
Article

The Crosstalk of Melatonin and Hydrogen Sulfide Determines Photosynthetic Performance by Regulation of Carbohydrate Metabolism in Wheat under Heat Stress

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Abstract: Photosynthesis is a pivotal process that determines the synthesis of carbohydrates required for sustaining growth under normal or stress situation. Stress exposure reduces the photosynthetic potential owing to the excess synthesis of reactive oxygen species that disturb the proper functioning of photosynthetic apparatus. This decreased photosynthesis is associated with disturbances in carbohydrate metabolism resulting in reduced growth under stress. We evaluated the importance of melatonin in reducing heat stress-induced severity in wheat plants (*Triticum aestivum* L.). The plants were subjected to 25 °C (optimum temperature) or 40 °C (heat stress) for 15 days at 6 hours time duration and then developed the plants for 30 days. Heat stress led to oxidative stress with increased production of TBARS and H₂O₂ content and reduced accrual of total soluble sugars, starch and carbohydrate metabolism enzymes which are reflected in reduced photosynthesis. Application of melatonin not only reduced oxidative stress through lowering TBARS and H₂O₂ content, through augmenting the activity of antioxidative enzymes but also increased the photosynthesis in plant and carbohydrate metabolism that is needed to provide energy and carbon skeleton to the developing plant under stress. However, the increase in these parameters with melatonin was mediated via hydrogen sulfide (H₂S), as the inhibition of H₂S by hypotaurine (HT; H₂S inhibitor) reversed the ameliorative effect of melatonin. This suggests a crosstalk of melatonin and H₂S in protecting heat stress-induced photosynthetic inhibition via regulation of carbohydrate metabolism.

Keywords: Carbohydrate; heat stress; hydrogen sulphide; hypotaurine; melatonin

1. Introduction

Heat stress is an emerging issue due to global rise in temperature which will adversely affect plants' growth, photosynthesis, crop production and sustainability. Hassan et al. [1] reported that heat stress harshly impacted the growth and quality of plant, ecosystem as well as food security. The expansion in the world's temperature because of anthropogenic activities is the significant worry for mankind [2], as this will lead to major losses in crop production worldwide [3]. The photosynthetic ability of plants gets

disturbed by the enhanced production of reactive oxygen species (ROS) under high temperature [4]. ROS adversely affect enzyme activity and membrane fluidity, resulting in altered growth and development of plants and yield loss [1]. The plants exposed to heat stress exhibit altered carbohydrate metabolism through modulation in expression of the involved genes. Transgenic plants with higher levels of sugars/ sugar alcohols like mannitol, fructans, trehalose and raffinose are tolerant to different stresses [5]. Plants accelerate inherent different strategies for better stress tolerance. More recently the role of melatonin in heat stress tolerance has been explored [6-9].

Melatonin is a pleiotropic signal molecule that is known to maintain balance between the different physiological processes and protection of plants besides numerous abiotic stresses responses [10,11] both through endogenous manipulation or exogenous application [8,12-14]. It works as an antioxidative molecule to scavenge ROS [15] and protects photosynthesis [16]. It is referred to work by means of an antioxidant that regulates abiotic stress tolerance [14]. Melatonin changes the cell layer penetrability arbitrated through ion transporters, which regulates the stomatal opening and closing in plants. Melatonin improves photosynthetic capacity of plants and upgrades the photosystem (PS) I and photosystem II, incites higher nitrogen and chlorophyll content and higher substance of dissolvable proteins and Rubisco [14,17]. Melatonin improved antioxidant system for enhancing tolerance to high temperature in tomato [18-19] and in wheat [9] by increasing transcription of stress-responsive genes that helped in enduring photosynthetic machinery and increasing resistance to heat stress. In view of its diverse roles, melatonin is considered as a master regulator [14,20]. Moreover, melatonin shows interaction with different phytohormones and signalling molecules [17,21]. He and He [22] have reported melatonin and nitric oxide (NO) may actuate different plant physiological responses through their association under both normal or stress conditions and hydrogen sulfide (H₂S) communicates with ROS and NO influencing the two ROS and responsive nitrogen species (RNS) uptake [23,24]. Mukherjee and Bhatla [25] found that both H₂S and melatonin ameliorated salt stress mediated decrease in tomato growth and exogenous melatonin modulated early H₂S signalling.

Hydrogen sulfide is the third important gasotransmitter that regulates various physiological responses under stress and play an important role in the damaging effect of heat stress [4]. It directs germination, development, senescence and plays protective role in plants [26]. Furthermore, it can diffuse to various cells parts and maintain equilibrium the antioxidant system pools by providing sulfur to cells or changing osmolytes levels, malondialdehyde substance or mechanism of H₂S biosynthesis and enzymes of antioxidants [27]. Yang et al. [28] found that H₂S played an important role in combating heat stress and sodium hydrosulfide (NaHS)-pretreated seedlings (a H₂S donor) reduced oxidative stress via enhancement in the activity and gene expression level of antioxidant enzymes along with soluble sugar levels and H₂S in wheat. In *Spinacia oleracea* seedlings, H₂S stimulates chloroplast biogenesis for the improvement of photosynthesis and the gene expression of photosynthetic enzymes and thiol redox modification [29].

During study of the photosynthetic process under heat stress, it is equally important to gather knowledge on the metabolism of carbohydrates under stress. Starch plays an important role in the tolerance of different abiotic stress [30], and works as a source-sink relations modulator [31]. Starch degradation is activated under stress and contributes to sugar accumulation [32,33]. Sugars may likewise aggregate from photosynthesis or on account of reduced demand, under development constraint of stress [30]. In transgenic plants with suppressed sucrose synthase expression, Zrenner et al. [34] in potato tubers and Tang and Sturm [35] in carrot taproots found reduced starch accumulation supporting connection between sucrose synthase and starch synthesis. Plants remobilize starch to afford carbon and energy under abiotic stress to reduce photosynthesis, but sugar released by starch degradation derived metabolites to support plant development, in addition to functioning as an osmoprotectant to mitigate stress adversities [36]. Exogenous NO and H₂S enhanced photosynthesis and reduced the glucose-arbitrated photosynthetic repression in wheat under heat stress [4]. The work of Dawood and ElAwadi [37] on *Vicia faba*,

and Wei et al. [11] on soybean elaborated that melatonin supplementation increased carbohydrates accumulation that helped in salt tolerance. Several studies report that besides acting as an antioxidant, melatonin manages worked in-related genes, comprising the metabolism of carbohydrate/fatty and amino acids [11, 38,39].

The present study was initiated to find how modulation of antioxidative enzymes and carbohydrate accumulation was related to the impact of melatonin and alleviation of heat stress and consequent upon the changes in plants growth and photosynthesis. Evolution of H₂S was also estimated and the relationship between melatonin and H₂S for heat tolerance in wheat was worked out. Wheat was chosen because it is one of the most important crops worldwide contributing to 20% of the total calories and proteins in the human intake [40]. There is high demand for wheat, but the crop is susceptible to climate change [41]. Therefore, it is required to work out its tolerance strategies under heat stress.

2. Materials and Methods

2.1 Plant Material and Growth Conditions

The present investigation was executed on wheat (*Triticum aestivum* L. cultivar WH 542). The method followed by Iqbal et al. [4] was adopted to carry out the experiment.

The details of the process adopted for experimentation and procedures for determinations are given in Supplementary file S1.

2.2 Treatments

Plants were subjected to heat stress by exposing them for 6 h to 40° C temperature daily during afternoon. The heat treatment was given after seedling emergence with 2-3 leaves and at 10 days after seed sowing. Heat treatment was maintained for 15 days at the same time and for the same duration. After the heat stress treatment, plants were allowed to grow at 25 °C (optimum temperature) for 5 days. The sampling time was 30 days. In the morning the plants were supplemented with Hoagland (nutrient solution) on alternate days. Throughout the experimental period the untreated control plants were provided an optimum temperature of 25 °C. A hand sprayer was used to supply 100 µM melatonin on foliage to both heat treated or control plants at 5 days after heat stress or 15 days after seeding. A Tween-20 (0.01%) was added with both melatonin and control. To substantiate the role of H₂S in melatonin-mediated heat tolerance, we applied 100 µM hypotaurine as H₂S inhibitor in all treatments and for all parameters studied. It was also applied at 15 days after the sowing. All concentrations taken are backed by the research of Kaya et al. [21]. Four replications were taken for each treatment.

2.3 Measurement of Growth and Photosynthetic Parameters

To measure dry weight of plants, the plants were pulled out with the root, washed to remove soil and dried on blotting paper. These plants were kept in oven at 80°C till a constant weight was obtained and weighed for dry mass determination. Leaf area was obtained through the use of leaf area meter (LA211 Systronics, New Delhi, India).

Gas exchange photosynthetic parameters (stomatal conductance, net photosynthetic rate, and intercellular CO₂ concentration) were measured with infrared gas analyzer (CID-340, Photosynthesis System, BioScience USA) in the full expanded upper leaves. During the measurement PAR was 780 µmol m⁻² s⁻¹ and atmospheric CO₂ concentrations was 390 ± 5 µmol mol⁻¹ (i.e., at light saturating intensity)

SPAD chlorophyll meter (502 DL PLUS Spectrum Technologies, USA) was used to determine chlorophyll content in the whole leaves. For chlorophyll fluorescence measurement, we used Junior-PAM chlorophyll fluorometer (Heinz Walz Germany).

2.4 Determination of H₂S, H₂O₂ and TBARS Content

The process demonstrated by Xie et al. [42] was used for measuring leaf H₂S content. The generation of methylene blue from dimethyl-phenylenediamine in HCl was used for estimation of H₂S content with slight modification.

The procedure adapted by Okuda et al. [43] was followed for the leaf H₂O₂ content determination. Peroxidase was added to initiate the reaction and the absorbance increase was noted at 590 nm.

Lipid peroxidation was done by estimating thiobarbituric acid reactive substance (TBARS) content through the method of Dhindsa et al. [44] and reported in Iqbal et al. [4].

2.5 Assay of Antioxidant Enzymes

The superoxide dismutase (SOD) activity was measured adopting Giannopolitis and Ries [45] and Beyer and Fridovich [46] method. To assay catalase (CAT), ascorbate peroxidase (APX) and glutathione reductase (GR) the method used was Aebi [47], Foyer and Halliwell [48] and Nakano and Asada [49], respectively with little modifications. The complete process is given earlier in the study of Fatma et al. [50].

2.6 Determination of Enzymes Involved in Carbohydrate Metabolism

Rubisco (EC 4.1.1.39) activity was determined using the method described by Usuda [51] and reported earlier (Iqbal et al. [4]). The reaction was started by addition of enzyme extract to the assay medium followed by conversion of 3-phosphoglycerate to glycerol 3-phosphate and at this stage NADH oxidation at 30 °C at 340 nm was monitored. Protein was estimated with bovine serum albumin as standard.

Activity of sucrose phosphate synthase enzyme (EC 2.4.1.14), sucrose synthase enzyme (EC 2.4.1.13), and soluble acid invertase was assayed adopting the method of Kalwade and Devarumath [52]. The activity of ADP-Glucose pyrophosphorylase was determined by the method of Kleczkowski et al. [53]. The pyrophosphorolytic activity of ADP-Glucose pyrophosphorylase was monitored by observing the increase in absorbance at 340 nm during the conversion of NADP to NADPH at 340 nm.

2.7 Determination of Starch, Sucrose and Total Soluble Carbohydrate

Starch estimation was done using the method described by Kuai et al. [54], while total soluble sugars and sucrose content were measured adopting the procedure of Xu et al. [55].

2.8 Statistical Analysis

Analysis of data was done statistically through analysis of variance (ANOVA) by SPSS 17.0 suggested for Windows. The data are represented as treatment mean with (\pm) SE ($n = 4$). Least significant difference (LSD) was calculated for the significant data at $p < 0.05$ and $p < 0.01$. Bars showing the same letter are not significantly different by LSD test at $p < 0.05$ or $p < 0.01$.

3. Results

3.1. Melatonin Improves Growth and Photosynthesis under Heat Stress via H₂S

We examined growth and photosynthesis parameters in wheat leaves treated with melatonin alone or in combination with heat stress in order to evaluate melatonin response under stress conditions. Substantial reductions were observed in leaf area and plant dry mass of plants by 32.8 and 52.5%, under heat stress in comparison to control. The application of melatonin individually increased plant dry mass by 62.6% and leaf area by 37.3% compared to control. The negative effect of heat stress on plant growth was altered with individual use of melatonin in heat stressed plants. The application of melatonin under heat stress proved more effective in increasing leaf area and plant dry mass by 21.7% and 43.4%, respectively, compared to control plants (Fig. 1). Supplementation of

hypotaaurine to melatonin and heat- stressed plants resulted in reduction in leaf area and growth significantly equal to heat stress again emphasizing that melatonin action in protecting growth under heat stress was H₂S dependent.

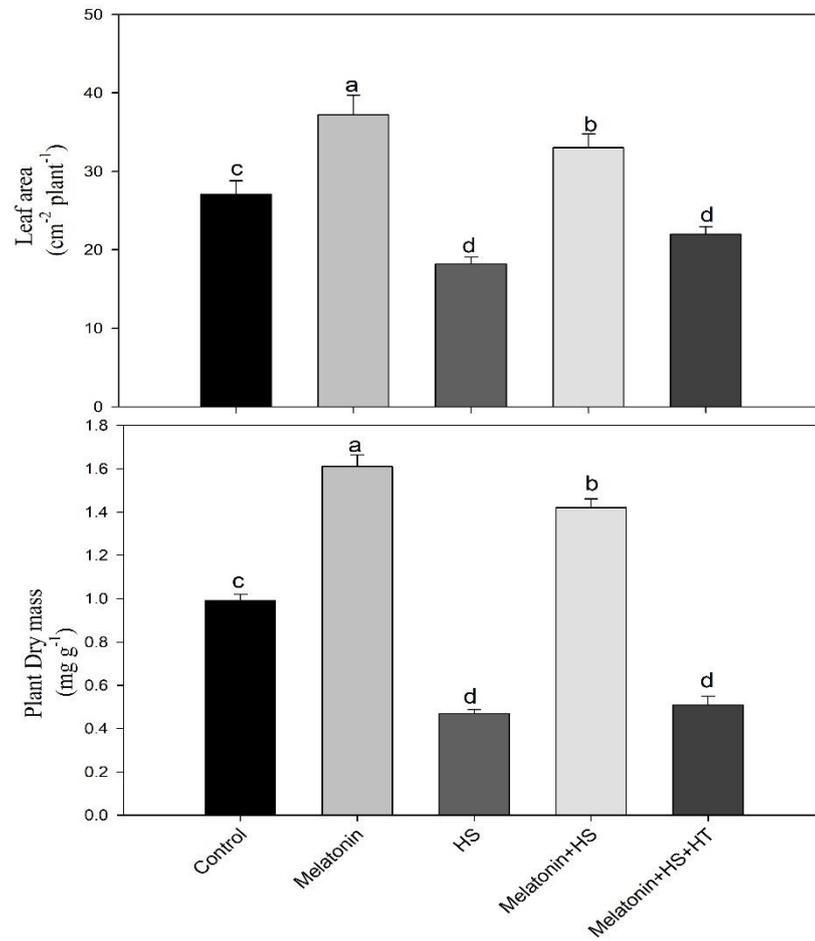


Figure 1. Leaf area and plant dry mass of wheat (*Triticum aestivum* L. var. WH 542) leaves at 30 days after sowing (DAS). Plants were treated with melatonin (100 μ M) and/or HT (100 μ M) in the presence (40 °C) or absence (25 °C) of heat stress. Data are presented as means \pm SE ($n = 4$). Data followed by same letter are not significantly different by LSD test at $p < 0.05$. HS, heat stress; HT; hypotaaurine.

Heat stress declined net photosynthetic rate, stomatal conductance, intercellular CO₂ concentration by 31.2, 38.4 and 29.5%, respectively in comparison to control. In contrast, melatonin increased net photosynthetic rate, stomatal conductance and intercellular CO₂ concentration by 18.7, 38.1 and 22.8% under no stress, and 11.7, 17.5 and 10.4%, respectively under heat stress in comparison with the untreated control plants (Fig. 2).

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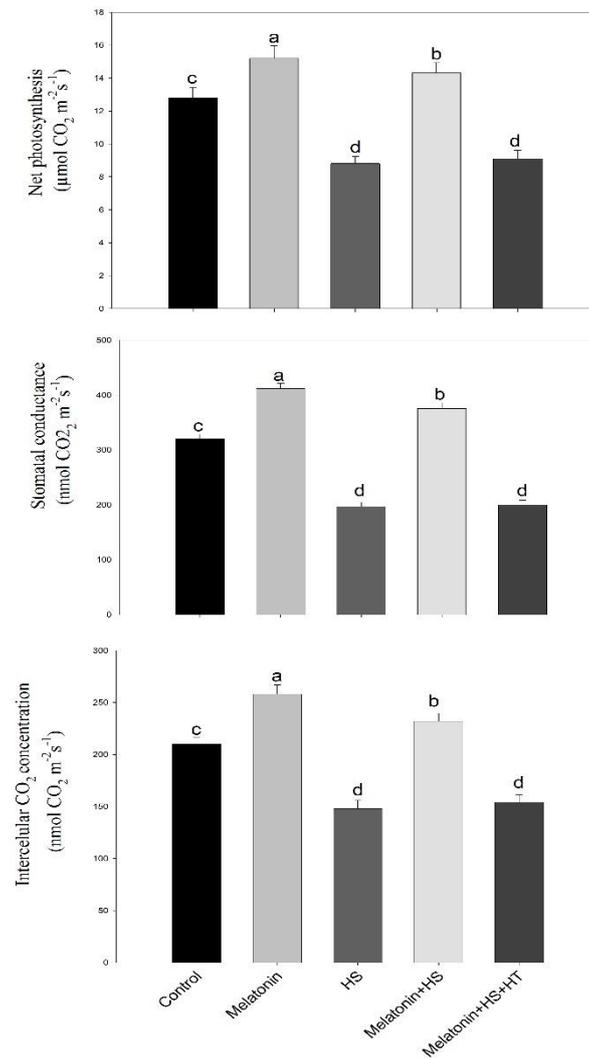


Figure 2. Net photosynthesis, intercellular CO₂ concentration, stomatal conductance of wheat (*Triticum aestivum* L. var. WH 542) leaves at 30 days after sowing (DAS). Plants were treated with melatonin (100 µM) and/or HT (100 µM) in the presence (40 °C) or absence (25 °C) of heat stress. Data are presented as means ± SE ($n = 4$). Data followed by same letter are not significantly different by LSD test at $p < 0.05$. HS, heat stress; HT; hypotaurine.

Similarly, Chl content and maximal PS II photochemical efficiency were also increased by 17.9 and 26.7%, respectively with melatonin under heat stress compared to control. The maximal increase was obtained with the melatonin alone in comparison to control. The results showed that melatonin strongly reduced the negative outcome of heat stress and improved the photosynthetic parameters (Fig. 3). Heat stress decreased Rubisco activity by 39.4%, while melatonin supplementation increased it by 20.2% under no stress and by 4.4% under heat stress conditions.

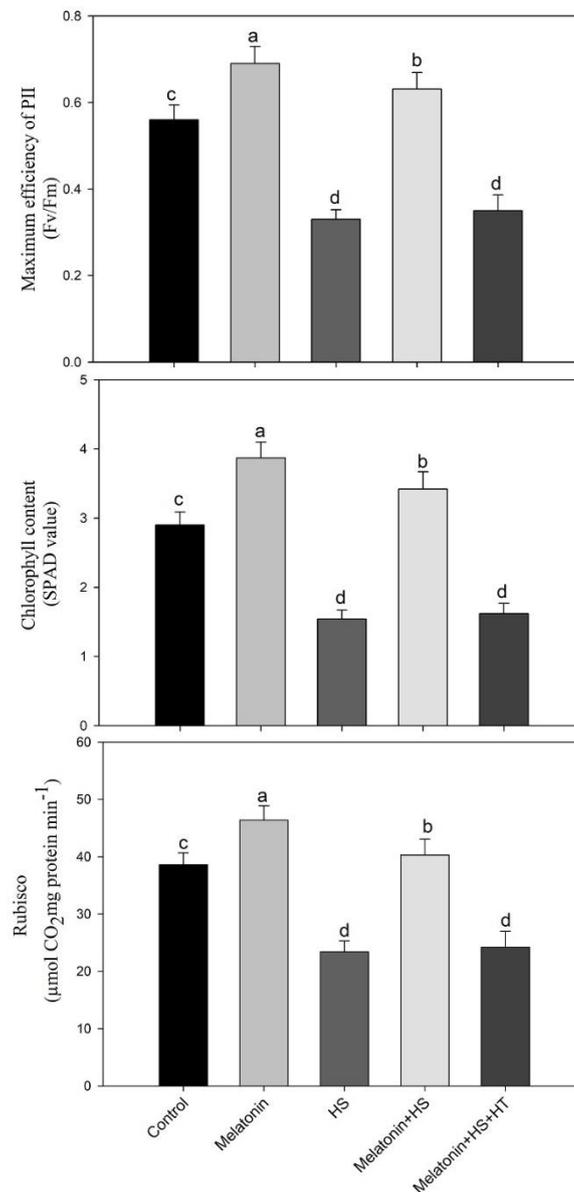


Figure 3. Maximal efficiency of PS II, chlorophyll content and Rubisco activity of wheat (*Triticum aestivum* L. var. WH 542) leaves at 30 days after sowing (DAS). Plants were treated with melatonin (100 µM) and/or HT (100 µM) in the presence (40 °C) or absence (25 °C) of heat stress. Data are presented as means ± SE ($n = 4$). Data followed by same letter are not significantly different by LSD test at $p < 0.05$. HS, heat stress; HT; hypotaurine.

Melatonin was thus efficient in enhancing plants photosynthetic potential under heat stress, but hypotaurine treatment inhibited increases in these photosynthetic parameters even in the presence of melatonin. This suggests the role of H₂S in melatonin mediated photosynthetic protection under heat stress. Possibly, melatonin functions upstream of H₂S in inducing heat stress signalling response.

3.2. Involvement of H₂S in Melatonin-Induced Reduction of Oxidative Damage Caused by Heat Stress via Increasing Antioxidant Systems

Exogenous melatonin was supplemented on heat-stressed wheat plants to evaluate its' effect on management of oxidative stress. Heat stress led to significant increases in H₂O₂ and TBARS content by 2.6 and 2.3-times, respectively over the control. Melatonin mitigated the oxidative damage caused by heat stress by decreasing H₂O₂ and TABRS content by 21.8 and 16.4%, respectively as compared to control (Fig. 4), and when compared to heat stressed plants, we found that H₂O₂ content was reduced by 70.7 and TBARS by 64.1%. Melatonin alone maximally reduced oxidative stress with H₂O₂ content reduction by 33.7 and TBARS by 32.7% compared to control. Hypotaurine, H₂S inhibitor, reduced the ameliorative effect and we observed that the contents of H₂O₂ and TBARS were significantly equal to heat stressed plants suggesting the importance of H₂S in melatonin-mediated reduction in oxidative stress.

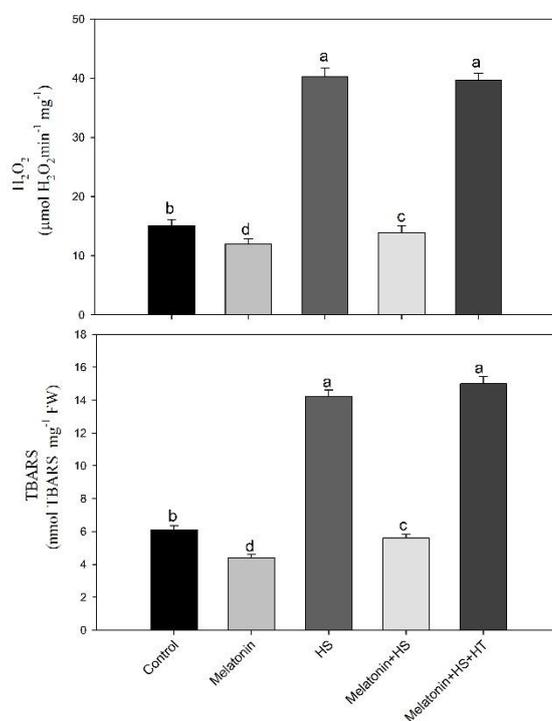


Figure 4. Content of H₂O₂ and TBARS in wheat (*Triticum aestivum* L. var. WH 542) at 30 days after sowing (DAS). Plants were treated with melatonin (100 µM) and/or HT (100 µM) in the presence (40 °C) or absence (25 °C) of heat stress. Data are presented as means ± SE ($n = 4$). Data followed by same letter are not significantly different by LSD test at $p < 0.05$. H₂O₂, hydrogen peroxide; HS, heat stress; HT; hypotaurine; TBARS, thiobarbituric acid reactive substances.

The effect of melatonin was also examined on antioxidant defense system under heat stress. Heat stress increased CAT, GR, APX and SOD by 25.4, 95.3, 70.9 and 46.7% respectively, compared to control plants. However, exogenous application of melatonin to non-stress plants increased the activity of CAT by 31.1, APX by 109.3% and GR by 79.7%, and also augmented the SOD activity by 59.8% compared with control. Melatonin enhanced CAT, APX, GR and SOD activities 44.3, 160, 94 and 80.7%, respectively in heat stressed plants compared to control (Fig. 5). On inhibition of H₂S with the supplementation of hypotaurine to melatonin-treated heat stressed plants, a reduction in the antioxidative

enzymes activities was observed signifying that melatonin needs H₂S signalling for initiating antioxidative enzymes mediated defense responses.

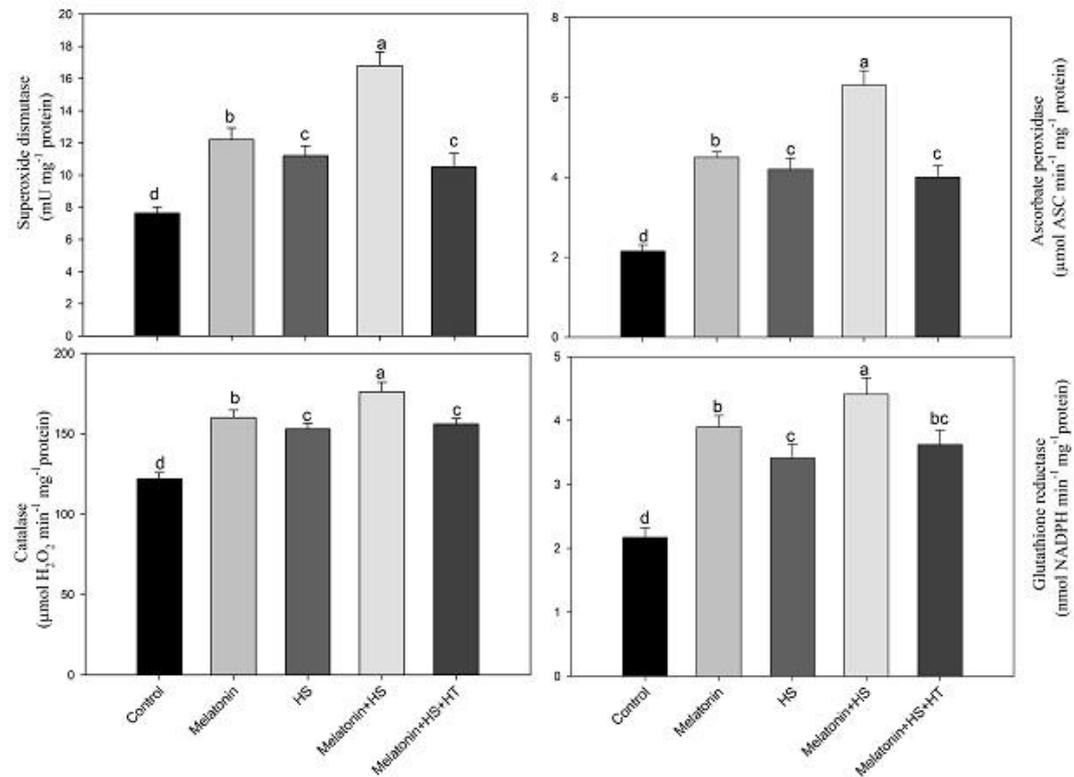


Figure 5. Activity of SOD, APX, CAT and GR of wheat (*Triticum aestivum* L. var. WH 542) leaves at 30 days after sowing (DAS). Plants were treated with melatonin (100 μM) and/or HT (100 μM) in the presence (40 °C) or absence (25 °C) of heat stress. Data are presented as means ± SE ($n = 4$). Data followed by same letter are not significantly different by LSD test at $p < 0.05$. APX, ascorbate peroxidase; CAT, catalase; GR, glutathione reductase; HS, heat stress; HT; hypotaurine; SOD, superoxide dismutase.

3.3. Melatonin Enhances H₂S under Heat Stress

Heat stress enhanced H₂S levels by 1.5-times compared with control plants. Exogenously applied melatonin further increased H₂S in plants, but H₂S level was not over than plants under heat stress (Fig. 6). However, the result was more significant with melatonin under heat stress by the evolution of leaf H₂S by 74.5 % compared to the control. These results indicate that melatonin-generated H₂S was involved in the reduction of oxidative stress.

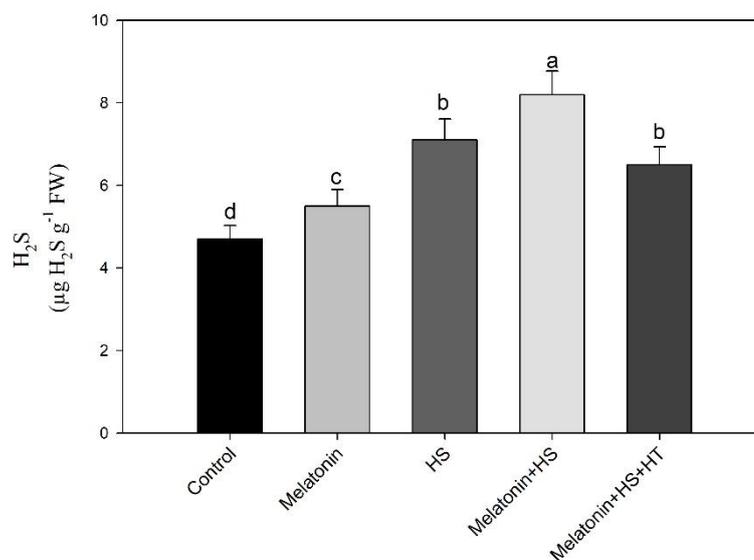


Figure 6. Evolution of H₂S content in wheat (*Triticum aestivum* L. var. WH 542) at 30 days after sowing (DAS). Plants were treated with melatonin (100 µM) and/or HT (100 µM) in the presence (40 °C) or absence (25 °C) of heat stress. Data are presented as means ± SE ($n = 4$). Data followed by same letter are not significantly different by LSD test at $p < 0.05$. HS, heat stress; HT; hypotaurine.

3.4 Involvement of H₂S in Melatonin-Induced Alteration of Soluble Sugars and Sucrose Content

Heat stress decreased the content of total soluble sugar by 7.2% compared with the control. Conversely, melatonin treatment either alone or under heat stress increased the total soluble sugar in comparison with the control. Melatonin increased the soluble sugar content by 22.8% under non stress and by 11.2% under heat stress conditions. Thus, melatonin treatment under heat stress was beneficial to plants adaptation to environmental changes and increased the soluble sugar content. These soluble sugar contents are used to analyze the defensive ability of wheat against stress responses. In the presence of H₂S inhibitor, we observed a decrease in soluble sugars which was significantly equal to heat stress plants even in the presence of melatonin signifying that melatonin increased soluble sugars under heat stress via H₂S.

The sucrose content was found to increase under heat stress by 2.7-times compared to the control. Exogenous melatonin enhanced sucrose content both under no stress or heat stress but in both the cases it was lower than heat-stressed plants. On hypotaurine treatment to heat treated melatonin supplemented plants, a decline in the content of sucrose below the level of control was noted and melatonin mediated increase was not observed. This suggests that melatonin requires H₂S for its action on carbohydrate metabolism (Table 1).

3.5. Requirement of H₂S for Melatonin-induced Effects on the Enzymes Activity in Sucrose Synthesis and Metabolism in Leaves

Heat stress had significant effects on the activities of sucrose synthase, sucrose phosphate synthase, soluble acid invertase related to sucrose synthesis and cleavage (Table 1). The activity of sucrose phosphate synthase increased by 45.8%, while decrease in sucrose

synthase by 26.7% and soluble acid invertase by 10.0% was observed in comparison to the control. The treatment of melatonin under non stress plants increased sucrose phosphate synthase by 21.2% compared to control though the increase was considerably equal to heat stressed plants. Maximum sucrose phosphate synthase activity was noted with heat stress plants supplemented with melatonin. Activity of sucrose synthase decreased by 7.8% and acid invertase by 16.5% compared to control under heat stress, while melatonin improved it both under heat stress or no stress conditions. The content of sucrose could be increased by improving sucrose phosphate synthase activity of plants, and provide explanation of the result of increase in sugar transport.

Table 1: Total soluble sugars, sucrose content, starch content, and activity of sucrose phosphate synthase, sucrose synthase, soluble acid invertase, and activity of ADP-Glucose phosphorylase in wheat (*Triticum aestivum* L. var. WH 542) at 30 days after sowing (DAS). Plants were treated with melatonin (100 μ M) and/or hypotaurine (HT, 100 μ M) in the presence of (40 °C) or absence (25 °C) of heat stress. Data are presented as means \pm SE ($n = 4$). Data followed by same letter are not significantly different by LSD test at $p < 0.05$. HS, heat stress; HT; hypotaurine.

Treatments	Soluble sugars	Sucrose content	Starch content	Sucrose phosphate synthase	Sucrose synthase	Soluble acid invertase	ADP glucose phosphorylase
	(mg g ⁻¹ DW)			(μmol g ⁻¹ protein min ⁻¹)		(nmol g ⁻¹ protein min ⁻¹)	
Control	62.31 \pm 1.69c	15.90 \pm 2.84d	25.82 \pm 0.67c	20.13 \pm 0.59d	66.75 \pm 2.68bc	146.87 \pm 4.68c	22.63 \pm 0.81c
Melatonin	76.55 \pm 3.22a	24.82 \pm 3.42b	30.26 \pm 0.82a	24.40 \pm 0.69bc	79.2 \pm 3.16a	171.10 \pm 6.84a	26.19 \pm 1.04a
HS	57.86 \pm 1.49d	42.3 \pm 1.56a	16.24 \pm 0.47d	29.36 \pm 0.84b	48.96 \pm 1.23d	132.1 \pm 3.32b	18.06 \pm 0.58d
Melatonin +HS	69.36 \pm 1.44b	20.3 \pm 1.83bc	27.40 \pm 0.89b	36.48 \pm 0.99a	72.04 \pm 2.44b	169.43 \pm 5.72a	24.37 \pm 0.69b
Melatonin +HS +HT	60.16 \pm 1.63d	11.1 \pm 1.59e	17.35 \pm 0.67d	13.20 \pm 0.48e	50.20 \pm 1.59d	139.57 \pm 3.44b	17.58 \pm 1.59d

Inhibition of H₂S by hypotaurine inhibited the increase in sucrose synthesis and degradation enzymes and the decrease was comparable or lower than heat treated plants even in the presence of melatonin. Thus, H₂S has a major role in affecting melatonin-mediated sucrose synthesis enzymes under heat stress.

3.6. Effect of H₂S in Melatonin-Induced Starch Accumulation and the Related Enzyme ADP-Glucose Phosphorylase

The effect of melatonin on starch accumulation and ADP-Glucose phosphorylase was examined in photosynthetic tissue of wheat plants under non stress and stress states. Heat stress decreased both accrual of starch and the activity of ADP-Glucose phosphorylase enzyme by 37.2 and 20.3%, respectively, compared to control. However, starch accumulation was increased by 18.6% in plants treated with melatonin alone and by 7.8% in plants treated with melatonin plus heat stress, as compared to control (Table 1). Similarly, the activity of ADP-Glucose phosphorylase enzyme was increased by melatonin in non-stressed plants by 15.5%, while an increase of 7.5% was observed with melatonin under heat stressed plants in comparison to control. Application of hypotaurine inhibited the

melatonin-induced increase in both starch and ADP-Glucose phosphorylase activity under heat stress.

4. Discussion

Plants initiate several mechanisms such as stimulation of antioxidant system, accumulation of osmolytes or secondary metabolites, heat shock proteins to resist the harmful effect of heat stress through maintenance of cellular homeostasis and repair of damaged membranes and proteins. These mechanisms are associated with production of various hormones and signalling molecules that help in adapting to heat-induced adversities. Kaya et al. [56] reported that interactive effect of H₂S and NO resulted in reducing oxidative stress and cadmium uptake by enhancing the antioxidative enzyme system and mineral nutrients uptake. At low concentration, H₂S affects plant development and growth by playing a major role in abiotic and biotic stress responses [27,29,57]. Hydrogen sulfide has gained much attention because it is a vital mode of sulfur metabolism in organisms and has increased special significance owed to its signalling properties, besides the essentiality of S requirement is known under both abiotic and biotic stress [58]. Under conditions of stress when ROS accumulation causes oxidative stress, the increased H₂S helps in reducing the ROS via enhancing the non-enzymatic and enzymatic ways. The rescue mechanism of H₂S also involves signal that regulates stomatal movement, increases GSH, redox ratio, along with GSH-associated genes expression under chilling stress [59]. H₂S relates through H₂O₂ and was found to upregulate ascorbate-glutathione cycle which acted as the downstream signal for H₂S regulation on ROS [60,61]. Exogenous H₂S application on plant seedlings was reported to inhibit oxidative stress by reducing the MDA activity, electrolyte leakage and proline [61-62] and protected photosynthesis and stabilized the chloroplast structure [63].

The interaction of H₂S with various other signalling molecules has also been reported. Hydrogen sulfide with Ca²⁺ and CaM effectively alleviated heat-induced damage to plants via increasing the H₂S accumulation [64] and H₂S-induced heat tolerance required Ca²⁺ transport toward cytoplasm then intracellular CaM coordinates it [65]. Methylglyoxal (MG) which resembles H₂S contributes in abiotic stress response and application of MG and/or NaHS enhanced maize seedlings response to heat stress and both showed positive interaction [66]. A crosstalk of H₂S signals with carbon monoxide, salicylic acid (SA), abscisic acid (ABA), and ethylene has also been reported [67-70], and these are found to prompt H₂S-producing enzymes activation and endogenous H₂S accretion under high-temperature stress. Hydrogen sulfide interacts with ethylene under osmotic stress [71], with ABA for closure of stomata [70], with NO for heat tolerance [4] with SA for lead tolerance [72], with H₂O₂ and brassinosteroid (BR) for stomatal closure [73] and with melatonin for salt and iron deficiency tolerance [21].

While undergoing various studies we found that melatonin interaction with H₂S under heat stress has not been worked out, although antioxidants and photosynthesis are shown to be influenced with H₂S application. We still do not know how H₂S affects carbohydrate metabolism under heat stress and its relationship with melatonin. Melatonin is known as a biopromoter as it regulates numerous physiological methods and improves heat stress resistance in plants [74,75]. It is a small molecular weight indoleamine hormone explicitly being measured as a candidate phytohormone due to its various responses under biotic and abiotic stresses. Its level increases upon plant's exposure to abiotic stresses and it scavenges ROS molecules and enhances activity of antioxidant enzyme, content of metabolite, photosynthetic efficiency and regulates stress transcription features and other metabolites and signalling molecules [76]. However, the information on how melatonin regulates carbohydrate metabolism and photosynthesis in heat stress in wheat and its interplay with H₂S in this mechanism is scanty. In the present study, melatonin enhanced plants' photosynthetic potential by increasing carbohydrate accumulation with enhanced antioxidative enzymes activity. The influence of melatonin was found to be dependent on

H₂S evolution as evidenced that the inhibition of H₂S inhibited photosynthesis and growth in wheat.

4.1. Melatonin Increases H₂S Evolution in Wheat under Heat Stress

Exogenous application of melatonin further increased H₂S evolution which was induced under heat stress. Previous studies have reported that heat stress induces H₂S evolution [4]. The interrelationship between melatonin and H₂S has been stated in some researches. Kaya et al. [55] found that melatonin imparted resistance to iron deficiency and salt-stress which was intervened through both NO and H₂S. These molecules went about as downstream signal molecule in melatonin actuated resilience. Exogenous melatonin treatment under salt stress modulated the endogenous H₂S growth and absolutely upregulated the L-cysteine desulfhydrase activity under salt stress in tomato seedlings [25]. Turk and Erdal [77] studied that melatonin application enhanced mineral element content in cold-stressed plants. Among studied nutrients, sulfur level also increased by melatonin under stress which acts as a building block of proteins, enzymes and vitamins. Sulfide is an important intermediate in sulfur metabolism and the amount of H₂S released has been correlated with sulfate supply to the plants [78].

4.2. Melatonin Decreases Heat Stress-Induced Oxidative Stress by Enhancing the Antioxidative Machinery: the Effect Mediated by H₂S

In this research, heat stress was found to increase oxidative stress supported by increased TBARS and H₂O₂ content which was reduced by the supplementation of melatonin. Melatonin supplementation increased the activity of antioxidative enzymes (CAT, SOD, APR, GR) which scavenges the excess generated ROS to reduce oxidative stress. Enhanced activity of antioxidative enzymes detoxifies excess ROS and reduces oxidative stress. Reports suggest that melatonin mediates several enzymes expression that detoxify excess H₂O₂ such as, CAT, glutathione/ascorbate reductases, peroxidases in addition to peroxiredoxins [17,79] and balances the ROS/RNS level through antioxidant action. Either endogenous or exogenous supplied melatonin induces its own synthesis presenting melatonin as specific regulator, an anti-stress manager and a plant master regulator [80,81]. Siddiqui et al. [82] reported that melatonin induces photosynthetic enzymes, antioxidative system, proline metabolism and carbohydrate level under salt stress. Since supplementation of melatonin increases H₂S therefore, it was thought that melatonin action was via H₂S. Kaya et al. [21] have studied crosstalk among H₂S and NO in melatonin-induced salt stress tolerance in pepper. NaHS pretreatment enhances cellular viability and was found to decrease MDA accumulation and electrolyte leakage [65]. H₂S diminishes the harmful effect of oxidative stress under heat stress [67]. In our study we found that SOD, CAT, APX, GR activity amplified in stress and more increase was received with melatonin, but inhibition of H₂S activity inhibited the increase in antioxidative enzymes. Similarly, Iqbal et al. [4] reported that H₂S enhances antioxidative enzymes activity and ascorbate-glutathione cycle to scavenge excess ROS and reduce oxidative stress in wheat under heat stress. Although not much has been discussed on the interaction between melatonin and H₂S under heat studies, various studies support for a regulatory interaction either directly or indirectly. The relationship of melatonin, H₂S and NO was considered in regulation of fruit ripening [83] and under salt stress by Kaya et al. [56]. However, the present reported study is the first pertaining to melatonin and H₂S under heat stress in wheat.

4.3. Impact of Melatonin on Photosynthesis and Carbohydrate Metabolism under Heat Stress: reversal of the Effect by H₂S Inhibitor

Heat stress affects the process of photosynthesis [4,84] by altering the activities of enzymes in metabolism of carbon and it also affects sucrose synthesis and starch accumulation by modulating explicit genes of the carbohydrate metabolism pathway [85]. The reduction in photosynthetic efficiency could be attributed to reduced activity of the Calvin cycle enzymes comprising Rubisco [86]. We found decreased Rubisco activity under heat stress which was restored by melatonin application. Reduction in chlorophyll content also leads to reduced photosynthesis probably due to increased chlorophyllase activity [87]. In the present study, the impact of melatonin was observed in inhibiting heat-induced photosynthetic reduction. Heat-stress reduced the content of chlorophyll, Rubisco activity in addition to efficiency of PSII and reduced stomatal conductance, intercellular CO₂ concentration and net photosynthesis. However, melatonin supply increased Rubisco, chlorophyll content and photosynthetic traits which were reduced by application of H₂S inhibitor suggesting the involvement of H₂S in melatonin-induced increase in photosynthesis under heat stress. Individually both melatonin and H₂S are reported to enhance photosynthesis but here we found melatonin effect to be mediated by H₂S.

We observed that heat stress led to decrease in accumulation of starch content and total soluble sugar. Exposure of plants to heat stress can be harmful to the starch synthesis enzymes activity and starch content [88]. In several studies, ADP-Glucose pyrophosphorylase activity in maize, wheat, barley and rice were repressed under heat stress which reduced starch biosynthesis [88,89]. The present work reveals that the activity of ADP-Glucose pyrophosphorylase, one of the core rate limiting starch biosynthesis enzymes decreased in heat-stressed plants. Conversely, study also suggests that heat stress primes to rise in content of carbohydrates that helps to uphold cell turgor, helps in cell layers' stabilization and prevents deterioration of protein [90]. Soluble sugar acts as a signalling molecule that controls growth and metabolic processes and defence linked genes to regulate plant growth [91,92]. Thus, enhanced sugar accumulation is required for potential tolerance of plants and we observed rise in sucrose under heat stress which was reduced by melatonin supplementation when the stress was relieved and photosynthesis and starch increased. The degradation of starch under heat stress was supported by reduced ADP-Glucose phosphorylase activity then increased sucrose phosphate synthase and sucrose synthase activity which helped in utilization of sugars for plant growth. Melatonin led to enhancement of starch and sucrose content to alleviate heat induced oxidative stress by maintaining an osmotic balance and providing energy and food for growth. These actions of melatonin were blocked by H₂S inhibitor which supported the role of H₂S in increasing carbohydrate accumulation under heat stress.

Heat tolerance requires the accumulation of sugars in plants and availability of carbohydrate (glucose and sucrose), is a significant physiological attribute for the resistance of heat [93]. They reported decreased carbohydrate content under heat stress that was more in heat-susceptible genotype compared to tolerant one. In our study the content of total soluble sugar decreased in heat stress, but by supplementation of melatonin under heat stress sugar accumulation increased. Figure 7 highlights the important changes that occur in the carbohydrate metabolism under heat stress. It may be said that plants under heat stress increase their sucrose content by favouring the degradation of starch. The conversion of sucrose to starch is reduced with reduced activity of ADP-Glucose phosphorylase but increased sucrose phosphate activity increases sucrose content. We observed that the activity of invertase and sucrose synthase reduced which resulted in reduced total soluble sugars, also with reduced starch content. The interconversion of starch-sugar in leaf is necessary for the tolerance in plants from abiotic stress [94].

Higher sucrose phosphate synthase activity under heat stress helps in shifting the carbon grade starch to sucrose synthesis [95]. Melatonin increased the activity of sucrose synthase and invertase under heat stress thus leading to increased mobilization of carbon source to demanding sink. Increased expression of sucrose phosphate synthase genes are reported to increase heat tolerance. Lafta and Lorenzen [96] reported that at high temperatures, improvement in the sucrose phosphate synthase genes expression reasons upsurge in the sucrose synthesis that improves tolerance of plants under slight heat stress. Alike

result was seen in our study by increased sucrose and sucrose phosphate synthase activity in heat stress. Sucrose phosphate synthase activity further increased with melatonin treatment under heat stress, however, sucrose content decreased on melatonin treatment compared to heat-stressed plants although they were greater than control. The reason for this could be the utilization of sucrose with melatonin for plants growth and development with the release of stress. Khan et al. [94] reported increased sucrose content to be aligned with decreased starch content under salt stress in tomato seedling roots. The melatonin-induced increase in carbohydrate was reduced when hypotaurine was supplemented with melatonin suggesting that this carbohydrate accumulation by melatonin require H₂S for its action under heat stress.

The increase in invertase activity with melatonin helps in heat tolerance and this increase was mediated through H₂S. Siddiqui et al. [82] stated that melatonin alleviated salt stress through osmoregulation via growing the content of total soluble carbohydrate and proline and upregulated carbohydrate metabolism. Increased invertase activity enhanced sucrose transport and was found to improve heat tolerance [65].

Hydrogen sulfide was found to increase glucose utilization under heat stress in wheat and decreased heat-induced reduction in photosynthesis [4]. NaHS treatment to *S. oleracea* leaves increased accumulation of glucose and sucrose though it was lower than control nevertheless trehalose and fructose contents considerably increased above the control suggesting the role of trehalose and fructose in drought tolerance. Increased photosynthesis and PSII efficiency in *S. oleracea* under drought stress on H₂S supplementation was due to modulation of sugar metabolism [97]. Ye et al. [66] found that H₂S increases photosynthesis by promoting Rubisco activity plus photosynthetic electron transport increases chlorophyll biosynthesis. It also modulates Fv/Fm ratio, chlorophyll content together with net photosynthetic rate, stomatal conductance, and transpiration rate [60,98]. Khan et al. [94] reported that potassium and endogenous H₂S regulated sugar metabolism for initiating adaptive responses under salinity stress. Hydrogen sulfide was found to alleviate salt stress and increased sucrose synthase, sucrose phosphate synthase, invertase activity and decreased starch content.

Although not much study has been done on the role of H₂S in regulating carbohydrate, here we observed that the inhibition of H₂S by its inhibitor resulted in decreased melatonin function on carbohydrate content suggesting the notion that melatonin effect on carbohydrate is via H₂S. Taken together, this is the first report that shows the importance of H₂S in melatonin-induced heat stress tolerance through enhancing carbohydrate metabolism under heat stress. It may be emphasized that melatonin modulated carbohydrate metabolism under heat stress to regulate tolerance of heat and caused reduction in oxidative stress and increase in antioxidative metabolism via H₂S. Such study of regulation of carbohydrate metabolism by H₂S in heat stress is not reported till date and this explores the possibility of H₂S application as heat stress mitigator in plants. In order to make the concept clear a diagrammatic representation of the effect of melatonin and H₂S on carbohydrate metabolism under heat stress is given (Fig. 7).

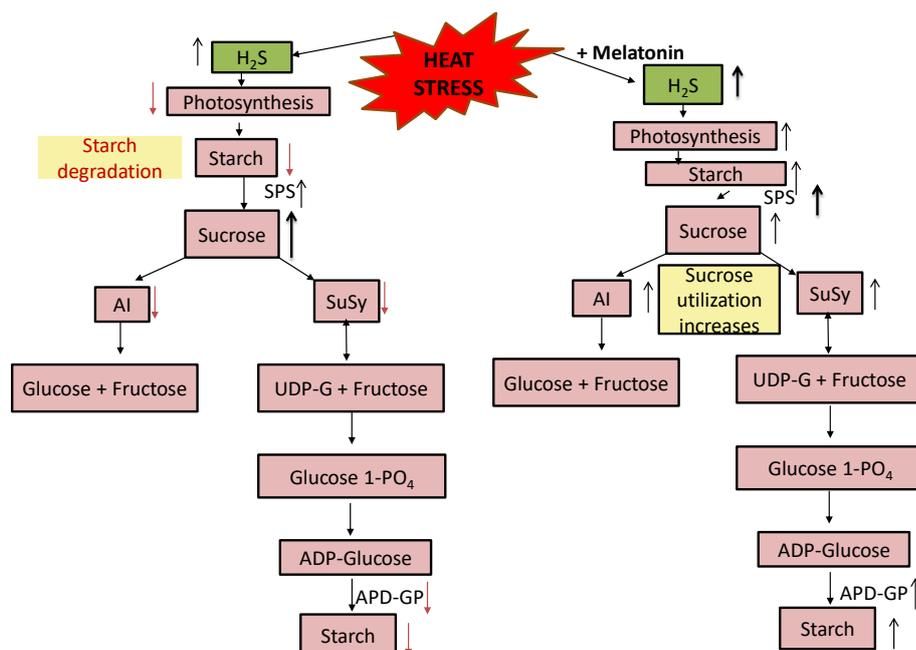


Figure 7. Influence of heat stress and melatonin on carbohydrate metabolism. Red arrow shows decrease and black arrow shows increase. Double arrow shows regulation in both the direction. Black thick arrow shows more increase. SPS, sucrose phosphate synthase; ADP-GP, ADP-Glucose phosphorylase; AI, acid invertase; SuSy, sucrose synthase; UDP-G, uridine diphosphate glucose.

5. Conclusions

Heat stress affects carbohydrate metabolism and reduces photosynthesis. This study emphasised that melatonin helps the plants under heat stress by not only decreasing the oxidative stress through enhancement in antioxidative enzymes activity but also increases photosynthesis and growth through increase in the carbohydrate utilization to support growth under adverse conditions. This effect of melatonin was found to be mediated via H₂S and inhibition of H₂S inhibited melatonin-mediated increase in heat tolerance by inhibiting the activity of antioxidative enzymes, photosynthesis and carbohydrate metabolism in wheat suggesting of downward signalling of H₂S in melatonin-induced heat tolerance.

Supplementary Materials: The following are available online at www.mdpi.com/xxx/s1, Material and methods details

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