#### **Title**

Towards a MicroRNAs-Based Biomarker Panel for AIS: A Meta-Analysis

### Running title

Towards a microRNAs-based biomarker panel for AIS

### **Authors & Affilations**

Nadia Alejandra Rivero-Segura. BSc, PhD. Dirección de Investigación, Instituto Nacional de Geriatría (INGER). Av. Contreras 424, San Jerónimo Lídice, La Magdalena Contreras, 10200 Ciudad de México, CDMX, México. Phone number: +52(1)-55 5573 9087. E-mail: nrivero@inger.gob.mx

**Juan Carlos Gomez-Verjan. Pharm. MSc. PhD.** Dirección de Investigación, Instituto Nacional de Geriatría (INGER), Av. Contreras 424, San Jerónimo Lídice, La Magdalena Contreras, 10200 Ciudad de México, CDMX, México. Phone number: +52(1)-55 5573 9087\_E-mail: <a href="mailto:jverjan@inger.gob.mx">jverjan@inger.gob.mx</a>

Ricardo Mathn Ramírez-Aldana. MSc. PhD. Dirección de Investigación, Instituto Nacional de Geriatría (INGER), Av. Contreras 424, San Jerónimo Lídice, La Magdalena Contreras, 10200 Ciudad de México, CDMX, México. Phone number: +52(1)-55 5573 9087. E-mail: ricardoramirezaldana@gmail.com

### \*Corresponding Author:

Rivero-Segura NA. Dirección de Investigación, Instituto Nacional de Geriatría, Av. Contreras 424, San Jerónimo Lídice, La Magdalena Contreras, 10200 Ciudad de México, CDMX, México. E-mail: nrivero@inger.gob.mx

#### **Conflict of Interest Statement**

The authors declare NO potential conflict of interest.



### Abstract

**Background**: Acute ischemic stroke is among the main causes of mortality worldwide; a rapid and opportune diagnosis is crucial to improve a patient's outcome. MicroRNAs are quite useful for a rapid and accurate diagnosis.

**Methods:** We perform both structural networks approach and a meta-analysis (using a random-effect model to evaluate the heterogeneity and risk bias, according to the PRISMA statement) to analyze the feasibility to develop a microRNA-based biomarker panel for an opportune AIS diagnosis.

**Results:** Structural networks identify a set of eight miRNAs (miR-16, miR-124-3p, miR-484, miR-15a, miR-4454, miR-107, miR-125b-5p and miR-320b) as preliminary microRNA-based biomarker panel, from these only three microRNAs are significantly associated with the main risk factors of AIS, (miR-107: hypertension, 95% CI 9.74-53.24 p<0.0001, type 2 Diabetes mellitus, 95% CI 2.18-19.26); p=0.0008; miR-16 hypertension, 95% CI 1.26-3.56 p=0.0046, smoking, 95% CI 1.07-3.54 p=0.0277; and miR-15a hypertension, 95% CI 1.26-3.56 p=0.0046; smoking, 95% CI 1.07-3.54 p=0.0277). However, the meta-analysis reveals that data is highly heterogeneous and biased; and only microRNAs isolated from plasma samples and further processed in microarrays are the most reliable to distinguish AIS patients.

**Conclusions:** Together our results show that although there are some miRNAs that seem to be associated with AIS, we are still far to develop a miRNA-based biomarker for AIS diagnosis and it is necessary to harmonize the protocols, results and include more populations for further studies otherwise we will remain throwing punches in the dark.

**Keywords:** miRNAs, stroke, acute ischemic stroke, biomarkers, meta-analysis.

#### 1. INTRODUCTION

Brain stroke is a major public health problem worldwide that could be divided into hemorrhagic and ischemic. The latter one in turn could be subdivided into transient stroke and acute ischemic stroke (AIS); which represents the 80-90% of cases all-over the world<sup>1</sup>. Additionally, data from the World Health Organization and from the Institute of Health Metrics and Evaluation, mentioned that AIS is the second cause of morbidity, incapacity, and mortality in individuals over 60 years old<sup>2</sup>.

The AIS results from the permanent local blockage in the arteries that supplies glucose and oxygen into the brain, that required of a rapid evaluation and treatment to achieve best outcomes<sup>3,4</sup>. Both endovascular and thrombolytic (recombinant tissue plasminogen activator, rtPA) therapies helps to the restoration of cerebral blood flow, however, rtPA administration induces symptomatic intracerebral hemorrhage in 3% of AIS-patients<sup>5</sup>, thus, clinicians are very cautious with its administration to avoid health complications. Additionally, AIS diagnosis is quite challenging since it could be confused with other types of stroke that need other type of treatments, this added to other factors such as deficits on the efficient triage of the emergency rooms, expenses and availability of experts, neuroimaging equipment and a scarce education among the general population to identify opportunely stroke, narrow the window of time to manage adequately the AIS<sup>6,7</sup>. Hence AIS diagnosis should be more efficient through the development of non-invasive, cheap, and highly sensitive strategies.

Recently, several studies have been focused on characterizing biomarkers<sup>8</sup> to differentiate among the most common stroke subtypes accurately. The non-coding RNAs such as microRNAs (miRNA's, small single-stranded non-coding RNA molecules from

~22 nucleotides which are endogenously expressed and regulate gene expression though different epigenetic mechanisms) have been proposed as novel biomarkers since they are highly stable and differentially expressed for specific conditions such as cancer, arthritis, osteoporosis, infectious diseases, cardiovascular diseases, neurodegenerative diseases, and AIS<sup>9–11</sup>; miRNA's are easily isolated from different liquid biopsies such as whole blood, plasma, serum, blood circulating exosomes, peripheral blood cells, or cerebrospinal fluid<sup>12</sup> with low invasiveness for the patients, additionally, it they could be easily measured with conventional labs with the minimum requirements on molecular biology<sup>13-17</sup>.

Particularly, the research in AIS offers several studies characterizing miRNA's profiles, which differentiate accurately from other stroke types<sup>18-22</sup>. However, there still a lack of consensus that may constitute a miRNA-based biomarker panel for AIS diagnosis. Therefore, in the present study we performed a structural network analysis followed by a traditional meta-analysis to identify whether among the studies published from 2015 to 2020 is possible to suggest a feasible miRNA-based biomarker panel for AIS diagnosis.

### 2. Methods

# 2.1 Study strategy and Selection

We follow the PRISMA statement<sup>23</sup> to perform this study, methods are submitted to the PROSPERO database with registration number **CRD42020206145**. The studies were explored on the SCOPUS database using MESH terms *microRNAs* AND *mirna* AND

acute ischemic stroke OR brain stroke. All relevant studies from 2015 to 2020 were searched in the Scopus database in July 2020 by two independent reviewers.

Selected studies fulfil the following eligibility criteria:

# 2.1.1 Inclusion criteria:

- 1. Studies reported or published between 2015 and 2020.
- 2. Studies that discuss miRNAs differentially expressed in AIS.
- 3. Studies were presenting both cases and controls groups.
- Studies performed in human samples such as whole blood, serum, plasma, exosomes, or blood cells; and did not exclude studies based on the ethnicity of study participants.
- 5. Only studies that validate AIS diagnosis by neuroimaging, such as computed tomography (CT) or magnetic resonance imaging (MRI), were included.
- 6. Studies conducted within 24 h of AIS symptoms.

# 2.1.2 Exclusion criteria:

- 1. Studies published in other languages different from English.
- 2. Narrative reviews, intervention studies, letter to editors, and non-original articles.
- 3. Unpublished data, incomplete datasets, or preprints.
- 4. Studies without available data.
- 5. Studies without controls.
- 6. Studies using duplicated data.
- 7. Studies performed in vivo.

- 8. Studies performed in vitro, even when these were human-derived.
- 9. Studies performed with already published databases.

# 2.2 Data extraction

We retrieved relevant information from each selected study as depicted in Table 1S from the supplementary material.

# 2.3 Network analysis

Structural networks were built using Cytoscape software v  $3.8^{24}$ . The most connected genes among the network were identify using the Cytohubba plugin<sup>25</sup>. Additionally, we use BinGO plugin of Cytoscape<sup>26</sup> to identify the principal signaling pathways altered by the miRNAs. The most significantly enriched pathways (p < 0.05, p-values) were corrected using the Benjamin-Hochberg procedure, according to previous reports<sup>27</sup>.

#### 2.4 Associations with clinical variables

The association between miRNA expression and the most common risk factors associated with AIS development (HTN, T2DM or smoking) were calculated by the odds ratio (OR) for the differentially expressed (DE) miRNAs that appear shared among tissues or geographical regions. The cases were considered individuals having AIS and one risk factor simultaneously; and the controls were the remaining individuals without AIS. Thus, we considered three types of cases, and consequently, three types of OR are 1) AIS in patients with HTN + DE-miRNA, 2) AIS in patients with T2DM + DE-miRNA, or 3) AIS in patients being active smokers + DE-miRNA.

# 2.5 Meta Analysis

We calculated the heterogeneity using  $\chi^2$  -tests based on the Q-test and the I-squared (I²) statistical tests. The pooled effect size (OR) was assessed based in the random-effect model, if heterogeneity was considered statistically significant (I² -value more than 50% and P < 0.05). To evaluate the specific effects, we also performed subgroup analyses of the data arranged by tissues, geographical origin, and technological platform. The complete meta-analyses of the retrieved data were performed using the *Metafor* R package in R-studio (Version 3.4), according to the methodology previously reported by  $^{28}$ . The complete description of the algorithm is in http://www.metafor-project.org/doku. php/metafor.

# 3. Results

We identify 678 different studies by applying the filters mentioned above. From these, we use 25 studies to construct the networks. According to the PRISMA statement, Figure 1 depicts in detail the article selection for the subsequent analyses.

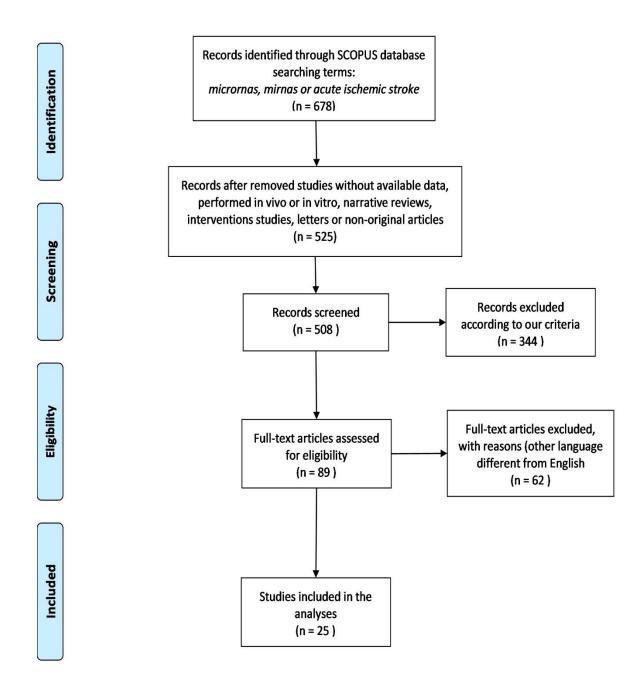
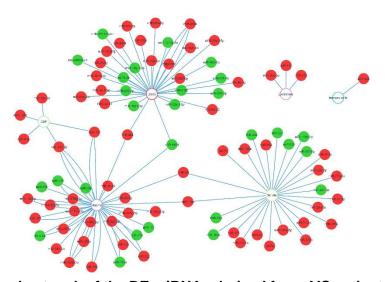


Figure 1. Flow diagram of article selection for the meta-analysis. Modified from <sup>23</sup>

Then we build a network (Figure 2) which demonstrates that despite a significant amount of the miRNAs identified on blood, plasma, CSF, serum, exosomes, and immune cells, only six miRNAs are shared among at least two of them (miR-16, miR-124-3p, miR-484, miR-15a, miR-4454, and miR-107). Interestingly, most miRNAs are upregulated, and plasma is the tissue that shares most of the targets; thus, it could be considered a hub in the network and suggest that plasma may be enriched in miRNAs compared to the other tissues.



**Figure 2. Structural network of the DE miRNAs derived from AIS patients according to sample origin.** This network organizes miRNAs (targets) according to the tissue (source) from which the samples derived (complete data are in Table 1S). DE miRNAs appear in red (upregulated) or green (downregulated) according to their expression; multiple edges indicate the number of independent studies that also report this target. Six miRNAs appear at the network center, representing that these are common in at least two different tissues.

Additionally, we try to answer which miRNAs are shared among the countries where the studies belong, so we built another network that depict such issue. The network from Figure 3 identifies that five miRNAs (miR-125b-5p, miR-320b, miR-124-3p, miR-484, and miR-107) shared among China, Denmark, Germany, and the USA. However, Spain stands apart from the network with the miR-638.

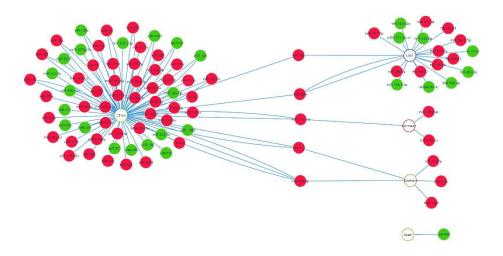
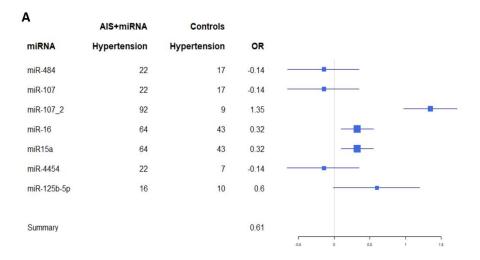


Figure 3. Structural network analysis of miRNAs differentially expressed in AIS from human samples according to the geographical distribution. Data were extracted from 25 selected articles in agreement with our inclusion criteria (Supplementary Information Table 1S). Network was built by using Cytoscape software (v.3.8.0). Nodes correspond to miRNAs (targets-according to its expression those appear in red (upregulated) or green (downregulated)) and countries (sources). Multiple edges indicate different studies joining the same miRNA with the same country.

miRNAs resulting from Figures 2 and 3 were submitted miRNet database (https://www.mirnet.ca/ accessed March 2021) to identify the genes that such miRNAs alter. Then, with the Cytohubba plug-in we identify the most connected nodes (genes) (CREBF, TUBB, CDK6, ABL2, ELK4, GANAB, FBXL18, CALU, CCNE, TNRC6B, and HNRNPA2B1) in the network (Supplementary material Figure 1S). Besides, the gene enrichment using the KEGG, GO:BP, Reactome databases and the BinGO plugin showed that miRNAs in AIS altered genes that are significantly associated to neurotrophin signaling pathway, cell cycle, cell adhesion, protein folding, apoptosis, angiogenesis, aging, oxidative stress, and mitochondrial membrane organization signaling, among others (Supplementary material Figure 2S).

Since HTN, T2DM, dyslipidemias, or being an active smoker are often associated to AIS occurrence, we estimated the association between the selected DE miRNAs identified by the networks in AIS and individuals with has HTN, T2DM, or being an active smoker. The results demonstrate that miR-107, miR-16 and miR-15a overexpression is significantly associated with individuals with both AIS and HTN (OR= 22.77 95% CI 9.74-53.24 p<0.0001, OR=2.12 95% CI 1.26-3.56 p=0.0046, respectively, Figure 4 A-C). Likewise, miR-107 is significantly associated with AIS in T2DN (OR= 6.48 95% CI 2.18-19.26; p=0.00277), and both miR-16 and miR-15a are significantly associated with AIS in active smokers (OR= 1.95 95% CI 1.07 -3.54; p=0.00277), suggesting such miRNAs as potential biomarkers of AIS. Complete data from the association analysis appear in Table-2S.



В

В				
	AIS+miRNA	Controls		
miRNA	T2DM	T2DM	OR	
miR-484	8	3	0.32	
miR-107	8	3	0.32	
miR-107_2	37	4	0.81	
miR-16	38	55	-0.18	
miR15a	38	55	-0.18	-
miR-4454	8	3	0.32	-
miR-125b-5p	4	0	1.04	
Summary			0.53	
				-0.5 0 0.5 1 1.5 2 2.5

C

	AIS+miRNA	Controls	
miRNA	Smoking	Smoking	OR
miR-484	9	9	-0.5
miR-107	9	9	-0.5
miR-107_2	49	17	0.25
miR-16	36	25	0.29
miR15a	36	25	0.29
miR-4454	9	9	-0.5
miR-125b-5p	8	9	-0.09
Summary			-0.01

**Figure 4. Forest plot for miRNA associations with risk factors.** Representative forest plot of the associations (OR) between the eight miRNAs shared among different tissues and geographical regions in individuals experiencing AIS simultaneously and (A)HTN (blue), (B)T2DM (red), or (C) being active smokers (green).

Concomitantly, we aimed to test the potential heterogeneity and the publication bias of the studies, to confirm our results from the network analysis and to suggest a reliable biomarker panel for AIS diagnosis. To achieve such a goal, we performed a meta-analysis with the data retrieved from 25 selected studies. Surprisingly, the results (Figure 5A) shows that there is no statistically significant difference between the DE miRNAS in AIS patients and controls (p-value = 0.0943), suggesting that the overall analysis of the raw data is not useful enough to differentiate between patients with AIS or controls. In addition, these results also show significant heterogeneity (Q= 10588.83, p-value <0.0001), high risk of publishing bias (Figure 5B), and high sampling variance (Figure 5C); probably due to the lack of consensus in protocols focused on characterized miRNAs expression in AIS patients.

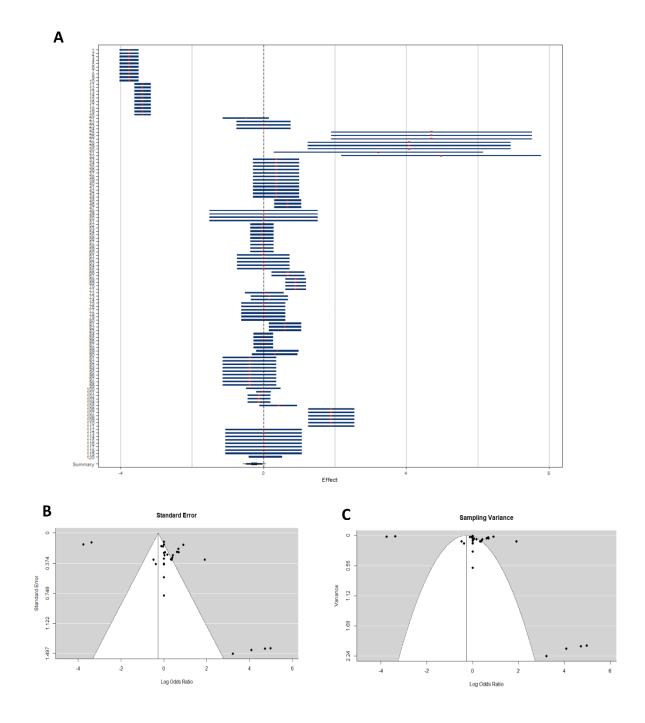


Figure 5. Meta-analysis of all the miRNAs retrieved from the 25 selected studies. (A) Forest plot for the differentially expressed (DE) miRNAs retrieved from the studies summarized in Tables 1S. The scattering shows two groups of miRNAs, ones that are positively associated with AIS and others that are not (associated with the controls, No-AIS). However, globally the results are not statistically significant (p-value 0.0943). (B) Funnel plot for all the DE miRNAs. Each dot represents a miRNA retrieved from the database from Table 1S. The outliers in the grey region represent the miRNAs that have publication bias, since they are out of the significance region (95% CI). (C) Sampling variance plot demonstrates

that the analyzed miRNAs have a significant variability suggesting that there is heterogeneity in the sampling.

Due to the results above, we tested whether meta-analysis among subgroups could be useful for AIS diagnosis. The results show that blood (Q= 1823.75 p-value <0.0001), plasma (Q= 111.92 p-value <0.0001), and serum (Q= 207.91 p-value <0.0001) are a reliable source of miRNAs for AIS in comparison with the data from CSF (p-value=1) and other (immune cells and exosomes, p-value= 0.07). Nevertheless, the results from the funnel plots reveal that except for the plasma-derived samples, the rest of the tissues have bias, probably due to the sampling variance (Figures 3S-5S).

Likewise, when data is analyzed by geographical regions the results demonstrate that only the studies performed in China (p-value <0.0001) and USA (p-value <0.0001) can identify AIS patients from the controls. However, the studies performed in both Europe and USA are significantly biased; in contrast to the data retrieved from the studies performed in China probably due to the small sampling variance (Figures 6S-8S) Finally, our results also suggest that among the available platforms to analyze miRNAs expression, microarrays are significantly useful to identification DE miRNAs in AIS patients (p-value=0.007, Figures 9S-11S).

### 4. Discussion and Conclusions

Despite the great technological advances in neuroimaging, both CT or MRI have limitations such as sensitivity in early stages of AIS, technical issues to view some brain regions, expensive costs, low equipment availability and expertise needed to distinguish among stroke subtypes; that unfortunately are often common in the public health centers,

doi:10.20944/preprints202107.0490.v1

and that delay both the opportune AIS diagnosis and treatment. Hence, developing novel strategies as molecular biomarkers may allow the efficient and accurate diagnosis of AIS. The research on biomarkers for AIS diagnosis or prognosis is an active and dynamic field that have been nourished from several studies worldwide, particularly those based on miRNAs profiles that seems to be a useful tool for the development of a reliable biomarker panel to distinguish among stroke subtypes <sup>29–31</sup>. Also, since the research on this field increases daily a meta-analysis is the best way to move forward and offer a potential biomarker-panel for AIS diagnosis.

At first sight our results show a potential set of miRNAs that may be used for AIS characterization, such miRNAs are involved in biological processes (excitotoxicity, neuronal death, inflammation, neurogenesis, and angiogenesis) often common during AIS <sup>32</sup>. However, when we performed the metanalysis, we found that such panel are poorly feasible since data have high risk of publication and sampling bias. Despite such observation when data were analyze per subgroup we found that that further studies seeking to characterize miRNA-based biomarkers for AIS diagnosis may be conducted in plasma, since this shows the lowest risk of publishing bias of all the analyzed tissues, as previously reported <sup>33</sup>,<sup>34</sup>.

Similarly, when data is analyzed by the geographical, the results show that despite the heterogeny of the data only China exhibits twenty-one miRNAs with both desirable features, they are statistically significant and with the lowest risk of publishing bias, from these miRNAs only five (miR-124-3p, miRNA-125b-5p, miR-107, miR-221-3p, and miR-

16) can distinguish AIS patients from controls. Nevertheless, since the largest number of studies derive from China this could represent a drawback for our analysis, so further analyses are required to validate these miRNAs in other populations.

Since the results from the last meta-analysis demonstrate that microarrays are significantly suitable to discriminate samples derived from AIS patients or from controls and have low risk of publishing bias; this technology may be considered as one of the best options to perform further studies that seek to characterize miRNAs profiles for AIS. These results are probably because most companies use the same databases to build microarrays, and in consequence the variance among the results decreases significantly. Despite these promising results, this technology does not solve the issue that represents the narrow window of time for AIS diagnosis; so, we should delve into other technologies that combine specificity, sensitivity, and efficiency for AIS diagnosis.

Notwithstanding, our study have limitations such as the number of studies included such limitation correspond to the heterogeneity and lack of relevant clinical data, including a complete survey about individual's lifestyles, the heterogeneity in protocols, sample collection, miRNA isolation methodologies, and the platforms used for miRNAs analysis. In addition, the lack of representation of Latin-American population, also contributes somehow to biasing the results. Hence, we suggest that further validation of the proposed miRNAs set is urgently required in other cohorts of AIS patients; likewise, it is necessary to work in harmonized protocols that help to decrease the heterogeneity. Despite such limitations, our study offers a robust perspective about the molecular analysis performed

at date on miRNAs and biomarkers for AIS diagnosis, this study also offers a complete insight about the crucial targets in AIS, providing a new direction for further therapeutic interventions to decrease the consequences of such conditions.

# 5. Acknowledgments

This research received no specific grant from any funding agency in the public, commercial, or not-for-profit sectors.

This review is part of a registered project at Instituto Nacional de Geriatría DI-PI-002-2020 (NARS).

# 6. Supplementary material

Table.1S. Complete data retrieved from the 25 selected studies.

Tables.2S. Complete data from the association analyses.

Fig 1S and 2S. Gene enrichment analysis and Pathway enrichment analysis.

Fig.3S-11S. Complete data from the metanalysis

### 7. References

- 1. Benjamin EJ, Muntner P, Alonso A, et al. Heart Disease and Stroke Statistics-2019 Update: A Report From the American Heart Association. Circulation 2019; 139: e56–e528.
- 2. Zhao H-P, Liu P, Xu C-M, et al. Unique MicroRNAs Signature of Lymphocyte of Yang and Yin Syndromes in Acute Ischemic Stroke Patients. Chinese Journal of Integrative Medicine 2019; 25: 590–597.
- 3. Hu X, De Silva TM, Chen J, et al. Cerebral Vascular Disease and Neurovascular Injury in Ischemic Stroke. Circulation Research 2017; 120: 449–471.

- 4. Fonarow GC, Zhao X, Smith EE, et al. Door-to-needle times for tissue plasminogen activator administration and clinical outcomes in acute ischemic stroke before and after a quality improvement initiative. JAMA 2014; 311: 1632–1640.
- 5. O'Carroll CB, Aguilar MI. Management of Postthrombolysis Hemorrhagic and Orolingual Angioedema Complications. Neurohospitalist 2015; 5: 133–141.
- 6. Gurav SK, Zirpe KG, Wadia RS, et al. Problems and limitations in thrombolysis of acute stroke patients at a tertiary care center. Indian J Crit Care Med 2015; 19: 265–269.
- 7. Gomolka RS, Chrzan RM, Urbanik A, et al. Quantification of image contrast of infarcts on computed tomography scans. Neuroradiol J 2017; 30: 15–22.
- 8. Rivero-Segura NA, Bello-Chavolla OY, Barrera-Vázquez OS, et al. Promising biomarkers of human aging: In search of a multi-omics panel to understand the aging process from a multidimensional perspective. Ageing Res Rev 2020; 64: 101164.
- 9. Maitiseyiti A, Ci H, Fang Q, et al. Identification of Novel Long Noncoding RNAs and Their Role in Abdominal Aortic Aneurysm. Biomed Res Int 2020; 2020: 3502518.
- 10. Beermann J, Piccoli M-T, Viereck J, et al. Non-coding RNAs in Development and Disease: Background, Mechanisms, and Therapeutic Approaches. Physiol Rev 2016; 96: 1297–1325.
- 11. Schulte C, Barwari T, Joshi A, et al. Noncoding RNAs versus Protein Biomarkers in Cardiovascular Disease. Trends Mol Med 2020; 26: 583–596.
- 12. Wijerathne H, Witek MA, Baird AE, et al. Liquid biopsy markers for stroke diagnosis. Expert Rev Mol Diagn 2020; 20: 771–788.
- 13. Kumar S, Vijayan M, Bhatti JS, et al. MicroRNAs as Peripheral Biomarkers in Aging and Age-Related Diseases. Prog Mol Biol Transl Sci 2017; 146: 47–94.

- 14. Oe S, Kimura T, Yamada H. Regulatory non-coding RNAs in nervous system development and disease. Front Biosci 2019; 24: 1203–1240.
- 15. Condrat CE, Thompson DC, Barbu MG, et al. miRNAs as Biomarkers in Disease: Latest Findings Regarding Their Role in Diagnosis and Prognosis. Cells; 9. Epub ahead of print 23 January 2020. DOI: 10.3390/cells9020276.
- 16. Chakraborty C, Sharma AR, Sharma G, et al. Therapeutic advances of miRNAs: A preclinical and clinical update. J Advert Res 2021; 28: 127–138.
- 17. Tiedt S, Dichgans M. Role of Non-Coding RNAs in Stroke. Stroke 2018; 49: 3098–3106.
- 18. Spinetti G, Fortunato O, Caporali A, et al. MicroRNA-15a and microRNA-16 impair human circulating proangiogenic cell functions and are increased in the proangiogenic cells and serum of patients with critical limb ischemia. Circ Res 2013; 112: 335–346.
- 19. Zhou J, Zhang J. Identification of miRNA-21 and miRNA-24 in plasma as potential early stage markers of acute cerebral infarction. Mol Med Rep 2014; 10: 971–976.
- 20. Dhiraj DK, Chrysanthou E, Mallucci GR, et al. miRNAs-19b, -29b-2\* and -339-5p show an early and sustained up-regulation in ischemic models of stroke. PLoS One 2013; 8: e83717.
- 21. Ouyang Y-B, Giffard RG. MicroRNAs affect BCL-2 family proteins in the setting of cerebral ischemia. Neurochem Int 2014; 77: 2–8.
- 22. Li G, Morris-Blanco KC, Lopez MS, et al. Impact of microRNAs on ischemic stroke: From pre- to post-disease. Prog Neurobiol 2018; 163-164: 59–78.
- 23. Moher D, Liberati A, Tetzlaff J, et al. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. BMJ 2009; 339: b2535–b2535.

- 24. Shannon P, Markiel A, Ozier O, et al. Cytoscape: a software environment for integrated models of biomolecular interaction networks. Genome Res 2003; 13: 2498–2504.
- 25. Chin C-H, Chen S-H, Wu H-H, et al. cytoHubba: identifying hub objects and subnetworks from complex interactome. BMC Syst Biol 2014; 8 Suppl 4: S11.
- 26. Maere S, Heymans K, Kuiper M. BiNGO: a Cytoscape plugin to assess overrepresentation of gene ontology categories in biological networks. Bioinformatics 2005; 21: 3448–3449.
- 27. Zhong X, Heinicke F, Rayner S. miRBaseMiner, a tool for investigating miRBase content. RNA Biology 2019; 16: 1534–1546.
- 28. Illescas O, Gomez-Verjan JC, García-Velázquez L, et al. Macrophage Migration Inhibitory Factor -173 G/C Polymorphism: A Global Meta-Analysis across the Disease Spectrum. Front Genet 2018; 9: 55.
- 29. Xu W, Gao L, Zheng J, et al. The Roles of MicroRNAs in Stroke: Possible Therapeutic Targets. Cell Transplant 2018; 27: 1778–1788.
- 30. Khoshnam SE, Winlow W, Farbood Y, et al. Emerging Roles of microRNAs in Ischemic Stroke: As Possible Therapeutic Agents. J Stroke Cerebrovasc Dis 2017; 19: 166–187.
- 31. Kalani MYS, Alsop E, Meechoovet B, et al. Extracellular microRNAs in blood differentiate between ischaemic and haemorrhagic stroke subtypes. J Extracell Vesicles 2020; 9: 1713540.

- 32. Bulygin KV, Beeraka NM, Saitgareeva AR, et al. Can miRNAs Be Considered as Diagnostic and Therapeutic Molecules in Ischemic Stroke Pathogenesis?-Current Status. Int J Mol Sci; 21. Epub ahead of print 14 September 2020. DOI: 10.3390/ijms21186728.
- 33. Bruno DCF, Donatti A, Martin M, et al. Circulating nucleic acids in the plasma and serum as potential biomarkers in neurological disorders. Braz J Med Biol Res 2020; 53: e9881.
- 34. He X-W, Shi Y-H, Liu Y-S, et al. Increased plasma levels of miR-124-3p, miR-125b-5p and miR-192-5p are associated with outcomes in acute ischaemic stroke patients receiving thrombolysis. Atherosclerosis 2019; 289: 36–43.
- 35. Salinas J, Lin H, Aparico HJ, et al. Whole blood microRNA expression associated with stroke: Results from the Framingham Heart Study. PLoS One 2019; 14: e0219261.
- 36. Luque A, Farwati A, Krupinski J, et al. Association between low levels of serum miR-638 and atherosclerotic plaque vulnerability in patients with high-grade carotid stenosis. J Neurosurg 2018; 131: 72–79.
- 37. Tiedt S, Prestel M, Malik R, et al. RNA-Seq Identifies Circulating miR-125a-5p, miR-125b-5p, and miR-143-3p as Potential Biomarkers for Acute Ischemic Stroke. Circ Res 2017; 121: 970–980.
- 38. He X-W, Shi Y-H, Zhao R, et al. Plasma Levels of miR-125b-5p and miR-206 in Acute Ischemic Stroke Patients After Recanalization Treatment: A Prospective Observational Study. J Stroke Cerebrovasc Dis 2019; 28: 1654–1661.
- 39. Rainer TH, Leung LY, Chan CPY, et al. Plasma miR-124-3p and miR-16 concentrations as prognostic markers in acute stroke. Clin Biochem 2016; 49: 663–668.

- 40. Tian C, Li Z, Yang Z, et al. Plasma MicroRNA-16 Is a Biomarker for Diagnosis, Stratification, and Prognosis of Hyperacute Cerebral Infarction. PLoS One 2016; 11: e0166688.
- 41. Yang Z-B, Li T-B, Zhang Z, et al. The Diagnostic Value of Circulating Brain-specific MicroRNAs for Ischemic Stroke. Intern Med 2016; 55: 1279–1286.
- 42. Yuan Y, Kang R, Yu Y, et al. Crosstalk between miRNAs and their regulated genes network in stroke. Sci Rep 2016; 6: 20429.
- 43. Jin F, Xing J. Circulating pro-angiogenic and anti-angiogenic microRNA expressions in patients with acute ischemic stroke and their association with disease severity. Neurol Sci 2017; 38: 2015–2023.
- 44. Jin F, Xing J. Circulating miR-126 and miR-130a levels correlate with lower disease risk, disease severity, and reduced inflammatory cytokine levels in acute ischemic stroke patients. Neurol Sci 2018; 39: 1757–1765.
- 45. Sørensen SS, Nygaard A-B, Carlsen AL, et al. Elevation of brain-enriched miRNAs in cerebrospinal fluid of patients with acute ischemic stroke. Biomark Res 2017; 5: 24.
- 46. Wang Y, Ma Z, Kan P, et al. The Diagnostic Value of Serum miRNA-221-3p, miRNA-382-5p, and miRNA-4271 in Ischemic Stroke. J Stroke Cerebrovasc Dis 2017; 26: 1055–1060.
- 47. Wu J, Fan C-L, Ma L-J, et al. Distinctive expression signatures of serum microRNAs in ischaemic stroke and transient ischaemic attack patients. Thromb Haemost 2017; 117: 992–1001.

- 48. Xiang W, Tian C, Lin J, et al. Plasma let-7i and miR-15a expression are associated with the effect of recombinant tissue plasminogen activator treatment in acute ischemic stroke patients. Thromb Res 2017; 158: 121–125.
- 49. Chen Z, Wang K, Huang J, et al. Upregulated Serum MiR-146b Serves as a Biomarker for Acute Ischemic Stroke. Cell Physiol Biochem 2018; 45: 397–405.
- 50. Cheng X, Kan P, Ma Z, et al. Exploring the potential value of miR-148b-3p, miR-151b and miR-27b-3p as biomarkers in acute ischemic stroke. Biosci Rep; 38. Epub ahead of print 21 December 2018. DOI: 10.1042/BSR20181033.
- 51. Li G, Ma Q, Wang R, et al. Diagnostic and Immunosuppressive Potential of Elevated Mir-424 Levels in Circulating Immune Cells of Ischemic Stroke Patients. Aging Dis 2018; 9: 172–181.
- 52. Wang W, Li D-B, Li R-Y, et al. Diagnosis of Hyperacute and Acute Ischaemic Stroke: The Potential Utility of Exosomal MicroRNA-21-5p and MicroRNA-30a-5p. Cerebrovasc Dis 2018; 45: 204–212.
- 53. Xue Y, Yin P, Li G, et al. Genome-wide Integration Study of Circulating miRNAs and Peripheral Whole-Blood mRNAs of Male Acute Ischemic Stroke Patients. Neuroscience 2018; 380: 27–37.
- 54. Zhou J, Chen L, Chen B, et al. Increased serum exosomal miR-134 expression in the acute ischemic stroke patients. BMC Neurol 2018; 18: 198.
- 55. Huang S, Lv Z, Wen Y, et al. miR-129-2-3p directly targets SYK gene and associates with the risk of ischaemic stroke in a Chinese population. J Cell Mol Med 2019; 23: 167–176.

- 56. Wu J, Du K, Lu X. Elevated expressions of serum miR-15a, miR-16, and miR-17-5p are associated with acute ischemic stroke. Int J Clin Exp Med 2015; 8: 21071–21079.
- 57. Chen Y, Song Y, Huang J, et al. Increased Circulating Exosomal miRNA-223 Is Associated with Acute Ischemic Stroke. Front Neurol 2017; 8: 57.
- 58. Gui Y, Xu Z, Jin T, et al. Using Extracellular Circulating microRNAs to Classify the Etiological Subtypes of Ischemic Stroke. Transl Stroke Res 2019; 10: 352–361.
- 59. Sørensen SS, Nygaard A-B, Nielsen M-Y, et al. miRNA expression profiles in cerebrospinal fluid and blood of patients with acute ischemic stroke. Transl Stroke Res 2014; 5: 711–718.
- 60. Sun M, Hou X, Ren G, et al. Dynamic changes in miR-124 levels in patients with acute cerebral infarction. Int J Neurosci 2019; 129: 649–653.