

The nerves to conduct a multiple sclerosis crime investigation

Sameeksha Chopra^{1,2,3}, Zoë Myers^{1,2,3}, Henna Sekhon^{1,2,3} and Antoine Dufour^{1,2,3}

¹ McCaig Institute for Bone and Joint Health, Calgary, Alberta, Canada

² Department of Physiology and Pharmacology, University of Calgary, Calgary, Alberta, Canada

³ Department of Biochemistry and Molecular Biology, University of Calgary, Calgary, Alberta, Canada

Abstract

Multiple sclerosis (MS) is a chronic inflammatory neurodegenerative autoimmune disease characterized by aberrant infiltration of immune cells into the central nervous system (CNS) and by the loss of myelin. Sclerotic lesions and various inhibitory factors hamper remyelination processes within the CNS. MS patients typically experience gradual cognitive and physical disabilities as the disease progresses. The etiology of MS is still unclear and emerging evidence suggests that microbiome composition could play a much more significant role in disease pathogenesis than was initially thought. Initially believed to be isolated to the gut microenvironment, we now know that the microbiome plays a much broader role in various tissues and is essential in the development of the immune system. Here, we present some of the unexpected roles that the microbiome plays in MS and discuss approaches for the development of next-generation treatment strategies.

Keywords: Multiple sclerosis (MS), microbiome, bacteria, virus, central nervous system (CNS), immunity.

Multiple sclerosis (MS) is a chronic inflammatory autoimmune disease characterized by aberrant infiltration of immune cells into the central nervous system (CNS) parenchyma¹. Immune cell infiltration is accompanied by the loss of myelin (demyelination) and neurodegeneration of the CNS^{1,2}. The accumulation of sclerotic lesions and inhibitory factors can significantly hamper remyelination processes within the CNS, causing MS patients to experience debilitating cognitive and physical disabilities as the disease progresses^{1,2}. While the etiology of MS is unknown, emerging evidence continues to suggest that the microbiome may play a significant role in disease pathogenesis^{1,3,4}. Autoimmune diseases, including MS, are more prevalent in countries that have a higher standard for sanitation and a greater use of antibiotics⁵. This phenomenon is described as the *hygiene hypothesis*⁶, which suggests that overly hygienic westernized lifestyles may result in gut dysbiosis and dysregulated immune responses. Several environmental factors such as diet and vitamin D exposure have been linked to an increased risk of MS and may help explain the distinct geographical distribution of the disease⁶. People moving from countries with low prevalence of MS to those with high prevalence of MS adopt a higher risk of developing the disease⁶⁻⁹, perhaps facilitated by the changes in lifestyle and diet that drive a shift in the composition of the intestinal flora. People with MS¹⁰⁻¹² and animal models of the disease¹³ have altered gut microbiota composition, which brings us to question whether the microbiome could be a driver of MS pathogenesis. There is an imminent need to better characterize changes in the microbiome composition of individuals living with MS and study the complex mechanisms by which microbiota can influence host biology and affect disease pathogenesis.

Box 1: What if the CNS of MS patients was compared to a crime scene where a detective would arrive at the crime scene to investigate the *damage* done and gather forensic evidence.

The victim: Central Nervous System

- Damage myelin coating on nerves
- Evidence of an activated immune response in the CNS that resulted in clearance of myelin debris and promotion of remyelination
- Bacterial contribution that impacted resident and peripheral immune cells in addition to modulation of CNS myelination

Investigating the Multiple sclerosis *Crime scene*

MS patients are monitored and diagnosed after presenting with lesions visible on an MRI that are disseminated in time and space. However, there is little known about the initiating cellular and molecular events that contribute to dysregulated inflammation prior to formation of MS lesions. There are likely drastic changes to the CNS tissue microenvironment that accompany neuroinflammation and the subsequent development of MS plaques. Many of the inhibitory factors that accumulate within plaques impair remyelination in the CNS and thus limit the organs endogenous tissue repair processes.

Breaking and Entering the Blood brain barrier

Homeostasis is maintained in a healthy CNS through the regulated entry of substances from peripheral circulation. Tight-junction proteins create a selective blood brain barrier (BBB) surrounding the delicate CNS tissue. However, BBB breakdown in MS patients is accompanied by increased peripheral immune cell infiltration into the CNS. There are many mechanisms proposed to cause a dysfunctional BBB including inflammatory cytokines, activated peripheral immune cells recognizing CNS antigens, secretion of proteases and microbial derived factors^{14,15}.

Alterations in the gut microbiome could affect the permeability of the BBB and have indications in neurological diseases such as MS. In a study that investigated the effects of antibiotics on BBB integrity in a germ-free mouse model of MS, a decrease gut microbiome diversity was positively correlated with an increase in BBB permeability and a downregulation of short chain fatty acids (SCFAs)-producing microbes was observed¹⁶. Several studies have reported the effect of gut microbes and their metabolites on BBB permeability¹⁷. For example, germ-free mice colonized with bacterial strains that produce SCFAs reinforced the integrity of the BBB by upregulating the expression of tight junction proteins¹⁵. Furthermore, resident microglial cells in the brain can also modulate BBB permeability¹⁸. As described next, the microbiome can influence the function of cell types resident to the CNS, highlighting the complex interplay between the gut and brain. In MS patients, a deficit of SCFAs-producing bacteria likely contributes to increased BBB permeability and subsequent entry of neuroinflammatory factors into the CNS. People with MS also often have elevated serum zonulin concentrations¹⁹, which is a protein known to be released in response to gut dysbiosis²⁰ capable of increasing intestinal permeability and breaking down the BBB through the regulation of tight junction proteins^{19,21}. Altering the microbiome or delivering BBB-reinforcing microbial metabolites may be a strategy to repair a damaged BBB, stop peripheral immune cell access into the CNS and thereby prevent the exacerbation of disease.

Is the Peripheral Immune System an accomplice?

The increase of immune cell infiltration into the CNS during MS relapses supports the notion that MS is an immune-mediated disease. However, whether neuroinflammation during the early stages of MS is a result of infiltrating immune cells activated in the periphery or is a process initiated within the CNS itself remains to be characterized. Gut bacteria transferred from MS patients into mice exacerbate the development of MS-like disease in various mouse models^{22,23}, showing that the intestinal microbiome is capable of influencing disease progression. Despite the less obvious connection between the brain and gut, growing evidence suggests that intestinal immune responses and neuroinflammation are interconnected (**Figure 1**).

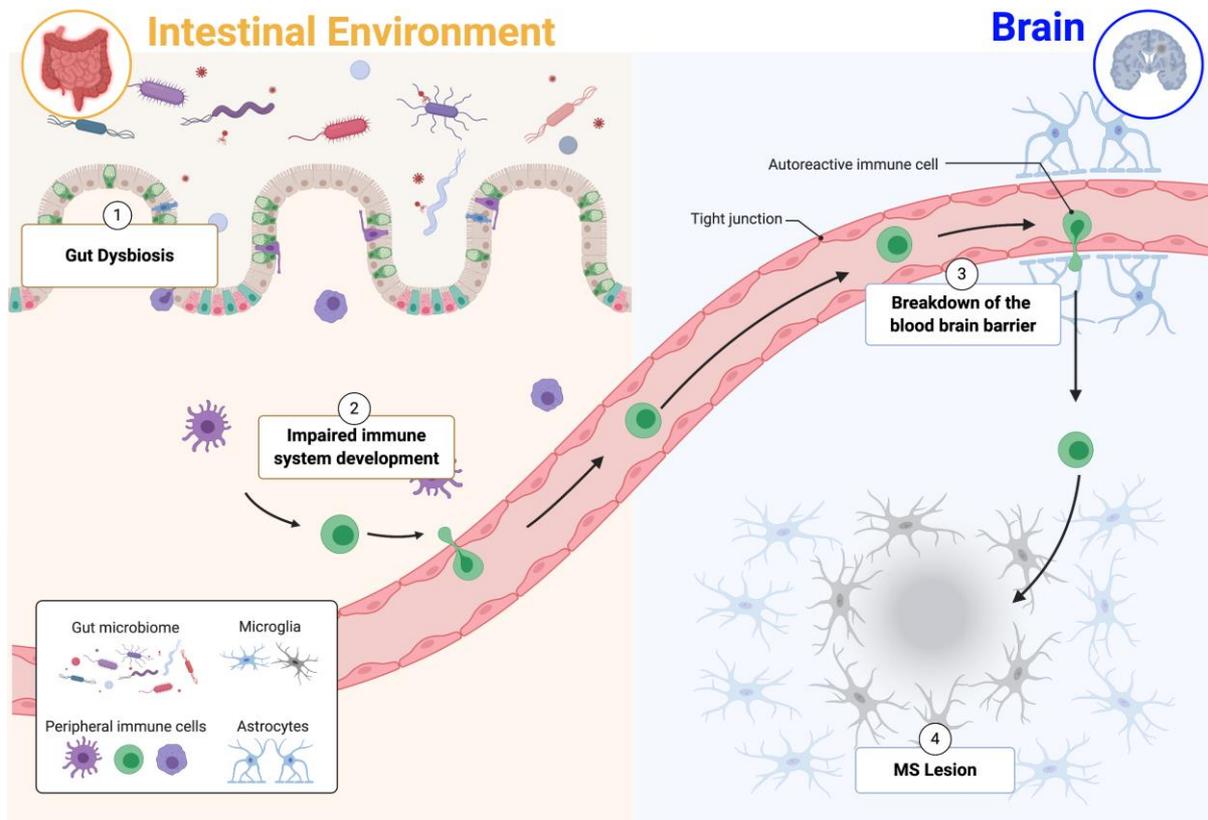


Figure 1: Gut-immune-brain connection. 1) Gut dysbiosis can impair mucosal immunity homeostasis. 2) Changes in immune system development are linked to multiple autoimmune diseases. 3) Blood brain barrier breakdown is a hallmark of multiple sclerosis (MS) and the gut microbiome is a key regulator of blood brain barrier permeability. 4) Peripheral immune cell infiltration and subsequent autoimmune responses contribute to the development and exacerbation of MS lesions.

Maternal and in utero effects on immune system development

The maternal microbiome has been shown to affect fetal immune system development²⁴, which may confer lifelong consequences on the offspring by modulating their risk of developing autoimmune diseases such as MS. Interestingly, fetal thymic CD4⁺ T cell and regulatory T cell (Treg) development is compromised in germ-free mice²⁵. Maternal supplementation of the intestinal bacterial metabolite acetate significantly rescued CD4⁺ T cell and Treg development by upregulating the autoimmune regulator (AIRE)²⁵, which is essential for self-tolerance induction and Treg development early in life. In humans, low maternal serum acetate levels are mirrored in the fetus, which may predispose the offspring towards impaired self-tolerance. The specific connection between *in utero* effects of bacterial-derived factors on immune development in offspring later in life, particularly in the context of multiple sclerosis, has yet to be explored in depth. A case-control study analyzed maternal vitamin D concentrations in the serum of pregnant women in the Finish Maternity Cohort whose children had developed MS²⁶. Maternal vitamin D deficiency during early pregnancy was associated with a 2-fold increase of MS risk in the offspring²⁶. Low concentrations of neonatal vitamin D are also associated with increased risk of MS²⁷. Interestingly, birth month influences the risk of developing MS later in life, with the risk of MS peaking in May and dropping in November

which may be linked to maternal vitamin D levels²⁸. Additionally, bacterial/viral infections during pregnancy may also contribute to the unique birth month pattern of MS risk. Cohort studies that assess bacterial metabolites using unbiased metabolomics may be insightful to understand the contribution of the maternal microbiome in modulating the risk of offspring developing MS.

Immune system development

Epidemiological data support the idea that exposure to environmental factors in childhood may be strong determinants of MS risk^{27,29-31}. As migration studies suggest, the incidence rate of MS in migrants tends to be somewhere between the incidence rate associated with their birthplace and the incidence rate associated with their final residence. If this migration happens in childhood, then the incidence rate tends to be closer to that associated with their final residence^{27,31,32}. For example, natives of the Caribbean islands and Asia do not experience a significant increased risk of developing MS upon migrating to the United Kingdom (UK), however high rates of MS are documented in their UK-born children³¹. This may suggest that individuals spending early life in low-risk areas tend to benefit from long-lasting protection against MS that is not transferred to their children^{27,31}, hinting at the possibility that these factors may include bacteria and viruses that shape the development of their immune system. Perhaps there are vulnerable periods in one's early life where environmental factors are particularly critical for immune system development affecting one's future risk of developing MS and other autoimmune diseases.

The gut microbiome plays an essential role in the development of the immune system³³. The gut microbes and the host exist in a symbiotic relationship and when this relationship becomes dysregulated, dysbiosis can occur, resulting in defects in the immune response, immune system disorders and triggering of autoimmune diseases. During fetal and infant development, the gut microbiome is developing alongside the immune system³⁴. As demonstrated in germ-free mice, the immune system requires the colonization of the gut bacteria to properly mature. Initial colonization of gut microbes occurs *in utero*, indicating a possible transfer of bacteria from the blood stream of the mother to the fetus, and this continues after birth as the infant breast feeds and begins eating solid foods³⁵. The method of birth delivery may affect the development of the gut microbiota as it is suggested that Caesarian delivered babies have low bacterial diversity within their gut³⁵. This makes these infants more susceptible to immune disorders like asthma and Celiac disease^{35,36}. Similar results were seen in infants who use baby formula rather than breastfeeding indicating that there are various environmental factors that play a role in the development of the gut microbiome and that a slightly impaired gut microbiota (decrease in diversity) may lead to immune disorders later on in life. Additionally, the immunogenicity of early colonizing intestinal bacteria may have profound impacts on the repertoire of immune responses later in life. Vatanen *et al.* followed gut microbiome development in 222 infants in Northern Europe from birth until the age of three³⁷. Bacteroides species that inhibit innate immune signalling were lowly abundant in infants from Russia, where early-onset autoimmune diseases are less prevalent as compared to Finland and Estonia³⁷. In line with the hygiene hypothesis, early colonization with immunologically silencing microbiota may impair immune education and maturation.

Short chain fatty acids (SCFAs) can modulate immunity

Gut bacteria play a crucial role in extracting key nutritional factors from a healthy diet. Non-digestible carbohydrates can be fermented by specific bacterial species to make metabolites that modulate host biology^{38,39}. SCFAs produced by the intestinal microbiota are critical in the development and maintenance of a healthy immune system^{38,40}. Importantly, SCFAs, such as acetate, propionate, butyrate and pentanoate, can exert anti-inflammatory effects on the host and are often downregulated in chronic inflammatory conditions⁴¹. In MS patients, serum concentrations of butyric acid (BA) are reduced and medium chain fatty acids (MCFAs), such as caproic acid (CA), are increased⁴². The altered BA/CA ratio correlates with the immunological profile of MS patients, as they present with an increase in Th1 and Th17 cells as well as a decrease in Treg lymphocytes⁴². Concurrently, the microbiota of MS patients is depleted of BA producers⁴². Decreased serum butyric acid levels correlate with increased intestinal permeability as measured by serum intestinal fatty acid binding protein (I-FABP) and bacterial lipopolysaccharide (LPS) in MS patients compared to healthy controls. The presence of these factors in the blood suggests impaired gut barrier integrity in MS. Pentanoate is a potent regulator of immunometabolism as it can reprogram the metabolic activity of lymphocytes. In experimental mouse models of multiple sclerosis, pentanoate-induced regulatory B cells protect against autoimmunity⁴³. Additionally, pentanoate induces IL-10 production in lymphocytes and reduces IL-17A production by epigenetically modifying CD4 T cells⁴³. Germ-free mice are highly resistant to the development of experimental autoimmune encephalomyelitis (EAE), a mouse model of MS. However, segmented filamentous bacteria (SFB) induce pathogenic Th17 immune response and promote CNS inflammation in SFB mono-colonized germ-free animals⁴³. Interestingly, administering pentanoate in SFB-colonized germ-free mice ameliorates EAE and decreases Th17 cells in the CNS⁴³.

The gut, an unexpected suspect in MS, modifies the autoimmune response

Macrophages and Microglia

Microglia are tissue resident immune cells in the CNS that can influence inflammation and neurodegeneration by secreting factors and coordinating with other cell types, such as astrocytes, oligodendrocytes, neurons and peripheral immune cells that have infiltrated into the CNS⁴⁴. A thorough characterization of how the microbiota influences the function of neural cell types in MS remains to be done. Gut microbiota can metabolize dietary tryptophan to produce aryl hydrocarbon receptor (AHR) agonists that can limit CNS inflammation by acting on AHR receptors expressed by microglial cells⁴⁵. Tryptophan metabolism has been of interest in autoimmunity research as metabolic products of the kynurenine pathway used to metabolize tryptophan are known to exert several effects on the immune system, including modulating immunotolerance⁴⁶. Several studies have investigated the levels of kynurenines in MS patients or in the EAE mouse model, demonstrating that aberrant kynurenine pathway activity is associated with increased severity of disease⁴⁶⁻⁴⁹. Tryptophan-derived metabolites, such as indole-3-propionic acid, are produced by the microbiota and are associated with MS relapses⁴⁶. However, further research is required to understand the specific pathophysiological role of tryptophan-derived microbial metabolites in MS. Bacterial products from members of the gut microbiota, such as SCFAs, play a substantial role in microglia homeostasis as temporal

eradication of the host microbiota or limited microbial complexity can significantly change microglia properties⁵⁰. Duscha *et al.* showed that supplementing MS patients with the SCFA propionic acid increases Tregs and can be a promising immunomodulatory supplement to MS drugs⁵¹. While the study did not assess the impact of SCFA on tissue resident immune cells in the CNS, it is likely that SCFA treatment modulates the maturation and activation of microglia as well⁵⁰. Ultimately, applying our understanding of microbiome-host interactions specifically pertaining to the CNS may help to design better therapeutic approaches for neurological illnesses.

T cells

The gut-brain axis plays a key role in neuroimmunology, with emerging evidence suggesting that encephalitogenic immune responses may be driven by intestinal dysbiosis in MS^{52,53}. Th17 cells are key mediators of CNS autoimmunity and their role in MS has been established in both humans and animal models⁵⁴. Specific microbiota modifications in the human intestinal microenvironment, including a higher *Firmicutes/Bacteroidetes* ratio, increased relative abundance of *Streptococcus*, and decreased *Prevotella* strains, have been implicated in promoting Th17 cell expansion and correlating with brain autoimmunity in MS patients with high disease activity⁵². In an adoptive transfer mouse model of MS, CNS-specific Th17 cells first migrate to the intestine to proliferate during disease pathogenesis prior to reaching the CNS and inducing neurological symptoms⁵⁴. The pro-inflammatory properties of encephalitogenic Th17 cells are strengthened in part by gut microbiota changes in mice. Blocking $\alpha 4\beta 7$ -integrin and its cognate ligand mucosal addressin cell adhesion molecule 1 (MAdCAM-1) disrupts Th17 cell intestinal homing and attenuates EAE severity^{54,55}. In another study, eradication of the gut microbiome using antibiotics prevented the induction of CNS inflammation by gut derived T cells and abolished the development of EAE⁵⁶. These studies implicate the intestine as a regulatory checkpoint for regulating autoimmune T cells. The gut microbiota may also modulate the disease course of MS. In the EAE model, differentially abundant intestinal bacteria determine susceptibility to chronic progressive versus relapse remitting forms of disease⁵⁷. The contribution of intestinal T cell subsets, including the presence of mucosal associated invariant T (MAIT) cells in brain lesions⁵⁸ and peripheral circulation of MS patients⁵⁹, is starting to be recognized in the pathophysiology of MS. Ultimately, studies in animal models and patient data suggest that the microbial composition of the gut may play a critical role in catalyzing CNS-specific autoimmune responses in MS.

B cells

Gut-associated B cells, most of which differentiate into IgA-secreting plasma cells (PC), are predominantly found in Peyer's patch⁶⁰. IgA antibodies are a critical first line of defence against antigens in the intestinal microenvironment. During EAE, there is a downregulation of IgA PCs in the gut and an upregulation of IgA PCs in the CNS⁶¹. PCs that egress from the gut into the CNS suppress the symptoms of EAE via the interleukin (IL)-10 pathway⁶¹. Therefore, intestinal derived IgA B cells may represent a population of regulatory cells that can be recruited to tissues to mediate inflammation independent of their receptor specificities. Also, intestinal PCs may be neuroprotective by migrating to and limiting neuroinflammation of the damaged CNS. Future studies should test the efficacy of treatments that mobilize IgA PCs in

MS. Overall, the presence and the functions of immunosuppressive IgA PCs in the CNS of MS patients needs to be further characterized.

Immune activation: What is the motive for the crime?

Autoimmune responses often exacerbate MS pathogenesis, however the specific triggers that initiate the activation of pathogenic self-reactive immune cells and lead to the onset of MS remain unidentified. One possible trigger is the cross-reactivity of immune cells to microbiota-derived peptides⁶². Genetic and environmental risk factors are likely both implicated in MS pathogenesis². Some of the most common environmental risk factors potentially involved in MS are viral infections². Viral models of MS, such as the Theiler's murine encephalomyelitis virus (TMEV) model⁶³, are commonly used to study MS pathology. The need to study viral contributions to disease is emerging as pathogenic human endogenous retroviruses (HERVs), normally dormant and comprising 8% of the human genome, are upregulated in response to environmental factors in MS⁶⁴⁻⁶⁶. Another virus implicated in MS pathogenesis is the Epstein-Barr virus (EBV), a member of the herpes virus family that causes infectious mononucleosis^{67,68}. While more than 95% of healthy individuals demonstrate an immune response to EBV, virtually all MS patients are seropositive for EBV^{69,70}. Libbey *et al.*⁷¹ hypothesize that MS could be triggered via molecular mimicry, whereby specific peptides secreted by pathogens have sequence and/or structural similarities to self-antigens. Previous studies have suggested the role of molecular mimicry in the pathogenesis of MS through Haemophilus influenzae viral infections⁷² and a potentially similar mechanism has been observed for EBV infection⁷³. The proposed cascade of events leading to CNS autoimmunity following EBV infection involve the proliferation and maturation of T-cells against Epstein-Barr virus nuclear antigen 1 (EBNA1)⁷⁴. Tengvall *et al.*⁷⁴ hypothesized that T cell responses against EBNA1 may cross-react with a protein known as anoctamin 2 (ANO2) expressed in the CNS. Interestingly, autoantibody levels against ANO2 have been shown to be increased in the cerebral spinal fluid (CSF) of MS patients⁷⁴. ANO2 is a calcium activated chloride channel important for ion transport and control of neuronal excitability that is expressed in hippocampal and cortical regions, and specifically detected near and inside of MS plaques⁷⁴. EBV remains a *priority suspect* as: 1) MS does not occur until people have developed an immune response to EBV, and 2) a history of mononucleosis doubles the risk for MS⁷⁴. Molecular mimicry remains the primary explanation for the link between EBV and MS.

Other viruses may have similar indications in the pathogenesis of MS. Human herpesvirus 6 (HHV6) is a member of the herpesvirus family and has two subtypes: HHV6A and HHV6B⁷⁵. Both types preferentially attack and/or affect the nervous system, but HHV6A has greater neurotropism which may implicate a closer link to MS⁷⁵. Opsahl and Kennedy detected significantly higher levels of HHV6 in post-mortem lesioned brain tissue from MS patients compared to the healthy controls⁷⁶. Another study reported that a high antibody response against the HHV6A-specific antigen immediate-early protein 1A (IE1A) positively correlates with MS⁷⁷. Individuals with strong humoral immunity against IE1A, particularly those younger than 20 years old, have a higher risk of developing MS later in life⁷⁷. The positive association between the IE1A antigen from HHV6-A and MS is intriguing considering that HHV6A can impede the migration of oligodendrocyte progenitor cells and establish latent infection in

oligodendrocytes⁷⁸, the myelin-producing cells presumed to be a target of the MS-related autoimmune responses. However, the mechanistic role of HHV6 in the pathology of disease remains unknown. Human endogenous retroviruses (HERV) are usually epigenetically silenced within our genome but certain triggers, including transactivation by exogenous viral infections such as EBV or other viruses, may lead to the re-expression of HERVs. HERV type W (HERV-W), formerly known as multiple sclerosis-associated retrovirus (MSRV), was discovered in the CSF of MS patients and is frequently detected in the brains of MS patients⁷⁹. In MS lesions, myeloid cells contain the envelope protein ENV from HERV-W, which has recently been shown to polarize microglia and contribute to neurodegeneration⁷⁹. A Phase IIb clinical trial (NCT02782858) tested the efficacy of GNBAC1 treatment, a humanized anti-ENV IgG₄ monoclonal antibody⁸⁰. GNBAC1 exerted significant neuroprotective effects in MS patients within 1 year of treatment including a 1) 31% reduction of cortical atrophy, 2) 72% reduction of thalamic atrophy and, 3) 63% reduction of T1 hypointense lesions correlating with permanent brain tissue damage^{79,80}. Several studies suggest an association between viral infections and MS, however there is still a need to delineate if there is a causal relationship between viruses and the onset of MS.

Another area of research that remains underexplored is the contribution of bacteriophages, viruses that infect bacteria, in MS. It is plausible that bacteriophages are the underlying cause of intestinal dysbiosis in MS patients, however they have also been implicated in mimicry of MS-relevant autoantigens. The *Synechococcus* phage is suggested to be a major contributor to the molecular mimicry phenomenon⁸¹, whereby antibodies raised to viral proteins cross react with self-antigens leading to autoimmune responses. Bacteriophages may also induce chronic inflammation by changing gut microbiome composition and in turn increasing intestinal permeability^{82,83}. Zhao *et al.* found less diversity in the intestinal virome of children who subsequently developed serum autoantibodies associated with progression to type 1 diabetes (cases) as compared to healthy controls⁸⁴. Populations of bacteriophages differed significantly between cases and controls⁸⁴. The disease-discriminating bacteriophage sequences identified also significantly correlated with different bacterial OTUs in both cases and controls, suggesting unique virome-bacterial microbiome interactions were present prior to development of autoantibodies⁸⁴. Ultimately, epidemiological evidence, genetic association and mechanistic data in humans is needed to elucidate the role of viruses in the pathogenesis of MS. To this end, a broader characterizing of the MS patient virome⁸⁵ will be valuable.

Deception in the interrogation room: Crosstalk between host and bacteria, not two distinct species, but one intermixed system

The investigation of the complex crosstalk between host and bacteria is still in its infancy. Bacteria derived factors have been shown to influence host proteases⁸⁶ and host proteins were shown to modulate the activity of bacterial proteases⁸⁷. In MS patients, the human protease inhibitor, cystatin C, is significantly upregulated in brain biopsies⁸⁸. A bacterial protease, IdeS, produced by the human pathogen *Streptococcus pyogenes* (Group A *Streptococcus* [GAS]) hijacks mechanisms deployed by the host to inhibit proteases by using cystatin C as a cofactor to increase its own activity⁸⁷. The intriguing finding that a host protease inhibitor may increase the activity of a bacterial protease further supports our lack of understanding of the complexity

of host-microbial interactions. Cystatin C is also highly expressed in the brains of EAE mice⁸⁸. Whether it interacts with bacterial proteases to influence disease pathogenesis in MS is currently unknown, however, it does seem to have a detrimental function in myelin oligodendrocyte glycoprotein 35-55-induced EAE in female mice, but not in male demonstrating a sex-specific role⁸⁸. An analysis of the fecal proteome of EAE mice during disease latency demonstrated a transient increase in host protease inhibitors that inversely correlated with disease severity⁸⁹. The administration of the antibiotic vancomycin attenuated EAE symptoms and elevated protease inhibitors⁸⁹ further supporting the idea that the microbiota modulates host biology.

Guo *et al.*⁸⁶ identified microbial metabolites with protease inhibitory activity produced by the gut bacteria of most healthy people. These metabolites impact cathepsins' activity in human cell proteomes⁸⁶, demonstrating new biological roles in interspecies signalling. Compared to healthy controls, MS patients present with a significant decrease in the abundance of certain bacteria, including species belonging to the genus *Clostridium*⁹⁰, which may produce microbial metabolites that inhibit host cell cathepsins⁸⁶. Accordingly, MS patients have increased cathepsin activity which may contribute to disease pathogenesis (**Figure 2**). Hypomethylation of the cathepsin Z locus and increased cathepsin Z transcripts were detected in the pathology-free regions of MS brains as compared to healthy controls⁹¹. EAE mice deficient in cathepsin Z exhibit attenuated neuroinflammation and demyelination⁹². Cathepsin Z deficiency dramatically reduced circulating interleukin 1 beta levels and comprised the ability of mice to develop Th17 responses critical for the development of EAE⁹². Perhaps restoring the balance of the gut microbiome may help to regulate the aberrant activity of host proteases such as cathepsins seen in chronic inflammatory diseases such as MS.

Metabolites produced by gut bacteria are also capable of influencing immune homeostasis⁹³, therefore alterations in gut bacteria composition may have drastic effects on immune responses (**Figure 2**). CD4⁺Foxp3⁺ Tregs are critical mediators of immune homeostasis and the development of this cell population is directly affected by interactions with gut microbiota⁹³. The spore-forming components of intestinal microbiota, specifically *Clostridia* clusters IV and XIVa, was shown to promote Tregs by providing an environment rich in transforming growth factor (TGF)- β to drive Treg development⁹⁴. The microbiome of MS patients has a reduction of bacteria belonging to *Clostridia* clusters IV and XIVa - potent producers of SCFAs that regulate inflammation^{90,95}. Mizuno *et al.*⁹⁵ demonstrated that administration of SCFAs suppresses lymphocyte-mediated systemic inflammation in EAE, thereby reducing EAE severity. Ultimately, further research characterizing changes in bacteria-host interactions will contribute to a stronger understanding of the molecular mechanisms affecting MS pathogenesis.

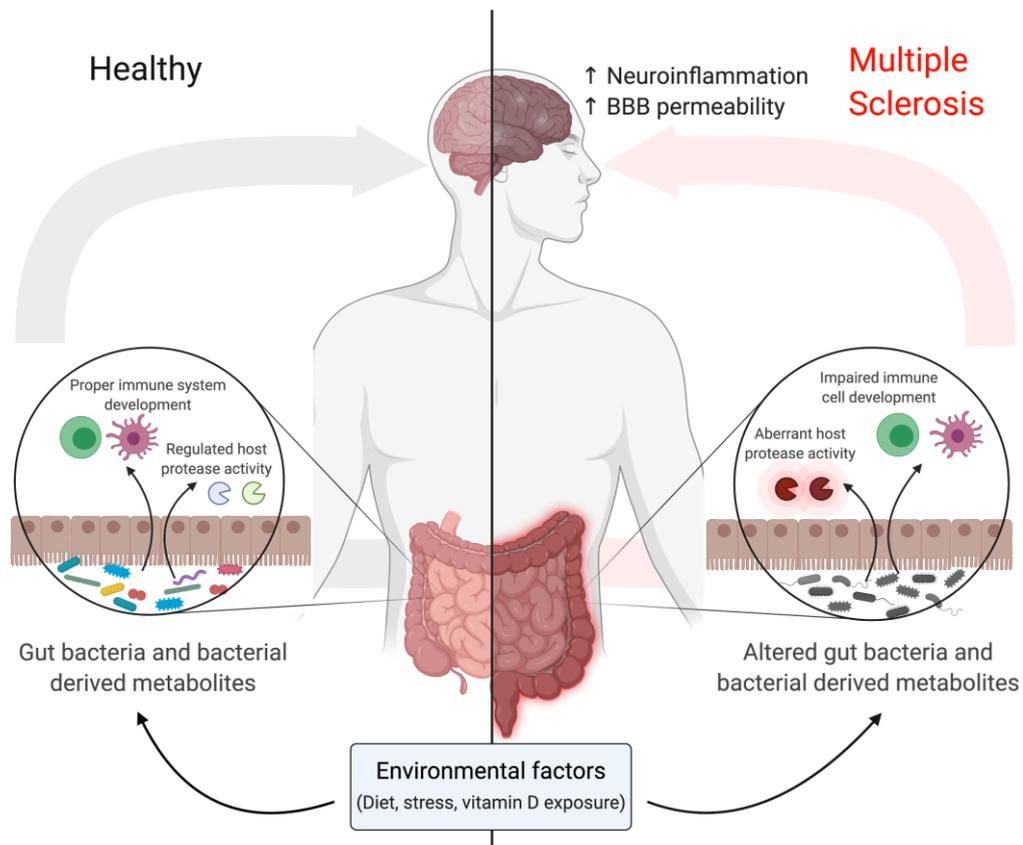


Figure 2: Gut microbiome can modulate host biology. There are known environmental factors such as diet, stress and vitamin D levels that can impact exacerbation of MS.

Interrogating the suspect: Is the gut behind the increased crime rate in the CNS?

Animal models of MS have provided novel insights into the effect of the gut microbiota on MS pathogenesis. In the EAE murine model of MS, perturbing the gut microbiota has been demonstrated to affect disease susceptibility via modulation of immune responses⁹⁶. Interestingly, germ-free (GF) mice are resistant to the development of EAE, and show attenuated signs of disease, reduced clinical scores and shorter duration of EAE symptoms⁹⁷, which is likely a result of broadly impaired immune system development in mice that lack an intestinal microbiome⁹⁸. While the molecular interactions between intestinal microbial organisms and the host was shown to affect the balance between pro- and anti-inflammatory immune responses, the immune system in return may further impact the composition of the microbiota⁹⁹. However, changing the gut microbiome composition through the administration of probiotics does not significantly improve remyelination potential in mice afflicted with EAE⁹⁹. Additional research is needed to delineate the cause-and-effect relationship between the role of the microbiome in MS. In cancer immunotherapy, intestinal bacteria have been associated with enhanced efficacy of immune checkpoint inhibitors¹⁰⁰. Future studies should consider assessing the efficacy of probiotic supplementation in conjunction with pharmacological inhibitors and lifestyle changes to elucidate potential synergistic effects of probiotic therapy on maximizing benefit afforded to patients living with MS. Additionally, the human gut commensal *Prevotella histicola* (*P. histicola*) was demonstrated to be as effective as the immunomodulatory drug glatiramer acetate in suppressing disease severity in a preclinical mouse model of MS as demonstrated by a lower cumulative EAE score¹⁰¹. While combination

of *P. histicola* and glatiramer acetate did not result in an additive effect on EAE disease severity¹⁰¹, it would be interesting to investigate whether *P. histicola* works synergistically with a different pharmacological drug approved for MS.

Profiling other tissues beyond the gut

While the gut has been implicating in the initial priming and proliferation of autoreactive CNS-specific immune cells, the lungs are another site potentially important for the reactivation of immune cells prior to disease relapse¹⁰². MS patients have elevated breath methane¹⁰ which is often attributed to the presence of methanogens such as *Methanobrevibacter* in the gut. Methanogens are also found in the respiratory tract^{103,104}, however, the lung microbiome is currently under characterized in health and disease. To better characterize the role that the microbiota plays in MS, it is important to study the microbiome of MS patients at sites other than the gut, such as the microbiota of the skin and lungs. Besides altered intestinal bacteria composition in chronic inflammatory diseases, the influence of other environmental factors, such as changes in the virome, fungome or parasitome, remain underexplored. Importantly, certain microbes and their products have also been implicated to exert protective effects in MS^{105–107}. Infection of MS patients or animal models of the disease with helminth parasites has been shown to be protective due to beneficial immune modulation and improvements in clinical symptoms¹⁰⁵. Unbiased *Omic*s technologies such as genomics, transcriptomics, proteomics and metabolomics, are valuable tools to enhance our understanding of the human microbiome as a whole¹⁰⁸.

Chasing a cure: MedXercise.

Exercise is known to exert a wide range of health benefits. A review by Lozinski and Yong presented the impact of exercise on structural and functional changes in the CNS¹⁰⁹. The therapeutic effects of exercise in MS may, in part, be mediated by direct immunomodulatory effects since exercise is known to promote anti-inflammatory immune responses¹¹⁰. However, exercise may also induce beneficial changes for patients with neuroimmune illnesses by shifting the composition of their microbiota^{111–116}. Whether these effects are solely attributed to physical activity or are influenced by other factors associated with healthier lifestyle, such as an altered diet, needs to be investigated¹¹⁷. It is important to note that the benefits of exercise may be dependent on the composition of the microbiota¹¹⁸, thus combined treatment targeting the microbiota and engaging in exercise may maximize therapeutic effects. Interestingly, a physically active lifestyle is associated with beneficial changes to the microbiome¹¹⁹ and it may prove to be useful in managing MS. Exercise has been shown to work in combination with pharmacological interventions such as clemastine¹²⁰, also known as meclastin, a drug that promotes remyelination and is a H1 histamine antagonist. In a toxin-induced model of focal demyelination, exercise promotes changes in the lesion microenvironment and makes the CNS more conducive to repair¹²⁰. Exercise was shown to be as equally efficacious as the pharmacological treatment clemastine at promoting remyelination of lysolecithin-induced lesions in mice¹²⁰. Furthermore, exercise and clemastine used in combination work synergistically to additively enhance remyelination¹²⁰. While exercise exerts direct neuroprotective effects by promoting anti-inflammatory innate immune responses and increasing phagocytosis of myelin debris¹²⁰, it would be interesting to explore whether these

changes are correlated with a shift in the microbiome. Nonetheless, the potential to couple pharmacological interventions with an adjunctive therapy like exercise, a concept termed MedXercise¹⁰⁹, is exciting as it provides a new means by which maximal benefit of treatments can be afforded.

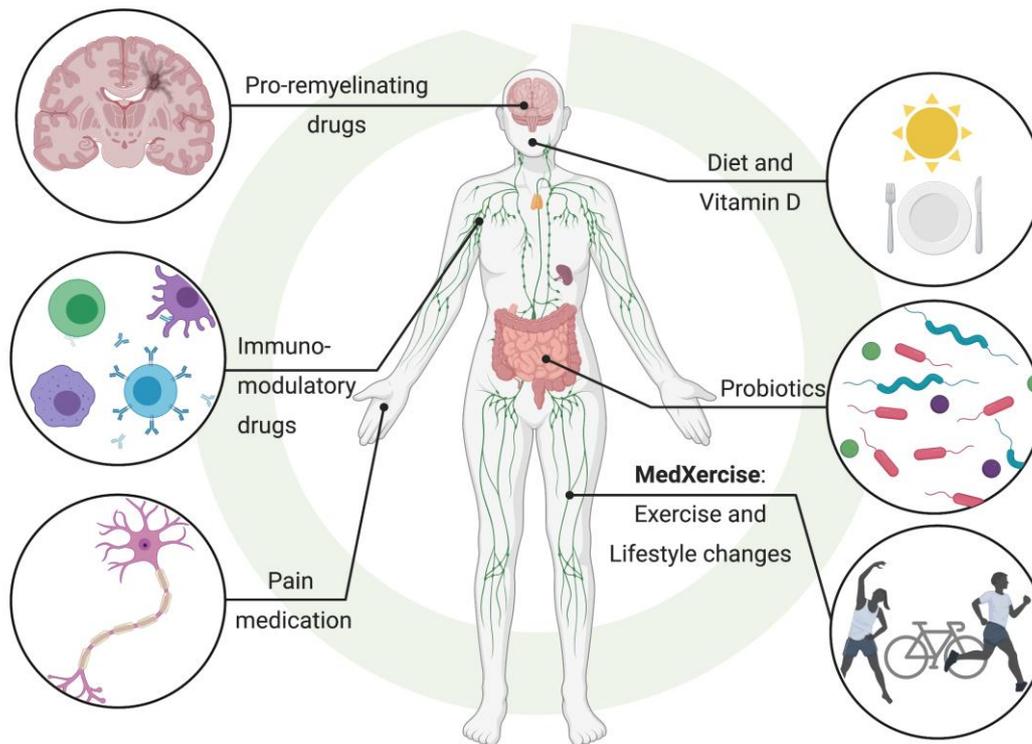


Figure 3: Multimodal approach to managing and treating MS and other chronic inflammatory diseases

The evidence speaks: Multifactorial diseases must be treated by multimodal means

For complex diseases, such as MS, individual drugs have proven to be ineffective at managing and treating MS and have failed to inhibit its progression. A multimodal approach may be more effective at addressing complex chronic diseases including MS and its comorbidities¹²¹. Administering a pro-remyelinating drug may not be as efficacious for long-term usage without reducing recurring chronic inflammation. Importantly, the immunomodulatory roles of the microbiota and other environmental or lifestyle factors should be considered. McMurrin *et al.* showed that inflammatory responses during remyelination depend on the composition of the microbiota, but interventions that modify the microbiota have minimal impact on endogenous CNS remyelination⁹⁹. Here, a multimodal approach of modulating the microbiota along with taking a pro-remyelinating drug is an optimal treatment strategy, as shown by Jensen and colleagues¹²⁰. Additionally, monoclonal antibodies, such as natalizumab approved for MS^{122,123}, have been shown to be efficacious at preventing leukocyte trafficking into the CNS¹²⁴, but whether this therapy can be combined with exercise or other means to modulate the microbiome to reduce inflammatory immune cells warrants investigation. While pharmacological treatment may focus on one aspect of disease, an effective “cure” will need

to address all dysregulated features of MS. It is likely that microbiota composition plays an influential role in MS pathogenesis. Correcting intestinal dysbiosis may also improve cognitive behaviour and depression¹²⁵, which is a common symptom of MS^{121,126}. Therefore, future treatments should consider how disease modifying treatments (DMTs) can be combined with therapies targeted at the microbiota, including lifestyle changes¹²⁷ such as exercise and/or modified diets, to maximize tissue repair and clinical benefit for MS patients (**Figure 3**). This may be of particular importance for MS patients residing in long-term care facilities as nursing home residents microbiomes are associated with decreased SCFA-producing organisms and increased intestinal dysbiosis¹²⁸ which may exacerbate MS pathogenesis. Multicenter studies across the world may help identify microbial strains common to MS patients independent of vastly different lifestyle and diets of patients from different countries. However, it is important to consider the effects of treatment and disease duration on the microbiome of MS patients, as several current/promising DMTs have antimicrobial effects including glatiramer acetate¹²⁹, minocycline¹³⁰, fingolimod¹³¹, teriflunomide¹³² and dimethyl fumarate¹³². It is likely that *the crime of MS* is committed by more than one criminal. Therefore, solving the cold case of MS will require the combination of multiple experts from various disciplines including microbiology, virology, neurology and immunology.

References

1. Filippi M, Bar-Or A, Piehl F, et al. Multiple sclerosis. *Nat Rev Dis Primer*. 2018;4(1):43. doi:10.1038/s41572-018-0041-4
2. Dendrou CA, Fugger L, Friese MA. Immunopathology of multiple sclerosis. *Nat Rev Immunol*. 2015;15(9):545-558. doi:10.1038/nri3871
3. Filyk HA, Osborne LC. The Multibiome: The Intestinal Ecosystem's Influence on Immune Homeostasis, Health, and Disease. *EBioMedicine*. 2016;13:46-54. doi:10.1016/j.ebiom.2016.10.007
4. Cryan JF, O'Riordan KJ, Sandhu K, Peterson V, Dinan TG. The gut microbiome in neurological disorders. *Lancet Neurol*. 2020;19(2):179-194. doi:10.1016/S1474-4422(19)30356-4
5. Okada H, Kuhn C, Feillet H, Bach J-F. The 'hygiene hypothesis' for autoimmune and allergic diseases: an update: The 'hygiene hypothesis': an update. *Clin Exp Immunol*. 2010;160(1):1-9. doi:10.1111/j.1365-2249.2010.04139.x
6. Bach J-F. The hygiene hypothesis in autoimmunity: the role of pathogens and commensals. *Nat Rev Immunol*. 2018;18(2):105-120. doi:10.1038/nri.2017.111
7. Ahlgren C, Lycke J, Odén A, Andersen O. High risk of MS in Iranian immigrants in Gothenburg, Sweden. *Mult Scler J*. 2010;16(9):1079-1082. doi:10.1177/1352458510376777
8. Wallin MT, Page WF, Kurtzke JF. Migration and multiple sclerosis in Alaskan military veterans. *J Neurol*. 2009;256(9):1413-1417. doi:10.1007/s00415-009-5123-5

9. Guimond C, Dymont DA, Ramagopalan SV, et al. Prevalence of MS in Iranian Immigrants to British Columbia, Canada. *J Neurol*. 2010;257(4):667-668. doi:10.1007/s00415-009-5417-7
10. Jangi S, Gandhi R, Cox LM, et al. Alterations of the human gut microbiome in multiple sclerosis. *Nat Commun*. 2016;7(1):12015. doi:10.1038/ncomms12015
11. Chen J, Chia N, Kalari KR, et al. Multiple sclerosis patients have a distinct gut microbiota compared to healthy controls. *Sci Rep*. 2016;6(1):28484. doi:10.1038/srep28484
12. Ventura RE, Iizumi T, Battaglia T, et al. Gut microbiome of treatment-naïve MS patients of different ethnicities early in disease course. *Sci Rep*. 2019;9(1):16396. doi:10.1038/s41598-019-52894-z
13. Johanson DM, Goertz JE, Marin IA, Costello J, Overall CC, Gaultier A. Experimental autoimmune encephalomyelitis is associated with changes of the microbiota composition in the gastrointestinal tract. *Sci Rep*. 2020;10(1):15183. doi:10.1038/s41598-020-72197-y
14. Almutairi MMA, Gong C, Xu YG, Chang Y, Shi H. Factors controlling permeability of the blood–brain barrier. *Cell Mol Life Sci*. 2016;73(1):57-77. doi:10.1007/s00018-015-2050-8
15. Braniste V, Al-Asmakh M, Kowal C, et al. The gut microbiota influences blood-brain barrier permeability in mice. *Sci Transl Med*. 2014;6(263):263ra158-263ra158. doi:10.1126/scitranslmed.3009759
16. Wu Q, Zhang Y, Zhang Y, et al. Potential effects of antibiotic-induced gut microbiome alteration on blood–brain barrier permeability compromise in rhesus monkeys. *Ann N Y Acad Sci*. 2020;1470(1):14-24. doi:10.1111/nyas.14312
17. Parker A, Fonseca S, Carding SR. Gut microbes and metabolites as modulators of blood-brain barrier integrity and brain health. *Gut Microbes*. 2020;11(2):135-157. doi:10.1080/19490976.2019.1638722
18. Haruwaka K, Ikegami A, Tachibana Y, et al. Dual microglia effects on blood brain barrier permeability induced by systemic inflammation. *Nat Commun*. 2019;10(1):5816. doi:10.1038/s41467-019-13812-z
19. Camara-Lemarroy CR, Silva C, Greenfield J, Liu W-Q, Metz LM, Yong VW. Biomarkers of intestinal barrier function in multiple sclerosis are associated with disease activity. *Mult Scler J*. 2020;26(11):1340-1350. doi:10.1177/1352458519863133
20. Fasano A. All disease begins in the (leaky) gut: role of zonulin-mediated gut permeability in the pathogenesis of some chronic inflammatory diseases. *F1000Research*. 2020;9:69. doi:10.12688/f1000research.20510.1
21. Rahman MT, Ghosh C, Hossain M, et al. IFN- γ , IL-17A, or zonulin rapidly increase the permeability of the blood-brain and small intestinal epithelial barriers: Relevance for neuro-inflammatory diseases. *Biochem Biophys Res Commun*. 2018;507(1-4):274-279. doi:10.1016/j.bbrc.2018.11.021

22. Berer K, Gerdes LA, Cekanaviciute E, et al. Gut microbiota from multiple sclerosis patients enables spontaneous autoimmune encephalomyelitis in mice. *Proc Natl Acad Sci.* 2017;114(40):10719-10724. doi:10.1073/pnas.1711233114
23. Cekanaviciute E, Yoo BB, Runia TF, et al. Gut bacteria from multiple sclerosis patients modulate human T cells and exacerbate symptoms in mouse models. *Proc Natl Acad Sci.* 2017;114(40):10713-10718. doi:10.1073/pnas.1711235114
24. Gomez de Agüero M, Ganai-Vonarburg SC, Fuhrer T, et al. The maternal microbiota drives early postnatal innate immune development. *Science.* 2016;351(6279):1296-1302. doi:10.1126/science.aad2571
25. Hu M, Eviston D, Hsu P, et al. Decreased maternal serum acetate and impaired fetal thymic and regulatory T cell development in preeclampsia. *Nat Commun.* 2019;10(1):3031. doi:10.1038/s41467-019-10703-1
26. Munger KL, Äivo J, Hongell K, Soilu-Hänninen M, Surcel H-M, Ascherio A. Vitamin D Status During Pregnancy and Risk of Multiple Sclerosis in Offspring of Women in the Finnish Maternity Cohort. *JAMA Neurol.* 2016;73(5):515. doi:10.1001/jamaneurol.2015.4800
27. Munk Nielsen N, Corn G, Frisch M, et al. Multiple sclerosis among first- and second-generation immigrants in Denmark: a population-based cohort study. *Brain.* 2019;142(6):1587-1597. doi:10.1093/brain/awz088
28. Disanto G, Watson CT, Meier UC, Ebers GC, Giovannoni G, Ramagopalan SV. Month of Birth and Thymic Output. *JAMA Neurol.* 2013;70(4):527. doi:10.1001/jamaneurol.2013.2116
29. Hawkes CH, Giovannoni G, Lechner-Scott J, Levy M, Waubant E. Multiple sclerosis and migration revisited. *Mult Scler Relat Disord.* 2019;34:A1-A2. doi:10.1016/j.msard.2019.08.001
30. Barnett MH, McLeod JG, Hammond SR, Kurtzke JF. Migration and multiple sclerosis in immigrants from United Kingdom and Ireland to Australia: a reassessment. III: risk of multiple sclerosis in UKI immigrants and Australian-born in Hobart, Tasmania. *J Neurol.* 2016;263(4):792-798. doi:10.1007/s00415-016-8059-6
31. Dean G, Elian M. Age at immigration to England of Asian and Caribbean immigrants and the risk of developing multiple sclerosis. *J Neurol Neurosurg Psychiatry.* 1997;63(5):565-568. doi:10.1136/jnnp.63.5.565
32. Berg-Hansen P, Celius EG. Socio-economic factors and immigrant population studies of multiple sclerosis. *Acta Neurol Scand.* 2015;132:37-41. doi:10.1111/ane.12429
33. Fung TC, Olson CA, Hsiao EY. Interactions between the microbiota, immune and nervous systems in health and disease. *Nat Neurosci.* 2017;20(2):145-155. doi:10.1038/nn.4476
34. Tanaka M, Nakayama J. Development of the gut microbiota in infancy and its impact on health in later life. *Allergol Int.* 2017;66(4):515-522. doi:10.1016/j.alit.2017.07.010

35. Laforest-Lapointe I, Arrieta M-C. Patterns of Early-Life Gut Microbial Colonization during Human Immune Development: An Ecological Perspective. *Front Immunol.* 2017;8:788. doi:10.3389/fimmu.2017.00788
36. Arrieta M-C, Stiemsma LT, Dimitriu PA, et al. Early infancy microbial and metabolic alterations affect risk of childhood asthma. *Sci Transl Med.* 2015;7(307):307ra152-307ra152. doi:10.1126/scitranslmed.aab2271
37. Vatanen T, Kostic AD, d’Hennezel E, et al. Variation in Microbiome LPS Immunogenicity Contributes to Autoimmunity in Humans. *Cell.* 2016;165(4):842-853. doi:10.1016/j.cell.2016.04.007
38. Haghikia A, Jörg S, Duscha A, et al. Dietary Fatty Acids Directly Impact Central Nervous System Autoimmunity via the Small Intestine. *Immunity.* 2015;43(4):817-829. doi:10.1016/j.immuni.2015.09.007
39. Dopkins N, Nagarkatti PS, Nagarkatti M. The role of gut microbiome and associated metabolome in the regulation of neuroinflammation in multiple sclerosis and its implications in attenuating chronic inflammation in other inflammatory and autoimmune disorders. *Immunology.* 2018;154(2):178-185. doi:10.1111/imm.12903
40. Ratajczak W, Rył A, Mizerski A, Walczakiewicz K, Sipak O, Laszczyńska M. Immunomodulatory potential of gut microbiome-derived short-chain fatty acids (SCFAs). *Acta Biochim Pol.* Published online March 4, 2019. doi:10.18388/abp.2018_2648
41. Silva YP, Bernardi A, Frozza RL. The Role of Short-Chain Fatty Acids From Gut Microbiota in Gut-Brain Communication. *Front Endocrinol.* 2020;11:25. doi:10.3389/fendo.2020.00025
42. Saresella M, Marventano I, Barone M, et al. Alterations in Circulating Fatty Acid Are Associated With Gut Microbiota Dysbiosis and Inflammation in Multiple Sclerosis. *Front Immunol.* 2020;11:1390. doi:10.3389/fimmu.2020.01390
43. Luu M, Pautz S, Kohl V, et al. The short-chain fatty acid pentanoate suppresses autoimmunity by modulating the metabolic-epigenetic crosstalk in lymphocytes. *Nat Commun.* 2019;10(1):760. doi:10.1038/s41467-019-08711-2
44. Dong Y, Yong VW. When encephalitogenic T cells collaborate with microglia in multiple sclerosis. *Nat Rev Neurol.* 2019;15(12):704-717. doi:10.1038/s41582-019-0253-6
45. Barroso A, Mahler JV, Fonseca-Castro PH, Quintana FJ. The aryl hydrocarbon receptor and the gut–brain axis. *Cell Mol Immunol.* Published online January 6, 2021. doi:10.1038/s41423-020-00585-5
46. Gaetani L, Boscaro F, Pieraccini G, et al. Host and Microbial Tryptophan Metabolic Profiling in Multiple Sclerosis. *Front Immunol.* 2020;11:157. doi:10.3389/fimmu.2020.00157

47. Mangalam A, Poisson L, Nemutlu E, et al. Profile of Circulatory Metabolites in an Animal Model of Multiple Sclerosis using Global Metabolomics. *J Clin Cell Immunol.* 2013;04(03). doi:10.4172/2155-9899.1000150
48. Rothhammer V, Borucki DM, Tjon EC, et al. Microglial control of astrocytes in response to microbial metabolites. *Nature.* 2018;557(7707):724-728. doi:10.1038/s41586-018-0119-x
49. Lim CK, Bilgin A, Lovejoy DB, et al. Kynurenine pathway metabolomics predicts and provides mechanistic insight into multiple sclerosis progression. *Sci Rep.* 2017;7(1):41473. doi:10.1038/srep41473
50. Erny D, Hrabě de Angelis AL, Jaitin D, et al. Host microbiota constantly control maturation and function of microglia in the CNS. *Nat Neurosci.* 2015;18(7):965-977. doi:10.1038/nn.4030
51. Duscha A, Gisevius B, Hirschberg S, et al. Propionic Acid Shapes the Multiple Sclerosis Disease Course by an Immunomodulatory Mechanism. *Cell.* 2020;180(6):1067-1080.e16. doi:10.1016/j.cell.2020.02.035
52. Cosorich I, Dalla-Costa G, Sorini C, et al. High frequency of intestinal T_H 17 cells correlates with microbiota alterations and disease activity in multiple sclerosis. *Sci Adv.* 2017;3(7):e1700492. doi:10.1126/sciadv.1700492
53. Choileáin SN, Kleinewietfeld M, Raddassi K, Hafler DA, Ruff WE, Longbrake EE. CXCR3+ T cells in multiple sclerosis correlate with reduced diversity of the gut microbiome. *J Transl Autoimmun.* 2020;3:100032. doi:10.1016/j.jtauto.2019.100032
54. Duc D, Vigne S, Bernier-Latmani J, et al. Disrupting Myelin-Specific Th17 Cell Gut Homing Confers Protection in an Adoptive Transfer Experimental Autoimmune Encephalomyelitis. *Cell Rep.* 2019;29(2):378-390.e4. doi:10.1016/j.celrep.2019.09.002
55. Berer K, Boziki M, Krishnamoorthy G. Selective Accumulation of Pro-Inflammatory T Cells in the Intestine Contributes to the Resistance to Autoimmune Demyelinating Disease. Linker RA, ed. *PLoS ONE.* 2014;9(2):e87876. doi:10.1371/journal.pone.0087876
56. Hauptelshofer S, Leichsenring T, Berg S, et al. Smad7 in intestinal CD4+ T cells determines autoimmunity in a spontaneous model of multiple sclerosis. *Proc Natl Acad Sci.* 2019;116(51):25860-25869. doi:10.1073/pnas.1905955116
57. Gandy KAO, Zhang J, Nagarkatti P, Nagarkatti M. The role of gut microbiota in shaping the relapse-remitting and chronic-progressive forms of multiple sclerosis in mouse models. *Sci Rep.* 2019;9(1):6923. doi:10.1038/s41598-019-43356-7
58. Held K, Bhonsle-Deeng L, Siewert K, et al. $\alpha\beta$ T-cell receptors from multiple sclerosis brain lesions show MAIT cell-related features. *Neurol - Neuroimmunol Neuroinflammation.* 2015;2(4):e107. doi:10.1212/NXI.000000000000107
59. Carnero Contentti E, Farez MF, Correale J. Mucosal-Associated Invariant T Cell Features and TCR Repertoire Characteristics During the Course of Multiple Sclerosis. *Front Immunol.* 2019;10:2690. doi:10.3389/fimmu.2019.02690

60. Reboldi A, Cyster JG. Peyer's patches: organizing B-cell responses at the intestinal frontier. *Immunol Rev.* 2016;271(1):230-245. doi:10.1111/imr.12400
61. Rojas OL, Pröbstel A-K, Porfilio EA, et al. Recirculating Intestinal IgA-Producing Cells Regulate Neuroinflammation via IL-10. *Cell.* 2019;176(3):610-624.e18. doi:10.1016/j.cell.2018.11.035
62. Planas R, Santos R, Tomas-Ojer P, et al. GDP-l-fucose synthase is a CD4⁺ T cell-specific autoantigen in DRB3*02:02 patients with multiple sclerosis. *Sci Transl Med.* 2018;10(462):eaat4301. doi:10.1126/scitranslmed.aat4301
63. Mecha M, Carrillo-Salinas FJ, Mestre L, Feliú A, Guaza C. Viral models of multiple sclerosis: Neurodegeneration and demyelination in mice infected with Theiler's virus. *Prog Neurobiol.* 2013;101-102:46-64. doi:10.1016/j.pneurobio.2012.11.003
64. Küry P, Nath A, Créange A, et al. Human Endogenous Retroviruses in Neurological Diseases. *Trends Mol Med.* 2018;24(4):379-394. doi:10.1016/j.molmed.2018.02.007
65. Perron H, Garson JA, Bedin F, et al. Molecular identification of a novel retrovirus repeatedly isolated from patients with multiple sclerosis. *Proc Natl Acad Sci.* 1997;94(14):7583-7588. doi:10.1073/pnas.94.14.7583
66. Morandi E, Tanasescu R, Tarlinton RE, et al. The association between human endogenous retroviruses and multiple sclerosis: A systematic review and meta-analysis. Ruprecht K, ed. *PLOS ONE.* 2017;12(2):e0172415. doi:10.1371/journal.pone.0172415
67. Handel AE, Williamson AJ, Disanto G, Handunnetthi L, Giovannoni G, Ramagopalan SV. An Updated Meta-Analysis of Risk of Multiple Sclerosis following Infectious Mononucleosis. Jacobson S, ed. *PLoS ONE.* 2010;5(9):e12496. doi:10.1371/journal.pone.0012496
68. Afrasiabi A, Parnell GP, Swaminathan S, Stewart GJ, Booth DR. The interaction of Multiple Sclerosis risk loci with Epstein-Barr virus phenotypes implicates the virus in pathogenesis. *Sci Rep.* 2020;10(1):193. doi:10.1038/s41598-019-55850-z
69. Pakpoor J, Disanto G, Gerber JE, et al. The risk of developing multiple sclerosis in individuals seronegative for Epstein-Barr virus: a meta-analysis. *Mult Scler J.* 2013;19(2):162-166. doi:10.1177/1352458512449682
70. Abrahamyan S, Eberspächer B, Hoshi M-M, et al. Complete Epstein-Barr virus seropositivity in a large cohort of patients with early multiple sclerosis. *J Neurol Neurosurg Psychiatry.* 2020;91(7):681-686. doi:10.1136/jnnp-2020-322941
71. Libbey JE, McCoy LL, Fujinami RS. Molecular Mimicry in Multiple Sclerosis. In: *International Review of Neurobiology.* Vol 79. Elsevier; 2007:127-147. doi:10.1016/S0074-7742(07)79006-2
72. Croxford JL, Olson JK, Anger HA, Miller SD. Initiation and Exacerbation of Autoimmune Demyelination of the Central Nervous System via Virus-Induced Molecular Mimicry: Implications for the Pathogenesis of Multiple Sclerosis. *J Virol.* 2005;79(13):8581-8590. doi:10.1128/JVI.79.13.8581-8590.2005

73. Jog NR, McClain MT, Heinlen LD, et al. Epstein Barr virus nuclear antigen 1 (EBNA-1) peptides recognized by adult multiple sclerosis patient sera induce neurologic symptoms in a murine model. *J Autoimmun.* 2020;106:102332. doi:10.1016/j.jaut.2019.102332
74. Tengvall K, Huang J, Hellström C, et al. Molecular mimicry between Anoctamin 2 and Epstein-Barr virus nuclear antigen 1 associates with multiple sclerosis risk. *Proc Natl Acad Sci.* 2019;116(34):16955-16960. doi:10.1073/pnas.1902623116
75. Tao C, Simpson S, Taylor BV, van der Mei I. Association between human herpesvirus & human endogenous retrovirus and MS onset & progression. *J Neurol Sci.* 2017;372:239-249. doi:10.1016/j.jns.2016.11.060
76. Opsahl ML, Kennedy PGE. Early and late HHV-6 gene transcripts in multiple sclerosis lesions and normal appearing white matter. *Brain.* 2005;128(3):516-527. doi:10.1093/brain/awh390
77. Engdahl E, Gustafsson R, Huang J, et al. Increased Serological Response Against Human Herpesvirus 6A Is Associated With Risk for Multiple Sclerosis. *Front Immunol.* 2019;10:2715. doi:10.3389/fimmu.2019.02715
78. Ahlqvist J, Fotheringham J, Akhyani N, Yao K, Fogdell-Hahn A, Jacobson S. Differential tropism of human herpesvirus 6 (HHV-6) variants and induction of latency by HHV-6A in oligodendrocytes. *J Neurovirol.* 2005;11(4):384-394. doi:10.1080/13550280591002379
79. Kremer D, Gruchot J, Weyers V, et al. pHERV-W envelope protein fuels microglial cell-dependent damage of myelinated axons in multiple sclerosis. *Proc Natl Acad Sci.* 2019;116(30):15216-15225. doi:10.1073/pnas.1901283116
80. GeNeuro SA. *An International, Double-Blind, Randomised, Placebo-Controlled Phase I/IIb Trial to Assess the Efficacy, Safety, and Pharmacokinetics of GNBAC1 in Patients With Relapsing Remitting Multiple Sclerosis.* clinicaltrials.gov; 2020. Accessed January 7, 2021. <https://clinicaltrials.gov/ct2/show/NCT02782858>
81. Carter CJ. Epstein–Barr and other viral mimicry of autoantigens, myelin and vitamin D-related proteins and of EIF2B, the cause of vanishing white matter disease: massive mimicry of multiple sclerosis relevant proteins by the *Synechococcus* phage. *Immunopharmacol Immunotoxicol.* 2012;34(1):21-35. doi:10.3109/08923973.2011.572262
82. Tetz G, Tetz V. Bacteriophage infections of microbiota can lead to leaky gut in an experimental rodent model. *Gut Pathog.* 2016;8(1):33. doi:10.1186/s13099-016-0109-1
83. Tetz GV, Ruggles KV, Zhou H, Heguy A, Tsirigos A, Tetz V. Bacteriophages as potential new mammalian pathogens. *Sci Rep.* 2017;7(1):7043. doi:10.1038/s41598-017-07278-6
84. Zhao G, Vatanen T, Droit L, et al. Intestinal virome changes precede autoimmunity in type I diabetes-susceptible children. *Proc Natl Acad Sci.* 2017;114(30):E6166-E6175. doi:10.1073/pnas.1706359114

85. Kumata R, Ito J, Takahashi K, Suzuki T, Sato K. A tissue level atlas of the healthy human virome. *BMC Biol.* 2020;18(1):55. doi:10.1186/s12915-020-00785-5
86. Guo C-J, Chang F-Y, Wyche TP, et al. Discovery of Reactive Microbiota-Derived Metabolites that Inhibit Host Proteases. *Cell.* 2017;168(3):517-526.e18. doi:10.1016/j.cell.2016.12.021
87. Vincents B, Vindebro R, Abrahamson M, von Pawel-Rammingen U. The Human Protease Inhibitor Cystatin C Is an Activating Cofactor for the Streptococcal Cysteine Protease IdeS. *Chem Biol.* 2008;15(9):960-968. doi:10.1016/j.chembiol.2008.07.021
88. Hoghooghi V, Palmer AL, Frederick A, et al. Cystatin C Plays a Sex-Dependent Detrimental Role in Experimental Autoimmune Encephalomyelitis. *Cell Rep.* 2020;33(1):108236. doi:10.1016/j.celrep.2020.108236
89. Gonzalez CG, Tankou SK, Cox LM, Casavant EP, Weiner HL, Elias JE. Latent-period stool proteomic assay of multiple sclerosis model indicates protective capacity of host-expressed protease inhibitors. *Sci Rep.* 2019;9(1):12460. doi:10.1038/s41598-019-48495-5
90. Miyake S, Kim S, Suda W, et al. Dysbiosis in the Gut Microbiota of Patients with Multiple Sclerosis, with a Striking Depletion of Species Belonging to Clostridia XIVA and IV Clusters. Wilson BA, ed. *PLOS ONE.* 2015;10(9):e0137429. doi:10.1371/journal.pone.0137429
91. Huynh JL, Garg P, Thin TH, et al. Epigenome-wide differences in pathology-free regions of multiple sclerosis-affected brains. *Nat Neurosci.* 2014;17(1):121-130. doi:10.1038/nn.3588
92. Allan ERO, Campden RI, Ewanchuk BW, et al. A role for cathepsin Z in neuroinflammation provides mechanistic support for an epigenetic risk factor in multiple sclerosis. *J Neuroinflammation.* 2017;14(1):103. doi:10.1186/s12974-017-0874-x
93. Arpaia N, Campbell C, Fan X, et al. Metabolites produced by commensal bacteria promote peripheral regulatory T-cell generation. *Nature.* 2013;504(7480):451-455. doi:10.1038/nature12726
94. Atarashi K, Tanoue T, Shima T, et al. Induction of Colonic Regulatory T Cells by Indigenous Clostridium Species. *Science.* 2011;331(6015):337-341. doi:10.1126/science.1198469
95. Mizuno M, Noto D, Kaga N, Chiba A, Miyake S. The dual role of short fatty acid chains in the pathogenesis of autoimmune disease models. Ashour HM, ed. *PLOS ONE.* 2017;12(2):e0173032. doi:10.1371/journal.pone.0173032
96. Gödel C, Kunkel B, Kashani A, Lassmann H, Arumugam M, Krishnamoorthy G. Perturbation of gut microbiota decreases susceptibility but does not modulate ongoing autoimmune neurological disease. *J Neuroinflammation.* 2020;17(1):79. doi:10.1186/s12974-020-01766-9

97. Lee YK, Menezes JS, Umesaki Y, Mazmanian SK. Proinflammatory T-cell responses to gut microbiota promote experimental autoimmune encephalomyelitis. *Proc Natl Acad Sci*. 2011;108(Supplement_1):4615-4622. doi:10.1073/pnas.1000082107
98. Belkaid Y, Hand TW. Role of the Microbiota in Immunity and Inflammation. *Cell*. 2014;157(1):121-141. doi:10.1016/j.cell.2014.03.011
99. McMurrin CE, Guzman de la Fuente A, Penalva R, et al. The microbiota regulates murine inflammatory responses to toxin-induced CNS demyelination but has minimal impact on remyelination. *Proc Natl Acad Sci*. 2019;116(50):25311-25321. doi:10.1073/pnas.1905787116
100. Mager LF, Burkhard R, Pett N, et al. Microbiome-derived inosine modulates response to checkpoint inhibitor immunotherapy. *Science*. 2020;369(6510):1481-1489. doi:10.1126/science.abc3421
101. Shahi SK, Freedman SN, Murra AC, et al. *Prevotella histicola*, A Human Gut Commensal, Is as Potent as COPAXONE® in an Animal Model of Multiple Sclerosis. *Front Immunol*. 2019;10:462. doi:10.3389/fimmu.2019.00462
102. Odoardi F, Sie C, Streyl K, et al. T cells become licensed in the lung to enter the central nervous system. *Nature*. 2012;488(7413):675-679. doi:10.1038/nature11337
103. Hassani Y, Brégeon F, Aboudharam G, Drancourt M, Grine G. Detection of *Methanobrevibacter smithii* and *Methanobrevibacter oralis* in Lower Respiratory Tract Microbiota. *Microorganisms*. 2020;8(12):1866. doi:10.3390/microorganisms8121866
104. Koskinen K, Pausan MR, Perras AK, et al. First Insights into the Diverse Human Archaeome: Specific Detection of Archaea in the Gastrointestinal Tract, Lung, and Nose and on Skin. Schleper CM, Jansson JK, eds. *mBio*. 2017;8(6):mBio.00824-17, e00824-17. doi:10.1128/mBio.00824-17
105. Dixit A, Tanaka A, Greer JM, Donnelly S. Novel Therapeutics for Multiple Sclerosis Designed by Parasitic Worms. *Int J Mol Sci*. 2017;18(10):2141. doi:10.3390/ijms18102141
106. Tanasescu R, Tench CR, Constantinescu CS, et al. Hookworm Treatment for Relapsing Multiple Sclerosis: A Randomized Double-Blinded Placebo-Controlled Trial. *JAMA Neurol*. 2020;77(9):1089. doi:10.1001/jamaneurol.2020.1118
107. Lund ME, Greer J, Dixit A, et al. A parasite-derived 68-mer peptide ameliorates autoimmune disease in murine models of Type 1 diabetes and multiple sclerosis. *Sci Rep*. 2016;6(1):37789. doi:10.1038/srep37789
108. Marzano V, Mancinelli L, Bracaglia G, et al. “Omic” investigations of protozoa and worms for a deeper understanding of the human gut “parasitome.” Blair D, ed. *PLoS Negl Trop Dis*. 2017;11(11):e0005916. doi:10.1371/journal.pntd.0005916
109. Lozinski BM, Yong VW. Exercise and the brain in multiple sclerosis. *Mult Scler J*. Published online October 30, 2020:135245852096909. doi:10.1177/1352458520969099

110. Nieman DC, Wentz LM. The compelling link between physical activity and the body's defense system. *J Sport Health Sci.* 2019;8(3):201-217. doi:10.1016/j.jshs.2018.09.009
111. Bermon S, Petriz B, Kajèniènè A, Prestes J, Castell L, Franco OL. The microbiota: an exercise immunology perspective. *Exerc Immunol Rev.* 2015;21:70-79.
112. Estaki M, Pither J, Baumeister P, et al. Cardiorespiratory fitness as a predictor of intestinal microbial diversity and distinct metagenomic functions. *Microbiome.* 2016;4(1):42. doi:10.1186/s40168-016-0189-7
113. Campbell SC, Wisniewski PJ, Noji M, et al. The Effect of Diet and Exercise on Intestinal Integrity and Microbial Diversity in Mice. Blachier F, ed. *PLOS ONE.* 2016;11(3):e0150502. doi:10.1371/journal.pone.0150502
114. Barton W, Penney NC, Cronin O, et al. The microbiome of professional athletes differs from that of more sedentary subjects in composition and particularly at the functional metabolic level. *Gut.* Published online March 30, 2017:gutjnl-2016-313627. doi:10.1136/gutjnl-2016-313627
115. Codella R, Luzi L, Terruzzi I. Exercise has the guts: How physical activity may positively modulate gut microbiota in chronic and immune-based diseases. *Dig Liver Dis.* 2018;50(4):331-341. doi:10.1016/j.dld.2017.11.016
116. Quiroga R, Nistal E, Estébanez B, et al. Exercise training modulates the gut microbiota profile and impairs inflammatory signaling pathways in obese children. *Exp Mol Med.* 2020;52(7):1048-1061. doi:10.1038/s12276-020-0459-0
117. Mitchell CM, Davy BM, Hulver MW, Neilson AP, Bennett BJ, Davy KP. Does Exercise Alter Gut Microbial Composition? A Systematic Review: *Med Sci Sports Exerc.* 2019;51(1):160-167. doi:10.1249/MSS.0000000000001760
118. Liu Y, Wang Y, Ni Y, et al. Gut Microbiome Fermentation Determines the Efficacy of Exercise for Diabetes Prevention. *Cell Metab.* 2020;31(1):77-91.e5. doi:10.1016/j.cmet.2019.11.001
119. Bressa C, Bailén-Andrino M, Pérez-Santiago J, et al. Differences in gut microbiota profile between women with active lifestyle and sedentary women. Dasgupta S, ed. *PLOS ONE.* 2017;12(2):e0171352. doi:10.1371/journal.pone.0171352
120. Jensen SK, Michaels NJ, Ilyntskyy S, Keough MB, Kovalchuk O, Yong VW. Multimodal Enhancement of Remyelination by Exercise with a Pivotal Role for Oligodendroglial PGC1 α . *Cell Rep.* 2018;24(12):3167-3179. doi:10.1016/j.celrep.2018.08.060
121. Magyari M, Sorensen PS. Comorbidity in Multiple Sclerosis. *Front Neurol.* 2020;11:851. doi:10.3389/fneur.2020.00851
122. Perumal J, Fox RJ, Balabanov R, et al. Outcomes of natalizumab treatment within 3 years of relapsing-remitting multiple sclerosis diagnosis: a prespecified 2-year interim analysis of STRIVE. *BMC Neurol.* 2019;19(1):116. doi:10.1186/s12883-019-1337-z

123. Butzkueven H, Kappos L, Wiendl H, et al. Long-term safety and effectiveness of natalizumab treatment in clinical practice: 10 years of real-world data from the Tysabri Observational Program (TOP). *J Neurol Neurosurg Psychiatry*. 2020;91(6):660-668. doi:10.1136/jnnp-2019-322326
124. Brandstadter R, Katz Sand I. The use of natalizumab for multiple sclerosis. *Neuropsychiatr Dis Treat*. 2017;Volume 13:1691-1702. doi:10.2147/NDT.S114636
125. Wang H, Lee I-S, Braun C, Enck P. Effect of Probiotics on Central Nervous System Functions in Animals and Humans: A Systematic Review. *J Neurogastroenterol Motil*. 2016;22(4):589-605. doi:10.5056/jnm16018
126. Feinstein A, Magalhaes S, Richard J-F, Audet B, Moore C. The link between multiple sclerosis and depression. *Nat Rev Neurol*. 2014;10(9):507-517. doi:10.1038/nrneurol.2014.139
127. Jakimovski D, Guan Y, Ramanathan M, Weinstock-Guttman B, Zivadinov R. Lifestyle-based modifiable risk factors in multiple sclerosis: review of experimental and clinical findings. *Neurodegener Dis Manag*. 2019;9(3):149-172. doi:10.2217/nmt-2018-0046
128. Haran JP, Bucci V, Dutta P, Ward D, McCormick B. The nursing home elder microbiome stability and associations with age, frailty, nutrition and physical location. *J Med Microbiol*. 2018;67(1):40-51. doi:10.1099/jmm.0.000640
129. Murphy R, Harrison J, Schelenz S, Davies J. M5 The multiple sclerosis drug, glatiramer acetate, acts as a resistance breaker with antibiotics from different classes against cystic fibrosis strains of pseudomonas aeruginosa. In: *The Epidemiology and Impact of Difficult Infections*. BMJ Publishing Group Ltd and British Thoracic Society; 2019:A236.3-A237. doi:10.1136/thorax-2019-BTSabstracts2019.413
130. Metz LM, Li DKB, Traboulsee AL, et al. Trial of Minocycline in a Clinically Isolated Syndrome of Multiple Sclerosis. *N Engl J Med*. 2017;376(22):2122-2133. doi:10.1056/NEJMoa1608889
131. Jia L, Zhang M, Liu H, Sun J, Pan L. Early-life fingolimod treatment improves intestinal homeostasis and pancreatic immune tolerance in non-obese diabetic mice. *Acta Pharmacol Sin*. Published online January 20, 2021. doi:10.1038/s41401-020-00590-4
132. Rumah KR, Vartanian TK, Fischetti VA. Oral Multiple Sclerosis Drugs Inhibit the In vitro Growth of Epsilon Toxin Producing Gut Bacterium, *Clostridium perfringens*. *Front Cell Infect Microbiol*. 2017;7. doi:10.3389/fcimb.2017.00011