Review

NSCLC Biomarkers to predict response to immunotherapy with checkpoint inhibitors (ICI): from the cells to *in-vivo* images

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Simple Summary: Lung cancer, and in particular non-small cell lung cancer (NSCLC), remains the leading cause of cancer-related death. The development of new therapeutic approaches, including immunotherapy, has led to substantial improvement in survival time and quality of life. However, clinical benefit of immunotherapy-based strategies is still limited to a minority of patients, reflecting the need to identify predictive biomarkers of response, which are any substance, structure, or process or its products that can be measured in the body, and influence or predict clinical response. In this work, we provide an overview of approved and most promising investigational biomarkers, assessed *in-vitro/ex-vivo* and *in-vivo*, to identify patients who could benefit the most from immunotherapy-based treatment.

Abstract: Lung cancer remains the leading cause of cancer-related death and it is usually diagnosed in advanced stages (stage III or IV). Recently, the availability of targeted strategies and of immunotherapy with checkpoint inhibitors (ICI) has favorably changed patient prognosis. Treatment outcome is closely related to tumor biology and interaction with the host immune microenvironment (TME). Whether for targeted therapies the response relies on the presence of specific genetic alterations in tumor cells, for ICI accurate biomarkers of response are lacking and clinical outcome likely depends on multiple factors, host and tumor-related. This paper is an overview of the ongoing research on predictive factors both from *in-vitrolex-vivo* analysis (ranging from conventional pathology to molecular biology) and *in-vivo analysis*, where molecular imaging is showing an exponential growth and use due to the technological advancement and to the new bioinformatics approaches applied to image analyses that allow specific features recovery in specific tumor subclones.

Keywords: immune checkpoint inhibitors, non-small cell lung carcinoma, PD-1, PD-L1, immune PET, immunotherapy, Radiomics, PET/CT

List of Abbreviations: CR = complete response; CT = computer tomography; CTLA-4 = cytotoxic T-lymphocyte–associated protein 4; EGFR = epidermal growth factor receptor: EMA = European Medicines Agency; FDA = Food and Drug Administration; [18F]FDG = 2-deoxy-2-[18F]fluoro-D-glucose; ICIs = immune checkpoint inhibitors; IHC = immunohistochemistry; irAE = immune-related adverse event; mAb = monoclonal antibody; MHC-I or -II = major histocompatibility

complex-I or -II; MR = magnetic resonance; OS = overall survival; PD = progressive disease; PD-1= programmed cell death protein 1; PD-L1 = programmed cell death ligand 1; PD-L2 = programmed cell death ligand 2; PERCIMT = PET Response Evaluation Criteria for Immunotherapy; PERCIST = positron emission tomography response criteria in solid tumors; PET = positron emission tomography; PFS = progression free survival; PR = partial response; RECIST = response evaluation criteria in solid tumors; SD = stable disease; SUV = standardized uptake value; TCR = T cell receptor; TKI = tyrosine kinase inhibitors; TMB = tumor mutational burden; TME = tumor microenvironment; TPS = tumor proportion score.

1. Introduction

Lung cancer is one of the most common tumors and the leading cause of cancer-related death worldwide, accounting for about 2.21 million new cases and 1.80 million deaths in 2020 [1,2]. The majority of lung cancers (80-90%) are non-small cell lung cancers (NSCLC), which are most commonly diagnosed at advanced stages of the disease (65%), with an estimated 5-year overall survival rate of 18% [3,4].

However, the recent availability of targeted strategies and of immunotherapy with immune-checkpoint inhibitors (ICI) is favorably changing prognosis. Indeed, the treatment of advanced stage NSCLC has evolved from the empiric use of chemotherapy to a armamentarium of tailored approaches, with subsets of patients treated according to the genetic alterations of their tumor and the status of programmed death ligand-1 (PD-L1), which predict the response to targeted therapies or immune checkpoint inhibitors, respectively [4].

Only 25% of NSCLC are oncogene-addicted, showing genetic mutations targetable with highly selective tyrosine kinase inhibitors [5]. To date, in the large majority of NSCLC cases no actionable driver mutations are known, and prognosis is usually poorer in these cases [6]. In recent years, immunotherapy with ICI has provided substantial survival benefit, across all lines of treatment[7].

Immune checkpoints proteins (ICs), such as cytotoxic T-lymphocyte associated protein 4 (CTLA-4) and programmed cell death protein 1 (PD-1) are co-inhibitory receptors present on activated T cells [8–14]. Overexpression of PD-1, a transmembrane protein receptor of the CD28 family, is a hallmark of T cell exhaustion, a process characterized by functional impairment of antigen-experienced CD8+ T cells [15,16]. PD-1 has two ligands, both members of the B7 family: PD-L1 and PD-L2. PD-L1 is constitutively expressed by many different cells of hematopoietic and non-hematopoietic origin and can be induced by type I and II IFN and other cytokines [17]. Upregulation of PD-L1 in response to IFN- γ has been described as a resistance mechanism to dampen ongoing immune responses and protect healthy tissues [18].

Cancer cells have learned to take advantage of the PD-L1 overexpression to evade immune surveillance. In fact, when PD-1 on activated CD8+ T cells interacts with its corresponding ligands on antigen presenting cells and/or tumor cells, the T cell receptor (TCR) signaling is attenuated, resulting in reduced T cell effector functions in a process that ultimately generate exhaustion [12]. In both chronic infections and human cancer therapeutic, blockade of PD-1/PD-L1 interaction with monoclonal antibodies has been demonstrated to reinvigorate exhausted CD8+ T cells, restoring T cell proliferation and production of effector molecules (perforin, granzymes and cytokines)[15,19]. PD-L1 expression in tumor cells/microenvironment has been correlated to response to treatment with PD-1 inhibitor, and to date represents the only available biomarker of response for single agent PD-1/PD-L1 blockade [20].

CTLA-4 is the second IC used in clinical practice for NSCLC (USA, only). It plays a central role in T cell priming and early activation in lymphoid organs. The inhibitory signal on TCR, generated upon interaction with its ligand CD80/86, for which it competes

with CD28, downregulates antigen-specific T cell expansion [21]. In addition, CTLA-4 is not only expressed on activated T effector cell but is also constitutively exposed on T regulatory cells. This suggests that CTLA-4 blockade exerts its anti-cancer activity mostly suppressing this inhibitory CD4+ T cell population [22,23]. On the other hand, direct effects of anti-CTLA-4 in increasing effector T cell functionality is still not completely clear in humans.

ICI with PD-1/PD-L1 blockade represents the current backbone of systemic treatment strategies for advanced non-oncogene addicted NSCLC [7,24]. The choice to use PD-1/PD-L1 blockade alone or in combination with other agents (chemotherapy/chemotherapy plus anti-angiogenetics/anti-CTLA4) is based on the line of treatment and PD-L1 expression on tumor cells [25], as shown in Table 1.

Table 1. Summary of the FDA and EMA-approved immune checkpoint inhibitors for Non-small-cell lung cancer (NSCLC) patients.

ICI class	DRUG	Stage	Line	FDA indication	EMA indication
CTLA-4 inhibitor	Ipilimumab	IV	1 st	 In combination with nivolumab if tumor PD-L1 ≥1% (no EGFR or ALK mutation) In combination with nivolumab and 2 cycles of platinum-doublet chemotherapy (no EGFR or ALK mutation) [https://www.accessdata.fda.gov/drug- 	In combination with nivolumab and 2 cycles of platinum-doublet chemotherapy (no EGFR or ALK mutation) [https://www.ema.europa.eu/en/documents/product-information/yervoy-epar-product-information_en.pdf]
				satfda docs/la- bel/2020/125377s110lbl.pdf]	
PD-1 inhibitors	Nivolumab	IV	1st	 In combination with ipilimumab if tumor PD-L1 ≥1% (no EGFR or ALK mutation) In combination with ipilimumab and 2 cycles of platinum-doublet chemotherapy (no EGFR or ALK mutation) 	In combination with ipilimumab and 2 cycles of platinum-doublet chemotherapy (no EGFR or ALK mutation) [https://www.ema.europa.eu/en/documents/product-information/opdivo-epar-product-information en.pdf]
				[https://www.accessdata.fda.gov/drug-satfda_docs/la-bel/2021/125554s090lbl.pdf]	* *
			2 nd - N	 As single agent if progression on or after platinum-based chemotherapy (no EGFR or ALK mutation) If EGFR or ALK mutation: as single agent if progression on corre- 	As single agent if progression on or after chemotherapy
				sponding FDA-approved therapy	[https://www.ema.europa.eu/en/docu-
				[https://www.accessdata.fda.gov/drug-satfda_docs/la-bel/2021/125554s090lbl.pdf]	ments/product-information/opdivo- epar-product-information_en.pdf]
	Pembrolizumab	III*-IV	1 st	 In combination with pemetrexed+platinum chemotherapy in non-squamous histology, (no <i>EGFR</i> or <i>ALK</i> mutation) In combination with carboplatin+nabpaclitaxel/paclitaxel in squamous histology 	 In combination with pemetrexed+platinum chemotherapy in non-squamous histology, (no <i>EGFR</i> or <i>ALK</i> mutation) In combination with carboplatin+nabpaclitaxel/paclitaxel in squamous histology

			2 nd - N	As a single agent if tumor PD-L1≥1% (no EGFR or ALK mutations) [https://www.accessdata.fda.gov/drugsatfda_docs/label/2021/125514s096lbl.pdf] As a single agent, if tumor PD-L1≥1% at progression on or after platinum-based chemotherapy If EGFR or ALK mutation: as single agent if tumor PD-L1≥1% and progression on corresponding FDA-	As a single agent if tumor PD-L1≥50% (no EGFR or ALK mutations) [https://www.ema.europa.eu/en/medicines/human/EPAR/keytruda#productinformation-section] As a single agent, if tumor PD-L1≥1% at progression after ≥1 prior chemotherapy If EGFR or ALK mutation: as single agent if tumor PD-L1≥1% and progression after ≥1 prior chemother-
				approved therapy [https://www.accessdata.fda.gov/drug-satfda_docs/la-bel/2021/125514s096lbl.pdf]	apy and target therapy [https://www.ema.europa.eu/en/medicines/human/EPAR/keytruda#productinformation-section]
	Cemiplimab	III*-IV	1 st	As single agent if tumor PD- L1≥50% (no EGFR, ROS-1 or ALK mutations) [https://www.accessdata.fda.gov/drug-	As single agent if tumor PD- L1≥50% (no EGFR, ROS-1 or ALK mutations) [https://www.ema.europa.eu/en/docu-
				satfda_docs/la- bel/2021/761097s007lbl.pdf]	ments/product-information/libtayo- epar-product-information_en.pdf]
PD-L1 inhibitors	Durvalumab	IIIA*-B	Consolidation after CHRT	Disease not progressed following concurrent platinum-based chemo- radiotherapy [https://www.accessdata.fda.gov/drug- satfda_docs/la-	 Disease not progressed following platinum-based chemo-radiother- apy if tumor PD-L1 on ≥ 1% [https://www.ema.europa.eu/en/documents/product-information/imfinzi-
	Atezolizumab	IV	1 st	 bel/2021/761069s029lbl.pdf] As single agent if tumor PD-L1≥ 50% or tumor-infiltrating immune cells PD-L1 covering≥ 10% of the tumor area (no EGFR or ALK mutations) In combination with bevacizumab+paclitaxel+carboplatin in non-squamous histologies (no EGFR or ALK mutations) In combination with carboplatin/nab-paclitaxel in non-squamous histologies (no EGFR or ALK mutations) Interpretation of the packet of the	 epar-product-information_en.pdf] As single agent if tumor PD-L1≥ 50% or tumor-infiltrating immune cells PD-L1 covering≥ 10% of the tumor area (no EGFR or ALK mutations) In combination with bevacizumab+paclitaxel+carboplatin in non-squamous histologies (no EGFR or ALK mutations) In combination with carboplatin/nab-paclitaxel in non-squamous histologies (no EGFR or ALK mutations) In tembination with carboplatin/nab-paclitaxel in non-squamous histologies (no EGFR or ALK mutations) [https://www.ema.europa.eu/en/documents/product-information/tecentriqepar-product-information_en.pdf] As single agent at progression during or following platinum-contain-
				 ing chemotherapy If EGFR or ALK mutation: as single agent at progression on FDA-approved therapy. 	 ing chemotherapy If EGFR or ALK mutation: as single agent at progression on chemotherapy and targeted therapy

	T	
	[https://www.accessdata.fda.gov/drug-satfda_docs/la-bel/2021/761034s033s034s035s036s037s0 38lbl.pdf]	[https://www.ema.europa.eu/en/docu- ments/product-information/tecentriq- epar-product-information_en.pdf]

^{*:} III stage ineligible for surgery of definitive chemo-radiotherapy

However, despite the undiscussed survival advantage demonstrated in phase III clinical trials, ICIs benefit is not universal and only a minority of patients experience a durable response. This is due, at least in part, to poor patients' selection and knowledge gap in biological underpinnings. Accumulating evidence has shown that PD-L1 alone, assayed with immunohistochemistry, is not accurate or reliable enough to predict response to ICIs [26,27], and currently the identification of alternative/companion biomarkers is a research priority. More likely, response to ICIs may depend on a set of factors, host and tumor related, involved in the complex network of immune anti-tumor control [26,27].

Here, we provide an overview on both *in-vitro* and *in-vivo* biomarkers, already in use or under development, that may enrich for clinical benefit in NSCLC patient's candidate to ICIs treatment.

2. Materials and Methods

We searched the PubMed, PMC, Scopus, Google Scholar, Embase, Web of Science, and Cochrane library databases (between January 2010 and July 2021), using the following terms (as text and as MeSH terms):

- "Immunotherapy", "ICI*", "response biomarkers", "predictive factors", "anti-PD-1", "anti-PD-L1"; "anti-CTLA-4"; "nivolumab", "pembrolizumab", "durvalumab", "cemiplimab", "atezolizumab", "ipilimumab".
- "Positron-Emission Tomography", "Positron Emission Tomography Computed Tomography", "PET/MR*", "PET/CT", "Lung Neoplasms", "Carcinoma, Non-Small-Cell Lung", "ICOS", "immune PET", "immune-PET", "ICI*", "Fluorodeoxyglucose F18", "radiomic*" and "Artificial Intelligence".

No language restriction was applied to the search, but only articles in English were reviewed. The systematic literature search returned 565 articles. According to the PRISMA flow-chart, after duplicate removal, 182 articles have been considered, fully read, analyzed, and extensively described according to their title and abstract, as previously described [28]. We also checked for further relevant articles in the references of the articles included in the retrieved literature.

3. In-vitro biomarkers for immunotherapy

The search for indicators of clinical benefit after ICI is a current hot topic, involving various fields of knowledge, from conventional pathology to molecular biology. Due to the technological advancement in genetic sequencing platform and the availability of bioinformatic tools to interpret complex set of data, putative response biomarkers have been explored in tumor cells, blood, and healthy tissues, at cellular and molecular level. Here reviewed approved and investigational predictive factors of response, grouped according to their relationship with tumor cells, immune microenvironment, and host-intrinsic variables.

3.1. Tumor-related biomarkers

3.1.1. PD-L1 TPS (Tumor Proportion Score)

^{**:} Unresectable. Abbreviations: CH-RT, chemo-radiotherapy.

Consistent with the current knowledge of PD-1/PD-L1 blockade mechanism of action, PD-L1 expression was the first response biomarker developed, and, so far, the most successful, guiding treatment choice in locally advanced and metastatic non-oncogene addicted NSCLC [7,24].

Robust clinical evidences from phase III trials support the ability of PD-L1 status in enriching for clinical benefit [20,29,30]. However, PD-L1 TPS (i.e the percentage of tumor cells with positive membrane staining over the whole amount of tumor cells in the specimen) assessment with immunohistochemistry (IHC) on tumor specimens presents several limitations. Mechanistically, PD-L1 expression on epithelial cells is a sign of an inflamed microenvironment, mainly induced by IFN-gamma, which is produced by surrounding dendritic cells and activated T lymphocytes [12,31]. Hence, by nature, its expression in tissue is highly variable in time and space, even in the context of the same tumor lesion.

Moreover, the predictive role of PD-L1 might be influenced also by tumor histology. Phase III trials testing nivolumab upon chemotherapy failure showed a correlation between PD-L1 expression and clinical benefit in lung adenocarcinoma, whereas in squamous NSCLC it was neither prognostic nor predictive of response [32,33].

Beyond this intrinsic limitation, some technical issues also affect PD-L1 reliability as a single biomarker. Indeed, each of the approved ICI has been accompanied by a corresponding IHC kit to assess PD-L1 expression. Potential differences in staining properties, and the use of various cutoff points for expression, have long been a matter of debate.

With the aim to harmonize results and clarify the relationship across these different assays, the Blueprint project compared the 4 assays most commonly used (22C3 for pembrolizumab, 28-8 for nivolumab, SP142 for atezolizumab, and SP263 for durvalumab) [34]. When compared to the other 3, which showed a similar analytical performance, the SP142 was found to be less sensitive. However, it must be specified that SP421 criteria for positivity differ from the other assays for detecting membrane PD-L1 not only in tumor cells, but also in tumor infiltrating immune cells [34]. In this respect, it is noteworthy the study by Herbst et al, which showed that PD-L1 scoring on both tumor and immune cells in tumor microenvironment may be more accurate in predicting response, compared to the sole PD-L1 assessment on tumor cells [35]. Among other relevant findings, this study, carried out in different tumor types, also demonstrated that further variables related to T cell biology, as T-helper type 1 gene expression (interferon-gamma, CD27, CXCR3, CD45RO, GZMB and CD8A), CTLA4 expression and the absence of fractalkine (CX3CL1) in baseline tumor specimens can significantly predict benefit after ICI. With regard to these latter factors, as opposed to what observed for other tumors like melanoma, their association with response is not always significant in NSCLC.

All considered, PD-L1 expression is a weak biomarker when used alone, and the search for alternative biomarkers to use for a composite evaluation is underway.

3.1.2 Tumor mutational burden

With the advent of novel bioinformatics tools, growing attention has been dedicated to biomarkers related to cancer neoantigen load [36]. Tumor mutational burden (TMB) is defined as the number of non-synonymous mutations within the coding portion of cancer genome, which possibly reflects in a higher source of neoantigens able to stimulate anticancer T cell response. When assessed with targeted Next Generation Sequencing (NGS) platforms, TMB can be considered as the best surrogate available for neoantigen load, in terms of applicability on a large scale [37,38].

Seminal studies applied whole exome sequencing to describe and highlight the predictive role of TMB in response to ICIs [38–40]. However, due to the high costs, long turnaround time and expertise needed, targeted panels have been used on both tissue and plasma specimens, demonstrating a close correlation with whole-exome sequencing [41,42]. Similarly, to what concerns PD-L1 testing, different cut-offs and panels have been used according to the ICIs tested. A further issues concern lung cancer tissue availability,

often scarce in clinical practice and insufficient for TMB evaluation on tissue. On the other hand, when TMB is tested on blood, it is the amount of circulating DNA to be critical.

As a newer candidate biomarker, compared to PD-L1 TPS, results from prospective validation are scarce and often inconsistent. Checkmate 227 trial established a clinically meaningful TMB cut-off (≥10 mutations per megabase, using the FoundationOne CDx assay), for progression free survival (PFS) benefit after double ICIs with nivolumab plus ipilimumab vs chemotherapy. Notably, PFS benefit in high TMB patients occurred irrespectively of PD-L1 expression [43]. However, when the same threshold was used to discriminate overall survival (OS) benefit upon the same regimen, TMB failed to achieve significant results over chemotherapy [44]. Likewise, the NEPTUNE trial, which prospectively assessed response to double ICIs (durvalumab plus tremelimumab) according to TMB level in blood (≥/<20 mutations per megabase), was negative [45]. Based on these data, it cannot be excluded that the performance of tissue and blood TMB still needs to be optimized, before entering clinical practice.

Despite this limitation, based on the overall response rates from a pre-planned retrospective analysis from the KEYNOTE-158 trial, in June 2020, FDA grated approval to pembrolizumab for patients affected by advanced solid tumors with high tissue TMB (≥10 mutations per megabase evaluable with the FoundationOneCDx or the MSK-IMPACT assays), upon failure to first line chemotherapy [46].

Interestingly, a refined version of TMB has emerged from a large meta-analysis on patients receiving ICI for various cancer types, including 76 NSCLC. By analyzing tumor and immune cells, it showed that clonal TMB (i.e. the number of non- synonymous mutations estimated to be present in every cancer cell) outperforms TMB as a whole in predicting sensitization to ICI [47]. Importantly, this work, which utilized standardized bioinformatics workflows and clinical outcome criteria, underlines the potential of multivariable prediction models integrating tumor- and T-cell-intrinsic mechanisms of response. As a limitation, it must be considered that these translational works are generally retrospective and designed with objective response as the endpoint for clinical outcome, instead of survival benefit.

An alternative approach to enhance the performance of blood TMB, could be the adjustment for allele frequency, as proposed by a retrospective report from the POPLAR and OAK trials. Here, low allele frequency blood TMB (mutation counts with an allele frequency < 5%) significantly predicted favorable OS, PFS, and objective response rate [48].

Another strategy to optimize tissue TMB performance integrates tumor mutations count with epigenetic alterations, showing that high TMB is more frequently associated to aberrant DNA methylation [49]. Yet, both approaches need prospective validation.

3.1.3. Tumor genotype

Whether it is well-acknowledged that actionable mutations in the driver oncogenes epidermal growth factor receptor (*EGFR*) and anaplastic lymphoma kinase (*ALK*) confer resistance to ICI [50,51], accumulating evidence suggests a possible positive or negative predictive role for other tumor mutations, involved in various cancer pathways, that may improve the predictive ability of both PD-L1 and TMB when combined.

The IMMUNOTARGET retrospective analysis evaluated response to ICI therapy in metastatic NSCLC patients with oncogenic driver alterations including Kirsten Rat Sarcoma virus (*KRAS*), *EGFR*, v-raf murine sarcoma viral oncogene homolog B1 (*BRAF*), human epidermal growth factor receptor 2 (*HER2*), mesenchymal epithelial transition factor (*MET*), mutations, and *ALK*, c-ros oncogene 1 (*ROS1*) and rearranged during transfection (*RET*) rearrangements. Evaluating the 12-month progression free survival (PFS), the best outcome to ICI therapy was found in NSCLC patients with *KRAS* and *MET* mutation reaching the highest 12-months PFS (25.6% and 23.4%, respectively), NSCLC patients with *HER2* mutation exhibited an intermediate outcome (18.0%) and those with *RET*, *EGFR*, *ALK* and *ROS1* mutation exhibited the poorest 12-months PFS outcomes (7.0%, 6.4%, 5.9% and unevaluable due to small numbers, respectively) [52].

In a recent report, Aggarwal et al. [53] evaluated baseline blood TMB and specific genetic mutations using a 500-genes NGS panel in 66 NSCLC patients receiving first-line pembrolizumab-based treatment. The combination of blood TMB \geq 16 and absence of negative predictor mutations, as STK11/KEAP1/PTEN and HER2 exon 20, was associated with PFS and overall survival (OS) benefit.

Another analysis by Lamberti et al. [54], examined in 909 non-squamous NSCLC, clinical, pathological, and genomic data associated with high (\geq 50%), low (1-49%) and negative (<1%) PD-L1 expression. Among other factors, compared to high PD-L1 tumors, negative PD-L1 tumors presented more often *STK11* (19% versus 5%; P < 0.001), *EGFR* (22% versus 11%; P < 0.001), *CTNNB1* (4% versus 0.4%; P < 0.001) and *APC* mutations (5% versus 1%; P= 0.005) respectively. In a smaller subset receiving ICIs, *STK11* or *EGFR* mutations were also significantly associated with shorter PFS and OS.

A similar study, by Schoenfeld et al. [55] on 1586 lung adenocarcinoma patients with paired PD-L1 and targeted-NGS data, not only confirmed the association of *EGFR* and *STK11* mutation with PD-L1 negativity, but also showed that *KRAS*, *TP53*, and *MET* mutations were significantly associated with high PD-L1 expression.

Furthermore, an exploratory analysis from the KEYNOTE-042 trial presented at the AACR 2020, assessed the prevalence of *STK11* and *KEAP1* mutations and their association with pembrolizumab efficacy over chemotherapy [56]. Despite confirming that *STK11* mutations occurred more often in low PD-L1, a higher frequency in high TMB patients has been reported. More importantly, response rate, PFS and survival with pembrolizumab were similar irrespective of *STK11* or *KEAP1* mutation, while chemotherapy efficacy was lower in *STK11* mutation carriers.

As for the *KRAS* mutations which account for 20-30% of lung adenocarcinoma, these represent a heterogeneous categories, with specific mutations having different responses to checkpoint inhibition [57].

An example, even though described in a different tumor type (pancreatic cancer), has been provided for the $KRAS^{G12D}$ mutation, which has been associated with production of granulocyte–macrophage colony-stimulating factor (GM-CSF), involved in recruitment of myeloid cells, thus maintaining an immunosuppressive environment. On the other hand, it is still unexplored if other KRAS single-nucleotide point mutations at exon 12 or 13 (G12V,G12A,G12C and G13D) also mediate GM-CSF production or if this is specific to the G12D variant [58].

Other translational studies have shown an increased immunogenicity of *KRAS*-mutant tumors [59,60]. However, it has been suggested that co-occurring mutations may flip the potential immune stimulating role of *KRAS*. Skoulidis et al. [61] retrospectively assessed 17 *KRAS* mutant NSCLC treated with nivolumab, showing poorer responses in case of *KRAS/STK11* co-mutation, as compared to the *KRAS/TP53* one. Moreover, in the same report, the association of *STK11* mutation with worse clinical outcome was also confirmed among the PD-L1 positive subgroup of *KRAS/STK11* mutant NSCLC patients treated with ICIs.

Favorable effect on ICIs outcome has been reported also for NSCLC harboring ARID1A mutation. In the context of double ICIs, an exploratory analysis from the MYSTIC trial evaluated the relationship between gene alterations, detected on circulating free DNA, and outcome to durvalumab plus tremelimumab treatment [62]. Whether mutations in the tumor suppressor genes KEAP1 and STK11 were found to be associated with poorer prognosis, regardless of the treatment received (chemotherapy or ICI), mutations in ARID1A were associated with longer survival only in patients receiving durvalumab plus tremelimumab. These findings suggest that, while the highly common mutations in KEAP1 and STK11 may have negative prognostic value in NSCLC, ARID1A may deserve further attention as a possible predictive factor of response. In addition, ARID1A was retrospectively investigated in 3403 patients affected by 9 tumor types, including NSCLC [63]. ARID1A mutations were significantly associated with microsatellite instability and high TMB. Of note, its favorable predictive role on PFS occurred independently from microsatellite or TMB status.

Among others, further mutant genes described as potential predictors of ICI outcome are *POLE* (presence) and *PTEN* (absence), the first involved in DNA replication and repairs, the second an acknowledged tumor suppressor, inhibiting the PI3K/mTOR/AKT pathway [64,65].

Despite promising and, possibly, easily to assess, the concept that single tumor mutations, or even a cluster, may predict clinical outcome to ICI needs further prospective and mechanistic studies.

3.2. Biomarkers related to tumor microenvironment (TME)

3.2.1. T Lymphocyte infiltration

Whether high levels of tumor-infiltrating T lymphocytes (both CD3+CD8+ and CD3+CD4+) in TME have long been recognized to have a favorable prognostic role [66,67], emerging insight into T cell biology indicates that the rescue of T cell homing at tumor site is also essential for ICIs efficacy.

Undoubtedly, T cell infiltration is a sign of immune recognition and of inflamed microenvironment.

Activated CD8⁺ T cells are the major actor involved in anti-cancer immunity, poised to recognize and selectively eliminate host cells expressing intracellular non-self or mutated-self antigens, arising from infection or cancer [12].

A recent multiomics analysis estimated CD8+T-cell abundance as the most predictive factor of response to anti–PD-1/PD-L1 therapy across 21 cancer types, including NSCLC [68].

Indeed, in the study by Herbst et al. [35], PD-L1 expression on immune cells in TME, including myeloid cells (macrophages, dendritic cells) and T cells seemed to correlate with response to ICI better than expression on tumor cells, as mentioned in the PD-L1 relative section.

Even though standard Hematoxylin and Eosin (H&E) staining may be sufficient to assess lymphocyte infiltration, IHC is the optimal tool, allowing the characterization of lymphocyte populations using specific markers [69].

The role of CD4+ T cells is more controversial, since these are a complex population, with either immune activating (T helpers 1 and 2) or suppressive features (T helper 17, T regulatory cells), thus requiring supplemental markers for association. For example, FOXP3 and CD25 are useful for identifying T regulatory cells [70], whose infiltration has been associated to poor prognosis in NSCLC [71]. Deeper insight should be gained into the potential functions of T regulatory cells in influencing response to ICI.

A practical limitation for the histological characterization T lymphocyte infiltration in NSCLC clinical practice is the scarce availability of tumor tissue. Indeed, in contrast to other tumor types more often amenable of surgical resection, in the majority of cases, NSCLC is diagnosed at advanced stages of disease, when radical approaches are not feasible and tumor tissue is limited to cytological samples from small biopsy.

3.2.2. Tumor infiltrating CD8+ T Cell: Phenotype and TCR clonality

The incorporation of novel sequencing and sorting strategies in laboratory and translational research has allowed to augment the resolution power on T cell, in terms of new phenotypic markers discovery and evolution tracking. Consequently, this progress in technology deepened our comprehension on how T cells respond to ICI and set the basis for the development of promising response biomarker.

Comparing transcriptional, metabolic, and functional signatures of intra-tumoral CD8+ T lymphocytes, based on different expression levels of PD-1, Thommen et al. [72] found that CD8+ T cells expressing the highest levels of PD-1 displayed the strongest reactivity against tumor cells. Moreover, compatibly with their exhausted state, PD-1high CD8+ T cells showed an impaired ability in cytotoxic cytokine production, and produced

CXCL13, which stimulated immune recruitment at tertiary lymphoid structure. Importantly, the presence of this phenotype was strongly predictive for both response and survival in a small cohort of NSCLC patients treated with PD-1 blockade.

Litchfield et al. [47] recently confirmed the relevant favorable role of CXCL13 expressed by CD8+ T cells on ICI response. This group performed deep transcriptome sequencing with single-cell RNA of tumor infiltrating CD8+ T cells reactive to a clonal cancer neoantigen (MTFR2) in a NSCLC patient, finding that CXCL13 was upregulated. In the same study, CXCL13 exhibited the most marked selective expressions in ICI responders across 1008 patients receiving ICI for different tumor types.

Single-cell RNA sequencing is particularly used for the analysis of T cell receptor (TCR) sequencing, thus enabling an indirect and quantitative characterization of antigen-specific T cells.

Clonal architecture of the intra-tumoral or peripheral T cell repertoire can be considered an indicator for tumor immunogenicity and its role in predicting ICI clinical benefit is under investigation. Diversity in TCR composition is usually quantified as for richness and clonality. To simplify, richness stands for the number of specific TCR sequences, whereas clonality takes into account the distribution of the frequencies of sequences. Clonality is defined as the complement of evenness (i.e., 1 – evenness), where high TCR clonality indicates an unequal sequence distribution, with the predominance of one or more oligoclonal populations [37,73]. Most of the available evidence derive from melanoma [18,73], reflecting, at least in part, the tissue availability constrains in NSCLC clinical practice mentioned above. Nevertheless, a thorough translational analysis on resected NSCLC specimens from a phase II trial testing nivolumab in the neoadjuvant setting also explored the dynamics of intra-tumoral and peripheral neoantigen-specific T-cell clonotypes. Tumors presenting major pathological response after resection showed a higher TCR clonality compared to tumors without a pathological response. Notably, the T cell clones at higher frequency after surgery were shared between blood and tissue and many of these clones were not detected in the peripheral blood before treatment, suggesting that ICIs may induce expansion of specific T cell clones [74].

However, studies in other tumor types and treatment settings showed conflicting results both for the positive or negative predictive role of high TCR clonality on ICIs outcome [75–77], indicating that these evidences are very preliminary. Additionally, it must be considered that TCR sequencing is an expensive technique, requires a relative long time and high expertise. So far, more work is needed to clarify the utility of T cell diversity as a response biomarker.

3.2.3. Vascularization in tumor microenvironment

T cell infiltration in tumor is mediated by the co-operation of a complex network involving numerous cell types and cytokines. In contrast with what occurs for infection site, T cells entry in TME is physically restrained by vascular architecture, which is irregular in tumor, because of the high interstitial fluid pressure and altered expression of adhesion molecules [78]. These characteristics, besides limiting efficacy of ICIs, have also discouraged the clinical development in solid tumors of T cell adoptive therapies, including chimeric antigen receptor (CAR) T-cell therapies [79].

Response biomarkers to ICIs related to neo-angiogenesis, in tumor or blood, are still in early phases of translational investigation. Yet, there is increasing interest in unravelling the complex TME network regulating tumor vascularization, in search for new therapeutic targets for immunotherapy-based combinations with the aim to overcome primary or acquired resistance to ICIs.

An increasing body of preclinical and, indirect, clinical evidence has suggested that the proangiogenic soluble factors vascular endothelial growth factor (VEGF) and angiotensin 2 (ANG2) act as underlying shared regulators of angiogenesis and immune suppression [80,81].

Findings on the role of VEGF in neo-angiogenesis made it a key therapeutic target for anti-cancer strategies and led to design monoclonal antibodies or multi-kinase inhibitors targeting VEGF-alpha or its cognate endothelial receptor tyrosine kinases, VEGFR1,2 and 3. The underlying rationale for anti-cancer therapy envisions tumor starvation for shortage of blood supply, as a consequence of the inhibition of vessel formation [80].

In NSCLC treatment, bevacizumab, the most widely prescribed anti-angiogenic drug, in combination with carboplatin/paclitaxel chemotherapy has long been one of the front-line standards for non-squamous histology. Upon failure of first-line chemotherapy, 2 anti-angiogenic agents, nintedanib and ramucirumab are licensed for use in combination with docetaxel, despite marginal survival/PFS benefit [7,24].

The discovery of the interconnected immune modulation raised renewed interest in anti-angiogenetic agents for combination with ICI. Recently, bevacizumab has been FDA and EMA approved in association with first-line chemo-immunotherapy for non-squamous NSCLC, based on the results of the Impower150 trial [82].

VEGF is generated under oxygen shortage, as in case of uncontrolled tumor proliferation, through HIF-1 alpha signaling. Cancer cells, but also various immune-suppressive cells, like tumor-associated macrophages, myeloid-derived suppressor cells (MDSCs), monocytes and immature dendritic cells, have been reported to produce VEGF [83].

Numerous preclinical works have shown that VEGF exerts its immune suppressive functions on different compartments. As for CD8⁺ T cells, increased VEGF can inhibit trafficking, proliferation, and effector function [84]. While inhibiting the maturation and antigen presentation of dendritic cells [85], VEGF also induces recruitment and proliferation of immunosuppressive cells, including T regulatory cells, MDSCs, and protumor associated macrophages [81]. More importantly, the aberrant tumor vasculature fostered by VEGF leads to hypoxia and to low pH, significantly hampering T cell infiltration and survival in TME [86].

Indirect evidence on the immune functions of VEGF also derives from clinical trials testing anti-angiogenetic drugs. Indeed, in renal cell carcinoma, the combo atezolizumab-bevacizumab enhanced T cell migration, and further evidence reported increased T cell infiltration into solid tumors, possibly due to a normalized microvasculature and restoration of homing molecules [87,88].

Notably, a study in patients with metastatic melanoma treated with immune-check-point inhibitors revealed that high levels of circulating ANG2 correlate with resistance to ICIs and worse prognosis, possibly reflecting hypoxia and immunosuppression in the TME [89].

Important open questions remain to be addressed in future studies, as for the antiangiogenetics optimal dose for vasculature normalization over disruption, with subsequent improvement of oxygen levels, drug delivery, and CD8+T cell infiltration. Furthermore, the most appropriate neo-angiogenesis biomarkers, and especially the most accurate techniques/assays to study these phenomena still remain to be determined.

3.3. Host-related biomarkers

3.3.1. Circulating lymphocyte population

The limited availability of biopsy tissue in NSCLC promoted the investigation and development of blood-based biomarkers for ICIs response. Besides the assessment of TMB, other promising circulating biomarkers are related to the host immune response, especially in terms of lymphocytes populations and proliferation.

Kamphorst et al. [90] showed that increased proliferation of PD-1⁺ CD8⁺ T cells in peripheral blood after the first or second immunotherapy cycle was associated with clinical benefit, in terms of improved objective responses. Notably, an increased proliferation was observed in about 70% of the 29 patients evaluated, where PD-1⁺ CD8⁺ T cells displayed an effector-like phenotype (HLA-DR⁺, CD38⁺, Bcl-2^{low}) and expressed costimulatory molecules (CD28, CD27, ICOS).

Another study, by Kim et al. [91], in epithelial thymic carcinoma and NSCLC patients, reported that a proliferative response of peripheral blood PD-1+CD8+T cells, measured as the fold-change in the percentage of Ki-67+ cells, as early as one week after ICI start, can predict clinical benefit in terms of PFS and OS.

Another perspective has been offered by Ferrara et al. [92], who showed that an immune senescent population of circulating CD8+T cells, characterized by low proliferation, CD28-, CD57+, KLRG1+, independently predicted poor outcome to ICIs, in terms of lower objective responses, and shorter PFS and OS. Lymphocyte phenotyping was performed at baseline and immune senescent CD8+T cells were found in up to 28% of the ICI cohort. Immune senescent phenotype did not play a prognostic or predictive role for response in the platinum-based chemotherapy control, where it was present in the 11% of the cohort.

In this context, the underlying limitation of this type of studies is the hurdle in tracking tumor antigen-specific T cells, not only because of technical challenges, but especially because of the highly heterogeneous and dynamic nature of cancer cells and of the related neo-antigens.

Clearly, (circulating) T cell phenotype as a biomarker is interesting because of the close connection with T cell and ICIs biology, nevertheless it is at early phases of development and perspective confirmation is needed.

3.3.2. Innate immune populations

The negative prognostic and predictive role of circulating neutrophils, and in particular of neutrophil/lymphocyte ratio, after ICIs, has been addressed in numerous retrospective studies, reporting a significant association. Whether neutrophil/lymphocyte ratio or derived neutrophil/lymphocyte ratio are easy to assess from routine blood cell count and reproducible, a clinically meaningful threshold is awaited for prospective validation [93,94].

Using multi-parametric flow cytometry, a recent study in NSCLC patients investigated, along with neutrophil/lymphocyte ratio, the possible association with ICIs response of HLA-DR monocytes and dendritic cells. Baseline elevated neutrophil/lymphocyte ratio and high frequency of HLA-DR monocytes and/or low frequency of dendritic cells of total leukocytes identified poor responders to ICIs. Interestingly, after ICIs, reduced neutrophil/lymphocyte ratio and HLA-DR monocytes frequency, as well increased dendritic cells, predicted responses and longer OS [95].

New high-throughput labelling technologies, like mass cytometry, have widened our horizon on the complex innate immune cell network that orchestrates the innate immune and cancer interaction, paving the way for future biomarkers discovery.

Even if dedicated studies have not been carried out specifically in NSCLC, valuable evidence from metastatic melanoma suggests a positive correlation between the frequency of CD14+CD16-HLA-DRhi monocytes in peripheral blood mononuclear cells, and PFS and OS upon ICIs [96].

MDSCs represent a heterogeneous population of immature myeloid cells that fail to differentiate into neutrophils (polymorphonuclear MDSC), macrophages, or dendritic cells (monocytic MDSC) and increase during cancer and chronic infection, inhibiting T cells and natural killers (NK), among others [97,98]. The negative predictive role for ICIs outcome has been proven mostly in melanoma [99,100]. However, in a study by Youn et al. [101], a decreased frequency of circulating Lox-1 polymorphonuclear MDSC following the first cycle of nivolumab predicted a better outcome in NSCLC. Frequency of NK showed an inverse pattern, where a higher ratio of NK/ Lox-1 polymorphonuclear MDSC strongly correlated with response, PFS, and OS.

Seminal studies recently highlighted how the composition of the intestinal commensal bacterial flora (microbiota) plays a pivotal role in tuning systemic immune functions, including cancer immunity, and in particular response to ICIs [102,103].

In the setting of metastatic melanoma, the analysis of fecal samples at baseline has revealed that the presence of specific gut bacteria (namely, *Faecalibacterium genus* for ipilimumab; *Bifidobacterium longum*, *Collinsella aerofaciens*, *and Enterococcus faecium* for anti-PD-1 agents) was significantly associated to ICIs benefit [104,105].

In other epithelial tumors, including NSCLC, the relative abundance of *Akkermansia muciniphila* identified ICI responders [106]. As well, in this study, a broader and more diverse microbiome composition was also associated with longer ICIs benefit. To corroborate these findings, fecal transplantation in "germ-free" mice from ICIs responders or enriched with *Akkermansia muciniphila*, conferred increased sensitivity to ICIs activity. Of note, this study also showed an association of antibiotics use and worse ICIs outcome at the end of therapy [106].

It is unclear whether underlying factors might have interfered with the findings observed, or how to apply these results in clinical practice. The connection between microbial ecosystem and anticancer immunosurveillance is unquestioned, nonetheless, inherent mechanistic may warrant further investigation.

3.3.4. Germline genetics

Since the ability to recognize and combat non-self-pathogens largely depends on the Human Leucocyte Antigen (HLA) system, increasing interest has been raised by its possible role in influencing ICIs response, especially for the class I compartment.

Each HLA-I molecule binds specific intracellular peptides for presentation on the cell surface to CD8+ T cells. The corresponding genes are highly polymorphic, where each variant binds a select repertoire of ligands. Homozygosity in at least one HLA class I locus would be expected to present a smaller, less diverse repertoire of tumor-derived neoantigens to CD8+ T cell, when compared to heterozygosity at each class I locus [107].

In a landmark study from 2018, Chowell et al. [108] determined the HLA class I genotype of 1535 advanced cancer patients treated with ICIs, including NSCLC, showing that HLA class I homozygosity in at least one locus was significantly associated with reduced OS, independently from mutation load, tumor stage, age, and ICIs class. Notably, the predictive ability of HLA zygosity was enhanced when coupled with TMB count. Moreover, this study also described specific HLA "supertype" associated with ICIs outcome in melanoma, where patients with *HLAB44* alleles was associated with longer OS and patients with *HLAB62* with a reduced one.

The large meta-analysis by Litchfield et al. [47] did not confirm a significant association with ICI response for the level of germline HLA-I evolutionary divergence, nor for maximal HLA heterozygosity, nor for HLA B62 supertype. HLA B44 supertype was found to be marginally non-significant (OR = 1.17 [1.00-1.37], p = 0.053). However, while not significant overall, HLA B44 supertype and germline HLA-I evolutionary divergence were significant in the melanoma anti-CTLA-4 cohorts. The same authors suggest that this latter association is potentially consistent with the increase in T cell receptor (TCR) diversity observed in anti-CTLA-4-treated patients.

Whether further investigation is warranted to clarify the relationship between HLA class I status and ICI outcome, recent evidence from melanoma - where single nucleotide polymorphisms in the $Fc\gamma R$ impacted response to ipilimumab [109] - suggests that polymorphisms in other genes involved in immune response (*i.e.*, HLA class II or genes involved in the antigen presentation and processing machinery) may also be implied in response to ICIs. Dedicated studied should be carried out to explore these possible host-related predictive biomarkers.

4. In-vivo biomarkers for immunotherapy: molecular imaging

Nowadays, noninvasive imaging techniques are getting an increasing role in screening, prognosis, baseline assessment, staging, restaging, response to treatment and follow-up of several diseases, especially oncological ones. In this context, positron emission to-mography (PET) is gaining a central role for several reasons, emerging as a predominant imaging modality. Technological advancement has allowed the development of new hybrid tomographs, in which PET is associated with the state of the art of computed tomography (CT- PET/CT) or magnetic resonance (MR- PET/MR) technologies, able to provide increasingly detailed information both on the functional (PET imaging) and on the morphological side (CT and MR imaging) [110,111].

In NSCLC, both anatomic evaluation with contrast-enhanced (ce)-CT and metabolic evaluation with 2-deoxy-2-[18F]fluoro-D-glucose ([18F]FDG) PET/CT are essential for patient's staging and management [112–114]. [18F]FDG has already reached a pivotal role in NSCLC, being used routinely to evaluate tumor metabolism in NSCLC patients at staging [115–117]. Although guidelines do not support the use of serial [18F]FDG PET/CT in NSCLC patients treated with ICIs in clinical practice, several studies are highlighting the potential role of metabolic information in both predictive (at baseline PET) and prognostic (at interim evaluation or time to radiological progression) perspectives.

Moreover, the development of new molecular imaging probes to answer specific biological questions has further enhanced the potential role of functional imaging to predict and monitor response to immunotherapy [118].

Here below we present an overview of the role of [18F]FDG and other functional imaging probes (*in-vivo* tools), currently under investigation in clinical trials or already approved to enhance clinician decision-making capabilities for immunotherapeutic regimens in the NSCLC setting, highlighting limitations and advantages.

4.1. 2-deoxy-2-[18F]fluoro-D-glucose ([18F]FDG) and immunotherapy in the NSCLC setting

4.1.1. [18F]FDG and immunotherapy response assessment

The NSCLC setting represent one of the most widely acknowledged indications for [18F]FDG PET/CT, which is considered particularly helpful for the detection of lymph node metastases and distant metastases, preventing unnecessary surgeries and improving cost-effectiveness in the management of these patients [119].

[18F]FDG PET/CT is recommended for staging NSCLC due to its excellent ability (sensitivity 93%, specificity 96%) to detect adrenal and bone metastases, if baseline CT is negative (level of evidence A). [18F]FDG PET/CT is also proposed in oligometastatic NSCLC patients potentially eligible for local treatment [112–114]. In the latest revision of the National Comprehensive Cancer Network (NCCN), [18F]FDG PET/CT is recommended for staging all patients with NSCLC, if not previously done, and in case of recurrence after therapy, but it is not routinely indicated for surveillance after therapy [120].

Despite that, [18F]FDG PET/CT has reached a pivotal role in the evaluation of response to immunotherapy in clinical routine, although the method has inherent limitations in this precise setting. Growth, survival, and proliferation of cancer cells depend on their peculiar metabolism: cancer cells, even in presence of ample oxygen, prefer to metabolize glucose and ferment glucose to lactate through "aerobic glycolysis" due to mitochondrial dysfunction. This phenomenon, known as the Warburg Effect [121], is the mechanism underlying the [18F]FDG uptake in tumor cells, such as non-small-cell lung cancer. However, immunotherapy results in an increased presence of tumor-infiltrating lymphocytes (TILs) and of the inflammatory response, which lead to a transient increase in tumor volume and increased [18F]FDG uptake in responding tumoral tissue. This phenomenon determines an underestimation of the response to therapy and can be confused with a progression, and it is called "pseudo-progression" [122–124].

Both morphological Response Evaluation Criteria in Solid Tumors (RECIST) 1.1 and functional PET Response Criteria in Solid Tumors version (PERCIST) 1.0 cannot address

pseudo-progression adequately [125-127]. Regarding functional imaging, in the evaluation criteria the visual interpretation (presence of disease, affected districts, target lesions and identification of new lesions) is supported and integrated by semi-quantitative interpretation, performed using standardized uptake value (SUV). SUV is calculated by normalizing the attenuation corrected FDG uptake in a lesion to the injected dose and body weight [128]. PERCIST 1.0 define an increase of at least 30% in the sum of SULpeak of all target lesions detected at baseline and/or the appearance of new lesions as a metabolic progression disease (MPD). However, as aforementioned, in pseudo-progression phenomenon, inflammatory changes could contribute to lesion enlargement. Therefore, in the last decade, as well as the immune-related response criteria (irRC) and the immunotherapy-modified Response Evaluation Criteria in Solid Tumors (imRECIST) have been developed for morphological response assessment to immunotherapy. Furthermore also new functional criteria have been developed to overcome this limit, namely the immunotherapy-modified PET Response Criteria in Solid Tumors (imPERCIST) and PET Response Evaluation Criteria for Immunotherapy (PERCIMT) [117,129-132]. In particular, imPERCIST incorporated two new patterns of response: the unconfirmed metabolic progression disease (UMPD), which is a possible MPD that needs to be confirmed after follow-up with interval studies, and the confirmed metabolic progression disease (CMPD), which is a real progression confirmed at the follow-up study after 4-8 weeks. At the follow-up evaluation, three different patterns can be found: - the UMPD is confirmed if the uptake status is not changed; - a decrease > 30% of the uptake defines a partial metabolic response (PMR), confirming that the UMPD was a pseudo-progression; - an increase or decrease in tumor burden different from the first two cases is defined as stable disease and durable stable disease may represent antitumoral activity. Finally, PERCIMT criteria tried to overcome the limits related to the appearance of new lesions, considering as a CMPD the appearance of ≥ 4 new lesions (< 1 cm in functional diameter) or ≥ 3 new lesions (> 1 cm in functional diameter) or \geq 2 new lesions (> 1.5 cm in functional diameter), measured in centimeters on the fused PET/CT images. The major radiological and functional response criteria to immunotherapy are summarized in **Table 2**.

Table 2. Summary of the radiological and functional immunotherapy-modified response criteria (irRC, imRECIST, imPERCIST, PERCIMT) to the immunotherapy in tumor compared with the non-immunotherapy-specific ones (RECIST 1.1 and PERCIST).

Radiological response cri- teria	RECIST 1.1 [133]	irRC [132]	imRECIST [129]
Complete response (CR)	Disappearance of all target and non-target lesions, without any new lesions. Any pathological lymph nodes must have reduction in short axis to <10 mm. Determined by two observations not less than 4 weeks apart.	Disappearance of all target lesions. Determined by two observations not less than 4 weeks apart.	Disappearance of all target and non-target lesions, without any new lesions. Any pathological lymph nodes must have reduction in short axis to <10 mm. Determined by two observations not less than 4 weeks apart.
Partial re- sponse (PR)	At least a 30% decrease of the sum of maximum diameters of target lesions; no new lesions; no progression of disease.	Sum of product of all lesions decreased by >50% for at least 4 weeks; no new lesions; no progression of any lesions.	At least a 30% decrease of the sum of maximum diameters of target lesions; no new lesions; no progression of disease.
Stable dis- ease (SD)	Does not meet the criteria for CR, PR or PD, taking as reference the smallest sum of maximum diameters of target lesions.	Sum of product of all lesions decreased by <50% or increased by <25% in the size of one or more lesions.	Does not meet the criteria for CR, PR or PD, taking as reference the smallest sum of maximum diameters of target lesions.
Progressive disease (PD)	Sum of the maximum diameter of lesions increased by >20% over the smallest achieved sum of maximum diameter. The appearance of one or more new lesions is always considered progression.	A single lesion increased by >25% (over the smallest measurement achieved for the single lesion) or the appearance of new lesions, that has to be confirmed in 2 consecutive observations at least 4 weeks apart.	Sum of the maximum diameter of lesions increased by >20% over the smallest achieved sum of maximum diameter. The appearance of new lesions and/or progression of non-target lesions are considered iUPD and must be confirmed 4-8 weeks later as iCPD. Progression is not confirmed in case of shrinkage of these lesions at 4-8 weeks and evaluation must be reset.

Functional response cri- teria	PERCIST [134]	imPERCIST [130]	PERCIMT [131]
Complete metabolic response (CMR)	Complete resolution of [18F]FDG uptake within all lesions, to a level of less than or equal to that of the mean liver activity and indistinguishable from the background (blood pool uptake).	Complete resolution of [18F]FDG uptake within all lesions, to a level of less than or equal to that of the mean liver activity and indistinguishable from the background (blood pool uptake).	Complete resolution of [18F]FDG uptake within all lesions, to a level of less than or equal to that of the mean liver activity and indistinguishable from the background (blood pool uptake).
Partial metabolic response (PMR)	Reduction of at least 30% in the sum of SULpeak of all target lesions detected at baseline and an absolute drop of 0.8 SULpeak units.	Reduction of at least 30% in the sum of SULpeak of all target lesions detected at baseline and an absolute drop of 0.8 SULpeak units.	Reduction of at least 30% in the sum of SULpeak of all target lesions detected at baseline and an absolute drop of 0.8 SULpeak units.
Stable metabolic disease (SMD)	Does not meet the criteria for CR, PR or PD.	Does not meet the criteria for CR, PR or PD.	Does not meet the criteria for CR, PR or PD.
Progressive metabolic disease (PMD)	Increased of at least 30% in the sum of SULpeak of all target lesions detected at baseline and an absolute increased of 0.8 SULpeak units. Or 75% increase in total lesions glycolysis (TLG), with no decrease in SUL. Or The appearance of one or more new FDG-avid lesions that are typical of cancer and not related to inflammation or infection is always considered progression.	Increased of at least 30% in the sum of SULpeak of all target lesions detected at baseline or new FDG-avid lesions are considered UPMD and must be confirmed 4-8 weeks later as CPMD. Progression is not confirmed in case of PMR or SMD at 4-8 weeks and evaluation must be reset.	Progressive disease if: ≥ 4 new lesions (< 1 cm in functional diameter); ≥ 3 new lesions (> 1 cm in functional diameter); ≥ 2 new lesions (> 1.5 cm in functional diameter).

Note: *CPMD* = confirmed progressive metabolic disease; *iCPD* = immune confirmed progressive disease; *imPERCIST* = immunotherapy-modified PET Response Criteria in Solid Tumors; *imRECIST* = immune-related response criteria; *iUPD* = immune unconfirmed progressive disease; *PERCIMT* = PET Response Evaluation Criteria for Immunotherapy; *PERCIST* = PET Response Criteria in Solid Tumors; *RECIST* = Response Evaluation Criteria in Solid Tumors; *VPMD* = unconfirmed progressive metabolic disease.

In a recent systematic review and metanalysis by Park et al. [135], the pooled incidence of pseudo-progression was calculated around 5.0% (95% CI: 3.4%, 6.7%) in patients with NSCLC, around 6.4% (95% CI: 4.6%, 8.3%) in patients with melanoma, around 7.0% (95% CI: 5.2%, 8.6%) in patients with genitourinary cancer and around 2% (one of 45; 95% CI: 0%, 6%) in patients with squamous cell cancer of the head and neck. Interestingly, they also found that the pooled incidence of pseudo-progression was higher in melanoma patients treated with CTLA-4 inhibitor (9.7%) comparing to studies of PD-1/PD-L1 inhibitor monotherapy (5.7%; 95% CI: 4.8%, 6.6%); in particular, in studies of NSLSC patients treated with PD-1 inhibitor, the incidence of pseudo-progression ranges from 3.4% to 6.9%. Indeed, recently Humbert et al. [136] reported that pseudo-progression occurs in more than one third of the NSCLC patients treated with Nivolumab or Pembrolizumab: 58% (29/50) showed a real PD, but more than one-third (11/29) were misclassified as they finally reached a durable clinical benefit (DCB). No standard PETinterim criteria has been found to accurately distinguish responding from non-responding patients.

Anyway, these criteria can help to manage the phenomenon of pseudo-progression. However, no criteria are now able to define "hyper-progression", a response pattern associated mainly with the use of anti-PD-1/PD-L1 agents, but also with the use of anti-CTLA-4 therapy. Hyper-progression is defined as a twofold or greater increase of tumor

volume growth rate during immunotherapy, and it is associated with a worse clinical outcome, high mortality, and median overall survival of 3-6 months. Some hypothesis has been developed, such as T regulatory cell expansion and T-cell exhaustion; however, the prevalence, the natural history, and the predictive factors of hyper-progression in patients with cancer treated by anti-PD-1/PD-L1 remain unknown [115].

In 2017, Champiat et al. [137] analyzed the medical records from all patients (N = 218) prospectively treated at Gustave Roussy with anti-PD-1/PD-L1 within phase I clinical trials. Hyper-progression was found in 9% of the population eligible for the study (9/131 patients), it was associated with a higher age (p < 0.05) and a worse outcome, but any association was found with higher tumor burden at baseline, nor with any specific tumor sub-type. More recently, other quantitative PET parameters such as metabolic tumor volume (MTV) and total lesion glycolysis (TLG), which is calculated by multiplying SU-Vmean to the MTV of the selected region of interest (ROI), have also been used to measure tumor burden, define as the total volume of tumor lesions with increased metabolic activity, and their use has been recently implemented in both scientific papers and clinical practice [128]. In 2020, Castello et al. [138] investigated the prevalence of hyper-progression phenomenon and its association with clinical variables and metabolic parameters by [18F]FDG PET/CT in 50 patients with NSCLC treated with ICIs and prospectively collected. Hyper-progression was found in 14/50 patients and, in contrast to the study of Champiat et al., a statistically significant association was found between hyper-progression and tumor burden, expressed by both TLG (756.1 cm3 for HPD vs. 475.6 cm3 for non-HPD, P = 0.042) and MTV (287.3 for HPD vs. 62.1 for non-HPD, P = 0.011). The median OS for patients with hyper-progression was of 4 months.

4.1.2. [18F]FDG and immune-related adverse events (irAEs)

As previously mentioned, consistent with the ICIs mechanism of action, the immune system activation responsible for the anti-tumor response may also be involved in the onset of autoimmune reactions, where several healthy tissues can be invaded by activated T cells resulting. IrAEs commonly manifest within the first 3 months of ICIs administration, even if they could appear at any time, and are most frequently associated with combination treatments (anti-CTLA-4 plus anti-PD-1 or anti-PD-L1), followed by anti-CTLA-4 drugs and finally by anti-PD-1 or anti-PD- L1 drugs. The onset of these immune-related adverse events (irAEs), such as hypophysitis, pneumonia, colitis, hepatitis, and thyroiditis, is unpredictable and, though mild in most cases, can still be life-threatening, thus requiring prompt recognition and appropriate immune-suppressive therapy.

Compared with morphologic imaging, [18F]FDG PET/CT images can early detect irAEs due to increased uptake of [18F]FDG at these sites. This potentially allows for a rapid intervention in life-threatening cases, such as severe colitis, pneumonitis and pancreatitis [115,139].

In 2018, Aide et al. [140] have compiled a checklist for researching immune-related side effects: - measure the spleen and the liver-to-spleen [18F]FDG uptake ratio, an inversion of this ratio reflecting immune activation preceding T cell proliferation; - consider whether the pattern of new nodal uptake suggests sarcoidosis (lambda sign); - refer to baseline scan when an organ frequently showing increased physiological uptake is thought to be involved by an immune-related side effect (thyroid, stomach); - check the pituitary gland; - considering that irAEs, such as colitis and pneumonitis, may need treatment withdrawal or corticosteroid treatment; - bilateral adrenal enlargement and increased uptake is probably due to adrenalitis.

In NSCLC patients, the most common irAEs are rash and diarrhea with nivolumab treatment and hypo- and hyperthyroidism with pembrolizumab treatment [141]. In particular, thyroiditis has an incidence of 5%–23% and [18F]FDG uptake in the thyroid gland can depict thyroid inflammation even before laboratory changes are observed and allows prediction of development of thyroid dysfunction, as demonstrated by the study of Eshgh et al. [142], performed on 18 patients with NSCL and treated with nivolumab.

Interestingly , it has been recently shown that the onset of irAEs may also predict ICIs clinical benefit [143–145]. Even though the clinical implications of this evidence are limited to date, in future, the identification of predictive factors of onset and especially of severity of irAE, may further guide treatment decision on how ICI should be administered to maximize response and safety together.

4.2. [18F]FDG, radiomic and AI

The utility of noninvasive imaging has been further enhanced by the introduction of radiomics and artificial intelligence (AI), which improved the interpretation of medical image datasets. Radiomic is a new innovative bioinformatic approach to the image's analysis, that allows to evaluate tumor heterogeneity and quantify parameters, namely radiomic features (RFs), using standardized mathematical based models. RFs include first-order statistical functions, such as conventional parameters (i.e.: the standard uptake value (SUV), the metabolic tumor volume (MTV) or the total lesion glycolysis (TLG)), histogram and shape PET parameters, and second-order and high-order statistical functions, which contain also information about the spatial relationships between the intensities of more than two voxels (such as texture parameters). RFs can be applied both in clinical practice (especially conventional parameters) and in clinical research, supporting and implementing the clinical decision support system (CDSS) [146-148]. More recently, AI-based approaches have been applied to medical imaging in combination with radiomics or alone, allowing relevant features to be identified with several fully automatic or semi-automatic approaches (machine learning and deep learning) [149-152]. These new approaches could implement the assessment of different aspects and stages of the lung cancer history: to differentiate histological subtypes of cancer, particularly, adenocarcinoma and squamous cell carcinoma, to predict EGFR mutation status or PD-L1 expression, for the risk stratification of NSCLC patients and to predict response to different kind of therapies [153].

4.2.1. Molecular profiling

Radiogenomic is one of the most interesting fields of application of radiomics, allowing the correlation of quantitative imaging features (representative of the phenotype of the genomic signature) with genomic profiles. Thus, radiogenomic represents a non-invasive, whole-body, and repeatable method for extracting molecular information from images. Indeed, several studies tried to identify radiomic signature of NSCLC patients trough [18F]FDG PET/CT images, especially regarding EGFR mutation.

As mentioned above, actionable mutations in the driver oncogenes *EGFR* and *ALK* confer resistance to ICIs [50,51]. For that reason, studies on [18F]FDG PET/CT images that aimed to identify the clinico-radiologic predictors of tumors with *KRAS*, *EGFR*, *BRAF*, *HER2*, *MET*, *ALK*, *ROS1*, and *RET* mutations in NSCLC patients are useful not only with regard to targeted therapies, but also for a possible predictive value of the response to ICI [154–161].

While several studies demonstrated how imaging phenotypes were connected to somatic mutations through an integrated analysis of NSCLC patients with somatic mutation testing and [18F]FDG PET/CT images using both radiomic and AI, only one study by Jiang at al. [162] explored the potential value of radiomics features from [18F]FDG PET/CT images in assessing different PD-L1 mutational status in 399 NSCLC. Models that comprised radiomic features from the CT, the PET, and the PET/CT images resulted in an AUC of 0.97, 0.61, and 0.97, respectively, for prediction of PD-L1 (SP142) expression level over 1% and in an AUC of 0.91, 0.75, and 0.88, respectively, for PD-L1 expression level over 50%.

4.2.2. Staging and prognostic risk stratification: prognostic and predictive value

Several studies have evaluated the prognostic and predictive role of conventional PET parameters (SUVmax or SUVmean) in NSCLC patients who are candidates to ICIs.

In 2016, Lopci et al. [163] investigated the correlation between baseline (prior to surgery) [18F]FDG PET/CT parameters and ICs tissue expression and other markers of tumor-related immunity in 55 resected NSCLC patients (stage IA-IIIB; 36 adenocarcinomas, 18 squamous cell carcinomas and 1 sarcomatoid carcinoma). Sampled surgical tumor specimens were analyzed by immunohistochemistry (IHC) for CD68-TAMs (tumor-associated macrophages), CD8-TILs (tumor infiltrating lymphocytes), PD-1-TILs, and PD-L1 tumor expression. Median SUVmax 11.3 (range: 2.3-32.5) resulted significantly higher in squamous cell carcinomas compared to other NSCLC histotypes (p = 0.007). Moreover, a statistically significant correlation between SUVmax and SUVmean with the expression of CD8+ TILs (rho = 0.31; p = 0.027) and PD-1 (rho = 0.33; p = 0.017 and rho = 0.36; p = 0.009, respectively) was found.

In 2019, Takada et al. [164] aimed to study the association between [18F]FDG PET/CT and the response to anti-PD-1 therapy in 89 NSCLC patients with advanced or recurrent disease. Among the 89 patients evaluated, 24 patients were classified as responders with an average SUVmax of 15.60, and 65 patients as non-responders with an average SUVmax of 11.61 (p=0.0016). The response rate in patients with SUVmax \geq 11.16 (41.3% [19/46]) was significantly higher compared to patients with SUVmax < 11.16 (11.6% [5/43], p=0.0012).

After these studies, Grizzi et al. [165] reported their preliminary data analysis on 27 NSCLC patients, stage IV or IIIB and ineligible for local therapy, treated with monoclonal IgG targeting PD-1 or PD-L1 as single agent (54 patients with nivolumab, 21 with pembrolizumab and 5 with atezolizumab, respectively). In contrast with the aforementioned study, their data documented how, classifying patients as "fast progressors" or "responders" after 8 weeks of treatment, in almost all cases with fast progression SUVmax was ≤17.1 (sensitivity 88.9%) or SUVmean ≤8.3 (sensitivity 100%).

More recently, new semiquantitative parameters, namely metabolic tumor volume (MTV) and total lesion glycolysis (TLG) rate, have been studied to evaluate the predictive and prognostic value of the metabolic tumor burden at baseline [18F]FDG PET/CT. Many of these studies have shown that new PET metabolic parameters are better in predicting prognosis in early and advanced NSCLC and SCLC patients than conventional PET parameters (SUVmax or SUVmean) [166,167].

Higher MTV was significantly related with worse PFS (P<0.001) and OS (P<0.001) in 61 patients with treated NSCLC [168]. The same group [169] also found that a 25-mL increase in MTV in the restaging PET of 19 patients with lung cancer (18 NSCLC and 1 SCLC) was associated with a statistically increased risk of progression (5.4-fold; P=0.0014) and death (7.6-fold; P=0.001), respectively.

In 2020, Polverari et al. [170] evaluated predictive factors of response to immunotherapy in 57 advanced NCSLC patients, demonstrating that patients with higher MTV and TLG had higher probability of disease progression, compared to those patients presenting with lower values. SUVmax did not show correlation with PD status, PFS and OS. Moreover, patients with high MTV, TLG and heterogeneity evaluated by radiomic features "skewness" and "kurtosis" had a higher probability of failing ICIs.

Moreover, radiomic and AI have been successfully used also for the prognostic risk stratification. In a study performed by Kim et al. [171], [18F]FDG PET/CT parameters, clinical, mRNA-seq, and whole exome-seq data of 11 adenocarcinoma and 20 squamous cell carcinoma from The Cancer Genome Atlas (TCGA) database were analyzed. In linear regression analysis, texture parameters such as low gray-level run emphasis (LGRE, R2=0.48, p<0.0001), short run low gray-level emphasis (SRLGE, R2=0.45, p<0.0001) and long run low gray-level emphasis (LRLGE, R2=0.41, p=0.0001) showed remarkable correlation with PD-L1 mRNA expression.

More recently, Valentinuzzi et al. [172] created a radiomics signature (iRADIOMICS) able to predict response of metastatic NSCLC (stage IV) to pembrolizumab compared to the clinical standards (iRECIST and PD-L1 immunohistochemistry). Thirty patients receiving pembrolizumab were scanned with [18F]FDG PET/CT at baseline, month 1 and 4. Multivariate baseline iRADIOMICS was found superior to the current standards in terms of predictive power, timewise (AUC (95% CI) and accuracy = 0.90 (0.78-1.00), 78% (18%)).

Other parameters, such as spleen-to-liver ratio (SLR) or bone marrow-to-liver (BLR) SUVmax ratio, were shown to represent surrogates (indirect index) of the hematopoietic tissue metabolism. Seban et al. [173] proposed a prognostic score combining baseline total metabolic tumor volume (TMTV) and the Derived Neutrophil-to-Lymphocyte Ratio (dLNR) to predict OS in 80 NSCLC patients receiving ICIs. Median follow-up was 11.6 months (95%CI 7.7-15.5). Progression and death occurred in 64 and 52 patients, respectively; disease clinical benefit (DCB) was reported in 40% of cases. Baseline tumor burden (TMTV) > 75 cm3 on [18F]FDG PET/CT scans and inflammatory status (dNLR) > 3 were associated with poor OS and absence of DCB for ICI treatment in advanced NSCLC patients, unlike tumor SUVmax, and might be used in the future in association to improve the selection of appropriate candidates.

In 2021, Bauckneht et al. [174] aimed to identify reliable biomarkers (systemic inflammation indexes and PET imaging parameters at the time of radiological progression) able to discriminate between responders and non-responders among 45 patients showing imaging progression (defined as RECIST 1.1 progressive disease) during nivolumab administration for advance NSCLC. Among all parameters collected, MTV and systemic inflammation index (SII) independently predicted OS. Moreover, they developed the immunemetabolic prognostic index (IMPI) based on these two parameters, categorizing the enrolled cases in three groups with different risk of progression as follows: low (neither MTV \geq 208.01 nor SII \geq 197.21, IMPI = 0, n = 11), intermediate (MTV \geq 208.01 or SII \geq 197.21, IMPI = 1, n = 23), and high IMPI (MTV \geq 208.01 and SII \geq 197.21, IMPI = 2, n = 11). Median OS was 17.5 month (95% CI 11.3–31.5 months), 9.4 months (95% CI 5.6–33.6 months), 3.2 months (95% CI 2.1–18.5 months) for the low, intermediate, and high IMPI groups, respectively (p < 0.0001). This interesting index, if validated, may allowed the identification of patients who might benefit from immunotherapy continuation, despite radiological progression.

4.3. Functional imaging probe beyond [18F]FDG

[18F]FDG PET/CT is still the mainstay in the evaluation of NSCLC patients treated with ICI and new technologies, radiomics and AI have expanded the ability of this radio-tracer to address open questions about ICIs mechanisms. Nevertheless, [18F]FDG is not able to assess the finer mechanisms underlying tumor ICs expression and their inhibition.

For this reason, in the last decades, new molecular imaging probes have been developed to improve our knowledge on TME, immune-system and ICI. Even if biopsy and tissue analysis are still mandatory and strictly necessary, *in-vivo* molecular imaging of ICs expression may be more accurate than *in-vitro* analysis. Moreover, inducible T-cell costimulatory receptor (ICOS) PET radiotracers may predict early response to therapy.

As aforementioned, biopsy specimen is the first fundamental step to identify PD-L1 tumor expression and/or PD-1 and CTLA-4 T cells expression and for therapy and patient selection. However, in clinical practice generally only a single-tumor biopsy is performed, leading to potential underestimation of tumor heterogeneity of ICs expression both spatial- (inter- and intra-tumoral) and time-related (more aggressive cell clones developing over time) [175,176].

As previously mentioned, the spatial immune infiltration patterns ('topography') across cancer entities and across various immune cell types is a central concept to understand the mechanisms underlying the response to ICIs immunotherapy [177].

Three distinct "immune topographies" have been identified: *hot tumors* characterized by lymphocytes infiltration, mixed with tumor cells in the tumor core; *cold tumors*, characterized by an absence of lymphocytes infiltrations (i.e., almost no lymphocytes can be seen on histological slides); and *immune-excluded tumors* characterized by an abundance of lymphocytes at the invasive edge of the tumor, but few to no lymphocytes in the tumor core [178].

ImmunoPET radiotracer, ICOS PET radiotracer and [18F]FDG give the possibility to systematically mapping the "Immune Topographies" in both spatial (whole-body) and

temporal distribution in NCSLC [118]. These techniques trough whole-body specific target expression probes could improve the current knowledge on TME and all the dynamic mechanisms behind tumor development and growth including the relationship between tumor cells and infiltrating immune cells in the TME, and grading heterogeneity among primary and metastasis.

If validated, in the future these new radiotracers could help clinicians' decision in clinical practice stratifying at baseline patients who are good candidates for immunotherapy, identifying earlier response assessment, distinguishing tumor progression from pseudo-progression (especially immunoPET radiotracers combined with [18F]FDG PET/CT images), monitoring "immune topographies" changes during ICIs, and identifying potential non-responding patients (patients which ICOS PET shows low or no uptake of activate T cells) and/or lesions.

New radiotracers that may have a role in the evaluation of NSCLC patients, who are candidates for immunotherapy, and are currently on study in clinical trials are described below.

4.3.1. PD-1/PD-L1 Pathways

Several techniques have been implemented to find the best radiotracer, specific for an antigen expression only on target cells labelled with an optimal radionuclide for imaging. For example, antibody fragments and small proteins labeled with [18F] and [68Ga] (half-life of 109.8 and 67.7 min, respectively) are preferred over immunoglobulins G (IgG) antibodies (mAbs) labeled with [111In], [89Zr] and [64Cu] (half-life of 67.2, 78.4 and 12.7 h, respectively) due to their shorter biological half-life [13,179,180].

Since 2015, after the development of the first [64Cu]DOTA labeled antibody (IgG) PD-1 radiotracer [181], several immune checkpoint radiotracers have been developed for PET imaging, but only few of them have been tested in human.

The first-in-human study was presented by Bensch at al. [182] in 2019. They studied [89Zr]atezolizumab PET/CT images in 22 patients with different cancers (metastatic bladder cancer, NSCLC, or triple-negative breast cancer). To prevent rapid clearance during imaging, which was performed on days 4 and 7, 10 mg of unlabeled atezolizumab was administered together with the radiotracer. [89Zr]atezolizumab uptake was found to be physiologically high in bone marrow, intestines, kidney, and liver and low in healthy brain, subcutaneous tissue, muscle, compact bone, and lung, while the uptake in lymphoid tissue (lymph nodes and spleen mainly) depends from the activation state of the immune system. They found that pre-treatment radiotracer uptake better correlated with PFS and OS, comparing to conventional IHC staining of PD-L1, highlighting the limitations of a single biopsy evaluation. Moreover, tumor [89Zr]atezolizumab uptake was generally high (mean SUVmax of 10.4), but often with an high intratumoral, intertumoral and interpatient heterogeneity distribution.

In 2018, Niemeijer et al. [183] studied 13 patients with advanced NSCLC before treatment with nivolumab using two new immuno-PET radiotracers: an anti-PD-1 radiotracer, [89Zr]nivolumab (images acquired after 7 days after injection) and an anti-PD-L1 radiotracer, [18F]labeled anti-PD-L1 fibronectin-based protein (adnectin), namely [18F]BMS-986192 (images acquired after 1 hour after injection). Both radiotracers showed favorable distribution in tumor, but with a high intratumoral, intertumoral and interpatient heterogeneity distribution. Higher uptake of both radiotracers was found in responder patients: [18F]BMS-986192 SUV peak (median 6.5 (responder) vs. 3.2 (non-responder), p = 0.03), and a similar association was found for [89Zr]nivolumab (median SUV peak 6.4 vs. 3.9, p = 0.019). Spleen uptake reflected the presence of immune cells, and both radiotracers were catabolized in the liver. The median [18F]BMS-986192 uptake was higher for lesions with $\geq 50\%$ tumor PD-L1 expression evaluated by IHC than for lesions with $\leq 50\%$ expression. Similarly, [89Zr]nivolumab uptake was higher in lesions with a

higher aggregates of PD-1 positive tumor-infiltrating immune cells at tumor biopsy. Finally, due to low central nervous system (CNS) tracer penetration, both tracers showed lower accumulation in brain metastases compared to the extracerebral lesions.

4.3.2. CD8+ T Lymphocytes

A great effort has been made in recent years to optimize ICOS PET radiotracers for the detection of CD8+ and CD4+ T cells in the TME.

The only ICOS PET radiotracer tested in human is a radiolabeled minibody against tumor-infiltrating CD8-positive (CD81) T lymphocytes, with a fast blood clearance due to its small size, which allows for early image acquisition.

In 2020, Pandit-Taskar et al. [184] assessed the safety and utility of [89Zr]IAB22M2C for targeted imaging of CD81 T cells in cancer patients (1 melanoma, 4 NSCLC, and 1 hepatocellular carcinoma). The highest uptake was observed in the spleen, followed by the bone marrow. The maximum uptake in normal lymph nodes was reached between 24–48h. Uptake in tumor lesions was seen already 2 h after injection, but the uptake in [89Zr]IAB22M2C–positive lesions increased until 24 h (SUV from 5.85 to 22.8). No side effects were registered, and serum clearance was biexponential.

4.3.3. Cancer-associated fibroblasts (CAFs)

As previously mentioned, the *tumor microenvironment* (TME) is composed by an organic and evolving complex of tumor, stromal and tumor-infiltrating immune cells, coevolving and growing within a protective extracellular matrix (ECM) with the help of cytokines, glycoproteins, proteoglycans and growth factors [118].

In this context, cancer-associated fibroblasts (CAFs) play an essential role in tumor growth, integrity of TME architecture and function, tumor progression, metastasis development, and therapeutic resistance through potent immunosuppressive activity, conferring resistance to immune-based therapies. CAFs are activated by tumor cells through the secretion of factors such as TGF- β , PDGF, EGF, CTGF, and FGF, or in response to damage-associated molecular patterns (DAMPs) released by damaged tissue or necrotic tumor cells [185].

Indeed, a greater understanding of how tumor stroma and TME comprehensively affect immunotherapy will provide important advances: CAF markers correlate with T-cell immunosuppression, blocking the effects of both CD8+ T cells and NK cells, especially through the release of various chemokines and cytokines (mainly IL-6 production), and leading to poor clinical outcome. It is for these reasons that CAF depletion and targeting CAF-dependent pathways may indirectly lead to malignant cell death through both immune-dependent and immune-independent mechanisms [186].

In this context, fibroblast activation protein (FAP), a transmembrane serine protease and marker of CAF activation, is exclusively synthesized and highly expressed by CAFs from almost all tumors, whereas it is relatively absent in normal stromal cells. FAP is an important CAF protein associated with the outcome of several immunotherapies and its presence in histological staining correlates with poor prognosis for most cancer patients.

A study has shown that in models of lung cancer where the presence of FAP is genetically depleted, this led to hypoxic necrosis in the tumor in addition to an induction of CD8+ T-cell infiltration [187],

Recently, researchers at the University of Heidelberg have introduced PET imaging of FAP expression in cancer, producing radiopharmaceuticals derived from quinoline peptidomimetics that bind with high affinity to FAP expressed on CAFs labeling with [68Ga] [188,189]. While [18F]FDG visualizes tumor cell glycolysis, [68Ga]-DOTA-FAPi predominantly depicts tumor stroma, which is influenced by the amount of tumor stroma and the level of FAP expression in the tumor stroma, and is extremely promising due to the excellent visualization of the tumor resulting from the high tumor-to-background ratio (TBR) [190,191].

FAPi PET can provide diagnostic, predictive, prognostic, and phenotypic informations. A combined [18F]FDG and FAPi PET imaging approach to determine the relationship between tumor and stroma can therefore be highly informative [191,192]. Moreover, FAPi can serve as theranostic: the high TBR can be exploited by labeling FAPi with alpha- or beta-emitting isotopes to generate a potentially potent therapy [193–195].

However, FAPI PET is still in a preliminary study phase and all the advantages and limitations of this radifarmaceutical are still being understood, so it will take some time before widespread adoption can become feasible [196]. For example, radiation may induce fibrosis, limiting the usefulness of FAPI PET for decision making after therapy. Moreover, fibroblast activation could be associated with chronic inflammation, with liver and lung fibrosis and with checkpoint inhibitor-associated myocarditis, reducing the specificity and limit the diagnostic value of FAPI PET and further studies are needed [197–199].

4.3.4. Other probes

Many other tracers may be useful in the evaluation and monitoring of immunotherapy candidate patients outside the triad immunoPET radiotracer, ICOS PET radiotracer and [18F]FDG.

New PET imaging tracers are currently being developed to assess the presence of hypoxia in tumor tissue, a hallmark of carcinogenesis and tumor resistance to systemic therapy. Currently, there are mainly three major radiotracer of hypoxia PET tracers:

- 18F-fluoromisonidazole ([18F]F-FMISO), which enters cells via passive diffusion, where it is reduced by nitroreductase enzymes under hypoxic conditions. [18F]F-FMISO PET/CT images are challenging because images must be acquired several hours after the radiotracer adminstration;
- 18F-fluoroazomycin arabinoside ([18F]F-FAZA) are more hydrophilic hypoxia probes. Unfortunately, imaging of this tracer is also challenging because image quality (resolution of tissue-hypoxia ratios) depends on renal clearance, which could lead to measurements variability;
- 64Cu-labeled diacetyl-bis(N-methylthiosemicarbazone) analogs ([64Cu]Cu-ATSM) is an alternative class of agents with low molecular weight and lipophilicity, resulting in high membrane permeability. Its use in the clinical setting is more handleable [200,201].

These radiotracers have been studied mainly to assess tumor resistance in chemotherapy and radiotherapy setting. However, as recently reported by Pietrobon et al. [202], the exclusion of lymphocytes from the tumor nest is a common phenomenon that limits the efficiency of ICI in solid tumors (*cold tumors*) and one hypothesis is that hypoxia, a hallmark of most solid tumors, may be a common biological determinant of immune exclusion in solid tumors.

To the best of our knowledge, to date, no study regarding hypoxia tracers in correlation with immunotherapy has been done in NSCLC. However, one study of Rühle et al. [203] aimed to investigate the interaction between tumor hypoxia dynamics and the PD-1/PD-L1 axis in 49 patients with head-and-neck cancer (HNSCC) undergoing chemoradiation and its relevance for patient outcomes in a prospective trial. Patients performed [18F]F-FMISO PET/CT at weeks 0, 2 and 5 during treatment; SUV index was defined as ratio of maximum tumor [18F]FMISO SUV to mean SUV in the contralateral sternocleido-mastoid muscle (i.e. tumor-to-muscle ratio). PET parameters and the PD-1/PD-L1 expression both on immune cells and on tumor cells obtained from pre-therapeutic tumor biopsies were analyzed. Rühle et al. interestingly found that patients with no hypoxia resolution between weeks 0 and 2 and PD-L1 expression on tumor cells (TPS of at least 1%) showed significantly worse locoregional control (LRC; HR = 3.374, p = 0.022) and a trend towards reduced PFS (HR = 2.752, p = 0.052), while the tumor cells' PD-L1 expression did not influence the outcomes of patients with early tumor hypoxia resolution.

As previous mentioned, *VEGF* and *VEGFR* are upregulated and tumor angiogenesis often produces blood vessels with aberrant morphology, excluding T-cell migration. Due to the therapeutic development of combination of anti-angiogenetic drugs with ICI, anti-*VEGF* or anti-*VEGFR* monoclonal antibodies labelled with diagnostic or therapeutic radionuclide-bound may become optimal radiotracers for PET images [204].

Bevacizumab is a IgG1 antibodies directed against *VEGF-A*. Radiolabeled bevacizumab, [64Cu]bevacizumab, is the only PET radiotracer of angiogenesis that has been tested in clinical studies, performed in patients with breast cancer, neuro-endocrine tumors (NET), renal cell carcinoma, and NSCLC [205]. In 2014, Bahce et al. [206] aimed to evaluate whether the uptake of [64Cu]bevacizumab in 7 patients with advanced NSCLC tumors could be visualized and quantified, prior to treatment with bevacizumab combined with carboplatin and paclitaxel. Zr-bevacizumab uptake (SUVpeak) was approximately four times higher in tumor tissues (primary tumor and metastases) than in nontumor tissues (healthy muscle, lung, and fat) on days 4 and 7. The level of tracer uptake was associated with improved progression free survival (PFS) and overall survival (OS). Moreover, tumor tracer uptake was blocked by adding non-labeled bevacizumab, supporting the notion of a specific binding.

Even if *VEGF* plays a prominent role in mediating T-cells infiltration into tumors and in regulating their function until now, no studies have been focused on angiogenesis PET radiotracer and immunotherapy,

5. Conclusions

In recent decades, the impressive improvement in translational medicine has led to a better understanding of the mechanisms behind tumor development and interaction with TME setting and to the implementation of ICI-based treatment options.

The way forward to create an accurate multivariable platform to predict ICI outcome is still long. Nonetheless, the continuous clinical and scientific collaboration between oncologists, pathologists, geneticists and nuclear medicine physicians, could be the winning strategy for the development of a composite assessment of tissue, circulating and in vivo biomarkers, refining our ability to predict response to ICI, ultimately improving patients' selection for more tailored ICI-based therapy.

6. Patents

Author Contributions: Substantial contributions to the conception or design of the work: DD, VL and AM; Literature search: VL and AM; Article selection: VL, AM and LR; Manuscript preparation: VL and AM; Drafting the work or revising it critically for important intellectual content: all authors. All authors have read and agreed to the published version of the manuscript.

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References

- 1. WHO (World Health Organization) Oesophageal Source: Globocan 2020 Number of new cases in 2020, both sexes, all ages; 2020;
- Bray, F.; Ferlay, J.; Soerjomataram, I.; Siegel, R.L.; Torre, L.A.; Jemal, A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA. Cancer J. Clin. 2018, 68, 394

 –424, doi:10.3322/caac.21492.
- 3. Reck, M.; Heigener, D.F.; Mok, T.; Soria, J.C.; Rabe, K.F. Management of non-small-cell lung cancer: Recent developments. Lancet

- 2013, 382, 709-719.
- 4. Zhang, N.; Wu, J.; Yu, J.; Zhu, H.; Yang, M.; Li, R. Integrating Imaging, Histologic, and Genetic Features to Predict Tumor Mutation Burden of Non–Small-Cell Lung Cancer. *Clin. Lung Cancer* **2020**, *21*, e151–e163, doi:10.1016/j.cllc.2019.10.016.
- 5. Herbst, R.S.; Morgensztern, D.; Boshoff, C. The biology and management of non-small cell lung cancer. Nature 2018, 553, 446-454.
- Bray, F.; Ferlay, J.; Soerjomataram, I.; Siegel, R.L.; Torre, L.A.; Jemal, A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA. Cancer J. Clin. 2018, 68, 394–424, doi:10.3322/caac.21492.
- 7. Planchard, D.; Popat, S.; Kerr, K.; Novello, S.; Smit, E.F.; Faivre-Finn, C.; Mok, T.S.; Reck, M.; Van Schil, P.E.; Hellmann, M.D.; et al. Metastatic non-small cell lung cancer: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann. Oncol.* **2018**, 29, iv192–iv237, doi:10.1093/annonc/mdy275.
- 8. Zhang, Y.; Zhang, Z. The history and advances in cancer immunotherapy: understanding the characteristics of tumor-infiltrating immune cells and their therapeutic implications. *Cell. Mol. Immunol.* 2020, *17*, 807–821.
- 9. Waldman, A.D.; Fritz, J.M.; Lenardo, M.J. A guide to cancer immunotherapy: from T cell basic science to clinical practice. *Nat. Rev. Immunol.* 2020, *20*, 651–668.
- 10. Galli, F.; Aguilera, J.V.; Palermo, B.; Markovic, S.N.; Nisticò, P.; Signore, A. Relevance of immune cell and tumor microenvironment imaging in the new era of immunotherapy. *J. Exp. Clin. Cancer Res.* 2020, 39.
- 11. Gonzalez, H.; Hagerling, C.; Werb, Z. Roles of the immune system in cancer: From tumor initiation to metastatic progression. *Genes Dev.* 2018, *32*, 1267–1284.
- 12. Hashimoto, M.; Kamphorst, A.O.; Im, S.J.; Kissick, H.T.; Pillai, R.N.; Ramalingam, S.S.; Araki, K.; Ahmed, R. CD8 T Cell Exhaustion in Chronic Infection and Cancer: Opportunities for Interventions. *Annu. Rev. Med.* 2018, 69, 301–318, doi:10.1146/annurev-med-012017-043208.
- 13. Bouleau, A.; Lebon, V.; Truillet, C. PET imaging of immune checkpoint proteins in oncology. *Pharmacol. Ther.* 2021, 222, 107786.
- 14. Sun, C.; Mezzadra, R.; Schumacher, T.N. Regulation and Function of the PD-L1 Checkpoint. *Immunity* 2018, 48, 434–452.
- 15. Barber, D.L.; Wherry, E.J.; Masopust, D.; Zhu, B.; Allison, J.P.; Sharpe, A.H.; Freeman, G.J.; Ahmed, R. Restoring function in exhausted CD8 T cells during chronic viral infection. *Nature* **2006**, *439*, 682–687, doi:10.1038/nature04444.
- Day, C.L.; Kaufmann, D.E.; Kiepiela, P.; Brown, J.A.; Moodley, E.S.; Reddy, S.; Mackey, E.W.; Miller, J.D.; Leslie, A.J.; DePierres,
 C.; et al. PD-1 expression on HIV-specific T cells is associated with T-cell exhaustion and disease progression. *Nature* 2006, 443, 350–354, doi:10.1038/nature05115.
- 17. Schildberg, F.A.; Klein, S.R.; Freeman, G.J.; Sharpe, A.H. Coinhibitory Pathways in the B7-CD28 Ligand-Receptor Family. *Immunity* 2016, 44, 955–972.
- 18. Tumeh, P.C.; Harview, C.L.; Yearley, J.H.; Shintaku, I.P.; Taylor, E.J.M.; Robert, L.; Chmielowski, B.; Spasic, M.; Henry, G.; Ciobanu, V.; et al. PD-1 blockade induces responses by inhibiting adaptive immune resistance. *Nature* **2014**, *515*, 568–571, doi:10.1038/nature13954.
- 19. Topalian, S.L.; Hodi, F.S.; Brahmer, J.R.; Gettinger, S.N.; Smith, D.C.; McDermott, D.F.; Powderly, J.D.; Carvajal, R.D.; Sosman, J.A.; Atkins, M.B.; et al. Safety, Activity, and Immune Correlates of Anti–PD-1 Antibody in Cancer. *N. Engl. J. Med.* 2012, 366, 2443–2454, doi:10.1056/nejmoa1200690.
- 20. Taube, J.M.; Klein, A.; Brahmer, J.R.; Xu, H.; Pan, X.; Kim, J.H.; Chen, L.; Pardoll, D.M.; Topalian, S.L.; Anders, R.A. Association of PD-1, PD-1 ligands, and other features of the tumor immune microenvironment with response to anti-PD-1 therapy. *Clin. Cancer Res.* **2014**, *20*, 5064–5074, doi:10.1158/1078-0432.CCR-13-3271.
- 21. Korman, A.J.; Peggs, K.S.; Allison, J.P. Checkpoint Blockade in Cancer Immunotherapy. Adv. Immunol. 2006, 90, 297–339.
- Simpson, T.R.; Li, F.; Montalvo-Ortiz, W.; Sepulveda, M.A.; Bergerhoff, K.; Arce, F.; Roddie, C.; Henry, J.Y.; Yagita, H.; Wolchok, J.D.; et al. Fc-dependent depletion of tumor-infiltrating regulatory t cells co-defines the efficacy of anti-CTLA-4 therapy against melanoma. *J. Exp. Med.* 2013, 210, 1695–1710, doi:10.1084/jem.20130579.
- 23. Selby, M.J.; Engelhardt, J.J.; Quigley, M.; Henning, K.A.; Chen, T.; Srinivasan, M.; Korman, A.J. Anti-CTLA-4 antibodies of IgG2a

- isotype enhance antitumor activity through reduction of intratumoral regulatory T cells. *Cancer Immunol. Res.* **2013**, *1*, 32–42, doi:10.1158/2326-6066.CIR-13-0013.
- 24. Hanna, N.H.; Schneider, B.J.; Temin, S.; Baker, S.; Brahmer, J.; Ellis, P.M.; Gaspar, L.E.; Haddad, R.Y.; Hesketh, P.J.; Jain, D.; et al. Therapy for stage IV non–small-cell lung cancer without driver alterations: ASCO and OH (CCO) joint guideline update. *J. Clin. Oncol.* 2020, *38*, 1608–1632, doi:10.1200/JCO.19.03022.
- 25. Vaddepally, R.K.; Kharel, P.; Pandey, R.; Garje, R.; Chandra, A.B. Review of indications of FDA-approved immune checkpoint inhibitors per NCCN guidelines with the level of evidence. *Cancers (Basel)*. 2020, *12*.
- 26. Fridman, W.H.; Zitvogel, L.; Sautès-Fridman, C.; Kroemer, G. The immune contexture in cancer prognosis and treatment. *Nat. Rev. Clin. Oncol.* 2017, *14*, 717–734.
- Zou, Y.; Hu, X.; Zheng, S.; Yang, A.; Li, X.; Tang, H.; Kong, Y.; Xie, X. Discordance of immunotherapy response predictive biomarkers between primary lesions and paired metastases in tumours: A multidimensional analysis. *EBioMedicine* 2021, 63, doi:10.1016/j.ebiom.2020.103137.
- 28. Liberati, A.; Altman, D.G.; Tetzlaff, J.; Mulrow, C.; Gøtzsche, P.C.; Ioannidis, J.P.A.; Clarke, M.; Devereaux, P.J.; Kleijnen, J.; Moher, D. The PRISMA statement for reporting systematic reviews and meta-analyses of studies that evaluate health care interventions: explanation and elaboration. In Proceedings of the Journal of clinical epidemiology; J Clin Epidemiol, 2009; Vol. 62, pp. e1–e34.
- 29. Hui, R.; Garon, E.B.; Goldman, J.W.; Leighl, N.B.; Hellmann, M.D.; Patnaik, A.; Gandhi, L.; Eder, J.P.; Ahn, M.J.; Horn, L.; et al. Pembrolizumab as first-line therapy for patients with PD-L1-positive advanced non-small cell lung cancer: A phase 1 trial. *Ann. Oncol.* **2017**, *28*, 874–881, doi:10.1093/annonc/mdx008.
- 30. Patel, S.P.; Kurzrock, R. PD-L1 expression as a predictive biomarker in cancer immunotherapy. Mol. Cancer Ther. 2015, 14, 847–856.
- 31. Chen, D.S.; Mellman, I. Elements of cancer immunity and the cancer-immune set point. *Nature* 2017, 541, 321–330.
- 32. Borghaei, H.; Paz-Ares, L.; Horn, L.; Spigel, D.R.; Steins, M.; Ready, N.E.; Chow, L.Q.; Vokes, E.E.; Felip, E.; Holgado, E.; et al. Nivolumab versus Docetaxel in Advanced Nonsquamous Non–Small-Cell Lung Cancer. *N. Engl. J. Med.* 2015, 373, 1627–1639, doi:10.1056/nejmoa1507643.
- 33. Brahmer, J.; Reckamp, K.L.; Baas, P.; Crinò, L.; Eberhardt, W.E.E.; Poddubskaya, E.; Antonia, S.; Pluzanski, A.; Vokes, E.E.; Holgado, E.; et al. Nivolumab versus Docetaxel in Advanced Squamous-Cell Non–Small-Cell Lung Cancer. *N. Engl. J. Med.* **2015**, 373, 123–135, doi:10.1056/nejmoa1504627.
- 34. Tsao, M.S.; Kerr, K.M.; Kockx, M.; Beasley, M.B.; Borczuk, A.C.; Botling, J.; Bubendorf, L.; Chirieac, L.; Chen, G.; Chou, T.Y.; et al. PD-L1 Immunohistochemistry Comparability Study in Real-Life Clinical Samples: Results of Blueprint Phase 2 Project. *J. Thorac. Oncol.* 2018, *13*, 1302–1311, doi:10.1016/j.jtho.2018.05.013.
- 35. Herbst, R.S.; Soria, J.C.; Kowanetz, M.; Fine, G.D.; Hamid, O.; Gordon, M.S.; Sosman, J.A.; McDermott, D.F.; Powderly, J.D.; Gettinger, S.N.; et al. Predictive correlates of response to the anti-PD-L1 antibody MPDL3280A in cancer patients. *Nature* **2014**, *515*, 563–567, doi:10.1038/nature14011.
- 36. Schumacher, T.N.; Schreiber, R.D. Neoantigens in cancer immunotherapy. Science (80-.). 2015, 348, 69-74.
- 37. Havel, J.J.; Chowell, D.; Chan, T.A. The evolving landscape of biomarkers for checkpoint inhibitor immunotherapy. *Nat. Rev. Cancer* 2019, *19*, 133–150.
- 38. Hellmann, M.D.; Nathanson, T.; Rizvi, H.; Creelan, B.C.; Sanchez-Vega, F.; Ahuja, A.; Ni, A.; Novik, J.B.; Mangarin, L.M.B.; Abu-Akeel, M.; et al. Genomic Features of Response to Combination Immunotherapy in Patients with Advanced Non-Small-Cell Lung Cancer. *Cancer Cell* 2018, 33, 843-852.e4, doi:10.1016/j.ccell.2018.03.018.
- 39. Alexandrov, L.B.; Nik-Zainal, S.; Wedge, D.C.; Aparicio, S.A.J.R.; Behjati, S.; Biankin, A. V.; Bignell, G.R.; Bolli, N.; Borg, A.; Børresen-Dale, A.L.; et al. Signatures of mutational processes in human cancer. *Nature* **2013**, *500*, 415–421, doi:10.1038/nature12477.
- Rizvi, N.A.; Hellmann, M.D.; Snyder, A.; Kvistborg, P.; Makarov, V.; Havel, J.J.; Lee, W.; Yuan, J.; Wong, P.; Ho, T.S.; et al. Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. *Science* (80-.). 2015, 348, 124–128, doi:10.1126/science.aaa1348.

- 41. Rizvi, H.; Sanchez-Vega, F.; La, K.; Chatila, W.; Jonsson, P.; Halpenny, D.; Plodkowski, A.; Long, N.; Sauter, J.L.; Rekhtman, N.; et al. Molecular determinants of response to anti-programmed cell death (PD)-1 and anti-programmed death-ligand 1 (PD-L1) blockade in patients with non-small-cell lung cancer profiled with targeted next-generation sequencing. *J. Clin. Oncol.* 2018, *36*, 633–641, doi:10.1200/JCO.2017.75.3384.
- 42. Gandara, D.R.; Paul, S.M.; Kowanetz, M.; Schleifman, E.; Zou, W.; Li, Y.; Rittmeyer, A.; Fehrenbacher, L.; Otto, G.; Malboeuf, C.; et al. Blood-based tumor mutational burden as a predictor of clinical benefit in non-small-cell lung cancer patients treated with atezolizumab. *Nat. Med.* **2018**, *24*, doi:10.1038/s41591-018-0134-3.
- 43. Hellmann, M.D.; Ciuleanu, T.-E.; Pluzanski, A.; Lee, J.S.; Otterson, G.A.; Audigier-Valette, C.; Minenza, E.; Linardou, H.; Burgers, S.; Salman, P.; et al. Nivolumab plus Ipilimumab in Lung Cancer with a High Tumor Mutational Burden. *N. Engl. J. Med.* 2018, 378, 2093–2104, doi:10.1056/nejmoa1801946.
- 44. Hellmann, M.D.; Paz-Ares, L.; Bernabe Caro, R.; Zurawski, B.; Kim, S.-W.; Carcereny Costa, E.; Park, K.; Alexandru, A.; Lupinacci, L.; de la Mora Jimenez, E.; et al. Nivolumab plus Ipilimumab in Advanced Non–Small-Cell Lung Cancer. N. Engl. J. Med. 2019, 381, 2020–2031, doi:10.1056/nejmoa1910231.
- 45. Update on the Phase III NEPTUNE trial of Imfinzi plus tremelimumab in Stage IV non-small cell lung cancer Available online: https://www.astrazeneca.com/media-centre/press-releases/2019/update-on-the-phase-iii-neptune-trial-of-imfinzi-plus-tremelimumab-in-stage-iv-non-small-cell-lung-cancer-21082019.html (accessed on Mar 30, 2020).
- 46. Research, C. for D.E. and FDA approves pembrolizumab for adults and children with TMB-H solid tumors. FDA 2020.
- 47. Litchfield, K.; Reading, J.L.; Puttick, C.; Thakkar, K.; Abbosh, C.; Bentham, R.; Watkins, T.B.K.; Rosenthal, R.; Biswas, D.; Rowan, A.; et al. Meta-analysis of tumor- and T cell-intrinsic mechanisms of sensitization to checkpoint inhibition. *Cell* **2021**, *184*, 596-614.e14, doi:10.1016/j.cell.2021.01.002.
- 48. Wang, Z.; Duan, J.; Wang, G.; Zhao, J.; Xu, J.; Han, J.; Zhao, Z.; Zhao, J.; Zhu, B.; Zhuo, M.; et al. Allele Frequency–Adjusted Blood-Based Tumor Mutational Burden as a Predictor of Overall Survival for Patients With NSCLC Treated With PD-(L)1 Inhibitors. *J. Thorac. Oncol.* **2020**, *15*, 556–567, doi:10.1016/j.jtho.2019.12.001.
- 49. Cai, L.; Bai, H.; Duan, J.; Wang, Z.; Gao, S.; Wang, D.; Wang, S.; Jiang, J.; Han, J.; Tian, Y.; et al. Epigenetic alterations are associated with tumor mutation burden in non-small cell lung cancer. *J. Immunother. Cancer* **2019**, 7, doi:10.1186/s40425-019-0660-7.
- 50. Gainor, J.F.; Shaw, A.T.; Sequist, L. V.; Fu, X.; Azzoli, C.G.; Piotrowska, Z.; Huynh, T.G.; Zhao, L.; Fulton, L.; Schultz, K.R.; et al. EGFR mutations and ALK rearrangements are associated with low response rates to PD-1 pathway blockade in non-small cell lung cancer: A retrospective analysis. *Clin. Cancer Res.* **2016**, *22*, 4585–4593, doi:10.1158/1078-0432.CCR-15-3101.
- 51. Bylicki, O.; Paleiron, N.; Margery, J.; Guisier, F.; Vergnenegre, A.; Robinet, G.; Auliac, J.B.; Gervais, R.; Chouaid, C. Targeting the PD-1/PD-L1 Immune Checkpoint in EGFR-Mutated or ALK-Translocated Non-Small-Cell Lung Cancer. *Target. Oncol.* **2017**, *12*, 563–569, doi:10.1007/s11523-017-0510-9.
- 52. Mazieres, J.; Drilon, A.; Lusque, A.; Mhanna, L.; Cortot, A.B.; Mezquita, L.; Thai, A.A.; Mascaux, C.; Couraud, S.; Veillon, R.; et al. Immune checkpoint inhibitors for patients with advanced lung cancer and oncogenic driver alterations: Results from the IMMUNOTARGET registry. *Ann. Oncol.* 2019, *30*, 1321–1328, doi:10.1093/annonc/mdz167.
- 53. Aggarwal, C.; Thompson, J.C.; Chien, A.L.; Quinn, K.J.; Hwang, W.T.; Black, T.A.; Yee, S.S.; Christensen, T.E.; LaRiviere, M.J.; Silva, B.A.; et al. Baseline Plasma Tumor Mutation Burden Predicts Response to Pembrolizumab-based Therapy in Patients with Metastatic Non–Small Cell Lung Cancer. Clin. Cancer Res. 2020, 26, 2354–2361, doi:10.1158/1078-0432.CCR-19-3663.
- 54. Lamberti, G.; Spurr, L.F.; Li, Y.; Ricciuti, B.; Recondo, G.; Umeton, R.; Nishino, M.; Sholl, L.M.; Meyerson, M.L.; Cherniack, A.D.; et al. Clinicopathological and genomic correlates of programmed cell death ligand 1 (PD-L1) expression in nonsquamous non-small-cell lung cancer. *Ann. Oncol.* 2020, *31*, 807–814, doi:10.1016/j.annonc.2020.02.017.
- 55. Schoenfeld, A.J.; Rizvi, H.; Bandlamudi, C.; Sauter, J.L.; Travis, W.D.; Rekhtman, N.; Plodkowski, A.J.; Perez-Johnston, R.; Sawan, P.; Beras, A.; et al. Clinical and molecular correlates of PD-L1 expression in patients with lung adenocarcinomas. *Ann. Oncol.* **2020**, *31*, 599–608, doi:10.1016/j.annonc.2020.01.065.

- 56. Cho, B.C.; Lopes, G.; Kowalski, D.M.; Kasahara, K.; Wu, Y.-L.; Castro, G.; Turna, H.Z.; Cristescu, R.; Aurora-Garg, D.; Loboda, A.; et al. Abstract CT084: Relationship between STK11 and KEAP1 mutational status and efficacy in KEYNOTE-042: pembrolizumab monotherapy versus platinum-based chemotherapy as first-line therapy for PD-L1-positive advanced NSCLC. In Proceedings of the Cancer Research; American Association for Cancer Research (AACR), 2020; Vol. 80, pp. CT084—CT084.
- 57. Aviel-Ronen, S.; Blackhall, F.H.; Shepherd, F.A.; Tsao, M.S. K-ras mutations in non-small-cell lung carcinoma: A review. *Clin. Lung Cancer* 2006, 8, 30–38.
- 58. Pylayeva-Gupta, Y.; Lee, K.E.; Hajdu, C.H.; Miller, G.; Bar-Sagi, D. Oncogenic Kras-Induced GM-CSF Production Promotes the Development of Pancreatic Neoplasia. *Cancer Cell* **2012**, *21*, 836–847, doi:10.1016/j.ccr.2012.04.024.
- 59. Dong, Z.Y.; Zhong, W.Z.; Zhang, X.C.; Su, J.; Xie, Z.; Liu, S.Y.; Tu, H.Y.; Chen, H.J.; Sun, Y.L.; Zhou, Q.; et al. Potential predictive value of TP53 and KRAS mutation status for response to PD-1 blockade immunotherapy in lung adenocarcinoma. *Clin. Cancer Res.* **2017**, *23*, 3012–3024, doi:10.1158/1078-0432.CCR-16-2554.
- Toki, M.I.; Mani, N.; Smithy, J.W.; Liu, Y.; Altan, M.; Wasserman, B.; Tuktamyshov, R.; Schalper, K.; Syrigos, K.N.; Rimm, D.L.
 Immune Marker Profiling and Programmed Death Ligand 1 Expression Across NSCLC Mutations. *J. Thorac. Oncol.* 2018, 13, 1884–1896, doi:10.1016/j.jtho.2018.09.012.
- 61. Skoulidis, F.; Goldberg, M.E.; Greenawalt, D.M.; Hellmann, M.D.; Awad, M.M.; Gainor, J.F.; Schrock, A.B.; Hartmaier, R.J.; Trabucco, S.E.; Gay, L.; et al. STK11/LKB1 mutations and PD-1 inhibitor resistance in KRAS-mutant lung adenocarcinoma. *Cancer Discov.* 2018, 8, 822–835, doi:10.1158/2159-8290.CD-18-0099.
- 62. Rizvi, N.; Cho, B.C.; Reinmuth, N.; Lee, K.H.; Luft, A.; Ahn, M.; Papadimitrakopoulou, V.; Heymach, J.; Scheuring, U.; Higgs, B.; et al. OA04.07 Mutations Associated with Sensitivity or Resistance to Immunotherapy in mNSCLC: Analysis from the MYSTIC Trial. *J. Thorac. Oncol.* **2019**, *14*, S217, doi:10.1016/j.jtho.2019.08.428.
- 63. Okamura, R.; Kato, S.; Lee, S.; Jimenez, R.E.; Sicklick, J.K.; Kurzrock, R. ARID1A alterations function as a biomarker for longer progression-free survival after anti-PD-1/PD-L1 immunotherapy. *J. Immunother. Cancer* **2020**, *8*, doi:10.1136/jitc-2019-000438.
- 64. Liu, L.; Ruiz, J.; O'Neill, S.S.; Grant, S.C.; Petty, W.J.; Yang, M.; Chen, K.; Topaloglu, U.; Pasche, B.; Zhang, W. Favorable outcome of patients with lung adenocarcinoma harboring POLE mutations and expressing high PD-L1. *Mol. Cancer* 2018, *17*.
- 65. Chen, H.; Chong, W.; Teng, C.; Yao, Y.; Wang, X.; Li, X. The immune response-related mutational signatures and driver genes in non-small-cell lung cancer. *Cancer Sci.* **2019**, *110*, 2348–2356, doi:10.1111/cas.14113.
- 66. Brambilla, E.; Le Teuff, G.; Marguet, S.; Lantuejoul, S.; Dunant, A.; Graziano, S.; Pirker, R.; Douillard, J.Y.; Le Chevalier, T.; Filipits, M.; et al. Prognostic effect of tumor lymphocytic infiltration in resectable non-small-cell lung cancer. *J. Clin. Oncol.* **2016**, *34*, 1223–1230, doi:10.1200/JCO.2015.63.0970.
- Zeng, D.Q.; Yu, Y.F.; Ou, Q.Y.; Li, X.Y.; Zhong, R.Z.; Xie, C.M.; Hu, Q.G. Prognostic and predictive value of tumor-infiltrating lymphocytes for clinical therapeutic research in patients with non-small cell lung cancer. *Oncotarget* 2016, 7, 13765–13781, doi:10.18632/oncotarget.7282.
- 68. Lee, J.S.; Ruppin, E. Multiomics Prediction of Response Rates to Therapies to Inhibit Programmed Cell Death 1 and Programmed Cell Death 1 Ligand 1. *JAMA Oncol.* **2019**, *5*, 1614–1618, doi:10.1001/jamaoncol.2019.2311.
- 69. Fridman, W.H.; Zitvogel, L.; Sautès-Fridman, C.; Kroemer, G. The immune contexture in cancer prognosis and treatment. *Nat. Rev. Clin. Oncol.* 2017, *14*, 717–734.
- 70. Gajewski, T.F.; Schreiber, H.; Fu, Y.X. Innate and adaptive immune cells in the tumor microenvironment. *Nat. Immunol.* 2013, *14*, 1014–1022.
- 71. Hori, S.; Nomura, T.; Sakaguchi, S. Control of regulatory T cell development by the transcription factor Foxp3. *J. Immunol.* **2017**, *198*, 981–985, doi:10.1126/science.1079490.
- 72. Thommen, D.S.; Koelzer, V.H.; Herzig, P.; Roller, A.; Trefny, M.; Dimeloe, S.; Kiialainen, A.; Hanhart, J.; Schill, C.; Hess, C.; et al. A transcriptionally and functionally distinct pd-1 + cd8 + t cell pool with predictive potential in non-small-cell lung cancer treated with pd-1 blockade. *Nat. Med.* **2018**, *24*, doi:10.1038/s41591-018-0057-z.

- 73. Riaz, N.; Havel, J.J.; Makarov, V.; Desrichard, A.; Urba, W.J.; Sims, J.S.; Hodi, F.S.; Martín-Algarra, S.; Mandal, R.; Sharfman, W.H.; et al. Tumor and Microenvironment Evolution during Immunotherapy with Nivolumab. *Cell* **2017**, *171*, 934-949.e15, doi:10.1016/j.cell.2017.09.028.
- 74. Forde, P.M.; Chaft, J.E.; Smith, K.N.; Anagnostou, V.; Cottrell, T.R.; Hellmann, M.D.; Zahurak, M.; Yang, S.C.; Jones, D.R.; Broderick, S.; et al. Neoadjuvant PD-1 Blockade in Resectable Lung Cancer. *N. Engl. J. Med.* **2018**, *378*, 1976–1986, doi:10.1056/nejmoa1716078.
- 75. Roh, W.; Chen, P.L.; Reuben, A.; Spencer, C.N.; Prieto, P.A.; Miller, J.P.; Gopalakrishnan, V.; Wang, F.; Cooper, Z.A.; Reddy, S.M.; et al. Integrated molecular analysis of tumor biopsies on sequential CTLA-4 and PD-1 blockade reveals markers of response and resistance. *Sci. Transl. Med.* **2017**, *9*, doi:10.1126/scitranslmed.aah3560.
- 76. Snyder, A.; Nathanson, T.; Funt, S.A.; Ahuja, A.; Buros Novik, J.; Hellmann, M.D.; Chang, E.; Aksoy, B.A.; Al-Ahmadie, H.; Yusko, E.; et al. Contribution of systemic and somatic factors to clinical response and resistance to PD-L1 blockade in urothelial cancer: An exploratory multi-omic analysis. *PLoS Med.* **2017**, *14*, doi:10.1371/journal.pmed.1002309.
- 77. Hopkins, A.C.; Yarchoan, M.; Durham, J.N.; Yusko, E.C.; Rytlewski, J.A.; Robins, H.S.; Laheru, D.A.; Le, D.T.; Lutz, E.R.; Jaffee, E.M. T cell receptor repertoire features associated with survival in immunotherapy-treated pancreatic ductal adenocarcinoma. *JCI insight* 2018, 3, doi:10.1172/jci.insight.122092.
- 78. Bellone, M.; Calcinotto, A. Ways to enhance lymphocyte trafficking into tumors and fitness of tumor infiltrating lymphocytes. *Front. Oncol.* 2013, *3 SEP*.
- 79. Beatty, G.L.; Moon, E.K. Chimeric antigen receptor T cells are vulnerable to immunosuppressive mechanisms present within the tumor microenvironment. *Oncoimmunology* **2014**, *3*, e970027-1-e970027-3, doi:10.4161/21624011.2014.970027.
- 80. Campesato, L.F.; Merghoub, T. Antiangiogenic therapy and immune checkpoint blockade go hand in hand. Ann. Transl. Med. 2017, 5.
- 81. Fukumura, D.; Kloepper, J.; Amoozgar, Z.; Duda, D.G.; Jain, R.K. Enhancing cancer immunotherapy using antiangiogenics: Opportunities and challenges. *Nat. Rev. Clin. Oncol.* 2018, *15*, 325–340.
- 82. Socinski, M.A.; Jotte, R.M.; Cappuzzo, F.; Orlandi, F.; Stroyakovskiy, D.; Nogami, N.; Rodríguez-Abreu, D.; Moro-Sibilot, D.; Thomas, C.A.; Barlesi, F.; et al. Atezolizumab for First-Line Treatment of Metastatic Nonsquamous NSCLC. *N. Engl. J. Med.* **2018**, *378*, 2288–2301, doi:10.1056/nejmoa1716948.
- 83. Rahma, O.E.; Hodi, F.S. The intersection between tumor angiogenesis and immune suppression. *Clin. Cancer Res.* 2019, *25*, 5449–5457.
- 84. Voron, T.; Colussi, O.; Marcheteau, E.; Pernot, S.; Nizard, M.; Pointet, A.L.; Latreche, S.; Bergaya, S.; Benhamouda, N.; Tanchot, C.; et al. VEGF-A modulates expression of inhibitory checkpoints on CD8++ T cells in tumors. *J. Exp. Med.* 2015, 212, 139–148, doi:10.1084/jem.20140559.
- 85. Gabrilovich, D.; Ishida, T.; Oyama, T.; Ran, S.; Kravtsov, V.; Nadaf, S.; Carbone, D.P. Vascular endothelial growth factor inhibits the development of dendritic cells and dramatically affects the differentiation of multiple hematopoietic lineages in vivo. *Blood* **1998**, *92*, 4150–4166, doi:10.1182/blood.v92.11.4150.
- 86. Huang, Y.; Yuan, J.; Righi, E.; Kamoun, W.S.; Ancukiewicz, M.; Nezivar, J.; Santosuosso, M.; Martin, J.D.; Martin, M.R.; Vianello, F.; et al. Vascular normalizing doses of antiangiogenic treatment reprogram the immunosuppressive tumor microenvironment and enhance immunotherapy. *Proc. Natl. Acad. Sci. U. S. A.* 2012, *109*, 17561–17566, doi:10.1073/pnas.1215397109.
- 87. Wallin, J.J.; Bendell, J.C.; Funke, R.; Sznol, M.; Korski, K.; Jones, S.; Hernandez, G.; Mier, J.; He, X.; Hodi, F.S.; et al. Atezolizumab in combination with bevacizumab enhances antigen-specific T-cell migration in metastatic renal cell carcinoma. *Nat. Commun.* 2016, 7, doi:10.1038/ncomms12624.
- 88. Lanitis, E.; Dangaj, D.; Irving, M.; Coukos, G. Mechanisms regulating T-cell infiltration and activity in solid tumors. *Ann. Oncol.* 2017, 28, xii18–xii32.
- 89. Wu, X.; Giobbie-Hurder, A.; Liao, X.; Connelly, C.; Connolly, E.M.; Li, J.; Manos, M.P.; Lawrence, D.; McDermott, D.; Severgnini, M.; et al. Angiopoietin-2 as a biomarker and target for immune checkpoint therapy. *Cancer Immunol. Res.* **2017**, *5*, 17–28,

- doi:10.1158/2326-6066.CIR-16-0206.
- 90. Kamphorst, A.O.; Pillai, R.N.; Yang, S.; Nasti, T.H.; Akondy, R.S.; Wieland, A.; Sica, G.L.; Yu, K.; Koenig, L.; Patel, N.T.; et al. Proliferation of PD-1+ CD8 T cells in peripheral blood after PD-1-targeted therapy in lung cancer patients. *Proc. Natl. Acad. Sci. U. S. A.* 2017, 114, 4993–4998, doi:10.1073/pnas.1705327114.
- 91. Kim, K.H.; Cho, J.; Ku, B.M.; Koh, J.; Sun, J.M.; Lee, S.H.; Ahn, J.S.; Cheon, J.; Min, Y.J.; Park, S.H.; et al. The first-week proliferative response of peripheral blood PD-1bCD8b T cells predicts the response to Anti-PD-1 therapy in solid tumors. *Clin. Cancer Res.* 2019, 25, 2144–2154, doi:10.1158/1078-0432.CCR-18-1449.
- 92. Ferrara, R.; Naigeon, M.; Auclin, E.; Duchemann, B.; Cassard, L.; Jouniaux, J.M.; Boselli, L.; Grivel, J.; Desnoyer, A.; Mezquita, L.; et al. Circulating T-cell immunosenescence in patients with advanced non⊎small cell lung cancer treated with single-agent PD-1/PD-L1 inhibitors or platinum-based chemotherapy. *Clin. Cancer Res.* 2021, 27, 492–503, doi:10.1158/1078-0432.CCR-20-1420.
- 93. Carnio, S.; Mariniello, A.; Pizzutilo, P.; Numico, G.; Borra, G.; Lunghi, A.; Soto Parra, H.; Buosi, R.; Vavalà, T.; Stura, I.; et al. ROC Analysis Identifies Baseline and Dynamic NLR and dNLR Cut-Offs to Predict ICI Outcome in 402 Advanced NSCLC Patients. *J. Mol. Pathol.* 2020, 1, 19–31, doi:10.3390/jmp1010004.
- 94. Mezquita, L.; Auclin, E.; Ferrara, R.; Charrier, M.; Remon, J.; Planchard, D.; Ponce, S.; Ares, L.P.; Leroy, L.; Audigier-Valette, C.; et al. Association of the lung immune prognostic index with immune checkpoint inhibitor outcomes in patients with advanced non-small cell lung cancer. *JAMA Oncol.* **2018**, *4*, 351–357, doi:10.1001/jamaoncol.2017.4771.
- 95. Möller, M.; Turzer, S.; Schütte, W.; Seliger, B.; Riemann, D. Blood Immune Cell Biomarkers in Patient with Lung Cancer Undergoing Treatment with Checkpoint Blockade. *J. Immunother.* **2020**, *43*, 57–66, doi:10.1097/CJI.00000000000000297.
- 96. Krieg, C.; Nowicka, M.; Guglietta, S.; Schindler, S.; Hartmann, F.J.; Weber, L.M.; Dummer, R.; Robinson, M.D.; Levesque, M.P.; Becher, B. High-dimensional single-cell analysis predicts response to anti-PD-1 immunotherapy. *Nat. Med.* **2018**, *24*, 144–153, doi:10.1038/nm.4466.
- 97. Kumar, V.; Patel, S.; Tcyganov, E.; Gabrilovich, D.I. The Nature of Myeloid-Derived Suppressor Cells in the Tumor Microenvironment. *Trends Immunol.* 2016, *37*, 208–220.
- 98. Weber, R.; Fleming, V.; Hu, X.; Nagibin, V.; Groth, C.; Altevogt, P.; Utikal, J.; Umansky, V. Myeloid-derived suppressor cells hinder the anti-cancer activity of immune checkpoint inhibitors. *Front. Immunol.* 2018, *9*.
- 99. Weide, B.; Martens, A.; Zelba, H.; Stutz, C.; Derhovanessian, E.; Di Giacomo, A.M.; Maio, M.; Sucker, A.; Schilling, B.; Schadendorf, D.; et al. Myeloid-derived suppressor cells predict survival of patients with advanced melanoma: Comparison with regulatory T cells and NY-ESO-1- or melan-A-specific T cells. *Clin. Cancer Res.* **2014**, *20*, 1601–1609, doi:10.1158/1078-0432.CCR-13-2508.
- 100. Meyer, C.; Cagnon, L.; Costa-Nunes, C.M.; Baumgaertner, P.; Montandon, N.; Leyvraz, L.; Michielin, O.; Romano, E.; Speiser, D.E. Frequencies of circulating MDSC correlate with clinical outcome of melanoma patients treated with ipilimumab. *Cancer Immunol. Immunother.* 2014, 63, 247–257, doi:10.1007/s00262-013-1508-5.
- 101. Youn, J.I.; Park, S.M.; Park, S.; Kim, G.; Lee, H.J.; Son, J.; Hong, M.H.; Ghaderpour, A.; Baik, B.; Islam, J.; et al. Peripheral natural killer cells and myeloid-derived suppressor cells correlate with anti-PD-1 responses in non-small cell lung cancer. *Sci. Rep.* **2020**, *10*, doi:10.1038/s41598-020-65666-x.
- 102. Sivan, A.; Corrales, L.; Hubert, N.; Williams, J.B.; Aquino-Michaels, K.; Earley, Z.M.; Benyamin, F.W.; Lei, Y.M.; Jabri, B.; Alegre, M.L.; et al. Commensal Bifidobacterium promotes antitumor immunity and facilitates anti-PD-L1 efficacy. *Science* (80-.). 2015, 350, 1084–1089, doi:10.1126/science.aac4255.
- 103. Iida, N.; Dzutsev, A.; Stewart, C.A.; Smith, L.; Bouladoux, N.; Weingarten, R.A.; Molina, D.A.; Salcedo, R.; Back, T.; Cramer, S.; et al. Commensal bacteria control cancer response to therapy by modulating the tumor microenvironment. *Science* (80-.). 2013, 342, 967–970, doi:10.1126/science.1240527.
- 104. Chaput, N.; Lepage, P.; Coutzac, C.; Soularue, E.; Le Roux, K.; Monot, C.; Boselli, L.; Routier, E.; Cassard, L.; Collins, M.; et al. Corrigendum: Baseline gut microbiota predicts clinical response and colitis in metastatic melanoma patients treated with ipilimumab (Annals of Oncology (2017) 28:6 (1368-1379) DOI: 10.1093/annonc/mdx108). *Ann. Oncol.* 2019, *30*, 2012.

- 105. Matson, V.; Fessler, J.; Bao, R.; Chongsuwat, T.; Zha, Y.; Alegre, M.L.; Luke, J.J.; Gajewski, T.F. The commensal microbiome is associated with anti-PD-1 efficacy in metastatic melanoma patients. *Science* (80-.). 2018, 359, 104–108, doi:10.1126/science.aao3290.
- 106. Routy, B.; Le Chatelier, E.; Derosa, L.; Duong, C.P.M.; Alou, M.T.; Daillère, R.; Fluckiger, A.; Messaoudene, M.; Rauber, C.; Roberti, M.P.; et al. Gut microbiome influences efficacy of PD-1-based immunotherapy against epithelial tumors. *Science* (80-.). **2018**, 359, 91–97, doi:10.1126/science.aan3706.
- 107. Parham, P.; Ohta, T. Population biology of antigen presentation by MHC class I molecules. *Science (80-.).* **1996**, *272*, 67–74, doi:10.1126/science.272.5258.67.
- 108. Chowell, D.; Morris, L.G.T.; Grigg, C.M.; Weber, J.K.; Samstein, R.M.; Makarov, V.; Kuo, F.; Kendall, S.M.; Requena, D.; Riaz, N.; et al. Patient HLA class I genotype influences cancer response to checkpoint blockade immunotherapy. *Science* (80-.). 2018, 359, 582–587, doi:10.1126/science.aao4572.
- 109. Vargas, F.A.; Furness, A.J.S.; Litchfield, K.; Joshi, K.; Rosenthal, R.; Ghorani, E.; Solomon, I.; Lesko, M.H.; Ruef, N.; Roddie, C.; et al. Fc Effector Function Contributes to the Activity of Human Anti-CTLA-4 Antibodies. *Cancer Cell* 2018, 33, 649-663.e4, doi:10.1016/j.ccell.2018.02.010.
- 110. Aide, N.; Lasnon, C.; Kesner, A.; Levin, C.S.; Buvat, I.; Iagaru, A.; Hermann, K.; Badawi, R.D.; Cherry, S.R.; Bradley, K.M.; et al. New PET technologies embracing progress and pushing the limits. *Eur. J. Nucl. Med. Mol. Imaging* **2021**, doi:10.1007/s00259-021-05390-4.
- 111. Luke, J.J.; Flaherty, K.T.; Ribas, A.; Long, G. V. Targeted agents and immunotherapies: Optimizing outcomes in melanoma. *Nat. Rev. Clin. Oncol.* 2017, *14*, 463–482.
- 112. Grootjans, W.; De Geus-Oei, L.F.; Troost, E.G.C.; Visser, E.P.; Oyen, W.J.G.; Bussink, J. PET in the management of locally advanced and metastatic NSCLC. *Nat. Rev. Clin. Oncol.* 2015, *12*, 395–407.
- 113. Salaün, P.Y.; Abgral, R.; Malard, O.; Querellou-Lefranc, S.; Quere, G.; Wartski, M.; Coriat, R.; Hindie, E.; Taieb, D.; Tabarin, A.; et al. Good clinical practice recommendations for the use of PET/CT in oncology. *Eur. J. Nucl. Med. Mol. Imaging* **2020**, *47*, 28–50, doi:10.1007/s00259-019-04553-8.
- 114. Fuchs, S.; Grössmann, N.; Ferch, M.; Busse, R.; Wild, C. Evidence-based indications for the planning of PET or PET/CT capacities are needed. *Clin. Transl. Imaging* 2019, 7, 65–81.
- 115. Costa, L.B.; Queiroz, M.A.; Barbosa, F.G.; Nunes, R.F.; Zaniboni, E.C.; Ruiz, M.M.; Jardim, D.; Marin, J.F.G.; Cerri, G.G.; Buchpiguel, C.A. Reassessing patterns of response to immunotherapy with pet: From morphology to metabolism. *Radiographics* **2021**, *41*, 120–143, doi:10.1148/rg.2021200093.
- 116. Aide, N.; De Pontdeville, M.; Lopci, E. Evaluating response to immunotherapy with 18F-FDG PET/CT: where do we stand? *Eur. J. Nucl. Med. Mol. Imaging* 2020, 47, 1019–1021.
- 117. Rossi, G.; Bauckneht, M.; Genova, C.; Rijavec, E.; Biello, F.; Mennella, S.; Dal Bello, M.G.; Cittadini, G.; Bruzzi, P.; Piva, R.; et al. Comparison Between 18F-FDG PET-Based and CT-Based Criteria in Non-Small Cell Lung Cancer Patients Treated with Nivolumab. J. Nucl. Med. 2020, 61, 990–998, doi:10.2967/jnumed.119.233056.
- 118. Liberini, V.; Laudicella, R.; Capozza, M.; Huellner, M.W.; Burger, I.A.; Baldari, S.; Terreno, E.; Deandreis, D. The future of cancer diagnosis, treatment and surveillance: a systemic review on immunotherapy and immuno-pet radiotracers. *Molecules* 2021, 26.
- 119. Kandathil, A.; Kay, F.U.; Butt, Y.M.; Wachsmann, J.W.; Subramaniam, R.M. Role of FDG PET/CT in the eighth edition of TNM staging of non–Small cell lung cancer. *Radiographics* **2018**, *38*, 2134–2149, doi:10.1148/rg.2018180060.
- 120. Ettinger, D.S.; Wood, D.E.; Chair, V.; Aisner, D.L.; Akerley, W.; Bauman, J.R.; Bharat, A.; Lurie, R.H.; Bruno, D.S.; Chang, J.Y.; et al. Continue NCCN Guidelines Panel Disclosures NCCN Guidelines Version 4.2021 Non-Small Cell Lung Cancer; 2021;
- 121. Liberti, M. V.; Locasale, J.W. The Warburg Effect: How Does it Benefit Cancer Cells? *Trends Biochem. Sci.* 2016, 41, 211–218.
- 122. Kaira, K.; Kuji, I.; Kagamu, H. Value of 18F-FDG-PET to predict PD-L1 expression and outcomes of PD-1 inhibition therapy in human cancers. *Cancer Imaging* 2021, 21, 11.
- 123. Cho, S.Y.; Huff, D.T.; Jeraj, R.; Albertini, M.R. FDG PET/CT for Assessment of Immune Therapy: Opportunities and Understanding

- Pitfalls. Semin. Nucl. Med. 2020, 50, 518-531.
- 124. Iravani, A.; Hicks, R.J. Imaging the cancer immune environment and its response to pharmacologic intervention, Part 1: The role of 18F-FDG PET/CT. *J. Nucl. Med.* **2020**, *61*, 943–950, doi:10.2967/JNUMED.119.234278.
- 125. Beer, L.; Hochmair, M.; Haug, A.R.; Schwabel, B.; Kifjak, D.; Wadsak, W.; Fuereder, T.; Fabikan, H.; Fazekas, A.; Schwab, S.; et al. Comparison of RECIST, iRECIST, and PERCIST for the evaluation of response to PD-1/PD-L1 blockade therapy in patients with non-small cell lung cancer. *Clin. Nucl. Med.* **2019**, *44*, 535–543, doi:10.1097/RLU.0000000000002603.
- 126. Mulkey, F.; Theoret, M.R.; Keegan, P.; Pazdur, R.; Sridhara, R. Comparison of iRECIST versus RECIST V.1.1 in patients treated with an anti-PD-1 or PD-L1 antibody: Pooled FDA analysis. *J. Immunother. Cancer* **2020**, *8*, doi:10.1136/jitc-2019-000146.
- 127. Tazdait, M.; Mezquita, L.; Lahmar, J.; Ferrara, R.; Bidault, F.; Ammari, S.; Balleyguier, C.; Planchard, D.; Gazzah, A.; Soria, J.C.; et al. Patterns of responses in metastatic NSCLC during PD-1 or PDL-1 inhibitor therapy: Comparison of RECIST 1.1, irRECIST and iRECIST criteria. *Eur. J. Cancer* 2018, 88, 38–47, doi:10.1016/j.ejca.2017.10.017.
- 128. Boellaard, R.; Delgado-Bolton, R.; Oyen, W.J.G.; Giammarile, F.; Tatsch, K.; Eschner, W.; Verzijlbergen, F.J.; Barrington, S.F.; Pike, L.C.; Weber, W.A.; et al. FDG PET/CT: EANM procedure guidelines for tumour imaging: version 2.0. *Eur. J. Nucl. Med. Mol. Imaging* 2015, 42, 328–354.
- 129. Stephen Hodi, F.; Ballinger, M.; Lyons, B.; Soria, J.C.; Nishino, M.; Tabernero, J.; Powles, T.; Smith, D.; Hoos, A.; McKenna, C.; et al. Immune-modified response evaluation criteria in solid tumors (imrecist): Refining guidelines to assess the clinical benefit of cancer immunotherapy. *J. Clin. Oncol.* 2018, 36, 850–858, doi:10.1200/JCO.2017.75.1644.
- 130. Ito, K.; Teng, R.; Schöder, H.; Humm, J.L.; Ni, A.; Michaud, L.; Nakajima, R.; Yamashita, R.; Wolchok, J.D.; Weber, W.A. 18 F-FDG PET/CT for monitoring of ipilimumab therapy in patients with metastatic melanoma. *J. Nucl. Med.* 2019, 60, 335–341, doi:10.2967/jnumed.118.213652.
- 131. Anwar, H.; Sachpekidis, C.; Winkler, J.; Kopp-Schneider, A.; Haberkorn, U.; Hassel, J.C.; Dimitrakopoulou-Strauss, A. Absolute number of new lesions on 18F-FDG PET/CT is more predictive of clinical response than SUV changes in metastatic melanoma patients receiving ipilimumab. *Eur. J. Nucl. Med. Mol. Imaging* **2018**, *45*, 376–383, doi:10.1007/s00259-017-3870-6.
- 132. Wolchok, J.D.; Hoos, A.; O'Day, S.; Weber, J.S.; Hamid, O.; Lebbé, C.; Maio, M.; Binder, M.; Bohnsack, O.; Nichol, G.; et al. Guidelines for the evaluation of immune therapy activity in solid tumors: Immune-related response criteria. *Clin. Cancer Res.* **2009**, *15*, 7412–7420, doi:10.1158/1078-0432.CCR-09-1624.
- Therasse, P.; Arbuck, S.G.; Eisenhauer, E.A.; Wanders, J.; Kaplan, R.S.; Rubinstein, L.; Verweij, J.; Van Glabbeke, M.; Van Oosterom, A.T.; Christian, M.C.; et al. New guidelines to evaluate the response to treatment in solid tumors. *J. Natl. Cancer Inst.* **2000**, *92*, 205–216, doi:10.1093/jnci/92.3.205.
- 134. Wahl, R.L.; Jacene, H.; Kasamon, Y.; Lodge, M.A. From RECIST to PERCIST: Evolving considerations for PET response criteria in solid tumors. *J. Nucl. Med.* 2009, *50*, 122S.
- 135. Park, H.J.; Kim, K.W.; Pyo, J.; Suh, C.H.; Yoon, S.; Hatabu, H.; Nishino, M. Incidence of pseudoprogression during immune checkpoint inhibitor therapy for solid tumors: A systematic review and meta-Analysis. *Radiology* 2020, 297, 87–96.
- 136. Humbert, O.; Cadour, N.; Paquet, M.; Schiappa, R.; Poudenx, M.; Chardin, D.; Borchiellini, D.; Benisvy, D.; Ouvrier, M.J.; Zwarthoed, C.; et al. 18FDG PET/CT in the early assessment of non-small cell lung cancer response to immunotherapy: frequency and clinical significance of atypical evolutive patterns. *Eur. J. Nucl. Med. Mol. Imaging* **2020**, *47*, 1158–1167, doi:10.1007/s00259-019-04573-4.
- 137. Champiat, S.; Dercle, L.; Ammari, S.; Massard, C.; Hollebecque, A.; Postel-Vinay, S.; Chaput, N.; Eggermont, A.; Marabelle, A.; Soria, J.C.; et al. Hyperprogressive disease is a new pattern of progression in cancer patients treated by anti-PD-1/PD-L1. *Clin. Cancer Res.* 2017, 23, 1920–1928, doi:10.1158/1078-0432.CCR-16-1741.
- 138. Castello, A.; Rossi, S.; Mazziotti, E.; Toschi, L.; Lopci, E. Hyperprogressive Disease in Patients with Non-Small Cell Lung Cancer Treated with Checkpoint Inhibitors: The Role of 18F-FDG PET/CT. *J. Nucl. Med.* **2020**, *61*, 821–826, doi:10.2967/jnumed.119.237768.
- 139. Donegani, M.I.; Ferrarazzo, G.; Marra, S.; Miceli, A.; Raffa, S.; Bauckneht, M.; Morbelli, S. Positron emission tomography-based response to target and immunotherapies in oncology. *Med.* 2020, *56*, 1–22.

- 140. Aide, N.; Hicks, R.J.; Le Tourneau, C.; Lheureux, S.; Fanti, S.; Lopci, E. FDG PET/CT for assessing tumour response to immunotherapy: Report on the EANM symposium on immune modulation and recent review of the literature. *Eur. J. Nucl. Med. Mol. Imaging* 2019, 46, 238–250.
- 141. Eze, C.; Schmidt-Hegemann, N.S.; Sawicki, L.M.; Kirchner, J.; Roengvoraphoj, O.; Käsmann, L.; Mittlmeier, L.M.; Kunz, W.G.; Tufman, A.; Dinkel, J.; et al. PET/CT imaging for evaluation of multimodal treatment efficacy and toxicity in advanced NSCLC—current state and future directions. *Eur. J. Nucl. Med. Mol. Imaging* 2021.
- Eshghi, N.; Garland, L.L.; Nia, E.; Betancourt, R.; Krupinski, E.; Kuo, P.H. 18F-FDG PET/CT can predict development of thyroiditis due to immunotherapy for lung cancer. *J. Nucl. Med. Technol.* **2018**, *46*, 260–264, doi:10.2967/jnmt.117.204933.
- 143. Rizwan, S.; Alhamad, K.; Abel, S.; Bakalov, V.; Rodriguez, R.R.; Sethi, A.; Khan, T.; Attah, A.A.; Yasir, M.; Shankar, K.; et al. Impact of immune-related adverse events on survival in patients with advanced non-small cell lung cancer treated with immune checkpoint inhibitors: A real-world perspective. *J. Clin. Oncol.* 2021, 39, e21213–e21213, doi:10.1200/jco.2021.39.15_suppl.e21213.
- Prigent, K.; Aide, N. 18F-Fludeoxyglucose PET/Computed Tomography for Assessing Tumor Response to Immunotherapy and Detecting Immune-Related Side Effects: A Checklist for the PET Reader. *PET Clin.* 2020, *15*, 1–10.
- 145. Nobashi, T.; Baratto, L.; Reddy, S.A.; Srinivas, S.; Toriihara, A.; Hatami, N.; Yohannan, T.K.; Mittra, E. Predicting Response to Immunotherapy by Evaluating Tumors, Lymphoid Cell-Rich Organs, and Immune-Related Adverse Events Using FDG-PET/CT. *Clin. Nucl. Med.* 2019, 44, e272–e279, doi:10.1097/RLU.000000000000002453.
- Lambin, P.; Leijenaar, R.T.H.; Deist, T.M.; Peerlings, J.; De Jong, E.E.C.; Van Timmeren, J.; Sanduleanu, S.; Larue, R.T.H.M.; Even, A.J.G.; Jochems, A.; et al. Radiomics: The bridge between medical imaging and personalized medicine Available online: https://www.ncbi.nlm.nih.gov/pubmed/?term=Lambin++Radiomics%3A+the+bridge+between+medical+imaging+and+personalized+m edicine (accessed on Feb 7, 2020).
- 147. Zwanenburg, A.; Leger, S.; Vallières, M.; Löck, S. Image biomarker standardisation initiative. 2016.
- 148. Sanduleanu, S.; Woodruff, H.C.; de Jong, E.E.C.; van Timmeren, J.E.; Jochems, A.; Dubois, L.; Lambin, P. Tracking tumor biology with radiomics: A systematic review utilizing a radiomics quality score. *Radiother. Oncol.* 2018, *127*, 349–360.
- 149. Mayerhoefer, M.E.; Materka, A.; Langs, G.; Häggström, I.; Szczypiński, P.; Gibbs, P.; Cook, G. Introduction to Radiomics. *J. Nucl. Med.* 2020, *61*, 488–495.
- 150. Gatta, R.; Depeursinge, A.; Ratib, O.; Michielin, O.; Leimgruber, A. Integrating radiomics into holomics for personalised oncology: from algorithms to bedside. *Eur. Radiol. Exp.* 2020, *4*.
- 151. Froelich, J.W.; Salavati, A. Artificial intelligence in PET/CT is about to make whole-body tumor burden measurements a clinical reality. *Radiology* 2020, *294*, 453–454.
- 152. Laudicella, R.; Comelli, A.; Stefano, A.; Szostek, M.; Crocè, L.; Vento, A.; Spataro, A.; Comis, A.D.; La Torre, F.; Gaeta, M.; et al. Artificial Neural Networks in Cardiovascular Diseases and its Potential for Clinical Application in Molecular Imaging. *Curr. Radiopharm.* **2020**, *13*, doi:10.2174/1874471013666200621191259.
- 153. Piñeiro-Fiel, M.; Moscoso, A.; Pubul, V.; Ruibal, Á.; Silva-Rodríguez, J.; Aguiar, P. A Systematic Review of PET Textural Analysis and Radiomics in Cancer. *Diagnostics (Basel, Switzerland)* **2021**, *11*, 380, doi:10.3390/diagnostics11020380.
- 154. Yoon, H.J.; Sohn, I.; Cho, J.H.; Lee, H.Y.; Kim, J.H.; Choi, Y. La; Kim, H.; Lee, G.; Lee, K.S.; Kim, J. Decoding tumor phenotypes for ALK, ROS1, and RET fusions in lung adenocarcinoma using a radiomics approach. *Med. (United States)* **2015**, *94*, doi:10.1097/MD.000000000001753.
- 155. Yip, S.S.F.; Kim, J.; Coroller, T.P.; Parmar, C.; Velazquez, E.R.; Huynh, E.; Mak, R.H.; Aerts, H.J.W.L. Associations between somatic mutations and metabolic imaging phenotypes in non-small cell lung cancer. *J. Nucl. Med.* **2017**, *58*, 569–576, doi:10.2967/jnumed.116.181826.
- Li, X.; Yin, G.; Zhang, Y.; Dai, D.; Liu, J.; Chen, P.; Zhu, L.; Ma, W.; Xu, W. Predictive Power of a Radiomic Signature Based on 18F-FDG PET/CT Images for EGFR Mutational Status in NSCLC. *Front. Oncol.* **2019**, *9*, doi:10.3389/fonc.2019.01062.
- 157. Jiang, M.; Zhang, Y.; Xu, J.; Ji, M.; Guo, Y.Y.; Guo, Y.Y.; Xiao, J.; Yao, X.; Shi, H.; Zeng, M. Assessing EGFR gene mutation status

- in non-small cell lung cancer with imaging features from PET/CT. *Nucl. Med. Commun.* **2019**, *40*, 842–849, doi:10.1097/MNM.000000000001043.
- Zhang, J.J.; Zhao, X.; Zhao, Y.; Zhang, J.J.; Zhang, Z.; Wang, J.; Wang, Y.; Dai, M.; Han, J. Value of pre-therapy 18F-FDG PET/CT radiomics in predicting EGFR mutation status in patients with non-small cell lung cancer. Eur. J. Nucl. Med. Mol. Imaging 2020, 47, 1137–1146, doi:10.1007/s00259-019-04592-1.
- 159. Koyasu, S.; Nishio, M.; Isoda, H.; Nakamoto, Y.; Togashi, K. Usefulness of gradient tree boosting for predicting histological subtype and EGFR mutation status of non-small cell lung cancer on 18F FDG-PET/CT. *Ann. Nucl. Med.* **2020**, *34*, 49–57, doi:10.1007/s12149-019-01414-0.
- 160. Liu, Q.; Sun, D.; Li, N.; Kim, J.; Feng, D.; Huang, G.; Wang, L.; Song, S. Predicting EGFR mutation subtypes in lung adenocarcinoma using 18F-FDG PET/CT radiomic features. *Transl. Lung Cancer Res.* **2020**, *9*, 549–562, doi:10.21037/tlcr.2020.04.17.
- 161. Yang, B.; Ji, H.S.; Zhou, C.S.; Dong, H.; Ma, L.; Ge, Y.Q.; Zhu, C.H.; Tian, J.H.; Zhang, L.J.; Zhu, H.; et al. 18F-fluorodeoxyglucose positron emission tomography/computed tomography-based radiomic features for prediction of epidermal growth factor receptor mutation status and prognosis in patients with lung adenocarcinoma. *Transl. Lung Cancer Res.* 2020, *9*, 563–574, doi:10.21037/tlcr-19-592.
- 162. Jiang, M.; Sun, D.; Guo, Y.; Guo, Y.; Xiao, J.; Wang, L.; Yao, X. Assessing PD-L1 Expression Level by Radiomic Features From PET/CT in Nonsmall Cell Lung Cancer Patients: An Initial Result. *Acad. Radiol.* 2020, 27, 171–179, doi:10.1016/j.acra.2019.04.016.
- Lopci, E.; Toschi, L.; Grizzi, F.; Rahal, D.; Olivari, L.; Castino, G.F.; Marchetti, S.; Cortese, N.; Qehajaj, D.; Pistillo, D.; et al. Correlation of metabolic information on FDG-PET with tissue expression of immune markers in patients with non-small cell lung cancer (NSCLC) who are candidates for upfront surgery. Eur. J. Nucl. Med. Mol. Imaging 2016, 43, 1954–1961, doi:10.1007/s00259-016-3425-2.
- Takada, K.; Toyokawa, G.; Yoneshima, Y.; Tanaka, K.; Okamoto, I.; Shimokawa, M.; Wakasu, S.; Haro, A.; Osoegawa, A.; Tagawa, T.; et al. 18F-FDG uptake in PET/CT is a potential predictive biomarker of response to anti-PD-1 antibody therapy in non-small cell lung cancer. *Sci. Rep.* **2019**, *9*, doi:10.1038/s41598-019-50079-2.
- 165. Grizzi, F.; Castello, A.; Lopci, E. Is it time to change our vision of tumor metabolism prior to immunotherapy? *Eur. J. Nucl. Med. Mol. Imaging* 2018, 45, 1072–1075.
- 166. Monaco, L.; Gemelli, M.; Gotuzzo, I.; Bauckneht, M.; Crivellaro, C.; Genova, C.; Cortinovis, D.; Zullo, L.; Ammoni, L.C.; Bernasconi, D.P.; et al. Metabolic parameters as biomarkers of response to immunotherapy and prognosis in non-small cell lung cancer (Nsclc): A real world experience. *Cancers (Basel)*. **2021**, *13*, doi:10.3390/cancers13071634.
- 167. Quartuccio, N.; Evangelista, L.; Alongi, P.; Caobelli, F.; Altini, C.; Cistaro, A.; Lambertini, A.; Schiorlin, I.; Popescu, C.E.; Linguanti, F.; et al. Prognostic and diagnostic value of [18F]FDG-PET/CT in restaging patients with small cell lung carcinoma: an Italian multicenter study. *Nucl. Med. Commun.* 2019, 40, 808–814, doi:10.1097/MNM.000000000001038.
- 168. Lee, P.; Weerasuriya, D.K.; Lavori, P.W.; Quon, A.; Hara, W.; Maxim, P.G.; Le, Q.T.; Wakelee, H.A.; Donington, J.S.; Graves, E.E.; et al. {A figure is presented} Metabolic Tumor Burden Predicts for Disease Progression and Death in Lung Cancer. *Int. J. Radiat. Oncol. Biol. Phys.* 2007, 69, 328–333, doi:10.1016/j.ijrobp.2007.04.036.
- 169. Lee, P.; Bazan, J.G.; Lavori, P.W.; Weerasuriya, D.K.; Quon, A.; Le, Q.T.; Wakelee, H.A.; Graves, E.E.; Loo, B.W. Metabolic tumor volume is an independent prognostic factor in patients treated definitively for nonsmall-cell lung cancer. *Clin. Lung Cancer* **2012**, *13*, 52–58, doi:10.1016/j.cllc.2011.05.001.
- 170. Polverari, G.; Ceci, F.; Bertaglia, V.; Reale, M.L.; Rampado, O.; Gallio, E.; Passera, R.; Liberini, V.; Scapoli, P.; Arena, V.; et al. 18F-FDG pet parameters and radiomics features analysis in advanced nsclc treated with immunotherapy as predictors of therapy response and survival. *Cancers (Basel).* 2020, 12, doi:10.3390/cancers12051163.
- 171. Kim, B.S.; Kang, J.; Jun, S.; Kim, H.; Pak, K.; Kim, G.H.; Heo, B.J.; Kim, Y.H. Association between immunotherapy biomarkers and glucose metabolism from F-18 FDG PET. *Eur. Rev. Med. Pharmacol. Sci.* **2020**, *24*, 8288–8295, doi:10.26355/eurrev 202008 22625.
- 172. Valentinuzzi, D.; Vrankar, M.; Boc, N.; Ahac, V.; Zupancic, Z.; Unk, M.; Skalic, K.; Zagar, I.; Studen, A.; Simoncic, U.; et al. FDG

- PET immunotherapy radiomics signature (iRADIOMICS) predicts response of non-small-cell lung cancer patients treated with pembrolizumab. *Radiol. Oncol.* **2020**, *54*, 285–294, doi:10.2478/raon-2020-0042.
- 173. Seban, R.D.; Mezquita, L.; Berenbaum, A.; Dercle, L.; Botticella, A.; Le Pechoux, C.; Caramella, C.; Deutsch, E.; Grimaldi, S.; Adam, J.; et al. Baseline metabolic tumor burden on FDG PET/CT scans predicts outcome in advanced NSCLC patients treated with immune checkpoint inhibitors. *Eur. J. Nucl. Med. Mol. Imaging* **2020**, *47*, 1147–1157, doi:10.1007/s00259-019-04615-x.
- 174. Bauckneht, M.; Genova, C.; Rossi, G.; Rijavec, E.; Bello, M.G.D.; Ferrarazzo, G.; Tagliamento, M.; Donegani, M.I.; Biello, F.; Chiola, S.; et al. The role of the immune metabolic prognostic index in patients with non-small cell lung cancer (Nsclc) in radiological progression during treatment with nivolumab. *Cancers (Basel)*. 2021, 13, doi:10.3390/cancers13133117.
- 175. Ziv, E.; Durack, J.C.; Solomon, S.B. The Importance of Biopsy in the Era of Molecular Medicine. *Cancer J. (United States)* 2016, 22, 418–422.
- 176. McQuerry, J.A.; Chang, J.T.; Bowtell, D.D.L.; Cohen, A.; Bild, A.H. Mechanisms and clinical implications of tumor heterogeneity and convergence on recurrent phenotypes. *J. Mol. Med.* 2017, *95*, 1167–1178.
- 177. Kather, J.N.; Suarez-Carmona, M.; Charoentong, P.; Weis, C.A.; Hirsch, D.; Bankhead, P.; Horning, M.; Ferber, D.; Kel, I.; Herpel, E.; et al. Topography of cancer-associated immune cells in human solid tumors. *Elife* **2018**, 7, doi:10.7554/eLife.36967.
- 178. Pietrobon, V.; Cesano, A.; Marincola, F.; Kather, J.N. Next Generation Imaging Techniques to Define Immune Topographies in Solid Tumors. *Front. Immunol.* **2021**, *11*, 3519, doi:10.3389/fimmu.2020.604967.
- 179. Abousaway, O.; Rakhshandehroo, T.; Van den Abbeele, A.D.; Kircher, M.F.; Rashidian, M. Noninvasive imaging of cancer immunotherapy. *Nanotheranostics* **2021**, *5*, 90–112, doi:10.7150/ntno.50860.
- 180. McCarthy, C.E.; White, J.M.; Viola, N.T.; Gibson, H.M. In vivo Imaging Technologies to Monitor the Immune System. *Front. Immunol.* 2020, *11*, 1067.
- 181. Natarajan, A.; Mayer, A.T.; Xu, L.; Reeves, R.E.; Gano, J.; Gambhir, S.S. Novel Radiotracer for ImmunoPET Imaging of PD-1 Checkpoint Expression on Tumor Infiltrating Lymphocytes. *Bioconjug. Chem.* 2015, 26, 2062–2069, doi:10.1021/acs.bioconjchem.5b00318.
- 182. Bensch, F.; van der Veen, E.L.; Lub-de Hooge, M.N.; Jorritsma-Smit, A.; Boellaard, R.; Kok, I.C.; Oosting, S.F.; Schröder, C.P.; Hiltermann, T.J.N.; van der Wekken, A.J.; et al. 89Zr-atezolizumab imaging as a non-invasive approach to assess clinical response to PD-L1 blockade in cancer. *Nat. Med.* 2018, 24, 1852–1858, doi:10.1038/s41591-018-0255-8.
- Niemeijer, A.N.; Leung, D.; Huisman, M.C.; Bahce, I.; Hoekstra, O.S.; van Dongen, G.A.M.S.; Boellaard, R.; Du, S.; Hayes, W.; Smith, R.; et al. Whole body PD-1 and PD-L1 positron emission tomography in patients with non-small-cell lung cancer. *Nat. Commun.* **2018**, *9*, 1–5, doi:10.1038/s41467-018-07131-y.
- Pandit-Taskar, N.; Postow, M.A.; Hellmann, M.D.; Harding, J.J.; Barker, C.A.; O'Donoghue, J.A.; Ziolkowska, M.; Ruan, S.; Lyashchenko, S.K.; Tsai, F.; et al. First-in-Humans Imaging with 89Zr-Df-IAB22M2C Anti-CD8 Minibody in Patients with Solid Malignancies: Preliminary Pharmacokinetics, Biodistribution, and Lesion Targeting. *J. Nucl. Med.* **2020**, *61*, 512–519, doi:10.2967/jnumed.119.229781.
- 185. Barrett, R.L.; Puré, E. Cancer-associated fibroblasts and their influence on tumor immunity and immunotherapy. *Elife* **2020**, *9*, 1–20, doi:10.7554/ELIFE.57243.
- 186. BH, S.; PM, C. Fibroblasts Influence the Efficacy, Resistance, and Future Use of Vaccines and Immunotherapy in Cancer Treatment. *Vaccines* **2021**, *9*, doi:10.3390/VACCINES9060634.
- 187. M, K.; PJ, B.; JN, A.; EW, R.; L, M.; JO, J.; A, G.; DA, T.; DT, F. Suppression of antitumor immunity by stromal cells expressing fibroblast activation protein-alpha. *Science* **2010**, *330*, 827–830, doi:10.1126/SCIENCE.1195300.
- 188. A, L.; T, L.; W, M.; J, D.; A, A.; D, J.; F, G.; C, K.; P, B.; C, R.; et al. A Tumor-Imaging Method Targeting Cancer-Associated Fibroblasts. *J. Nucl. Med.* **2018**, *59*, 1423–1429, doi:10.2967/JNUMED.118.210435.
- 189. C, K.; P, F.; T, L.; L, A.; A, A.; W, M.; S, A.; H, R.; M, R.; H, W.; et al. 68 Ga-FAPI PET/CT: Tracer Uptake in 28 Different Kinds of Cancer. J. Nucl. Med. 2019, 60, 801–805, doi:10.2967/JNUMED.119.227967.

- 190. SNMMI Image of the Year: Novel Radiotracer Detects 28 Cancer Types, Paving the Way for Development of New Therapies SNMMI Available online: https://www.snmmi.org/NewsPublications/NewsDetail.aspx?ItemNumber=32020 (accessed on Aug 15, 2021).
- 191. Calais, J.; Mona, C.E. Will FAPI PET/CT Replace FDG PET/CT in the Next Decade? Point—An Important Diagnostic, Phenotypic, and Biomarker Role. https://doi.org/10.2214/AJR.20.24302 2020, 216, 305–306, doi:10.2214/AJR.20.24302.
- 192. H, C.; Y, P.; J, W.; L, Z.; B, H.; J, W.; J, W.; S, W.; L, Z.; Z, L.; et al. Comparison of [68 Ga]Ga-DOTA-FAPI-04 and [18 F] FDG PET/CT for the diagnosis of primary and metastatic lesions in patients with various types of cancer. *Eur. J. Nucl. Med. Mol. Imaging* 2020, 47, 1820–1832, doi:10.1007/S00259-020-04769-Z.
- 193. T, W.; Y, L.; K, K.-N.; Y, S.; T, L.; K, O.; A, T.; K, N.; E, S.; U, H.; et al. Theranostics Targeting Fibroblast Activation Protein in the Tumor Stroma: 64 Cu- and 225 Ac-Labeled FAPI-04 in Pancreatic Cancer Xenograft Mouse Models. *J. Nucl. Med.* **2020**, *61*, 563–569, doi:10.2967/JNUMED.119.233122.
- 194. Kuyumcu, S.; Kovan, B.; Sanli, Y.; Buyukkaya, F.; Has Simsek, D.; Özkan, Z.G.; Isik, E.G.; Ekenel, M.; Turkmen, C. Safety of Fibroblast Activation Protein—Targeted Radionuclide Therapy by a Low-Dose Dosimetric Approach Using 177Lu-FAPI04. *Clin. Nucl. Med.* 2021, 46, 641–646, doi:10.1097/RLU.0000000000003667.
- 195. Eryilmaz, K.; Kilbas, B. Fully-automated synthesis of 177Lu labelled FAPI derivatives on the module modular lab-Eazy. *EJNMMI Radiopharm. Chem.* **2021**, *6*, doi:10.1186/S41181-021-00130-3.
- 196. Moradi, F.; Iagaru, A. Will FAPI PET/CT Replace FDG PET/CT in the Next Decade? Counterpoint—No, Not So Fast! https://doi.org/10.2214/AJR.20.23794 2020, 216, 307–308, doi:10.2214/AJR.20.23794.
- 197. M, S.; M, K.; F, G.; F, F.; N, G.; A, C. State-of-the-art of FAPI-PET imaging: a systematic review and meta-analysis. *Eur. J. Nucl. Med. Mol. Imaging* **2021**, doi:10.1007/S00259-021-05475-0.
- 198. Popescu, B.A.; Ameri, P.; Lehmann, L.H.; Finke, D.; Heckmann, M.B.; Herpel, E.; Katus, H.A.; Haberkorn, U.; Leuschner, F. Early Detection of Checkpoint Inhibitor-Associated Myocarditis Using 68 Ga-FAPI PET/CT. Front. Cardiovasc. Med. | www.frontiersin.org 2021, 8, 614997, doi:10.3389/fcvm.2021.614997.
- 199. EJ, H.; FM, K.; S, T.; O, S.; MD, G. Understanding fibroblast activation protein (FAP): substrates, activities, expression and targeting for cancer therapy. *Proteomics. Clin. Appl.* **2014**, *8*, 454–463, doi:10.1002/PRCA.201300095.
- 200. Theodoropoulos, A.S.; Gkiozos, I.; Kontopyrgias, G.; Charpidou, A.; Kotteas, E.; Kyrgias, G.; Tolia, M. Modern radiopharmaceuticals for lung cancer imaging with positron emission tomography/computed tomography scan: A systematic review. SAGE Open Med. 2020, 8, 205031212096159, doi:10.1177/2050312120961594.
- 201. Padhani, A. PET imaging of tumour hypoxia. Cancer Imaging 2006, 6, S117.
- 202. Pietrobon, V.; Marincola, F.M. Hypoxia and the phenomenon of immune exclusion. J. Transl. Med. 2021, 19.
- 203. Rühle, A.; Grosu, A.L.; Wiedenmann, N.; Mix, M.; Stoian, R.; Niedermann, G.; Baltas, D.; Werner, M.; Weber, W.A.; Kayser, G.; et al. Hypoxia dynamics on FMISO-PET in combination with PD-1/PD-L1 expression has an impact on the clinical outcome of patients with Head-and-neck Squamous Cell Carcinoma undergoing Chemoradiation. *Theranostics* **2020**, *10*, 9395–9406, doi:10.7150/thno.48392.
- 204. Masłowska, K.; Halik, P.K.; Tymecka, D.; Misicka, A.; Gniazdowska, E. The role of vegf receptors as molecular target in nuclear medicine for cancer diagnosis and combination therapy. *Cancers (Basel)*. 2021, *13*, 1–46.
- 205. Bahce, I.; Yaqub, M.; Smit, E.F.; Lammertsma, A.A.; van Dongen, G.A.M.S.; Hendrikse, N.H. Personalizing NSCLC therapy by characterizing tumors using TKI-PET and immuno-PET. *Lung Cancer* **2017**, *107*, 1–13, doi:10.1016/j.lungcan.2016.05.025.
- 206. Bahce, I.; Huisman, M.C.; Verwer, E.E.; Ooijevaar, R.; Boutkourt, F.; Vugts, D.J.; van Dongen, G.A.M.S.; Boellaard, R.; Smit, E.F. Pilot study of 89Zr-bevacizumab positron emission tomography in patients with advanced non-small cell lung cancer. *EJNMMI Res.* **2014**, *4*, 1–7, doi:10.1186/s13550-014-0035-5.