Remiero

Thyroid Hormone Receptor Beta as Tumor Suppressor: Untapped Potential in Treatment and Diagnostics in Solid Tumors

Cole D. Davidson 1,2, Noelle E. Gillis 1,2, and Frances E. Carr 1,2*

- Department of Pharmacology, Larner College of Medicine, University of Vermont Burlington VT 05405 USA
- ² University of Vermont Cancer Center, Burlington VT 05401 USA cole.d.davidson@uvm.edu negillis@uvm.edu
- * Correspondence: frances.carr@med.uvm.edu Tel.: 802-656-1318

Simple Summary: Dysregulation of the thyroid hormone receptor beta (TR β) is characteristic of many solid and endocrine-related tumors. Despite a recognized role as a tumor suppressor, the mechanisms by which TR β regulates tumor growth are not yet clear. As a transcription factor that responds to changes in thyroid hormone levels, TR β plays a key role in regulating many cell signalling nodes that are important for maintenance of normal cell identity and tumor progression. This review will address the need for a deeper understanding of TR β tumor suppressor mechanisms to inform the development of more effective thyroid cancer diagnostics and therapies.

Abstract: There is compelling evidence that the nuclear receptor $TR\beta$, a member of the thyroid hormone receptor (TR) family, is a tumor suppressor in thyroid, breast and other solid tumors. Cell-based and animal studies reveal that the liganded $TR\beta$ induces apoptosis, reduces an aggressive phenotype, decreases stem cell populations, and slows tumor growth through modulation of a complex interplay of transcriptional networks. $TR\beta$ -driven tumor suppressive transcriptomic signatures include repression of known drivers of proliferation such as PI3K/Akt pathway and activation of novel signaling (JAK1/STAT1) and metabolic reprogramming in both thyroid and breast cancers. The presence of $TR\beta$ is also correlated with a positive prognosis and response to therapeutics in BRCA+ and triple-negative breast cancers respectively. Ligand activation of $TR\beta$ enhances sensitivity to chemotherapeutics. $TR\beta$ co-regulators and bromodomain-containing chromatin remodeling proteins are emergent therapeutic targets. This review considers $TR\beta$ as a potential biomolecular diagnostic and therapeutic target.

Keywords: TR β , tumor suppression, co-regulators, therapeutics

1. Introduction

Altered gene expression programming in cancer cells is often a consequence of a loss of function of cell-type specific transcriptional control mechanisms. Genetic mutations and epigenetic silencing of thyroid hormone receptor beta ($TR\beta$) is characteristic of a number of solid tumors, and can be a marker for dedifferentiation [1-5]. $TR\beta$ is a ligand-dependent transcription factor that responds primarily to triiodothyronine (T_3). $TR\beta$ is recognized as a tumor suppressor and a positive prognostic indicator, however the mechanisms by which it regulates tumor growth remain unclear[6-9]. Recent studies indicate that $TR\beta$ tumor suppressive effects are mediated in part through intracellular signaling pathways including PI3K/Akt, Ras/MAPK, and JAK-STAT pathways, and induction of the mesenchymal-to-epithelial transition. Mutations in $TR\beta$ that lead to thyroid hormone resistance have also been shown to be oncogenic [10]. Transcriptional regulation by $TR\beta$ is critical for its function as a tumor suppressor because it acts as both a signal transducer and facilitator of long-term epigentic



programming for maintenance of cell identity. The purpose of this review is to discuss recent advances in our understanding of $TR\beta$ tumor suppression, and highlight its potential utility as a diagnostic indicator and therapeutic target.

2.1 TRβ Satisfies the Criteria for a Tumor Suppressor

There is an abundance of evidence both in vivo and at the molecular level that highlight $TR\beta$ tumor suppressor function. Loss of the transcription factor $TR\beta$, a member of the thyroid hormone receptor (TR) family, through mutation or epigenetic silencing is characteristic of thyroid and other endocrine-related cancers [1, 2, 6-9]. Restoration of $TR\beta$ function in malignant cells decreases tumor growth in xenograft studies, supporting a tumor suppressor role for $TR\beta$ [7, 11, 12]. In order to designate a particular factor as a tumor suppressor there are criteria that need to be met: 1) loss of the factor must result in cancer growth, and 2) restoration of the factor must reduce cancer growth [13, 14]. Over the course of the last two decades, it has been demonstrated through multiple studies that $TR\beta$ does indeed meet these criteria (Figure 1).

One of the first reports of potential TR β tumor suppressor activity was from a study of resistance to thyroid hormone [10]. Mice with a point mutation in the ligand binding domain of TR β that renders TR β unable to bind ligand (TR β^{PV}) unexpectedly exhibited enlarged thyroid glands, in addition to the symptoms of resistance to thyroid hormone syndrome. Further examination of the enlarged thyroid glands in TR β^{PV} mutant mice by histology revealed that the mice had developed thyroid cancer. Further studies from the same group demonstrated that thyroid-specific knockout mice also spontaneously develop thyroid cancer [15]. Histological sections from tissues of TR β knockout mice showed evidence of anaplasia, capsular invasion, vascular invasion, and metastatic lesions in the lung. In human thyroid cancer tissues, loss of TR β expression is correlated with dedifferentiation [16]. Normal thyroid epithelial cells have the highest TR β expression, while TR β expression is lowest in anaplastic thyroid cancer cells, the most aggressive form of thyroid cancer. Combined, these results establish that loss of TR β results in cancer growth.

Restoration of TR β signaling in TR β -low or TR β -null cell lines has been shown to slow tumor growth *in vivo*. This was first shown in MDA-MB-468 triple negative breast cancer cells with TR β restored, in a nude mouse xenograft model [6]. In the same study, SK-hep1 hepatocarcinoma cells with TR β restored showed reduced tumor growth [6]. Cells with restored TR β expression were also shown to have less metastatic potential than control cells. A separate study showed that FTC-133 follicular thyroid cancer cells with TR β restored show reduced growth in a xenograft study [9]. These TR β -expressing tumors showed evidence of reduced PI3K-Akt signaling and less blood vessel formation compared to tumors without TR β expression. Most recently, TR β restoration has been shown to suppress growth and migration in colorectal cancer cells [17], and block cancer stem cell out growth in luminal A breast cancer cell lines [18]. Our lab demonstrated that restoration of TR β in anaplastic thyroid cancer cells re-programs the transcriptome, promotes apoptosis, and suppresses many of their aggressive phenotypic traits [19]. Taken together, these results establish that restoration of TR β slows cancer growth.

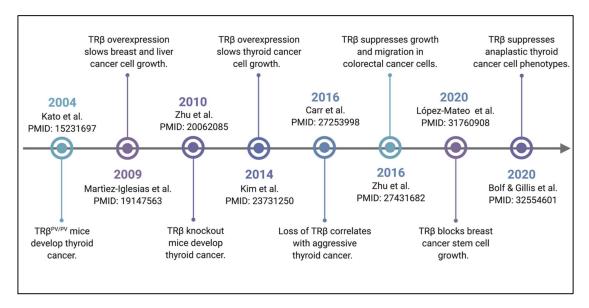


Figure 1. Timeline of seminal studies of $TR\beta$ tumor suppression. $TR\beta$ function was first linked to cancer growth when it was shown that a expression of ligand-binding domain mutant (TRB^{PV}) in mice led to spontaneous development of thyroid tumors. Increasing numbers of studies have demonstrated over the following two decades that $TR\beta$ is a classically-defined tumor suppressor.

2.2 TR\$ Attenuates the PI3K-Akt Signaling Pathway via Genomic and Nongenomic Mechanisms

Phosphoinositide 3-kinase (PI3K) signaling is a potent tumor activating pathway that is implicated in many solid and hematologic tumors [20-24]. PI3K is recruited to ligand-bound, phosphorylated receptor tyrosine kinases (RTKs) and phosphorylates the membrane lipid phosphatidylinositol 4,5-bisphosphate (PIP2) to phosphatidylinositol (3,4,5)-trisphosphate (PIP3). PIP3 is a docking lipid that anchors protein kinase B (Akt) for phosphorylation and activation by phosphoinositide-dependent kinase-1 (PDK1) on thr308 and mammalian target of rapamycin complex 2 (mTORC2) on ser473. Akt is a multi-substrate kinase that phosphorylates targets involved in apoptosis regulation, cell cycle progression, angiogenesis, and metabolism. An important function of Akt is the indirect activation of mTORC1, which phosphorylates a myriad of targets such as p70S6K and eukaryotic translation initiation factor 4B (eIF4B) to increase cell metabolism and protein translation. PI3K-Akt pathway activation is frequently overactive due to gain-of-function mutations in PI3K, loss of expression of phosphatase and tensin homolog (PTEN), or amplifications in RTKs and Akt [21, 23, 25].

TRβ has long been understood to directly alter the PI3K pathway through nongenomic mechanisms. Simoncini et al. first reported on the potential for hormone receptors to increase PIP3 content in endothelial cells [26]. Estrogen, dexamethasone, and thyroid hormone receptors all increased PIP3 levels following respective hormone treatment, suggesting a shared yet noncanonical role for hormone receptors in PI3K activation. In other normal cells such as human fibroblasts, treatment with 15 min of 10 nM T3 induced Akt phosphorylation, and phosphorylation of mTORC2 and p70S6K was observed after 30 min T3 exposure[27]. This effect was abrogated with PI3K inhibitors LY294002 and wortmannin. This rapid impact of T3 was unlikely due to TRβ-meditated transcription; indeed TRβ was shown to directly bind to the RTK localization subunit of PI3K, p85 α , independently of ligand (Figure 2) [27]. Interestingly, 100 nM of T3 did result in reduced TRβ-p85 α binding and increase in PI3K activity in fibroblasts [27]. These findings were confirmed in GH4C1 and CHO cells treated with 100 nM T3 for only five minutes [28], suggesting a rapid and conserved nongenomic function of TR β across diverse mammalian cell types.

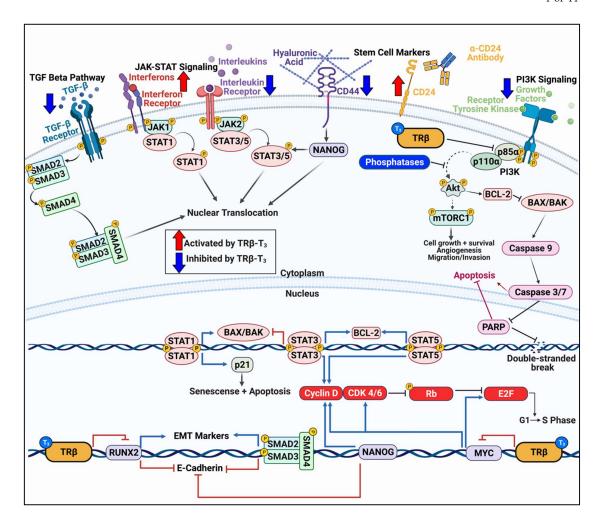


Figure 2. TR β regulates various oncogenic signaling pathways in cancer models that govern cell proliferation, migration, and apoptosis. TR β has shown to be a significant regulator of various oncogenic cell signaling pathways in diverse models of normal and cancer cells. TR β decreases TGF- β signaling resulting in decreased SMAD phosphorylation and transcription of EMT markers. TR β differentially regulates JAK-STAT signaling pathways which results in decreased STAT3 signaling and enhanced STAT1 response to induce apoptosis. TR β activation can reduce the cancer stem cell population as evident by decreased levels of CD44 and increased CD24 in breast and thyroid cancer models, leading to attenuated NANOG levels. TR β also has potent inhibitory effects on PI3K signaling whether via direct binding to the p85 α subunit or through genomic mechanisms to increase transcripts of phosphoinositol phosphatases and decrease receptor tyrosine kinases. TR β also modulates cell cycle genes in various cancer models to enhance expression and phosphorylation of Rb to stall the cell cycle. These potent effects on cancer cell transcriptional reprogramming allow for enhanced efficacy of targeted inhibitors on the PI3K pathway and cell cycle.

In cancer models however, the role of TR β on modulating PI3K appears to be more nuanced. TR β first appeared to have a role in regulating PI3K with the TR $\beta^{PV/PV}$ mouse model [29], in which Akt was hyperphosphorylated. It was later revealed that TR $\beta^{PV/PV}$ bound to p85 α with a higher affinity compared to wildtype TR β [30]. This is likely due to the C terminal frameshift in the ligand-binding domain resulting in higher p85 α binding affinity. Zhu et al. reported that absence of thyroid hormone receptors correlated with higher levels of phosphorylated Akt, mTORC1, and p70S6K in thyroid cancer [15]. Moriggi et al. showed that TR β could complex with p85 α in four of the six cancer cell lines investigated which resulted in either an increase or decrease in Akt phosphorylation [31]. This may reflect the levels of endogenous TR β in cell lines as well as individual genetic backgrounds of the cells.

Notably, the experiments were conducted at 24 and 48 hours of T_3 exposure, indicating that gene expression could have been at play in conjunction with p85 α binding. In MDA-MB-468 and SK-Hep1 cells transduced with TR β , pAkt was blunted in the presence of Insulin-like growth factor 1 (IGF-1) compared to control cells, suggesting a protective effect of TR β on the PI3K-Akt pathway [6]. Indeed, qPCR revealed a reduction in the RTKs epidermal growth factor receptor 1 (*EGFR1*), HER3 (*ERBB3*), and *IGFR1* in both cell lines [6].

In vivo studies using the follicular thyroid cancer cell lines FTC-133 and FTC-236 transfected with TR β revealed a decrease in pAkt, mTORC1, p70S6K, and eIF4B [9]. There was also a decrease in vascular endothelial growth factor (VEGF) levels which stimulates endothelial cells to promote angiogenesis by way of the PI3K pathway, further implicating a broad-spectrum tumor suppressive role of TR β . Additionally, TR β transduction resulted in reduced autocrine signaling in an MCF-7 model [18]. TR β -T $_3$ decreased expression of vascular endothelial growth factor receptor 9 (FGFR9) and cognate ligands FGF3 and FGF4, while blunting estrogen-mediated induction of FGF9. While PI3K activity itself was not measured, this study confirmed an additional mechanism of TR β modulation of players in the PI3K-Akt pathway.

There were similar findings in colorectal cancer cells in which long-term T_3 exposure resulted in reduced ser473 Akt phosphorylation by an unknown mechanism [32]. However, as we have recently described, $TR\beta$ may play a critical genomic role in PI3K regulation in anaplastic thyroid cancer (ATC) [33]. While short term (30 min) T_3 exposure failed to reduce pAkt levels, long term exposure (24 hours) reduced pAkt and pmTORC1 levels concordantly with changes in gene expression. T_3 - $TR\beta$ in ATC cells increased phosphatase levels such as phosphatidylinositol 4,5-bisphosphate 5-phosphatase A (*INPP51*), inositol polyphosphate 4-phosphatase type II (*INPP4B*), and PH domain and leucine rich repeat protein phosphatase 1 (*PHLPP1*). Importantly, INPP4B is a potent tumor suppressor of thyroid cancer in vivo [34]. Conversely, $TR\beta$ - T_3 decreased levels of receptor tyrosine kinases *ERBB3* (HER3), *FGFR3*, and *FGFR4*. This impact on PI3K pathway attenuation resulted in increased sensitivity to the PI3K inhibitors LY294002 and buparlisib, providing a provocative implication on the relationship between $TR\beta$ status in cancer patients and response to therapies.

A major consequence of PI3K-Akt signaling in cancer cells is the increase in cell metabolism. While the impact of TR β on normal metabolism is well studied [35, 36], there is little known on the tumor suppressor's role in cancer cell metabolism. We were able to determine that TRβ and T₃ potently modulate key metabolic pathways in triple negative breast cancer and ATC cells. Stearic acid is known to exhibit tumor suppressor functions in breast cancer by inducing apoptosis[37]. In our MDA-MB-468 cells transduced with TR β , T₃ induced expression of enzymes in the stearic acid synthesis pathway including Acetyl-CoA synthetase short chain family member 2 (ACSS2), glutaminase, and 6-phosphofructo-2-kinase/fructose-2,6-biphosphatase 3 (PFKFB1). In ATC cells, we noticed the potential for TRβ and T₃ to regulate glycogen metabolism, an oncogenic metabolic pathway that's been observed in many cancer models such as breast, colorectal, and pancreatic cancer[38-41]. Glycogen is a storage form of glucose for cancer cells to advantageously breakdown via glycogen phosphorylase in times of low cellular energy or oxidative stress (Figure 3)[42]. TR β and T₃ decreased expression of the brain isoform of glycogen phosphorylase (PYGB) and differentially regulated expression of key signaling proteins that activate PYGB such as cell migration inducing hyaluronidase (CEMIP), and the beta inhibitory subunit of phosphorylase kinase (PHK)[43]. Since these studies highlight two specific metabolic pathways regulated by TR β in cancer cells, further exploration will be required to ascertain the extent of how TR β induces global metabolic changes in cancer cells.

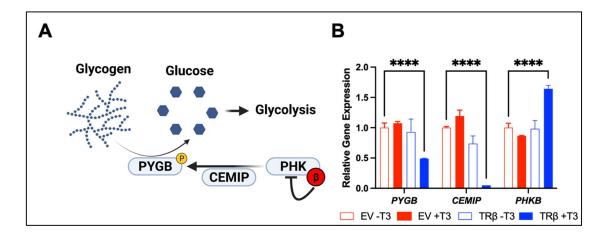


Figure 3. TR β may regulate glycogen metabolism in anaplastic thyroid cancer cells. RNA-sequencing[19] revealed a novel function of liganded TR β on metabolism in a cancer cell model. TR β and T3 induced significant changes in gene expression in the glycogen pathway to decrease levels of PYGB and the glycogen signaling protein CEMIP. TR β also enhanced the inhibitory beta subunit of PHK which phosphorylates and activates PYGB to enhance glycogen breakdown and potentially cell survival.

2.3 TRβ Differentially Influences JAK-STAT Signaling

Another important signaling cascade in cancer is the JAK-STAT pathway. Immune modulators such as interferons and interleukins activate cognate receptors to recruit Janus kinases (JAKs) to the membrane to phosphorylate specific signal transducer and activator of transcription proteins (STATs)[44]. Although the JAK-STAT pathways are best studied in immune cells to promote cell growth and the immune response, cancer cells can take advantage of the signaling pathway to contribute to malignant phenotypes[45-47]. STAT3 and STAT5 typically demonstrate tumor promoting activity by inhibiting apoptosis through repression of BAX and BAK transcription along with induction of BCL2 transcription (Figure 2)[48-50]. Conversely, JAK1 and STAT1 have shown to induce apoptosis in a variety of cell models, likely through BCL2 repression and CDKN1A (p21) transcription[51-54]. TRβ has been shown to specifically regulate activity of certain JAK-STAT pairs. For example, Guignon et al. showed that TRβ^{PV/PV} in a mouse model of breast cancer resulted in enhanced prolactin expression and sustained phosphorylation of STAT5[55], which is known to inhibit apoptosis and encourage cell cycle progression by promoting BCL2 and CCND1 (cyclin D) transcription, respectively[50, 56]. Introduction of wildtype TRB attenuated prolactin-induced p-STAT5 levels which were further reduced with addition of T₃. Park et al. showed that TRβ decreased JAK2, STAT3, and STAT5 phosphorylation and activation in MCF-7-TRβ cells[8]. This led to a decrease in tumor size and proliferation with an increase in apoptosis. In an ATC model with stably-transduced TRβ, we observed differential regulation in opposing JAK-STAT pathways[19]. STAT3 signaling was downregulated in the ATC cells while STAT1 signaling was induced compared to control cells. STAT1 activation led to induction of apoptosis as evident from Caspase 3 and poly (ADP-ribose) polymerase (PARP) cleavage. Importantly, we were able to activate STAT1 independently of TR β with 2-(1,8-Naphthyridin-2-ly)phenol. This study revealed a novel target in ATC, highlighting the importance of better understanding the tumor suppressor program of TRβ.

2.4 TRB Regulation of Cell Cycle Progression

The cell cycle is a crucial target for aggressive cancers (Figure 2). Perez-Juste et al. first observed that overexpression of $TR\beta$ with T_3 had profounds effects on neuron development in murine neural crest-derived cells[57]: $TR\beta$ activation led to a decrease in MYC and cyclin D1 expression with a concomitant increase in the cell cycle regulator p27. Since p27 can directly inhibit cyclin-dependent kinases (CDKs), the investigators unsurprisingly observed a decrease in phosphorylated

retinoblastoma protein (Rb), resulting in cell cycle arrest. The authors later confirmed the direct relationship of TR β to the cell cycle promoter MYC by elucidating a negative thyroid response element (TRE) on the *MYC* promoter[58]. Porlan et al. expanded on these observations by showing that TR β in 3T3 cells inhibited proliferation and the cell cycle via decreasing pRb, cyclin D 1, 2, and 3, and cyclin E[59]. Yen et al. showed similar findings in HepG2 cells using 10 nM T $_3$ for multiple days; there was an increase in p21 and decrease in cyclin E and pRb[60]. In pancreatic adenocarcinoma cell lines, TR β decreased cyclins D1 and E but increased p21, which led to an increase in p27 [61]. This excitingly allowed for an enhanced response to the antiproliferative agents gemcitabine and cisplatin. The impact of TR β on cell cycle has also been observed in vivo; Martinez et al. observed a decrease in cyclin E in hypothyroid patients with hepatocellular carcinoma or breast cancer[62]. These authors also noted increased p27 expression in their SK-TR β mouse model.

Lin et al. expanded on the mechanism by showing that TR β with T $_3$ increases endoglin expression to stabilize p21 protein in HCC cells[63]. Cell cycle arrest was also measured in MCF-7-TR β cells with long term T $_3$ exposure; transcriptomic analysis revealed a decrease in cell cycle related gene transcripts including MYC and members of the E2F family[18]. Finally, we recently performed RNA-sequencing to capture the full cell cycle signaling pathway with TR β and 24 hours of T $_3$ [19]. We observed a decrease in CCND1 and MYC expression and an increase in CDKN1A, highlighting the importance of TR β on cell cycle regulation. Importantly, we showed that TR β enhanced the efficacy of palbociclib, a CDK4/6 inhibitor. Palbociclib is frequently used to treat ER $^+$ and HER2 $^+$ breast cancers and is in clinical trials for several solid tumors[64-67]. However, resistance to CDK inhibition often develops, highlighting a potentially useful diagnostic and role for TR β in predicting response to CDK inhibitors.

2.5 Impact of TRβ on TGF-β Signaling

Another important signaling pathway in cancer is the transforming growth factor beta (TGF- β) signaling cascade. TGF- β stimulates its cognate receptor to induce dimerization and autophosphorylation to phosphorylate mothers against decapentaplegic homolog (SMADs) [68-70]. The SMAD complex is translocated to the nucleus to regulate gene transcription (Figure 2). In normal cells, TGF- β signaling regulates the cell cycle and can promote apoptosis. In cancer cells however, mutations in TGF- β , SMADs, or SMAD binding partners can induce a tumor promoting gene expression program[68-70]. While TR α and T $_3$ positively regulated TGF- β signaling in liver cancer cells[60], TR β and T $_3$ appear to negatively regulate TGF- β . This was first observed in GH $_4$ C1 cells as well as an in vivo model[71]. TR β and T $_3$ downregulated TGF- β signaling induction, partly by directly competing for SMAD binding sites, resulting in decreased fibrosis. In transduced MCF-7 cells, López-Mateo et al. noted that TR β -T $_3$ could blunt TGF- β -induced SMAD2 and SMAD3 phosphorylation in MCF-7 cells, leading to a decrease in both SMAD2 and SMAD3 transcriptional activity [18]. TGF- β signaling attenuation is a notable and conserved feature of liganded TR β that further highlights its function as a broad-spectrum, potent tumor suppressor.

2.6 TRβ Inhibits Epithelial–Mesenchymal Transition

Aggressive cancers display hallmarks of epithelial–mesenchymal transition (EMT) as the cell becomes more metastatic. $TR\beta$ has been shown to reduce EMT in several cancer models (Figure 2). First, Martinez-Iglesias et al. demonstrated that $TR\beta$ reduced expression of vimentin, beta catenin, and matrix metallopeptidase 1 and 9 (MMP1 and MMP9) in breast and liver cancer models, leading to a decrease in malignancy in vivo[6]. Dentice et al. further showed an increase in E cadherin with T_3 in colon cancer cells [72]. We also showed that there is a direct and positive relationship between $TR\beta$ expression/activation and mesenchymal—epithelial transition (MET) in thyroid cancer[73]. $TR\beta$ directly regulates the expression of the RUNX family transcription factor 2 (RUNX2), a master EMT transcription factor. There was an inverse relationship observed between $TR\beta$ and RUNX2 expression in thyroid cancer from normal cells to the highly dedifferentiated anaplastic thyroid cancer. T_3 reduced RUNX2 expression in normal and ATC cells, and $TR\beta$ knockdown increased RUNX2, increasing the expression of MMP2, MMP13, cyclin D1, osteopontin (OPN), and cadherin 6. Transfected $TR\beta$ also repressed RUNX2 and EMT markers in MDA-MB-231 cells; conversely, $TR\beta$ -

knockdown in breast epithelial-like MCF10A cells caused an increase in RUNX2 and EMT markers [74]. López-Mateo et al. confirmed a decrease in EMT genes such as vimentin and snail family transcriptional repressor 2 (SLUG) in estrogen receptor alpha positive (ER α ⁺) breast cancer cells[18]. We recently observed more evidence of MET in a transduced ATC model in which TR β and T₃ increased transcript levels of E cadherin and decreased vimentin mRNA [19].

$2.2\ TR\beta$ Promotes Cancer Cell Re-Differentiation

EMT is closely correlated with dedifferentiation, a phenotype that does not resemble the original tissue of origin but more closely resembles a stem cell [75]. Dedifferentiation is a crucial process in the cancer cell to evade the immune system, enhance proliferation, promote angiogenesis, and develop drug resistance [76, 77]. Modulation in expression of specific tissue markers is often correlated with dedifferentiation. For example, breast cancer cells increaseexpression of cytokeratins, proteins that are secreted into the extracellular matrix (ECM) or attached at the cell surface [78, 79]. These keratins are advantageous modulators of the ECM that allow for enhanced cancer cell migration and invasion [80]. TR β decreased the expression of keratins 8 and 18 in MCF-7 cells [6, 62]. These two keratins, amongst other keratin isoforms, were also reduced in stem cell models of ER α + breast cancer [18]. Furthermore, we observed a decrease in mRNA and protein levels of keratins 5 and 14 in a triple negative breast cancer model. It may be possible that TR β canonically regulates keratin expression, as this has been observed in organisms such as *Xenopus laevis* [81].

TR β appears to induce re-differentiation in cells to encourage expression of normal cell markers, agnostic of cell type. Perra et al. noted a striking difference in rats with preneoplastic liver lesions treated with the selective TR β agonist sobetirome (GC-1): TR β activation resulted in loss of dedifferentiation markers and reacquisition of differentiated liver proteins [82]. This phenomenon was also detected in colon cancer cells, in which T $_3$ induced robust expression of normal colon markers sucrase isomaltase and intestinal alkaline phosphatase and slowed the proliferation of the cancer cells[72].

In addition to breast and colon cancer models, $TR\beta$ has shown to induce re-differentiation in thyroid cancer cells. We recently demonstrated that $TR\beta$ and T_3 induced the re-expression of several key thyroid specific genes that are lost in dedifferentiated thyroid cancer[19]. These included iodothyronine deiodinase 2 (DIO2), dual oxidase 1 (DUOX1), thyroid peroxidase (TPO), and thyroglobulin (TG). Excitingly, we were also able to induce expression of these genes plus six other thyroid specific markers by using the potent $TR\beta$ -specific analog GC-1 to activate the low level of $TR\beta$ expressed in ATC [83]. The sodium iodide symporter (NIS) transcript and protein level were increased using GC-1, which allowed for a significantly higher intake of iodide in cell culture models. These studies not only demonstrate the role of $TR\beta$ in re-differentiation programming but could have functional significance as induction of NIS could be exploited for radioactive iodide treatment in thyroid and breast cancers.

Finally, in the course of dedifferentiation, aggressive cancers become more stemlike. This allows for unlimited replicative potential and evasion from the immune system [75]. TR β appears to reduce the stem cell population in both breast and thyroid cancers. López-Mateo et al. showed that TR β activation in MCF-7 cells reduced the mammosphere population[18]. This was associated with a decrease in the breast stem cell markers aldehyde dehydrogenase (ALDH1) and CD44, with an increase in the monolayer marker CD24. An increase in CD24 expression is not only notable for demonstrating a decrease in the stem cell population, but it can also be used as a drug target for promoting tumor cell clearing by macrophages in the tumor microenvironment[84, 85]. They also observed a decrease in SRY-box transcription factor 2 (SOX2) and NANOG, which are downstream of CD44 and TGF- β signaling. We also observed a decline in the stem cell population in both ATC and MDA-MB-468 breast cancer cells by manipulating TR β levels[19]. We noted that ALDH, POU5F1 (encodes OCT3/4), CD44, FUT4 (encodes SSEA-1), and PROM1 expression were significantly downregulated in cancer cells transduced with TR β . We have also recently been able to see this change in ATC cells with GC-1 that resulted in stem cell death, increase in CD24 expression, and decrease in CD44 expression [83]. We also observed an enhanced efficacy to the inhibitors buparlisib

(PI3K), sorafenib (MAPK), and palbociclib (cell cycle) in the stem cell population. The use of $TR\beta$ screening and specific activation may help reduce the stem cell population and increase the efficacy of clinically relevant therapeutics in ATC patients.

2.8 TRβ interactions with Epigenetic Modulators are Key to Tumor Suppression

In its capacity as a transcription factor, $TR\beta$ is a hub for incoming molecular signals that include fluctuations in hormone levels, input from cellular signaling pathways, and post-translational modifications. It must integrate all of these signals to recruit the necessary coregulators and excute a transcriptional response. $TR\beta$ has a diverse repertoire of potential binding partners in normal cells, as evidenced by immunoprecipitation to mass spectrometry studies [86, 87], and by our own proximity ligation assays [88]. By complexing with a variety of co-regulators $TR\beta$ acts to coordinate complex gene regulatory events that have variety of implications in maintenance of cellular homeostasis and tumor suppression. Disruption of these crucial interactions in cancer cells may lead to either a loss of response to T_3 or to abberant transcription.

Many coactivators have been implicated in T₃-dependent gene activation by TRβ, including the steroid receptor coactivator (SRC), p300/CBP histone acetyl transferases, and the mediator-like TR associated proteins (TRAP)/DRIP) [89]. SRC interacts directly with liganded TRβ and serves as an adapter molecule to facilitate recruitment of p300/CBP to acetylate histones and interact with components of basal transcriptional machinery. The TRAP complex is a mutlisubunit coactivator complex that interacts with liganded TRs and recruits RNA polymerase II to promoters. Chromatin immunoprecipitation experiments have demonstrated that upon T₃ binding, TRβ first recruits SRC proteins and p300, resulting in histone acetylation, followed by the TRAP complex [90]. Together these coactivators facilitate transcriptional activation through a stepwise process of acetylation of histones to decompact the local chromatin and subsequent recruitement of the basal transcriptional machinery. Compounds that target the specific interactions between nuclear hormone receptors and SRC have been developed, and have shown early signs of therapeutic benefit *in vivo* [91].

TR β also complexes with a variety of nuclear co-repressors. Specifically, evidence suggests that TR β represses gene expression via the recruitment of either nuclear co-repressor 1 (NCoR1) or silencing mediator for retinoid or thyroid-hormone receptors (SMRT, NCoR2) [92-94]. NCoR1 and SMRT are highly homologous and contain three similar nuclear receptor interaction domains. Furthermore, both NCoR1 and SMRT bind to TR β , as well as other nuclear hormone receptors via similar mechanisms at specific residues [95]. The seminal article that first identified NCoR1 as a crucial regulatory protein found that it bound TR β at amino acid residues 203-230 but that amino acid residues 230-260 act to stabilize this interaction [96]. NCoR1 and SMRT directly recruit and interact with Class II histone deacetylases (HDAC), and recruit Class I HDAC's via linker proteins such as Sin3a or Sin3b [97-100]. NCoR1 itself has been demonstrated to be critical for suppression of breast cancer growth in coordination with TR β [101]. HDAC inhibitors may be useful in combinatiton with hormone therapy, particularly in the context of anti-estrogen or anti-androgen [102, 103] resistance. The future use of compounds that can promote interactions between TR β and its coregulators, or block interactions with negative regulators, may be an attractive way to enhance the beneficial effects of thyroid hormone or thyromimetics.

3. Conclusions

Despite a recognized role as a tumor suppressor, the potential for TR β as a therapeutic target and diagnostic indicator remains untapped. This is in part because, until recently, the mechanisms by which TR β regulates tumor growth were unclear. TR β is a ligand-dependent nuclear receptor that mediates the effects of T $_3$ on many biological processes, and therefore has potent effects throughout the cell as disussed in this review. At the genomic level, TR β mediates the effects of T $_3$ via the regulation of gene expression through the recruitment of co-regulators and chromatin remodeling complexes to genomic regulatory elements to alter target gene transcription. Disruption of TR β is therefore expected to alter assembly of co-regulator complexes needed for initiation of gene transcription. TR β -selective thyromimetics have been developed [104] and have been shown to elicit

the same transcriptional response as T₃[105]. Current work in our group is focused on stimulation of endogenous TR β , even when it is expressed at the low levels in cancer cells, to elicit an anti-tumor response [83]. Our work suggests TR β agonists can be combined with modulators of other pathways to enhance their efficacy, and opens many new avenues for exploration of TR β tumor suppressive action.

Author Contributions: Conceptualization, CDD, NEG, and FEC; writing—original draft preparation, CDD and NEG.; visualization, CDD and NEG; writing—review and editing, CDD, NEG, FEC. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by grants from National Institutes of Health U54 GM115516 for the Northern New England Clinical and Translational Research Network; National Cancer Institute 1F99CA245796-01; UVM Cancer Center-Lake Champlain Cancer Research Organization (C3) 12577-21; and UVM Larner College of Medicine.

Acknowledgements: Biorender vector graphics were used to construct the figures.

Conflicts of Interest: The authors declare no conflict of interest.

References

- 1 Kim WG, Cheng SY. Thyroid hormone receptors and cancer. *Biochimica et biophysica acta* 2013; 1830: 3928-3936.
- Aranda A, Martinez-Iglesias O, Ruiz-Llorente L, Garcia-Carpizo V, Zambrano A. Thyroid receptor: roles in cancer. *Trends in endocrinology and metabolism: TEM* 2009; 20: 318-324.
- Landa I, Ibrahimpasic T, Boucai L, Sinha R, Knauf JA, Shah RH et al. Genomic and transcriptomic hallmarks of poorly differentiated and anaplastic thyroid cancers. J Clin Invest 2016; 126: 1052-1066.
- Joseph B, Ji M, Liu D, Hou P, Xing M. Lack of mutations in the thyroid hormone receptor (TR) alpha and beta genes but frequent hypermethylation of the TRbeta gene in differentiated thyroid tumors. *The Journal of clinical endocrinology and metabolism* 2007; 92: 4766-4770.
- Puzianowska-Kuznicka M, Krystyniak A, Madej A, Cheng SY, Nauman J. Functionally impaired TR mutants are present in thyroid papillary cancer. *The Journal of clinical endocrinology and metabolism* 2002; 87: 1120-1128.
- Martínez-Iglesias O, Garcia-Silva S, Tenbaum SP, Regadera J, Larcher F, Paramio JM *et al.* Thyroid hormone receptor beta1 acts as a potent suppressor of tumor invasiveness and metastasis. *Cancer Res* 2009; 69: 501-509.
- 7 Kim W, Zhu X, Kim D, Zhang L, Kebebew E, Cheng S. Reactivation of the silenced thyroid hormone receptor B gene expression delays thyroid tumor progression., vol. 154: Endocrinology, 2013, pp 25-35.
- Park JW, Zhao L, Cheng SY. Inhibition of estrogen-dependent tumorigenesis by the thyroid hormone receptor beta in xenograft models. *Am J Cancer Res* 2013; 3: 302-311.
- 9 Kim WG, Zhao L, Kim DW, Willingham MC, Cheng SY. Inhibition of tumorigenesis by the thyroid hormone receptor beta in xenograft models. *Thyroid* 2014; 24: 260-269.
- 10 Kato Y, Ying H, Willingham MC, Cheng SY. A tumor suppressor role for thyroid hormone beta receptor in a mouse model of thyroid carcinogenesis. *Endocrinology* 2004; 145: 4430-4438.
- 11 Martinez-Iglesias O, Garcia-Silva S, Tenbaum S, Regadera J, Larcher F, Paramio J *et al*. Thyroid hormone receptor β 1 acts as a potent suppressor of tumor invasiveness and metastasis., vol. 69: Cancer Res, 2009, pp 501-509.
- Park J, Zhao L, Cheng S. Inhibition of estrogen-dependent tumorigenesis by the thyroid hormone receptor B in xenograft models., vol. 3: Am J Cancer Res, 2013, pp 302-311.
- 13 Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. Cell 2011; 144: 646-674.
- 14 Weinberg RA. Oncogenes and tumor suppressor genes. CA Cancer J Clin 1994; 44: 160-170.
- Zhu XG, Zhao L, Willingham MC, Cheng SY. Thyroid hormone receptors are tumor suppressors in a mouse model of metastatic follicular thyroid carcinoma. *Oncogene* 2010; 29: 1909-1919.
- Carr FE, Tai PW, Barnum MS, Gillis NE, Evans KG, Taber TH *et al*. Thyroid Hormone Receptor-β (TRβ) Mediates Runt-Related Transcription Factor 2 (Runx2) Expression in Thyroid Cancer Cells: A Novel Signaling Pathway in Thyroid Cancer. *Endocrinology* 2016; 157: 3278-3292.
- Zhu L, Tian G, Yang Q, De G, Zhang Z, Wang Y *et al*. Thyroid hormone receptor β1 suppresses proliferation and migration by inhibiting PI3K/Akt signaling in human colorectal cancer cells. *Oncol Rep* 2016; 36: 1419-1426.

- 18 López-Mateo I, Alonso-Merino E, Suarez-Cabrera C, Park JW, Cheng SY, Alemany S *et al.* Thyroid Hormone Receptor β Inhibits Self-Renewal Capacity of Breast Cancer Stem Cells. *Thyroid* 2020; 30: 116-132.
- Bolf EL, Gillis NE, Davidson CD, Rodriguez PD, Cozzens L, Tomczak JA *et al.* Thyroid Hormone Receptor Beta Induces a Tumor-Suppressive Program in Anaplastic Thyroid Cancer. *Molecular Cancer Research* 2020; 18: 1443-1452.
- 20 Carnero A, Blanco-Aparicio C, Renner O, Link W, Leal JF. The PTEN/PI3K/AKT signalling pathway in cancer, therapeutic implications. Curr Cancer Drug Targets 2008; 8: 187-198.
- 21 Martini M, De Santis MC, Braccini L, Gulluni F, Hirsch E. PI3K/AKT signaling pathway and cancer: an updated review. *Annals of medicine* 2014; 46: 372-383.
- 22 Revathidevi S, Munirajan AK. Akt in cancer: mediator and more. Seminars in cancer biology 2019.
- Jiang N, Dai Q, Su X, Fu J, Feng X, Peng J. Role of PI3K/AKT pathway in cancer: the framework of malignant behavior. *Molecular biology reports* 2020; 47: 4587-4629.
- 24 Hoxhaj G, Manning BD. The PI3K-AKT network at the interface of oncogenic signalling and cancer metabolism. *Nature reviews Cancer* 2020; 20: 74-88.
- Wang Y, Hou P, Yu H, Wang W, Ji M, Zhao S *et al*. High prevalence and mutual exclusivity of genetic alterations in the phosphatidylinositol-3-kinase/akt pathway in thyroid tumors. *The Journal of clinical endocrinology and metabolism* 2007; 92: 2387-2390.
- Simoncini T, Hafezi-Moghadam A, Brazil DP, Ley K, Chin WW, Liao JK. Interaction of oestrogen receptor with the regulatory subunit of phosphatidylinositol-3-OH kinase. *Nature* 2000; 407: 538-541.
- Cao X, Kambe F, Moeller LC, Refetoff S, Seo H. Thyroid hormone induces rapid activation of Akt/protein kinase B-mammalian target of rapamycin-p70S6K cascade through phosphatidylinositol 3-kinase in human fibroblasts. *Molecular endocrinology (Baltimore, Md)* 2005; 19: 102-112.
- Storey NM, Gentile S, Ullah H, Russo A, Muessel M, Erxleben C *et al.* Rapid signaling at the plasma membrane by a nuclear receptor for thyroid hormone. *Proc Natl Acad Sci U S A* 2006; 103: 5197-5201.
- 29 Kim CS, Vasko VV, Kato Y, Kruhlak M, Saji M, Cheng SY et al. AKT activation promotes metastasis in a mouse model of follicular thyroid carcinoma. Endocrinology 2005; 146: 4456-4463.
- Furuya F, Lu C, Willingham MC, Cheng SY. Inhibition of phosphatidylinositol 3-kinase delays tumor progression and blocks metastatic spread in a mouse model of thyroid cancer. *Carcinogenesis* 2007; 28: 2451-2458.
- Moriggi G, Verga Falzacappa C, Mangialardo C, Michienzi S, Stigliano A, Brunetti E *et al*. Thyroid hormones (T3 and T4): dual effect on human cancer cell proliferation. *Anticancer Res* 2011; 31: 89-96.
- 32 Zhu L, Tian G, Yang Q, De G, Zhang Z, Wang Y *et al*. Thyroid hormone receptor β1 suppresses proliferation and migration by inhibiting PI3K/Akt signaling in human colorectal cancer cells. *Oncology reports* 2016; 36: 1419-1426.
- Davidson CD, Bolf EL, Gillis NE, Cozzens LM, Tomczak JA, Carr FE. Thyroid Hormone Receptor Beta Inhibits PI3K-Akt-mTOR Signaling Axis in Anaplastic Thyroid Cancer via Genomic Mechanisms. *Journal of the Endocrine Society* 2021; 5.
- Li Chew C, Lunardi A, Gulluni F, Ruan DT, Chen M, Salmena L *et al.* In Vivo Role of INPP4B in Tumor and Metastasis Suppression through Regulation of PI3K-AKT Signaling at Endosomes. *Cancer Discov* 2015; 5: 740-751.
- Pramfalk C, Pedrelli M, Parini P. Role of thyroid receptor β in lipid metabolism. *Biochimica et biophysica acta* 2011; 1812: 929-937.
- 36 Master AN, A. THRB (Thyroid Hormone Receptor, Beta). Atlas Genet Cytogenet Oncol Haematol 2014; 18: 400-433.
- Evans LM, Cowey SL, Siegal GP, Hardy RW. Stearate preferentially induces apoptosis in human breast cancer cells. *Nutr Cancer* 2009; 61: 746-753.
- Favaro E, Bensaad K, Chong MG, Tennant DA, Ferguson DJ, Snell C *et al*. Glucose utilization via glycogen phosphorylase sustains proliferation and prevents premature senescence in cancer cells. *Cell metabolism* 2012; 16: 751-764.
- Pelletier J, Bellot G, Gounon P, Lacas-Gervais S, Pouyssegur J, Mazure NM. Glycogen Synthesis is Induced in Hypoxia by the Hypoxia-Inducible Factor and Promotes Cancer Cell Survival. *Frontiers in oncology* 2012; 2: 18.
- 40 Lee WN, Guo P, Lim S, Bassilian S, Lee ST, Boren J et al. Metabolic sensitivity of pancreatic tumour cell apoptosis to glycogen phosphorylase inhibitor treatment. British journal of cancer 2004; 91: 2094-2100.
- 41 Davidson CD CF. Review of pharmacological inhibition of thyroid cancer metabolism. . *J Cancer Metastasis Treat* 2021; 7:[Accept]. http://dx.doi.org/10.20517/2394-4722.2021.77.
- 42 Dauer P, Lengyel E. New Roles for Glycogen in Tumor Progression. Trends Cancer 2019; 5: 396-399.

- Terashima M, Fujita Y, Togashi Y, Sakai K, De Velasco MA, Tomida S *et al.* KIAA1199 interacts with glycogen phosphorylase kinase beta-subunit (PHKB) to promote glycogen breakdown and cancer cell survival. *Oncotarget* 2014; 5: 7040-7050.
- 44 Harrison DA. The Jak/STAT pathway. Cold Spring Harb Perspect Biol 2012; 4.
- 45 Seif F, Khoshmirsafa M, Aazami H, Mohsenzadegan M, Sedighi G, Bahar M. The role of JAK-STAT signaling pathway and its regulators in the fate of T helper cells. *Cell Commun Signal* 2017; 15: 23.
- 46 Owen KL, Brockwell NK, Parker BS. JAK-STAT Signaling: A Double-Edged Sword of Immune Regulation and Cancer Progression. Cancers (Basel) 2019; 11.
- 47 Brooks AJ, Putoczki T. JAK-STAT Signalling Pathway in Cancer. Cancers (Basel) 2020; 12.
- 48 Kamran MZ, Patil P, Gude RP. Role of STAT3 in cancer metastasis and translational advances. *BioMed research international* 2013; 2013: 421821.
- 49 Huynh J, Chand A, Gough D, Ernst M. Therapeutically exploiting STAT3 activity in cancer using tissue repair as a road map. *Nature reviews Cancer* 2019; 19: 82-96.
- 50 Halim CE, Deng S, Ong MS, Yap CT. Involvement of STAT5 in Oncogenesis. *Biomedicines* 2020; 8.
- 51 Chin YE, Kitagawa M, Kuida K, Flavell RA, Fu XY. Activation of the STAT signaling pathway can cause expression of caspase 1 and apoptosis. *Mol Cell Biol* 1997; 17: 5328-5337.
- 52 Stephanou A, Latchman DS. STAT-1: a novel regulator of apoptosis. Int J Exp Pathol 2003; 84: 239-244.
- 53 Sironi JJ, Ouchi T. STAT1-induced apoptosis is mediated by caspases 2, 3, and 7. *The Journal of biological chemistry* 2004; 279: 4066-4074.
- 54 Su Q, Wang F, Dong Z, Chen M, Cao R. IFN-γ induces apoptosis in human melanocytes by activating the JAK1/STAT1 signaling pathway. *Molecular medicine reports* 2020; 22: 3111-3116.
- 55 Guigon CJ, Kim DW, Willingham MC, Cheng SY. Mutation of thyroid hormone receptor-β in mice predisposes to the development of mammary tumors. *Oncogene* 2011; 30: 3381-3390.
- 56 Debierre-Grockiego F. Anti-apoptotic role of STAT5 in haematopoietic cells and in the pathogenesis of malignancies. *Apoptosis* 2004; 9: 717-728.
- 57 Perez-Juste G, Aranda A. The cyclin-dependent kinase inhibitor p27(Kip1) is involved in thyroid hormone-mediated neuronal differentiation. *The Journal of biological chemistry* 1999; 274: 5026-5031.
- Pérez-Juste G, García-Silva S, Aranda A. An element in the region responsible for premature termination of transcription mediates repression of c-myc gene expression by thyroid hormone in neuroblastoma cells. *The Journal of biological chemistry* 2000; 275: 1307-1314.
- Porlan E, Vega S, Iglesias T, Rodríguez-Peña A. Unliganded thyroid hormone receptor beta1 inhibits proliferation of murine fibroblasts by delaying the onset of the G1 cell-cycle signals. *Oncogene* 2004; 23: 8756-8765.
- Yen CC, Huang YH, Liao CY, Liao CJ, Cheng WL, Chen WJ et al. Mediation of the inhibitory effect of thyroid hormone on proliferation of hepatoma cells by transforming growth factor-beta. Journal of molecular endocrinology 2006; 36: 9-21.
- Michienzi S, Bucci B, Verga Falzacappa C, Patriarca V, Stigliano A, Panacchia L et al. 3,3',5-Triiodo-L-thyronine inhibits ductal pancreatic adenocarcinoma proliferation improving the cytotoxic effect of chemotherapy. The Journal of endocrinology 2007; 193: 209-223.
- 62 Martinez-Iglesias O, Garcia-Silva S, Regadera J, Aranda A. Hypothyroidism enhances tumor invasiveness and metastasis development. *PLoS One* 2009; 4: e6428.
- Lin YH, Huang YH, Wu MH, Wu SM, Chi HC, Liao CJ *et al.* Thyroid hormone suppresses cell proliferation through endoglin-mediated promotion of p21 stability. *Oncogene* 2013; 32: 3904-3914.
- Turner NC, Ro J, André F, Loi S, Verma S, Iwata H *et al.* Palbociclib in Hormone-Receptor-Positive Advanced Breast Cancer. *The New England journal of medicine* 2015; 373: 209-219.
- Karasic TB, O'Hara MH, Teitelbaum UR, Damjanov N, Giantonio BJ, d'Entremont TS et al. Phase II Trial of Palbociclib in Patients with Advanced Esophageal or Gastric Cancer. Oncologist 2020; 25: e1864-e1868.
- Sepúlveda-Sánchez JM, Gil-Gil M, Alonso-García M, Vaz Salgado M, Vicente E, Mesía Barroso C et al. Phase II Trial of Palbociclib in Recurrent Retinoblastoma-Positive Anaplastic Oligodendroglioma: A Study from the Spanish Group for Research in Neuro-Oncology (GEINO). Target Oncol 2020; 15: 613-622.
- 67 Serra F, Lapidari P, Quaquarini E, Tagliaferri B, Sottotetti F, Palumbo R. Palbociclib in metastatic breast cancer: current evidence and real-life data. *Drugs Context* 2019; 8: 212579.
- 68 Hata A, Chen YG. TGF-β Signaling from Receptors to Smads. Cold Spring Harb Perspect Biol 2016; 8.
- Morikawa M, Derynck R, Miyazono K. TGF-β and the TGF-β Family: Context-Dependent Roles in Cell and Tissue Physiology. *Cold Spring Harb Perspect Biol* 2016; 8.
- 70 Vander Ark A, Cao J, Li X. TGF-β receptors: In and beyond TGF-β signaling. *Cellular signalling* 2018; 52: 112-120.

- Alonso-Merino E, Martín Orozco R, Ruíz-Llorente L, Martínez-Iglesias OA, Velasco-Martín JP, Montero-Pedrazuela A *et al.* Thyroid hormones inhibit TGF-β signaling and attenuate fibrotic responses. *Proc Natl Acad Sci U S A* 2016; 113: E3451-3460.
- Dentice M, Luongo C, Ambrosio R, Sibilio A, Casillo A, Iaccarino A *et al.* β-Catenin regulates deiodinase levels and thyroid hormone signaling in colon cancer cells. *Gastroenterology* 2012; 143: 1037-1047.
- Carr FE, Tai PW, Barnum MS, Gillis NE, Evans KG, Taber TH *et al*. Thyroid Hormone Receptor-beta (TRbeta) Mediates Runt-Related Transcription Factor 2 (Runx2) Expression in Thyroid Cancer Cells: A Novel Signaling Pathway in Thyroid Cancer. *Endocrinology* 2016; 157: 3278-3292.
- Bolf EL, Gillis NE, Barnum MS, Beaudet CM, Yu GY, Tomczak JA *et al*. The Thyroid Hormone Receptor-RUNX2 Axis: A Novel Tumor Suppressive Pathway in Breast Cancer. *Hormones & cancer* 2020; 11: 34-41.
- 75 Nassar D, Blanpain C. Cancer Stem Cells: Basic Concepts and Therapeutic Implications. *Annu Rev Pathol* 2016; 11: 47-76.
- 76 Liu J. The dualistic origin of human tumors. Seminars in cancer biology 2018; 53: 1-16.
- 77 Wang H, Unternaehrer JJ. Epithelial-mesenchymal Transition and Cancer Stem Cells: At the Crossroads of Differentiation and Dedifferentiation. *Dev Dyn* 2019; 248: 10-20.
- Vora HH, Patel NA, Rajvik KN, Mehta SV, Brahmbhatt BV, Shah MJ *et al.* Cytokeratin and vimentin expression in breast cancer. *Int J Biol Markers* 2009; 24: 38-46.
- Alshareeda AT, Soria D, Garibaldi JM, Rakha E, Nolan C, Ellis IO *et al.* Characteristics of basal cytokeratin expression in breast cancer. *Breast Cancer Res Treat* 2013; 139: 23-37.
- 80 Karantza V. Keratins in health and cancer: more than mere epithelial cell markers. Oncogene 2011; 30: 127-138.
- 81 Mathisen PM, Miller L. Thyroid hormone induces constitutive keratin gene expression during Xenopus laevis development. Mol Cell Biol 1989; 9: 1823-1831.
- Perra A, Kowalik MA, Pibiri M, Ledda-Columbano GM, Columbano A. Thyroid hormone receptor ligands induce regression of rat preneoplastic liver lesions causing their reversion to a differentiated phenotype. *Hepatology* 2009; 49: 1287-1296.
- Gillis NE, Davidson CD, Cozzens LM, Wilson E, Bolf EL, Tomczak JA *et al.* A Thyroid Hormone Receptor Beta Specific Agonist Suppresses Anaplastic Thyroid Cancer Cell Phenotype and Increases Efficacy of Therapeutic Agents. *bioRxiv* 2021: 2021.2006.2009.447689.
- Salnikov AV, Bretz NP, Perne C, Hazin J, Keller S, Fogel M *et al*. Antibody targeting of CD24 efficiently retards growth and influences cytokine milieu in experimental carcinomas. *British journal of cancer* 2013; 108: 1449-1459.
- Barkal AA, Brewer RE, Markovic M, Kowarsky M, Barkal SA, Zaro BW *et al.* CD24 signalling through macrophage Siglec-10 is a target for cancer immunotherapy. *Nature* 2019; 572: 392-396.
- 86 Hahm JB, Schroeder AC, Privalsky ML. The two major isoforms of thyroid hormone receptor, TRα1 and TRβ1, preferentially partner with distinct panels of auxiliary proteins. *Mol Cell Endocrinol* 2014; 383: 80-95.
- Fozzatti L, Lu C, Kim D-W, Cheng S-y. Differential Recruitment of Nuclear Coregulators Directs the Isoform-Dependent Action of Mutant Thyroid Hormone Receptors. *Molecular Endocrinology* 2011; 25: 908-921.
- 688 Gillis NE, Boyd JR, Tomczak JA, Frietze S, Carr FE. Thyroid Hormone Dependent Transcriptional Programming by TRβ Requires SWI/SNF Chromatin Remodelers. *bioRxiv* 2021: 2021.2003.2022.436429.
- 89 Lee KC, Li J, Cole PA, Wong J, Kraus WL. Transcriptional Activation by Thyroid Hormone Receptor-β Involves Chromatin Remodeling, Histone Acetylation, and Synergistic Stimulation by p300 and Steroid Receptor Coactivators. *Molecular Endocrinology* 2003; 17: 908-922.
- 90 Sharma D, Fondell JD. Ordered recruitment of histone acetyltransferases and the TRAP/Mediator complex to thyroid hormone-responsive promoters in vivo. *Proceedings of the National Academy of Sciences* 2002; 99: 7934-7939.
- 91 Skowron KJ, Booker K, Cheng C, Creed S, David BP, Lazzara PR *et al.* Steroid receptor/coactivator binding inhibitors: An update. *Mol Cell Endocrinol* 2019; 493: 110471.
- Astapova I, Hollenberg AN. The in vivo role of nuclear receptor corepressors in thyroid hormone action. *Biochim Biophys Acta* 2013; 1830: 3876-3881.
- 93 Yen PM. Physiological and molecular basis of thyroid hormone action. *Physiol Rev* 2001; 81: 1097-1142.
- 94 Astapova I. Role of co-regulators in metabolic and transcriptional actions of thyroid hormone. J Mol Endocrinol 2016; 56: 73-97.
- 95 Hu X, Lazar MA. The CoRNR motif controls the recruitment of corepressors by nuclear hormone receptors. *Nature* 1999; 402: 93-96.

- 96 Horlein AJ, Naar AM, Heinzel T, Torchia J, Gloss B, Kurokawa R et al. Ligand-independent repression by the thyroid hormone receptor mediated by a nuclear receptor co-repressor. Nature 1995; 377: 397-404.
- 97 Aranda A, Pascual A. Nuclear hormone receptors and gene expression. Physiol Rev 2001; 81: 1269-1304.
- Zhang Y, Dufau ML. Dual mechanisms of regulation of transcription of luteinizing hormone receptor gene by nuclear orphan receptors and histone deacetylase complexes. The Journal of steroid biochemistry and molecular biology 2003; 85: 401-414.
- 99 Yao YL, Yang WM. The metastasis-associated proteins 1 and 2 form distinct protein complexes with histone deacetylase activity. J Biol Chem 2003; 278: 42560-42568.
- 100 Fleischer TC, Yun UJ, Ayer DE. Identification and characterization of three new components of the mSin3A corepressor complex. Mol Cell Biol 2003; 23: 3456-3467.
- 101 Martínez-Iglesias O, Olmeda D, Alonso-Merino E, Gómez-Rey S, González-López AM, Luengo E *et al.* The nuclear corepressor 1 and the thyroid hormone receptor β suppress breast tumor lymphangiogenesis. *Oncotarget* 2016; 7: 78971-78984.
- 102 Kaushik D, Vashistha V, Isharwal S, Sediqe SA, Lin MF. Histone deacetylase inhibitors in castration-resistant prostate cancer: molecular mechanism of action and recent clinical trials. *Ther Adv Urol* 2015; 7: 388-395.
- Hodges-Gallagher L, Valentine CD, Bader SE, Kushner PJ. Inhibition of histone deacetylase enhances the anti-proliferative action of antiestrogens on breast cancer cells and blocks tamoxifen-induced proliferation of uterine cells. *Breast Cancer Res Treat* 2007; 105: 297-309.
- Elbers LP, Kastelein JJ, Sjouke B. Thyroid Hormone Mimetics: the Past, Current Status and Future Challenges. *Curr Atheroscler Rep* 2016; 18: 14.
- Yuan C, Lin JZ, Sieglaff DH, Ayers SD, Denoto-Reynolds F, Baxter JD et al. Identical gene regulation patterns of T3 and selective thyroid hormone receptor modulator GC-1. Endocrinology 2012; 153: 501-511.