
Article

Carbon, Nitrogen Dynamics and CO₂ efflux in the Calcareous Sandy Loam Soil Treated with Chemically Modified Organic Amendments

Ahmed Mohammed-Nour^{1,2}, Mohamed Alsewailem¹, Ahmed H El-Naggar^{1,3,4,*}, Mohamed H. EL-Saeid¹, Anwar A. Aly^{1,5} and Jamal Elfaki^{1,6,7}

¹ Department of Soil Science, College of Food and Agriculture Sciences, King Saud University, P. O. Box 2460, Riyadh 11451, Saudi Arabia.

² Soil and Water Research Centre, Agricultural Research Corporation, P. O. BOX 126, Wad-Medani, Gezira State 21111, Sudan

³ Sustainable Natural Resources Management Section, International Centre for Biosaline Agriculture (ICBA), Dubai 14660, United Arab Emirates. a.naggar@biosaline.org.ae

⁴ Department of Soil Sciences, Faculty of Agriculture, Ain Shams University, Cairo 11241, Egypt.

⁵ Soil and Water Science Department, Alexandria University, Alexandria 21545, Egypt.

⁶ Faculty of Agriculture, Nile Valley University, River Nile State, Sudan.

⁷ Desert Farming Techniques and Agricultural Biotechnology Programs, Sultan Qaboos Chair in Desert Farming, Arabian Gulf University, Manama, kingdom of Bahrain.

* Correspondence: a.naggar@biosaline.org.ae; Tel.: (+971551099647)

Abstract: In Saudi Arabia, more than 335,000 tons of cow manure is produced every year from dairy farming. However, the produced cow manure is usually added to the agricultural soils as raw or composted manure; the recovery of ammonia from cow manure is a promising technique for producing concentrated nitrogen fertilizer and reducing nitrogen losses from raw manure. The byproduct effluents from the recovery process are characterized by different chemical properties from the raw manure. The effect of using the recovery effluents as soil amendments on the soil carbon and nitrogen dynamics is not apparent. Therefore, A 90-day incubation experiment was conducted to study the impact of these effluents on CO₂ efflux, organic C, microbial biomass C, available NH₄⁺, and NO₃⁻ when added to agricultural soil. In addition to the two types of effluents (produced at pH 9 and pH 12), cow manure CM, cow manure compost CMC, cow manure biochar CMB, and control were used for comparison. The application of CM resulted in a considerable increase in soil available nitrogen, CO₂ efflux compared to other treatments. Cow manure biochar showed the lowest CO₂ efflux. Cumulative CO₂ effluxes of cow manure effluents were lower than CM; this possibly due to the relatively high C: N ratio of manure effluent. The content of P, Fe, Cu, Zn, and Mn decreased as incubation time increased. Soil microbial biomass C for soil treated with cow manure effluents (pH 12 and 7) was significantly higher than the rest of soil amendments and control.

Keywords: organic wastes; cow manure; CO₂ effluxes; ammonia stripping; C: N ratio

1. Introduction

Agriculture is considered the primary source of ammonia emission globally (Bouwman and Van Der Hoek, 1997) and regionally in, e.g., Europe (Sutton et al., 2011) and USA (Pinder et al., 2004). Ammonia and its inorganic derivatives, nitrite, and nitrate are easily percolated to the ground and surface water, resulting in a deterioration of water quality and risk hazards in drinking water (Dubrovsky and Hamilton, 2010).

Regarding CO₂ efflux from soil treated with organic amendments, FAO (2014) showed that the total annual emissions of greenhouse gases (GHG) from agriculture to the atmosphere in 2011 were 5,335 Mt CO₂ eq. Twenty-five percent of this amount is released into the atmosphere due to manure storage, management, and amendments. Soil CO₂ efflux, as described by Kuzyakov (2006) is efflux that comes from the root and

rhizomicrobial respiration, decomposition of plant residues, the priming effect induced by root exudation or by addition of plant residues, and basal respiration by microbial decomposition of soil organic matter (SOM). Many key factors control the CO₂ efflux from the soil, such as climatic factors, temperature and water content (Sallam, 2002), clay content (FAO, 2014), water holding capacity (Schjønning et al., 1999), C: N ratio (Kowalska et al., 1994). Risse et al. (2006) reported that among many applied fertilizers to maize in China, CO₂ and NO₂ emissions were significantly correlated to pig manure and inorganic fertilizer due to their low C: N ratio. An incubation experiment (Yuan et al., 2014) showed that the biochar amendments to soils reduced N-gases volatilization and decreased CO₂ emissions from soils due to the low C: N ratio in the used biochar.

Therefore several pretreatments have been suggested to reduce the C: N ratio of N-rich wastes such as manure and produce organic amendments with a reduced C: N ratio and higher soil carbon stability. These methods include composting (Paré et al., 1998), pyrolysis process to produce biochar (Ameloot, et al., 2015), and ammonia recovery from manure (Mohammed-Nour et al., 2019).

Composting animal manure and nitrogen-rich wastes is a technique to reduce nitrogen release from organic materials to soils. During composting, the C/N of mixed organic materials decreases due to the biochemical oxidation of organic matter (van Vliet et al., 2007). Composting of 'struvite' food waster reduced N loss by 18% compared to the use of raw wastes (SAAF, 2007). In a lysimeter study, it was found that the application of poultry manure and paper mill sludge blends resulted in a pulse of NO₃-N (170 and 156 mg N/L) that occurred three months following application. At the same time, compost treatments showed no such pulse (Dere and Stehouwer, 2011). Another study by Sparks et al. (1996) found that composting of cattle manure doubled the soil humic substances compared with the raw manure. They concluded that composting of cattle manure resulted in more stable and less decomposable organic compounds in soils. On the other hand, during the composting process, an appreciable amount of nitrogen is lost through the volatilization of ammonia (Steiner et al., 2010).

Another method for increasing the C/N of the organic material is the pyrolysis of wastes to produce biochar. The pyrolysis process involves the heating of plant biomass in the absence of oxygen gas (Lehmann, 2007). The pyrolysis process increased carbon percentage due to the increasing degree of carbonization. However, hydrogen and oxygen content tends to decline (Al-Wabel et al., 2013). Enders et al. (2012) found that during the pyrolysis of different feedstock to produce biochar, fixed carbon significantly increased. The N recovery was negatively correlated to a pyrolysis temperature.

The conversion of plant residues to biochar is an attractive strategy for mitigating atmospheric carbon dioxide emissions and enhancing carbon storage in soil (Xiang et al., 2015).

Sigurnjak (2019) recently showed that nitrogen recovered from wastes could be used as a nitrogen fertilizer for crop cultivation. Mohammed-Nour et al., (2019) achieved a 90% recovery of the ammonia in cow manure through alkalization and thermal treatment. They suggested the use of ammonia stripping technique to reduce the environmental risks associated with ammonium volatilization from manure. The chemical properties of the produced effluents from the ammonia recovery process are different from manure. Therefore, its chemical behaviour as a soil amendments is not clear.

Soil stable organic carbon is one of the parameters used to evaluate the sustainability and efficiency of soil carbon sequestration. Bastida et al. (2012) mentioned that different carbon fractions, such as the total organic carbon, water-soluble carbon, and microbial biomass carbon, were increased in amended soils compared to the control. Also, Lima et al. (2009) stated an increase in lignin and lignin-like products in the soil amended with compost.

The study's primary objectives were to investigate the effects of cow manure stripped ammonia effluents (CMSAEs) on temporal changes in soil CO₂ flux, nitrogen forms (NH₄⁺,

NO_3^- and total N), estimate the microbial biomass C as an indicator for microbial activity during incubation.

2. Materials and Methods

2.1. Soil and Organic Materials Characterization

2.1.1. Soil

Topsoil (0–30 cm) was collected from an agricultural field located in Aloyyna, Riyadh, Saudi Arabia (24°54'27.36" N and 46°23'35.49" E). Collected soil was homogenized, and sieved to less than 2 mm, then it was air-dried for the physicochemical characterization according to standard methods (Estefan, 2017). pH was determined in 1:5 water extract (w/v) using pH meter, Orion star A211, Electrical conductivity (EC) was measured in the filtrated extracts using YSI, USA.

2.1.2. Soil amendments

2.1.2.1 Cow manure (CM)

Cow manure (CM) was collected from ALSAFI dairy farm situated in Al-Kharj, Saudi Arabia.

2.1.2.2. Cow manure stripped ammonia effluents (CMSAEs)

CMSAEs are the byproduct of ammonia recovery from cow manure through the alkalization and distillation process (Mohammed-Nour et al., 2019). In this experiment, two effluents from different alkalization degrees (pH 9 and 12) were studied.

Effluents were prepared by treating 100 g of fresh CM with either 2.44 or 0.5 ml of 15 N KOH to bring pH to 12 or 9. Then 100 ml of deionized water was added to manure paste in a one-lit round glass bottle. Mixtures were heated up to 95 °C for 5 h. Ammonia was recovered in 25ml of 0.5 N sulfuric acid. CMSAEs were collected, then slowly cooled down to 25 °C.

2.1.2.3. Composted Cow manure (CMC)

Composted cow manure (CMC) was obtained from (ALSAFI dairy products facility, Riyadh, Saudi Arabia), CMC was stored at 4°C.

2.1.2.4. Cow manure biochar (CMB)

One kg of dry cow manure (CM) was pyrolyzed at 400°C for 4 h. Cow manure biochar (CMB) was produced as mentioned by Al-Wabel et al. (2013).

CMC and CMB were used in the incubation experiment as reference material to evaluate the C stability from CMSAEs of CM. Before application time, all organic materials were air-dried, grained, and sieved to 1mm.

2.2. Chemical analysis of soil amendments

Total nitrogen and carbon were determined using a EuroVector Elemental Analyzer EA3000 equipped with Callidus software (EuroVectorSpA, Milan, Italy). To determine total phosphorus, potassium, calcium, magnesium, and other micronutrients in manure, 0.2 g of dried manure were treated with 10 mL of concentrated nitric acid and digested according to the procedure of Creed et al., (1994) using microwave digestion (MARS, CEM Corporation, USA). The total concentration was determined in the digestate using an inductivity coupled plasma optical emission spectrophotometer (ICP-OES, PerkinElmer Optima 4300 DV, USA).

2.3. Incubation Experiment

A long-term incubation experiment was carried out under laboratory conditions for 90 days. The experiment consists of six organic materials; CM, CMC, CMB, plus the control (pH12 and T95°C) CMSAE and (pH9 and T95 °C) CMSAE. CMC and CMB were used to compare C, and N dynamics with that of CMSAEs. 100 g of 2mm sieved calcareous soil (texture: sandy loam), placed into 250-mL jars. Experimental soil was pre-incubated for 15 days at 25°C to allow the soil to equilibrate after sieving and handling. The moisture of the soil samples was initially adjusted to 75% of the water holding capacity (WHC) through the addition of deionized H₂O at regular intervals (1 to 2 weeks). The WHC was determined by saturating a sample of soil in filter paper placed in a glass funnel. Then, the water was drained for two h before the gravimetric soil moisture content (for 100% WHC) was determined by drying for 24 h at 105°C. Amendments were added to the soil at a 50 mg C.g⁻¹ soil based on the elemental C analysis of the used amendments—total added N amendments estimated before the experiment. A blank without N and C addition was used as a control. The jars were fastened airtight and incubated in a growth chamber at 30°C. The samples' moisture was periodically adjusted to the value of field capacity (27% v/w). The CO₂ efflux from soil was measured at 1, 2, 4, 5, 10, 13, 18, 30, 40, 50, 60, 70, 80, and 90 days, using 10 mL of 1N NaOH solutions as the captured solution. At all sampling time, 10 g soil was extracted with 50 mL of 0.5 M K₂SO₄ before and after fumigation with chloroform. The extracts were titrated with Fe (NH₄)₂(SO₄) 0.2 N to determine the amount of microbial biomass carbon during the 90 day incubation period, according to (Anderson and Domsch, 1978). For available nitrogen determinations, 10 g of fresh soil were extracted with 2 M KCl, distilled with Kjeldahl instrument, the nitrogen was received at 3% boric acid. Standard H₂SO₄ 0.01N were used to back titrate boric acid and have mineral nitrogen content (NH₄⁺ and NO₃⁻) for pH and EC determination suspension of (1:5) soil: water was made, Orion Star (A211) pH meter was used to determine soil pH. Electrical conductivity (EC) was measured in the filtrated extracts using an EC meter (YSI, USA). Finally, for available phosphorus and micronutrients, an extract of 0.005 M ammonium bicarbonate (DTPA) was prepared, and the inductivity coupled plasma optical emission spectrophotometer (ICP-OES, PerkinElmer Optima 4300 DV, USA) were used to estimate the available concentration of extracting solution.

2.4. Statistical analysis

The presented data are averages of three replicates. The measured soil chemical properties, carbon efflux rate, and microbial carbon were statistically compared using Duncan's multiple range test. Nutrient release from different organic amendments at different time intervals was statistically analyzed by completely randomized design under two (days of incubation and organic amendment) factorial arrangement. Statistical analysis of the data, simple correlation, and regression analyses were performed using SPSS 19.0 software. The significance test was conducted at 5 and 1% level of significance ($p \leq 0.05$ and $p \leq 0.01$).

3. Results and Discussion

3.1. Experimental Soil

The soil used in this study was agricultural soil with sandy loam texture, pH of 8.5; electrical conductivity of 0.6 dS m⁻¹, total carbon 4.8%, total nitrogen (N) content of 0.1% and calcium carbonate content was 29.9%.

3.1.1. Soil pH

There were significant differences ($p \leq 0.01$) in soil pH among the organic amendments and incubation time. Almost all treatments increased pH after seven days of incubation except for CMB (Figure1). The highest pH value was 9.7, which was obtained by CMSAE (pH12&T95°C), while the lowest pH was 8, which was obtained by CMSAE

(pH9&T95°C). The soil pH of CM, CMC CMB, and CMSAE (pH9&95°C) was lower than form 7 days was increased and the pH was stable afterward. Higher pH values of CMSAE (pH12&T95°C) could be explain by the addition of more KOH before ammonia stripping, compared to CMSAE (pH9&T95°C). At the start, the CMB had the highest pH. The increase of basic metals in the biochar might be the reason to raise soil pH after biochar application (Kookana et al., 2011), (Streubel et al., 2011) reported 0.8 to 0.1 unit rise in the soil pH as a result of biochar application to the soil.

Soil pH after 20 days tends to increase then fall at 60 days except for CMC, decreasing earlier at 20 days incubation time. This was also reported by McCauley et al., (2009) they stated that the cation exchange capacity increases as a result of organic matter decomposition, therefore increasing the soil buffering capacity. The high buffering capacity of the soil increased by the increasing decomposition of organic material, releasing of OH⁻ and CO₂ and pH increased. Hence the lack of effect of the organic materials on soil pH may most likely be approved by the findings of Gramss et al., (2003). At 90 days pH tends to equilibrate, while row caw manure and by CMSAE (pH12 and T 95°C) tend to increase. The second pH increase could be attributed to the breakdown of organic matter, which results in H⁺ ions to be released into the soil from the functional groups which led to pH decrease (Van Ranst, 2006).

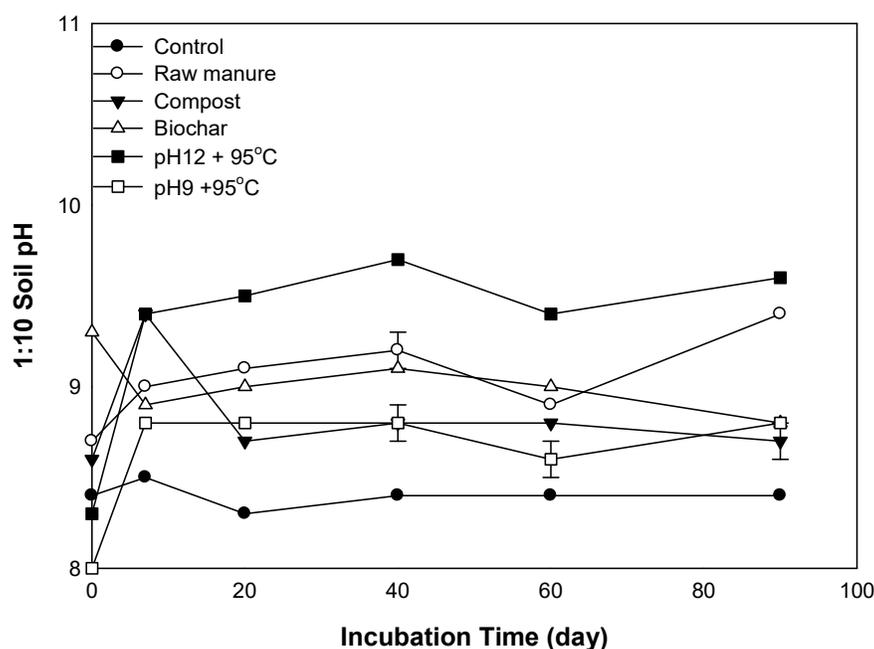


Figure 1. Variation in soil pH treated with different organic amendments derived from cow manure at incubation times.

3.1.2. soil EC

There was a statistically significant difference in soil EC due to the addition of the different amendments. The incubation time did not affect soil EC. Application of various organic amendments resulted in an initial rise in soil EC followed by steady afterward. All amendments EC were less than two dS m⁻¹ except for CMC of 3 dS m⁻¹ (Figure 2). The soil incubated with CMC showed the highest EC. The soil EC of CMB was increased during 0- 40 days. The CMSAEs showed less EC as compared to CMC but similar to that of the CMB. Higher CM soil EC related to initial EC. The increase of soil salinity due to cow manure application was reported by Hao and Chang (2003). They considered manure as an essential source of soil salinity. The same observation was reported by Goff (2006), who found that huge manure quantities induced soil salinity. These quantities increased salt

content and soil EC to be increased. CM can increase salinity due to the presence of water-soluble nutrients ammonium, Na, Ca, Mg, K, Cl, SO_4 , and HCO_3 and the use of nutritional salts (NaCl).

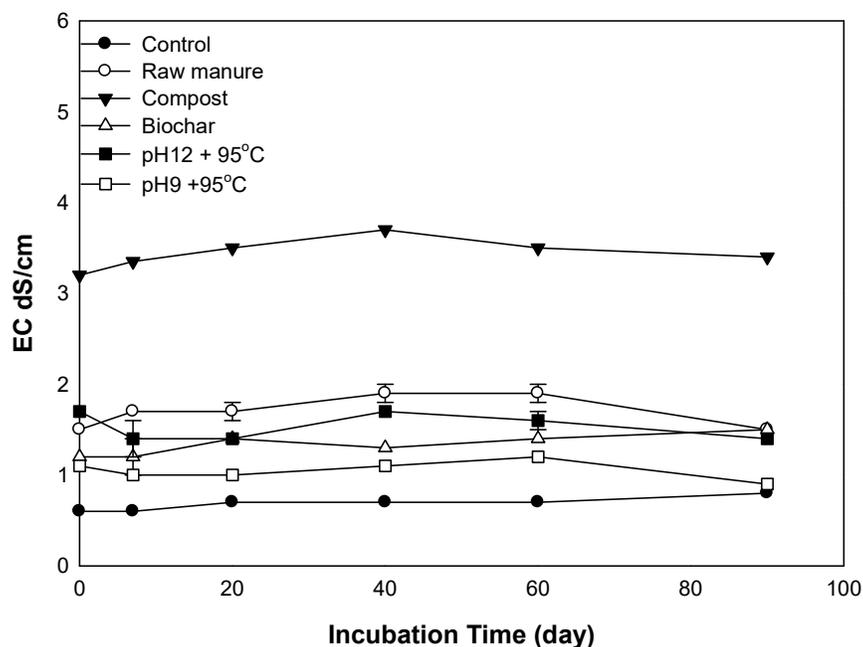


Figure 2. Variation in soil EC as affected by different organic amendments derived from cow manure at different incubation time.

3.1.3. Available phosphorus

The results for available phosphorus (P) affected by different organic amendments derived from CM were shown in Figure 3. There was a statistically significant difference in available P as a result of organic amendments and incubation time. The available P in soils treated with CM, CMC and CMSAE (pH12&T95°C) decreased between 0 to 7 days and then tended to increase as the incubation time increases. This decrease may result from $CaCO_3$, which is reported to retain P as dicalcium phosphate and octacalcium phosphate (Von Wandruszka, 2006). Soil treated with CMSAE (pH12&T95°C) showed the highest available P between days 20 to 90. This increase in soil available P might be due to the high humic acid content in manure effluents compared to CM and CMB, as found by (Delgado et al., 2002). They indicated that the presence of humic acid slows the precipitation of poorly soluble Ca phosphates.

There were irritating results for P as general this could be explained as P adsorption, and desorption biochar and other carbonaceous materials are governed mainly by soil pH (Initiative, 2012). In addition, P availability in soil is controlled by Ca and Mg ions under high pH conditions. Thus, the decline in the availability of P mainly qualified to the high content of $CaCO_3$ and high soil pH by forming less soluble compounds.

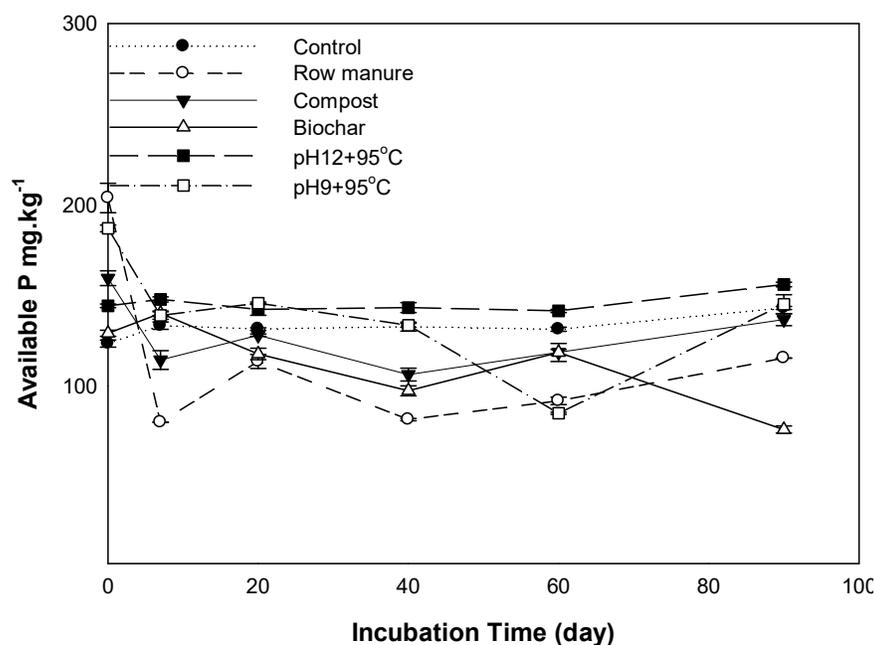


Figure 3. Available phosphorus is affected by different organic amendments derived from cow manure at incubation times.

3.1.4. Available iron

There was a significant difference in soil available Fe as a result of organic amendments addition. The Fe declined as incubation time decreased (Figure 4). The Fe content values are plotted in two areas: sharp Fe decrease in CMC and CMB, gradual Fe decrease: in CMSAEs and CM. After 60 days, Fe content depleted to the lowest value for all treatments. The Fe content of soils treated with CM and CMSAEs was higher compared to other amendments. This may be related to the high Fe content of CM, and CMASEs had high humic substance content that may contribute to Fe availability via the formation of water-soluble Fe- Humic substance complexes, which quickly move in the soil (García-Mina et al., 2004). Uptake of ^{59}Fe from ^{59}Fe -complex was measured even at pH values compatible with those found in calcareous soils (Colombo et al., 2012). Although calcareous soils tend to reduce Fe availability due to high pH in this soil (Abadía et al., 2011; Bavaresco and Poni, 2003). The high pH increases the hydroxyl functional group, and subsequently, Fe hydroxide precipitated (Zeng et al., 2014).

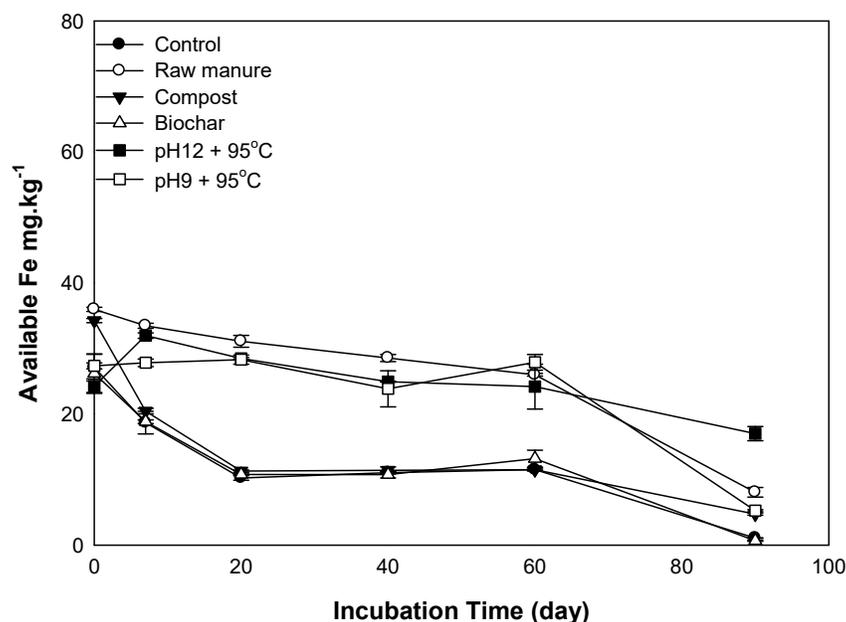


Figure 4. Soil available iron as affected by different organic amendments derived from cow manure at different incubation times.

3.1.5. Available copper

There was a significant difference in Cu content as result of organic amendments. The Cu content declined with increasing incubation time. The Cu values were also plotted in two areas: a sharp decrease in CMC and CMB, a gradual decrease in CM and CMSAEs. The Cu content of 0 to 20 days decreased in all amendments with the exception of CM; afterward, Cu contents were almost stable until 60 days, and then its content decreased at 90 days (Figure 5). The highest Cu content at start explained by low soil pH as showed in (Figure1). The Cu content CMSAEs was higher than CMC and CMB; the reason behind this was the temperature of CMSAEs preparation, which promoted Cu containing tissue of CM to degradation, consequently produce more Cu in soil solution (Aaltonen et al., 2019).

(Cuske et al., 2013) stated copper could be binding to low molecular weight organic acids into a shape of movable complexes. The low Cu values at the end of the experiment might be due to the buffering capacity of this soil.

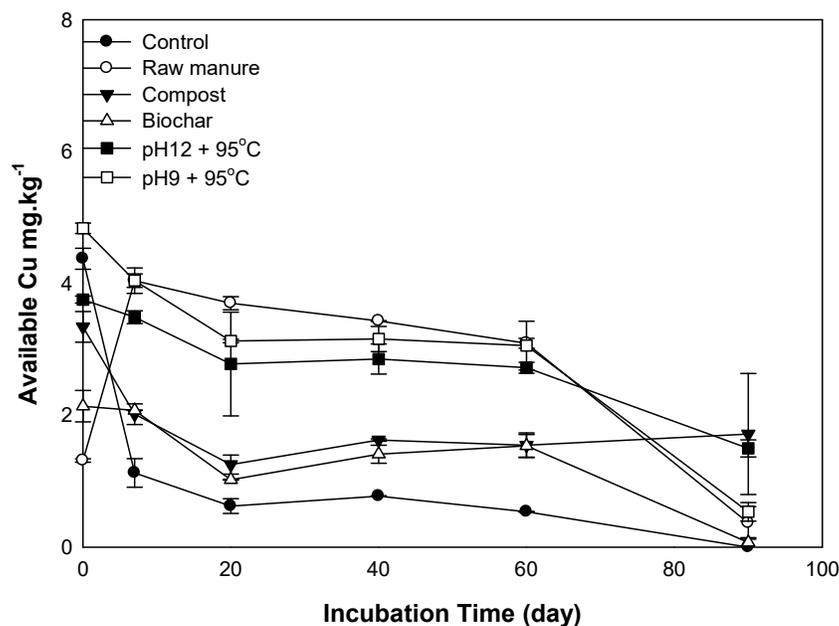


Figure 5. Soil available copper as affected by different organic amendments derived from cow manure at different incubation times.

3.1.6. Available zinc

There were significant differences in Zn content was observed as a result of organic amendments application. Zn content generally declined with increasing incubation time (Figure 6). The highest recorded Zn content was (39.7 mg kg^{-1}) for CMC treatment, while the lowest Zn content was (1.1 mg kg^{-1}) in control soils after 90 days. Higher soil pH and CO_3 content negatively affected Zn content and decreased its availability. The results of (Smith, 2009) showed that aerobic composting processes increase the heavy metals complex with organic materials by forming strongly bound to the compost, affecting their solubility.

High pH values increase soil hydroxyl functional groups, and subsequently, zinc hydroxide and zinc carbonate are expected to be precipitate and reduce Zn availability (Zeng et al., 2014).

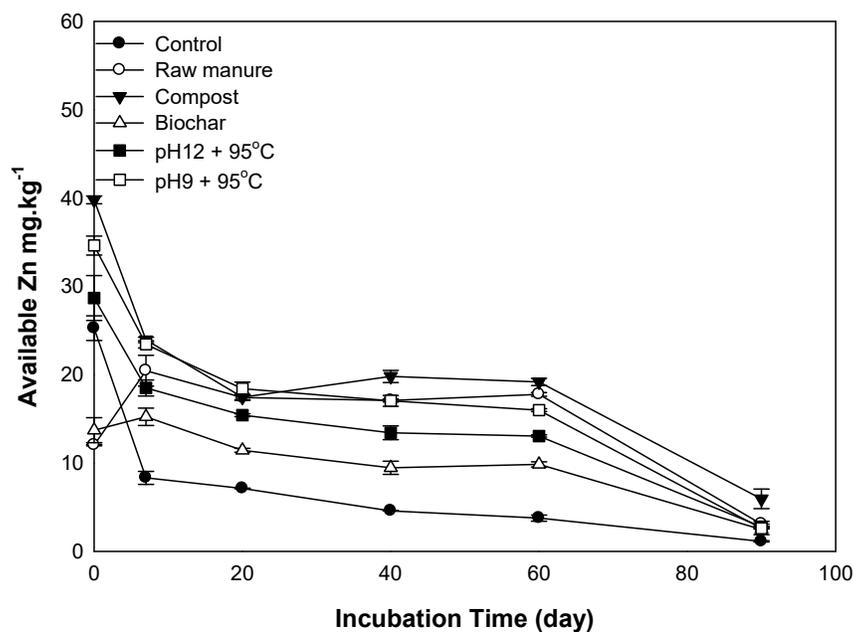


Figure 6. Soil available Zinc as affected by different organic amendments derived from cow manure at different incubation times.

3.1.7. Available manganese

Higher Mn content was observed at the start of the incubation. Afterward, Mn concentration tends to sharply decrease for all organic amendments in 20 days, except for a gradual decline for CMSAE (pH9&T95°C), which reached 9.6 mg kg⁻¹ at 40 days. The highest Mn content was 86 mg kg⁻¹ which was observed at CMSAE (pH9&T95°C), while the lowest Mn concentration was 0.38 mg kg⁻¹ which was observed at CMB with 90 days. Mn concentration was reduced in CMB three times than CMSAE (pH9&T95°C) at seven days incubation time. At the same time, its content decreased by (1:8) ratio at 20 days (Figure 7). Low Mn content may have resulted from the mass losses that happened after the decomposition of organic matter (Ingelmo et al., 2012). Similar results were also obtained by (Zeng et al., 2014), who found the Mn content was below the critical level when soil pH was increased.

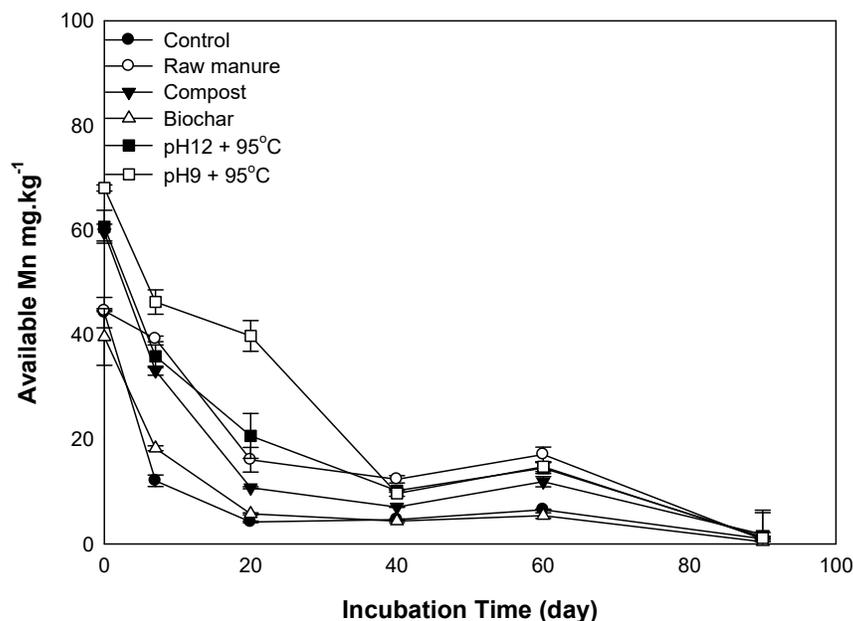


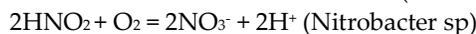
Figure 7. Soil available Mn as affected by different organic amendments derived from cow manure at different incubation times.

3.2. Nitrogen dynamic

3.2.1. Available nitrate

Nitrate -nitrogen is considered one of the measurements for soil nitrogen availability. The results showed that the addition of different organic materials caused an increase in the KCl extract NO_3^- of nitrogen concentrations as incubation time increased. The maximum nitrification was recorded at 60 and 90 days after incubation. The maximum NO_3^- accounted for 1124 ± 4.2 and 870 ± 2 mg kg^{-1} soil for CM and CMSAE (pH12 + 95°C) respectively (Figure 8).

Generally incubation time increased NO_3^- content. After 20 days increase in soil NO_3^- might be related to biological oxidation of ammonia to nitrate through nitrification process (Grunditz and Dalhammar, 2001) and (Vadivelu et al., 2007)



Environmental factors that affect ammonification and nitrification are temperature, water status, C: N ratio (Agehara and Warncke, 2005); (Kampschreur et al., 2007). Ammonium nitrification could be a reason for the increase in NO_3^- the content of soil incubated with an organic compound. The rise in nitrification at 60 days was also found by Usman et al. (2013) stated that the poultry manure addition to soil resulted in NO_3^- accumulation at 60 days. This was mainly due to the increased activity of nitrifying bacteria. These findings agree with those results found by Preusch et al. (2002), who found that the ammonium content extracted from soil treated with fresh poultry litter declined during the first 30 days of incubation there was a rapid increase in NO_3^- contents.

Also, high soil pH could be a reason for the inhibition of nitrifying bacteria growth, resulting in incremental NO_3^- content (Pichtel, 1990). It is essential to highlight the importance of NO_3^- management while applying cow manure to prevent NO_3^- pecculation through the soil profile. The CMC and CMSAE (pH12 + 95°C) showed the highest NO_3^- at incubation. This increase is due to the appreciable content of NO_3^- in CMC. Paul and Beauchamp (1994) found that the soil amended with CMC contains more than 206 mg N per kg soil compared to fresh CM. Hence, if CM is added to the soil under field conditions,

we suggest that the best management practice would be to avoid the environmental risk of $\text{NO}_3\text{-N}$ leaching.

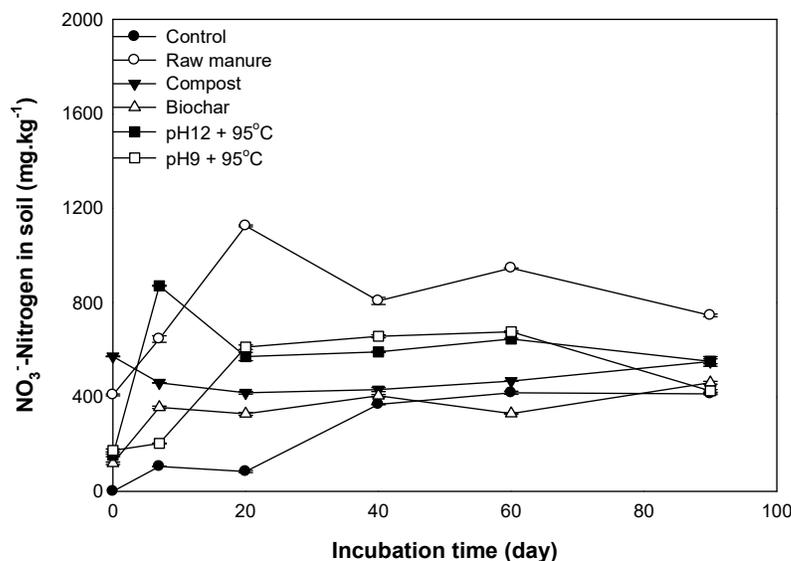


Figure 8. Available $\text{NO}_3\text{-N}$ (mg.kg^{-1}) as affected by different organic amendments derived from cow manure at different incubation time.

3.2.2. Available ammonium

Ammonium nitrogen is considered one of the measurements for soil nitrogen availability to plants. The results showed that the addition of different organic materials caused a significant difference in extracted NH_4^+ nitrogen content. At zero time, the available ammonium was relatively high, as much as incubation time, increased the NH_4^+ content tend to decrease, these results point out the added manures increased available NH_4^+ . Extracted ammonium increased immediately after manure applications, which was also found by Whalen et al., (2000), who reported a concentration of 1100 mg.kg^{-1} ammonium following manure application. Microbes quickly degrade simple organic compounds such as simple carbohydrate and amino acids in a short time. Therefore, soil available ammonium increases (Cai et al., 2019).

Over time, ammonium content decreased up to 60 days. This reduction was mainly due to the nitrification process where ammonium is converted to NO_3^- as stated by (Hampel et al., 2018). The NH_4^+ content was high due to CM addition results showed by (Nyawade et al., 2019) had the same manner when organic matter was added to cultivated soil. This may be due to dissolved organic matter, which increased the ammonium content (Haroon et al., 2018). These findings are consistent with those of Preusch et al., (2002); they found that the $\text{NH}_4\text{-N}$ concentration extracted from soil treated with fresh manure declined during the first 30 days of incubation, but that there was a rapid increase in $\text{NO}_3\text{-N}$ concentrations. $\text{NH}_4\text{-N}$ immobilization by soil microbes and/or N losses such as ammonia volatilization might also be responsible for the rapid decrease in net N mineralization after 20 days of incubation (Wang et al., 2018). The reduction in ammonium content may also be attributed to wide C/N, as stated by (Kirckner et al., 1993). Also (Sharifnia et al., 2016) noted that the retention of ammonium into the negative charge of clay might be attributed to lower NH_4^+ content.

The maximum NH_4^+ content was observed at 90 days, with the exception of CMC and CMB (Figure 9). This seconded increase may be explained by the decomposition of the resistant material of CM and CMSAE such as protein, amino acids hemicelluloses (Narita et al., 2005) and produce NH_4^+ .

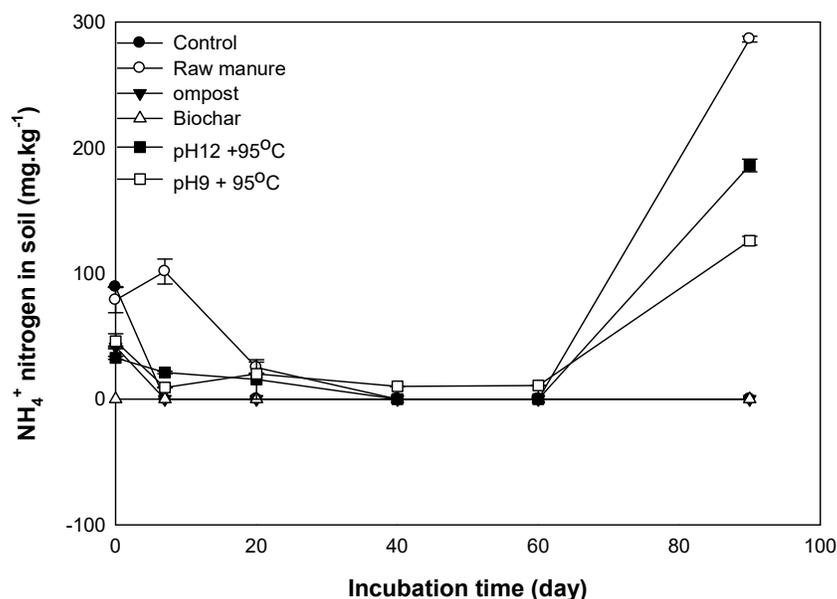


Figure 9. Available NH_4^+ mg kg^{-1} as affected by different organic amendments derived from cow manure at different incubation time.

3.3. Changes in microbial biomass carbon (MBC)

Microbial biomass carbon considered as a measurement for living component mass of the soil organic matter. The microbial biomass carbon plays a huge role in availability and transformation of soil organic matter and plants nutrients. (Figure 10) shows the effect organic materials and incubation time on microbial biomass carbon for 90 days. Most treatments showed increase of MBC at 7 days with exception of CMB and CMSAE (pH 9 and 95°C). The CMSAE produced at pH12 showed the highest MBC at 7 days of incubation, the value was $11481 \pm 61 \text{ mg kg}^{-1}$. The MBC for control and CMB were the lowest as compare to other treatments.

The increase of MBC indicated the presence of inorganic nitrogen and consequently high microbial activity. Studies by Jara-Samaniego et al., (2017) showed the positive effect of organic materials addition which led to increase the MBC. The soil microorganisms largely depend on easy degradable materials to increase their numbers and decompose more organic material. After 20 days of incubation MBC content decline, this probably due to consuming soluble organic carbon by microbes. The CM reported to have amino-sugars, amino-acids, proteins and short-chain organic acids (Morvan and Nicolardot, 2009) that were reported to decompose fast in soil (Glanville et al., 2012 and Kögel-Knabner, 2002). The lowest Biochar MBC might be attributed to preparation temperature caused loss of many organic compounds that are consumed by microorganisms (Jindo et al., 2012).

The second stage of MBC have also been observed at 60 days it's also found at in other studies on sheep dung decomposition, and it is proposed that in the first peak of CO_2 emission (high MBC) is due to the decomposition of labile C from soil and easily degradable dung fractions, while in the second phase, the decomposers attack more recalcitrant material (Ma et al., 2013) so the MBC increased.

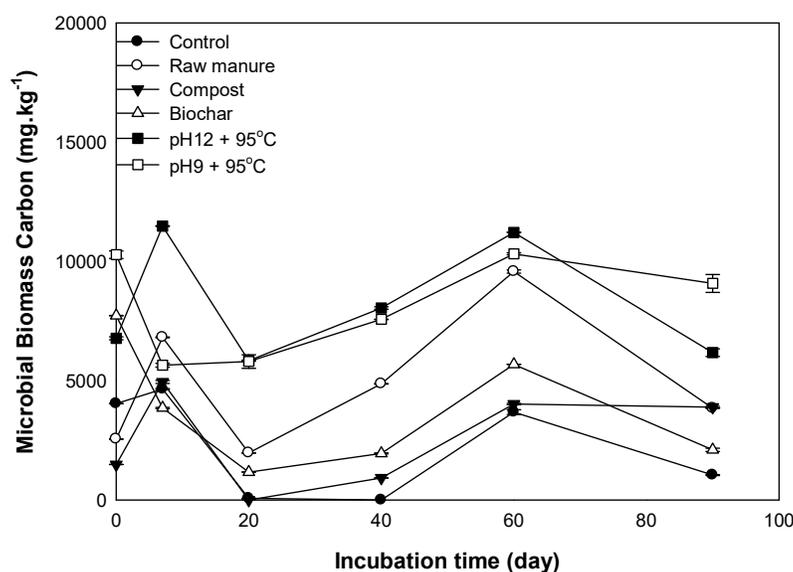


Figure 10. Extracted potassium sulfate microbial biomass carbon from soil as affected by different organic amendments derived from cow manure at different incubation time.

3.4. Changes in CO₂ efflux and cumulative CO₂

The maximum CO₂ efflux values were observed at the first seven days, after 20 days the values of the CO₂ efflux was decreased with increasing incubation time. The high values of CO₂ flux at start may be explained by addition of organic manures provide the soil with variety of essential nutrient for soil microorganisms activity and growth. The soil CO₂ efflux ranged from 0.231- 0.001 g kg⁻¹ soil. The high CO₂ fluxes were observed in the CM it was 0.231 g kg⁻¹ soil (Figure 11). The organic manures added the high soil pH expected to decrease for instance, although between 0 to 7 days no soil samples were extracted. It's known that the high CaCO₃ conditions and high pH conditions inhibits some of the neutral pH microorganisms growth (Rietz and Haynes, 2003). The addition of organic material in the short term provides additional substrates for the soil microorganisms and resulted in an increase in microorganism's activity. This substrate may also relieve osmotic and pH stress on the soil microorganisms, while improving soil chemical conditions (Chander et al., 1994).

The results reveal that the CMSAEs significantly increased the accumulated CO₂ efflux (Figure 12). The highest cumulative CO₂ flux values of CM almost similar 3.28 g kg soil⁻¹, while the CMB showed the smallest flux value was 2.7 g kg soil⁻¹. Our study thus clearly shows that the CMSAE didn't decrease the accumulative CO₂ flux.

The low CO₂ flux rate possibly related to the low levels of MBC from 20 to 40 days. The previous studies found that the microbial population densities in the initial days at organic matter addition were higher as compared to the final days (Martin-Olmedo and Rees, 1999). It was observed that the CMB and CMC were proved to have less CO₂ efflux as compared to the CMSAE this is properly due to the high C/N ratio of these materials as well illustrated in literature (Cayuela et al., 2013; Huang et al., 2004). It was very clear that the CMSAEs efflux values were higher as compared to the other organic materials including the CM which expected to have low CO₂ flux, the reason behind this is mainly because the row manure was collected from the barn floor which is differ than that used to prepare CMSAE (fresh manure), furthermore the preparation temperature at ammonia string process may be attributed to promote the CMSAEs decomposition when added to the soil. The increase in MBC of soil treated with different organic material enhanced microbial activity.

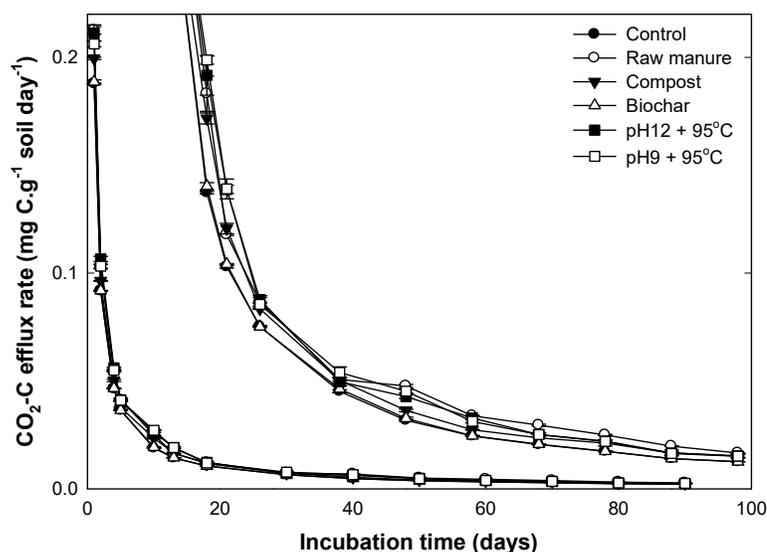


Figure 1. CO₂ efflux rate (mg C/g soil/day) as affected by different organic amendments derived from cow manure at different incubation time.

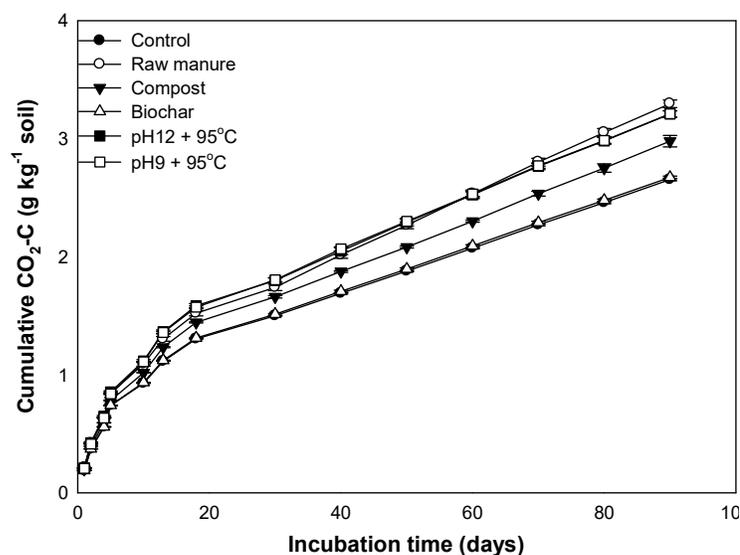


Figure 2. Cumulative CO₂ efflux (g/kg soil) as affected by different organic amendments derived from cow manure at different incubation time.

4. Conclusions

In this experiment, a calcareous sandy loam soil was treated with different organic amendments derived from cow manure for 90 days. The results showed that the addition of different organic materials caused an increase in the available NO₃⁻ nitrogen contents after 20 days of incubation time and decreased the available NH₄⁺ concentration after seven days from the incubation start. MBC was significantly increased ($p \leq 0.05$) as the incubation time increased. The cow CMSAE produced at pH12 and 95°C, CM and CMC showed the highest MBC; these values accounted for 11481 ±61 and 6815 ±14 mg kg⁻¹, respectively. At the same time, control and CMB reached their lowest MBC values after seven days.

Furthermore, the soil CO₂ efflux ranged from 0.231- 0.001 g kg⁻¹ soil. The high CO₂ efflux was observed in the raw CM and CMSAEs of pH12 and T95°C; the values accounted

for 0.231 and 0.211, respectively. Moreover, the results reveal that the CMSAEs at pH 12 and pH 9 with 95°C decreased the accumulated CO₂ efflux, while the CMB showed the smallest efflux value (19.8). Our study thus clearly indicates that the CMSAE didn't reduce the accumulative CO₂ efflux. Finally, the organic materials significantly affected the soil micronutrient cations (Fe, Cu, Zn and Mn). The contents of available micronutrients were high at the beginning of incubation then decreased over time.

Acknowledgments: This Work was funded by the National Plan for Science, Technology and Innovation (MAARIFAH), King Abdul-Aziz City for Science and Technology, Kingdom of Saudi Arabia, grant Number ENV-1970-02

Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study, in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

References

1. Aaltonen, H., Palviainen, M., Zhou, X., Köster, E., Berninger, F., Pumpanen, J., and Köster, K. (2019). Temperature sensitivity of soil organic matter decomposition after forest fire in Canadian permafrost region. *Journal of Environmental Management* 241, 637-644.
2. Abadía, J., Vázquez, S., Rellán-Álvarez, R., El-Jendoubi, H., Abadía, A., Álvarez-Fernández, A., and López-Millán, A. F. (2011). Towards a knowledge-based correction of iron chlorosis. *Plant Physiology and Biochemistry* 49, 471-482.
3. Agehara, S., and Warncke, D. (2005). Soil moisture and temperature effects on nitrogen release from organic nitrogen sources. *Soil Science Society of America Journal* 69, 1844-1855.
4. Al-Wabel, M. I., Al-Omran, A., El-Naggar, A. H., Nadeem, M., and Usman, A. R. A. (2013). Pyrolysis temperature induced changes in characteristics and chemical composition of biochar produced from conocarpus wastes. *Bioresource Technology* 131, 374-379.
5. Ameloot, N., Sleutel, S., Das, K. C., Kanagaratnam, J., & De Neve, S. (2015). Biochar amendment to soils with contrasting organic matter level: effects on N mineralization and biological soil properties. *Gcb Bioenergy*, 7, 135-144.
6. Anderson, J., and Domsch, K. (1978). A physiological method for the quantitative measurement of microbial biomass in soils. *Soil biology and biochemistry* 10, 215-221.
7. Bavaresco, L., and Poni, S. (2003). Effect of calcareous soil on photosynthesis rate, mineral nutrition, and source-sink ratio of table grape. *Journal of plant nutrition* 26, 2123-2135.
8. Bouwman, A. F., and Van Der Hoek, K. W. (1997). Scenarios of animal waste production and fertilizer use and associated ammonia emission for the developing countries. *Atmospheric Environment* 31, 4095-4102.
9. Cai, Y., He, Y., He, K., Gao, H., Ren, M., and Qu, G. (2019). Degradation mechanism of lignocellulose in dairy cattle manure with the addition of calcium oxide and superphosphate. *Environmental Science and Pollution Research*.
10. Cayuela, M. L., Sánchez-Monedero, M. A., Roig, A., Hanley, K., Enders, A., and Lehmann, J. (2013). Biochar and denitrification in soils: when, how much and why does biochar reduce N₂O emissions? *Scientific reports* 3, 1732.
11. Chander, K., Goyal, S., and Kapoor, K. (1994). Effect of sodic water irrigation and farmyard manure application on soil microbial biomass and microbial activity. *Applied Soil Ecology* 1, 139-144.
12. Colombo, C., Palumbo, G., Sellitto, V. M., Rizzardo, C., Tomasi, N., Pinton, R., and Cesco, S. (2012). Characteristics of insoluble, high molecular weight iron-humic substances used as plant iron sources. *Soil Science Society of America Journal* 76, 1246-1256.
13. Creed, J. T., Brockhoff, C. A., and Martin, T. D. (1994). Method 2008: Determination of trace elements in waters and wastes by inductively-coupled plasma-mass spectrometry. Environmental Monitoring Systems Laboratory, Office of Research and Development, US Environmental Protection Agency, Cincinnati, OH, Rev, 5.
14. Cuske, M., Gersztyn, L., and Karczewska, A. (2013). The influence of pH on solubility of copper in soils contaminated by copper industry in Legnica. *Civil and Environmental Engineering Reports*.
15. Delgado, A., Madrid, A., Kassem, S., Andreu, L., and Del Campillo, M. D. C. (2002). Phosphorus fertilizer recovery from calcareous soils amended with humic and fulvic acids. *Plant and Soil* 245, 277-286.
16. Dere, A. L., and Stehouwer, R. C. (2011). Labile and stable nitrogen and carbon in mine soil reclaimed with manure-based amendments. *Soil Science Society of America Journal* 75, 890-897.
17. Dubrovsky, N. M., and Hamilton, P. A. (2010). "Nutrients in the Nation's streams and groundwater: National Findings and Implications." US Geological Survey.
18. Estefan, G. (2017). *Methods of Soil, Plant, and Water Analysis: A manual for the West Asia and North Africa Region*. International Center for Agricultural Research in the Dry Areas (ICARDA).
19. FAO (2014). "Agriculture, Forestry and Other Land Use Emissions by Sources and Removals by Sinks: 1990-2011 Analysis," The Food and Agriculture Organization of the United Nations, Rome, Italy, Viale delle Terme di Caracalla 00153 Rome, Italy.
20. Garcia-Mina, J., Antolin, M., and Sanchez-Diaz, M. (2004). Metal-humic complexes and plant micronutrient uptake: a study based on different plant species cultivated in diverse soil types. *Plant and Soil* 258, 57-68.

21. Glanville, H., Rousk, J., Golyshin, P., and Jones, D. (2012). Mineralization of low molecular weight carbon substrates in soil solution under laboratory and field conditions. *Soil Biology and Biochemistry* 48, 88-95.
22. Goff, J. P. (2006). Macromineral physiology and application to the feeding of the dairy cow for prevention of milk fever and other periparturient mineral disorders. *Animal feed science and technology* 126, 237-257.
23. Gramss, G., Voigt, K. D., and Bergmann, H. (2004). Plant availability and leaching of (heavy) metals from ammonium-, calcium-, carbohydrate-, and citric acid-treated uranium-mine-dump soil. *Journal of Plant Nutrition and Soil Science* 167, 417-427.
24. Grunditz, C., and Dalhammar, G. (2001). Development of nitrification inhibition assays using pure cultures of *Nitrosomonas* and *Nitrobacter*. *Water research* 35, 433-440.
25. Hampel, J. J., McCarthy, M. J., Gardner, W. S., Zhang, L., Xu, H., Zhu, G., and Newell, S. E. (2018). Nitrification and ammonium dynamics in Taihu Lake, China: seasonal competition for ammonium between nitrifiers and cyanobacteria. *Biogeosciences* 15, 733-748.
26. Hao, X., and Chang, C. (2003). Does long-term heavy cattle manure application increase salinity of a clay loam soil in semi-arid southern Alberta? *Agriculture, Ecosystems & Environment* 94, 89-103.
27. Haroon, B., Abbasi, A., Faridullah, An, P., Pervez, A., and Irshad, M. (2018). Chemical Characterization of Cow Manure and Poultry Manure after Composting with Privet and Cypress Residues. *Communications in Soil Science and Plant Analysis* 49, 2854-2866.
28. Huang, G. F., Wong, J., Wu, Q., and Nagar, B. (2004). Effect of C/N on composting of pig manure with sawdust. *Waste management* 24, 805-813.
29. Initiative, I. (2012). Standardized product definition and product testing guidelines for biochar that is used in soil. IBI biochar Stand.
30. Jara-Samaniego, J., Pérez-Murcia, M., Bustamante, M., Pérez-Espinosa, A., Paredes, C., López, M., López-Lluch, D., Gavilanes-Terán, I., and Moral, R. (2017). Composting as sustainable strategy for municipal solid waste management in the Chimborazo Region, Ecuador: Suitability of the obtained composts for seedling production. *Journal of cleaner production* 141, 1349-1358.
31. Jindo, K., Sánchez-Monedero, M. A., Hernández, T., García, C., Furukawa, T., Matsumoto, K., Sonoki, T., and Bastida, F. (2012). Biochar influences the microbial community structure during manure composting with agricultural wastes. *Science of the total environment* 416, 476-481.
32. Kampschreur, M. J., Tan, N. C., Kleerebezem, R., Picioreanu, C., Jetten, M. S., and Loosdrecht, M. C. v. (2007). Effect of dynamic process conditions on nitrogen oxides emission from a nitrifying culture. *Environmental science & technology* 42, 429-435.
33. Kirckner, M., Wollum, A., and King, L. (1993). Soil microbial populations and activities in reduced chemical input agroecosystem. *J. Soil Sci. Am. Society* 57, 1289-1295.
34. Kögel-Knabner, I. (2002). The macromolecular organic composition of plant and microbial residues as inputs to soil organic matter. *Soil biology and biochemistry* 34, 139-162.
35. Kookana, R. S., Sarmah, A. K., Van Zwieten, L., Krull, E., and Singh, B. (2011). Biochar application to soil: agronomic and environmental benefits and unintended consequences. In "Advances in agronomy", Vol. 112, pp. 103-143. Elsevier.
36. Kowalska, M., Güler, H., and Cocke, D. L. (1994). Interactions of clay minerals with organic pollutants. *Science of The Total Environment* 141, 223-240.
37. Kuzyakov, Y. (2006). Sources of CO₂ efflux from soil and review of partitioning methods. *Soil Biology and Biochemistry* 38, 425-448.
38. Lehmann, J. (2007). A handful of carbon. *Nature* 447, 143-144.
39. Ingelmo, F., Molina, M. J., Soriano, M. D., Gallardo, A., and Lapeña, L. (2012). Influence of organic matter transformations on the bioavailability of heavy metals in a sludge based compost. *Journal of environmental management* 95, S104-S109.
40. Ma, X., Ambus, P., Wang, S., Wang, Y., and Wang, C. (2013). Priming of soil carbon decomposition in two inner mongolia grassland soils following sheep dung addition: a study using ¹³C natural abundance approach. *PloS one* 8, e78578.
41. Martin-Olmedo, P., and Rees, R. (1999). Short-term N availability in response to dissolved-organic-carbon from poultry manure, alone or in combination with cellulose. *Biology and Fertility of Soils* 29, 386-393.
42. McCauley, A., Jones, C., and Jacobsen, J. (2009). Soil pH and organic matter. *Nutrient management module* 8, 1-12.
43. Morvan, T., and Nicolardot, B. (2009). Role of organic fractions on C decomposition and N mineralization of animal wastes in soil. *Biology and Fertility of Soils* 45, 477-486.
44. Narita, H., Zavala, M. A. L., Iwai, K., Ito, R., and Funamizu, N. (2005). Transformation and characterisation of dissolved organic matter during the thermophilic aerobic biodegradation of faeces. *Water research* 39, 4693-4704.
45. Nyawade, S. O., Karanja, N. N., Gachene, C. K., Gitari, H. I., Schulte-Geldermann, E., and Parker, M. L. (2019). Short-term dynamics of soil organic matter fractions and microbial activity in smallholder potato-legume intercropping systems. *Applied Soil Ecology*.
46. Paré, T., Dinel, H., Schnitzer, M., & Dumontet, S. (1998). Transformations of carbon and nitrogen during composting of animal manure and shredded paper. *Biology and fertility of soils*, 26, 173-178.
47. Paul, J., and Beauchamp, E. (1994). Short-term nitrogen dynamics in soil amended with fresh and composted cattle manures. *Canadian Journal of Soil Science* 74, 147-155.
48. Pichtel, J. (1990). Microbial respiration in fly ash/sewage sludge-amended soils. *Environmental Pollution* 63, 225-237.
49. Pinder, R. W., Strader, R., Davidson, C. I., and Adams, P. J. (2004). A temporally and spatially resolved ammonia emission inventory for dairy cows in the United States. *Atmospheric Environment* 38, 3747-3756.

50. Preusch, P., Adler, P., Sikora, L., and Tworkoski, T. (2002). Nitrogen and phosphorus availability in composted and uncomposted poultry litter. *Journal of Environmental Quality* 31, 2051-2057.
51. Rietz, D., and Haynes, R. (2003). Effects of irrigation-induced salinity and sodicity on soil microbial activity. *Soil Biology and Biochemistry* 35, 845-854.
52. Risse, L., Cabrera, M., Franzluebbers, A., Gaskin, J., Gilley, J. E., Killorn, R., Radcliffe, D., Tollner, W., and Zhang, H. (2006). Land application of manure for beneficial reuse.
53. SAAF (2007). "Space Image Atlas of the Kingdom of Saudi Arabia," Prince Sultan Research Center for Environment, Water and Desert.
54. Sallam, A. S. (2002). Micronutrients status of Mollisols (southwestern mountainous region, Saudi Arabia). *J Saudi Soc Agr Sci* 1, 1-22.
55. Schjøning, P., Thomsen, I. K., Møberg, J. P., de Jonge, H., Kristensen, K., and Christensen, B. T. (1999). Turnover of organic matter in differently textured soils: I. Physical characteristics of structurally disturbed and intact soils. *Geoderma* 89, 177-198.
56. Sharifnia, S., Khadivi, M. A., Shojaeimehr, T., and Shavisi, Y. (2016). Characterization, isotherm and kinetic studies for ammonium ion adsorption by light expanded clay aggregate (LECA). *Journal of Saudi Chemical Society* 20, S342-S351.
57. Sigurnjak, I., Brienza, C., Snauwaert, E., De Dobbelaere, A., De Mey, J., Vaneekhaute, C., ... & Meers, E. (2019). Production and performance of bio-based mineral fertilizers from agricultural waste using ammonia (stripping-) scrubbing technology. *Waste Management*, 89, 265-274.
58. Smith, S. R. (2009). A critical review of the bioavailability and impacts of heavy metals in municipal solid waste composts compared to sewage sludge. *Environment international* 35, 142-156.
59. Steiner, C., Das, K. C., Melear, N., and Lakly, D. (2010). Reducing Nitrogen Loss during Poultry Litter Composting Using Biochar. *Journal of Environmental Quality* 39, 1236-1242.
60. Streubel, J., Collins, H., Garcia-Perez, M., Tarara, J., Granatstein, D., and Kruger, C. (2011). Influence of contrasting biochar types on five soils at increasing rates of application. *Soil Science Society of America Journal* 75, 1402-1413.
61. Sutton, M. A., Oenema, O., Erisman, J. W., Leip, A., van Grinsven, H., and Winiwarter, W. (2011). Too much of a good thing. *Nature* 472, 159-161.
62. Usman, A. R., Almaroai, Y. A., Ahmad, M., Vithanage, M., and Ok, Y. S. (2013). Toxicity of synthetic chelators and metal availability in poultry manure amended Cd, Pb and As contaminated agricultural soil. *Journal of hazardous materials* 262, 1022-1030.
63. Vadivelu, V., Keller, J., and Yuan, Z. (2007). Free ammonia and free nitrous acid inhibition on the anabolic and catabolic processes of *Nitrosomonas* and *Nitrobacter*. *Water Science and Technology* 56, 89-97.
64. Van Ranst, E. (2006). Properties and management of soils in the tropics. Chapter 2, 31-32.
65. van Vliet, P. C. J., Reijs, J. W., Bloem, J., Dijkstra, J., and de Goede, R. G. M. (2007). Effects of Cow Diet on the Microbial Community and Organic Matter and Nitrogen Content of Feces. *Journal of Dairy Science* 90, 5146-5158.
66. Von Wandruszka, R. (2006). Phosphorus retention in calcareous soils and the effect of organic matter on its mobility. *Geochemical transactions* 7, 6.
67. Wang, X.-L., Park, S.-H., Lee, B.-R., Jeong, K.-H., and Kim, T.-H. (2018). Changes in Nitrogen Mineralization as Affected by Soil Temperature and Moisture. *Journal of the Korean Society of Grassland and Forage Science* 38, 196-201.
68. Whalen, J. K., Chang, C., Clayton, G. W., and Carefoot, J. P. (2000). Cattle manure amendments can increase the pH of acid soils. *Soil Science Society of America Journal* 64, 962-966.
69. Xiang, J., Liu, D., Ding, W., Yuan, J., and Lin, Y. (2015). Effects of biochar on nitrous oxide and nitric oxide emissions from paddy field during the wheat growth season. *Journal of Cleaner Production* 104, 52-58.
70. Yuan, H., Lu, T., Wang, Y., Huang, H., and Chen, Y. (2014). Influence of pyrolysis temperature and holding time on properties of biochar derived from medicinal herb (*radix isatidis*) residue and its effect on soil CO₂ emission. *Journal of Analytical and Applied Pyrolysis* 110, 277-284.
71. Zeng, W., M. Z., H. Z., H. L., Q. X., and F. L. (2014). The effects of soil pH on tobacco growth. *Journal of Chemical and Pharmaceutical Research* 6, 5.