

Article

Predictive Value of Baseline [18F]FDG PET/CT for Response to Systemic Therapy in Patients with Advanced Melanoma

Virginia Liberini^{1,2}, Marco Rubatto³, Riccardo Mimmo⁴, Roberto Passera¹, Francesco Ceci⁵, Paolo Fava³, Luca Tonella³, Giulia Polverari^{1,6}, Adriana Lesca¹, Marilena Bellò¹, Vincenzo Arena⁶, Simone Ribero³, Pietro Quaglino³, Désirée Deandreis¹

- 1 Department of Medical Science, Division of Nuclear Medicine, University of Turin, Italy. v.liberini@gmail.com; roberto.passera@unito.it, giulia.polverari@unito.it, adriana.lesca@unito.it, marilena.bello@unito.it, desiree.deandreis@unito.it;
- 2 Nuclear Medicine Department, S. Croce e Carle Hospital, Cuneo, Italy. v.liberini@gmail.com;
- 3 Department of Medical Sciences, Section of Dermatology, University of Turin, C.so Dogliotti, 14 - 10126 Torino Italy. paolo.fava@unito.it, marco.rubatto@unito.it, luca.tonella@unito.it, simone.ribero@unito.it, pietro.quaglino@unito.it;
- 4 UNITO - University of Turin, Torino, Italy. riccardomimmo93@gmail.com;
- 5 Division of Nuclear Medicine, IEO European Institute of Oncology IRCCS, Milan, Italy. francesco.cec83@gmail.com;
- 6 PET Center, Affidea IRMET, Torino, Italy. giulia.polverari@unito.it, vincenzo.arena@affidea.it

* Correspondence: v.liberini@gmail.com

Abstract:

Background/Aim: To evaluate the association between baseline [18F]FDG-PET/CT tumor burden parameters and disease progression rate after first-line target therapy or immunotherapy in advanced melanoma patients.

Materials and Methods: 44 melanoma patients, who underwent [18F]FDG-PET/CT before first-line target therapy (28/50) or immunotherapy (16/50), were retrospectively analyzed. Whole-body and per-district metabolic tumor volume (MTV) and total lesion glycolysis (TLG) were calculated. Therapy response was assessed according to RECIST 1.1 on CT scan at 3 (early) and 12 (late) months. PET parameters were compared with Mann-Whitney test. Optimal cut-offs for predicting progression were defined using the ROC curve. PFS and OS were studied using Kaplan-Meier analysis.

Results: Median(IQR) MTVwb and TLGwb were 13.1 mL and 72.4 respectively. Non-responders patients were 38/44, 26/28 and 12/16 at early evaluation, and in 33/44, 21/28 and 12/16 at late evaluation in the whole-cohort, target and immunotherapy subgroup respectively. At late evaluation, MTVbone and TLGbone were higher in non-responders compared to responder patients (all $p < 0.037$) in the whole-cohort and target subgroup and also MTVwb and TLGwb (all $p < 0.022$) in target subgroup. No significant differences were found for immunotherapy subgroup. No metabolic parameters were able to predict PFS. Controversy, MTVlfn, TLGlfn, MTVsoft+lfn, TLGsoft+lfn, MTVwb and TLGwb were significantly associated (all $p < 0.05$) with OS in both the whole-cohort and target therapy subgroup.

Conclusion: Higher values of whole-body and bone metabolic parameters were correlated with poorer outcome, while higher values of whole-body, lymph node and soft tissue metabolic parameters were correlated with OS.

Keywords: immune checkpoint inhibitors, PD-1, PD-L1, CTLA-4, immunotherapy, target therapy, BRAF, MET, melanoma, [18F]FDG PET/CT.

1. Introduction

Cutaneous malignant melanoma (CMM), a malignancy of melanocyte cells, has been strongly increased in the last 40 years, with about 287700 new cases each year (3.1 cases per 100000 inhabitants-year) globally in the world [1–3]. CMM is one of the most aggressive type of skin cancer and is still associated with poor outcome, causing 90% of skin cancer mortality [4].

Recently, the introduction of two major systemic therapies has revolutionized the treatment of advanced melanoma, reducing the mortality in treated metastatic melanoma patients: molecular targeted therapy and immunotherapy [5]. Target therapies are mainly based on the use of small molecule inhibitors for v-raf murine sarcoma viral oncogene homolog B1 (BRAF)- and/or mitogen-activated protein kinase (MEK)-mutated melanomas, which specifically inhibit the most common oncogenic driver mutation responsible for melanoma cell proliferation and survival [6–8]. On the other hand, immune checkpoint inhibitors (ICIs) are based on the use of monoclonal antibodies targeting immunomodulatory receptors such as cytotoxic T-lymphocyte associated protein 4 (CTLA-4) or programmed cell death protein 1 (PD-1), administered alone or in combination. Anti-CTLA-4 therapies, such as ipilimumab, and anti-PD1 therapies, such as nivolumab, are able to draw cytotoxic T cells onto tumor cells, by blocking these inhibitory checkpoints of the immune system [2,5,9,10].

In spite of the high impact of systemic therapies on melanoma patient outcomes, still a significant percentage of patients do not achieve a response or relapse after treatment; the reasons of response heterogeneity and tumor relapse are still not clear and optimal biomarkers to predict response assessment have not yet been identified [11–14]. A personalized approach to select the right therapy, to predict the response to treatment and to avoid unnecessary toxicities appears necessary. Indeed, beside the clinical extension of disease, several predictive biomarkers have been already explored, such as histopathology's biomarkers of the primary tumor, circulating biomarkers and clinical aspect of the patient, as well as immunological and molecular markers [5,15–20].

Since 2019, 18F-fluorodeoxyglucose ([18F]FDG) positron emission tomography/computed tomography (PET/CT) has gained a leading role in malignant (stage III and IV) melanoma staging and for response prediction and response monitoring of these two major systemic therapies in melanoma [8,21,22].

Semiquantitative parameters, such as the maximum standardized uptake value (SUV_{max}), has been widely explored in the oncology field. However, in recent years interest has shifted more toward parameters that allow assessment of tumor metabolic burden (MTB) from [18F]FDG PET/CT images. MTB calculation is based on two PET parameters: metabolic tumor volume (MTV), which indicates the volume of metabolically active tumor, and total lesion glycolysis (TLG), which is the product of SUV_{mean} and MTV and provides information about average total tumor glycolysis. Moreover, these new parameters provide both global and district assessment of MTB, allowing to evaluate the prognostic weight of the involvement of certain districts, such as bone or liver [23].

Indeed, we aimed to investigate whether semiquantitative parameters and metabolic tumor parameters on baseline [18F]FDG PET/CT scans are able to: 1- predict response to target- or immunotherapy at early (three months) and late evaluation (twelve months) after initiation of systemic therapy in a cohort of metastatic melanoma patients; 2- to predict progression-free survival (PFS) and overall survival (OS).

2. Materials and Methods

2.1. Patient selection

This was a retrospective, observational, non-interventional, multicenter study, conducted at the AOU Città della salute e della Scienza di Torino.

We retrospectively analyzed a cohort of 236 metastatic melanoma patients, treated with either immunotherapy (checkpoint-inhibitors, such as anti-PD-1/anti-CTLA-4) or target therapies (BRAF and/or MEK inhibitors) between January 2018 and January 2020.

Only patients with documented willingness to the use of their medical data for research were then included in this retrospective, observational study. The study was conducted in compliance with ICH-GCP rules and the Declaration of Helsinki and approved by the Institutional Ethics Committee of University of Turin as part of the project TESEO ("Traguardi di Eccellenza nelle Scienze mediche Esplorando le Omiche" - protocol code D15D18000410001).

Eligible patients matched all the following inclusion criteria: (a) histologically proven melanoma; (b) immunotherapy or target therapy administered as the first-line treatment; (c) pre-treatment [18F]FDG PET/CT (time-point 0, TP0) performed within four months prior to systemic therapies; (d) availability of baseline CT, first post-treatment CT (time-point 1, TP1) within 3 months after the start of systemic therapy and a second post-treatment CT (time-point 2, TP2) within 12 months after the start of systemic therapy.

Exclusion criteria were: (a) they were under 18 years of age; (b) lack of follow-up/baseline imaging and clinical data; (c) patients with others concomitant oncological pathology; (d) patients treated with previous cycles of systemic therapies prior to undergoing study therapy; (e) patients enrolled for systemic therapy as neoadjuvant treatment.

2.2. Clinical evaluation and Melanoma Classification

The following characteristics of the patients selected for the study were retrieved from the clinical database of the Dermatology Department of AOU Città della Salute e della Scienza: age, sex, genetic mutations, TNM stage at the time of PET scan, tumor staging according to AJCC VIIIth edition [24], previous, ongoing and during follow-up therapies.

2.3. PET/CT acquisition

All included patients underwent a [18F]FDG PET/CT scan in a dedicated tomograph: - Philips Gemini Dual-slice EXP (Philips Medical Systems, Cleveland, OH, USA) at AOU Città della Salute e della Scienza; - Discovery 610 and Discovery IQ (GE Healthcare, Chicago, IL, USA) at Affidea-IRMET.

Patients were instructed to fast for at least 6 h before the scan, and blood glucose levels were measured before the injection of [18F]FDG. Patients were excluded if their blood glucose levels at the time of the scans exceeded 150 mg/dL (median(IQR)=5.1 (4.7|6.9) mmol/L). The intravenous injected tracer activity was of 2.5–3 MBq/kg of [18F]FDG (median(IQR)=230.0 (210.0|269.0) MBq), according to EAMN procedure guideline [25].

After an uptake time of 60 minutes (median(IQR)=73.0 (57.0|112.0) min) and following CT acquisition both for attenuation correction and anatomical correlation (from the vertex of the skull to the feet), PET data were acquired, covering the identical anatomical region of the CT. The PET scans were reconstructed with ordered subset expectation maximization (OSEM) algorithms. The tomographs results were validated for a proper quantification and quality of the images recorded.

2.4. Quantitative imaging analysis

All PET/CT images were qualitative analyzed with dedicated workstation (Advantage; GE Healthcare) and were interpreted by one nuclear medicine physicians (VL), aware of clinical data.

For each primary lesion were evaluated PET semi-quantitative parameters, including maximum standardized uptake value (SUV_{max}), metabolic tumor volume (MTV) and total lesion glycolysis (TLG). All FDG-avid lesions were semi-automatically segmented by one nuclear medicine physicians using LIFEx v. 6.0 (IMIV/CEA, Orsay, France).

LIFEx software provide the possibility to perform a semi-automatic whole-body MTV and TLG, performing the following steps: - process initialization by selecting regions with a standard uptake value (SUV) above a predefined threshold ($SUV > 2.5$) and applying a threshold set at 41% of maximum standard uptake value (SUV_{max}); - automatic calculation of the whole-body metabolic tumor volume (MTV_{wb}) and total lesion glycolysis (TLG_{wb}); - visual analysis of the resulting automated volume segmentation to remove background physiologic uptake.

For each lesion, the MTV and TLG were extracted. TLG was calculated by multiplying the MTV of each lesion with its corresponding SUV_{mean} value. Each FDG-avid lesion with clear delineation of the tumor were used for MTB parameters calculation. Each volume of interest (VOI) has been classified according to its site including soft tissue (st), lymph node (ln), lung, liver and bone.

At the end of this process, the whole-body MTV and TLG (MTV_{wb} and TLG_{wb}) has been calculated, defined as the sum of all MTV and TLG of each lesion, respectively.

We also calculated the corresponding MTV (MTV_{st} , MTV_{ln} , MTV_{lung} , MTV_{liver} and MTV_{bone}) and TLG (TLG_{st} , TLG_{ln} , TLG_{lung} , TLG_{liver} and TLG_{bone}) according to each tumor site.

For all patients, fixed VOI's were drawn over the right lobe of liver and aortic arch to evaluate blood pool and parenchymal organ background, respectively, measuring as mean standard uptake value (SUV_{mean}).

2.5. Assessment of therapy response – Endpoints

Therapy response (TR) was routinely assessed on a lesion-individual level, using response evaluation criteria in solid tumors (RECIST) 1.1 criteria [26] and comparing lesion diameter at three different time-points based on CT images: baseline (TP0), first follow-up at 3 months (TP1), second follow-up at 12 months (TP2). When CT evaluation was not feasible for fast disease progression or the worsening of patient clinical condition, the response to therapy was performed by clinical and laboratory evaluation only.

Regarding immunotherapy sub-cohort, pseudo progression (PP) was defined as a diameter increase by $\geq 20\%$ at TP1, followed by a decrease to $< 20\%$ at TP2 compared to TP0. True progressive disease (TPD) was defined as an increase by $\geq 20\%$ on both TP1 and TP2 compared to TP0.

Based on CT response assessment, patients were classified as “responder” in case of clinical benefit “including complete response, partial response, and stable disease or “non-responder” in case of disease progression at first follow-up at 3 months (TP1) and second follow-up at 12 months (TP2), respectively.

Overall survival (OS) was defined as the time from treatment initiation to the date of death. Progression-free survival (PFS) was defined as the time from treatment initiation to the date of first progression/appearance of new lesions.

Study workflow is shown in Figure S1.

2.6. Statistical analysis

Descriptive statistics for quantitative variables were expressed as median and inter-quartile range (IQR), while those for categorical variables as absolute/relative frequencies. The inferential tests were based on the Mann-Whitney one for continuous variables and the Fisher's exact test for categorical ones.

For PET parameters (whole-body and districts MTV and TLG), we used a ROC analysis to determine the best cut-off allowing to predict the patient's outcome using the Youden index [20]. The area under the curves (AUC), sensitivity, specificity and accuracy were reported.

The survival analysis was carried out using the Kaplan-Meier curves and the log-rank test to compare PFS and OS between the two groups (“responders” versus “non-responders”).

Statistical significance was considered for $p < 0.05$. Statistical analyses were performed using IBM SPSS version 26.0 (IBM, Armonk, NY, USA) [27].

3. Results

3.1. Patient and primary tumor characteristics.

Out of the 236 malignant melanoma patients who had undergone FDG PET/CT (retrospectively analyzed from the dermatology department database of our University Center), a total of 44 patients naive to systemic therapies were included in the final analysis (refer to CONSORT diagram, Figure 1).

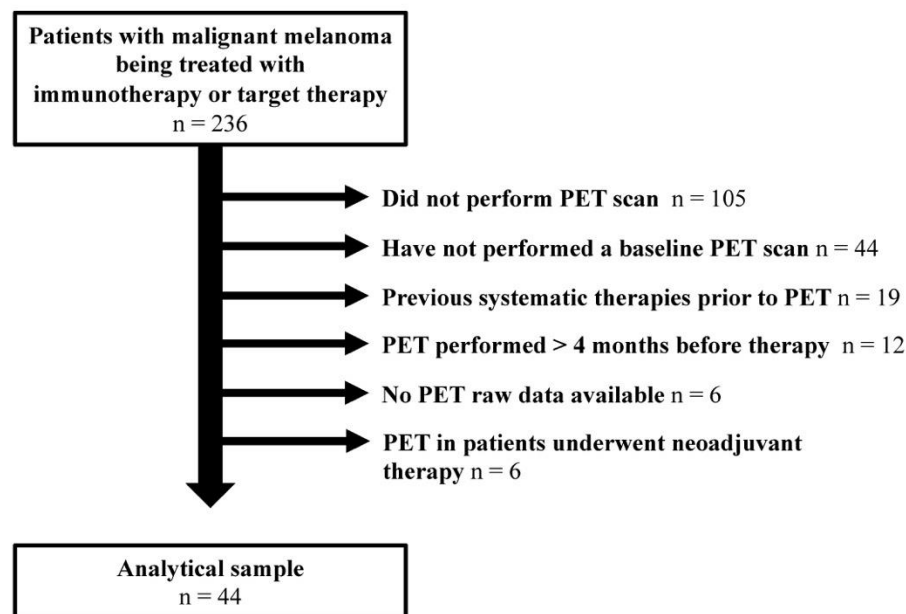


Figure 1. CONSORT DIAGRAM of patient inclusion/exclusion.

Before PET/CT, all 44 patients (28M, 16F) had already undergone surgery (primary tumor and/or lymph node surgery). Of the 44 patients, 28/44 (63.6%) underwent target therapy and 16/44 (36.4%) underwent immunotherapy. Patient and tumor characteristics are listed in Table 1.

Table 1. Patient and primary tumor characteristics.

Patient characteristics	
<i>Gender, n (%)</i>	
Male	28.0 (63.6)
Female	16.0 (36.4)
<i>Age (years), median (IQR)</i>	
62.0 (49.7-75.0)	
Primary melanoma characteristics	
<i>Type, n (%)</i>	
Superficial spreading melanoma (SSM)	12 (27.3)
Lentigo malignant melanoma (LMM)	2 (4.5)
Acral lentiginous melanoma (ALM)	0 (0.0)

<i>Nodular melanoma (NM)</i>	16 (36.4)
<i>Unknown</i>	8 (18.2)
<i>Location, n (%)</i>	
<i>Head and neck</i>	9 (20.4)
<i>Torso</i>	19 (43.3)
<i>Limbs</i>	10 (22.7)
<i>Unknown</i>	6 (13.6)
<i>PET stage, n (%)</i>	
<i>III</i>	11 (25.0)
<i>IV</i>	33 (75.0)
<i>Breslow (mm), median (IQR)</i>	4.5 (2.0-5.0)
<i>Ulceration, n (%)</i>	
<i>Yes</i>	8 (18.2)
<i>No</i>	11 (25.0)
<i>Unknown</i>	25 (56.8)
<i>BRAF mutation, n (%)</i>	
<i>Yes</i>	29 (65.9)
<i>No</i>	15 (34.1)

Note: BRAF = *v-Raf murine sarcoma viral oncogene homolog B*; MBq = megabecquerel; PET = positron emission tomography.

3.2. Semi-quantitative PET images results – Metabolic tumor burden

The median(IQR) MTVwb was 13.1(4.3|30.9) mL in the entire cohort, 12.1(3.9|28.6) mL in the subgroup of patients treated with target therapy and 14.9(5.4|46.2) mL in the subgroup of patients treated with immunotherapy. The median (IQR) TLGwb was 72.4 (11.6|17.5) in the entire cohort, 65.2(10.2|257.8) in the subgroup of patients treated with target therapy and 94.6 (22.7|263.6) in the subgroup of patients treated with immunotherapy. Details of the semi-quantitative parameters extrapolated for each district from the PET images are summarized in Table S1.

As showed in Figure 2, PET/CT images showed soft tissue, lymph node, lung, liver, and bone involvement: - in respectively 25.0%, 70.5%, 38.6%, 6.8% and 13.6% of cases in the whole cohort; - in 28.6%, 78.6%, 17.9%, 7.1% and 14.3% of cases respectively in the target therapy subgroup; - in 18.9%, 56.3%, 75%, 6.3% and 12.5% of cases in respectively in the immunotherapy subgroup.

[18F]FDG PET/CT semiquantitative parameters: MTV and TLG (median-IQR)

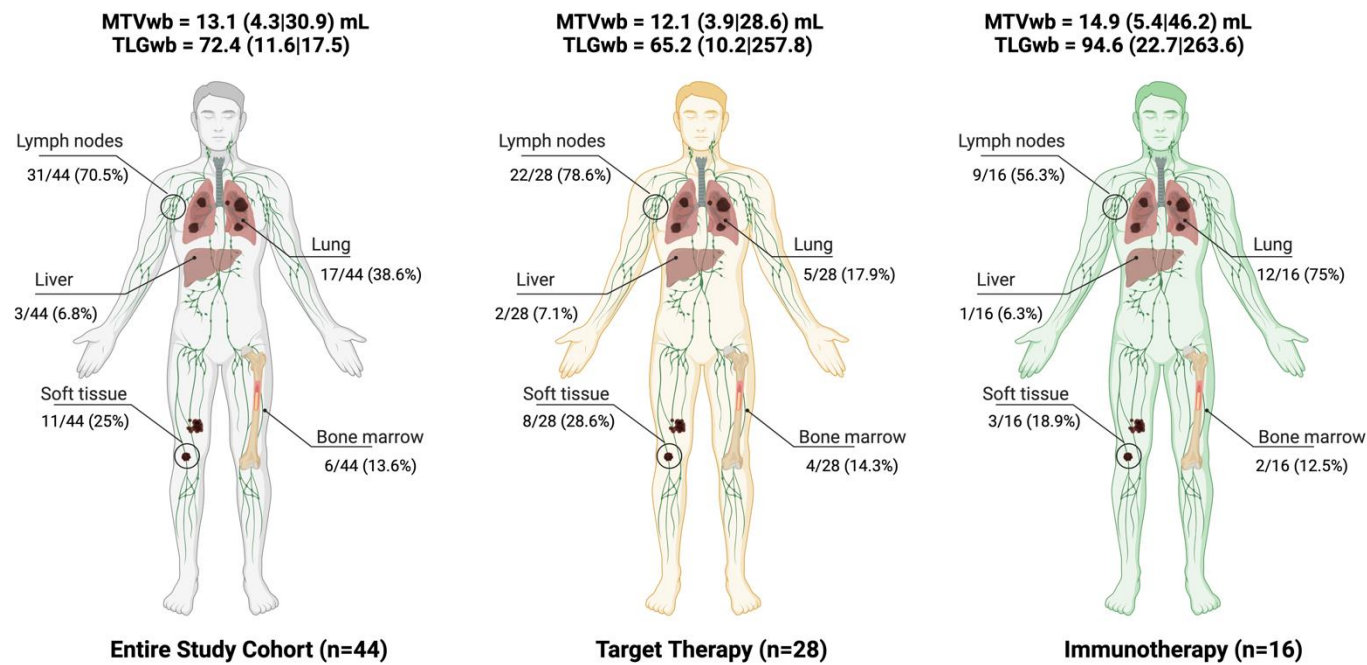


Figure 2. MTV and TLG whole body and Metabolic active lesions per district (Interest expressed in percentage) for the entire cohort, the target therapy subgroup, and the immunotherapy subgroup respectively.

Stratifying patients according to stage at PET scan time (11/44 stage III and 33/44 stage IV), in the entire cohort, the median(IQR) MTVwb and TLGwb was 4.3(2.6|30.2) mL and 12.3(5.3|408.2) respectively for the stage III, and 16.0(7.3|33.6) mL and 79.1(28.8|225.8) respectively for the stage IV. However, these differences were not statistically significant (p=0.178 for MTVwb and p=0.406 for TLGwb).

3.3. Early and late response assessment

According to RECIST 1.1 criteria, the favorable outcome (complete response + partial response + stable disease = 'patients with clinical benefit') in the entire cohort, in the target therapy subgroup and in the immunotherapy subgroup was respectively identified in 6/44 (13.6%), 2/28 (7.1%) and 4/16 (25%) of cases at 3 months and in 11/44 (25%), 7/28 (25%) and 4/16 (25%) of cases at 12 months. For the immunotherapy subgroup, no pseudoprogression events occurred at 3 months, as all progressions at 3 months were confirmed at 12 months.

On the Mann-Whitney test, no semi-quantitative PET parameter was associated to response to therapy at 3 months in either in the whole-cohort or the two subgroups.

Conversely, at the 12-month evaluation, higher values of skeletal metabolic tumor burden in the entire cohort and in the target therapy subgroup (MTVbone, TLGbone and SUVmax-bone; all p<0.037) and higher values of total metabolic tumor burden (MTVwb and TLGwb; all p<0.022) in the target therapy subgroup were associated with radiological disease progression (non-responder patients). Data are summarized in Table S2.

For the above-mentioned parameters, optimal cut-offs to predict responder vs non-responder patients at 12 months were defined using the receiver operating characteristic (ROC) curve, the results of which are shown in Table 2 and Figure S2.

Table 2. Optimal cut-offs of semi-quantitative parameters associated to responder vs non-responder patients at 12 months defined using the receiver operating characteristic (ROC) curve.

Parameters at 12 months	Optimal cut-off	Sensitivity	Specificity	AUC	p-value
Entire cohort					
Bone MTV (mL)	6.1	36%	97%	0.713	0.037
Bone TLG	18.8	45%	97%	0.716	0.034
Bone SUVmax	4.1	36%	97%	0.716	0.034
Target therapy cohort					
Bone MTV (mL)	13.1	42%	100%	0.786	0.027
Bone TLG	18.8	57%	100%	0.786	0.027
Bone SUVmax	5.7	42%	100%	0.786	0.027
Target therapy cohort					
Whole body MTV (mL)	24.6	71%	85%	0.814	0.015
Whole body TLG	208.4	71%	85%	0.793	0.023

3.4. Patients' outcome results

Median (IQR) follow up for the whole cohort and for the target therapy and the immunotherapy sub-groups was 21.0(13.2|35.7), 22.5(15.0|37.7) and 16.5(7.5|34.7), respectively. In the entire cohort, there were 11/44 (25%) deaths and 22/44 (50%) progression events; OS was 24.2 (range: 2.0–59.0; IQR: 13.2–35.7), while PFS was 21.0 (range: 2.0–53.0; IQR: 10.2–32.7). In the target therapy sub-cohort, there were 6/28 (21.4%) deaths and 13/28 (46.4%) progression events; OS was 26.8 (range: 9.0–59.0; IQR: 15.0–37.7), while PFS was 22.6 (range: 3.0–53.0; IQR: 14.0–31.7). In the immunotherapy sub-cohort, there were 5/16 (31.3%) deaths and 9/16 (56.3%) progression events; OS was 19.6 (range: 2.0–42.0; IQR: 7.5–34.7), while PFS was 18.0 (range: 2.0–42.0; IQR: 3.2–33.7).

On the Mann-Whitney test, no semi-quantitative PET parameter was associated to progression either in the whole cohort or in the two subgroups.

Conversely, the following semi-quantitative PET parameter was able to predict OS: - lymph nodes (MTVln, $p=0.011$; TLGln, $p=0.005$; SUVmax-ln, $p=0.036$), soft tissue + lymph nodes (MTVsoft+ln, $p=0.036$; TLGsoft+ln, $p=0.018$; SUVmax-soft+ln; all $p=0.047$) and whole body (MTVwb, $p=0.044$; TLGwb, $p=0.009$; SUVmax-wb, $p=0.029$) metabolic tumor burden in the entire cohort; - lymph nodes (MTVln, $p<0.020$; TLGln, $p=0.005$), soft tissue + lymph nodes (MTVsoft+ln, $p=0.010$; TLGsoft+ln, $p=0.004$) and whole body (MTVwb, $p=0.045$; TLGwb, $p=0.008$) metabolic tumor burden in the target therapy sub-group. For the above-mentioned parameters, optimal cut-offs to predict OS were defined using the receiver operating characteristic (ROC) curve, the results of which are shown in Table 3 and Figure S3.

Table 3. Optimal cut-offs of semi-quantitative parameters associated to OS defined using the receiver operating characteristic (ROC) curve.

Parameters for OS	Optimal cut-off	Sensitivity	Specificity	AUC	p-value
Entire cohort					
Lymph nodes MTV (mL)	10.6	63%	76%	0.755	0.012
Lymph nodes TLG	55.1	63%	76%	0.777	0.006
Lymph nodes SUVmax*	9.7	72%	64%	0.713	0.036
Entire cohort					
Soft tissue + LFN MTV (mL)	10.6	72%	67%	0.713	0.036
Soft tissue + LFN TLG	66.1	63%	70%	0.738	0.019
Soft tissue + LFN SUVmax*	9.2	81%	61%	0.702	0.046
Entire cohort					
Whole body MTV (mL)	14.8	72%	61%	0.705	0.043
Whole body TLG	86.4	72%	64%	0.760	0.010
Whole body SUVmax*	11.6	72%	61%	0.720	0.030

Target therapy cohort					
Lymph nodes MTV (mL)	10.9	66%	73%	0.811	0.022
Lymph nodes TLG	137.4	66%	82%	0.864	0.007
Target therapy cohort					
Soft tissue +LFN MTV (mL)	14.6	66%	82%	0.841	0.012
Soft tissue + LFN TLG	132.1	66%	78%	0.871	0.006
Target therapy cohort					
Whole body MTV (mL)	17.6	66%	69%	0.773	0.044
Whole body TLG	158.1	66%	78%	0.848	0.010

In the entire cohort and target therapy subgroup, Kaplan–Meier curves showed a marginal trend in predicting disease progression among patients with MTVlfn, TLGlfn, MTVsoft+lfn, TLG soft+lfn, MTVwb and TLGwb lower or higher the median values. (Figure 5 for the entire cohort and Figure 6 for the target therapy cohort).

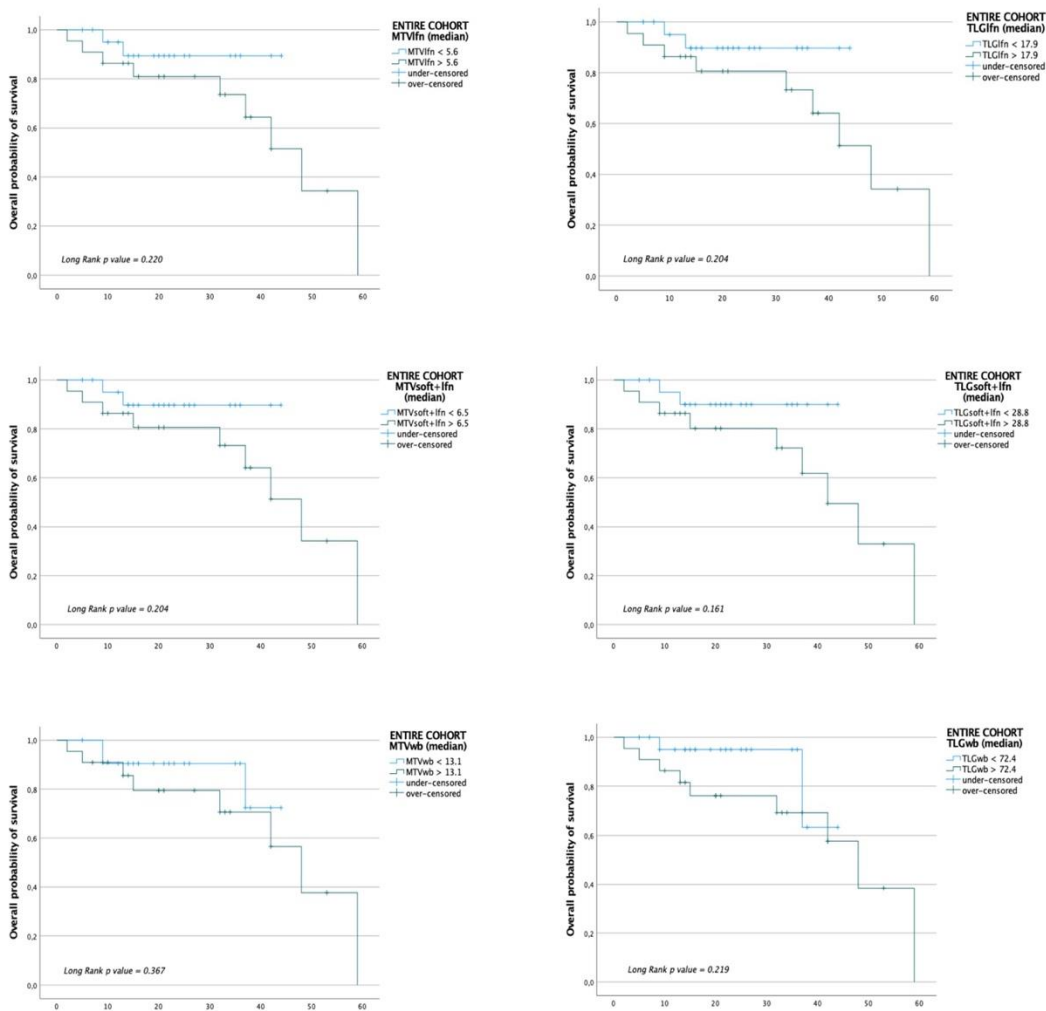


Figure 3. Kaplan–Meier plot analysis for whole-body and district MTV and TLG with overall survival (OS) in the entire cohort. Population was grouped by the median value.

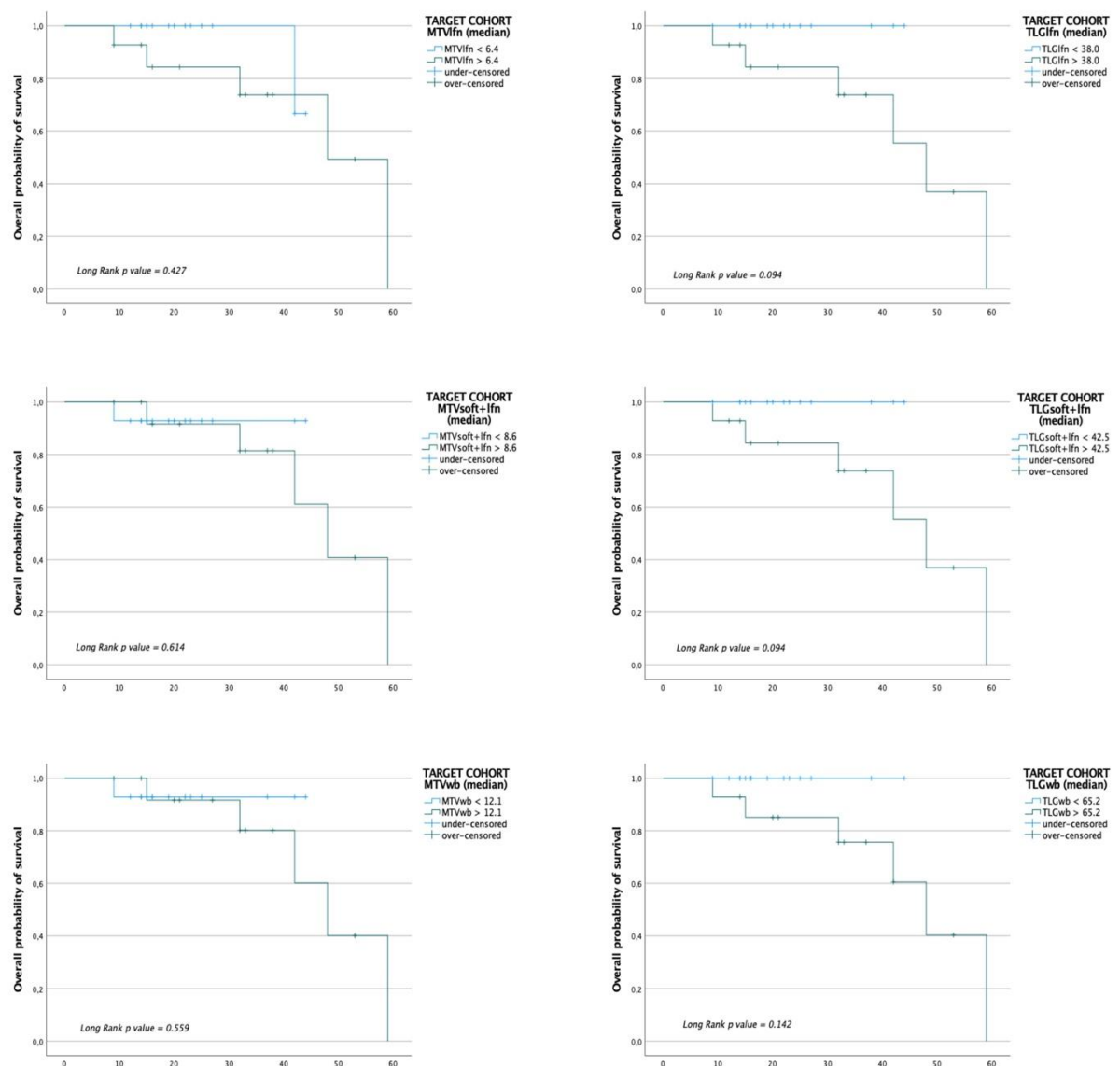


Figure 4. Kaplan-Meier plot analysis for whole-body and district MTV and TLG with overall survival (OS) in the target cohort. Population was grouped by the median value.

4. Discussion

Over the past decade [18F]FDG PET/CT has gained a central role in stage III and IV melanoma staging and become widely used in this clinical scenario, even if limited and heterogeneous data are currently available regarding the role of PET/CT semi-quantitative parameters as predictors of patient outcome [8,21,22].

Whole-body MTV and TLG are the PET-derived parameters that have been previously applied to assess the tumor metabolic activity in malignant melanoma patients [22,28–31]. To the best of our knowledge, this is the first study that attempts to identify PET parameters to assess their predictive value of disease progression and survival not only based on whole body involvement (whole-body tumor metabolic burden), but also as district-based disease involvement.

The most significant results of our study were as follows: we observed that (1) PET semi-quantitative parameters were not significantly correlated with radiological progression at three months (2) higher MTVbone and TLGbone value (all $p < 0.037$) in both the entire cohort and in the target therapy subgroup as well as higher MTVwb and TLGwb value (all $p < 0.022$) in the target therapy subgroup were correlated with radiological progression at twelve months ; (3) higher MTVlfn, TLGlfn, MTVsoft+lfn, TLGsoft+lfn, MTVwb and TLGwb value (all $p < 0.05$) in both the entire cohort and in the target therapy subgroup were correlated with OS ; (4) no correlation was found between PET semi-quantitative parameters and both radiological progression and OS in patients receiving immunotherapy.

From these results, we can deduce that [18F]FDG baseline PET/CT may have a role in predicting disease progression in patients with high tumor burden, especially in patients candidates to target therapy.

Compatibly with the small sample size, our study suggests that the bone involvement could be a predictor of worse response to new generation systemic therapies. The skeleton is the fourth site of metastasis in malignant melanoma (occurring in about 11–18% of patients); however, the impact of bone disease in melanoma has been scarcely investigated [32]. However, our results seem in line with data collected from the SEER (Surveillance, Epidemiology and End Results) database, which showed how bone metastases are frequently associated with poor prognosis [33]. Recently, in their retrospective survey on bone metastasis in melanoma, Mannavola et al. [32] confirmed the unfavorable impact of bone metastases on patient survival. The authors found a direct correlation between prognosis and skeletal tumour burden (≥ 5 bone lesions); moreover, their analysis revealed that patients receiving ICIs and/or targeted agents showed a better prognosis (9.0–16.5 months) than those undergoing chemotherapy (4.0 months). Finally, Kudura et al. [31] have recently showed how melanoma metastases located in bone structures had a negative influence on the outcome at [18F]FDG PET/CT scans performed six months after immunotherapy start in 111 metastatic melanoma patients, particularly in women.

With regarding to the negative results obtained in the immunotherapy subgroup patients, the small sample size certainly represents a limitation for the statistical analysis and future studies with inclusion of a larger sample are needed to evaluate the possible association of PET district-based semi-quantitative parameters with outcome. However, although several studies have already shown a correlation between PET parameters (MTVwb and TLGwb) and outcome [34], our results may however be in line with what has recently been shown by several works. Tumor burden is not the only factor impacting the response to immunotherapy, indeed tumor microenvironments and the patient's immune response seem to play a predominant role [10]. Recently, in a retrospective study on 112 metastatic melanoma patients treated with immune checkpoint inhibition, Basler et al. [29] assessed that a combined blood biomarkers (LDH+S100) and non-invasive [18F]FDG PET/CT-based radiomic models are promising biomarkers for early differentiation of pseudo-progression. In the previous mentioned work of Kudura et al. [31] on 111 melanoma patients treated with immunotherapy, the initial metastatic volume identified at baseline [18F]FDG PET/CT scans was not a predictive biomarker of response. Hence, the incorporation of circulating biomarkers in predictive models of immunotherapy response has become crucial. Recent studies suggest that immune analysis of tumor samples and, more specifically, degree of T cell infiltration in tumor environment have a pivotal role to understand the tumor response to immunotherapy; since ICI are supposed to trigger the T cells activity [16,17].

Regarding the analysis of overall survival, significant correlations were found in the stratification of the analyzed semi-quantitative parameters. Patients with higher metabolic tumor burden (MTVwb > 13.1 mL and TLGwb > 72.4 in the entire cohort; MTVwb > 12.1 mL and TLGwb > 65.2 in the target therapy subgroup) showed a worse prognosis.

These results agree with those found in the previous works of Seban et al. [28] and Ito et al. [35] performed respectively on 56 and 142 patients with malignant melanoma undergoing immunotherapy; although in both studies the median cut-off value identified was higher for MTV (25 mL and 26.8 mL respectively, versus 13.1 mL), higher for TLG in the study of Seban et al. (TLG cut-off = 258) and comparable with the study of Ito et al. (TLG cut-off = 78.7 versus 72.4 at our study). These differences could be justified both by the limited sample of our study and by the different setting of patients evaluated (both target and immunotherapy in our study, only immunotherapy in the other two studies mentioned).

Finally, probably the most interesting finding in our work relates to the impact of lymph node and “soft tissue + lymph node” tumor burden on overall survival in both the entire cohort (MTV = 5.6 mL and 6.5 mL respectively; TLG = 17.9 and 28.8 respectively) and in the target therapy subgroup (MTV = 6.4 mL and 8.6 mL respectively; TLG = 38.0 and 42.5 respectively).

In 2015, Beasley et al [36] reported a 5-year survival rate of 59% in patients without regional lymph node disease compared to 19% for those with lymph node disease. In addition, cutaneous and subcutaneous metastases of melanoma are known to be associated with the development of lymph node and/or systemic metastases [37,38] and with poor prognosis [39–45]. Knowing in which patients the lymph node and soft tissue tumor burden assessed at baseline [18F]FDG PET/CT scan may be associated with a worse prognosis could be extremely useful, especially for patients candidates to target therapy, in which acquired resistance could eventually develop in most patients due to several secondary events including mutations that evolve in response to treatment [22] and it is important to recognize that development of new cutaneous lesions with high FDG uptake can reflect accelerated growth.

Limitation

This work is not exempt from limitations. Due to the retrospective design of the study, the population selected was not homogeneous (e.g., different sample size between target and immunotherapy subgroup, range of time between PET scan and start of systemic therapy). However, this is a real-world scenario cohort, represented by patients who generally are referred to [18F]FDG PET/CT before systemic therapy in daily clinical practice.

This analysis was performed in a relatively small sample size. A larger cohort would be preferable. In our study, we observed an association of MTV and TLG with both responder vs. non-responder status and OS. Nevertheless, this association was not maintained in the Kaplan–Meier analysis and, despite a non-negligible trend, MTV and TLG were not statistically significantly associated with PFS. The limited number of events at the end of follow-up influenced the correlation between these metabolic parameters and responder status and OS. It is probable that our statistical model might reach significance in a larger cohort, which would also allow stratification of patients according to clinical stage and risk of progression (based on clinical, laboratory and radiological data).

5. Conclusions

According to our results, baseline [18F]FDG PET/CT performed before the start of systemic therapy might be an important tool to predict response to treatment in patients with advanced melanoma.

Our study, despite far from definitive evidence, encouraged to consider the importance of whole-body tumor metabolic burden, together with single district involvement of metabolically active disease, especially bone metabolic parameters for disease progression and lymph node and soft tissue metabolic parameters for the OS and especially in patients treated with target therapy. These data should in future be validated in prospective studies with a larger patient population.

6. Patents

Supplementary Materials: The following are available online at www.mdpi.com/xxx/: Figure S1, Table S1, Table S2, Table S3 and Table S4.

Author Contributions: Conceptualization, V.L. and D.D.; methodology, V.L.; software, V.L. and R.M.; formal analysis, V.L., R.M. and R.P.; investigation, resources, and data curation, V.L. and R.M.; writing—original draft preparation, V.L.; writing—review and editing, F.C., P.F., M.R., L.T., G.P., S.R., M.B., V.A., P.Q. and D.D.; visualization, supervision and project administration, P.Q. and D.D. All authors have read and agreed to the published version of the manuscript.

Funding: This work was partially funded with the TESEO project "Precision medicine in malignancies using omics and big data - Strategic Project of Departmental Excellence" of the Department of Medical Sciences, University of Turin.

Institutional Review Board Statement: The study was conducted according to the guidelines of the Declaration of Helsinki and approved by the Institutional Ethics Committee of University of Turin as part of the project TESEO ("Traguardi di Eccellenza nelle Scienze mediche Esplorando le Omiche" protocol code D15D18000410001 and date of approval 31 January 2018).

Informed Consent Statement: Written informed consent has been obtained from the patient(s) to publish this paper.

Acknowledgments: We would like to express our gratitude to the anonymous patients on whom this work is based and the staff of the Department of Nuclear Medicine for their excellent technical support. Figure 2 has been created with BioRender.com.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Tripp, M.K.; Watson, M.; Balk, S.J.; Swetter, S.M.; Gershenwald, J.E. State of the science on prevention and screening to reduce melanoma incidence and mortality: The time is now. *CA. Cancer J. Clin.* **2016**, *66*, 460–480, doi:10.3322/caac.21352.
2. Schadendorf, D.; Fisher, D.E.; Garbe, C.; Gershenwald, J.E.; Grob, J.J.; Halpern, A.; Herlyn, M.; Marchetti, M.A.; McArthur, G.; Ribas, A.; et al. Melanoma. *Nat. Rev. Dis. Prim.* **2015**, *1*, 15003, doi:10.1038/nrdp.2015.3.
3. Ferlay, J.; Colombet, M.; Soerjomataram, I.; Mathers, C.; Parkin, D.M.; Piñeros, M.; Znaor, A.; Bray, F. Estimating the global cancer incidence and mortality in 2018: GLOBOCAN sources and methods. *Int. J. Cancer* **2019**, *144*, 1941–1953.
4. Bray, F.; Ferlay, J.; Soerjomataram, I.; Siegel, R.L.; Torre, L.A.; Jemal, A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA. Cancer J. Clin.* **2018**, *68*, 394–424, doi:10.3322/caac.21492.
5. Bomar, L.; Senithilnathan, A.; Ahn, C. Systemic Therapies for Advanced Melanoma. *Dermatol. Clin.* **2019**, *37*, 409–423.
6. Wong, D.J.L.; Ribas, A. Targeted therapy for melanoma. In *Cancer Treatment and Research*; Cancer Treat Res, 2016; Vol. 167, pp. 251–262.
7. Long, G. V.; Stroyakovskiy, D.; Gogas, H.; Levchenko, E.; De Braud, F.; Larkin, J.; Garbe, C.; Jouary, T.; Hauschild, A.; Grob, J.J.; et al. Dabrafenib and trametinib versus dabrafenib and placebo for Val600 BRAF-mutant melanoma: A multicentre, double-blind, phase 3 randomised controlled trial. *Lancet* **2015**, *386*, 444–451, doi:10.1016/S0140-6736(15)60898-4.
8. Wong, A.N.M.; McArthur, G.A.; Hofman, M.S.; Hicks, R.J. The Advantages and Challenges of Using FDG PET/CT for Response Assessment in Melanoma in the Era of Targeted Agents and Immunotherapy. *Eur. J. Nucl. Med. Mol. Imaging* **2017**, *44*, 67–77.
9. Wolchok, J.D.; Kluger, H.; Callahan, M.K.; Postow, M.A.; Rizvi, N.A.; Lesokhin, A.M.; Segal, N.H.; Ariyan, C.E.; Gordon, R.-A.; Reed, K.; et al. Nivolumab plus Ipilimumab in Advanced Melanoma. *N. Engl. J. Med.* **2013**, *369*, 122–133, doi:10.1056/nejmoa1302369.
10. Liberini, V.; Laudicella, R.; Capozza, M.; Huellner, M.W.; Burger, I.A.; Baldari, S.; Terreno, E.; Deandreis, D. The future of cancer diagnosis, treatment and surveillance: a systemic review on immunotherapy and immuno-pet radiotracers. *Molecules* **2021**, *26*.
11. Luke, J.J.; Flaherty, K.T.; Ribas, A.; Long, G. V. Targeted agents and immunotherapies: Optimizing outcomes in melanoma. *Nat. Rev. Clin. Oncol.* **2017**, *14*, 463–482.
12. Hamid, O.; Robert, C.; Daud, A.; Hodi, F.S.; Hwu, W.-J.; Kefford, R.; Wolchok, J.D.; Hersey, P.; Joseph, R.W.; Weber, J.S.; et al. Safety and Tumor Responses with Lambrolizumab (Anti-PD-1) in Melanoma. *N. Engl. J. Med.* **2013**, *369*, 134–144, doi:10.1056/nejmoa1305133.
13. Topalian, S.L.; Hodi, F.S.; Brahmer, J.R.; Gettinger, S.N.; Smith, D.C.; McDermott, D.F.; Powderly, J.D.; Carvajal, R.D.; Sosman, J.A.; Atkins, M.B.; et al. Safety, Activity, and Immune Correlates of Anti-PD-1 Antibody in Cancer. *N. Engl. J. Med.* **2012**, *366*, 2443–2454, doi:10.1056/nejmoa1200690.
14. Tonella, L.; Pala, V.; Ponti, R.; Rubatto, M.; Gallo, G.; Mastorino, L.; Avallone, G.; Merli, M.; Agostini, A.; Fava, P.; et al. Prognostic and predictive biomarkers in stage iii melanoma: Current insights and clinical implications. *Int. J. Mol. Sci.* **2021**, *22*.
15. Ugurel, S.; Schadendorf, D.; Horny, K.; Sucker, A.; Schramm, S.; Utikal, J.; Pföhler, C.; Herbst, R.; Schilling, B.; Blank, C.; et al. Elevated baseline serum PD-1 or PD-L1 predicts poor outcome of PD-1 inhibition therapy in metastatic melanoma. *Ann. Oncol. Off. J. Eur. Soc. Med. Oncol.* **2020**, *31*, 144–152, doi:10.1016/j.annonc.2019.09.005.
16. Diazi, S.; Tartare-Deckert, S.; Deckert, M. Bad neighborhood: Fibrotic stroma as a new player in melanoma resistance to targeted therapies. *Cancers (Basel)*. **2020**, *12*.
17. Falcone, I.; Conciatori, F.; Bazzichetto, C.; Ferretti, G.; Cognetti, F.; Ciuffreda, L.; Milella, M. Tumor microenvironment:

- Implications in melanoma resistance to targeted therapy and immunotherapy. *Cancers (Basel)*. 2020, 12, 1–26.
18. Saadani, H.; Van der Hiel, B.; Aalbersberg, E.A.; Zavrakidis, I.; Haanen, J.B.A.G.; Hoekstra, O.S.; Boellaard, R.; Stokkel, M.P.M. Metabolic biomarker-based BRAFV600 mutation association and prediction in melanoma. *J. Nucl. Med.* **2019**, *60*, 1545–1552, doi:10.2967/jnumed.119.228312.
 19. Burns, D.; George, J.; Aucoin, D.; Bower, J.; Burrell, S.; Gilbert, R.; Bower, N. The Pathogenesis and Clinical Management of Cutaneous Melanoma: An Evidence-Based Review. *J. Med. Imaging Radiat. Sci.* 2019, *50*, 460–469.e1.
 20. Franken, M.G.; Leeneman, B.; Gheorghe, M.; Uyl-de Groot, C.A.; Haanen, J.B.A.G.; van Baal, P.H.M. A systematic literature review and network meta-analysis of effectiveness and safety outcomes in advanced melanoma. *Eur. J. Cancer* 2019, *123*, 58–71.
 21. Wright, C.L.; Miller, E.D.; Contreras, C.; Knopp, M. V. Precision Nuclear Medicine: The Evolving Role of PET in Melanoma. *Radiol. Clin. North Am.* 2021, *59*, 755–772.
 22. Wong, A.N.M.; McArthur, G.A.; Hofman, M.S.; Hicks, R.J. The Advantages and Challenges of Using FDG PET/CT for Response Assessment in Melanoma in the Era of Targeted Agents and Immunotherapy. *Eur. J. Nucl. Med. Mol. Imaging* 2017, *44*, 67–77.
 23. Sarikaya, I.; Sarikaya, A. Assessing PET Parameters in Oncologic 18F-FDG Studies. *J. Nucl. Med. Technol.* 2020, *48*, 278–282.
 24. Gershenwald, J.E.; Scolyer, R.A. Melanoma Staging: American Joint Committee on Cancer (AJCC) 8th Edition and Beyond. *Ann. Surg. Oncol.* 2018, *25*, 2105–2110.
 25. Boellaard, R.; Delgado-Bolton, R.; Oyen, W.J.G.; Giammarile, F.; Tatsch, K.; Eschner, W.; Verzijlbergen, F.J.; Barrington, S.F.; Pike, L.C.; Weber, W.A.; et al. FDG PET/CT: EANM procedure guidelines for tumour imaging: version 2.0. *Eur. J. Nucl. Med. Mol. Imaging* 2015, *42*, 328–354.
 26. Eisenhauer, E.A.; Therasse, P.; Bogaerts, J.; Schwartz, L.H.; Sargent, D.; Ford, R.; Dancey, J.; Arbuck, S.; Gwyther, S.; Mooney, M.; et al. New response evaluation criteria in solid tumours: Revised RECIST guideline (version 1.1). *Eur. J. Cancer* **2009**, *45*, 228–247, doi:10.1016/j.ejca.2008.10.026.
 27. IBM Corp. Released 2017. IBM SPSS Statistics for Windows, Version 25.0. Armonk, NY: IBM Corp.
 28. Seban, R.D.; Nemer, J.S.; Marabelle, A.; Yeh, R.; Deutsch, E.; Ammari, S.; Moya-Plana, A.; Mokrane, F.Z.; Gartrell, R.D.; Finkel, G.; et al. Prognostic and theranostic 18F-FDG PET biomarkers for anti-PD1 immunotherapy in metastatic melanoma: association with outcome and transcriptomics. *Eur. J. Nucl. Med. Mol. Imaging* **2019**, *46*, 2298–2310, doi:10.1007/s00259-019-04411-7.
 29. Basler, L.; Gabryś, H.S.; Hogan, S.A.; Pavic, M.; Bogowicz, M.; Vuong, D.; Tanadini-Lang, S.; Förster, R.; Kudura, K.; Huellner, M.W.; et al. Radiomics, Tumor Volume, and Blood Biomarkers for Early Prediction of Pseudoprogression in Patients with Metastatic Melanoma Treated with Immune Checkpoint Inhibition. *Clin. Cancer Res.* **2020**, *26*, 4414–4425, doi:10.1158/1078-0432.CCR-20-0020.
 30. Ito, K.; Teng, R.; Schöder, H.; Humm, J.L.; Ni, A.; Michaud, L.; Nakajima, R.; Yamashita, R.; Wolchok, J.D.; Weber, W.A. 18 F-FDG PET/CT for monitoring of ipilimumab therapy in patients with metastatic melanoma. *J. Nucl. Med.* **2019**, *60*, 335–341, doi:10.2967/jnumed.118.213652.
 31. Kudura, K.; Dimitriou, F.; Basler, L.; Förster, R.; Mihic-Probst, D.; Kutzker, T.; Dummer, R.; Mangana, J.; Burger, I.A.; Kreissl, M.C. Prediction of early response to immune checkpoint inhibition using fdg-pet/ct in melanoma patients. *Cancers (Basel)*. **2021**, *13*, doi:10.3390/cancers13153830.
 32. Mannavola, F.; Mandala, M.; Todisco, A.; Sileni, V.C.; Palla, M.; Minisini, A.M.; Pala, L.; Morgese, F.; Di Guardo, L.; Stucci, L.S.; et al. An Italian Retrospective Survey on Bone Metastasis in Melanoma: Impact of Immunotherapy and Radiotherapy on Survival. *Front. Oncol.* **2020**, *0*, 1652, doi:10.3389/FONC.2020.01652.
 33. Abdel-Rahman, O. Clinical correlates and prognostic value of different metastatic sites in patients with malignant melanoma

- of the skin: a SEER database analysis. *J. Dermatolog. Treat.* **2018**, *29*, 176–181, doi:10.1080/09546634.2017.1360987.
34. Bisschop, C.; de Heer, E.C.; Brouwers, A.H.; Hospers, G.A.P.; Jalving, M. Rational use of 18F-FDG PET/CT in patients with advanced cutaneous melanoma: A systematic review. *Crit. Rev. Oncol. Hematol.* **2020**, *153*.
 35. Ito, K.; Schöder, H.; Teng, R.; Humm, J.L.; Ni, A.; Wolchok, J.D.; Weber, W.A. Prognostic value of baseline metabolic tumor volume measured on 18 F-fluorodeoxyglucose positron emission tomography/computed tomography in melanoma patients treated with ipilimumab therapy. *Eur. J. Nucl. Med. Mol. Imaging* **2019**, *46*, 930–939, doi:10.1007/s00259-018-4211-0.
 36. Beasley, G.; Tyler, D. In-transit Melanoma Metastases: Incidence, Prognosis, and the Role of Lymphadenectomy. *Ann. Surg. Oncol.* **2014**, *22*, 358–360.
 37. Gershenwald, J.E.; Scolyer, R.A.; Hess, K.R.; Sondak, V.K.; Long, G. V.; Ross, M.I.; Lazar, A.J.; Faries, M.B.; Kirkwood, J.M.; McArthur, G.A.; et al. Melanoma staging: Evidence-based changes in the American Joint Committee on Cancer eighth edition cancer staging manual. *CA. Cancer J. Clin.* **2017**, *67*, 472–492, doi:10.3322/caac.21409.
 38. Liberini, V.; Messerli, M.; Husmann, L.; Kudura, K.; Grünig, H.; Maurer, A.; Skawran, S.; Orita, E.; Pizzuto, D.A.; Deandreis, D.; et al. Improved detection of in-transit metastases of malignant melanoma with BSREM reconstruction in digital [18F]FDG PET/CT. *Eur. Radiol.* **2021**, doi:10.1007/s00330-021-07852-7.
 39. Pararajasingam, A.; Goodwin, R. In-transit metastases: the migration of melanoma. *Br. J. Hosp. Med.* **2020**, *81*, 1–1, doi:10.12968/hmed.2020.0202.
 40. Clemente-Ruiz de Almiron, A.; Serrano-Ortega, S. Factores de riesgo de metástasis en tránsito en pacientes con melanoma cutáneo. *Actas Dermosifiliogr.* **2012**, *103*, 207–213, doi:10.1016/j.ad.2011.06.002.
 41. Marcoval, J.; Ferreres, J.R.; Penín, R.M.; Piulats, J.M.; Caminal, J.M.; Fabra, À. Análisis descriptivo de los patrones de recidiva cutánea en los pacientes con melanoma. *Actas Dermosifiliogr.* **2011**, *102*, 791–796, doi:10.1016/j.ad.2011.04.006.
 42. Rao, U.N.M.; Ibrahim, J.; Flaherty, L.E.; Richards, J.; Kirkwood, J.M. Implications of microscopic satellites of the primary and extracapsular lymph node spread in patients with high-risk melanoma: Pathologic corollary of eastern cooperative oncology group trial E1690. *J. Clin. Oncol.* **2002**, *20*, 2053–2057, doi:10.1200/JCO.2002.08.024.
 43. León, P.; Daly, J.M.; Synnestvedt, M.; Schultz, D.J.; Elder, D.E.; Clark, W.H. The Prognostic Implications of Microscopic Satellites in Patients with Clinical Stage I Melanoma. *Arch. Surg.* **1991**, *126*, 1461–1468, doi:10.1001/archsurg.1991.01410360031006.
 44. Read, R.L.; Haydu, L.; Saw, R.P.M.; Quinn, M.J.; Shannon, K.; Spillane, A.J.; Stretch, J.R.; Scolyer, R.A.; Thompson, J.F. In-transit Melanoma Metastases: Incidence, Prognosis, and the Role of Lymphadenectomy. *Ann. Surg. Oncol.* **2015**, *22*, 475–481, doi:10.1245/s10434-014-4100-0.
 45. Weide, B.; Faller, C.; Büttner, P.; Pflugfelder, A.; Leiter, U.; Eigentler, T.K.; Bauer, J.; Forschner, A.; Meier, F.; Garbe, C. Prognostic Factors of Melanoma Patients with Satellite or In-Transit Metastasis at the Time of Stage III Diagnosis. *PLoS One* **2013**, *8*, doi:10.1371/journal.pone.0063137.