Article

Effects of durum wheat cultivars with different degrees of FHB susceptibility grown under different meteorological conditions on the contamination of regulated, modified and emerging mycotoxins.

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Abstract: The enhancement of Fusarium head blight (FHB) resistance is one of the best options to reduce mycotoxin contamination in wheat. This study has aimed to verify that the genotypes with high tolerance to deoxynivalenol could guarantee an overall minimization of the sanitary risk, by evaluating the contamination of regulated, modified and emerging mycotoxins on durum wheat *cvs* with different degrees of FHB susceptibility, grown under different meteorological conditions, in 8 growing seasons in North-West Italy. The years which were characterized by frequent and heavy rainfall in spring, were also those with the highest contamination of deoxynivalenol, zearalenone, moniliformin and enniatins. The most FHB resistant genotypes resulted in the lowest contamination of all the mycotoxins but showed the highest deoxynivalenol-3-glucoside/deoxynivalenol ratio and moniliformin/deoxynivalenol ratio. An inverse relationship between the amount of deoxynivalenol and the deoxynivalenol-3-glucoside/deoxynivalenol ratio was recorded for all the *cvs* and all the years. Conversely, the enniatins/deoxynivalenol ratio had a less intense relationship with *cv* tolerance to FHB. In conclusion, even though the more tolerant *cvs*, showed higher relative relationships between modified/emerging mycotoxins and native/target mycotoxins than the susceptible ones, they showed lower absolute levels of contamination of both emerging and modified mycotoxins.

Keywords: 3-acetyldeoxynivalenol; deoxynivalenol; deoxynivalenol-3-glucoside; enniatins; moniliformin; zearalenone

1. Introduction

Crop productions will be affected by global warming at various rates in different parts of the world, particularly in the most critical and vulnerable geographic areas, like the Mediterranean Basin [1]. Certain regions in the Mediterranean Basin and North America (i.e. Canada) produce about 60% of the world's durum wheat, which is mainly used for human consumption as pasta and other food products, such as bulgur, couscous and some types of bread. Despite its relatively high adaptability to marginal and drought affected environments, durum wheat could be negatively influenced by the impacts of climate change and is highly sensitive to climatic and environmental variations [2]. Furthermore, as a result of global warming (i.e. less severe winter temperatures and more severe dry periods during summer, which could limit the cultivation of summer crops), the geographic distribution of winter wheat might change, especially toward northern latitudes, and it might adapt better to these growing areas [3]. Moreover, in some countries, such as Italy, the pasta industry is looking for national durum wheat productions with high traceability and quality levels, as far as a high grain protein content is concerned. These qualitative targets can be achieved through the cultivation of winter durum wheat in more temperate and fertile growing areas, such as the Po Valley in Northern Italy. However,

durum wheat in temperate areas is characterized by a generally higher susceptibility to fungal diseases than the common one, thus limiting its potential cultivation in these areas [4]. In addition to the direct effects on crop species [5], climate change may act indirectly on the security and quality of food by altering interactions with pests [6,7], as well as impacting the disease triangle involving host, pathogen and environment [8]. Therefore, wheat producers need to face a variety of factors, including abiotic and biotic stresses, which could influence both yields and quality. One of the most serious biotic threats is related to the fact that Fusarium head blight (FHB) is able to affect both the yield potential and the sanitary risk of wheat to a great extent [9]. Moreover, among the most important hazards likely to be affected by climate change, and which is linked to FHB, is contamination by mycotoxins [10]. These compounds are secondary metabolites, which are harmful to both humans and animals, that are produced by filamentous fungi. Deoxynivalenol (DON), a type-B trichothecene produced mainly by F. graminearum sensu stricto and F. culmorum, is the most prevalent toxin in small cereal crops throughout the world and in Europe [11]. Regulatory limits have been set by the European Commission (EC) to protect humans from exposure to this mycotoxin through cereal consumption (EC Regulation No. 1881/2006, with a limit of 1750 μg kg⁻¹ in unprocessed durum wheat) [12]. However, recent quantitative estimations have shown that increased DON contamination can be expected in cereals in certain regions in Europe as a result of future climate change [13,14]. Although the contamination of wheat grains by DON depends mainly on the meteorological conditions, particularly at flowering [15], an important role is also played by agronomic factors [16]. At present, the most effective approach adopted to minimize the occurrence of DON in wheat is the use of preventive agronomic practices to reduce the pathogen infection and development, and the accumulation of mycotoxins in grains. Thus, an increased tolerance to Fusarium infection and mycotoxin contamination should remain a priority for the genetic improvement of wheat cvs. Although breeding programs are hindered by the fact that the resistance to FHB is under polygenic inheritance, the efforts to genetically improve of durum wheat have recently been directed toward releasing cvs on the market that are more tolerant to FHB [17] and thus which have been designed for cultivation in cropping systems in temperate growing areas. Furthermore, the selection of genotypes that are less prone to FHB is generally made by only focusing on DON, as it is the target mycotoxin currently considered by supply chains and in the regulatory limits. To the best of our knowledge, few research efforts have been made to investigate the influence of durum wheat genotypes, and their interaction with environmental conditions, on the occurrence and the relative amount of emerging and modified mycotoxins in grains.

Emerging mycotoxins are commonly defined as "mycotoxins which are neither routinely determined nor legislatively regulated; however, the evidence of their incidence is rapidly increasing" [11]. Aflatoxin precursors, ergot alkaloids, enniatins (ENNs), beauvericin (BEA) and moniliformin (MON) are those that are more commonly mentioned in this group [18]. EFSA, the European Authority for the Safety of Food, has recently been paying more attention to this group of mycotoxins, and risk assessment studies are still in progress. The term "modified mycotoxins" was introduced by Rychlik et al. in 2014 [19] to refer to any mycotoxin whose structure has been changed during a chemical/biochemical reaction [20]. Plants play a key role in biologically modifying mycotoxins as a primary plant mechanism against fungal infection and mycotoxin accumulation, producing the so-called "masked mycotoxins". As far as DON is concerned, the conjugation of DON with glucose (DON-3-glucoside, DON-3-G) is the wheat mechanism (phase II metabolism) that is involved the most in the resistance to DON accumulation and it is closely linked to the wheat genotypes [21].

Nowadays, both emerging and modified mycotoxins are receiving increasing attention by the scientific community, and by governments and regulators, due to the presence of some of them in high concentrations in food and feeds, and because of their toxic effect [22]. The drafting of legislation that will set limits for some of these mycotoxins is expected

in the near future. Since the occurrence of masked mycotoxins is a strategy that is implemented by a plant to limit the accumulation of their free native forms, it is advisable to verify the effect of cv selection considering the total risk associated with these toxic compounds. Moreover, since emerging mycotoxins, such as ENNs and MON, could be produced by other Fusarium species than those primary responsible for DON occurrence, more detailed knowledge on the environmental and agronomic conditions that promote their occurrence is essential, in order to set up field programs that would be able to minimize the overall sanitary risk.

Therefore, the aim of the present research has been to evaluate the effects of durum wheat *cvs* with different degrees of FHB susceptibility and the meteorological conditions that are able to lead to different degrees of disease pressure on the contamination of regulated, modified and emerging mycotoxins in 8 growing seasons in North-West Italy. The practical objective, in a food supply chain framework, is to verify that the genotypes selected because of their high tolerance to DON are overall able to guarantee a minimization of the sanitary risk, according to a holistic approach that also considers the modified DON forms and the emerging mycotoxins.

2. Materials and Methods

2.1. Design of the field experiment and samples

The effects of the choice of a durum wheat cultivar (cv) and the meteorological conditions on the contamination of regulated, modified and emerging mycotoxins was studied in North-West Italy over a period of 8 years at Cigliano (45° 18′ N, 8° 01′ E; altitude 237 m), on plants grown in a sandy-loam soil (Typic Hapludalfs). This growing area is characterized by a Cfa climate, that is, a humid subtropical climate according to the Köppen climate classification [23] and by a probable high FHB pressure, due to the environmental and agronomic conditions (frequent rotation with maize). The daily temperatures and precipitation were measured at a meteorological station near the experimental area. Two durum wheat cvs with different degrees of susceptibility to DON contamination, that is, Saragolla (classified as susceptible) and SY Cysco (classified as moderately tolerant), were compared each year under naturally-infected field conditions. Secolo, Odisseo and Fuego cvs, which are characterized by an intermediate susceptibility to DON contamination, were also compared, albeit only over four years (2016-2019). All the cvs were provided by Syngenta Italia Spa Milano, Italy.

The agronomic growing technique commonly adopted in the area was applied. Briefly, the previous crop was maize, the field was ploughed each year, incorporating the debris into the soil, and this was followed by disk harrowing to prepare a suitable seedbed. Planting was conducted in 12 cm wide rows at a seeding rate of 450 seeds m⁻² in October or November. A total of 170 kg N ha-1 was applied, 130 at wheat tillering [growth stage (GS) 23] [24] as ammonium sulfate-nitrate with nitrification inhibitors and 40 kg N ha-1 at heading (GS 51) as an ammonium nitrate fertilizer. A strobilurin fungicide (azoxystrobin active ingredient, applied at 0.25 kg ha-1, produced by Amistar®, Syngenta Italia Spa, formulation: suspension concentrate) was applied at the booting stage (GS 45) to control foliar diseases. No fungicide was applied at flowering (GS61-65) to control FHB infection. The sowing and harvest dates are reported in Table 1, together with the dates of the main growth stages, for each growing season. Treatments were assigned to an experimental unit using a completely randomized block design with three replicates. The plots measured 7 x 1.5 m. The grain yields were obtained by harvesting the whole plot using a Walter Wintersteiger cereal plot combine-harvester. A subsample was taken from each plot to determine the grain moisture and the test weight (TW). The TW was determined using a Dickey-John GAC2000 grain analysis meter, according to the supplied program. The grain yield results were adjusted to a 13% moisture content. The harvested grains were mixed thoroughly, and 4 kg grain samples were taken from each plot to analyze the mycotoxin content.

| Year | | N fertili | zation | Anthesis | Harvest |
|------|------------------|--------------------|--------------------|--------------------|--------------|
| | Sowing date | GS 23 ¹ | GS 51 ¹ | GS 65 ¹ | date |
| 2013 | 6 November 2012 | 11 March 2013 | 9 May 2013 | 17 May 2013 | 10 July 2013 |
| 2014 | 27 October 2013 | 7 March 2014 | 23 April 2014 | 5 May 2014 | 10 July 2014 |
| 2015 | 8 November 2014 | 12 March 2015 | 4 May 2015 | 10 May 2015 | 29 June 2015 |
| 2016 | 6 November 2015 | 23 February 2016 | 3 May 2016 | 10 May 2016 | 1 July 2016 |
| 2017 | 4 November 2016 | 7 March 2017 | 28 April 2017 | 12 May 2017 | 27 June 2017 |
| 2018 | 31 October 2017 | 23 February 2018 | 7 May 2018 | 11 May 2018 | 3 July 2018 |
| 2019 | 16 November 2018 | 6 March 2019 | 30 April 2019 | 16 May 2019 | 1 July 2019 |
| 2020 | 6 November 2019 | 5 March 2020 | 23 April 2020 | 12 May 2020 | 29 June 2020 |

Table 1. Main trial information for the field experiments conducted in Cigliano (NW Italy) in the 2012 - 2020 period.

2.2. FHB symptoms

FHB incidence and severity were recorded for each plot, by carrying out visual evaluations of the disease at the soft dough stage (GS 85). FHB head blight incidence was calculated as the percentage of ears with symptoms when 200 ears per plot were analyzed.

FHB severity was calculated as the percentage of kernels per ear with symptoms. A scale of 1 to 7 was used in which each numerical value corresponded to a percentage interval of surfaces exhibiting visible symptoms of the disease, according to the following schedule: 1 = 0.5%, 2 = 6.15%, 3 = 16.30%; 4 = 31.50%, 5 = 51.75%, 6 = 76.90%, 7 = 91.100% [25]. The FHB severity scores were converted into percentages of the ear exhibiting symptoms, replacing each score with the mid-point of the interval.

2.3. Multi-mycotoxin LC-MS/MS analysis

2.3.1. Extraction and sample preparation

The extraction and sample preparation were performed according to the dilute-and-shoot method reported by Scarpino et al. in 2019 [26]. Briefly, 5 g of wheat flour was extracted, by means of mechanical shaking, with 20 mL of CH₃CN/H₂O/CH₃COOH (79/20/1, v/v/v). The filtered extract was diluted with the same volume of CH₃CN/H₂O/CH₃COOH 20/79/1, v/v/v, vortexed and filtered again through 15 mm diameter, 0.2 μ m regenerated cellulose (RC) syringe filters (Phenex-RC, Phenomenex, Torrance, CA, USA) and 20 μ L of the diluted filtered extract was analyzed without any further pre-treatment.

2.3.2. LC-MS/MS analysis

LC-MS/MS analysis was carried out on a Varian 310 triple quadrupole (TQ) mass spectrometer (Varian, Italy), equipped with an electrospray ionization (ESI) source, a 212 LC pump, a ProStar 410 AutoSampler and dedicated software. Liquid chromatography (LC) separation was performed on a Gemini-NX C_{18} 100 × 2.0 mm i.d., 3 μ m particle size, 110 Å, equipped with a C_{18} 4 × 2 mm security guard cartridge column (Phenomenex, Torrance, CA, USA), using water (eluent A) and methanol (eluent B), both acidified with 0.1% v/v CH₃COOH, as eluents that were delivered at 200 μ L min⁻¹. The chromatographic and mass spectrometric conditions were described in detail by Scarpino et al. [26].

The results pertaining to the linearity range, the limit of detection (LOD), the limit of quantification (LOQ), the apparent recovery R_A (%), the matrix effects obtained through the evaluation of the signal suppression/enhancement SSE (%) and the recovery of the extraction R_E (%) were reported by Scarpino et al. [26].

2.4. Statistical analysis

¹Growth stage [24].

The normal distribution and homogeneity of variances were verified by performing the Kolmogorov–Smirnov normality test and the Levene test, respectively. An analysis of variance (ANOVA) was utilized to compare grain yield, TW, FHB incidence and severity, using a completely randomized block design, in which the year and the durum wheat *cvs*, with different degrees of FHB susceptibility, were the independent variables. Moreover, ANOVA was conducted separately for each year to evaluate the effect of the durum wheat *cvs*, with different FHB susceptibility, on the contamination of regulated, modified and emerging mycotoxins, using a completely randomized block design. In some analyses, the mycotoxin concentrations were transformed, using the y'=ln(x+1) equation, to normalize the residuals. SPSS for Windows, Version 26.0, (SPSS Inc., Chicago), was used for the statistical analysis.

3. Results

3.1. Meteorological data

The 8 growing seasons showed different meteorological trends, as far as both rainfall and temperature (expressed as growing degree days, GDDs) are concerned (Table 2).

Table 2. Monthly rainfall and growing degree days (GDD) for each growing season compared to data from 1990-2010.

| Parameters | Month | Average 1990-2010 | 2012-13 | 2013-14 | 2014-15 | 2015-16 | 2016-17 | 2017-18 | 2018-19 | 2019-20 |
|-----------------------------------------------|------------|----------------------|---------|---------|---------|---------|---------|---------|---------|---------|
| Rainfall | November | 85 | 182 | 68 | 438 | 5 | 158 | 48 | 124 | 314 |
| (mm) | December | 48 | 11 | 139 | 89 | 4 | 45 | 33 | 11 | 132 |
| | January | 38 | 17 | 117 | 36 | 14 | 4 | 107 | 6 | 5 |
| | February | 35 | 40 | 129 | 75 | 120 | 45 | 60 | 43 | 1 |
| | March | 38 | 118 | 71 | 88 | 52 | 69 | 109 | 17 | 62 |
| | April | 88 | 165 | 138 | 75 | 37 | 34 | 93 | 116 | 81 |
| | May | 105 | 101 | 84 | 96 | 171 | 79 | 188 | 178 | 122 |
| | June | 66 | 16 | 144 | 86 | 83 | 149 | 35 | 40 | 113 |
| _ | Nov - June | 503 | 650 | 890 | 982 | 486 | 582 | 673 | 535 | 830 |
| GDD 1 | November | 205 | 272 | 251 | 303 | 270 | 238 | 224 | 292 | 249 |
| $(\Sigma {}^{\circ}\text{C} d^{\text{-1}})$ | December | 110 | 113 | 169 | 184 | 164 | 144 | 113 | 151 | 193 |
| | January | 99 | 142 | 149 | 159 | 119 | 97 | 178 | 141 | 168 |
| | February | 142 | 110 | 179 | 143 | 158 | 152 | 113 | 195 | 229 |
| | March | 240 | 208 | 335 | 304 | 252 | 349 | 223 | 314 | 285 |
| | April | 337 | 398 | 428 | 409 | 405 | 415 | 456 | 393 | 414 |
| | May | 510 | 480 | 515 | 569 | 490 | 554 | 583 | 478 | 579 |
| | June | 594 | 620 | 637 | 659 | 624 | 673 | 665 | 667 | 624 |
| _ | Nov - June | 2239 | 2343 | 2662 | 2728 | 2481 | 2622 | 2554 | 2632 | 2741 |

¹ Accumulated growing degree days for each month using a 0°C base value.

Overall, the recent growing seasons showed higher temperatures than the average temperature for the 1990-2010 period, in particular from November to January, and in April and June. On average, the rainfall was also higher than in 1990-2010 (+200 mm), although the trends were highly variable, in terms of total precipitation and their distribution between seasons.

Frequent rainfall (> 120 mm) from wheat heading to the end of flowering (May) occurred in 2016, 2018, 2019 and 2020. In 2018 and 2020, the high rainfall in May was combined with high temperatures, while in 2016 and 2019, the GDDs during this period were

lower than in the other years. In 2015 and 2017, the limited rainfall and high temperature levels in May and June led to a rapid ripening and an early harvesting time.

3.2. Yield parameters

The recorded grain yields (on average 5.9 t ha⁻¹) were in agreement with the yield expected for durum wheat in growing areas without disease control at flowering. The moderately tolerant SY Cysco *cv* resulted in a significantly higher yield (+6.8%) and TW (+5.8%) than Saragolla (Table 3). The Fuego, Odisseo and Secolo *cv* grain yields did not differ from that of Saragolla, although these varieties all resulted in a significantly higher TW (Table 4). The TW was very low (< 77 kg hL⁻¹) in 2016 and in 2018.

The interaction between *cv* and year was significant for both the grain yield and TW (Tables 3 and 4). A greater difference was recorded for the yield parameters between SY Cysco and Saragolla in 2016, 2017 and 2019.

Table 3. Effects of the growing season and durum wheat cultivar with different degrees of FHB susceptibility on the grain yield (Yield), test weight (TW), FHB incidence (FHB Inc) and severity (FHB Sev) in NW Italy during the 2012 - 2020 period.

| Factor | Source of | Grain yield | TW | FHB Incidence 1 | FHB Severity ² | |
|----------------|------------------------------|-----------------------|------------------------|-----------------|---------------------------|--|
| Tuctor | variation | (t ha ⁻¹) | (kg hL ⁻¹) | (%) | (%) | |
| | 2013 | 5.89 cd | 80.77 a | 47.5 cd | 4.6 cd | |
| | 2014 | 7.27 a | 77.83 b | 28.6 e | 2.0 d | |
| | 2015 | 4.30 f | 80.53 a | 39.3 de | 1.4 d | |
| V = = = | 2016 | 6.52 bc | 76.38 c | 73.6 a | 20.9 a | |
| Year | 2017 | 6.07 bc | 77.83 b | 46.2 cd | 3.8 cd | |
| | 2018 | 5.00 e | 74.81 d | 44.7 cd | 10.2 b | |
| | 2019 | 5.27 de | 79.74 a | 53.6 bc | 6.7 bc | |
| | 2020 | 6.71 ab | 79.25 ab | 63.9 ab | 6.7 bc | |
| | <i>p</i> -value ³ | < 0.001 | < 0.001 | < 0.001 | < 0.001 | |
| | sem 4 | 2.56 | 5.54 | 36.3 | 16.6 | |
| Carltinan (an) | Saragolla | 5.63 b | 76.21 b | 71.3 a | 12.4 a | |
| Cultivar (cv) | SY Cysco | 6.01 a | 80.62 a | 27.2 b | 1.7 b | |
| | <i>p</i> -value ³ | 0.015 | < 0.001 | < 0.001 | < 0.001 | |
| | sem ⁴ | 0.22 | 2.96 | 30.9 | 7.5 | |
| Year * cv | <i>p</i> -value ³ | 0.006 | < 0.001 | 0.071 | < 0.001 | |
| rear " co | sem 4 | 1.25 | 3.89 | 18.1 | 18.9 | |

The reported data are the average of 3 replications.

3.3. FHB symptoms

FHB incidence and severity were higher in 2016, followed by 2018, 2019 and 2020 (Table 3).

The medium tolerant cv SY Cysco resulted in a significantly lower FHB incidence (-44.1%) and severity (-10.7%) than the susceptible Saragolla cv. The Secolo, Odisseo and Fuego *cvs* confirmed intermediate behavior between Saragolla and SY Cysco, as far as the FHB symptoms are concerned (Table 4). On average, Fuego did not differ from SY Cysco, while Secolo resulted in a higher amount of disease symptoms.

¹ FHB incidence was calculated as the percentage of ears with FHB damage, considering 200 ears per sample.

² FHB severity was calculated as the percentage of kernels per ear with FHB damage, considering 200 ears per sample.

³ *p*-value = level of significance of ANOVA.

⁴ sem = standard error of the means.

The interaction between *cv* and year was always significant. The differences between susceptible *cv* Saragolla and the other varieties were higher in 2016, 2018 and 2019.

Table 4. Effects of the growing season and durum wheat cultivar, with different degrees of FHB susceptibility, on the grain yield (Yield), test weight (TW), FHB incidence and severity in NW Italy during the 2015 - 2019 period.

| Factor | Source of variation | Grain Yield | TW | FHB Incidence 1 | FHB Severity ² | |
|---------------|------------------------------|-------------|-----------|-----------------|---------------------------|--|
| Factor | | (t ha-1) | (kg hL-1) | (%) | (%) | |
| | 2016 | 6.28 a | 76.32 c | 70.7 a | 26.4 a | |
| Year | 2017 | 5.75 b | 77.31 b | 40.2 b | 4.3 c | |
| rear | 2018 | 4.96 c | 76.29 c | 41.7 b | 8.4 b | |
| | 2019 | 5.25 c | 80.35 a | 42.5 b | 4.6 c | |
| | <i>p</i> -value ³ | < 0.001 | < 0.001 | < 0.001 | < 0.001 | |
| | sem 4 | 1.010 | 3.510 | 26.405 | 18.324 | |
| | Saragolla | 5.29 b | 74.51 d | 77.9 a | 17.9 b | |
| | Secolo | 5.58 b | 78.11 b | 52.1 b | 24.9 a | |
| Cultivar (cv) | Odisseo | 5.48 b | 77.36 c | 42.4 c | 6.4 c | |
| | Fuego | 5.31 b | 78.09 bc | 38.9 cd | 3.8 cd | |
| | SY Cysco | 6.08 a | 80.26 a | 31.1 d | 2.4 d | |
| | <i>p</i> -value ³ | < 0.001 | < 0.001 | < 0.001 | < 0.001 | |
| | sem ⁴ | 0.629 | 4.318 | 37.767 | 18.299 | |
| Year * cv | <i>p</i> -value ³ | 0.029 | < 0.001 | 0.001 | < 0.001 | |
| | sem ⁴ | 1.252 | 3.347 | 35.426 | 47.322 | |

The reported data are the average of 3 replications.

3.4. Mycotoxin contamination

About 10 mycotoxins were detected at the same time in at least one of the analyzed samples: 3-acetyldeoxynivalenol (3-ADON), deoxynivalenol (DON), deoxynivalenol-3-glucoside (DON-3-G), enniatin A (ENN A), enniatin A₁ (ENN A₁), enniatin B (ENN B), enniatin B₁ (ENN B₁), moniliformin (MON) and zearalenone (ZEA).

DON was detected for all of the years and in all of the cvs (Figures 1 and 2). The content of this mycotoxin was clearly related to the meteorological conditions, particularly close to anthesis, in each growing season. The DON contamination for the Saragolla and SY Cysco cvs was on average low in 2013, 2015 and 2017 (1888, 1233, 1453 $\mu g \ kg^{-1}$, respectively). The average contamination was clearly higher in 2014 (3081 $\mu g \ kg^{-1}$), 2016 (5573 $\mu g \ kg^{-1}$), 2019 (4540 $\mu g \ kg^{-1}$) and 2020 (3856 $\mu g \ kg^{-1}$). The highest contamination level was detected in 2018 (10815 $\mu g \ kg^{-1}$). As far as the modified DON forms are concerned, 3-ADON and DON-3-G were on average 5% and 11% of the total DON, respectively. However, 15-ADON was never detected. SY Cysco resulted in a significantly lower contamination of the total DON (-77%) than Saragolla, except for in 2020 (Figure 1).

The DON levels in the Saragolla cv were above the current EU regulatory limits (EC Regulation No. 1881/2006, with a limit of 1750 μ g kg) [12] for all the considered years. On the other hand, the cultivation of a moderately tolerant cv, SY Cysco, allowed the limits to be complied with, without the need of any fungicide application, for half of the considered growing seasons. Secolo, Odisseo and Fuego resulted in significantly lower levels than Saragolla in all the considered experiments (Figure 2). Their DON content was higher

¹ FHB incidence was calculated as the percentage of ears with FHB damage, considering 200 ears per sample.

² FHB severity was calculated as the percentage of kernels per ear with FHB damage, considering 200 ears per sample.

 $^{^{3}}$ *p*-value = level of significance of ANOVA.

⁴ sem = standard error of the means.

than SY Cysco in 2016, 2017 and 2019, while only in the 2018, with the highest total DON (DON TOT = sum of DON, DON-3-G and 3-ADON) was no difference in contamination observed between these *cvs*. In 2017 and 2019, Fuego showed a significant lower total DON content than Odisseo and Secolo, while Odisseo showed a lower contamination in 2016.

ZEA was detected in 2014, 2016, 2018 and 2019, albeit only in Saragolla, and in both Saragolla and SY Cysco in 2020 (Figure 1). The ZEA contamination in Saragolla was on average 178 μg kg⁻¹ in the aforementioned growing seasons. In these years, ANOVA showed a significantly lower ZEA content in SY Cysco than in Saragolla, except in 2019. No difference was recorded between SY Cysco and Secolo, Odisseo and Secolo in these years (Figure 2).

The highest contamination of MON for the Saragolla and SY Cysco cvs was detected in 2016 (514 μg kg⁻¹), followed by 2015 (251 μg kg⁻¹), 2020 (238 μg kg⁻¹) and 2018 (231 μg kg⁻¹). The content of this emerging mycotoxin was always below 200 μg kg⁻¹ in 2013, 2014, 2017 and 2019 (Figure 1). In all the years, SY Cysco showed a significantly lower MON contamination than Saragolla (-57%). Secolo, Odisseo and Fuego also had a significantly lower MON content than Saragolla, except for in 2018, while only in 2017 a significant difference between these cvs and SY Cysco was recorded (Figure 2).

ENNs (ENN TOT = the sum of ENN A, ENN A₁, ENN B, ENN B₁, ENN B₂) was the mycotoxin group with the highest content after DON and its modified forms. In the same way as for MON, 2016 was the growing season with the highest average contamination of ENN (2287 μ g kg⁻¹). The high content of these mycotoxins recorded in both 2015 (1002 μ g kg⁻¹) and in 2020 (1452 μ g kg⁻¹) highlight the absence of any relationship with the DON levels. The lowest ENN contaminations were observed in 2013 (49 μ g kg⁻¹) and 2017 (145 μ g kg⁻¹) (Figure 1). Significant differences between Saragolla and SY Cysco were only observed in 2015 (-62%), 2016 (-86%), 2019 (-43%) and 2020 (-56%), although greater differences than -80% were also recorded in 2013 and 2014. Secolo, Odisseo and Fuego had significantly lower ENN contents than Saragolla for all the considered growing seasons, although they did not differ from Sy Cysco, except for a lower contamination of Secolo in 2019 (Figure 2).

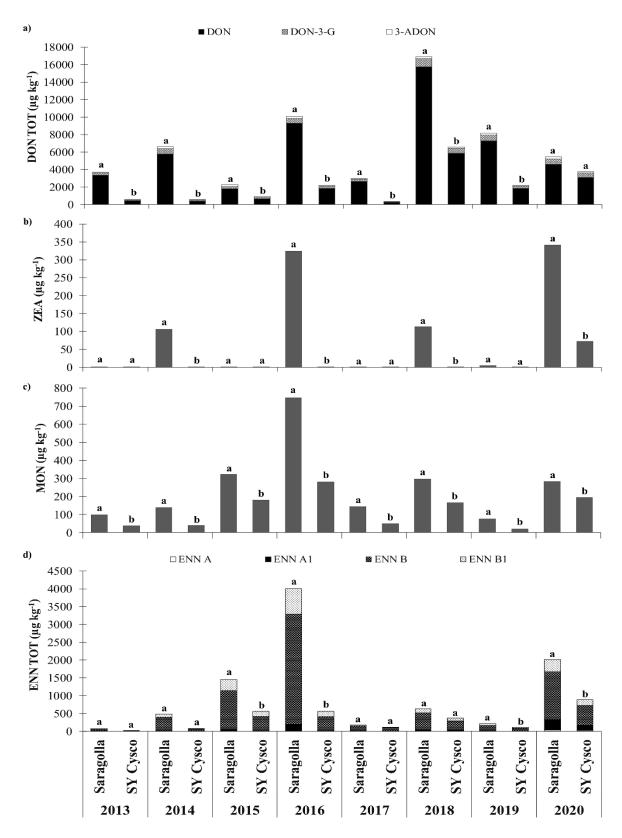


Figure 1. Effects of the growing season and durum wheat cultivar with different degrees of FHB susceptibility on the mycotoxin content in NW Italy during the 2012 - 2020 period: (a) Total deoxynivalenol content (DON TOT = sum of deoxynivalenol, DON; deoxynivalenol-3-glucoside, DON-3-G; and 3-acetyldeoxynivalenol, 3-ADON); (b) Zearalenone (ZEA) content; (c) Moniliformin (MON) content; (d) Total enniatin content (ENN TOT = sum of enniatin A, ENN A; enniatin A₁, ENN A₁; enniatin B, ENN B; and enniatin B₁, ENN B₁). Different letters above the bars indicate significant differences between cultivars for each growing season (p < 0.05). The reported data are the average of 3 replications. Statistical analysis was performed on the transformed [T; $y' = \ln(x + 1)$] mycotoxin concentration values.

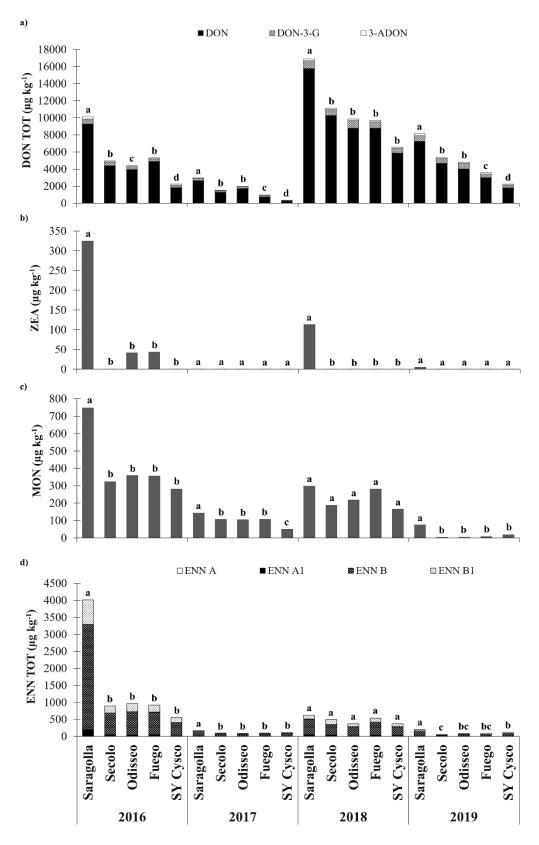


Figure 2. Effects of the growing season and durum wheat cultivar with different degrees of FHB susceptibility on the mycotoxin content in NW Italy during the 2015 - 2019 period: (a) Total deoxynivalenol content (DON TOT = sum of deoxynivalenol, DON; deoxynivalenol-3-glucoside, DON-3-G; and 3-acetyldeoxynivalenol, 3-ADON); (b) Zearalenone (ZEA) content; (c) Moniliformin (MON) content; (d) Total enniatin content (ENN TOT = sum of enniatin A, ENN A; enniatin A1, ENN A1; enniatin B, ENN B; and enniatin B1, ENN B1). Different letters above the bars indicate significant differences between cultivars for each growing season (p < 0.05). The reported data are the average of 3 replications.

3.5. Ratio between the modified and emerging mycotoxins and DON

As far as the ratio between the modified mycotoxins and DON is concerned, the DON-3-G/DON molar ratio (MR), was significantly higher for SY Cysco than for Saragolla over all the considered years, except for 2015 and 2020. Thus, SY Cysco showed a twotimes higher modifying ability of the parent form of DON than Saragolla (Figure 3). Secolo, Odisseo and Fuego never showed any differences, pertaining to their DON-3-G/DON MR from SY Cysco, except for Fuego in 2016 and Secolo in 2019, when significantly lower ratios were observed (Figure 4). Similarly, SY Cysco had a higher MON/DON ratio than Saragolla in 2013, 2014, 2016, 2017 and 2018, and almost two-times higher values were on average recorded in these years (Figure 3). Secolo, Odisseo and Fuego always showed a significantly lower MON/DON ratio than SY Cysco, except in 2018 (Figure 4). Conversely, the ENN TOT/DON TOT ratio showed variable behavior. Indeed, this ratio was only significantly higher for SY Cysco than for Saragolla in 2017 and in 2019, while it was significantly lower in 2020 (Figure 3). Like Saragolla, the Secolo, Odisseo and Fuego cvs also had a significantly lower ENN TOT/DON TOT ratio than SY Cysco in 2017 and 2019. The cvs that showed the significantly lowest ENN TOT/DON TOT ratios were Odisseo and Secolo in 2017 and 2019, respectively.

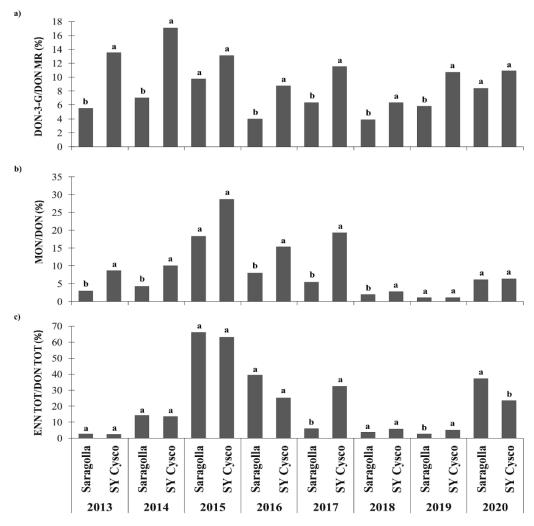


Figure 3. Effects of the growing season and durum wheat cultivar with different degrees of FHB susceptibility on the mycotoxin content ratios in NW Italy during the 2012 - 2020 period: (a) Deoxynivalenol-3-glucoside/deoxynivalenol molar ratio (DON-3-G/DON MR); (b) Moniliformin/Deoxynivalenol content ratio (MON/DON); (c) Total enniatin/Total deoxynivalenol content ratio (ENN TOT/DON TOT: ENN TOT = sum of enniatin A, ENN A; enniatin A1, ENN A1; enniatin B, ENN B; and enniatin B1, ENN B1; DON TOT = sum of deoxynivalenol, DON; deoxynivalenol-3-glucoside, DON-3-G; and 3-acetyldeoxynivalenol, 3-ADON). Different letters above the bars indicate significant differences between cultivars for each growing season (p < 0.05). The reported data are the average of 3 replications.

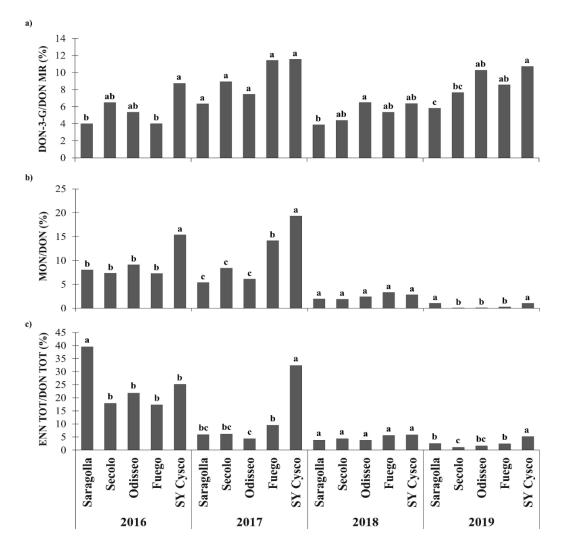


Figure 4. Effects of the growing season and durum wheat cultivar with different degrees of FHB susceptibility on the mycotoxin content ratios in NW Italy during the 2015 - 2019 period: (a) Deoxynivalenol-3-glucoside/deoxynivalenol molar ratio (DON-3-G/DON MR); (b) Moniliformin/Deoxynivalenol content ratio (MON/DON); (c) Total enniatin/Total deoxynivalenol content ratio (ENN TOT/DON TOT: ENN TOT = sum of enniatin A, ENN A; enniatin A₁, ENN A₁; enniatin B, ENN B; and enniatin B₁, ENN B₁; DON TOT = sum of deoxynivalenol, DON; deoxynivalenol-3-glucoside, DON-3-G; and 3-acetyldeoxynivalenol, 3-ADON). Different letters above the bars indicate significant differences between cultivars for each growing season (p < 0.05). The reported data are the average of 3 replications.

4. Discussion

The data collected in the present research highlight, for the first time ever, co-contamination by regulated (DON and ZEA), modified (DON-3-G and 3-ADON) and emerging *Fusarium* mycotoxins (MON and ENN A, A₁, B, B₁) of naturally-infected durum wheat *cvs* with different degrees of susceptibility to FHB over 8 growing seasons, in NW Italy. Previously, only a few studies had reported the co-occurrence of the aforementioned mycotoxins and these studies always referred to just a few years, mainly in areas of Central and Southern Italy [21,27,28], France [29], Poland [30], Canada [31] and Argentina [32], without comparing *cvs* with different degrees of susceptibility to FHB under field conditions in order to evaluate their contamination by emerging mycotoxins. In such a context, it is important to underline that the recent increase in demand for pasta products has also led to an expansion of the growth of winter durum wheat to non-traditional growing areas (such as Northern Italy, Austria, Germany and France), where there are more humid climatic conditions [4,28,33]. These factors have resulted in a higher risk of FHB infections, and, consequently, of mycotoxin contamination, thereby limiting their potential cultivation in these areas [4,28,33].

In the present study, which was specifically carried out in a location highly susceptible to FHB to allow an effective varietal screening, almost all the detected mycotoxins showed higher levels of contamination than those reported for trials conducted in Central and Southern Italy in the same years [27,28]. In relation to this aspect, the data highlight that the growing seasons characterized by frequent and heavy rainfall (> 120 mm), from wheat heading to the end of flowering (May), such as the 2016 and 2018 growing seasons, were also the years with the highest contamination of DON, ZEA, MON and ENNs.

Moreover, in the last few years, FHB has significantly increased on small-grain cereals, due to changes in crop management practices, minimum or reduced tillage, the intensification of maize in crop rotation and to weather patterns with more humidity and warm temperatures during anthesis as a result of climate change effects [34,35].

Like DON, the most susceptible *cv*, that is, Saragolla, exceeded the maximum level fixed for ZEA by EC Regulation No. 1881/2006 (100 µg kg⁻¹) [12]. in 2014, 2016, 2018 and 2020, reaching the highest levels of contamination of 325 and 341 µg kg⁻¹, respectively, in 2016 and 2020. ZEA is mainly produced by *F. graminearum* and the related species (i.e. *F. culmorum*) in cereals [36]. Therefore, its presence is commonly related to DON production [37]. ZEA has been shown to cause reproductive disorders in laboratory animals. Although the toxicity of ZEA in humans has not been conclusively established, the limited evidence available would seem to indicate that ZEA can cause the hyper estrogenic syndrome [38].

As far as the sanitary risk of modified and emerging mycotoxins is concerned, DON-3-G represents an additional risk to DON for human and animal health. In fact, this associated form could be hydrolyzed in the digestive tract of mammals, thereby contributing to the total dietary exposure of individuals to DON [39]. For this reason, together with its native form and with 3-ADON and 15-ADON, it should also be taken into account for correct risk assessments and food safety [40-42]. Nevertheless, to date, no regulatory limits have been established concerning the presence of MON and ENNs. Jonsson et al. [43] reported a high acute toxicity of MON in rats, with the LD50 value being at the same level as that of T-2 and HT-2 toxins, the most toxic of the *Fusarium* mycotoxins. In addition, Prosperini et al. [44] reported that, although in the last decade novel findings about a potential therapeutic action of ENN B have been presented, several in vitro and in vivo studies have revealed that ENN B interacts with primary target molecules, affects the biological response of cell defenses, promotes cell damage and produces potential interactions between food contaminants (particularly other mycotoxins), thus leading to abnormally high responses and to other molecular events underlying ENN B toxicity.

The development of more tolerant varieties is currently the most effective approach for controlling FHB and the occurrence of mycotoxins. The present data clearly show that the *cvs* with lower FHB susceptibility resulted in the lowest occurrence of DON, but also its modified forms (DON-3-G), ZEA and emerging mycotoxins, such as MON and ENNs.

Furthermore, in agreement with Berthiller et al., Dall'Asta et al. and Lemmens et al. [45-47], the most FHB resistant lines showed relatively more DON glycosylated (up to 30%) than susceptible cvs. In fact, the present data point out that the amount of DON-3-G relative to DON contamination varied between 5 and 15% and was influenced by both the genotype and the environment. Lemmens et al. [47] pointed out that since introgressing FHB resistance reduces both DON and DON-3-G levels in grain, although this reduction is lower for masked toxins, the specific resistance QTL (e.g. Fhb1) may enhance the speed or rate of DON detoxification. However, variances regarding the DON-3-G content, compared to its native form, and its distribution trend could also be explained by considering the different metabolic properties of each genotype in relation to that of fungi [48]. In addition to the genotype, the environment and the meteorological trend during ripening also play fundamental roles in establishing the extent and the relative amount of the contamination by co-occurring mycotoxins. In fact, depending on when a plant goes into the senescence period, its metabolism can be almost deactivated, thus preventing it from producing the glucoside form of DON [48]. In agreement with this statement, the highest DON-3-G/DON ratios were here recorded in 2013 and 2014, and these are also the years that were characterized by the longest ripening periods, as shown by the later harvesting time than for the other years, as a consequence of well distributed rainfall during ripening. Furthermore, the fungi, which developed to a greater extent on the peripheral tissues of the caryopsis, were still able to produce mycotoxins, as long as the moisture content during the dry-down process remained above 20%.

As far as the genotype × environment interaction is concerned, regardless of the pressure of the FHB disease, an inverse relationship between the level of contamination of DON and the DON-3-G/DON ratio was always recorded for all the *cvs* and years. These data suggest that the plant process of DON conjugation to glucose occurs with an intensity that is barely influenced by the occurrence of the free mycotoxin form, but which instead appears closely connected to the metabolic rate and the threshold biosynthetic ability of each *cv*. Thus, when the conditions (meteorological trend in spring and *cv* susceptibility) favor a high and quick increase in DON content during ripening, the DON-3-G/DON ratio tends to decrease, mainly because of high denominator values and an unequal proportional conversion to DON-3-G.

The absolute levels of the emerging mycotoxin contaminations highlight that after DON and its modified forms, ENNs represent the mycotoxin group with the highest content. As confirmed from the collected data, MON and ENNs are present at remarkably high contamination levels, in particular when adverse meteorological events occurred. In 2016, and considering the most susceptible cv Saragolla, the MON and ENNS were 747 and 4014 µg kg⁻¹, respectively. To the best of the authors' knowledge, the relative ratios between the emerging MON and ENN mycotoxins and the target mycotoxins of the FHB in wheat, DON, have never been reported before. These ratios indirectly indicate the variable resistance of the compared genotypes to the infections by the different Fusarium species that are able to produce their own characteristic mycotoxins. As already reported in the literature, although DON is mainly produced by *F. graminearum* and *F. culmorum* [11], F. avenaceum is the Fusarium species that is most able to biosynthesize depsipetides, such as ENN analogues and MON [18]. Like the DON-3-G/DON ratio, the MON/DON ratio confirms that, although the absolute MON levels of contamination for the more tolerant cvs were always the lowest, they showed higher MON biosynthetic ability than the susceptible ones, probably as a consequence of being relatively more prone to *F. avenaceum* infection. This ratio remained constantly higher in the FHB tolerant cvs in the years with both high (2016) and low (2017) DON contamination. Conversely, the ENN TOT/DON TOT ratio showed more variable behavior, as a function of the different years, and a less intense relationship with cv tolerance to FHB. This behavior could be linked to either a greater relative infection of F. avenaceum than F. graminearum, or a greater toxigenesis in relation to the environmental conditions that could favor *F. avenaceum vs F. graminearum* in different ways. However, only considering the presence of mycotoxins, it could be assumed that there is no real effect of flora inversion [49], because DON remained the main toxin, and the more tolerant cvs were also the least contaminated by all the other mycotoxins, such as the MON and ENNs mainly produced by *F. avenaceum*.

In conclusion, the present data underline that, even though the relative relationships between modified/emerging mycotoxins and native/target mycotoxins (DON, or DON TOT) could be higher in the more tolerant *cvs* than in susceptible ones, the genotypes that have a lower FHB susceptibility and/or DON biosynthetic ability also show lower absolute levels of contamination by other emerging and modified mycotoxins. In accordance with the consumers' and supply chains' request to reduce the use of pesticides, and as promoted in public policies (e.g. the *farm-to-fork strategy* in the EU), the selection of wheat genotypes resistant to FHB seems to be the most effective approach for controlling the overall mycotoxin risk, as confirmed in this experiment over several growing seasons with different meteorological trends.

Nevertheless, the most tolerant *cvs* that are also able to combine other food chain requirements, such as an adequate yield capacity, a high protein content, compliance with other qualitative parameters and a low susceptibility to other diseases (e.g. septoria leaf

blotch), should be inserted into cropping system and breeding programs designed to control not only the native mycotoxins, but also their modified forms and the emerging mycotoxins.

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References

- 1. Handmer, J.; Honda, Y.; Kundzewicz, Z.W.; Arnell, N.; Benito, G.; Hatfield, J.; Mohamed, I.F.; Peduzzi, P.; Wu, S.; Sherstyukov, B.; Takahashi, K.; Yan, Z. Changes in impacts of climate extremes: human systems and ecosystems. In *Managing the Risks of Extreme Events and Disasters to Advance Climate Change Adaptation*, A Special Report of Working Groups I and II of the Intergovernmental Panel on Climate Change (IPCC); Field, C.B., Barros, V., Stocker, T.F., Qin, D., Dokken, D.J., Ebi, K.L., Mastrandrea, M.D., Mach, K.J., Plattner, G.-K., Allen, S.K., Tignor, M., Midgley, P.M., Eds.; Cambridge University Press, Cambridge, UK, and New York, NY, USA, 2012, pp. 231-290.
- Porter, J.R.; Semenov, M.A. Crop responses to climatic variation. *Phil. Trans. R. Soc. B* 2005, 360, 2021-2035. https://doi.org/10.1098/rstb.2005.1752
- 3. Medina, A.; González-Jartín, J.M.; Sainz, M.J. Impact of global warming on mycotoxins. *Curr. Opin. Food Sci.* **2017**, *18*, 76-81. https://doi.org/10.1016/j.cofs.2017.11.009
- 4. Blandino, M.; Scarpino, V.; Sulyok, M.; Krska, R.; Reyneri, A. Effect of agronomic programmes with different susceptibility to deoxynivalenol risk on emerging contamination in winter wheat. *Europ. J. Agron.* **2017**, *85*, 12-24. https://doi.org/10.1016/j.eja.2017.01.001
- 5. Trnka, M.; Rötter; R.P.; Ruiz-Ramos, M.; Kersebaum, K. C.; Olesen, J. E.; Žalud, Z.; Semenov, M. A. Adverse weather conditions for European wheat production will become more frequent with climate change. *Nat. Clim. Chang.* **2014**, *4*, 637-643. https://doi.org/10.1038/nclimate2242
- 6. Chakraborty, S.; Tiedemann, A.V.; Teng, P.S. Climate change: potential impact on plant diseases. *Environ. Pollut.* **2000**, *108*, 317-326. https://doi.org/10.1016/s0269-7491(99)00210-9
- 7. Ramesh, K.; Matloob, A.; Aslam, F.; Florentine, S.K.; Chauhan, B.S. Weeds in a changing climate: vulnerabilities, consequences, and implications for future weed management. *Front. Plant Sci.* **2017**, *8*, 95. https://doi.org/10.3389/fpls.2017.00095
- 8. Jeger, M.J.; Pautasso, M. Plant disease and global change the importance of long-term data sets. *New Phytol.* **2008**, 177, 2-5. https://doi.org/10.1111/j.1469-8137.2007.02312.x
- 9. Beres, B.L.; Rahmani, E.; Clarke, J.M.; Grassini, P.; Pozniak, C.J.; Geddes, C.M.; Porker, K.D.; May, W.E.; Ransom, J.K. A Systematic Review of Durum Wheat: Enhancing Production Systems by Exploring Genotype, Environment, and Management (G × E × M) Synergies. Front. Plant Sci. 2020, 11, 568657. https://doi.org/10.3389/fpls.2020.568657
- 10. Miraglia, M.; Marvin, H.J.P.; Kleter, G.A.; Battilani, P.; Brera, C.; Coni, E.; Cubadda, F.; Croci, L.; De Santis, B.; Dekkers, S.; Filippi, L.; Hutjes, R.W.A.; Noordam, M.Y.; Pisante, M.; Piva, G.; Prandini, A.; Toti, L.; van den Born, G.J.; Vespermannh, A. Climate change and food safety: an emerging issue with special focus on Europe. *Food Chem. Toxicol.* **2009**, *47*, 1009-1021. https://doi.org/10.1016/j.fct.2009.02.005
- 11. Agriopoulou, S.; Stamatelopoulou, E.; Varzakas, T. Advances in Occurrence, Importance, and Mycotoxin Control Strategies: Prevention and Detoxification in Foods. *Foods* **2020**, *9*, 13. https://doi.org/10.3390/foods9020137
- 12. European Commission. Commission Regulation (EC) No. 1881/2006 setting maximum levels for certain contaminants in food-stuffs. *Off. J. Eur. Union* **2006**, *364*, 5-24.
- 13. Van der Fels-Klerx, H.J.; Liu, C.; Battilani, P. Modelling climate change impacts on mycotoxin contamination. *World Mycotoxin J.* **2016**, *9*, 717-726. https://doi.org/10.3920/WMJ2016.2066
- 14. Blandino, M.; Badeck, F.W.; Giordano, D.; Marti, A.; Rizza, F.; Scarpino, V.; Vaccino, P. Elevated CO₂ Impact on Common Wheat (*Triticum aestivum* L.) Yield, Wholemeal Quality, and Sanitary Risk. *J. Agric. Food Chem.* **2020**, *68*, 10574-10585. https://doi.org/10.1021/acs.jafc.0c02975

- 15. Van der Fels-Klerx, H.; Van Asselt, E.D.; Madsen, M.S.; Olesen, J.E. Impact of climate change effects on contamination of cereal grains with deoxynivalenol. *PloS One* **2013**, *8*, e73602. https://doi.org/10.1371/journal.pone.0073602
- 16. Blandino, M.; Haidukowski, M.; Pascale, M.; Plizzari, L.; Scudellari, D.; Reyneri, A. Integrated strategies for the control of Fusarium head blight and deoxynivalenol contamination in winter wheat. *Field Crop. Res.* **2012**, *133*, 139-14. http://dx.doi.org/10.1016/j.fcr.2012.04.004
- 17. Haile, J.K.; N'Diaye, A.; Walkowiak, S.; Nilsen, K.T.; Clarke, J.M.; Kutcher, H.R.; Steiner, B.; Buerstmayr, H.; Pozniak, C.J. Fusarium Head Blight in Durum Wheat: Recent Status, Breeding Directions, and Future Research Prospects. *Phytopathology* **2019**, 109, 1664-1675. http://dx.doi.org/10.1094/PHYTO-03-19-0095-RVW
- 18. Jestoi, M. Emerging *Fusarium*-mycotoxins fusaproliferin, beauvericin, enniatins, and moniliformin—a review. *Crit. Rev. Food Sci. Nutr.* **2008**, *48*, 21-49. http://dx.doi.org/10.1080/10408390601062021
- Rychlik, M.; Humpf, H.; Marko, D.; Dänicke, S.; Mally, A.; Berthiller, F., Klaffke, H.; Lorenz, N. Proposal of a comprehensive definition of modified and other forms of mycotoxins including "masked" mycotoxins. *Mycotoxin Res.*, 2014, 30, 197-205. https://doi.org/10.1007/s12550-014-0203-5
- 20. Khaneghah, A.M.; Martins, L.M.; von Hertwig, A.M.; Bertoldo, R.; Sant'Ana, A.S. Deoxynivalenol and its masked forms: Characteristics, incidence, control and fate during wheat and wheat based products processing-A review. *Trends Food Sci. Technol.* **2018**, *71*, 13-24. https://doi.org/10.1016/j.tifs.2017.10.012
- 21. Cirlini, M.; Generotti, S.; Dall'Erta, A.; Lancioni, P.; Ferrazzano, G.; Massi, A.; Galaverna, G.; Dall'Asta, C. Durum Wheat (*Triticum Durum* Desf.) Lines Show Different Abilities to Form Masked Mycotoxins under Greenhouse Conditions. *Toxins* **2014**, 6, 81-95. https://doi.org/10.3390/toxins6010081
- 22. Fraeyman, S.; Croubels, S.; Devreese, M.; Antonissen, G. Emerging *Fusarium* and *Alternaria* mycotoxins: occurrence, toxicity and toxicokinetics. *Toxins* **2017**, *9*, 228. http://dx.doi.org/10.3390/toxins9070228
- 23. Beck, H.E.; Zimmermann, N.E.; McVicar, T.R.; Vergopolan, N.; Berg, A.; Wood, E.F. Present and future Köppen-Geiger climate classification maps at 1-km resolution. *Sci. Data* **2018**, *5*, 180214. http://dx.doi.org/10.1038/sdata.2018.214
- 24. Zadoks, J.C.; Chang, T.T.; Konzak, C.F. A decimal code for the growth stages of cereals. *Weed Res.* **1974**, *14*, 415-421. https://doi.org/10.1111/j.1365-3180.1974.tb01084.x
- 25. Parry, D.W.; Jenkinson, P.; McLeod, L. Fusarium ear blight (scab) in small grain cereal A Review. *Plant Path.* **1995**, *44*, 207-238. https://doi.org/10.1111/j.1365-3059.1995.tb02773.x
- Scarpino, V.; Reyneri A.; Blandino M. Development and Comparison of Two Multiresidue Methods for the Determination of 17
 Aspergillus and Fusarium Mycotoxins in Cereals Using HPLC-ESI-TQ-MS/MS. Front. Microbiol. 2019, 10, 1-12.
 https://doi.org/10.3389/fmicb.2019.00361
- 27. Juan, C.; Covarelli, L.; Beccari, G.; Colasante, V.; Mañes, J. Simultaneous analysis of twenty-six mycotoxins in durum wheat grain from Italy. *FoodControl* **2016**, 62, 322-329. https://doi.org/10.1016/j.foodcont.2015.10.032
- 28. Beccari, G.; Prodi, A.; Senatore, M.T.; Balmas, V.; Tini, F.; Onofri, A.; Pedini, L.; Sulyok, M.; Brocca, L.; Covarelli, L. Cultivation Area Affects the Presence of Fungal Communities and Secondary Metabolites in Italian Durum Wheat Grains. *Toxins* **2020**, *12*, 97. https://doi.org/10.3390/toxins12020097
- 29. Orlando, B.; Grignon, G.; Vitry, C.; Kashefifard, K.; Valade, R. Fusarium species and enniatin mycotoxins in wheat, durum wheat, triticale and barley harvested in France. *Mycotoxin Res.* **2019**, *35*, 369-380. https://doi.org/10.1007/s12550-019-00363-x
- 30. Gorczyca, A., Oleksy, A., Gala-Czekaj, D.; Urbaniak, M.; Laskowska, M.; Waśkiewicz, A.; Stępień, L. *Fusarium* head blight incidence and mycotoxin accumulation in three durum wheat cultivars in relation to sowing date and density. Sci. Nat. **2018**, *105*, 2. https://doi.org/10.1007/s00114-017-1528-7
- 31. Tittlemier, S.A.; Blagden, R.; Chan, J.; Gaba, D.; Mckendry, T.; Pleskach, K.; Roscoe, M. Fusarium and Alternaria mycotoxins present in Canadian wheat and durum harvest samples. Can. J. Plant Pathol. 2019, 41, 403-414. https://doi.org/10.1080/07060661.2019.1592784
- 32. Palacios, S.A.; Erazo, J.G.; Ciasca, B.; Lattanzio, V.M.T.; Reynoso, M.M.; Farnochi, M.C.; Torres, A.M. Occurrence of deoxynivalenol and deoxynivalenol-3-glucoside in durum wheat from Argentina. *Food Chem.* **2017**, 230, 728-734. https://doi.org/10.1016/j.foodchem.2017.03.085
- 33. Steiner, B.; Michel, S.; Maccaferri, M.; Lemmens, M.; Tuberosa, R.; Buerstmayr, H. Exploring and exploiting the genetic variation of *Fusarium* head blight resistance for genomic-assisted breeding in the elite durum wheat gene pool. *Theor. Appl. Genet.* **2019**, 132, 969-988. https://doi.org/10.1007/s00122-018-3253-9
- 34. McMullen, M.; Bergstrom, G.; De Wolf, E.; Dill-Macky, R.; Hershman, D.; Shaner, G.; Van Sanford, D. A unified effort to fight an enemy of wheat and barley: *Fusarium* head blight. *Plant Dis* 2012, 96, 1712-1728. https://doi.org/10.1094/PDIS-03-12-0291-FE
- 35. Juroszek, P.; von Tiedemann, A. Linking plant disease models to climate change scenarios to project future risks of crop diseases: a review. *J. Plant Dis. Prot.* **2015**, 122, 3-15. https://doi.org/10.1007/BF03356525
- 36. Desjardins, A.E. Fusarium mycotoxins. Chemistry, genetics, and biology; APS Press, St. Paul, MN, USA, 2006; pp. 268.
- 37. Van der Fels-Klerx, H.J.; Klemsdal, S.; Hietaniemi, V.; Lindblad, M.; Ioannou-Kakouri, E.; Van Asselt, E.D. Mycotoxin contamination of cereal grain commodities in relation to climate in North West Europe. *Food Addit. Contam. Part A* **2012**, 29, 1581-1592. https://doi.org/10.1080/19440049.2012.689996
- 38. Rai, A.; Das, M.; Tripathi, A. Occurrence and toxicity of a fusarium mycotoxin, zearalenone. *Crit. Rev. Food Sci. Nutr.* **2020**, *60*, 2710-2729. https://doi.org/10.1080/10408398.2019.1655388
- 39. Berthiller, F.; Krska, R.; Domig, K. J.; Kneifel, W.; Juge, N.; Schuhmacher, R.; Adam, G. Hydrolytic fate of deoxynivalenol-3-glucoside during digestion. *Toxicol. Lett.* **2011**, 206, 264-267. https://doi.org/10.1016/j.toxlet.2011.08.006

- JECFA. Summary and Conclusions. In Joint Food and Agriculture Organization/World health Organization Expert Committee
 on food Additives, Proceedings of Joint FAO/WHO Expert Committee on food Additives Seventy-second Meeting, Rome, Italy,
 16-25 February 2010. http://www.who.int/foodsafety/chem/summary72 rev.pdf
- 41. Codex Alimentarius Commission. Report of the Fifth Session of the Codex Committee on Contaminants. In Foods, The Hague, The Netherlands, 2011. http://www.codexalimentarius.net/download/report/758/REP11 CFe.pdf
- 42. Lorenz, N.; Dänicke, S.; Edler, L.; Gottschalk, C.; Lassek, E.; Marko, D.; Rychlik, M.; Mally, A. A critical evaluation of health risk assessment of modified mycotoxins with a special focus on zearalenone. *Mycotoxin Res.* **2019**, *35*, 27-46. https://doi.org/10.1007/s12550-018-0328-z
- 43. Jonsson, M.; Atosuo, J.; Jestoi, M.; Nathanail, A. V.; Kokkonen, U.-M.; Anttila, M.; Koivisto, P.; Lilius, E.-M.; Peltonen, K. Repeated dose 28-day oral toxicity study of moniliformin in rats. *Toxicol. Lett.* **2015**, 233, 38-44. https://doi.org/10.1016/j.tox-let.2014.11.006
- 44. Prosperini, A.; Berrada, H.; Ruiz, M.J.; Caloni, F.; Coccini, T.; Spicer, L.J.; Perego, M.C.; Lafranconi, A. A Review of the Mycotoxin Enniatin B. *Front. Public Health* **2017**, *5*, 304. https://doi.org/10.3389/fpubh.2017.00304
- 45. Berthiller, F.; Dall'asta, C.; Corradini, R.; Marchelli, R.; Sulyok, M.; Krska, R.; Adam, G.; Schuhmacher, R. Occurrence of deoxynivalenol and its 3-β-D-glucoside in wheat and maize. *Food Addit Contam Part A* **2009**, 26, 507-511. https://doi.org/10.1080/02652030802555668
- 46. Dall'Asta, C.; Dall'Erta, A.; Mantovani, P.; Massi, A.; Galaverna, G. Occurrence of deoxynivalenol and deoxynivalenol-3-glucoside in durum wheat. *World Mycotoxin J.* **2013**, *6*, 83-91. https://doi.org/10.3920/WMJ2012.1463
- 47. Lemmens, M.; Steiner, B.; Sulyok, M.; Nicholson, P.; Mesterhazy, A.; Buerstmayr, H. Masked mycotoxins: does breeding for enhanced Fusarium head blight resistance result in more deoxynivalenol-3-glucoside in new wheat varieties? *World Mycotoxin J.* 2016, 9, 741-754. https://doi.org/10.3920/WMI2015.2029
- 48. Spaggiari, M.; Righetti, L.; Galaverna, G.; Giordano, D.; Scarpino, V.; Blandino, M.; Dall'Asta, C. HR-MS profiling and distribution of native and modified *Fusarium* mycotoxins in tritordeum, wheat and barley whole grains and corresponding pearled fractions. *J. Cereal Sci.* **2019**, *87*, 178-184. https://doi.org/10.1016/j.jcs.2019.03.009
- Scarpino, V.; Reyneri, A.; Sulyok, M.; Krska, R.; Blandino, M. Impact of the insecticide application to maize cultivated in different ent environmental conditions on emerging mycotoxins. *Field Crops Res.* 2018, 217, 188-198. https://doi.org/10.1016/j.fcr.2017.12.018