

Article

Marker Assisted Selection of malic-consuming *Saccharomyces cerevisiae* strains for winemaking. Efficiency and limits of a QTL's driven breeding program

Charlotte Vion ^{#1,2}, Emilien Peltier ^{#1,2,3}, Margaux Bernard ^{1,2}, Maitena Muro ^{1,2}, Philippe Marullo ^{*1,2}.

1 Unité de recherche Œnologie EA 4577, USC 1366 INRAE, Bordeaux INP ; ISVV, Villenave d'Ornon, FRANCE

2 Biolauffort, Bordeaux, FRANCE

3 Université de Strasbourg, CNRS, GMGM UMR 7156, Strasbourg, France

* Correspondence: philippe.marullo@u-bordeaux.fr;

Abstract: Background Natural *Saccharomyces cerevisiae* yeast strains exhibit very large genotypic and phenotypic diversity. Breeding programs taking advantage of this characteristic, are widely used for yeast selection in the wine industry, especially in the recent years when winemakers need to adapt their production to climate change. The aim of this work was to evaluate a Marker Assisted Selection (MAS) program to improve malic acid consumption capacity of *Saccharomyces cerevisiae* in grape juice. **Methods** Optimal individuals of two unrelated F1-hybrids were crossed to get a new genetic background carrying many “malic consumer” loci. Then, eleven QTLs already identified were used for implementing the MAS breeding program. **Results** By this way, extreme individuals able to consume more than 70% of malic acid in grape juice were selected. These individuals were tested in different enological matrixes and compared to their original parental strains. They greatly reduced the malic acid content at the end of alcoholic fermentations, they appeared to be robust to the environment and accelerate the ongoing of malo-lactic fermentations by *Oenococcus oeni*. **Conclusions** This study illustrates how MAS can be efficiently used for selecting industrial *Saccharomyces cerevisiae* strains with outlier properties for winemaking.

...

Keywords: Wine yeast; malic acid; pH; breeding; Malo Lactic Fermentation; Marker Assisted Selection

1. Introduction

The yeast *Saccharomyces cerevisiae* is involved in many biotechnological processes including bioethanol, brewery, bakery and wine making [1]. This species displays a wide genetic diversity that generates a high phenotypic variability for many traits of interest. In the context of wine fermentation, the selection of industrial starters is particularly relevant since yeast modulates the final quality of the product [2,3]. Indeed, numerous studies have demonstrated that commercial *S. cerevisiae* strains are different regarding their ability to complete the alcoholic fermentation, their fermentation kinetics and their contribution to the final composition of volatile and non-volatile compounds [4–9].

Wine industry has developed basic genetic selection for making the best of phenotypic variability and proposing optimal starters. Indeed, clonal selection [10] and yeast breeding programs [9,11,12] are nowadays widely used for developing new wine starters; for documented reviews see [2,13,14]. Beside this applicative aspect, the genetic basis of trait variability has been studied for several decades by forward [15,16,25–27,17–24] and reverse [5,28–30] genetic approaches allowing the detection of numerous genes and allelic forms that possibly impact phenotypes of interest. Recently, we reviewed the phenotypic

impact of 153 genes controlling traits of industrial interest [31]. The identification of such allelic variations and the steady decreasing cost of genotyping methods paved the avenue for efficiently combining Marker Assisted Selection (MAS) with classical breeding (cross and segregation). Despite its very efficient use in plant selection [32] and animal breeding [33], MAS has been poorly used in order to optimize fungi. Indeed, only few authors implemented this approach in the past for accelerating breeding programs [34–37]. In that case, alleles used had a high penetrance and controlled a large part of phenotypic variation. However, most of allelic variations have a low penetrance and a small effect on the phenotype [39]. This is particularly true when MAS is implemented in outbred backgrounds [40] due to modifier alleles that drastically reduce QTL expressivity. The aim of this work is to evaluate MAS efficiency for improving a complex quantitative trait in a four-parental genetic background which represents an applied case of yeast breeding program.

We focused our investigation on the L-malic acid consumption by *S. cerevisiae* during the alcoholic fermentation, that is a highly variable trait among wine isolates (Peltier 2018). The final concentration of this organic acid, originally present in grape, strongly contributes to the acidic pH of wine, which has a significant impact on its organoleptic qualities. Therefore, the control of its consumption is important for the choice of winemaking routes. *S. cerevisiae* is unable to consume all the L-malic acid of grape juice due to the lack of an active transporter [44]. Indeed, *S. cerevisiae* strains consuming more than 50% of total L-malic present in a grape juice were never reported [43,45]. During the winemaking process, malic acid can be transformed in L-lactic acid by wine-specific lactic bacteria *Oenococcus oeni* [41]. This biotransformation step, called Malo-Lactic Fermentation (MLF), mostly occurs in red wines after or during the alcoholic fermentation. A part of L-malic acid can be also consumed by the Malo-Ethanolic Fermentation (MEF) as well as by the fumarase activity [42,43].

Recently, the genetic determinism of malic acid consumption by *Saccharomyces cerevisiae* has been partially unraveled [46]. Numerous Quantitative Trait Loci (QTL) related to the malic acid consumption were precisely mapped in the SBxGN background. Additional loci controlling the consumption of malic acid in *S. cerevisiae* have been also reported [47]. The functional exploration of these QTLs reveals the role of seven genes (*MAE1*, *PMA1*, *PYC2*, *PNC1*, *YBL036c*, *PTC5*, and *SDH2*) by reciprocal hemizygosity assay [46,47]. In this present work, a large-scale breeding program was implemented for addressing how much, the rational use of MAS can be implemented for boosting malic acid consumption during the wine alcoholic fermentation. First, optimal individuals of two unrelated F1-hybrids were crossed in order to get a new genetic background carrying many “malic consumer” loci. Eleven unlinked QTLs previously identified were used for implementing a MAS program. By this way, optimal individuals consuming more than 70% of the malic acid of grape juice were obtained. Such individuals were tested in different enological matrixes; they lower the malic acid content at the end of the alcoholic fermentation and accelerate the ongoing of inoculated Malo-Lactic Fermentations.

2. Materials and Methods

2.1 . Yeast strain used and culture methods

Yeast (*Saccharomyces cerevisiae*) were propagated on YPD 2% (1% peptone, 1% yeast extract, 2% glucose) at 28°C in both liquid and plate cultures (2% agar). Long term storage at -80°C was achieved by adding one volume of glycerol to YPD 2% overnight cultures. The main yeast strains used were described in the Table 1 and are derived from the two F1-hybrids (M2xF15 and SBxGN) used in previous QTL mapping studies [26,48]. All the strains are homozygous for the *HO* locus and are therefore diploids. The strains GS-28b and GS-41b are meiotic spore clones of SBxGN while FM-8d is a meiotic spore clone of M2xF15. These three strains were used as allele donors in a breeding program aiming to fix in few individuals most of the alleles promoting the consumption of malic acid during the alcoholic fermentation. The pedigree of the strains generated is shown in Table 1.

Table 1. Main *Saccharomyces cerevisiae* strains used.

Strain	Description	Reference	WDCM-791 (CRB ISVV, Bordeaux)
M2	parental strain, a meiotic spore clone from Enoferm M2 (Lallemand, Canada)	[26]	CRBO L2010
F15	parental strain, a meiotic spore clone from Zymaflore F15 (Laffort, France)	[26]	CRBO L2011
GN	parental strain, a meiotic spore clone from Zymaflore VL1 (Laffort, France)	[48]	CRBO L2002
SB	parental strain, a meiotic spore clone from Zymaflore BO213 (Laffort, France)	[48]	CRBO L2001
M2xF15	F1-hybrid (M2 x F15)	[26]	CRBO L2005
SBxGN	F1-hybrid (SB x GN)	[48]	CRBO L2003
GS-28b	meiotic clone of SBxGN	this work	CRBO L2012
GS-41b	meiotic clone of SBxGN	this work	CRBO L2013
FM-8d	meiotic clone of M2xF15	this work	CRBO L2014
FMGS-1	F1-hybrid (GS-28b x FM-8d)	this work	CRBO L2015
FMGS-1-647	meiotic clone of FMGS-1, malic acid consumer strain	this work	CRBO L2016
FMGS-2	F1-hybrid (GS-41b x FMGS-647)	this work	CRBO L2017
FMGS-1-889	meiotic clone of FMGS-1, malic acid consumer strain	this work	CRBO L2018
FMGS-1-215	meiotic clone of FMGS-1, Malic acid consumer strain	this work	CRBO L2019
FMGS-2-107	meiotic clone of FMGS-2, Malic acid consumer strain	this work	CRBO L2020
FMGS-2-265	meiotic clone of FMGS-2, Malic acid consumer strain	this work	CRBO L2021

2.2 Spore mating and purification

Since all the strains are homothallic, F1-hybrids (FMGS-1 and FMGS-2) were obtained by spore to spore mating on YPD 2% using a routine method previously described [9]. After few hours, zygotes (trilobed shaped) were observed and manually isolated; their heterozygous genotype was verified by Mass Array genotyping (see below). Spore purification was carried out by using the hydrophobic properties of the spore cell wall. Each F1-hybrids was sporulated by cultivating 10⁹ cells in 5-mL of potassium acetate 1% (ACK) during three days at 24°C. This high-density spore culture was digested with cytohelicase (10 mg/mL) for 2 hours at 28°C under permanent shaking. Spore preparation was then washed twice (osmotic water) and the pellet was recovered in 100 µL in one centrifuge tube (1.5 mL). The suspension was vigorously vortexed for 2 minutes and the suspension was removed by pipetting. The remaining mixture of spores and cells was purified by adding/removing 1 mL of osmotic water two times. The remaining spores were unstacked from the tube wall by adding 1 mL of sterile Nonidet P40 (0.01%). The purification rate obtained ranged between 85 and 95% of spore clones. The purified spores were then sonicated (30 min), incubated 2h on YPD (28°C) and plated on several Petri dishes in order to obtained more than 1000 isolated colonies per preparation.

2.3 Cell culture and DNA extraction in microplates

Each single colony picked up from YPD-agar plate was cultivated in 96-Well microplate (Greiner Bio-one, France) coated with gas-permeable sheets (BREATHseal™, Greiner Bio-one, France) allowing CO₂ production. Genomic DNA was extracted by using a Li-Ac SDS protocol previously adapted for microplate handling [49]. Broadly, 5.10⁶ cells (200 µL of overnight cultures) were centrifuged and incubated with 50 µL of 200 mM LiAc/1% SDS at 70°C for 5 min. Genomic DNA was extracted by mixing cell lysates with 150 µL of pure ethanol and vortexed for 15 s. After a brief spin (2 min, 4000 rpm) the pellet

was washed with 70% ethanol and was solubilized in 200 μ L of milliQ water at 60°C for 5 min. Some liquid handling steps were assisted by the robotic platform NxP (Beckman Coulter, France).

2.4 Mass ARRAY genotyping

Genotyping was mostly carried out by using Mass ARRAY technology (Agena Bioscience, CA), which allows large multiplex SNP detection as previously described [50]. Less than 15 ng of DNA were used for genotyping the strain. The Online Resource 1 lists the identity of wanted SNP that possibly enhance the malic acid consumption according to a wide QTL mapping study recently published. SNPs were encoded according to the international nomenclature [51]. The genotypes of each parental strain founder (M2, F15, SB, GN) and the sequence of primers used were also provided. Primers were designed using the tool MassARRAY® Assay Design (version 4.0.0.2) in order to amplify short fragments (<115 bases) with a molecular weight differences ranging between 16 and 80 Da. By this method, 11 bi allelic loci were simultaneously tracked corresponding to the following QTLs (*II_661*, *IV_31*, *IV_360*, *IV_414*, *VII_427*, *VII_480*, *XI_382*, *XI_631*, *XII_53*, *XV_491*, *XV_1052*) which control malic acid consumption in the SBxGN background. The SNP calling was carried out by the MassARRAY® System suite with an average SNP calling of 94.3%. Control strains (SB, GN, F15, M2) were genotyped several times giving in any case the identical SNP calling. All clones showing a fully homozygous genotype and 10 or more loci assigned were included in the analysis, which corresponds to 593 and 797 clones for FMGS-1 and FMGS-2, respectively.

2.5 Alcoholic fermentation assays

2.5.1. Grape must

The grape musts Merlot 2015 (M15) and Sauvignon Blanc 2016 (SB16) used were provided by Vignobles Ducourt (Ladaux, France) and stored at -20°C. Grape musts were sterilized by membrane filtration (nitrate cellulose 0.45 μ m, Millipore, France). In order to test the phenotypic robustness of some relevant strains, these musts were spiked with different amounts of L-malic acid and nitrogen source (Table S2). The mixture of amino acid used mimics the average composition of organic sources in a grape wine as previously described [12]. The composition of the different grape musts in fermenting sugar, nitrogen sources and malic acid content as well their pH are listed in Table S2.

2.5.2. Alcoholic fermentation monitoring

Small-volume alcoholic fermentations were implemented in screwed vials fermentations according to the general procedure described in [52]. Rapidly, 20 mL-screwed vials (Fisher Scientific, France) were filled with 12 mL of filtered grape must and were tightly closed with screw cap-magnetic (Agilent Technologies, hdsp cap 18 mm PTFE / sil 100 pk, France) perforated by hypodermic needles (G26–0.45 x 13 mm, Terumo, Shibuya, Tokyo, Japan) for allowing the CO₂ release. Vessel was inoculated by 2.10^6 viable cell.mL⁻¹ precultured in liquid YPD for 24h. Cellular concentration and viability was estimated by flow cytometry using a Cell Lab Quanta apparatus (Beckman Coulter, France) as described by Zimmer et al. (2014). Fermentation took place at 24°C or 28°C in shaken vials by using an orbital shaker (SSL1, Stuart, Vernon Hills, Illinois, USA) at 175 rpm. Fermentation kinetics were estimated by monitoring manually (2–3 times per day) the weight loss caused by CO₂ release using a precision balance with automatic weight recording (LabX system, Mettler Toledo, France). The amount of CO₂ released according to time was modeled by local polynomial regression fit [52] allowing the estimation of the time necessary to reach the maximum CO₂ produced.

2.6 Malolactic fermentations monitoring

At the end of the alcoholic fermentation, vials were placed for 24 hours at 4°C for settling yeast lees. Sedimented part were then solidified by soaking vials in a thin layer of liquid nitrogen. Thus, yeasts were eliminated in order to avoid lees impact on MLF and to reproduce enological-like conditions. Ten mL of wine were transferred into 10 mL cylindrical vials (Fischer Scientific, France, ref: 11981523), closed with screw caps (Agilent Technologies, hdsp cap 18 mm PTFE / sil 100 pk, France). The wines were inoculated with Lactoenos® B7 Direct rehydrated lactic bacteria at a concentration of 10 mg.L⁻¹, as recommended by the supplier (Laffort, France). Fermentation samples were taken every 2 to 5 days for measuring L-malic acid decrease as a proxy of the MLF progress. The MLF ongoing was fitted by a 3 parameters logistic function using the R package (*drm* function of *drc* package), four kinetics parameters were extracted: *t*_{MLF_lag}, and *t*_{MLFend}, correspond to the time to initiate and to achieve the MLF, respectively. *t*_{MLF50} corresponds to the time for consuming 50% of malic acid, and *r*_{max_MLF} is the maximal malic acid consumption rate (g.L⁻¹.h⁻¹). Data fit and MLF parameters are illustrated in the Table S3.

2.7 Enzymatic assay of wine

At the end of the alcoholic fermentation, a sample volume of 800 µL was manually transferred in Micronics tubes (Novazine, Lyon, France, ref: MP32033L) and stored at -20°C. The concentrations of the following organic metabolites were measured: acetic acid, glycerol, malic acid, pyruvate, acetaldehyde and total SO₂ using the respective enzymatic kits: K-ACETGK, K-GCROLGK, K-LMAL-116A, K-PYRUV, K-ACHYD, K-TSULPH, (Megazyme, Bray, Ireland) following the instructions of the manufacturer. Glucose and fructose were assayed by using the enzymatic method described by Stitt et al. (1989). All the enzymatic assays were performed by a robotic platform using the Bordeaux metabolomics facilities (<http://metabolome.cgfb.u-bordeaux.fr/>). Only the strains able to complete the fermentations (<1.5 g.L⁻¹ of residual sugars) were retained.

2.8. Use of previous phenotypic datasets

Two previously published datasets were used for estimating the heritability and the efficiency of this marker assisted selection program. Fermentation phenotypes were measured in the same conditions and in the same grape musts allowing to compare the data. Malic Acid Consumption (MAC) was the ratio of malic acid consumed ($[\text{L-malic acid}]_{\text{initial}} - [\text{L-malic acid}]_{\text{final}} / [\text{L-malic acid}]_{\text{initial}}$ expressed in %. The MAC values of the 35 enological strains and the 94 progenies of SBxGN hybrid were extracted from Peltier et al. (2018a) and Peltier et al. (2020), respectively. The MAC values of M2xF15 offspring were measured in the same conditions described by Peltier et al. (2018b) but are only released in this present work.

2.5.6. Statistical analyses

All the statistical and graphical analyses were carried out using R software [54]. Plots were computed using the *ggplot2* package and analyses of variance were carried out using the *car* package. Four linear models were applied: LM1 investigated the effect of QTL numbers on MAC according to the formula :

$$y_i = m + EA_i + \epsilon_i, \quad (1)$$

where *y* is the MAC value for a strain carrying *i* *Enhancers Alleles* (*i*=1...11) and ϵ_i the residual, *m* is the overall mean of the 154 FMGSs progenies phenotyped. The LM2 model estimated the effects of the following factors *must*, *strain*, *temperature*, *nitrogen*, and *malic acid* as well as the first order interaction of all the factors on MAC value according to the formula:

$$y_{ijklm} = m + \text{strain}_i + \text{must}_j + \text{temp}_k + \text{nitro}_l + \text{malic}_m + \text{inter}_{ijklm}^2 + \epsilon_{ijklm}, \quad (2)$$

where y_{ijklm} is the MAC value for a strain i ($i=1\dots6$) fermented in a must j ($j=1,2$) at the temperature k ($k=1,2$) and containing the nitrogen level l ($l=1,2$) and the malic acid level m ($m=1,2,3$). Levels taken by each factor are given in the Online Resource 2. The term inter_{ijklm}^2 represents the first order interactions for each factor and ϵ_{ijklm} the residual, m is the overall mean of the 84 fermentations carried out in triplicate in an equilibrated fractional design. This model was refined with the models LM3 and LM4 allowing the estimation of each factors in the SB16 and M15 must, respectively.

The analysis of variance of models LM2, 3 and 4 allows the estimation of the primary effect of the strain, the grape must, the temperature, the nitrogen and the malic acid concentration as well as their primary interaction effects. The normal distribution of residues as well as the homoscedasticity of the variances were tested by Shapiro test and Levene test (*car* package).

1 Results

1.1. A basic breeding strategy for improving malic acid consumption

The main goal of this study consists to significantly increase the *S. cerevisiae* consumption of malic acid during alcoholic fermentation. Previously, it has been reported that wine strains were unable to consume more than 50 % of the malic acid grape juice in controlled conditions. This overall feature was previously confirmed in our laboratory by measuring the residual malic acid concentration at the end of the alcoholic fermentation for 35 strains in 5 grape juices [52]. Since the initial amount of malic acid varied according to the grape juice, we computed the percentage of malic acid consumed (MAC) for comparing strains within multiple environmental conditions. On average, MAC ranged between 30.0 and 53.0% within a population of 31 commercial starters plus the four founder strains of this study (SB, GN, M2, F15), that are haploid derivative from commercial starters. Strong differences between grape musts were found (Table S3). The continuous distribution of this trait was illustrated for a red grape juice Merlot 2015 in the Figure 1a. Strains SB and GN had extreme phenotypes, metabolizing between 40.0% and 3.5% of malic acid, respectively. Strains M2 and F15 had an intermediate phenotype, consuming up to 20% of malic acid in this condition. This analysis demonstrated that MAC is a quantitative trait with important variation among commercial starters and therefore we decided to took advantage of this natural variation by initiating a selection program with a breeding approach.

We decided to use two previously generated mapping populations as starting material. These populations originated from two hybrids, resulting from the mating of SB with GN (SBxGN) and M2 with F15 (M2xF15) that were sporulated to generate 94 and 96 segregants, respectively. The rationale being that meiosis segregation would reveal transgressive phenotypes. SBxGN offspring was phenotyped for MAC in a previous study [48] and the phenotypic characterization of M2xF15 offspring was achieved in the same conditions but the MAC values are only released in this present work (Table S3). The fermentation conditions applied (grape juice, fermentation volume, temperature) are the same in all the studies. In both F1-hybrids, the meiotic segregation yields few clones consuming up to 60% of malic acid (Figure 1 panels b and c). Thanks to this first approach, we were able to select two progeny clones GS-28B from SBxGN offspring and FM-8D from M2xF15 one that had high MAC value of 58.0 % and 45.4 %, respectively. These high values correspond to the reported limits of *Saccharomyces cerevisiae* physiology [45,55–57].

To pursue trait improvement, strains GS-28B and FM-8D were mated by manually pairing spores of both strains. The resulting F1-hybrid (FMGS-1) should have inherited from both genetic backgrounds independent genetic factors controlling MAC that can be recombined through meiosis. Therefore, FMGS-1 was sporulated and a high number of

spores were purified by taking advantage of the hydrophobicity properties of spore wall. A spore preparation of 95% was obtained and around 1000 clones were isolated on solid YPD and stored at -80°C . Fifty randomly picked individuals were fermented in the M15 grape juice and their MAC was measured. In this population, the consumption levels of malic acid were on average significantly much higher than those previously observed in SBxGN and M2xF15 progenies as well as in the commercial starters panel (Figure 1d). This result demonstrates that the simple cross of two optimal individuals derived from different background is very efficient for optimizing complex phenotypes. Such strategy is routinely used by microbiologists for improving yeast strains and have been reported and reviewed by many authors [11,14,58–60].

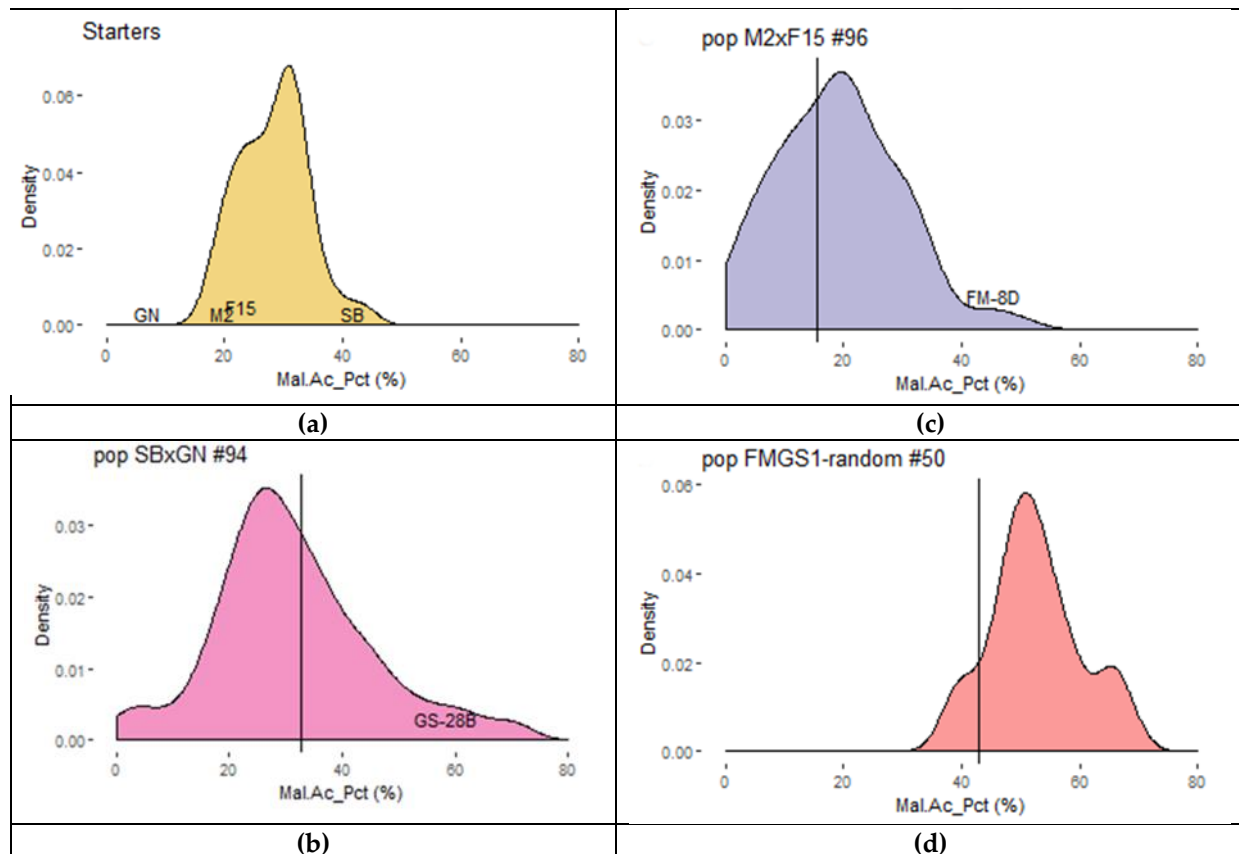
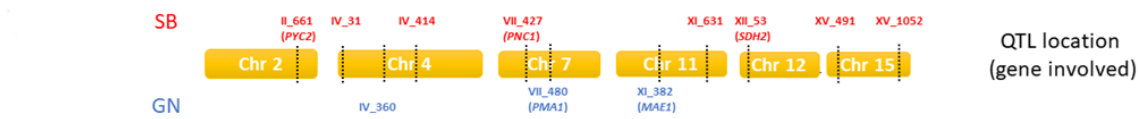


Figure 1 Malic Acid Consumed (MAC) at the end of alcoholic fermentation in a merlot grape juice. **(a)** Phenotypic distribution of 31 starters and relative position of the 4 founder strains SB, GN, M2, F15 (data previously published by [52]). **(b)** Phenotypic distribution of SBxGN hybrid and 94 related progeny clones including GS-28B (data previously published [46]). **(c)** Phenotypic distribution of M2xF15 hybrid and 96 related progeny clones including FM-8D (this work). All fermentations were carried out in the same conditions. **(d)** Phenotypic distribution of FMGS1 hybrid and 50 related progeny clones. The vertical bar indicated the value of the F1-hybrid of each progeny.

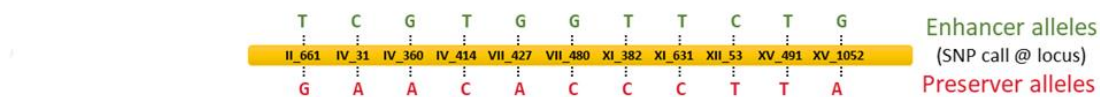
3.2. Use of Marker Assisted Selection (MAS) for enhancing malic acid consumption

To go further, we used at a large scale genetic markers for selecting optimal individual prior any phenotypic tests. Indeed, getting genotypic data is generally easier than setting up a phenotypic characterization. For testing the relevance of this strategy, we first defined a set of eleven genetic makers that are nucleotide variations genetically linked to QTL controlling MAC and identified in SBxGN background [46,47]. The Figure 2 a and b summarize SNP position and *enhancer* or *preserver* alleles that were define for each QTL. The genotype of the founder strains (SB, GN, M2 and F15) and the parental strains GS-28B and FM-8D is given panel c. We applied this genotypic screening on the ~1,000 meiotic spores previously isolated from FMGS-1 and sought for meiotic spore clones having

preferably inherited Quantitative Trait Loci (QTL) allele promoting the malic acid con-



(a)



(b)

Strain	II_661	IV_31	IV_360	IV_414	VII_427	VII_480	XI_382	XI_631	XII_53	XV_491	XV_1052
SB	T	C	A	T	G	C	C	T	C	C	G
GN	G	A	G	C	A	G	T	C	T	T	A
M2	G	A	G	T	A	G	T	T	T	T	A
F15	G	A	G	T	A	G	C	T	T	T	G
GS-28B	G	C	G	T	G	G	T	T	C	C	G
FM-8D	G	A	G	T	A	G	T	T	T	T	A

(c)

sumption. As detailed in the material and method section, each QTL was tracked by using the MASS-array technology able to discriminate in a unique experiment eleven pairs of biallelic Single Nucleotide Polymorphisms (SNP) (Table S1). The inheritance of the 11 loci for 924 FMGS-1 progenies was interrogated. SNP calling was higher than 94% and most of the strains were fully genotyped. A first segregation analysis confirmed that spore purification was very efficient for removing residual F1-hybrids (fully heterozygous). Indeed, most of the clones (77%) were fully homozygous while a minor fraction (15.5%) harbored both homozygous and heterozygous loci and could be considered as sibling-pair hybrids (F2) (Figure S1 left panel). Only fully homozygous strains with less than one missing genotype were analyzed representing 583 individuals. According to parental genotypes, the segregation of six loci was expected (Figure 2). Indeed, 5 *enhancer* alleles were fixed (IV_360G, IV_414T, VII_480G, XI_382T, and XI_631T), one *preserver* allele was fixed (II_661G), and the six other loci segregated in a mendelian fashion (χ^2 test $p>0.05$) (Figure 3a).

Figure 2 QTLs tracked, and Mass-Array markers used. **(a)** Genetic position of the eleven QTLs mapped modulating malic acid consumption in SBxGN hybrid. The parental allele (GN or SB inheritance) promoting a stronger consumption was indicated for each locus. **(b)** SNP corresponding to the enhancer and preserver allele, the exact position is given in Table S1. **(c)** SNP call in the four founder and in the parental strains GS-28B and FM-8D.

We next investigated the phenotypic impact of markers segregation. The progeny population was split in two groups according to the number of *enhancer* alleles detected. 122 individuals harbor 9 or 10 *enhancer* alleles while 125 individuals harbor only 5 or 6 *enhancer* alleles (Figure 3b). Each group represents nearly 20% of the total population genotyped. In order to test QTLs effect in the FMGS-1 background, 30 and 23 individuals belonging to each group were randomly selected and phenotyped in the Merlot grape juice. Strains enriched for *enhancer* alleles (9 to 10 optimal loci) consumed more malic acid than the control group (Figure 3c). On average, the *enhancer* group consumes 53.7% of malic acid while the control only 48.0% of malic acid (Wilcoxon test $p < 0.05$). Although the phenotypic gain observed (+5.7% of malic acid) was moderated, this demonstrates that MAS allows the blind selection of highly performing individuals without any phenotypic screening.

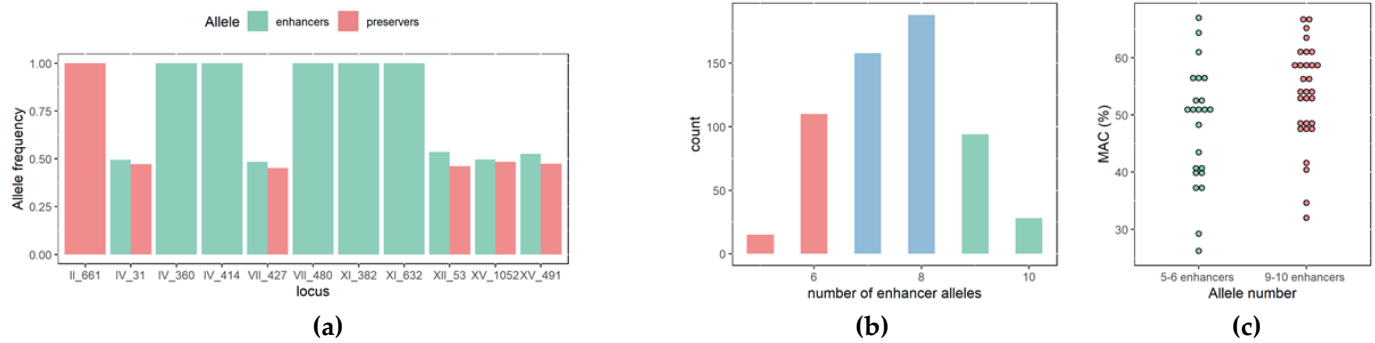


Figure 3. Marker assisted selection of FMGS-1 progeny for five segregating alleles. **(a)** Allele frequency in the progeny, enhancer and preserver alleles are shown in green and red, respectively. **(b)** distribution of the number enhancer alleles counted in the FMGS-1 progeny (593 fully genotyped progenies). **(c)** phenotypic difference between progenies having enriched and minimized number of enhancer alleles.

3.3. Construction and characterization of a backcrossed FMGS-2 population.

A possible cause of the moderate efficiency of MAS in the FMGS-1 background would be the fact that many QTLs were fixed (*i.e.* have a homozygous inheritance). Therefore, a second hybrid (FMGS-2) was obtained in order to increase the number of segregating loci. A progeny clone of FMGS-1 (FMGS-1#647) was crossed with GS-41B, one segregant of SBxGN hybrid. FMGS-1#647 has an optimal MAC (65.7%) and harbor 8 out of 11 *enhancer* alleles. GS-41B has a moderated MAC value (40.4%) but allows the introduction of other enhancer alleles in the resulting hybrid. Indeed, in the FMGS-2 hybrid, all but two QTLs (VII_480 and XII_53) were heterozygous (Figure 4a). From FMGS-2, 1232 spore clones were isolated and genotyped using the same procedure than for FMGS-1 offspring. Mass-array analysis confirmed the Mendelian segregation of 9 out of the 11 markers typed (χ^2 test $p > 0.05$). The genotyping of these clones allows the identification of nearly 12% of residual F1-hybrids and F2-sibling pair hybrids due to a less efficient spore purification. In the further analysis, 797 fully homozygous meiotic segregants were used. In order to test MAS efficiency in this new background, the same approach than for FMGS-1 was used. Fifty randomly selected individuals were phenotyped for characterizing the overall performance of FMGS-2 progeny. Random progenies of FMGS-1 and 2 have similar average values (52.6 and 48.7) (Table 3 and Figure S2), however the phenotypic variability (variance) in the FMGS-2 offspring is two folds higher (130 vs 58). This result is likely due to the higher number of QTLs in segregation in FMGS-2 population. Beside this difference, the proportion of clones with optimal malic acid consumption is broadly the same (Table 2).

Table 2. Biometric comparison of the populations used.

Populations	Number	Mean	Variance	Quantiles			% of strains with MAC higher than	
				50%	75%	95%	65%	70%
Starters	31	28.3	33.3	28.2	31.5	36.5	<i>no</i>	<i>no</i>
pop SBxGN	93	31.0	206.6	28.6	36.9	58.5	2.2	2.2
pop M2xF15	94	17.8	138.9	18.6	24.8	34.1	<i>no</i>	<i>no</i>
random FMGS-1	50	52.6	58.6	51.7	57.2	65.7	12.0	<i>no</i>
selected FMGS-1 ^a	30	53.7	77.0	54.1	59.2	65.8	10	<i>no</i>
random FMGS2-1	50	48.7	130.4	49.0	56.6	64.3	4.0	2.0
selected FMGS2-2 ^b	17	53.7	121.2	54.2	57.1	72.3	17.6	11.7

^a genetically selected for 9-10 enhancer alleles^b genetically selected for 10-11 enhancer alleles
no not observed

Similarly, a MAS was carried out by selecting individuals according to their genotype. Seventeen individuals that have inherited of 10 and 11 *enhancer* loci were selected among the 797 FMGS-2 progeny genotyped. This *enhancer*-rich group was compared to a small group of 10 strains harboring only 2 to 4 *enhancers* loci. On average, the *enhancer* group consumes +8% of malic acid at the end of the alcoholic fermentation (53.7 % *vs* 45.8%, Wilcoxon test $\alpha=0.05$) (Figure 4c). This also demonstrates that, in the FMGS-2 background, MAS is a convenient way for selecting extreme individuals in a small group of strains. Indeed, within the 17 genetically selected strains, two strains consumed more than 70% of malic acid which represents 11 % of the population (Table 2); this proportion is significantly higher than those expected in the random population (hyper geometric test $\alpha=0.05$).

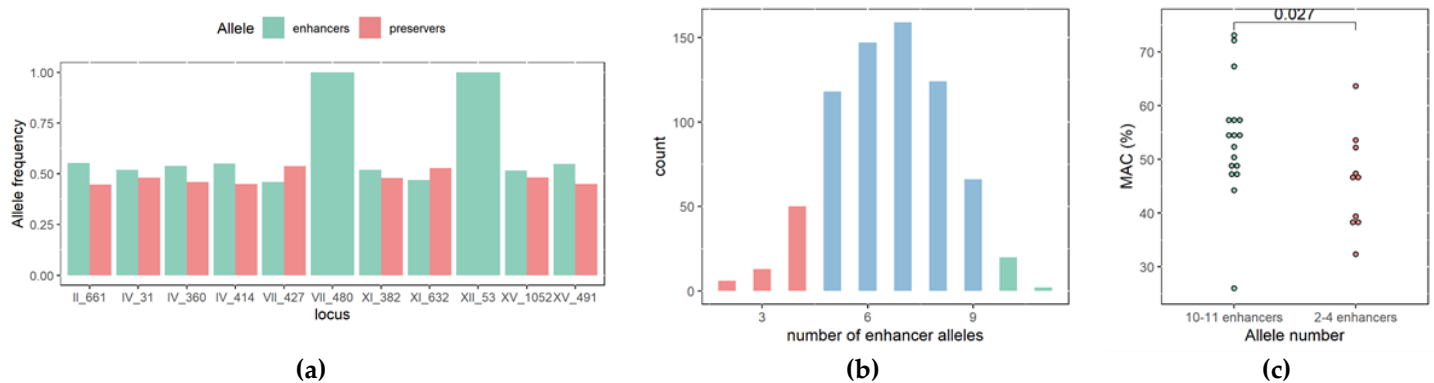


Figure 4 Marker assisted selection of FMGS-2 progeny for nine segregating alleles. **(a)** Allele frequency in the progeny, enhancer and preserver alleles are shown in green and red, respectively. **(b)** Distribution of the number enhancer alleles counted in the FMGS-2 progeny (797 fully genotyped progenies). **(c)** Phenotypic difference between progenies having enriched and minimized number of enhancer alleles.

3.4. Study of QTL penetrance in FMGS-1 and FMGS-2 populations

Although a significant difference was found between *enhancers* and *preservers* groups (Figure 3c and 4c), many individuals with a nearly optimal genotype reach a moderate MAC value. Encompassing the random and marker-selected strains of each FMGS populations, a total of 154 strains were phenotypically and genetically characterized (Table S4). This set of strains was used for estimating the cumulative impact *enhancer* allele on the malic acid consumption. A linear model confirms that, the more the number of *enhancer* alleles, the stronger the percentage of malic acid consumed (Figure 5a). Although, this

correlation is very significative (Spearman test, $\rho=0.23$, $p\text{-value}<0.0001$); a great residual variation within extreme groups is still observed. Indeed, the number of QTLs explained only 4.2% of the total variance observed in the linear model (LM1 see methods). This result illustrates the low penetrance level of the QTL pool used, which is a frequent case in MAS programs [40].

The effect of each QTL on malic acid consumption was tested and compared to their effect observed in the SBxGN background that was used for their identification. When a significative difference was observed (one-way Anova $\alpha=0.05$), the part of variance explained was indicated (Table 3). Since all the loci did not segregated in both progenies this test was in some cases not applied.

Table 3: QTL penetrance in various populations ^a

Population	II_661	IV_31	IV_360	IV_414	VII_427	VII_480	XI_382	XI_631	XII_53	XV_491	XV_1052
Gene validated	<i>PYC2</i>				<i>PNC1</i>	<i>PMA1</i>	<i>MAE1</i>		<i>SDH2</i>	<i>PTC5</i>	
SBxGN (n=94)	3.87	4.22	ns	ns	ns	8.20	2.97	ns	4.45	7.61	ns
FMGSs (n=154)	-	7.14	-	-	ns	-	-	-	-	ns	4.92
FMGS-1 (n=82)	-	9.31	-	-	ns	-	-	-	ns	ns	9.74
FMGS-2 (n=72)	ns	6.41	ns	ns	ns	-	ns	3.10	-	ns	ns

^a Part of variance explained in %, one-way analysis of variance, $p\text{-value}<0.05$.

ns: not significative, -: not relevant

Among the eleven QTLs used, only six have a significative effect in the SBxGN population (Anova $\alpha=0.05$). This result is explained by the fact that those QTLs were detected in three different environments [46] which is not the case in the present analysis of variance. Surprisingly, although the effect of the causative genes *PNC1* has been molecularly validated [46], the inheritance of the marker *VII_427* did not impact MAC in the population. When the two FMGSs populations were pooled, two of the four segregating QTLs had a significative effect (*IV_31* and *XV_1052*). The locus *IV_31* has a complete penetrance since is significative in the three populations. The locus *XV_1052* has a more contrasted effect; it accounted for 9.7 % of total variance observed in FMGS-1 progeny but did not significantly impact the malic acid consumption in FMGS-2 or SBxGN populations. Finally, the locus *XI_631* has a significative effect only in the FMGS2 population (Table 3). These results show that most of the QTLs tracked have an incomplete penetrance in the FMGS hybrids. This result can be explained by at least three causes: (i) the limited number of progenies tested (less than 100), (ii) the epistatic relations withing QTLs, (iii) the presence of modifier loci. We tested possible pair-wise interactions within the nine segregating QTLs in the FMGS-2 population (Table S5). Interestingly, two significative interactions were found (*XV_1052 vs XI_382*) and (*II_661 vs XV_491*). In the first case, the inheritance of the allele *XI_382^C* impairs the enhancer effect of the allele *XV_1052^G* (Figure 5b). This explains why *XV_1052^G* allele has a much higher penetrance in the FMGS-1 progeny where the *XI_382^T* allele is fixed (Figure 2). Concerning the *II_661* and *XV_491* interaction, the best combination expected *II_661^G/XV_491^T* has a similar effect than the worst one *II_661^T/XV_491^C* (Figure 5c); suggesting a possible metabolic trade-off between their associated genes *PYC2* and *PTC5*.

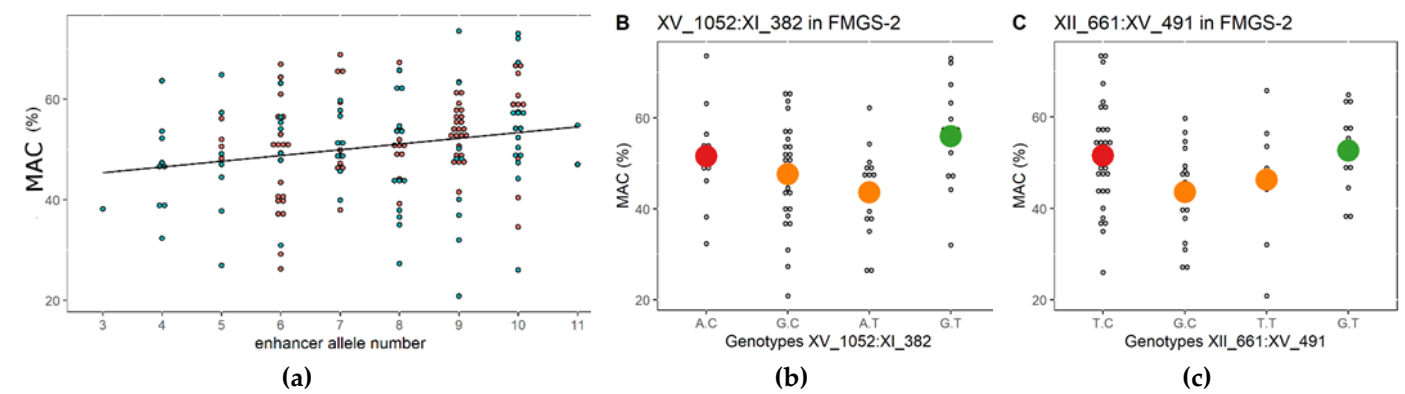


Figure 5 Overall QTL effect and pair wise interactions. **(a)** Linear relationship between the number of enhancer alleles and the MAC value in the 154 FMGS’ progenies tested; the red and blue dots represent the FMGS-1 and FMGS-2 progenies, respectively. **(b)** and **(c)** Genetic interactions between QTLs observed in the FMGS-2 population (72 individuals). The red, orange, and green dots are the average values observed for the bad, intermediate, and good combinations expected.

3.5. Phenotypic characterization of high malic acid consuming strains

Four malic consumer strains belonging to both FMGS’ populations (FMGS_215, FMGS_889, FMGS2_107, FMGS2_265) were selected in order to be phenotypically characterized in various environmental conditions. These strains were compared to SB and GN which represent the extreme limits of the starter population (Figure 1a). Small-volume alcoholic fermentations were carried out in two grape musts (M15 and SB16) adjusted with different concentrations of malic acid and nitrogen and incubated at two different temperatures (see methods). The objective was to assess the impact of several basic environmental factors in enology that could affect MAC. The part of variance of each factor as well as their first-order interactions were estimated by applying a fractional experimental design (see methods). The effect of each factor is represented in the Figure 6a and summarized in Table S6.

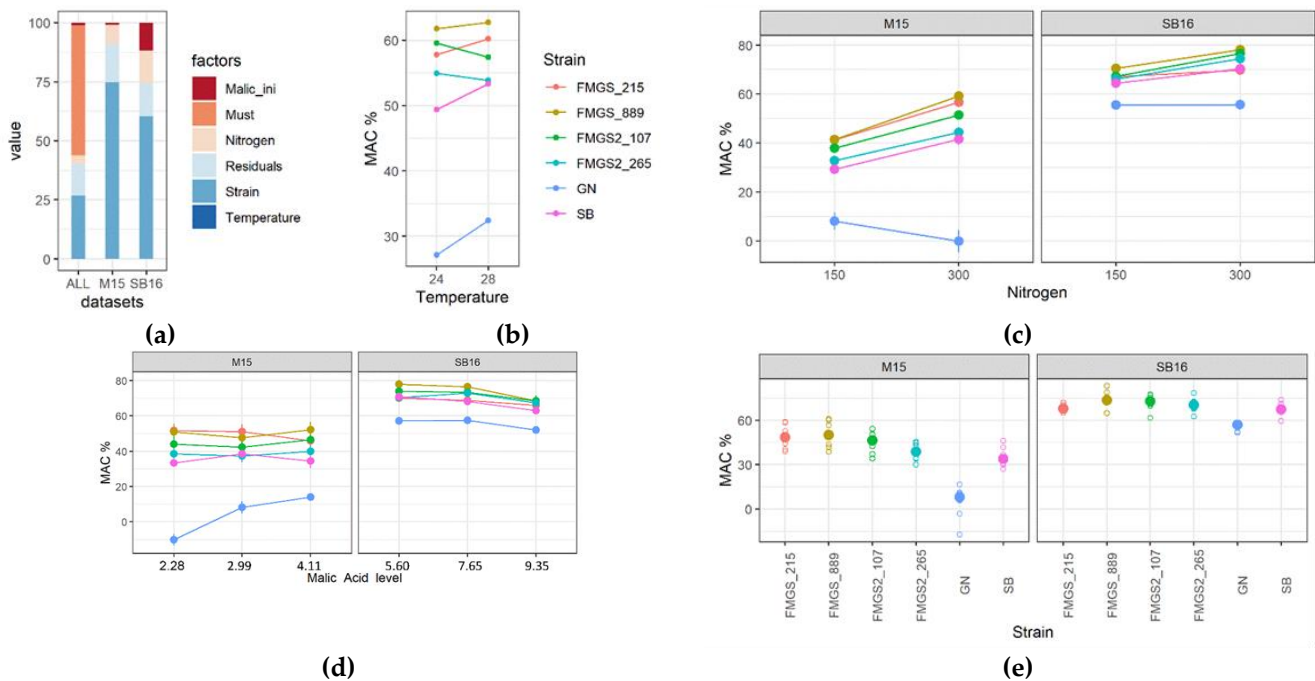


Figure 6 Biotic and abiotic factors affecting the variation of malic acid consumption. **(a)** Bar graphs indicate the part of variance explained by the different factors considered by the linear model applied: malic acid concentration, must origin, nitrogen content, strain origin, and temperature. **(b)** Temperature effect according to the strains used, **(c)** Nitrogen effect according to

the strains used, (d) initial malic acid concentration effect according to the strains used, (e) strain effect in both grape juices. For panels (b) to (e) the full dots represent the average values.

All conditions combined; MAC varies mainly with the nature of the must (55.2% of total variance explained). Indeed, yeast strains consumed in average 32.5 % more malic acid in white must compared to red must (Figure 6a). In contrast, temperature had a low impact on MAC and can be considered as poorly significant in the present study (Figure 6b). The important must-effect observed is likely due to the highest malic acid concentrations found in white must (SB16) compared to those of the red one (M15). This difference strongly impacts the pH of the matrix and may affect the diffusion of malic acid through cell membrane since it is protonated below pH=3.4 [61].

In order to dissociate this matrix effect, a second set of ANOVA was carried out by splitting the dataset according to the must. The new models shed light on the effect of nitrogen and malic acid concentrations that were partially hidden by the must effect. Nitrogen concentrations impacted malic acid consumption in both musts (Figure 6c). In our experiment, malic acid consumed was 10% higher in low nitrogen condition (150 mg/L N). However, this trend was not observed for the strain GN that consumes more malic acid (+4%) in rich nitrogen condition (300 mg/L N).. The impact of grape juice concentration of malic acid on MAC was also strongly contrasted. As shown on the Figure 6d, the malic acid level accounts for 11.8% of the total variance explained in the white wine but only 0.9% of the total variance in the red one. In the white grape juice, the MAC decreases (-8%) for the highest malic acid concentration (9.39 g/L) while this trait is steadier in red grape juice. Again, the response of GN differs from the malic consumer strains. Indeed, in red must, the impact of initial malic acid was stronger, with an increased MAC with higher initial malic acid concentration.

Finally, this experiment demonstrates that in both musts, the factor “strain” has the major impact explaining 74.8 and 60.5 % of “red” and “white” models, respectively (Figure 6e). As expected, all the strains selected by genetic markers consumed significantly more malic acid than the control GN. The strongest consumer is the strain FMGS_889 that metabolized 32 % and 11% more malic acid than the strains GN and SB, respectively (Wilcoxon test $p < 0.01$). Although minor variations were attributed to nitrogen and malic acid content, those strains steady consume malic acid which is not the case of the strain GN. The founder strain SB belongs to the malic consuming group but has always a lower MAC than the four individuals selected. As a direct consequence, the resulting pH of wine at the end of the alcoholic fermentation was drastically impacted (Figure S2). Indeed, in the most discriminating conditions, the pH difference measured in produced wine between extreme strains (GN and FMGS_889) was of 0.42 units (M15_high_initial_24), demonstrating the strong impact of *S. cerevisiae* strains in the modulation of wine pH.

3.6. Ongoing of Malo-Lactic Fermentation.

During the winemaking process, most of the red wines and some white wines underwent the Malo-Lactic Fermentation (MLF) due to the development of the lactic bacteria *Oenococcus oeni* [62]. The malic acid is one key energetic substrate of the MLF and its content in wine at the end of the alcoholic fermentation has an obvious impact on the process. It has been reported that L-malic acid concentration delays the MLF duration but does not drastically affect the *O. oeni* fermentation rate [63,64]. In addition, low pH has a negative impact on MLF initiation and efficiency [62]. Since strains selected clearly impacted malic acid concentration (Figure 6e) and wine pH (Figure S2), their influence on MLF was evaluated by inoculating the different wines produced with a malolactic starter (see methods). MLF ongoing was monitored by measuring the L-malic acid degradation by an enzymatic assay. MLF kinetics were fitted by a logistic function and four kinetics parameters were extracted (t_{MLF50} , t_{MLFend} , t_{MLFlag} , r_{max_MFL}) (Figure S3). For the malic consuming yeast strains, the MLFs were completed between 10 and 15 days. In contrast MLFs carried out in wines fermented by GN were much slower and were sometimes not achieved (Figure 7a).

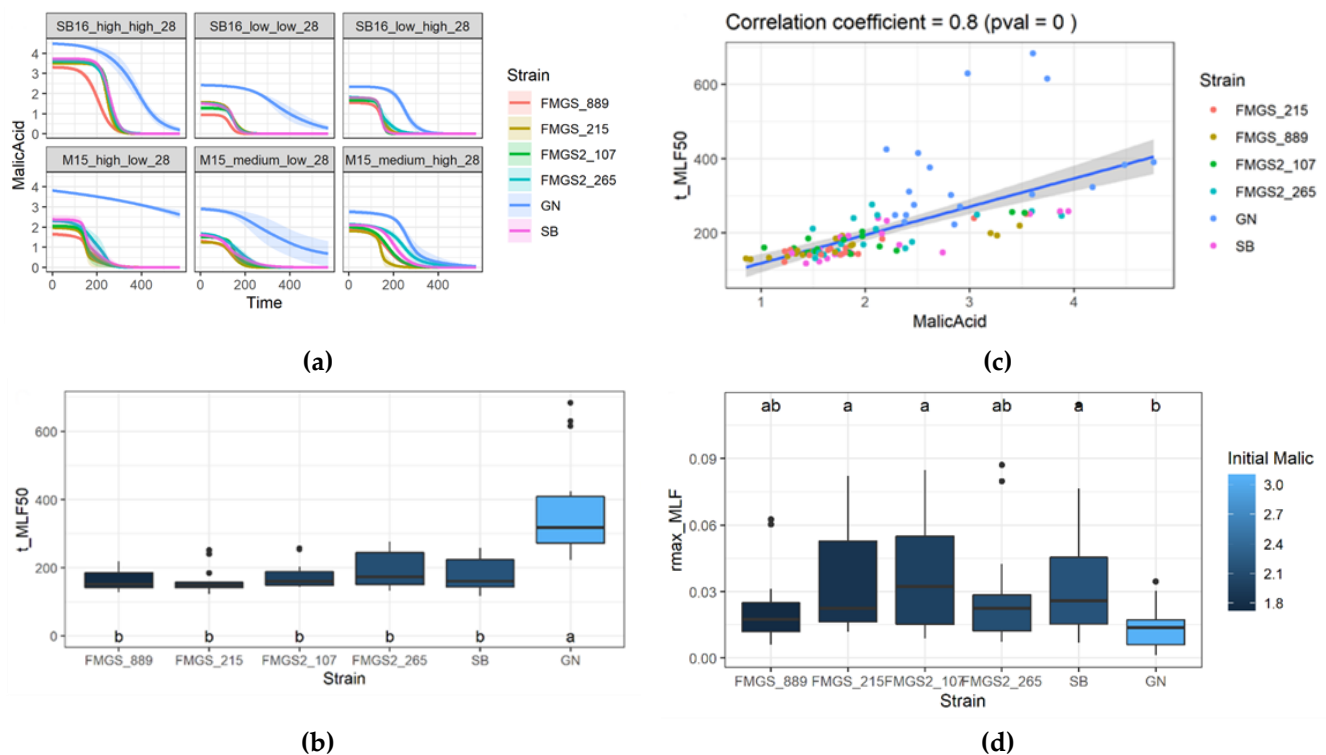


Figure 7 Malo-lactic fermentations with high malic consumer strains. (a) Malic acid concentration (g/L) according to the time (h) for each stain used in the 6 inoculated wines. **(b)** Time (h) to achieve 50 % of the MLF according to the strains used. **(c)** Correlation between the time at 50% of the MLF (t_{MLF50}) and the malic acid concentration (g/L). **(d)** Maximum fermentation rate (g/L/h) according to the strains used.

A correlation analysis performed on the 36 wines (6 strains and 6 conditions) clearly demonstrated a positive correlation between the initial malic acid content and the time for consuming 50% of malic acid (t_{MLF50}) (Figure 7b). Indeed, the amount of malic acid in wines fermented with GN was 1.1 g.L⁻¹ higher, on average, than those from other strains (Figure 7c). GN was different from the other strains (Kruskal-Wallis test, $\alpha=0.05$) and MLFs of wines fermented with GN required twice more time for consuming 50% of total malic acid. In contrast, the maximum fermentation rate (r_{max_MLF}) was quite similar between the strains. Indeed, strains FMGS_889 and GN have similar r_{max_MLF} but opposed t_{MLF50} values (Figure 7d). Thus, the time for completing MLF (t_{MLFend}) proved to be faster (less than 300 hours) when malic acid concentration was below 2 g.L⁻¹. In addition, a low initial concentration in malic acid does not seem to impair the initiation of MLF since the lag time of MLF (t_{MLFlag}) is quite similar in all the conditions tested. Altogether, these results suggested that malic consuming strains are compatible with sequential inoculation of lactic acid bacteria and should facilitate the MLF by increasing wine pH and reducing the malic acid concentration to consume.

4 Discussion

4.1. Assesment of a wide MAS program for improving a complex trait

Marker Assisted Selection (MAS) constitutes the applied side of quantitative genetics and has been widely used for improving plant varieties and animal races [32,65,66]. Since two decades, *Saccharomyces cerevisiae* has been raised as a perfect model for addressing fundamental questions in quantitative genetics [67–70]. Furthermore, the budding yeast is also the prime industrial microorganism involved in bioethanol and fermented foods

production. Although methods for identifying QTLs are perfectly controlled [31], their use for improving yeast strains for industrial applications is poorly documented. In the present study, we aimed to increase the malic acid consumption of wine starters in order to propose new strains with outlier features. To achieve this program, we get a new F1-hybrid (FMGS-1) resulting from the cross of two optimal unrelated strains (GS-28b and FM-8d). Then, we implemented for the first time a wide MAS program using 11 QTLs controlling the same trait.

First, a robust protocol was developed for isolating and genotyping a wide number of meiotic progenies derived from prototrophic and homothallic yeast. Although these features complexify breeding operations [14], the purification of FMGSs hybrids yielded more than 80% of spore clones (doubled haploids) or sibling-pair hybrids (F2) (Figure S1). The use of robotic-assisted DNA extraction and Mass Array genotyping allow a SNP call average of 94% and most of the strains were unambiguously genotyped. This technique allows the parallel genotyping of up to 40 markers which would strongly decrease the genotyping cost per SNP.

Second, we demonstrated that individuals carrying a high proportion of *enhancer* alleles consumes statistically more malic acid than those carrying a high proportion of *preserver* alleles in both FMGS progenies (Figure 3c and 4c). This result confirms that a blind preselection of strains can be achieved on the basis of their genotype. Nevertheless, the overall effect of this pool of alleles is limited regards to the general cross effect observed. Indeed, many FMGS-1 and FMGS-2 progenies reach high MAC value whatever their genotype. This result indicates that the simple selection of optimal outbred parental strains is very efficient for optimizing quantitative phenotypes. Indeed, the average malic acid consumed by randomly selected FMGS-1 progenies (50 strains) is statistically similar to average value of the FMGS-1 *enhancer* group (52.6% vs 53.7%). This is explained by the fact that five *enhancer* alleles are homozygous (fixed) in FMGS-1 hybrid and only five QTLs are in segregation (Figure 2a) reducing the proportion of non-optimal genotypes. Thus, 36% of randomly selected segregants (18) carry at least 8 *enhancer* alleles of the 11 possible one. These strains include FMGS-647, FMGS-215 and FMGS-889 that are among the top malic consumers. A second blind selection was then carried out in the FMGS-2 progeny. Since 10 of 11 QTL segregated, the difference of MAC between *enhancer* and *preserver* groups was larger (8%). A tight significative difference was found between random and *enhancer* enriched groups (48.7% vs 53.7%, respectively, Wilcoxon test < 0.1). In addition, the proportion of optimal clones consuming more than 65 or 70% is higher than in random population (Table 2). This highest proportion supports the idea that MAS would be efficient for the blind selection of many optimal individuals and should be used as a pre-screening step. Since many other quantitative traits segregate independently in FMGS's offspring, MAS may allow the selection of a collection of top-degrading malic acid strains to be used in downstream selection programs focusing on other traits (aromatic, fermentation performances, temperature adaptation...).

4.2. Possible causes of incomplete QTLs penetrance

Although it was successfully applied in many organisms, the efficiency of MAS is strongly depending on the number of marker used, the type of breeding design, the complexity, and the number of traits to improve [32,40,66,71]. Indeed, when QTLs detected in a specific background are introduced in an outbred individual, their expressivities are much lower than those observed in the initial QTL population [40]. This general trend was also observed here since only three QTLs of the eleven used has a significant effect of FMGSs hybrids (Table 2). This incomplete penetrance can be explained by several causes. First, the 11 QTLs used were detected in a multiple environmental design which increase the detection power of QTLs [46,47]. Thus, it is not surprising that minor QTLs showing GxE does not raise the significant threshold imposed in the subset of 94 SBxGN progenies. Second, possible alleles controlling MAC% in the M2xF15 background were not

considered at all. Since 50% of FMGS-1 genome is constituted by M2 or F15 alleles, QTLs governing the phenotypic variability of M2xF15 background are likely influencing the MAC% of FMGS's progeny. Third, the lack of penetrance observed for the marker VII_427 related to the causative gene *PNC1* would result to the complex genetic structure of the chromosome VII that contains many causative genes including *PMA1* a major contributor of MAC% [45]. Finally, some QTLs seems to show epistatic interactions (Figure 5b and c) that modulate their expressivity in a non-predicable manner.

4.3. Outlier strains for specific enological applications. Lowering the acidity of rich malic wines and shortening the MLF of red wines.

From an enological point of view, this study provides new strains for managing atypical vinification conditions. Indeed, *S. cerevisiae* strains consuming more than 50% of the grape juice malic acid are quite rare and have been scarcely described [55,72–74]. Such strains would be useful to better managing wine acidity and may have two distinct enological applications. First, in very acidic grape juice (cold climate white grapes), the excess malic acid is a concern since the taste of related wines is quite sour [75]. Therefore, the biotechnological reduction of malic acid would contribute to provide smoother wines (less aggressive) from a tasty point of view. In the Sauvignon blanc grape juice containing 9.39 g/L of L-malic acid, the difference in final L-malic acid concentration reached 1.3 g/L (Figure 7a) representing a pH shift close to 0.2 pH unit (Figure S2) with a possible impact on the acidity perception. Second, such strains would contribute to facilitate malo-lactic fermentation by increasing wine pH and reducing L-malic concentration. Indeed, we demonstrated that none of the FMGSs strains selected has a negative impact MLF ongoing inoculated with a commercial starter (Lactoenos® B7 Direct, Laffort oenology, FRANCE). In contrast, the malic acid reduction and the pH increase promote the MLF by reducing its duration. Therefore, whatever the residual amount of L-malic acid at the end of the alcoholic fermentation, the MLF is initiated readily without any significative impact on the consumption rate of bacteria.

By measuring the MAC% in several musts and conditions (temperature, nitrogen composition and L-malic concentration) we demonstrated that the high consumption properties of FMGS's strains are quite robust to environmental changes. Temperature did not impact this trait and the main factor is the nature of grape juice (Figure 6a). Although the composition of the two grape juices are not chemically defined, a possible cause of this discrepancy should be the pH of the media that affects the charge and the transport of malic acid [73]. However, in a specific must, L-malic acid concentration has moderated effect (Figure 6c). Beside the nature of the must, the impact of nitrogen seems to be the most important factor with strain specific interactions that has been previously reported [57]. Indeed, malic-consumer strains achieved a stronger MAC in nitrogen rich conditions which is not the case of GN. The GN's behavior was previously reported for other wine yeast strains [57]. This suggest that the high malic-consumer strains selected would have a particular response to nitrogen level.

Although significantly improved, the consumption of the total amount of malic acid by *S. cerevisiae* seems to be illusive. Indeed, the strain FMGS_889 consumed 84% of malic acid in SB16 and 67% in M15. In both cases, the final concentration of malic acid fall to 0.9 g/L constituting a kind of physiological threshold. This limit is likely due to the absence of an active malic acid transporter able to pump in this organic acid through the cell membrane [76]. A further improvement of malic acid consuming ability would require the enhancement of malic acid uptake. Beside the expression of recombinant proteins [77] this new skill might be acquired by activating quiescent transmembrane transporter by mutagenesis as achieved for xylulose import [78].

Supplementary Materials: The following are available online at www.mdpi.com/xxx/s1, Table S1: SNPs detected by mass array, Table S2: Composition of grape must, pH and nitrogen sources added, Table S3: MAC measured in 31 starters and the four founder strains [52], Figure S1: Purification rate

of progeny clones of FMGS-1 and FMGS-2 hybrids, Table S4: FMGS-1 and FMGS-2 genotypes and phenotypes, Table S5: Part of variance of pair-wise interactions in FMGS-2 population, Table S6: Analysis of variance of environmental and strain effect in selected strains, Figure S2: pH of wines at the end of the alcoholic fermentation, Figure S3: MFL parameters.

Author Contributions: PM, CV and EP conceived and designed study, MB, MM, CV and EP Performed research, EP, CV and PM Analyzed the data, PM, CV wrote the paper. All authors have read and agreed to the published version of the manuscript.”

Funding: “This research was funded by Aquitaine Region, grant number SESAM”

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable

Data Availability Statement: In this section, please provide details regarding where data supporting reported results can be found, including links to publicly archived datasets analyzed or generated during the study. Strains used have been deposited on the WDCM-791 (CRB ISVV, Bordeaux).

Conflicts of Interest: All the authors belong to the Biolaffort company; the funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

Acknowledgments: “Mass array assay was performed at the Genome Transcriptome Facility of Bordeaux (grants from the Conseil Régional d’Aquitaine n°20030304002FA and 20040305003FA, from the European Union FEDER n°2003227 and from Investissements d’Avenir ANR-10-EQPX-16-01), with the help of Erwan Guichoux”. Enzymatic assays were performed at the metabolomic Facility of Bordeaux (grants from PHENOME-ANR-INBS-0012) with the help of Yves Gibon.

References

1. Sicard, D.; Legras, J.L. Bread, beer and wine: Yeast domestication in the *Saccharomyces sensu stricto* complex. *Comptes Rendus - Biol.* **2011**, *334*, 229–236, doi:10.1016/j.crv.2010.12.016.
2. Rainieri, S.; Pretorius, I.I.S. Selection and improvement of wine yeasts. *Ann. Microbiol.* **2000**, *50*, 15–31.
3. Marullo, P.; Dubourdieu, D. Yeast selection for wine flavour modulation. In *Managing wine quality Vol 2*; Reynolds, A.G., Ed.; Woodhead Publishing Limited: Cambridge, 2010; pp. 296–320 ISBN 9781845697983.
4. Gammacurta, M.; Marchand, S.; Albertin, W.; Moine, V.; De Revel, G. Impact of yeast strain on ester levels and fruity aroma persistence during aging of bordeaux red wines. *J. Agric. Food Chem.* **2014**, *62*, 5378–5389, doi:10.1021/jf500707e.
5. Marchal, A.; Marullo, P.; Durand, C.; Moine, V.; Dubourdieu, D. Fermentative Conditions Modulating Sweetness in Dry Wines: Genetics and Environmental Factors Influencing the Expression Level of the *Saccharomyces cerevisiae* HSP12 Gene. *J. Agric. Food Chem.* **2015**, *63*, 304–311, doi:10.1021/jf504408t.
6. Murat, M.-L.; Masneuf, I.; Darriet, P.; Lavigne, V.; Tominaga, T.; Dubourdieu, D. Effect of *Saccharomyces cerevisiae* Yeast Strains on the Liberation of Volatile Thiols in Sauvignon blanc Wine. *Am. J. Enol. Vitic.* **2001**, *52*, 136–139.
7. Julien, a.; Roustan, J.L.; Dulau, L.; Sablayrolles, J.M. Comparison of nitrogen and oxygen demands of enological yeasts: Technological consequences. *Am. J. Enol. Vitic.* **2000**, *51*, 215–222.
8. Berthels, N.J.; Cordero Otero, R.R.; Bauer, F.F.; Thevelein, J.M.; Pretorius, I.S. Discrepancy in glucose and fructose utilisation during fermentation by *Saccharomyces cerevisiae* wine yeast strains. *FEMS Yeast Res.* **2004**, *4*, 683–689, doi:10.1016/j.femsyr.2004.02.005.
9. Marullo, P.; Mansour, C.; Dufour, M.; Albertin, W.; Sicard, D.; Bely, M.; Dubourdieu, D. Genetic improvement of thermo-tolerance in wine *Saccharomyces cerevisiae* strains by a backcross approach. *FEMS Yeast Res.* **2009**, *9*, 1148–1160, doi:10.1111/j.1567-1364.2009.00550.x.
10. Esteve-Zarzoso, B.; Gostincar, a; Bobet, R. Selection and molecular characterization of wine yeasts isolated from the “El Penedès” area (Spain). *Food ...* **2000**, *17*, 553–562, doi:10.1006/fmic.2000.0347.
11. Romano, P.; Soli, M.G.; Suzzi, G. Improvement of a wine *Saccharomyces cerevisiae* strain by a breeding program. *Appl. Environ. Microbiol.* **1985**, *50*, 1064–1067.
12. Marullo, P.; Bely, M.; Masneuf-Pomarède, I.; Pons, M.; Aigle, M.; Dubourdieu, D.; Masneuf-Pomarede, I.; Pons, M.; Aigle, M.; Dubourdieu, D. Breeding strategies for combining fermentative qualities and reducing off-flavor production in a wine yeast model. *FEMS Yeast Res.* **2006**, *6*, 268–279, doi:10.1111/j.1567-1364.2006.00034.x.
13. De Vero, L.; Solieri, L.; Giudici, P. Evolution-based strategy to generate non-genetically modified organisms *Saccharomyces cerevisiae* strains impaired in sulfate assimilation pathway. *Lett. Appl. Microbiol.* **2011**, *53*, 572–575, doi:10.1111/j.1472-765X.2011.03140.x.
14. Steensels, J.; Snoek, T.; Meersman, E.; Nicolino, M.P.; Voordeckers, K.; Verstrepen, K.J. Improving industrial yeast strains: exploiting natural and artificial diversity. *FEMS Microbiol. Rev.* **2014**, 1–49, doi:10.1111/1574-6976.12073.
15. Marullo, P.; Durrens, P.; Peltier, E.; Bernard, M.; Mansour, C.; Dubourdieu, D. Natural allelic variations of *Saccharomyces cerevisiae* impact stuck fermentation due to the combined effect of ethanol and temperature; a QTL-mapping study. *BMC Genomics* **2019**, *20*, 680, doi:10.1186/s12864-019-5959-8.
16. Marullo, P.; Aigle, M.; Bely, M.; Masneuf-Pomarède, I.; Durrens, P.; Dubourdieu, D.; Yvert, G. Single QTL mapping and nucleotide-level resolution of a physiologic trait in wine *Saccharomyces cerevisiae* strains. *FEMS Yeast Res.* **2007**, *7*, 941–952, doi:10.1111/j.1567-1364.2007.00252.x.
17. Zimmer, A.; Durand, C.; Loira, N.; Durrens, P.; Sherman, D.J.; Marullo, P. QTL dissection of lag phase in wine fermentation reveals a new translocation responsible for *Saccharomyces cerevisiae* adaptation to sulfite. *PLoS One* **2014**, *9*, 37–39, doi:10.1371/journal.pone.0086298.

18. Martí-Raga, M.; Peltier, E.; Mas, A.; Beltran, G.; Marullo, P. Genetic Causes of Phenotypic Adaptation to the Second Fermentation of Sparkling Wines in *Saccharomyces cerevisiae*. *G3 Genes| Genomes| Genet.* **2017**, *7*, 399–412, doi:10.1534/g3.116.037283.
19. Ambroset, C.; Petit, M.; Brion, C.; Sanchez, I.; Delobel, P.; Guerin, C.; Chiapello, H.; Nicolas, P.; Bigey, F.; Dequin, S.; et al. Deciphering the Molecular Basis of Wine Yeast Fermentation Traits Using a Combined Genetic and Genomic Approach. *G3 Genes| Genomes| Genetics* **2011**, *1*, 263–281, doi:10.1534/g3.111.000422.
20. Steyer, D.; Ambroset, C.; Brion, C.; Claudel, P.; Delobel, P.; Sanchez, I.; Erny, C.; Blondin, B.; Karst, F.; Legras, J.-L. QTL mapping of the production of wine aroma compounds by yeast. *BMC Genomics* **2012**, *13*, 573, doi:10.1186/1471-2164-13-573.
21. Cubillos, F.A.; Parts, L.; Salinas, F.; Bergström, A.; Scovacicchi, E.; Zia, A.; Illingworth, C.J.R.; Mustonen, V.; Ibstedt, S.; Warringer, J.; et al. High-resolution mapping of complex traits with a four-parent advanced intercross yeast population. *Genetics* **2013**, *195*, 1141–55, doi:10.1534/genetics.113.155515.
22. Jara, M.; Cubillos, F. a; García, V.; Salinas, F.; Aguilera, O.; Liti, G.; Martínez, C. Mapping genetic variants underlying differences in the central nitrogen metabolism in fermenter yeasts. *PLoS One* **2014**, *9*, e86533, doi:10.1371/journal.pone.0086533.
23. Salinas, F.; Cubillos, F.A.; Soto, D.; Garcia, V.; Bergström, A.; Warringer, J.; Ganga, M.A.; Louis, E.J.; Liti, G.; Martinez, C. The genetic basis of natural variation in oenological traits in *Saccharomyces cerevisiae*. *PLoS One* **2012**, *7*, e49640, doi:10.1371/journal.pone.0049640.
24. Brice, C.; Sanchez, I.; Bigey, F.; Legras, J.-L.; Blondin, B. A genetic approach of wine yeast fermentation capacity in nitrogen-starvation reveals the key role of nitrogen signaling. *BMC Genomics* **2014**, *15*, 495, doi:10.1186/1471-2164-15-495.
25. Roncoroni, M.; Santiago, M.; Hooks, D.O.; Moroney, S.; Harsch, M.J.; Lee, S. a.; Richards, K.D.; Nicolau, L.; Gardner, R.C. The yeast IRC7 gene encodes a b-lyase responsible for production of the varietal thiol 4-mercapto-4-methylpentan-2-one in wine. *Food Microbiol.* **2011**, *28*, 926–935, doi:10.1016/j.fm.2011.01.002.
26. Huang, C.; Roncoroni, M.; Gardner, R.C. MET2 affects production of hydrogen sulfide during wine fermentation. *Appl. Microbiol. Biotechnol.* **2014**, *98*, 7125–7135, doi:10.1007/s00253-014-5789-1.
27. Noble, J.; Sanchez, I.; Blondin, B. Identification of new *Saccharomyces cerevisiae* variants of the MET2 and SKP2 genes controlling the sulfur assimilation pathway and the production of undesirable sulfur compounds during alcoholic fermentation. *Microb. Cell Fact.* **2015**, *14*, 68, doi:10.1186/s12934-015-0245-1.
28. Cordente, A.G.; Heinrich, A.; Pretorius, I.S.; Swiegers, J.H. Isolation of sulfite reductase variants of a commercial wine yeast with significantly reduced hydrogen sulfide production. *FEMS Yeast Res.* **2009**, *9*, 446–459, doi:10.1111/j.1567-1364.2009.00489.x.
29. Linderholm, A.L.; Findleton, C.L.; Kumar, G.; Hong, Y.; Bisson, L.F. Identification of genes affecting hydrogen sulfide formation in *Saccharomyces cerevisiae*. *Appl. Environ. Microbiol.* **2008**, *74*, 1418–1427, doi:10.1128/AEM.01758-07.
30. Linderholm, A.; Dietzel, K.; Hirst, M.; Bisson, L.F. Identification of MET10-932 and Characterization as an Allele Reducing Hydrogen Sulfide Formation in Wine Strains of *Saccharomyces cerevisiae*. **2010**, *76*, doi:10.1128/AEM.01666-10.
31. Peltier, E.; Friedrich, A.; Schacherer, J.; Marullo, P. Quantitative Trait Nucleotides Impacting the Technological Performances of Industrial *Saccharomyces cerevisiae* Strains. *Front. Genet.* **2019**, *10*, 683, doi:10.3389/fgene.2019.00683.
32. Xu, Y.; Crouch, J.H. Marker-Assisted Selection in Plant Breeding: From Publications to Practice. *Crop Sci.* **2008**, *48*, 391–407, doi:10.2135/cropsci2007.04.0191.
33. Dekkers, J.C.M. Marker-assisted selection for commercial crossbred performance1. *J. Anim. Sci.* **2007**, *85*, 2104–2114, doi:10.2527/jas.2006-683.
34. Yao, F.-J.; Lu, L.-X.; Wang, P.; Fang, M.; Zhang, Y.-M.; Chen, Y.; Zhang, W.-T.; Kong, X.-H.; Lu, J.; Honda, Y. Development of a Molecular Marker for Fruiting Body Pattern in *Auricularia auricula-judae*. *Mycobiology* **2018**, *46*, 72–78, doi:10.1080/12298093.2018.1454004.

35. Dufour, M.; Zimmer, A.; Thibon, C.; Marullo, P. Enhancement of volatile thiol release of *Saccharomyces cerevisiae* strains using molecular breeding. *Appl. Microbiol. Biotechnol.* **2013**, *97*, 5893–5905, doi:10.1007/s00253-013-4739-7.
36. Marullo, P.; Yvert, G.; Bely, M.; Aigle, M.; Dubourdieu, D. Efficient use of DNA molecular markers to construct industrial yeast strains. *FEMS Yeast Res.* **2007**, *7*, 1295–1306, doi:10.1111/j.1567-1364.2007.00281.x.
37. Javelot, C.; Girard, P.; Colonna-Ceccaldi, B.; Vladescu, B. Introduction of terpene-producing ability in a wine strain of *Saccharomyces cerevisiae*. *J. Biotechnol.* **1991**, *21*, 239–251, doi:https://doi.org/10.1016/0168-1656(91)90045-W.
38. Blondin, B.; NOBLE, J.; Sanchez, I.; Agronomique, I.N.D.L.R.; Supagro), C.I.D.S.E.S.A. (Montpellier *Méthode de contrôle de la production de sulfites, d'hydrogène sulfureux et d'acétaldéhyde par des levures*; 2013;
39. Mackay, T.F.C. The Genetic Architecture of Quantitative Traits. *Annu. Rev. Genet.* **2001**, *35*, 303–339, doi:10.1146/annurev.genet.35.102401.090633.
40. Lecomte, L.; Duffé, P.; Buret, M.; Servin, B.; Hospital, F.; Causse, M. Marker-assisted introgression of five QTLs controlling fruit quality traits into three tomato lines revealed interactions between QTLs and genetic backgrounds. *Theor. Appl. Genet.* **2004**, *109*, 658–668, doi:10.1007/s00122-004-1674-0.
41. Ribéreau-Gayon, P.; Dubourdieu, D.; Donèche, B.; Lonvaud, A. *Handbook of Enology, Vol. 1: The Microbiology of Wine and Vinifications*; 2nd editio.; Wiley, 2006; ISBN 9780470010341.
42. Volschenk, H.; Vuuren, H.J.J. van; Viljoen-Bloom, M. Malo-ethanolic fermentation in *Saccharomyces* and *Schizosaccharomyces*. *Curr. Genet.* **2003**, *43*, 379–391, doi:10.1007/s00294-003-0411-6.
43. Redzepovic, S.; Orlic, S.; Majdak, A.; Kozina, B.; Volschenk, H.; Viljoen-Bloom, M. Differential malic acid degradation by selected strains of *Saccharomyces* during alcoholic fermentation. *Int. J. Food Microbiol.* **2003**, *83*, 49–61, doi:10.1016/S0168-1605(02)00320-3.
44. Volschenk, H.; Viljoen, M.; Grobler, J.; Petzold, B.; Bauer, F.; Subden, R.E.; Young, R. a; Lonvaud, a; Denayrolles, M.; van Vuuren, H.J. Engineering pathways for malate degradation in *Saccharomyces cerevisiae*. *Nat. Biotechnol.* **1997**, *15*, 253–257, doi:10.1038/nbt0397-253.
45. Delcourt, F.; Taillandier, P.; Vidal, F.; Strehaiano, P. Influence of pH, malic acid and glucose concentrations on malic acid consumption by *Saccharomyces cerevisiae*. *Appl. Microbiol. Biotechnol.* **1995**, *43*, 321–324, doi:10.1007/BF00172832.
46. Peltier, E.; Vion, C.; Abo Saada, O.; Friedrich, A.; Schacherer, J.; Marullo, P. Flor yeast rewire the central carbon metabolism during wine alcoholic fermentation. **2021**, doi: 10.1101/2021.02.27.433177
47. Peltier, E.; Marullo, P.; Coulon, J. Levures œnologiques permettant le contrôle de la teneur en acide des vins et méthodes de sélection de telles levures. 2020.
48. Peltier, E.; Sharma, V.; Raga, M.M.; Roncoroni, M.; Bernard, M.; Yves Gibon; Marullo, P.; Jiranek, V.; Gibon, Y.; Marullo, P. Dissection of the molecular bases of genotype x environment interactions: a study of phenotypic plasticity of *Saccharomyces cerevisiae* in grape juices. *BMC Genomics* **2018**, *19*, 772, doi:10.1186/s12864-018-5145-4.
49. Chernova, M.; Albertin, W.; Durrens, P.; Guichoux, E.; Sherman, D.J.; Masneuf-Pomarede, I.; Marullo, P. Many interspecific chromosomal introgressions are highly prevalent in Holarctic *Saccharomyces uvarum* strains found in human-related fermentations. *Yeast* **2018**, *35*, 141–156, doi:10.1002/yea.3248.
50. Gabriel, S.; Ziaugra, L.; Tabbaa, D. SNP genotyping using the sequenom massARRAY iPLEX Platform. *Curr. Protoc. Hum. Genet.* 2009.
51. Dunnen, J.T. den; Antonarakis, S.E. Mutation nomenclature extensions and suggestions to describe complex mutations: A discussion. *Hum. Mutat.* **2000**, *15*, 7–12, doi:10.1002/(SICI)1098-1004(200001)15:1<7::AID-HUMU4>3.0.CO;2-N.
52. Peltier, E.; Bernard, M.; Trujillo, M.; Prodhomme, D.D.; Barbe, J.-C.; Gibon, Y.; Marullo, P. Wine yeast phenomics: a standardized fermentation method for assessing quantitative traits of *Saccharomyces cerevisiae* strains in enological conditions. *PLoS One* **2018**, *13*, 191353, doi:10.1101/191353.
53. Stitt, M.; McC., L.R.; Gerhardt, R.; Heldt, H.W. Metabolite Levels in Specific Cells and Subcellular Compartments of Plant

- Leaves. *Methods Enzymol.* **1989**, *174*, 518–552.
54. R Development Core Team, R. R: A Language and Environment for Statistical Computing. *R Found. Stat. Comput.* 2011, *1*, 409.
 55. Taillandier, P.; Gilis, M.; Strehaiano, P.; Taillandier, P.; Gilis, M.; Strehaiano, P. Deacidification by Schizosaccharomyces : interactions with Saccharomyces. *J. Biotechnol.* **1995**, *40*, 199–205.
 56. Yéramian, N.; Chaya, C.; Suárez Lepe, J.A. L-(-)-malic acid production by Saccharomyces spp. during the alcoholic fermentation of wine (1). *J. Agric. Food Chem.* **2007**, *55*, 912–919, doi:10.1021/jf061990w.
 57. Vilanova, M.; Ugliano, M.; Varela, C.; Siebert, T.; Pretorius, I.S.; Henschke, P.A. Assimilable nitrogen utilisation and production of volatile and non-volatile compounds in chemically defined medium by Saccharomyces cerevisiae wine yeasts. *Appl. Microbiol. Biotechnol.* **2007**, *77*, 145–157, doi:10.1007/s00253-007-1145-z.
 58. Marullo, P.; Yvert, G.; Bely, M.; Aigle, M.; Dubourdieu, D. Genetic mapping and introgression of QTL as a selection strategy in industrial yeasts. In Proceedings of the Genetics of Industrial Microorganisms; Pragua, 2006.
 59. Mortimer, R.K.; Romano, P.; Suzzi, G.; Polsinelli, M. Genome Renewal: A New Phenomenon Revealed From a Genetic Study of 43 Strains of Saccharomyces cerevisiae Derived From Natural Fermentation of Grape Musts. *YEAST* **1994**, *10*, 1543–1552.
 60. Giudici, P.; Solieri, L.; Pulvirenti, A.M.; Cassanelli, S. Strategies and perspectives for genetic improvement of wine yeasts. *Appl. Microbiol. Biotechnol.* **2005**, *66*, 622–628, doi:10.1007/s00253-004-1784-2.
 61. Salmon, J. l-Malic-acid permeation in resting cells of anaerobically grown Saccharomyces cerevisiae. *Biochim. Biophys. Acta - Biomembr.* **1987**, *901*, 30–34, doi:10.1016/0005-2736(87)90253-7.
 62. Ribéreau-Gayon, P., Dubourdieu, D., Donéche, B. & A.L. *Handbook of Enology Volume 1 The Microbiology of Wine and Vinifications 2 nd Edition Handbook of Enology Volume 1 The Microbiology of Wine and Vinifications 2 nd Edition*; 2006; Vol. 1; ISBN 9780470010341.
 63. Vailiant, H.; Formisyn, P.; Gerbaux, V. Malolactic fermentation of wine: study of the influence of some physico-chemical factors by experimental design assays. *J. Appl. Bacteriol.* **1995**, *79*, 640–650, doi:10.1111/j.1365-2672.1995.tb00949.x.
 64. Versari, A.; Parpinello, G.P.; Cattaneo, M. Versari- et al- (1999)- Leuconostoc oenos and malolactic fermentation in wine: a review-Journal of Industrial Microbiology and Biotechnology . *J. Ind. Microbiol. Biotechnol.* **1999**, *447–455*, doi:10.1038/sj.jim.2900733.
 65. Hospital, F.; Charcosset, A. Marker-assisted introgression of quantitative trait loci. *Genetics* **1997**, *147*, 1469–1485.
 66. Bouchez, A.A.; Hospital, F.F.; Causse, M.; Gallais, A.A.; Charcosset, A. Marker-assisted introgression of favorable alleles at quantitative trait loci between maize elite lines. *Genetics* **2002**, *162*, 1945–1959, doi:10.2135/cropsci2012.04.0219.
 67. Bloom, J.S.; Kottenko, I.; Sadhu, M.J.; Treusch, S.; Albert, F.W.; Kruglyak, L. Genetic interactions contribute less than additive effects to quantitative trait variation in yeast. *Nat. Commun.* **2015**, *6*, 8712, doi:10.1038/ncomms9712.
 68. Steinmetz, L.M.; Sinha, H.; Richards, D.R.; Spiegelman, J.I.; Oefner, P.J.; McCusker, J.H.; Davis, R.W. Dissecting the architecture of a quantitative trait locus in yeast. *Nature* **2002**, *416*, 326–330, doi:10.1038/416326a.
 69. Sinha, H.; David, L.; Pascon, R.C.; Clauser-Münster, S.; Krishnakumar, S.; Nguyen, M.; Shi, G.; Dean, J.; Davis, R.W.; Oefner, P.J.; et al. Sequential Elimination of Major-Effect Contributors Identifies Additional Quantitative Trait Loci Conditioning High-Temperature Growth in Yeast. *Genetics* **2008**, *180*, 1661–1670, doi:10.1534/genetics.108.092932.
 70. Liti, G.; Louis, E.J. Advances in Quantitative Trait Analysis in Yeast. *PLoS Genet.* **2012**, *8*, doi:10.1371/journal.pgen.1002912.
 71. Bonnett, D.G.; Rebetzke, G.J.; Spielmeier, W. Strategies for efficient implementation of molecular markers in wheat breeding. *Mol. Breed.* **2005**, *15*, 75–85, doi:10.1007/s11032-004-2734-5.
 72. Fatichenti, F.; Farris, G.A.; Deiana, P.; Ceccarelli, S. Malic acid production and consumption by selected of Saccharomyces cerevisiae under anaerobic and aerobic conditions. *Appl. Microbiol. Biotechnol.* **1984**, *19*, 427–429, doi:10.1007/BF00454382.
 73. Ramon-Portugal, F.; Seiller, I.; Taillandier, P.; Favarel, J.L.; Nepveu, F.; Strehaiano, P. Kinetics of Production and Consumption of Organic Acids during Alcoholic Fermentation by Saccharomyces cerevisiae. *Food Technol. Biotechnol.* **1999**,

37, 235–240.

74. Vilanova, M.; Ugliano, M.; Varela, C.; Siebert, T.; Pretorius, I.S.; Henschke, P.A. Assimilable nitrogen utilisation and production of volatile and non-volatile compounds in chemically defined medium by *Saccharomyces cerevisiae* wine yeasts. *Appl. Microbiol. Biotechnol.* **2007**, *77*, 145–157, doi:10.1007/s00253-007-1145-z.
75. Kunkee, R.E. Some roles of malic acid in the malolactic fermentation in wine making. *FEMS Microbiol. Lett.* **1991**, *88*, 55–71, doi:10.1016/0378-1097(91)90696-8.
76. Volschenk, H.; van Vuuren, H.J.J.; Viljoen-Bloom, M. Malic Acid in Wine: Origin, Function and Metabolism during Vinification. *South African J. Enol. Vitic.* **2006**, *27*, doi:10.21548/27-2-1613.
77. Volschenk, H.; Viljoen, M.; Grobler, J.; Petzold, B.; Bauer, F.; Subden, R.E.; Young, R.A.; Lonvaud, A.; Denayrolles, M.; Van Vuuren, H.J.J. Engineering pathways for malate degradation in *saccharomyces cerevisiae*. *Nat. Biotechnol.* **1997**, *15*, 253–257, doi:10.1038/nbt0397-253.
78. Jeppsson, H.; Yu, S.; Hahn-Hägerdal, B. Xylulose and glucose fermentation by *Saccharomyces cerevisiae* in chemostat culture. *Appl. Environ. Microbiol.* **1996**, *62*, 1705–1709.