high temperatures are avoided. Serum Separator cards can be used to automatically separate serum from DBS, further reducing the effort required to process these samples (64).

While DBS allow several markers to be tested from a low volume of blood within a single sample, there is no standard platform or procedure for running and vetting results from multiplex antibody assays using DBS. In addition, platforms such as Luminex that are used for running multiplex assays are often only available in national or regional labs and require regular calibration and use of positive controls for consistency. Both individual and multiplex assays will need to be compared and validated before use in an integrated platform, including comparisons between DBS and venous blood samples. It will also be important for these protocols and methods to be shared through the platform, especially as new technologies (e.g., rapid or point of care tests, microarrays, fieldable instruments, phage-display approaches) become available.

### Logistics and resources

As described above, there will be various logistical challenges in implementing new serological surveys. Central reference labs could help with adoption, dissemination, and local capacity building. Public-private initiatives could also be leveraged. Within Africa, engaging the Africa CDC and WHO-AFRO will be important to ensure shared vision across the continent. Importantly, care should be taken that these efforts do not divert budgets and skilled technicians from the health care system.

One way to address logistical challenges is to integrate the platform within existing active and passive surveillance systems (17,18). Existing surveys that could be leveraged to accommodate multiplex testing include the Demographic and Health Survey (DHS), Malaria and AIDS Indicator Surveys, and NTD transmission assessment surveys (15), though the latter are often targeted to narrow geographic units and ages. Another potential source is remnants of samples from routine blood draws, which have been used for estimating SARS-CoV-2 seroprevalence (65). The most appropriate survey or surveillance tool will depend on timing of the surveys within each country and, importantly, on continued sources of funding.

The CIBS and an International Coordinating Center (ICC) could also provide guidance and oversight through coordination with a National Survey Program (NSP) (Box 2). In this framework, the NSP could be part of the Ministry of Health and would coordinate all activities in country and liaise with survey staff and researchers. Survey staff would include community relays or public health workers (participant recruitment, demographic questionnaires, GPS positioning, incentives distribution, logistics, and feedback), community health workers (specimen collection, participants information, and feedback), and regional and central laboratory personnel. Support from local health authorities would be critical for this type of program, and funding would need to be secured at international and national levels (16), with comprehensive roadmaps developed, including paths to sustainability.

**Box 2. Functions of an international coordinating committee**

Functions of the International Coordinating Committee (ICC)
✓ Oversight of program,

- ✓ Mobilize scientific expertise,
- ✓ Prepare program documentation,
- ✓ Secure funding,
- ✓ Provide guidance for assay validation,
- ✓ Oversee quality control activities,
- ✓ Centralize and share data,
- ✓ Centralize and dispatch specimens to specialized and research labs,
- ✓ Coordinate data analysis,
- ✓ Establish general and country/community-specific guidelines,
- ✓ Provide feedback to national disease programs,
- ✓ Organize international communication and dissemination of results.

In addition to logistical challenges described above, appropriate supply chains need to be developed to ensure availability of data collection and transport devices, including their transport into communities. National public health labs could take the lead in these efforts. Since purchasing these tools may be difficult in many settings, existing resources should be evaluated, strengthened, and used, where possible. When these tools are not available, alternative solutions/organizations could be identified through the NSP. This is another area where an integrated platform could provide critical support by creating opportunities for resource sharing between existing programs.

### Monitoring and evaluation

Monitoring and evaluation are critical components of any new surveillance program to ensure effective use of resources. We propose two key areas for evaluation. First, pilot studies to assess

the feasibility of a new platform, including proficiency testing with blinded test samples. Among criteria considered should also be the degree of community knowledge regarding disease prevention and intervention. Second, analyses of whether results from integrated surveillance studies led to a change in policy (e.g., whether a new program was started or changed, whether it triggered an intervention, or changed a clinical diagnosis) and whether it helped improve understanding of disease patterns. Long term, changes in disease patterns and reductions in disease burden should be evaluated by the implementing country.

As sero-epidemiology is resource-intensive, an additional area for evaluation is cost-effectiveness of preventing outbreaks. The potential savings of identifying immunity gaps and tailoring interventions should be modelled to evaluate whether investment in, for example, additional vaccination, would be a better use of funds.

### Advocacy

Local advocacy will be essential. Guidelines regarding communicating data that are considered sensitive by local and national authorities need to be established. Potential stigmatization of communities that fail to efficiently implement interventions should be considered. For advocacy efforts to be successful, they must actively engage Ministries of Health, national disease programs, political authorities, community leaders, civil society, and religious authorities.

When disease burden reduction has been achieved, it may be difficult to justify asking health authorities and communities to give blood or to spend additional limited resources. Thus, the

monitoring and evaluation approaches described above will be critical to evaluate the continued utility of an integrated platform and advocate for funding as needed.

Evidence-based arguments supporting the efficiency and cost-effectiveness of integrated surveys will also support advocacy internationally. Given the global interrelatedness of old and new emerging infectious diseases, there is a critical need to have well-coordinated responses (66). In this capacity, the WHO plays an essential role supporting national public health programs. Partners such as the Mérieux Foundation, the Global Fund, GAVI, and the BMGF should work with the WHO and local partners to further strengthen public health laboratory performance. Such organizations also have the financial and logistical resources to support a strong biobank, which would increase the impact of an integrated platform.

### Areas for innovation

Innovation is needed at several levels to successfully implement an integrated platform, as is research funding to support these efforts. Technologically, new biomarkers, existing biomarkers, and combinations of biomarkers need to be identified and validated. It will also be important to evaluate the best specimen for broad application, including new devices that reduce pain, increase acceptance by participants, and improve ease of storage and transportation. These devices must address multi-parameter testing from a single specimen, safe storage in degraded conditions (temperature, dust, moisture), space in transport packages, and cost.

Innovation is also needed to address logistical and resources issues. New technologies such as drones may be useful to transport specimen and supplies to and from remote locations (67).



Existing transportation capacities such as commercial companies involved in persons or goods transportation, pharmaceutical distribution, or other surveillance schemes should be evaluated.

Innovation in study design and analysis is also critical for defining pathogen priorities and sampling frames, as well as providing clear results and recommendations to disease programs. As in any survey, it will be important to carefully consider epidemiological components (e.g., age, sex, family environment, geographical environment, sample size, and GPS positioning). For an integrated platform, study designs must also be harmonized and optimized across diverse disease and surveillance priorities. Analysis pipelines that create informative results from a single assay for various diseases need to be developed, such as disease burden maps that overlay high burden populations for multiple pathogens simultaneously. Digital health solutions could help overcome some of these challenges. For example, tools like rapid diagnostic tests linked to cloud servers (68) could be developed for sending test results to a national dashboard, providing real-time disease maps and trends to health authorities.

## Conclusions

If implemented effectively, integrated serosurveillance platforms have the potential to dramatically expand our knowledge of pathogens circulating in populations locally and worldwide. In this paper, we have described gaps that must be filled for this type of platform to be feasible and effective. Biomarkers must be identified and validated, and technologies with sufficient sensitivity and specificity developed. Methods to select survey populations and analyze data from diverse pathogens must be optimized to inform disease-specific priorities and use-cases. Innovative specimen collection and transport methods are needed. Surveillance efforts

must be scalable and cost-effective, and there are numerous logistical issues that will need to be addressed in the field. Conducting studies in human populations necessitates addressing ethical issues and engaging with public health authorities and diverse members of civil society. Finally, this effort must have sustained funding.

The current COVID-19 pandemic has enabled advancement in many of these areas (69). It has also highlighted the importance of integrating serological and other types of surveillance data across human and animal health programs for preventing and controlling disease emergence. Given this momentum and the importance of integrated surveillance systems for responding to future infectious threats, the time is now to move forward with filling in the remaining gaps. Ultimately, this will enable better, more comprehensive data that can be used for designing interventions to reduce the burden of endemic and emerging diseases.

### Acknowledgements

The following are members of the Collaboration on Integrated Biomarkers Surveillance, who attended the meeting titled, “Expert Meeting on a platform for biomarker-based surveillance in communities to guide disease control interventions,” on July 25 to 27, 2018, at Les Pensieres Center for Global Health, Veyrier-du-Lac, France: Jon Andrus (George Washington University, Washington, DC, USA), Ben Arnold (UC Berkeley, Berkeley, CA, USA), Lawrence Ayong (Centre Pasteur du Cameroun, Yaoundé, Cameroon), Andrew Azman (Johns Hopkins University, Baltimore, MD, USA), Christine Bain (Advanced Biosciences Laboratory, Lyon, France), Kathryn Banke (Bill and Melinda Gates Foundation, Seattle, WA, USA), Bob Black (Johns Hopkins University, Baltimore, MD, USA), Sarah Browne (FDA, Silver Spring, MD,

USA), Juliet Bryant (Fondation Mérieux, Lyon, France), AC Camacho (DTRA, Washington, DC, USA), Jean-Sebastien Casalengo (Université Claude Bernard, Lyon, France), Adriana Costero Saint Denis (NIH, Rockville, MD, USA), Jane Cunningham (WHO, Geneva, Switzerland), Sabine Dittrich (FIND, Geneva, Switzerland), Marc Essodaigui (Advanced Biosciences Laboratory, Lyon, France), Alison Evarts (Merieux Foundation, Washington, DC, USA), Matthew Ferrari (Penn State University, University Park, PA, USA), Lia Florey (USAID, Washington, DC, USA), Dean Garrett (ICF, Silver Spring, MD, USA), Françoise Gay-Andrieu (bioMérieux, Marcy-l'Etoile, France), Bryan Greenhouse (UCSF, San Francisco, CA, USA), Robert Hall (NIH, Rockville, MD, USA), Kyla Hayford (Johns Hopkins University, Baltimore, MD, USA), Shelley Hossenlopp (Spot-On-Sciences, Austin, TX, USA), Rolf Kramer (ECDPC, Stockholm, Sweden), Daniel Leung (University of Utah, Salt Lake City, UT, USA), Nafissatou Leye (IRESSEF, Dakar, Senegal), Francisco Luquero (Epicentre, Geneva, Switzerland), Ivalda Macicame (NIH, Maputo, Mozambique), Yuka Manabe (Johns Hopkins, Baltimore, MD, USA), Henshaw Mandi (CEPI, Oslo, Norway), Christine Markwalter (Duke University, Durham, NC, USA), Anne Martin (Akros, Lusaka, Zambia), Elhadji Mbaye (IRESSEF, Dakar, Senegal), Souleymane Mboup (IRESSEF, Dakar, Senegal), David McGregor (LSHTM, London, UK), Martin Mengel (GMX, Valencia, Spain), Mark Miller (NIH, Bethesda, MD, USA), Michael Mina (Harvard School of Public Health, Cambridge, MA, USA), Marie Moroso (Fondation Mérieux, Lyon, France), Ivo Mueller (The Walter and Eliza Hall Institute of Medical Research, Parkville, Australia), Abdoulaye Nikiema (ASLM, Addis-Abeba, Ethiopia), Berthe Marie Njanpop (Paris, France), Tom Nutman (NIH, Rockville, MD, USA), David Olson (WHO, Geneva, Switzerland), Emily Penrose (Merieux Foundation USA, Washington, DC, USA), Jessica Radzio-Basu (Penn State University, University Park, PA, USA), Olivier Raynaud (Bill

and Melinda Gates Foundation, Seattle, WA, USA), Franck Remoue (IRD, Montpellier, France), Isabel Rodriguez-Barraquer (UCSF, San Francisco, CA, USA), Kolawole Salami (CEPI, Oslo, Norway), Amadou Sall (Institut Pasteur, Dakar, Senegal), Richard Schoske (DTRA, Washington, DC, USA), William Evan Secor (CDC, Atlanta, GA, USA), Yvan Sergeant (Quanterix, Belgium), Shash Shashidhar (Penn State University, University Park, PA, USA), Daniel Sikkema (Quanterix, USA, Lexington, MA, USA), Benjamin Svartzkopf (Luminex, Austin, TX, USA), Kay van der Horst (Penn State University, University Park, PA, USA), Fiona van der Klis (RIVM, Utrecht, the Netherlands), Jessica Vanhomwegen (Institut Pasteur, Paris, France), Guy Vernet (Merieux Foundation USA, Washington, DC, USA), Djibril Wade (IRESSEF, Dakar, Senegal), Joseph Wu (School of Public Health, The University of Hong Kong, Hong Kong SAR, China), Lindsey Wu (LSHTM, London, UK)

## Funding

The “Expert Meeting on a platform for biomarker-based surveillance in communities to guide disease control interventions,” was organized and supported by the Mérieux Foundation USA, with additional support from the Bill & Melinda Gates Foundation, the Coalition for Epidemic Preparedness Innovations (CEPI), the U.S. Defense Threat Reduction Agency (DTRA), Luminex Corporation, and Spot On Sciences. The funders had no role in the preparation or the decision to submit the work for publication.

## Biographical Sketch

Dr. Wiens is a postdoctoral fellow in the Department of Epidemiology at the Johns Hopkins Bloomberg School of Public Health. Her research interests include seroepidemiology and infectious disease dynamics and control.

## Address for Correspondence

Daniel T. Leung, Division of Infectious Diseases, University of Utah, 26 North Medical Drive, Wintrobe 517, Salt Lake City, UT 84132 USA

## References

1. Vos T, Lim SS, Abbafati C, Abbas KM, Abbasi M, Abbasifard M, et al. Global burden of 369 diseases and injuries in 204 countries and territories, 1990–2019: a systematic analysis for the Global Burden of Disease Study 2019. *The Lancet*. 2020 Oct 17;396(10258):1204–22.
2. Metcalf CJE, Farrar J, Cutts FT, Basta NE, Graham AL, Lessler J, et al. Use of serological surveys to generate key insights into the changing global landscape of infectious disease. *The Lancet*. 2016 Aug 13;388(10045):728–30.
3. Van Kerkhove MD, Ferguson NM. Epidemic and intervention modelling – a scientific rationale for policy decisions? Lessons from the 2009 influenza pandemic. *Bull World Health Organ*. 2012 Apr 1;90(4):306–10.

4. Mao B, Chheng K, Wannemuehler K, Vynnycky E, Buth S, Soeung SC, et al. Immunity to polio, measles and rubella in women of child-bearing age and estimated congenital rubella syndrome incidence, Cambodia, 2012. *Epidemiol Infect.* 2015 Jul;143(9):1858–67.
5. Drakeley CJ, Corran PH, Coleman PG, Tongren JE, McDonald SLR, Carneiro I, et al. Estimating medium- and long-term trends in malaria transmission by using serological markers of malaria exposure. *PNAS.* 2005 Apr 5;102(14):5108–13.
6. Scobie HM, Mao B, Buth S, Wannemuehler KA, Sørensen C, Kannarath C, et al. Tetanus immunity among women aged 15 to 39 years in Cambodia: a national population-based serosurvey, 2012. *Clinical and Vaccine Immunology.* 23(7):546–54.
7. Fritzell C, Rousset D, Adde A, Kazanji M, Kerkhove MDV, Flamand C. Current challenges and implications for dengue, chikungunya and Zika seroprevalence studies worldwide: A scoping review. *PLOS Neglected Tropical Diseases.* 2018 Jul 16;12(7):e0006533.
8. Li Z, Wang J, Cheng X, Hu H, Guo C, Huang J, et al. The worldwide seroprevalence of DENV, CHIKV and ZIKV infection: A systematic review and meta-analysis. *PLOS Neglected Tropical Diseases.* 2021 Apr 28;15(4):e0009337.
9. Azman AS, Lessler J, Luquero FJ, Bhuiyan TR, Khan AI, Chowdhury F, et al. Estimating cholera incidence with cross-sectional serology. *Science Translational Medicine [Internet].* 2019 Feb 20 [cited 2020 Oct 7];11(480). Available from: <https://stm.sciencemag.org/content/11/480/eaau6242>
10. Mathieu-Daudé F, Claverie A, Plichart C, Boulanger D, Mphande FA, Bossin HC. Specific human antibody responses to *Aedes aegypti* and *Aedes polynesiensis* saliva: A new

- epidemiological tool to assess human exposure to disease vectors in the Pacific. *PLOS Neglected Tropical Diseases*. 2018 Jul 24;12(7):e0006660.
11. Doucoure S, Mouchet F, Cournil A, Goff GL, Cornelié S, Roca Y, et al. Human antibody response to *Aedes aegypti* saliva in an urban population in Bolivia: a new biomarker of exposure to dengue vector bites. *The American Journal of Tropical Medicine and Hygiene*. 2012 Sep 5;87(3):504–10.
  12. Noukpo MH, Damien GB, Elanga-N'Dille E, Sagna AB, Drame PM, Chaffa E, et al. Operational Assessment of long-lasting insecticidal nets by using an anopheles salivary biomarker of human–vector contact. *The American Journal of Tropical Medicine and Hygiene*. 2016 Dec 7;95(6):1376–82.
  13. Drame PM, Poinsignon A, Besnard P, Mire JL, Dos-Santos MA, Sow CS, et al. Human antibody response to *Anopheles gambiae* saliva: an immuno-epidemiological biomarker to evaluate the efficacy of insecticide-treated nets in malaria vector control. *The American Journal of Tropical Medicine and Hygiene*. 2010 Jul 6;83(1):115–21.
  14. Brosseau L, Drame PM, Besnard P, Toto J-C, Foumane V, Mire JL, et al. Human antibody response to anopheles saliva for comparing the efficacy of three malaria vector control methods in Balombo, Angola. *PLOS ONE*. 2012 Sep 24;7(9):e44189.
  15. Arnold BF, Scobie HM, Priest JW, Lammie PJ. Integrated serologic surveillance of population immunity and disease transmission. *Emerging Infectious Diseases* [Internet]. 2018 Jul [cited 2021 Apr 23];12(7). Available from: [https://wwwnc.cdc.gov/eid/article/24/7/17-1928\\_article](https://wwwnc.cdc.gov/eid/article/24/7/17-1928_article)

16. Frenk J. The global health system: strengthening national health systems as the next step for global progress. *PLOS Medicine*. 2010 Jan 12;7(1):e1000089.
17. Multiplex bead assay for integrated serological surveillance of communicable diseases in the region of the Americas. Report of the third regional meeting (Cuernavaca, 4-5 March 2020) - PAHO/WHO | Pan American Health Organization [Internet]. [cited 2021 Jun 9]. Available from: <https://www.paho.org/en/documents/multiplex-bead-assay-integrated-serological-surveillance-communicable-diseases-region>
18. Hatherell H-A, Simpson H, Baggaley RF, Hollingsworth TD, Pullan RL. Sustainable surveillance of neglected tropical diseases for the post-elimination era. *Clinical Infectious Diseases*. 2021 Jun 15;72(Supplement\_3):S210–6.
19. Pinsent A, Solomon AW, Bailey RL, Bid R, Cama A, Dean D, et al. The utility of serology for elimination surveillance of trachoma. *Nat Commun*. 2018 Dec 21;9(1):5444.
20. Martin DL, Saboyà-Díaz MI, Abashawl A, Alemayeh W, Gwyn S, Hooper PJ, et al. The use of serology for trachoma surveillance: Current status and priorities for future investigation. *PLoS Negl Trop Dis*. 2020 Sep;14(9):e0008316.
21. Woodhall SC, Gorwitz RJ, Migchelsen SJ, Gottlieb SL, Horner PJ, Geisler WM, et al. Advancing the public health applications of *Chlamydia trachomatis* serology. *Lancet Infect Dis*. 2018 Dec;18(12):e399–407.
22. Plotkin SA. Correlates of protection induced by vaccination. *Clin Vaccine Immunol*. 2010 Jul;17(7):1055–65.



23. Sommen C, Commenges D, Vu SL, Meyer L, Alioum A. Estimation of the distribution of infection times using longitudinal serological markers of HIV: implications for the estimation of HIV incidence. *Biometrics*. 2011 Jun;67(2):467–75.
24. Brookmeyer R, Laeyendecker O, Donnell D, Eshleman SH. Cross-sectional HIV incidence estimation in HIV prevention research. *J Acquir Immune Defic Syndr*. 2013 Jul;63(02):S233–9.
25. Woldesenbet S, Kufa-Chakezha T, Lombard C, Manda S, Cheyip M, Ayalew K, et al. Recent HIV infection among pregnant women in the 2017 antenatal sentinel cross-sectional survey, South Africa: Assay-based incidence measurement. *PLOS ONE*. 2021 Apr 14;16(4):e0249953.
26. Kassanjee R, McWalter TA, Bärnighausen T, Welte A. A new general biomarker-based incidence estimator. *Epidemiology*. 2012 Sep;23(5):721–8.
27. Vu SL, Pillonel J, Semaille C, Bernillon P, Strat YL, Meyer L, et al. Principles and uses of HIV incidence estimation from recent infection testing - a review. *Eurosurveillance*. 2008 Sep 4;13(36):18969.
28. Arnold BF, Martin DL, Juma J, Mkocha H, Ochieng JB, Cooley GM, et al. Enteropathogen antibody dynamics and force of infection among children in low-resource settings. Ferguson NM, Jit M, White M, Leung DT, Azman A, editors. *eLife*. 2019 Aug 19;8:e45594.
29. Gergen PJ, McQuillan GM, Kiely M, Ezzati-Rice TM, Sutter RW, Virella G. A population-based serologic survey of immunity to tetanus in the United States [Internet].

<http://dx.doi.org/10.1056/NEJM199503233321201>. Massachusetts Medical Society; 2009

[cited 2021 Jul 13]. Available from:

<https://www.nejm.org/doi/10.1056/NEJM199503233321201>

30. Travassos MA, Beyene B, Adam Z, Campbell JD, Mulholland N, Diarra SS, et al. Immunization coverage surveys and linked biomarker serosurveys in three regions in Ethiopia. *PLOS ONE*. 2016 Mar 2;11(3):e0149970.
31. Priest JW, Jenks MH, Moss DM, Mao B, Buth S, Wannemuehler K, et al. Integration of multiplex bead assays for parasitic diseases into a national, population-based serosurvey of women 15-39 years of age in Cambodia. *PLOS Neglected Tropical Diseases*. 2016 May 3;10(5):e0004699.
32. Ximenes RA de A, Martelli CMT, Amaku M, Sartori AMC, de Soárez PC, Novaes HMD, et al. Modelling the Force of Infection for Hepatitis A in an urban population-based survey: a comparison of transmission patterns in Brazilian macro-regions. *PLoS One*. 2014 May 20;9(5):e94622.
33. Azman AS, Paul KK, Bhuiyan TR, Koyuncu A, Salje H, Qadri F, et al. Hepatitis E in Bangladesh: insights from a national serosurvey. *The Journal of Infectious Diseases* [Internet]. 2021 Sep 22 [cited 2021 Oct 1];(jiab446). Available from: <https://doi.org/10.1093/infdis/jiab446>
34. Bohm K, Strömpl J, Krumbholz A, Zell R, Krause G, Sievers C. Establishment of a highly sensitive assay for detection of hepatitis E virus-specific immunoglobulins. *Journal of Clinical Microbiology*. 58(2):e01029-19.

35. Bahl S, Estívariz CF, Sutter RW, Sarkar BK, Verma H, Jain V, et al. Cross-sectional serologic assessment of immunity to poliovirus infection in high-risk areas of Northern India. *The Journal of Infectious Diseases*. 2014 Nov 1;210(suppl\_1):S243–51.
36. Falkenhorst G, Simonsen J, Ceper TH, van Pelt W, de Valk H, Sadkowska-Todys M, et al. Serological cross-sectional studies on salmonella incidence in eight European countries: no correlation with incidence of reported cases. *BMC Public Health*. 2012 Jul 16;12(1):523.
37. Simonsen J, Mølbak K, Falkenhorst G, Krogfelt KA, Linneberg A, Teunis PFM. Estimation of incidences of infectious diseases based on antibody measurements. *Statistics in Medicine*. 2009;28(14):1882–95.
38. Lammie P, Solomon A, Secor E, Peeling R. Diagnostics needs for NTD programs [Internet]. *The Causes and Impacts of Neglected Tropical and Zoonotic Diseases: Opportunities for Integrated Intervention Strategies*. National Academies Press (US); 2011 [cited 2021 Jul 13]. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK62529/>
39. Njenga SM, Kanyi HM, Arnold BF, Matendechero SH, Onsongo JK, Won KY, et al. Integrated cross-sectional multiplex serosurveillance of IgG antibody responses to parasitic diseases and vaccines in coastal Kenya. *Am J Trop Med Hyg*. 2020 Jan;102(1):164–76.
40. Requena-Méndez A, Chiodini P, Bisoffi Z, Buonfrate D, Gotuzzo E, Muñoz J. The laboratory diagnosis and follow up of strongyloidiasis: a systematic review. *PLOS Neglected Tropical Diseases*. 2013 Jan 17;7(1):e2002.

41. Wilking H, Thamm M, Stark K, Aebischer T, Seeber F. Prevalence, incidence estimations and risk factors of *Toxoplasma gondii* infection in Germany: a representative, cross-sectional, serological study. *Sci Rep*. 2016 Mar 3;6(1):22551.
42. Azman AS, Lauer SA, Bhuiyan TR, Luquero FJ, Leung DT, Hegde ST, et al. *Vibrio cholerae* O1 transmission in Bangladesh: insights from a nationally representative serosurvey. *The Lancet Microbe*. 2020 Dec 1;1(8):e336–43.
43. Harris JB, LaRocque RC, Chowdhury F, Khan AI, Logvinenko T, Faruque ASG, et al. Susceptibility to *Vibrio cholerae* infection in a cohort of household contacts of patients with cholera in Bangladesh. *PLOS Neglected Tropical Diseases*. 2008 Apr 9;2(4):e221.
44. Haney DJ, Lock MD, Simon JK, Harris J, Gurwith M. Antibody-based correlates of protection against cholera: analysis of a challenge study of a cholera-naïve population. *Clin Vaccine Immunol* [Internet]. 2017 Aug 1 [cited 2021 Mar 3];24(8). Available from: <https://cvi.asm.org/content/24/8/e00098-17>
45. de Melker HE, Versteegh FGA, Schellekens JFP, Teunis PFM, Kretzschmar M. The incidence of *Bordetella pertussis* infections estimated in the population from a combination of serological surveys. *Journal of Infection*. 2006 Aug 1;53(2):106–13.
46. Cherry JD, Gornbein J, Heininger U, Stehr K. A search for serologic correlates of immunity to *Bordetella pertussis* cough illnesses. *Vaccine*. 1998 Dec;16(20):1901–6.
47. Swart EM, Gageldonk PGM van, Melker HE de, Klis FR van der, Berbers G a. M, Mollema L. Long-term protection against diphtheria in the Netherlands after 50 years of vaccination: results from a seroepidemiological study. *PLOS ONE*. 2016 Feb 10;11(2):e0148605.

48. Kutty PK, Kruszon-Moran DM, Dayan GH, Alexander JP, Williams NJ, Garcia PE, et al. Seroprevalence of antibody to mumps virus in the US population, 1999–2004. *The Journal of Infectious Diseases*. 2010 Sep 1;202(5):667–74.
49. Stringhini S, Zaballa M-E, Pullen N, Perez-Saez J, Mestral C de, Loizeau A, et al. Seroprevalence of anti-SARS-CoV-2 antibodies six months into the vaccination campaign in Geneva, Switzerland [Internet]. 2021 Aug [cited 2021 Oct 1] p. 2021.08.12.21261929. Available from: <https://www.medrxiv.org/content/10.1101/2021.08.12.21261929v1>
50. Heaney CD, Pisanic N, Randad PR, Kruczynski K, Howard T, Zhu X, et al. Comparative performance of multiplex salivary and commercially available serologic assays to detect SARS-CoV-2 IgG and neutralization titers. *medRxiv* [Internet]. 2021 Feb 1 [cited 2021 May 17]; Available from: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7852272/>
51. Rosado J, Pelleau S, Cockram C, Merkling SH, Nekkab N, Demeret C, et al. Multiplex assays for the identification of serological signatures of SARS-CoV-2 infection: an antibody-based diagnostic and machine learning study. *The Lancet Microbe*. 2021 Feb 1;2(2):e60–9.
52. Salje H, Cauchemez S, Alera MT, Rodriguez-Barraquer I, Thaisomboonsuk B, Srikiatkachorn A, et al. Reconstruction of 60 years of chikungunya epidemiology in the Philippines demonstrates episodic and focal transmission. *The Journal of Infectious Diseases*. 2016 Feb 15;213(4):604–10.
53. Yoon I-K, Srikiatkachorn A, Alera MT, Fernandez S, Cummings DAT, Salje H. Pre-existing chikungunya virus neutralizing antibodies correlate with risk of symptomatic

- infection and subclinical seroconversion in a Philippine cohort. *International Journal of Infectious Diseases*. 2020 Jun 1;95:167–73.
54. Flamand C, Bailly S, Fritzell C, Berthelot L, Vanhomwegen J, Salje H, et al. Impact of zika virus emergence in French Guiana: a large general population seroprevalence survey. *The Journal of Infectious Diseases*. 2019 Nov 6;220(12):1915–25.
55. Salje H, Paul KK, Paul R, Rodriguez-Barraquer I, Rahman Z, Alam MS, et al. Nationally-representative serostudy of dengue in Bangladesh allows generalizable disease burden estimates. Cooper B, Ferguson NM, Cooper B, Brady O, editors. *eLife*. 2019 Apr 8;8:e42869.
56. Imai N, Dorigatti I, Cauchemez S, Ferguson NM. Estimating dengue transmission intensity from sero-prevalence surveys in multiple countries. *PLOS Neglected Tropical Diseases*. 2015 Apr 16;9(4):e0003719.
57. Bailly S, Rousset D, Fritzell C, Hozé N, Ben Achour S, Berthelot L, et al. Spatial distribution and burden of emerging arboviruses in French Guiana. *Viruses*. 2021 Jul;13(7):1299.
58. Hozé N, Salje H, Rousset D, Fritzell C, Vanhomwegen J, Bailly S, et al. Reconstructing Mayaro virus circulation in French Guiana shows frequent spillovers. *Nat Commun*. 2020 Jun 5;11(1):2842.
59. Greenhouse B, Smith DL, Rodríguez-Barraquer I, Mueller I, Drakeley CJ. Taking sharper pictures of malaria with CAMERAs: combined antibodies to measure exposure recency assays. *The American Journal of Tropical Medicine and Hygiene*. 2018 Oct 8;99(5):1120–7.

60. Helb DA, Tetteh KKA, Felgner PL, Skinner J, Hubbard A, Arinaitwe E, et al. Novel serologic biomarkers provide accurate estimates of recent *Plasmodium falciparum* exposure for individuals and communities. *PNAS*. 2015 Aug 11;112(32):E4438–47.
61. Flamand C, Bailly S, Fritzell C, Berthelot L, Vanhomwegen J, Salje H, et al. Impact of zika virus emergence in French Guiana: a large general population seroprevalence survey. *The Journal of Infectious Diseases*. 2019 Nov 6;220(12):1915–25.
62. Randad PR, Pisanic N, Kruczynski K, Manabe YC, Thomas D, Pekosz A, et al. COVID-19 serology at population scale: SARS-CoV-2-specific antibody responses in saliva. *medRxiv* [Internet]. 2020 May 26 [cited 2021 May 17]; Available from: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7273305/>
63. Amini F, Auma E, Hsia Y, Bilton S, Hall T, Ramkhelawon L, et al. Reliability of dried blood spot (DBS) cards in antibody measurement: A systematic review. *PLoS One*. 2021;16(3):e0248218.
64. Iyer AS, Azman AS, Bouhenia M, Deng LO, Anderson CP, Graves M, et al. Dried blood spots for measuring *Vibrio cholerae*-specific immune responses. *PLOS Neglected Tropical Diseases*. 2018 Jan 29;12(1):e0006196.
65. Routledge I, Epstein A, Takahashi S, Janson O, Hakim J, Duarte E, et al. Citywide serosurveillance of the initial SARS-CoV-2 outbreak in San Francisco using electronic health records. *Nat Commun*. 2021 Jun 11;12(1):3566.

66. Mina MJ, Metcalf CJE, McDermott AB, Douek DC, Farrar J, Grenfell BT. A Global Immunological Observatory to meet a time of pandemics. Rosen CJ, Thakrar K, White M, editors. eLife. 2020 Jun 8;9:e58989.
67. July 2017 CC// 14. Are innovation labs delivering on their promise? [Internet]. Devex. 2017 [cited 2021 May 17]. Available from: <https://www.devex.com/news/sponsored/are-innovation-labs-delivering-on-their-promise-89045>
68. Mudanyali O, Dimitrov S, Sikora U, Padmanabhan S, Navruz I, Ozcan A. Integrated rapid-diagnostic-test reader platform on a cellphone. Lab Chip. 2012 Aug 7;12(15):2678–86.
69. Bryant JE, Azman AS, Ferrari MJ, Arnold BF, Boni MF, Boum Y, et al. Serology for SARS-CoV-2: Apprehensions, opportunities, and the path forward. Sci Immunol. 2020 May 19;5(47).