Review

Synthetic mRNAs; their analogue caps and contribution to disease

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Abstract: The structure of synthetic mRNAs as used in vaccination against cancer and infectious diseases contain specifically designed caps followed by sequences of the 5’ untranslated repeats of β-globin gene. The strategy for successful design of synthetic mRNAs by chemically modifying their caps aims to increase resistance to the enzymatic deccapping complex, offer a higher affinity for binding to the eukaryotic translation initiation factor 4E (elF4E) protein and enforce increased translation of their encoded proteins. However, the cellular homeostasis is finely balanced and obeys to specific laws of thermodynamics conferring balance between complexity and growth rate in evolution. An overwhelming and forced translation even under alarming conditions of the cell during a concurrent viral infection, or when molecular pathways are trying to circumvent precursor events that lead to autoimmunity and cancer, may cause the recipient cells to ignore their differential sensitivities which are essential for keeping normal conditions. The elF4E which is a powerful RNA regulon and a potent oncogene governing cell cycle progression and proliferation at a post-transcriptional level, may then be a great contributor to disease development. The mechanistic target of rapamycin (mTOR) axis manly inhibits the elF4E to proceed with mRNA translation but disturbance in fine balances between mTOR and elF4E action may provide a premature step towards oncogenesis, ignite pre-causal mechanisms of immune deregulation and cause maturation (aging) defects.

Keywords: synthetic mRNA; analogue caps; elF4E; mTORC1; autophagy; immunity deregulation; maturation defects; autoimmunity; cancer.

1. Introduction
Considering the living organisms as open systems maintaining a state far from of equilibrium, they have to obey specific laws of nature that direct intra and intermolecular information [1]. Evolution offers complexity that governs ontogenesis, embryogenesis, growth rate and morphogenesis at low entropy states [1, 2]. In other words, a reduction of entropy offering the macromolecular structures existence equals to an increase of degree of complexity during evolution to give sustainability of life [2]. In contrast to life requirements and unexpectedly, the inter-molecular interactions between synthetic caps of messenger RNAs with yeast [3] and mammalian [4], eukaryotic elongation factor 4E (elF4E), are both entropic and enthalpic, while the enthalpy of inter-molecular reaction rises with the increase in structural complexity of synthetic caps [3, 4].

It is the same natural evolution’s complexity and diversification that has provided the special quality control of mRNA capping to ensure for a) an efficient protein translation b) the stabilization of mRNAs from decomposition and c) the normality of cellular functions [5]. The genetic control due to mRNA caps elevates in complexity with respect to the progress of evolution of species and is mostly evidently complex in eukaryotic mammalian species [5, 6]. The main distinction between the different caps used for mRNA translation in higher eukaryotes is on the differential methylation of 7-methyl-queanosine-ppp (triphosphate linkage)-guanosine -5′ linkage present on the
starting position of mRNAs. At the primary co-transcriptional and co-splicing events within the nucleus, a series of enzymatic activities by polymerase II retain the cap of mammalian mRNA unmethylated at the 2′O position, which is described as the m7GpppG (Cap O) structure [6]. The Cap O has the addition of a methyl group to the N7 amine of the quinine group by guanine N7 methylase, whereas further methylation of the first ribonucleotide at the 2′O position of the ribose by m7G-specific 2′O methyltransferase results to the m7GpppGm (Cap 1) structure [6]. Central to the differential methylation of mRNA caps is the Cap 2′O methyltransferase (CMTR1) enzyme, which adds the methyl group to the first nucleoside of the cap 1 structure [6]. Further methylation by the CMTR2 [7] which adds an extra methyl group at the second nucleotide results in the m7GpppGmNm Cap 2 structure [7]. The enzyme activities of CMTR1 are within the nucleus whereas CMTR2 activities are both within the nucleus and the cytoplasm [7]. Complementary to the capping processes, the cytoplasm’s re-capping is a sophisticated self-preservation process of mRNA inactivation and reactivation regulation system of protein synthesis [5]. The re-capping of mRNAs is a frequent recycling event of mRNAs in mammalian cells [8], it is a highly frequent event in cancer cells [5] and helps to control and stabilize the cellular translation and stability of genome expression being a part of the transcriptome [9]. Complementary to the stabilization of mRNA translation, the multiple mRNA methylations of mRNA caps have evolved as a part of the innate immune defense system [10]. The cap structures and capping processes become more and more specific for cells of different higher eukaryotic species and provide the first alarm for distinction of “self to non self” mRNA recognition under circumstances of cellular evasion of foreign genetic material as in the case of a viral infection, thereby offering essential signaling for interferon responses to recognize and encounter the viral infections [11]. Overall, the cap formations correspond to the regulation of mRNA translation under cell specific differential autonomy variables and signaling specificities. Importantly, the numerous methylation sites on mRNA are associated to pathology conditions [6] and the mRNA caps are differentially methylated by nature’s evolution processes to circumvent cellular homeostatic imbalances [10, 11]. Therefore, the investigation of potential associations between the analogue caps of synthetic mRNA caps, regularly used for the purpose of vaccination enhancement of stability and translation of immunizing proteins, and further the investigation of affiliations due to increased elF4E binding, with potential disease progression due to cellular hyper-sensitivities and cryptic pathologies that may be triggered, as well as the possible molecular pathogenesis pathways associated with clinical conditions are underscored in this review. In this respect, the out of cellular purpose continuation of phosphorylation capacity of the mTORC1 pathway is identified as a possible candidate able to induce autoimmunity, cancer and other related pathologies, and this may arise as a consequence of the stabilization elements present in synthetic mRNAs used in vaccines against infectious diseases and where their use is required on a regular basis and for a large population. Therefore, the possible pathology and clinical implications of pathogenesis induced by an mTOR deregulation due to analogue caps (stability elements of synthetic mRNAs) in vaccines becomes important.

2. The mRNA caps

1.1 The caps used in synthetic mRNA formations for vaccination purposes

The overall design of mRNA genetic vaccine structures can be found in excellent reviews elsewhere [12, 13]. An extensive summary of nature’s mRNA cap formations and synthetic cap structures used in construction of mRNAs in vaccines is presented in table 1. During the almost five-decade history of mRNA vaccine development, which was initially destined to cure cancer [13], the restriction for use of the synthetic mRNAs was
mainly due to their molecular instability, which has been resolved by the sequential experimental progress of the technology of capping. Historically, in the early 1970's, the quanyyltrasferase (GTPase) and the mRNA-quanine-7-methyltrasferase (Mtase) enzymes where isolated from vaccinia virions [14]. GTPase enzyme transfers GMP from GTP to 5' terminus of unmethylated mRNA and sequentially the MTase transfers a methyl group from S-adenosylmethionine to the position 7 of quanosine to form the Cap O structure [5]. However, during the natural cap processes, about one third of mRNA remains uncapped. This is due to the addition of m7GpppG in the reversed orientation so that the m7G nucleotide instead of constituting the cap becomes the first transcribed nucleotide [5]. In fact the researchers Pasquinelli et al 1995 [15] have never observed methylated dinucleotides of m7GpppGm being incorporated in RNA in the normal orientation. Also, the reverse capped mRNAs apart from attracting an antibody response from antibodies specific to the m7GpppG structure [15] are only scarcely translated [8]. The research data on the reverse capped small nuclear RNA (snRNA) reveal a clear-cut deficiency for nuclear import due to the inability of reverse capped RNAs to form RNA – protein (RNP) complexes and for this reason they become hypermethylated to form m-2-2-7G caps; this contrasts the mRNAs that have caps with normal orientation [5, 8, 9, 15]. Further, the reverse capping of RNAs creates a cellular translational disturbance as the snRNAs form dynamic RNA-RNA complexes that catalytically intervene with the normal spliceosomal activities of nuclear pre-mRNA maturation events [16].

The vaccinia capping complex however, although seemingly functional in nature’s processes, has not been used by the industry to overcome capping problems by reverse added orientation due to additional enzymatic step requirements that may have complicated large-scale production [12]. Instead, the discovery of anti-reverse-caps (ARCAs), [17] which are chemically synthesized and added to the 5’ end of mRNA favored their industrial use since they contribute to a 2.3 to 2.6 fold higher protein translation than normal. Indeed, the finding of increase in protein translation has made ever since the ARCA structures the cornerstone of synthetic mRNA production [12, 13]. The ARCA structures are chemically synthesized caps and are made to possess a substitution at 3’–OH of 7’ methyl guanine ribose with hydrogen (H+) or –OCH3 moieties constituting respectively the 7-methyl (3’O-deoxy) GpppG and 7-methyl (3’O-methyl) GpppG capping structures [17]. These chemical adaptations of mRNA caps apart from stabilizing the synthetic mRNA “deceive” the eukaryotic initiation complex and the decapping complex [5] to favorably forward the translation of synthetic mRNAs. Complementary to the ARCAs the synthetic capping technology has devised structures of caps that have insertions of moieties like the imidodiphospate within the tri or tetra-phosphate bridges of ARCA structures to yield the NH, CH2, CCl2 and CF2 analogs [18]. These cap structures show an even more increase in resistance to the enzymatic decapping complex and also a higher binding affinity to eukaryotic translation initiation factor 4E (eIF4E) protein (table 1) [18]. Moreover, the insertions of 1,2-dithiodiphosphate within the phosphate bridges of ARCAs yield the SH analogs which show even higher affinities for binding to the eIF4E [19].

The general principle on the progress of devising caps for synthetic mRNAs focus on the chemical modifications of the caps that will confer, when added to the synthetic mRNAs, a) a decrease of the susceptibility to degradation of the synthetic mRNA by the decapping complex and b) an increase to the binding efficiency of the capped synthetic mRNA to the eIF4E. Furthermore, in order to bypass the cellular translation restriction of 2’O methylation and therefore the rejection of synthetic mRNAs as “non self” mRNA, a 2’-O-methyltransferase capping enzyme is used to methylate ARCA caps of synthetic mRNAs and denote the “self” Cap 1 structure to be recognized adequately by the recipient cellular translation machinery [20]. This may mean however, that even under prob-
lematic conditions of homeostatic imbalance [6, 10, 13] seen during a concurrent viral infection [13], the recipient cells will be forced to translate the synthetic mRNAs due to the capping they possess, even if their requirements for keeping cellular homeostasis are different. Notably, in the natural processes any cap modification has its own physiologic consequences and these arise due to the influence of the affinity of the cap to various cofactors involved in the specific translator machinery of gene regulation [21]. Furthermore, in the higher eukaryotic organization of protein translation, the mRNA capping enzymes have evolved to become sensitized by specific regulatory signals that sequentially correspond to a) their change of behavior in terms of genome expression, b) their differential localization within the compartments of the cell, and c) their divergence of activities [21]. These sensitizations eventually determine the overall gene expression and have a direct impact on the normality of the cell functions and thus on the cell’s fate [21]. In table 1, the naturally existing caps for mRNAs and the chemically modified analogue caps for synthetic mRNAs are listed.

Table 1. The naturally existing caps of mRNAs and the synthesized caps used for mRNA vaccination.

<table>
<thead>
<tr>
<th>A/ The most common natural quanosine methylated mRNA cap modifications and their biological functions</th>
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<tbody>
<tr>
<td>Name of cap modifications and enzymes involved</td>
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<tr>
<td>---------------------------------------------------------------</td>
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<tr>
<td>Cap O: Sequential methylation of the first quanosine nucleotide [5,6]</td>
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<tr>
<td>RNA TPase</td>
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<tr>
<td>GTase</td>
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<tr>
<td>guanine-N7 MTase</td>
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<tr>
<td>Cap N6A: Substitution of first transcribed guanosine nucleotide by adenosine methylated at 6N position [20,21,22]</td>
</tr>
<tr>
<td>Multi component protein complex consisting of catalytic subunit Methyl-</td>
</tr>
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<td></td>
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<tr>
<td>Transferase Like 3 (METTL3)</td>
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<tr>
<td>-----------------------------</td>
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<tr>
<td>Used by selected viruses including Influenza A virus (IAV), HCV, HBV, HIV, Simian virus 40 (CV40), and enterovirus 71</td>
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| Transcription start site (TSS) signaling. |
| Epitranscriptomic gene regulation. |
| Regulation of viral infection and host immune response. |
| Promotion of RNA Decay. |
| Cap independent mRNA translation. |

| Cap 1: Methylation of the +1 ribonucleotide at the 2’O position of the ribose [6] |
| m7G-specific 2’O methyltransferase (2’O MTase) |
| cap methyltransferase 1 (CMTR1) |

| m7G(5’)|ppp(5’)|Gm |
| Lower and higher eukaryotes (mouse and human) |
| Selected viruses |

| Eukaryotes: nuclear co-transcriptional modification |
| Restriction of cap O dependent initiation of translation “as non self” during cellular evasion of foreign mRNA. |
| Promotion of antiviral response by induction of interferon stimulated gene (ISG) proteins |
| Type 1 interferon signaling leads to expression of interferon-induced proteins with tetratricopeptide repeats (IFIT). |
| Binding of IFIT 1 to cap O instead of eIF4E-p attenuates mRNA translation and replication of viruses that do not encode their own 2’O Mtase e.g SARS-CoV, West Nile virus. |
| Viruses: post transcriptional modification at cytoplasm. |
| Evasion of recognition by the innate immune response of the host and interferon response by viruses using a) host mRNA capping pathways (conventional capping) e.g herpesviruses and retro- |
| Cap 2: Methylation of the +2 ribonucleotide at the 2’O position of the ribose [7] | m7G(5’)ppp(5’)GmNm | Nuclear and cytoplasmic |
| | Higher eukaryotes (human) | Independent from N7 methylation of guanosine (Cap O) or from methylation of first nucleotide 2’O ribose (cap1). Greater affiliation for cap2 methylation in cap1 mRNAs, |
| | | Used for efficient pre-mRNA splicing (small nuclear RNAs) |
| | | Promotion of cap 1 dependant mRNA restriction of translation as “non self” against foreign (viral) mRNA. |

| Cap NAD+: 5’ end NAD+ [39] | NAD(5’)pNp | Nuclear during initiation of transcription, ~3-4 fold increase in mRNA stability in bacteria. |
| | Bacteria | Non canonical initiating nucleotide (NCIN)-mediated initiation of transcription |
| | Yeasts | Mammalian cells are equipped with a distinct NAD+ capping mechanism from transcription initiation that does not support cap-dependant translation. |
| | Human | Combination of de-NADing proteins in human control the mRNA decay mechanisms and the gene expression |

CMTR-2
### B/ The synthetic mRNA capping systems

<table>
<thead>
<tr>
<th>Biochemical synthesis pathways</th>
<th>Structure</th>
<th>Biological properties</th>
</tr>
</thead>
</table>
| **5’ terminal mRNA modification by vaccinia virus enzymes [14]**  
  Guanylyl transferase and S adenosylmethionine: mRNA (guanine-7) methyltransferase | G(5’)ppp(5’)Gp  
  G(5’)ppp(5’)Ap  
  under presence of S adenosylmethionine  
  m7G(5’)ppp(5’)Gmp  
  m7G(5’)ppp(5’)Amp | Post-transcriptional modification  
  No sequence specificity apart from terminal purine. Acting by function of poly(A) as a substrate for all methylation reactions. |
| **Anti-reverse cap analogs (ARCA) [17]**  
  Biochemical modifications using pyrophosphate bond formation reactions | 7 methyl(3-deoxy) GpppG  
  7 methyl(3-O-methyl)GpppG  
  7 methyl(3-O-methyl)GpppG  
  m7G(5’)ppp(5’)G  
  m7G(5’)ppp(5’)G  
  m7G(5’)ppp(5’)G  | Inhibit reverse capping of m7GpppGm by bacteriophage polymerases.  
  Increase 2.3-2.6 fold of mRNA translation compared to m7GpppG cap in a rabbit reticulocyte lysate system  
  Increase protein translation in a dose dependant manner in EL4 cancer cells  
  In combination with optimum poly(A) length and β-globin 3’ untranslated regions significantly improve RNA stability in immature dendritic cells (human). |
<p>| <strong>Capping enzyme system and 2’O-methyltrasferase to generate cap 1 [20]</strong> | 7-methylGpppGm | Contribution to enhancement of transgene expression in stimulated T cells (human) in combination with optimized UTRs and poly(A) lengths. |</p>
<table>
<thead>
<tr>
<th>Equivalent in translation efficiency to the ARCA capping system</th>
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<tbody>
<tr>
<td><strong>Imidodiphosphate : NH analogues, [18]</strong></td>
<td><strong>Sequential increase in binding affinity to the elf4E</strong></td>
</tr>
<tr>
<td>Methylenebisphosphonate : CH2 analogues, [18]</td>
<td><strong>Decrease degradation by DcpS (decapping scavenger in exosome) and decapping complex Dcp1–Dcp2</strong></td>
</tr>
<tr>
<td>Dihalogenmethylenebisphosphonate: CCl2 and CF2 analogues, [18]</td>
<td></td>
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<tr>
<td>Biochemical moiety substitution within the 5’,5’-tri- or tetraphosphate bridge of mRNA caps (including ARCAs)</td>
<td></td>
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<tr>
<td>...5’pNHp5’…</td>
<td></td>
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<tr>
<td>...5’pCH2p5’…</td>
<td></td>
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<tr>
<td>...5’pCl2p5’…</td>
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<tr>
<td>...5’pCF2p5’…</td>
<td></td>
</tr>
<tr>
<td><strong>Cap analogues with 1,2-dithiodiphosphate moieties [19]</strong></td>
<td><strong>Dramatic increase in ElF4E-p binding affinity</strong></td>
</tr>
<tr>
<td>Biochemical dithiodiphosphate modified nucleotide synthesis applied to ARCA synthesis 2S analogs: phosphorothioate (O-to-S) substitution inside the triphosphate bridge 2S ARCA analogs: modified by the presence of a 2’-O-methyl group</td>
<td>Decrease of decapping susceptibility by SpDcp1/2 enzyme complex Increasing efficiency of mRNA translation in immature dendritic cells (D1 to D1D2) D1: Gold standard for the treatment of melanoma</td>
</tr>
<tr>
<td>7,2’-O-dimethylGppSpG1D1</td>
<td></td>
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<tr>
<td>7,2’-O-dimethylGppSpG1D1D2</td>
<td></td>
</tr>
<tr>
<td>7,2’-O-dimethylGppSpG1D1D2</td>
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### 2.2. The mRNA capping during normal and oncogenic conditions

The initial isolation and identification of GTPase and Mtase enzymes from vaccinia virus grown in Hela cells [14] was fundamental for the capping technology synthesis and the final manufacturing of mRNAs as vaccines. The identification of these enzymes was probably feasible to achieve due to the regular epigenetic burst vastly present in cancer cells [5]. However, and particularly in Hela cells, the abundant N6-methyladenosine (m6A) modifications of mRNA by m6A methyltransferases induce the expression of Snail mRNA in a cap independent manner [21, 22, 23], as the modifications of mRNA by the m6A methyltransferases constitute a dominant epitranscriptomic level of gene regulation [18]. The switching of the cap dependent to the cap independent mode of translation is often for cells under stress conditions. The reasoning that the cap O and cap 1 structures universally predominate in all cell functions appears to be been misleading for many years as it has been shown by recent advances of NAD+ cap identification and mRNA decay surveillance mechanisms [22]. The frequent presence of O’ 2 methylation on the ribose of the second transcribed nucleotide “the so-called cap 2 formation”, and the first nucleotide, adenosine N-6 methylation, prognosticate for the differential translation efficiency in different cell lineages and under specific stimuli [18,19]. In the experimentation by Holtkamp S et al 2006, quite rightly the authors conclude that an evident enhanced
RNA stability and translational efficiency was devoted to the presence of globin untranslated regions (UTRs) and also to the concurrent presence of a 120-length poly A tail incorporated to the synthetic mRNAs and not to the presence of the cap O structures that had been inserted to the tested synthetic mRNAs [24]. In this experimentation, [24] which is considered as fundamental for the optimization of synthetic mRNA technology, [12, 13] the detection of optimum synthetic mRNA expression was measured by means of antigen presentation on EL4 mouse malignant T-cell lymphoma cells. This can also be a misguidance as these cancer cells are often used in these kind of experimentations because they stably express fluorescent proteins and are thus a useful experimental tool for cancer biology research than may not reflect normal conditions (personal communication). In respect to the protein translation efficiency, the overwhelming research data from investigations of mRNA translation processes that cells follow during normal conditions and conditions of oncogenesis, reveal the presence of differential sensitivities of caps and the elF4E binding protein that further dictate the differential expression of numerous sets of genes. For example, the messenger RNAs by genes required for normal cellular functions like the glyceraldehyde 3-phosphate dehydrogenase (GAPDH), and the β-actin, are less sensitive to elF4E activity as compared to mRNAs from oncogenesis genes involved in cell growth proliferation and immune responses such as the C-MYC, the BCL-2, the vascular endothelial growth factor (VEGF), the cyclins and others [25]. In particular, in nature, the elF4E sensitivity is attributed to the 5’UTR structures located near the caps that dictate the cellular choices for a cap dependent and/or independent mRNA translation rather than the caps themselves [26]. Moreover, the excessive presence of elF4E concentrations above the required threshold during normal conditions is vital for the proper cellular functions and maintenance of homeostasis. Furthermore, during the burst of reactive oxygen species (oxidative stress) which is a hallmark of oncogenesis, the evolutionary guided excess of elF4E protein which otherwise is present ubiquitously during normal conditions is consumed to promote the cancerous cell transformations and the survival of malignant cells [27].

2.3. The elF4E dependence can be provocation to commencement of disease

The elF4E protein has independent roles in the nuclear export of mRNA and its cytoplasmic translation and nevertheless in most cases if not all the elF4E protein expression is a powerful regulon and in most conditions is regarded as a potent oncogene governing cell cycle progression and proliferation at a post-transcriptional level [28]. The unregulated binding affinity of elF4E has profound physiological consequences that can lead to a disease. For example, the elF4E efficient binding to the mRNA caps recruits the elF4G and elF4A proteins to form the elF4F binding complex. Once the elF4F complex becomes intact, the translation of mRNAs, especially those mRNAs that are equipped with the highly stable structures of 5’ UTRs stabilizing the β-globin gene translation [29] and which are fundamentally used in the manufacturing of synthetic mRNAs used for vaccination [24], becomes highly efficient [12, 13, 24]. The increased binding affinity of caps used in synthetic mRNAs [17, 18, 19, 20] and thereafter the increase in translation efficiency of synthetic mRNAs of vaccines means that the elF4E is rendered to become more readily bound to the elF4F complex for a prolonged time than normal and also that may avoid forming complex with the 4E binding proteins (4EBPs). Since the free Elf4E protein is known to attract more easily mRNAs with short unstructured 5’UTRs [26] like a) those from DNA regions encoding for proteins readily involved in angiogenesis, as is the VEGF involved in tumor development, b) those mRNAs translating the cyclin D1 protein which is a significant co-transcription regulator determining the cell cycle progression and tumorigenesis,
and c) those mRNAs translating the BCL-2 proto-oncogene, the chances of over expression of these tumor promoting and enhancing proteins by the prolonged binding of eIF4F complex may become relatively enhanced. This is illustrated in figure 1.

For many important reasons the eIF4E is appraised as an oncogene [31]. It has long been postulated that the activated eIF4F complex induces a deregulation of phosphorylation between eIF4E and the 4E binding protein 1 (4EBP-1) that leads to tumorigenesis [32]. On the other hand, this is the reason that the inhibition of unbound or bound forms of eIF4E on the cap initiation complex (m7GTP–eIF4E interaction), and therefore the inhibition of cap-dependent translation is considered as a potent anti-tumor therapeutic target [32]. It is due to the above that the 4EBPs (1, 2 and 3) sequestering eIF4E have been assessed for their anti-oncogenic potential [31]. However, the abundance of phosphorylations between the mechanistic target of rapamycin (mTOR) protein kinase and 4EBPs causing release of BP-1 from eIF4E do not identify clearly the actual biologic role of the 4EBPs. The data obtained by one 4EBP may not apply to other 4EBP and research investigations accumulate data declaring distinct cell-type specificities with respect to the role of 4EBPs [31]. Although both 4EBP1 and 4EBP2 are modestly decomposed by phosphorylation and are released from the eIF4E binding and set Elf4E free to promote cap dependent initialization of translation, and these are ubiquitously expressed in all mammalian cells, their amounts present are significantly different between organs [32]. For example, the 4EBP-1 is abundant in the skeletal, muscle, adipose tissue and the pancreas, whereas the 4EBP-2 is present in higher amounts in the central nervous system [32]. Remarkably, the activity of p53 proto-oncogene inhibitor activation on the diminishment of protein translation by de-phosphorylation of 4EBP-1 leads to the subsequent attenuation of eIF4E cap binding translation via the mTOR pathway [33].

The over expression and phosphorylation of 4EBP-1 and 4EBP-2 is encountered in many cancers [33] and systemic autoimmunity conditions [34]. As the mTOR kinase is also a major coordinator of the T helper cells differentiation and regulates their cellular fate decisions, the loss of mTOR control and specifically the loss of the mTORC1 dependent pathways can lead to the disorganization of protein synthesis and T cell dysfunction predisposing to immune irregularities [34]. In particular, through the p13/AKT/mTORC1 pathway as illustrated in figure 1, when the 4EBP is over expressed, it becomes hyperphosphorylated and is thus incapable of controlling mRNA expression [31]. On the other hand, the depletion of 4EBP1 expression is oncostatic against tumor progression via the AKT dependent translation pathway [35].

**Figure1.** Oncogenesis via the pathway of serine 209 eIF4E continuous phosphorylation. The eIF4E has long been regarded as a potent oncogenetic factor. The increase of synthetic mRNA cap affinity for eIF4E maintains the eIF4E binding and thereafter the eIF4E in its phosphorylated form ensures prolonged translation. The mTOR1 pathway keeps on the capacity to phosphorylate eIF4E. The maintenance of eIF4E in its phosphorylated form drives MYC and AKT activation pathways though ISR GCN2 sensing a) protruding autoimmunity and oncogenesis, and b) progressing cellular proliferation and cancer aggressiveness. Adapted from references [30-36].
2.4. The eIF4E serine 209 phosphorylation is important for disease onset

Whilst the 4EBPs are phosphorylated by the mTORC1 kinases, the eIF4E is phosphorylated in the irreplaceable site of serine 209 (human sequence) by the bonded mitogen actine protein kinases (MARK), kinases MNK1 and MNK2 [31]. This double way of 4EBP and eIF4E phosphorylation results in the dissociation of phosphorylated 4EBP thereon called p-4EBP, from the 4EBP-eIF4E complex and thus the initiation of translation by the phosphorylated eIF4E (p-ELF4E) and the assembly of the eIF4F complex is triggered [31-33].

So far, the actual role of serine 209 phosphorylation of eIF4E remained controversial [31]. However, recent experimental evidence supports the notion of p-eIF4E dependent translation in MYC and ATF 4 driven oncogenic initiation progression and aggressiveness of cancer in a rate-limited manner [36]. Latest scientific evidence [29], suggests that the p-eIF4E maintains the 4EBP-eIF4E binding [31] (figure 1), and by using the most ancient arm of integrated stress response (ISR) in mammals and the general control of nonderepressible 2 (GCN2), the p-ElF4E maintains the AKT/4EBP-1 signaling and the stress response by ATF4 transcription factor. In this process, the mTORC1 activity is maintained to phosphorylate 4EBP-1 and thus the anti-oncogenic potential of mTOR silencing is inactivated [27]. In this respect, any excess of cellular mRNA translation, as is in the case of overloaded synthetic capped mRNAs in vaccines, and especially when present in initiative and progressive conditions of autoimmunity [34] and oncogenesis [35, 36], the surplus drive of the p-elF4E-cap-dependant translation is far more than desirable for the maintenance of cellular homeostasis. It must be re-emphasized that synthetic mRNAs are designed to have solid capping structures and UTRs to ensure for efficient and long-lasting translation [10, 12, 13, 17-19]. Thus, an elevated cap-dependent protein translation under circumstances of PI3K/AKT pathway activation which causes MYC and ELF4E genome amplifications [37], and active translation of synthetic mRNA vaccine bearing caps with increased affinity to eLF4E than normal [10-13, 17, 18, 20], may drive even more the potential of cells for autoimmunity and oncogenesis. This is especially more important as the mammalian species have evolved mechanisms of eIF4E surplus in the cell for their normal development [27]. For example, the translation of ferritin heavy chain 1 (Fth1) is highly sensitive to eIF4E expression levels in a dose depended manner [27]. Moreover the synthetic mRNAs are modified structurally to carry the 5’ untranslated
repeats (URTs) of β-globin gene in order to confer translational efficiency and stability [12, 13, 17, 20, 24]. The above normal limits regular presence of 5’ UTRs and the overwhelming attractiveness by the long lasting presence of p-eIF4E through the regular binding of iron-responsive element binding protein / iron regulatory protein (IRE/IRP) regulatory network, could also possibly drive the activation of nuclear erythroid-derived 2-like 2 (NRF2) transcription factor targeting the gene of FTH1 [38, 40, 41]. According to specific cellular vulnerabilities [38]), this may also cause a disturbance in the cellular iron availability and in the regulation of ferroptosis of the cells that are induced to prolong translation by the synthetic mRNAs as illustrated in figure 1.

3. The role of mTOR

3.1. The origin of mTOR and its links to disease

The mechanistic target of rapamycin, or otherwise mTOR, is a kinase enzyme namely, the FK506-binding protein 12-rapamycin-associated protein 1 (FRAP1) [42]. The mTOR is a pivotal regulator for cell growth, metabolism, and aging, firstly identified in yeasts in 1991 [42-44]. At the cellular level, mTOR is involved in a) the regulation of transcription and translation, b) the control and stability of a plethora of mRNA transcripts, and c) the actin cytoskeletal organization, all activities by performing epigenetic regulation through regulation of autophagy [45-48]. In the yeast cells, there are two mTORs, namely the Tor1p (tor1), and the Torp2 (tor2), which are the mammalian orthologs of mTOR1 and mTOR2 [43]. The tor1 assembles solely into TORC1 and tor2 assembles preferentially to the TORC2 large protein complexes. These complexes are also conserved in mammalian species and human. The mTORC1 complex contains both tor1 and tor2. The mTOR as a whole structure, interacts with diverse cell signaling pathways, and stimulate protein synthesis through phosphorylation of crucial translation regulators, such as the ribosomal S6 kinase which is the major protein S6 kinase (S6K) in mammalians [46, 47, 49], and the 4EBPs (both 1, 2, and 3 4EBPs). The phosphorylation of S6K reinforces the mRNA translation. Furthermore, as the ribosomal proteins and other regulators related to the reinforced translation by the S6K also regulate the encoding of related genes of these translated mRNAs, the whole process constitutes a signaling cascade [49, 50]. The other important targets of mTOR are the 4EBPs, and specifically the 4EBP1, which binds to and inhibits the EIF4E from binding to the 5’end caps of eukaryotic mRNA transcripts as illustrated in figure 1 [51]. The phosphorylation of 4EBP1 by mTOR subsequently results in the separation of the 4EBP1 from the eIF4E [51]. As a result, the surplus of liberated elf4E released from binding to the 4EBP1 enhances the translation of mRNAs (in a rate-limited manner) of the key growth-promoting proteins, the cyclin D1, the c-Myc, and the VEGF as illustrated in figure 1 [30]. The regulation of cell growth is mediated by mTOR through two of the key extracellular and intracellular signaling factors a) the growth factors (insulin and insulin-like) and b) the nutrients such as amino acids and glucose [52, 53].

The dysfunction and the deregulated signaling of mTORs are implicated in metabolic, neurodegenerative, and inflammatory disorders and malignancy [44, 45]. In general, the mTOR deregulation is strongly associated with tumorigenesis. As mTOR inhibits autophagy under normal cellular conditions, its deregulation increases cell proliferation instead of driving cells to normal death as it is illustrated in figure 2 [43]. By the loss of function of autophagy regulation by mTOR (where by inhibition of mTOR the autophagy is induced) [54], and once the autophagic cell death is reduced, the activation of onco-genes is then enhanced [39]. The induction of autophagic cell death (or otherwise called as the type 2 cell death) is tightly connected to: a) the expression of Beclin-1 tumor sup-
pressor gene (present in many cancers), b) the induction of death-associated protein kinase (DAPK) expression (which otherwise is oncostatic and is being reduced by the restriction of autophagy), and c) the expression of the dual phosphatase and tensin homolog deleted on chromosome 10 (PTEN), which is also downregulated during the inhibition of autophagy [54, 55]. Nevertheless, another important enzymatic activity comes from the domain of the death-associated protein kinase 1 (DAPK1) which is a member of the family of serine/threonine kinases. The association of DAPK1 with the tuberous sclerosis 2 (TSC2) interferes with the binding between TSC1 and TCS2 by inhibiting TSC2 and thus the ability of TSC1 to halt the signaling activity of mTOR is abolished [56], as illustrated in figure 2. On the other hand however, the DAPK induces phosphorylation within the bona fide 3 (BH3) domain of Beclin1 protein and thus promotes the dissociation of Beclin-1 from Bcl-2 and its inhibitor of the BclXL a case that leads to the induction of autophagy [57] as illustrated in figure 2. Although at present, the knowledge about mTOR and the Beclin-1 and/or DAPK, and the loss of autophagy regulation is limited [56], it is clinically important to consider the deregulation of mTOR to autophagy and its implications in the pathogenesis of a series of tumors and neurodegenerative disorders as a result of an external stimulus. For example, it is strongly hypothesized that the Huntington disease and the spinocerebellar ataxias are related to the mTOR activity without having yet a clear mechanism been identified [43].

Figure 2. The pathways where mTORC1 can remain activated and can predispose to autoimmunity and cancer. Adapted from references [39, 43, 53-57].

3.2. Hamartomas and mTOR deregulation

Hamartomas are mainly benign tumors caused by mutations in several tumor-suppressor genes and can provide a fine example of mTOR deregulation. The hamartoma syndromes are divided into two categories [43]. The first category is directly associated with the dysregulation of mTOR and the second category, which is clinically similar to the first, has also its eventual causality and pathogenesis attributed to the mTOR deregulation [43]. The tuberous sclerosis complex (TSC), the PTEN-related hamartoma and the Peutz-Jeghers syndromes belong in the first category. Moreover, the Lhermitte-Duclos disease, the Proteus syndrome, the Cowden disease and the Bannayan-Riley-Ruvalcaba
syndrome are all autosomal-dominant benign tumors linked to the TSC pathway of pathogenesis. The tumor-suppressor gene of phosphatidylinositol 3,4,5-trisphosphate 3-phosphatase (PTEN) is mutated in all of these four pathologies. Furthermore, the already mentioned TSC1 and TSC2 protein complexes are the signaling regulators for the expression of mTOR. When these dysfunction or loose activity and/or when the over-expression of mTOR coincides, these are the key pathogenesis events that lead to TSC disease [58, 59]. The Neurofibromatosis type 1 (NF1) disease leads to malignancy when the rat sarcoma (RAS) suppressor gene NF1 that encodes neurofibromin is homozygote mutated [60]. The RAS hyperactivation activates subsequently the AKT/mTOR pathway along with the Raf/MEK/ERK pathway leading to inhibition of autophagy, increase of cellular proliferation, and subsequent development of the NF1 syndrome [61, 62]. The Von Hippel-Lindau syndrome (VHL) causes multiple tumors in numerous organs. Mutations of the Von Hippel-Lindau Tumor suppressor gene cause loss of the products that otherwise inhibit the ubiquitylation and the proteasomal degradation of the hypoxia-inducible factor-1-α (HIF-1α) [63], a case that involves numerous nuclear translocation and intranuclear hypoxia dependent signals. As mTOR is an active positive regulator of HIF gene products it is also strongly implicated in the causality of VHL syndrome [43, 64, 65].

The representative syndromes that belong to the second category of hamartomas are the family adenomatous polyposis (FAP) and juvenile polyposis syndrome (JPS). FAP is caused by germline mutations in the adenomatous polyposis coli (APC) gene that results in the triggering of upper and lower gastrointestinal polyps and carcinomas, hepatocellular carcinoma and hepatoblastoma (less frequently) [66]. Further, the activation of β-catenin signaling is linked to the loss of APC function that leads to promotion of cellular survival and inhibition of cell death [66]. The β-catenin works through the Wnt/β-catenin pathway to promote cancer [67]. This is important for mTORC1 deregulation since the PI3K/AKT/mTORC1 pathway works closely with the Wnt/β-catenin pathway regulating one another (thus constituting a common therapeutic target against the colorectal cancers that they produce), and therefore the activating signals of mTOR are considered as predisposing conditions for FAP development [43, 67, 68]. The JPS which is also characterized by multiple polyps along the gastrointestinal tract has predominantly (to a 50% – 60% of patients), germline mutations in the SMAD4 or BMPR1A genes [69]. Although mTOR has not yet been identified as a contributor to the development of cancer from JPS mutations, there is interlinking evidence that Bmpr1a phosphorylation regulates Smad activation in response to Bmp and associatively the Bmp activates the cellular differentiation of osteoblast through Smad4 and mTORC1 pathways [70].

3.3. The mTOR and the immune system regulation

The mTOR has a central role in the mammalian immune response and metabolism [71]. The mTOR activity, by connecting the extracellular with the intracellular environment, contributes to the adaptive immune system regulation, having a main role in the cell differentiation, maturation, migration, and antigen-presentation [71, 72]. The metabolic pathways are strongly related to the immune pathways, and mTOR is a regulator in various immune lineages such as the mast cells, neutrophils, natural killer cells, γδ T cells, macrophages, dendritic cells (DCs), T and B cells. The upstream and downstream diverse signaling of mTOR, regulates and interact with numerous key factors such as growth factors and nutrients [45].

The mTORC1 signaling enhancement induces the T cell activation and reversibly the mTORC1 inhibition during immune response, which promotes T cell anergy. This pathway is essential for performing cancer immunotherapy since tumors evade the im-
The immune surveillance defense mechanisms and in this respect, the activation of mTORC1, can constitute a target for anticancer therapy [73]. In addition, mTORC1 is involved in the T cell maturation. The research teams of Pollizzi et al., 2016 and Verbist et al., 2016 provide scientific evidence showing that the mTORC1 activity is elevated in the “effector-like” daughter cells and is reduced in the “memory-like” daughter cells. [74, 75]. The class I Phosphoinositide-3 kinases (PI3K) are important cellular signaling mediators inducing cell growth and proliferation with the main representative being the p110α protein which is involved actively in oncogenesis [76]. Notably, as the cytokine and the growth factor receptors induce the activation of mTOR, the CD28 precisely promotes the activation of PI3K and subsequently the mTORC1 in T cells [68, 71]. Also in this respect, the interleukin 2 and 4 (IL2 and IL4), activate the mTORC1 pathway through the activation of PI3K [71, 77, 78, 79]. Moreover, the stimulation of mTOR to produce IL-12 and IFN-γ prolong further the activation of mTOR in CD8+ T cells whereas the production of IL-1 induces the generation of Th17 type of immune response through the stimulation of mTOR [71, 80]. The Th17 T cells are a distinct subtype of CD 4+ cells holding a unique type of immune response, the Th17 type of immune response, which holds a central role in contributing to the pathogenesis of autoimmune and inflammatory disorders [71, 77, 81].

3.4 The Wnt signaling pathway, the mTOR, and the immune regulation

The Wnt (name that arises from the name of Drosophila gene wingless and the name of a vertebrate homolog gene int-1), constitutes an evolutionary ancient signaling pathway formatted by growth stimulated proteins and encoded by the highly conserved wnt gene from lower vertebrates to human [82]. Wnt is a fundamental signaling pathway that has, as protagonists, a plethora of secreted glycoproteins that follow complex cascades to determine the cells fate, migration, polarity, and organogenesis in an early embryonic life [78, 82]. When the wnt signaling pathway works in close relation with the β-catenin protein, it is termed as the wnt/β-catenin dependent pathway, or otherwise when β-catenin protein is not involved it is termed as the wnt/β-catenin independent pathway. However, there are yet more wnt signaling pathways not associated with or without the β-catenin and mTOR seems to be implicated in at least one of them [53]. As long as the TCS1/TSC2 tumor suppressing complex remains intact, the mTORC1 remains inactive and protein synthesis remains halted (figure 2). However, there are cases, where, and through wnt inactivation of GSK3β kinase (that regulates the silencing of the mTORC1 activity especially under conditions of energy deprivation and hypoxia), the GSK3β inhibition is bypassed and wnt pathway activates mTORC1 independently from the kinase Akt and RSK inhibitory activities. These cases constitute an important step leading to many cancers and neoplasms directly via the wnt activation of mTORC1 as illustrated in figure 2 [83]. Notably, the mTOR is also activated potently by amino acid induction, independently from the wnt pathway [53].

Under the presence of suitable conditions such as under presence of oxygen, amino acids, growth factors and lipids, the mTOR remains in its active form and induces translation, lipid synthesis, and mitochondrial biogenesis [51, 58]. However, by the lack of these conditions, the mTOR signaling is inhibited and thereby autophagy is enhanced, creating a feedback loop. The conversion of a resting T cell to an active T cell demands nutrients and active glycolysis reactions. This activation pathway is potently related to the Akt/PI3K axis. Respectively, the CD28 promotes the PI3K activation resulting to Akt activation, thereby resulting to the inhibition of lymphocyte metabolism that inhibits in turn the T cells from functioning properly [84]. Nevertheless, CD28 is known as the major stimulation pathway for naïve T-cell activation, and the CD28/B7 co-stimulatory complex
is mostly relevant in the combat of immune defense against clinically challenging conditions [84]. During CD28 dependent co-stimulatory signaling, the Akt kinase is repressed since the levels of ATP are high and hence the low levels of AMP do not allow AMP kinase to phosphorylate TSC2 and thereafter repress mTORC1 as illustrated in figure 2 [84, 85].

In many ways the role of mTOR in T cell differentiation and in relation to CD28 co-stimulatory signaling remains problematic [85]. However, the CD28 dependent pathway is synergizing with the T cell receptor (TCR) and the IL-2 synthesis and the inhibition of CD28 and IL2b result in low mTORC1 activity, ineffective clonal differentiation, and cellular anergy [55]. Importantly, the mTOR-deficient T cells can not differentiate to Th1, Th7, or Th2 effector cells in vivo or in vitro under potent polarization [75]. The environmental factors in this respect play a central role in the activation of mTOR and consequent metabolic and immunologic activation that lead to T cell differentiation (as the mTOR controls numerous transcription factors that regulate T cell differentiation by antigen recognition) that become important [86]. The mTOR phosphorylation through ribosomal S6K [87] is induced by antigen recognition in CD8+ T cells and through the MAP-kinase signaling. Complementary, the development of memory cells is depended on the expression of IL-7 (CD127) and IL-2 receptor beta chain (CD122) during a contraction phase [88]. These CD8+ T cells act as effectors and mTOR contributes to the generation of these stimulators. In this way, the inhibition of mTOR activity leads to the loss of CD8+ memory T cells activities. As the above activities of mTOR (which are related to various factors) show the dependence of numerous immune cell functions to the mTOR activation or inhibition, the destabilization of normality functions of mTOR may lead to immune dysregulation. In most senses, when the molecular breaks of mTOR activity are deregulated, the orchestra of the immune system contributors is deregulated too, leading the immune cells to become prone in developing reactions that may lead to inflammation, autoimmunity, and tumorigenesis as illustrated in figure 1.

3.5. Other clinically important implications arising by mTOR improper signaling

The role of mTOR in the normal brain’s functioning is essential since mTOR proper functioning contributes to the development and feeding of the brain neurons and the circuit formation. The mTORC1 activity inducing the mRNA translation near the synapses of brain neurons is vital for the neuronal system development. Notably, the deregulation and overexpression of mTORC1 is high in patients with TSC and this is accompanied by epilepsy (90%), autistic features (50%), and benign brain tumors [89].

As autophagy is interlinked with induction of the mTORC1 signaling [43, 44], mTORC1 plays also a key role in the glucose homeostasis in the liver. Specifically in the fasting periods when the glucose intake is deprived, mTORC1 is under functioning due to the inhibition of AMP kinase dependent and independent pathways [90]. Moreover, not unexpectedly, the mTORC1 is involved in the control of hepatic ketone body accumulation. During low glucose presence, the mTORC1 activity is reduced and so ketogenesis becomes under functioning. However, when mTORC1 signaling is persistently activated, as for example in mice with an activated TSC1 due to persistent mTORC1-dependent suppression of peroxisome proliferator-activated receptor alpha (PPAR-α), autophagy is impeded, and as a consequence a) amino acids are not produced and b) gluconeogenesis is blocked. This failed response to gluconeogenesis due to mTORC1 deregulation results in fatal hypoglycemia [91]. Furthermore mTOR, activity is vital for the β-cells of Langherans islets in the endocrine region of pancreas, as mTOR is a central positive regulator of β-cell growth, proliferation and function [46, 47]. In mice, the mTORC1 deregulation contributes to reduced blood glucose levels, induces glucose intolerance and therefore
hyperinsulinemia. This subsequently induces the constitutive increase of the mass and number of β-cells (increase in their proliferation and production). In this regard, mTORC1 is a regulator of β-cells mass and function. Further, the action of mTORC1 in mice is dependent on the S6K1 and by the absence of S6K1 the size of β-cells becomes smaller, the cells become glucose-intolerant with an increased insulin resistance due to an impaired insulin secretion. By the chronic insulin resistance, the pancreatic β-cells react with a compensating mechanism. The constitutive insulin resistance elevates the pressure in β-cells of Langerhans islet and after a period of time the β-cells become overloaded. This compensates with the observed β-cell hypertrophy, the increased β-cell proliferation, and subsequent formation of progenitor cells due to the attempt of compensation and due to the increased demands of insulin production and secretion. The aforementioned condition, if not reversible in a given time, leads to type 2 diabetes condition due to the steady insulin resistance. In the mice being genetically obese or in the high-fat-fed mice, mTORC1 activity is increased. The axis mTORC1/S6K1 has a key signaling role in the regulation of glucose homeostasis. If this axis remains activated over a time threshold, the pancreatic β-cells lose their mass through apoptosis and due to the feedback inhibition of IRS1 and IRS2, which promotes cell-death. Because of the sustained cellular apoptosis, the pancreas looses β-cells, a condition leading to β-cell compensated hypertrophy and elevated insulin resistance, hypoinsulinemia and hyperglycemia [47, 48]. In this regard, the known antidiabetic drug metformin, can improve the glucose homeostasis through the partial inhibition of mTORC1 signaling.

The neurodegenerative disorders a) Parkinson disease, b) multiple sclerosis, and c) Alzheimer disease are also associated with the hyperactivity of mTORC1 [45]. A central shared pathway of this disease group is the accumulation of toxic proteins, the so-called inclusion phenomenon, which constitutes the failure of neurons to clear mutated or misfolded proteins that subsequently causes cell death. The autophagy and the ubiquitin-proteasome system, which are avoided due to the mTORC1 hyperactivity, are the main pathways in the intracellular protein degradation system. Centrally, the defects associated with the autophagy activation contribute to the pathogenesis of neurodegenerative disorders in most cases. For example, another pathological pathway of neurodegeneration observed in mice, is the deletions in autophagy genes Atg5 and/or Atg7, which lead to the aggregation of polyubiquitinated proteins without a necessary presence of any disease associated protein mutation [92, 93]. In this regard, autophagy is a pivotal survival mechanism of the neurons and its impairment results in neurodegeneration. In most sense, the normal functioning of mTORC1 is fundamental for the neural homeostatic balance.

Research studies on the role of rapamycin as an inhibitor of mTORC1 activity have shown that the inhibition of mTORC1 impairs cellular aging and subsequently prevents age-related diseases such as cancer, neurogenerative disorders, type 2 diabetes and stem cell dysfunction pathologies. Therefore, the mTORC1 is also regarded as key negative regulator of cellular lifespan and its false signaling activation causes aging pathologies. In aging mice, the mTORC1 signaling pathway is elevated in the hematopoietic stem cells (HSC) [94] and the activation of mTORC1 by Tsc1 loss causes premature aging in HSC. Furthermore, the inhibition of mTORC1 signaling, for instance through rapamycin targeting, promotes HSC renewal and improves hematopoietic function, by improving the longevity of cells [95, 96]. Finally, the mTORC1-S6K1 and mTORC1-4E-BP1 axes have an important role in the signaling pathways which can cause a diverse range of pathogenesises. In this respect, the inhibition of these two axes can possibly expand the life expectancy of cells [97]. The mTORC1 may also activate complementary mechanisms without having an association with protein synthesis. Nevertheless, the autophagy activation linked to mTORC1 inhibition, may contribute to some of the effects of cellular ex-
tension of life span. Notably, as the mTORC1-4E-BP1 axis mainly inhibits the eIF4E to proceed with mRNA translation, defaults in fine balances between mTOR and eIF4 action may constitute a premature step of oncogenesis and ignite pre-causal mechanisms that can lead to stem cell related disorders and aging defects [95-97].

5. Conclusions

The deregulation of phosphorylation between eIF4E and the 4EBP-1 leads to tumorigenesis. Although for many years the role of serine 209 phosphorylation of eIF4E remained controversial, it has been recently elicited that p-eIF4E supports the notion of mTORC1 maintenance of activation by using the most ancient arm of integrated stress response (ISR) in mammals, the general control of nonderepressible 2 proteins. Therefore, there may be cases where the excess cellular mRNA translation, as in the case of overloaded synthetic capped mRNAs in vaccines bearing robust analogue caps, and especially when present in initiate and progressive conditions of autoimmunity and oncogenesis, that can be asserted as a predisposing factor for disease. The synthetic mRNAs used for vaccination against cancer and infectious disease are optimized by their caps to stabilize and increase the translation of their encoded proteins in order to immunize better by producing their encoded proteins. Their caps however create an increase of eIF4E binding dependence for fast translation and divergence from natural cellular decomposition events. This seems to be against the natural laws of thermodynamics offering the optimization of structural complexity of macromolecule interactions in evolution when measured in their entropy and enthalpy released. The enthalpic increase and entropic change between synthetic cap interactions with mammalian eIF4E as well as with the eIF4E of lower eukaryotic species may be in contrast to the elevation of complexity of living organisms in terms of growth rate requirements and compatibility with health (and life) [1-4]. Our analysis reflects the potentiality to disease onsets by eIF4E and mTOR deregulations. The internal variables of a living organism trying to keep the internal state unchangeable (homeostasis) may be disturbed by the analogue caps of synthetic mRNAs and the eIF4E interactions to an extend that has to be analyzed by further superior physical chemistry, biochemistry and explicit toxicity evaluation research. Particularly during sensitive circumstances, as during deregulation of fine balance of cellular homeostasis (conditions of mTORC1 deregulation that is primarily given as an example in our analysis) and as of this consequence, due to the autophagy deregulation, this can cause immune dysfunction irregularities, cellular maturation incompatibilities and predispose to various autoimmune disorders and malignancies. Foremost the loss of cap regulating innate defense of cells may prone the organism to susceptibility of further viral and bacterial infections.

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