

Techniques of Transcriptomic and Proteomic Data Integration

Janardan Patil*, Li Len, Abhinav Bharat, and Xi Li

Department of Computer Science, Nagpur University, India 560987

*Email: jpatilnagpuredu@gmail.com

Phone: +91 9877659876

Abstract

As of not long ago, understanding the administrative conduct of cells has been sought after through autonomous investigation of the transcriptome or the proteome. In view of the focal creed, it was commonly accepted that there exist an immediate correspondence between mRNA records and produced protein articulations. In any case, late examinations have demonstrated that the relationship among's mRNA and Protein articulations can be low because of different factors, for example, unique half lives and post record apparatus. In this manner, a joint investigation of the transcriptomic and proteomic information can give valuable experiences that may not be translated from singular examination of mRNA or protein articulations. This article audits the current significant methodologies for joint investigation of transcriptomic and proteomic information. We order the various methodologies into eight primary classes dependent on the underlying calculation and last investigation objective. We further present analogies with different spaces and talk about the current exploration issues around there.

Keywords: Transcriptome, Proteome, modeling.

INTRODUCTION

Two significant observational classifications includes estimation of transcriptomic profiles through procedures, for example, microarray, RNA-seq and so forth and estimation of proteomic profiles through strategies, for example, gel electrophoresis and mass spectrometry. A portion of the information estimation strategies may include annihilation of the living cell and along these lines joint estimation of the two records and proteins in a solitary cell won't be plausible by such techniques. Moreover, a few methodologies may give articulation information on the normal conduct of an assortment of cells and not the articulation dissemination of the cells. Subsequently, understanding the restrictions and suppositions in the information estimation procedures utilized for estimating the transcriptomic and proteomic profiles is basic prior to directing a joint examination of the two information sources. The subsequent stage in building up a joint model of the two spaces includes fathoming the distinctions in the statement of the mRNAs and proteins. Studies [1-5] have demonstrated that there can be helpless relationship among's mRNA and protein articulation information from same cells under comparable conditions. The current audit centers around revealing the essential classes of approaches that have been proposed for combination of transcriptomic and proteomic information. In examination, existing audits on joint transcriptomic and proteomic profiling centers around explicit parts of consolidated investigation. For example, Catherine Hack [6] centers around various measurable techniques for relationship among's transcriptomic and proteomic datasets. Cox et al. [7] audits various strategies for correlation of microarray and proteomic datasets alongside bunching and combining choices for these datasets. Nie et al. [8] centers around endeavors to create different factual devices for improving the odds of catching a connection among transcriptomic and proteomic information alongside various

change and standardization strategies for information, consequences for estimation blunders and difficulties of missing qualities in datasets. A huge piece of the paper by Hecker et al. [9] surveys ways to deal with fabricate dynamic models of transcriptomic and additionally proteomic network. Simon Rogers [10] depicted the accessible factual devices for crossing over multi-omics information. This can be considered as one of the most clear incorporation types.

METHOD & RESULTS

Approaches identified with this sort for the most part consider an association of two distinctive informational collections (proteomic information and transcriptomic information; not from a similar example) and afterward make a reference informational collection. The reference informational indexes have at times demonstrated new bits of knowledge and uncovered beforehand undetected wonder or upheld another marvel when contrasted with the individual informational indexes. There are various methodologies identified with this [11-30]. A work on *Bradyrhizobium japonicum* bacteroid digestion in soybean root knobs by Nathanael et al. [31] can be a case of this technique. In this examination, creators have aggregated a reference dataset by joining (association) transcriptomic and proteomic information. In light of the reference dataset, they have found huge number of proteins identified with a few sorts of bacterial digestion that were absent in the dataset from proteomic concentrate alone. Segment 5.1 quickly surveys the methodology considered by Nathanael et al.

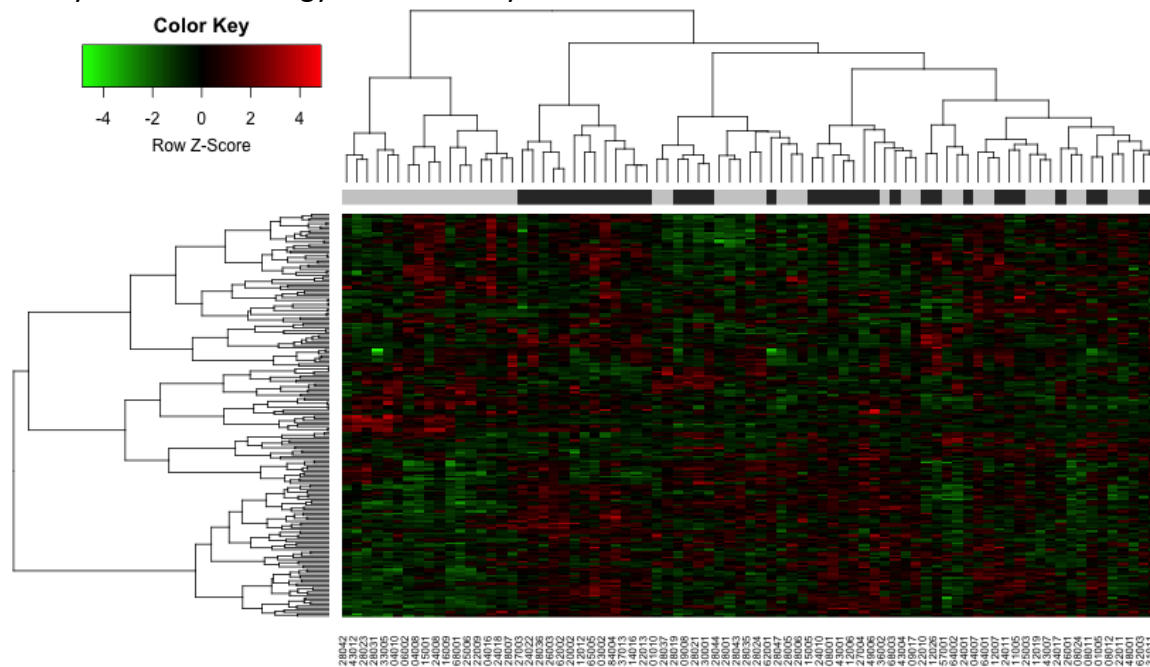


Figure 1: Heatmap comparing 50 samples transcriptomic clustering results for selected genes.

For different reasons [32-55], transcriptomic and proteomic information might not have direct cover in highlights (here element alludes to various qualities for records and proteins). In any case, includes on transcriptomic and proteomic level may have a similar utilitarian setting. These utilitarian settings may allude to various organic cycles or pathways in which highlights from the two records and proteins are enhanced. In this methodology, the regular utilitarian settings are

separated through the investigation of both transcriptomic and proteomic datasets fair and square of protein communication organizations. This methodology was distributed in 2010 by Paul et al. [56-70] which is examined in segment 5.2. Creators of this distribution likewise produced omicsNET for discovering reliance between highlights of proteomics and transcriptomics.

A comparative sort of approach (utilitarian examination) was applied for incorporating transcriptomic and proteomic assessment of gentamicin nephrotoxicity in rodents by Com et al. in 2011 [71-100]. Be that as it may, the practical examination was finished by GO-Browser 2 (an in-house Gene Ontology based explanation instrument) with assistance of Ingenuity Pathway Analysis software³. In view of the practical examination, some quality cosmology natural cycles were chosen which were improved by the highlights of the transcriptomic and proteomic dataset with Fisher $p \leq 0.05$. This incorporation by useful examination uncovers a putative model of harmfulness [39] in the kidney of rodents.

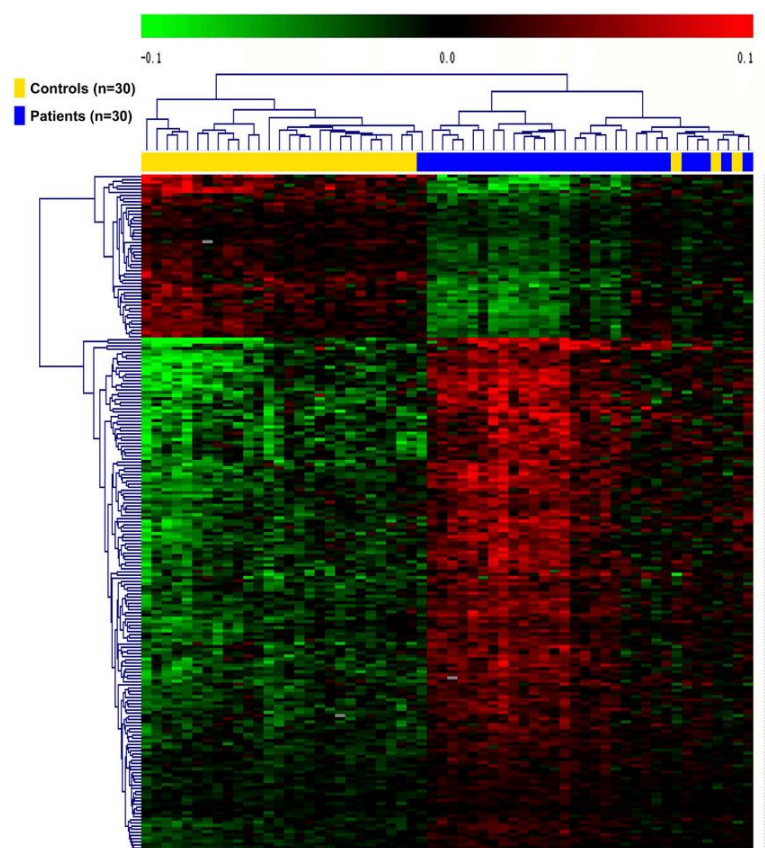


Figure 2: Heatmap comparing 60 samples transcriptomic clustering results for all significant genes.

Topological organization techniques (over-association examination, concealed hub investigation, rank collection and organization examination) have been utilized to clarify the regular controllers (transcriptional components and receptors) from two distinct kinds of informational indexes (transcriptomic and proteomic) by Eleonora Piruzian et al. [37]. This class of approach alludes to finding upstream controllers of mRNA and proteins independently and gathering the basic controllers in both the organizations for a consolidated flagging pathway. Topological and network investigation was utilized in discovering singular record factors (TF) of mRNAs and Proteins. The

TFs that were not regular in transcriptomic and proteomic profiles were disregarded and the normal TFs were utilized to locate the most powerful receptors that could trigger maximal conceivable transcriptional reaction. Among the receptors found from joint investigation, some of them were never revealed as psoriasis markers in prior examinations while some of them have been accounted for previously. In another as of late distributed examination [76], a coordinated quantitative proteomic, transcriptomic, and network investigation approach was talked about which additionally uncovers atomic highlights of tumorigenesis and clinical backslide. Segment 5.3 talks about the methodology of Eleonora et al..

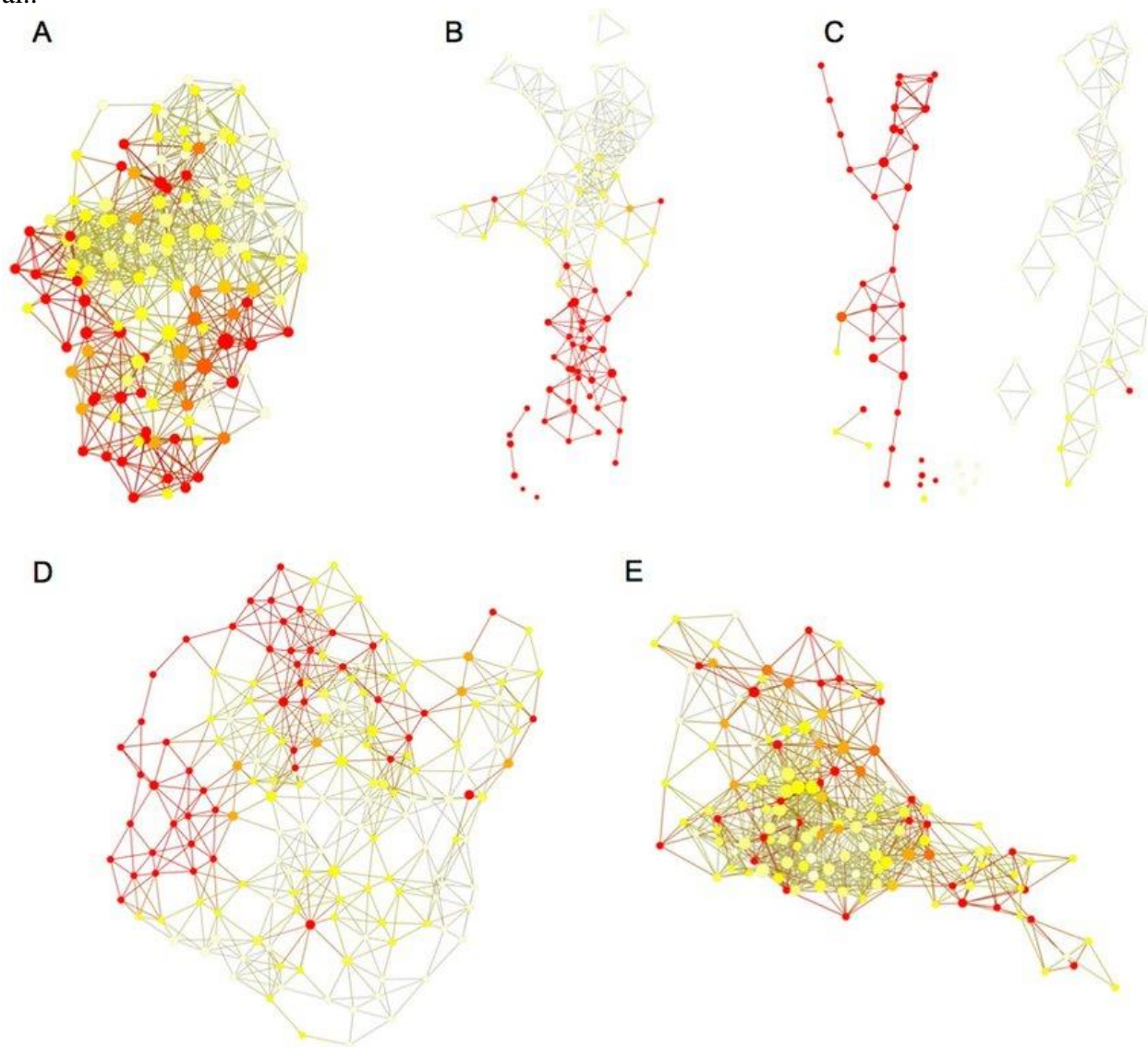


Figure 3: Topological organization of selected proteins.

Figure 4 combination consolidates various proteomic informational collections into a blended proteomic informational collection alongside joining numerous transcriptomic informational collections into a reference transcriptomic informational index. The transcriptomic and proteomic datasets that are blended can be made by various transcriptomic and proteomic profiling

separately. Subsequent to blending the datasets, relationship investigation is directed between these 2 combined informational collections and it is indicated that the coefficient of connection is superior to the one without consolidating. Moreover, explicit subsets of the consolidated informational collections can have higher coefficient of relationship. Dov Greenbaum et al. [67] utilized such a methodology in their distribution in 2003 which is talked about in segment 5.4.

This class of combination utilizes non-straight or direct enhancement to foresee missing estimations of proteomic information. It expands a target capacity to discover the associations among transcriptomic and proteomic networks. Notwithstanding, they don't bring about a powerful model ready to anticipate the wealth of next time point yet rather, they can foresee the protein articulation simultaneously point. A genuine case of non-straight advancement is a strategy portrayed in Wandaliz Torres-Garcia et al. [77] for an investigation of *Desulfovibrio vulgaris* distributed in 2009. The technique depends on stochastic inclination boosting tree (GBT) proposed by Friedman et al. [78]. Stochastic GBT enhancement method was likewise utilized in an investigation of *Shewanella oneidensis* in 2011 [79]. Counterfeit neural organization approach was applied to locate the missing estimations of the proteins utilizing the relations among transcriptomic and proteomic information in a different report distributed in 2011 [80]. In segment 5.5, we quickly survey the methodology made by Garcia et al. [77] in their *Desulfovibrio vulgaris* study.

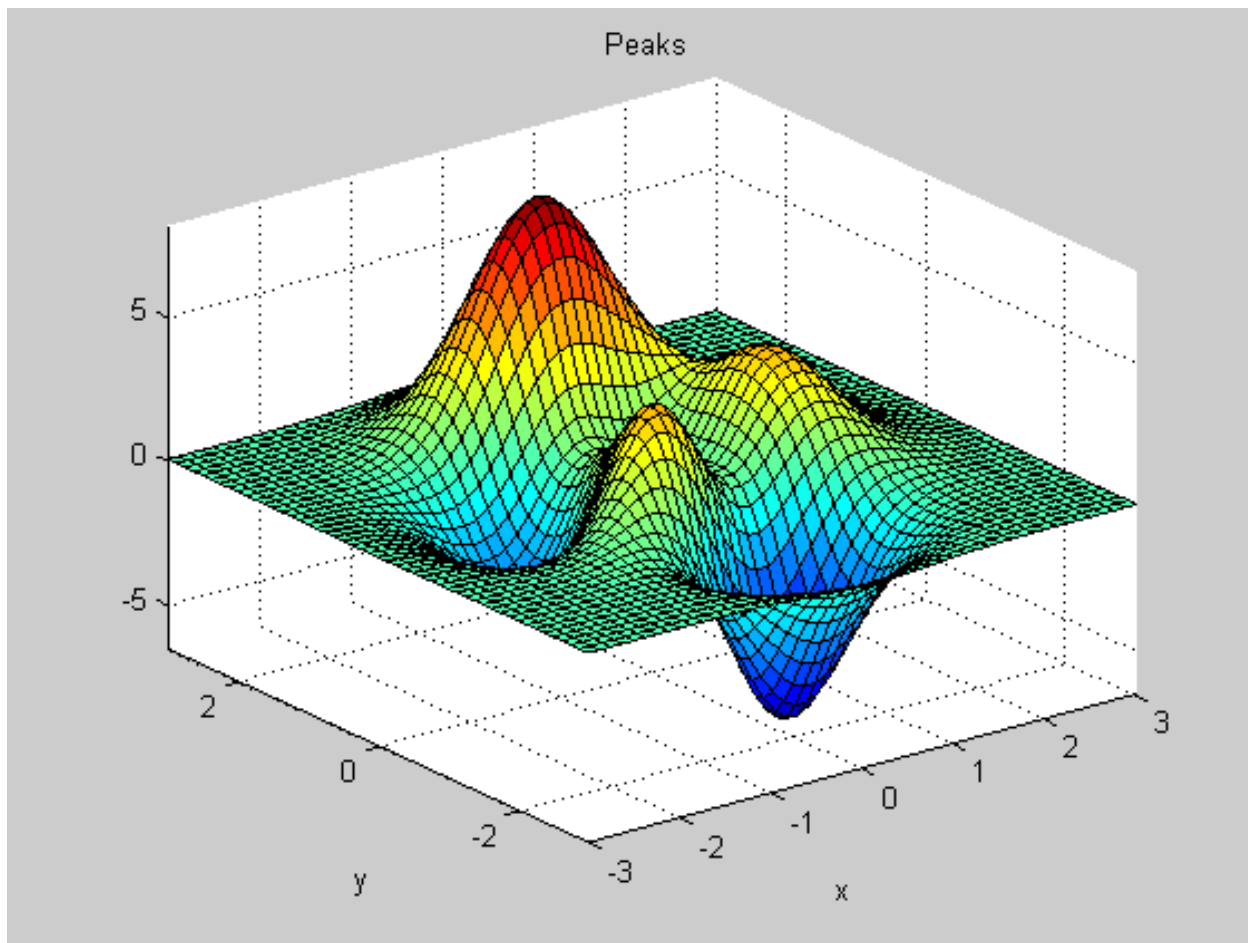


Figure 4: Transcriptional transformation index.

Protein plentitude isn't simply identified with comparing mRNA wealth yet additionally relies upon other natural and substance factors (named as covariates). Hence, the possibility of numerous relapse examination is utilized to relate attributes of various covariates of every individual quality with the mRNA-protein relationship. The different relapse approach can give a superior clarification of protein changeability than customary single relapse strategy. Impact of numerous succession highlight (one sort of covariate) on mRNA-protein relationship was examined by Nie et al. in 2006 [81] where they have utilized numerous relapse examination. Case of another direct relapse model can be Poisson's straight relapse model which has been utilized by Lie Nie et al. [47] to explain the relationship model of transcriptomic and proteomic networks. In area 5.6, we quickly clarify the numerous relapse examination utilized in [81].

Grouping mRNA and protein wealth datasets exclusively and finding likenesses (and subsequently relationship) between's the individual bunches doesn't create promising outcomes (as clarified in segment 5.7). This disappointment prompts the suspicion that connecting the proteomic and transcriptomic datasets and afterward grouping the linked dataset may not be a smart thought either (subtleties in segment 5.7). In view of these perceptions, another bunching technique called coupled grouping was executed by Rogers et al. [82]. Couple bunching makes certain number of proteomic and transcriptomic groups and gives the restrictive likelihood of a quality to be in a protein group given that it is in a mRNA bunch. These contingent probabilities can uncover the social unpredictability of mRNA and protein information. Rogers et al. utilized time arrangement transcriptomic and proteomic information extricated under same exploratory conditions. Segment 5.7 examines coupled bunching approach. We would need to accentuate that this kind of approach is likewise not a unique demonstrating approach that can give worldly forecasts.

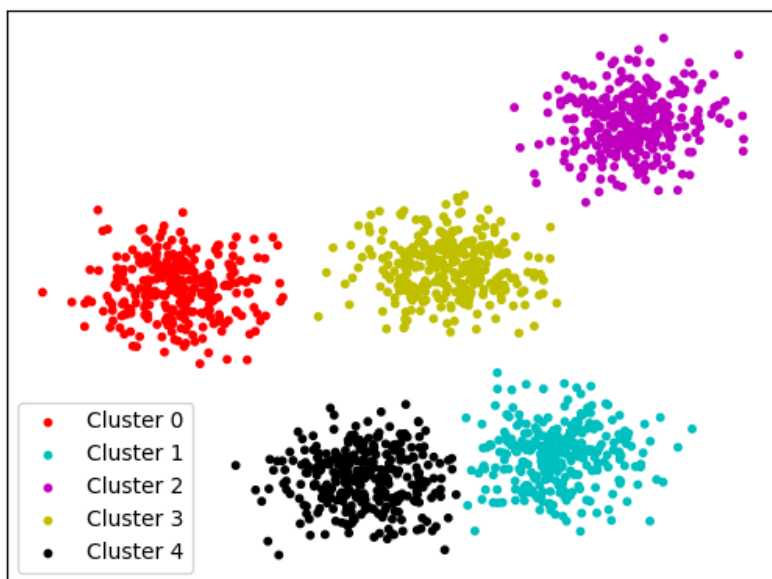


Figure 5: Clustering of selected proteins.

Various investigations announced in the writing have deduced dynamic models (, for example, Boolean organization, direct models, differential condition models, Bayesian organizations and so on) of GRNs from time arrangement transcriptomic information alone. For instance, Liang et al. [83] utilized REVEAL calculation for surmising of Boolean organization model from time arrangement mRNA articulation information. A fundamental straight demonstrating has been proposed by D'haeseleer [84]. GRN Models comprising of differential conditions was utilized by Guthke et al. [100-120]. Approval of deduction methods of GRN was examined by Edward R Dougherty [86]. Friedman utilized Bayesian organizations to dissect and show quality articulation information [121-152]. Among the current organization models, Bayesian organizations can be applied to consolidate heterogeneous information and earlier natural information. For instance, Nairai et al. [88] utilized protein-protein connection network information for refining the Bayesian Network model of the GRN created by mRNA information alone. Yu Zhang et al. [89] utilized transcriptional factor restricting site information and quality articulation information (transcriptomic) to show GRN utilizing Bayesian organization approach. Werhli et al. [90] coordinated various wellsprings of earlier natural information (TF restricting area) with microarray articulation information to create a Bayesian organization model.

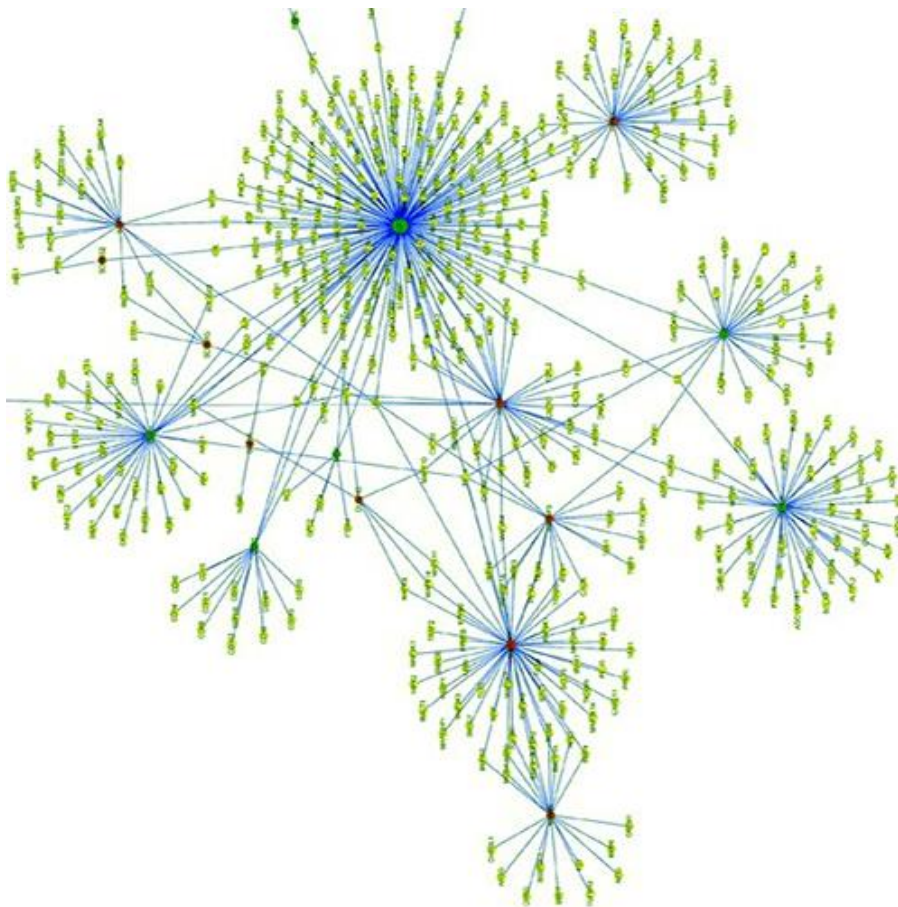


Figure 6: Interactome organization of selected proteins.

DISCUSSION

In view of the focal creed, it was commonly accepted that there exist an immediate correspondence between mRNA records and produced protein articulations. In any case, late examinations have demonstrated that the relationship among's mRNA and Protein articulations can be low because of different factors, for example, unique half lives and post record apparatus. In this manner, a joint investigation of the transcriptomic and proteomic information can give valuable experiences that may not be translated from singular examination of mRNA or protein articulations. This article audits the current significant methodologies for joint investigation of transcriptomic and proteomic information. We order the various methodologies into eight primary classes dependent on the underlying calculation and last investigation objective. We further present analogies with different spaces and talk about the current exploration issues around there.

CONFLICT OF INTEREST

The author(s) confirm that this article content has no conflicts of interest.

REFERENCES

1. Guoan Chen , Tarek G Gharib, Chiang-Ching Huang , Jeremy M G Taylor, David E Misek, Sharon L R Kardia, Thomas J Giordano, Gharib Chiang-Ching Huang Jeremy M G, Mark D Iannettoni, Mark B Orringer, Samir M Hanas, David G Beer. "Discordant protein and mrna expression in lung adenocarcinomas". *Molecular and Cellular Proteomics*. 2002;1(4):304–313.
2. Laura E Pascall , Lawrence D True , Eric W Deutsch , David S Campbell , Michael Risk, Ilsa M Coleman , Lillian J Eichner , Peter S Nelson , Alvin Y Liu. "Correlation of mrna and protein levels: Cell type-specific gene expression of cluster designation antigens in the prostate". *BMC Genomics*. 2008;9 (246)
3. Gygi SP, Rochon Y, Franz A, Aebersold R. "Correlation between protein and mRNA abundance in yeast" *Mol Cell Biol*. 1999;19 :1720–1730.
4. Edward Yeung S. "Genome-wide correlation between mrna and protein in a single cell" *Angewandte Chemie International Edition*. 2011;50 (3):583–585.
5. Anatole Ghazalpour, Brian Bennett, Vladislav A Petyuk, Luz Orozco, Raffi Hagopian, Imran N Mungrue, Charles R Farber, Janet Sinsheimer, Hyun M Kang, Nicholas Furlotte, Christopher C Park, Ping-Zi Wen, Heather Brewer, Karl Weitz, David G Camp II, Calvin Pan, Roumyana Yordanova, Isaac Neuhaus, Charles Tilford, Nathan Siemers, Peter Gargalovic, Eleazar Eskin, Todd Kirchgesner, Desmond J Smith, Richard D Smith, Aldons J Lusis. "Comparative analysis of proteome and transcriptome variation in mouse". *PLoS Genet*. 2011 Jun;7(6):e1001393.
6. Catherine Jane Hack. "Integrated transcriptome and proteome data: The challenges ahead". *Briefings in functional genomics and proteomics*. 2004;3 :212–219.
7. Brian Cox, Thomas Kislinger, Andrew Emili. "Integrating gene and protein expression data: pattern analysis and profile mining" *Methods*. 2005;35 :303–314.
8. Lei Nie, Gang Wu, David E Culley , Johannes C M Scholten , Weiwen Zhang. "Integrative analysis of transcriptomic and proteomic data: challenges, solutions and applications" *Critical Reviews In Biotechnology*. 2007;27 (2):63–75.
9. Michael Hecker, Sandro Lambeck, Susanne Toepfer, Eugene van Someren, Reinhard Guthke. "Gene regulatory network inference: Data integration in dynamic models—a review" *Biosystems*. 2009;96 (1):86–103.
10. Simon Rogers. "Statistical methods and models for bridging omics data levels" *Methods in Molecular Biology*. 2011;719 (1):133–151.
11. Michael Heller J. "Dna microarray technology: Devices, systems, and applications". *Annual Review of Biomedical Engineering*. 2002;4(1):129–153.
12. Ed M Southern. "Blotting at 25" *Trends in Biochemical Sciences*. 2000;25 (12):585–588.
13. Pieter Vos, Rene Hogers, Marjo Bleeker, Martin Reijans, Theo van de Lee, Miranda Hornes, Adrie Friters, Jerina Pot, Johan Paleman, Martin Kuiper, Marc Zabeau. "Aflp: a new technique for dna fingerprinting" *Nucleic Acids Research*. 1995;23 (21):4407–4414.
14. PUNCHAPAT SOJIKUL, PANIDA KONGSAWADWORAKUL, UNCHERA VIBOONJUN, JITRAWAN THAIPRASIT, BURAPAT INTAWONG, JARUNYA NARANGAJAVANA, MOM RAJAWONG JISNUSON SVASTI. "Aflp-based transcript profiling for cassava genome-wide expression analysis in the onset of storage root formation" *Physiologia Plantarum*. 2010; 140(2):189–298.
15. Ahsan N, Lee DG, Kim KH, Alam I, Lee SH, Lee KW, Lee H, Lee BH (2010) Analysis of arsenic stress-induced differentially expressed proteins in rice leaves by two-dimensional gel electrophoresis coupled with mass spectrometry. *Chemosphere* 78:224–231
16. Ahsan N, Nakamura T, Komatsu S (2012) Differential responses of microsomal proteins and metabolites in two contrasting cadmium (Cd)-accumulating soybean cultivars under Cd stress. *Amino Acids* 42:317–327

17. Akashi K, Nishimura N, Ishida Y, Yokota A (2004) Potent hydroxyl radical-scavenging activity of drought-induced type-2 metallothionein in wild watermelon. *Biochem Bioph Res Commun* 323:72–78
18. Al Mahmud J, Hasanuzzaman M, Nahar K, Rahman A, Hossain MS, Fujita M (2017) Maleic acid assisted improvement of metal chelation and antioxidant metabolism confers chromium tolerance in *Brassica juncea* L. *Ecotoxicol Environ Saf* 144:216–226
19. Ali H, Khan E, Sajad MA (2013) Phytoremediation of heavy metals- concepts and applications. *Chemosphere* 91:869–881
20. Alves M, Moes S, Jenö P, Pinheiro C, Passarinho J, Ricardo CP (2011) The analysis of *Lupinus albus* root proteome revealed cytoskeleton altered features due to long-term boron deficiency. *J Proteome* 74:1351–1363
21. Anjum NA, Gill SS, Duarte AC, Pereira E, Ahmad I (2013) Silver nanoparticles in soil plant systems. *J Nanopart Res* 15:1–26. <https://doi.org/10.1007/s11051-013-1896-7>
22. Pawar, S., Donthamsetty, S., Pannu, V., Rida, P., Ogden, A., Bowen, N., Osan, R., Cantuaria, G., and Aneja, R. (2014) KIFCI, a novel putative prognostic biomarker for ovarian adenocarcinomas: delineating protein interaction networks and signaling circuitries. *J. Ovarian Res.* 7, 53
23. Batra, H.; Pawar, S.; Bahl, D. Curcumin in combination with anti-cancer drugs: A nanomedicine review. *Pharm. Res.* 2018, 139, 91–105.
24. Mittal, K., Choi, D.H., Klimov, S. et al. A centrosome clustering protein, KIFCI, predicts aggressive disease course in serous ovarian adenocarcinomas. *J Ovarian Res* 9, 17 (2016). <https://doi.org/10.1186/s13048-016-0224-0>
25. Anjum NA, Hasanuzzaman M, Hossain MA, Thangavel P, Roychoudhury A, Gill SS, Rodrigo MAM, Adam V, Fujita M, Kizek R, Duarte AC, Pereira E, Ahmad I (2015) Jacks of metal/metalloid chelation trade in plants-an overview. *Front Plant Sci* 6:192. <https://doi.org/10.3389/fpls.2015.00192>
26. Arenhart RA, Lima JC, Pedron M, Carvalho FE, Silveira JA, Rosa SB, Caverzan A, Andrade CM, Schünemann M, Margis R, Margis-Pinheiro M (2013) Involvement of ASR genes in aluminium tolerance mechanisms in rice. *Plant Cell Environ* 36:52–67
27. Baker AJM, McGrath SP, Reeves RD, Smith JAC (2000) Metal hyperaccumulator plants: a review of the ecology and physiology of a biological resource for phytoremediation of metal polluted soils. In: Terry N, Banuelos G (eds) *Phytoremediation of contaminated soil and water*. Michael Lewis publishers, Boca Raton, pp 85–107
28. Barcelo J, Poschenrieder C (1990) Plant water relations as affected by heavy metal stress: a review. *J Plant Nutr* 13:1–37. <https://doi.org/10.1080/01904169009364057>
29. Bernhard WR, Kagi JH (1987) Purification and characterization of a typical cadmium-binding polypeptides from *Zea mays*. *Experientia Suppl* 52:309–315
30. Ashraf, M. I., Ong, S. K., Mujawar, S., Pawar, S., More, P., Paul, S., et al. (2018). A side-effect free method for identifying cancer drug targets. *Sci. Rep.* 8:6669. doi: 10.1038/s41598-018-25042-2
31. Lahiri, C., Pawar, S., Sabarinathan, R., Ashraf, M. I., Chand, Y., and Chakravorty, D. (2014). Interactome analyses of *Salmonella* pathogenicity islands reveal SicA indispensable for virulence. *J. Theor. Biol.* 363, 188–197. doi: 10.1016/j.jtbi.2014.08.013
32. Bona E, Marsano M, Massa M, Cattaneo C, Cesaro P, Argese E, Toppi LS, Cavaletto M, Berta G (2011) Proteomic analysis as a tool for investigating arsenic stress in *Pteris vittata* roots colonized or not by arbuscular mycorrhizal symbiosis. *J Proteome* 74:1338–1350
33. Burkhead JL, Reynolds KA, Abdel-Ghany SE, Cohu CM, Pilon M (2009) Copper homeostasis. *New Phytol* 182:799–816. <https://doi.org/10.1111/j.1469-8137.2009.02846.x>
34. Carvajal M, Cooke DT, Clarkson DT (1996) Responses of wheat plants to nutrient deprivation may involve the regulation of water-channel function. *Planta* 199:372–381
35. Casterline JL, Barnett NM (1982) Cadmium-binding components in soybean plants. *Plant Physiol* 69:1004–1007
36. Castrillo G, Sánchez-Bermejo E, de Lorenzo L, Crevillén P, Fraile-Escanciano A, Tc M, Mouriz A, Catarecha P, Sobrino-Plata J, Olsson S, Leo Del Puerto Y, Mateos I, Rojo E, Hernández LE, Jarillo JA, Piñero M, Paz-Ares J, Leyva A (2013) WRKY6 transcription factor restricts arsenate uptake and transposon activation in *Arabidopsis*. *Plant Cell* 25:2944–2957
37. Chen Z, Pan Y, Wang S, Ding Y, Yang W, Zhu C (2012) Overexpression of a protein disulfideisomerase-like protein from *Methanothermobacter thermoautotrophicum* enhances mercury tolerance in transgenic rice. *Plant Sci* 197:10–20
38. Pawar S, Ashraf MI, Mujawar S, Mishra R and Lahiri C (2018) In silico Identification of the Indispensable Quorum Sensing Proteins of Multidrug Resistant *Proteus mirabilis*. *Front. Cell. Infect. Microbiol.* 8:269. doi: 10.3389/fcimb.2018.00269
39. Michel Claverie, Marlène Souquet, Janine Jean, Nelly Forestier-Chiron, Vincent Lepitre, Martial Pr , John Jacobs, Danny Llewellyn, Jean-Marc Lacape. “cdna-afp-based genetical genomics in cotton fibers” *TAG Theoretical and Applied Genetics.* 2012;124 :665–683.
40. Ge Xiaomeng, Chen Weihua, Song Shuhui, Wang Weiwei, Hu Songnian, Yu Jun. “Transcriptomic profiling of mature embryo from an elite super-hybrid rice *lyp9* and its parental lines” *BMC Plant Biology.* 2008;8(114)
41. MD Adams, JM Kelley, JD Gocayne, M Dubnick, MH Polymeropoulos, H Xiao, CR Merril, A Wu, B Olde, RF Moreno, et al. “Complementary dna sequencing: expressed sequence tags and human genome project” *Science.* 1991;252(5013):1651–1656.
42. Victor E Velculescu, Lin Zhang , Bert Vogelstein, Kenneth W Kinzler. “Serial analysis of gene expression” *Science.* 1995;270 (5235):484–487.
43. Colleen D Hough , Cheryl A Sherman-Baust, Ellen S Pizer, F J Montz , Dwight D Im , Neil B Rosenshein , Kathleen R Cho , Gregory J Riggins , Patrice J Morin. “Large-scale serial analysis of gene expression reveals genes differentially expressed in ovarian cancer” *Cancer Researc.* 2000;60 (22):6281–6287.

44. Pawar S, Stanam A, Chaudhari M, Rayudu D. Effects of temperature on COVID-19 transmission. medRxiv. doi: <https://doi.org/10.1101/2020.03.29.20044461>
45. S Brenner, M Johnson, J Bridgham, G Golda, DH Lloyd, D Johnson, S Luo, S McCurdy, M Foy, M Ewan, R Roth, D George, S Eletr, G Albrecht, E Vermaas, S R Williams, K Moon, T Burcham, M Pallas, RB DuBridg, J Kirchner, K Fearon, J Mao, K Corcoran. "Gene expression analysis by massively parallel signature sequencing (mpss) on microbead arrays" *Nature Biotechnology*. 2000;18 (6):630 – 634.
46. Pawar, S.; Lahiri, C. Quorum sensing: An imperative longevity weapon in Bacteria. *Afr J Microbiol Res* 2018, 12, 96-104. DOI: 10.5897/AJMR2017.8751.
47. Deana Erdner, Donald Anderson. "Global transcriptional profiling of the toxic dinoflagellate alexandrium fundyense using massively parallel signature sequencing" *BMC Genomics*. 2006;7(1):88.
48. Rachael Natrajan, Alan Mackay, Maryou B Lambros, Britta Weigelt, Paul M Wilkerson, Elodie Manie, Anita Grigoriadis, Roger A'Hern, Petra van der Groep, Iwanka Kozarewa, Tatiana Popova, Odette Mariani, Samra Turajlic, Simon J Furney, Richard Marais, Daniel-Nava Rodrigues, Adriana C Flora, Patty Wai, Vidya Pawar, Simon McDade, Jason Carroll, Dominique Stoppa-Lyonnet, Andrew R Green, Ian O Ellis, Charles Swanton, Paul van Diest, Olivier Delattre, Christopher J Lord, William D Foulkes, Anne Vincent-Salomon, Alan Ashworth, Marc Henri Stern, Jorge S Reis-Filho. "A whole-genome massively parallel sequencing analysis of brca1 mutant oestrogen receptor-negative and -positive breast cancers" *The Journal of Pathology*. 2012;227 (1):29–41.
49. Zhong Wang, Mark Gerstein, Michael Snyder. "Rna-seq: a revolutionary tool for transcriptomics" *Nature Reviews. Genetics*. 2009;10(1):57–63.
50. Ugrappa Nagalakshmi, Zhong Wang, Karl Waern, Chong Shou, Debasish Raha, Mark Gerstein, Michael Snyder. "The transcriptional landscape of the yeast genome defined by rna sequencing" *Science*. 2008;320 (5881):1344–1349.
51. Brian T Wilhelm, Samuel Marguerat, Stephen Watt, Falk Schubert, Valerie Wood, Ian Goodhead, Christopher J Penkett, Jane Rogers, Jürg B hler. "Dynamic repertoire of a eukaryotic transcriptome surveyed at single-nucleotide resolution" *Nature*. 2008;453 (7199):1239–1243.
52. Ali Mortazavi, Brian A Williams, Kenneth McCue, Lorian Schaeffer, Barbara Wold. "Mapping and quantifying mammalian transcriptomes by rna-seq" *Nature Methods*. 2008;5 (7):621–628.
53. Pawar, S., Ashraf, M. I., Mehata, K. M., and Lahiri, C. (2017). "Computational identification of indispensable virulent proteins of Salmonella Typhi CT18," in *Current Topics in Salmonella and Salmonellosis*, ed M. Mares (InTech Publishers), 21–39.
54. Ryan Lister, Ronan C O'Malley, Julian Tonti-Filippini, Brian D Gregory, Charles C Berry, A Harvey Millar, Joseph R Ecker. "Highly integrated single-base resolution maps of the epigenome in Arabidopsis" *Cell*. 2008;133(3):523–536.
55. Nicole Cloonan, Alistair R R Forrest, Gabriel Kolle, Brooke B A Gardiner, Geoffrey J Faulkner, Mellissa K Brown, Darrin F Taylor, Anita L Steptoe, Shivangi Wani, Graeme Bethel, Alan J Robertson, Andrew C Perkins, Stephen J Bruce, Clarence C Lee, Swati S Ranade, Heather E Peckham, Jonathan M Manning, Kevin J McKernan, Sean M Grimmond. "Stem cell transcriptome profiling via massive-scale mrna sequencing" *Nature Methods*. 2008;5 (7):613–619.
56. Duan GL, Hu Y, Lui WJ, Kneer R, Zhao FJ, Zhu YG (2011) Evidence for a role of phytochelatin in regulating arsenic accumulation in rice grains. *Environ Exp Bot* 71:416–421. <https://doi.org/10.1016/j.envexpbot.2011.02.016>
57. Elbaz B, Shoshani-Knaani N, David-Assael O, Mizrachi-Dagri T, Mizrahi K, Saul H, Brook E, Berezin I, Shaul O (2006) High expression in leaves of the zinc hyperaccumulator Arabidopsis halleri of AhMHX, a homolog of an Arabidopsis thaliana vacuolar metal/proton exchanger. *Plant Cell Environ* 29:1179–1190
58. Enger MD, Tesmer JG, Travis GL, Barham SS (1986) Clonal variation of cadmium response in human-tumor cell-lines. *Am J Phys* 250:C256–C263
59. Feleafel MN, Mirdad ZM (2013) Hazard and effects of pollution by lead on vegetable crops. *J Agric Environ Ethic* 26:547–567. <https://doi.org/10.1007/s10806-012-9403-1>
60. Freisinger E (2011) Structural features specific to plant metallothioneins. *J Biol Inorg Chem* 16:1035–1045. <https://doi.org/10.1007/s00775-011-0801-z>
61. Führs H, Behrens C, Gallien S, Heintz D, Van Dorsselaer A, Braun HP, Horst WJ (2010) Physiological and proteomic characterization of manganese sensitivity and tolerance in rice (*Oryza sativa*) in comparison with barley (*Hordeum vulgare*). *Ann Bot* 105:1129–1140
62. Pawar, S., Ashraf, M. I., Mujawar, S., Mishra, R., and Lahiri, C. (2018). In silico identification of the indispensable quorum sensing proteins of multidrug resistant *Proteus mirabilis*. *Front. Cell. Infect. Microbiol.* 8:269. doi: 10.3389/fcimb.2018.00269
63. Chandrajit L, Pawar S, Sabarinathan R, Ashraf Md, Yamini C, Dipshikha C (2012). Identifying indispensable proteins of the type III secretion systems of *Salmonella enterica* serovar Typhimurium strain LT2. *BMC Bioinform.* 13:A10.
64. Fukao Y, Ferjani A, Tomioka R, Nagasaki N, Kurata R, Nishimori Y, Fujiwara M, Maeshima M (2011) iTRAQ analysis reveals mechanisms of growth defects due to excess zinc in Arabidopsis. *Plant Physiol* 155:1893–1907
65. Gabbrielli R, Pandolfini T, Espen L, Palandri MR (1999) Growth, peroxidase activity and cytological modifications in *Pisum sativum* seedlings exposed to Ni²⁺ toxicity. *J Plant Physiol* 155:639–645. [https://doi.org/10.1016/s0176-1617\(99\)80066-2](https://doi.org/10.1016/s0176-1617(99)80066-2)
66. Gallego SM, Pena LB, Barcia RA, Azpilicueta CE, Iannone MF, Rosales EP, Zawoznik MS, Groppa MD, Benavides MP (2012) Unravelling cadmium toxicity and tolerance in plants: insight into regulatory mechanisms. *Environ Exp Bot* 83:33–46. <https://doi.org/10.1016/j.envexpbot.2012.04.006>
67. Gao J, Sun L, Yang X, Liu JX (2013) Transcriptomic analysis of cadmium stress response in the heavy metal hyperaccumulator *Sedum alfredii*Hance. *PLoS One* 8:e64643

68. Pawar S, Davis CD, Rinehart CA (2011) Statistical analysis of microarray gene expression data from a mouse model of toxoplasmosis. *BMC Bioinform* 12(Suppl 7):A19
69. Gao J, Luo M, Zhu Y, He Y, Wang Q, Zhang C (2015) Transcriptome sequencing and differential gene expression analysis in *Viola yedoensis* Makino (Fam. Violaceae) responsive to cadmium (Cd) pollution. *Biochem Biophys Res Commun* 459:60–65
70. Gautam N, Verma PK, Verma S, Tripathi RD, Trivedi PK, Adhikari B (2012) Genome-wide identification of rice class I metallothionein gene: tissue expression patterns and induction in response to heavy metal stress. *Funct Integr Genomics* 12:635–647. <https://doi.org/10.1007/s10142-012-0297-98>
71. Gekeler W, Grill E, Winnacker EL, Zenk MH (1989) Survey of the plant kingdom for the ability to bind heavy metals through phytochelatin. *Z Naturforsch* 44:361–369
72. Gothberg A, Greger M, Holm K, Bengtsson BE (2004) Influence of nutrient levels on uptake and effects of mercury, cadmium, and lead in water spinach. *J Environ Qual* 33:1247–1255. <https://doi.org/10.2134/jeq2004.1247>
73. Mujawar, S., Mishra, R., Pawar, S., Gatherer, D., Lahiri, C.: Delineating the plausible molecular vaccine candidates and drug targets of multidrug-resistant *acinetobacter baumannii*. *Front. Cell. Infect. Microbiol.* 9, 203 (2019)
74. Pawar, S. D., Freas, C., Weber, I. T., & Harrison, R. W. (2018). Analysis of drug resistance in HIV protease. *BMC Bioinformatics*, 19, 362. <https://doi.org/10.1186/s12859-018-2331-y>.
75. John C Marioni , Christopher E Mason , Shrikant M Mane , Matthew Stephens, Yoav Gilad. "Rna-seq: An assessment of technical reproducibility and comparison with gene expression arrays" *Genome Research*. 2008;18(9):1509–1517.
76. Nathan Blow. "Transcriptomics: The digital generation". *Nature*. 2009;458 (7235):239–242.
77. Nicole C Roy, Eric Altermann , Zaneta A Park , Warren C McNabb. "A comparison of analog and next-generation transcriptomic tools for mammalian studies" *Briefings in Functional Genomics*. 2011;10 (3):135–150.
78. Kristin Schirmer, Beat B Fischer , Danielle J Madureira , Smitha Pillai. "Transcriptomics in ecotoxicology" *Analytical and Bioanalytical Chemistry*. 2010;397 (3):917–923.
79. Henriques A, Gonzalez De Aguilar JL. "Can transcriptomics cut the gordian knot of amyotrophic lateral sclerosis?" *Current Genomics*. 2011;12 (7):506–515.
80. Mittal K, Choi DH, Klimov S, Pawar S, Kaur R, Mitra A, Gupta MV, Sams R, Cantuaria G, Rida PCG, Aneja R (2016b) Evaluation of centrosome clustering protein KIF1C as a potential prognostic biomarker in serous ovarian adenocarcinomas. *J Clin Oncol* 34(15_suppl):e17083–e17083
81. Pawar S. (2019) Web-Based Application for Accurately Classifying Cancer Type from Microarray Gene Expression Data Using a Support Vector Machine (SVM) Learning Algorithm. In: Rojas L, Valenzuela O., Rojas F., Ortuño F. (eds) *Bioinformatics and Biomedical Engineering. IWBBIO 2019. Lecture Notes in Computer Science*, vol 11466. Springer, Cham. https://doi.org/10.1007/978-3-030-17935-9_14
82. Zhan Zhou, Jianying Gu, Yi-Ling Du, Yong-Quan Li, Yufeng Wang. "The -omics era- toward a systems-level understanding of streptomyces" *Current Genomics*. 2011;12 (6):404–416.
83. S Michael Rothenberg, Jeff Settleman. "Discovering tumor suppressor genes through genome-wide copy number analysis" *Current Genomics*. 2010;11(5):297–310.
84. Thierry Rabilloud, Cécile Lelong. "Two-dimensional gel electrophoresis in proteomics: A tutorial". *Journal of Proteomics*. 2011;74 (10):1829–1841.
85. Eleonora Piruzian, Sergey Bruskin, Alex Ishkin, Rustam Abdeev, Sergey Moshkovskii, Stanislav Melnik, Yuri Nikolsky, Tatiana Nikolskaya. "emphIntegrated network analysis of transcriptomic and proteomic data in psoriasis" *BMC Systems Biology* . 2010;4(41)
86. Dov Greenbaum, Ronald Jansen, Mark Gerstein. "Analysis of mRNA expression and protein abundance data: an approach for the comparison of the enrichment of features in the cellular population of proteins and transcripts" *Bioinformatics* . 2002;18 (4):585–596.
87. Emmanuelle Com, Eric Boitier, Jean-Pierre Marchandea, Arnd Brandenburg, Susanne Schroeder, Dana Hoffmann, Angela Mally, Jean-Charles Gautier. "Integrated transcriptomic and proteomic evaluation of gentamicin nephrotoxicity in rats" *Toxicology and Applied Pharmacology*. 2012;258 (1):124–133.
88. S. Pawar, A. Stanam and Y. Zhu, "Evaluating the computing efficiencies (specificity and sensitivity) of graphics processing unit (GPU)-accelerated DNA sequence alignment tools against central processing unit (CPU) alignment tool", *J. Bioinf. Sequence Anal.*, vol. 9, no. 2, pp. 10-14, 2018.
89. Jun X Yan , Angelica T Devenish , Robin Wait, Tim Stone, Steve Lewis, Sue Fowler. "Fluorescence two-dimensional difference gel electrophoresis and mass spectrometry based proteomic analysis of escherichia coli" *PROTEOMICS*. 2002;2 (12):1682–1698.
90. Rita Marouga, Stephen David, Edward Hawkins. "The development of the dige system: 2d fluorescence difference gel analysis technology". *Analytical and Bioanalytical Chemistry*. 2005;382:669–678.
91. Julien Franck, Karim Arafah, Mohamed Elayed, David Bonnel, Daniele Vergara, Amélie Jacquet, Denis Vinatier, Maxence Wisztorski, Robert Day, Isabelle Fournier, Michel Salzet. "Maldi imaging mass spectrometry" *Molecular and Cellular Proteomic*. 2009;8(9):2023–2033.
92. M Reid Groseclose, Pierre P Massion, Pierre Chaurand, Richard M Caprioli. "High-throughput proteomic analysis of formalin-fixed paraffin-embedded tissue microarrays using maldi imaging mass spectrometry" *PROTEOMICS* . 2008;8(18):3715–3724.
93. Mareike Elsner, Sandra Rauser, Stefan Maier, Cédrik Sch ne, Benjamin Balluff, Stephan Meding, Gerhard Jung, Martin Nipp, Hakan Sarioglu, Giuseppina Maccarrone, Michaela Aichler, Annette Feuchtinger, Rupert Langer, Uta Jütting, Marcus Feith,

- Bernhard Küster, Marius Ueffing, Horst Zitzelsberger, Heinz Höfler, Axel Walch. "Maldi imaging mass spectrometry reveals *cox7a2*, *tagln2* and *s100-a10* as novel prognostic markers in barrett's adenocarcinoma" *Journal of Proteomics*. 2012
94. Guihua Yue, Quanzhou Luo, Jian Zhang, Shiaw-Lin Wu, Barry L Karger. "Ultratrace lc/ms proteomic analysis using 10- μ m-i porous layer open tubular poly(styrene-divinylbenzene) capillary columns". *Analytical Chemistry*. 2007;79 (3):938–946.
95. Dwayne A Elias , Matthew E Monroe , Matthew J Marshall , Margaret F Romine , Alexander S Belieav , James K Fredrickson , Gordon A Anderson , Richard D Smith , Mary Lipton S. "Global detection and characterization of hypothetical proteins in *shewanella oneidensis* mr-1 using lc-ms based proteomics" *PROTEOMICS* . 2005;5 (12):3120–3130.
96. Lei Nie, Gang Wu, Fred J Brockman, Weiwen Zhang. "Integrated analysis of transcriptomic and proteomic data of *desulfovibrio vulgaris*: zero-inflated poisson regression models to predict abundance of undetected proteins" *Bioinformatics* . 2006;22 (13):1641–1647.
97. Ravindra Varma Polisetty, Poonam Gautam, Rakesh Sharma, H C Harsha, Sudha C Nair, Manoj >Kumar Gupta, Megha S Uppin , Sundaram Challa, Aneel Kumar Puligopu, Praveen Ankathi, Aniruddh K Purohit, Giriraj R Chandak , Akhilesh Pandey, Ravi Sirdeshmukh. "Lc-ms/ms analysis of differentially expressed glioblastoma membrane proteome reveals altered calcium signalling and other protein groups of regulatory functions" *Molecular and Cellular Proteomics*. 2012
98. Nathanael Delmotte, Christian H Ahrens, Claudia Knief, Ermir Qeli, Marion Koch, Hans-Martin Fischer , Julia A Vorholt, Hauke Hennecke, Gabriella Pessi. "An integrated proteomics and transcriptomics reference data set provides new insights into the *Bradyrhizobium japonicum* bacteroid metabolism in soybean root nodules" *Proteomics* . 2010;10 :1391–1400.
99. Shrikant Pawar, Tuck Onn Liew, Aditya Stanam, Chandrajit Lahiri. Common cancer biomarkers of breast and ovarian types identified through artificial intelligence. *Chemical Biology and Drug Design*, 995-1004, <https://doi.org/10.1111/cbdd.13672>
100. Lahiri, Chandrajit * and Pawar, Shrikant and Mishra, Rohit (2019) Precision medicine and future of cancer treatment. *Precision Cancer Medicine*. ISSN 2617-2216
101. Pawar, S., Stanam, A. A Six-Gene-Based Prognostic Model Predicts Survival in Head and Neck Squamous Cell Carcinoma Patients. *J. Maxillofac. Oral Surg.* 18, 320–327 (2019). <https://doi.org/10.1007/s12663-019-01187-z>
102. Lau Sennels, Mogjiborahman Salek, Lee Lomas, Egisto Boschetti, Pier Giorgio Righetti, Juri Rappsilber. "Pro-teomic analysis of human blood serum using peptide library beads" *Journal of Proteome Research* . 2007; 6 (10):4055–4062.
103. Leann M Mikesh, Beatrix Ueberheide, An Chi , Joshua J Coon , John EP, Syka Jeffrey Shabanowitz, Donald F Hunt . "The utility of etd mass spectrometry in proteomic analysis" *Biochimica et Biophysica Acta (BBA) - Proteins and; Proteomics*. 2006;1764 (12):1811–1822.
104. Henrik Molina, David M Horn, Ning Tang , Suresh Mathivanan, Akhilesh Pandey. "Global proteomic profiling of phosphopeptides using electron transfer dissociation tandem mass spectrometry" *Proceedings of the National Academy of Sciences*. 2007;104 (7):2199–2204.
105. Danielle L Swaney , Craig D Wenger , James A Thomson, Joshua J Coon. "Human embryonic stem cell phosphoproteome revealed by electron transfer dissociation tandem mass spectrometry" *Proceedings of the National Academy of Sciences*. 2009;106 (4):995–1000.
106. Brett Spurrier, Sundhar Ramalingam, Satoshi Nishizuka. "Reverse-phase protein lysate microarrays for cell signaling analysis" *Nat. Protocols* . 2008;3 (11):1796–1808.
107. Satoshi Nishizuka, Lu Charboneau, Lynn Young, Sylvia Major, William C Reinhold, Mark Waltham, Hosein Kouros-Mehr, Kimberly J Bussey, Jae K Lee, Virginia Espina, Peter J Munson, Emanuel Petricoin, Lance A Liotta, John N Weinstein . "Proteomic profiling of the nci-60 cancer cell lines using new high-density reverse-phase lysate microarrays" *Proceedings of the National Academy of Sciences*. 2003;100(24):14229–14234.
108. Stacy M Cowherd , Virginia A Espina , Emanuel F Petricoin, III, Lance A Liotta. "Proteomic analysis of human breast cancer tissue with laser-capture microdissection and reverse-phase protein microarrays" *Clinical Breast Cancer* . 2004;5 (5):385–392.
109. Y Baskin, T Yigitbasi. "Clinical proteomics of breast cancer" *Current Genomics*. 2010;11 (7):528–536.
110. Pawar, S., Yao, X. & Lu, C. Spermine and oxacillin stress response on the cell wall synthesis and the global gene expression analysis in Methicillin-resistance *Staphylococcus aureus*. *Genes Genom* 41, 43–59 (2019). <https://doi.org/10.1007/s13258-018-0735-8>
111. Shine J, Dalgarno L. "The 3'-terminal sequence of *escherichia coli* 16s ribosomal rna: complementarity to nonsense triplets and ribosome binding sites" *Proc Natl Acad Sci U S A* . 1974;71 :1342–1346.
112. Shine J, Dalgarno L. "Determinant of cistron specificity in bacterial ribosomes" *Nature* . 1975;254:34–38.
113. Alistair HA Bingham, Sreenivasan Ponnambalam , Bernard Chan, Stephen Busby. "Mutations that reduce expression from the p2 promoter of the *escherichia coli* galactose operon" *Gene* . 1986;41 (1):67–74.
114. Grossman AD, Zhou YN, Gross C, Heilig J, Christie GE, Calendar R. "Mutations in the *rpoh* (*htpr*) gene of *escherichia coli* k-12 phenotypically suppress a temperature-sensitive mutant defective in the sigma 70 subunit of rna polymerase" *J Bacteriol*. 1985;161(3):939–943.
115. Claes Gustafsson, Sridhar Govindarajan, Jeremy Minshull. "Codon bias and heterologous protein expression" *Trends in Biotechnology* . 2004;22 (7):346–353.
116. Pawar S., Stanam A., Lahiri C. (2020) Clustering Reveals Common Check-Point and Growth Factor Receptor Genes Expressed in Six Different Cancer Types. In: Rojas I, Valenzuela O., Rojas F., Herrera L., Ortuño F. (eds) *Bioinformatics and Biomedical Engineering. IWBBIO 2020. Lecture Notes in Computer Science*, vol 12108. Springer, Cham. https://doi.org/10.1007/978-3-030-45385-5_52

117. Paul M Sharp , Wen-Hsiung Li. "The codon adaptation index-a measure of directional synonymous codon usage bias, and its potential applications" *Nucleic Acids Research* . 1987;15 (3):1281–1295.
118. Gila Lithwick, Hanah Margalit. "Hierarchy of sequence-dependent features associated with prokaryotic translation" *Genome Research* . 2003;13(12):2665–2673.
119. Eldad N, Arava Y. "A ribosomal density-mapping procedure to explore ribosome positions along translating mrnas" *Methods Mol Biol* . 2008;419 :231–242.
120. Nicholas T Ingolia , Sina Ghaemmaghami, John R S Newman, Jonathan S Weissman. "Genome-wide analysis in vivo of translation with nucleotide resolution using ribosome profiling" *Science*. 2009;324 (5924):218–223.
121. Dov Greenbaum, Christopher Colangelo, Kenneth Williams, Mark Gerstein. "Comparing protein abundance and mRNA expression levels on a genomic scale" *Genome Biology*. 2003;4(9):117+.
122. Raymond J Cho , Michael J Campbell , Elizabeth A Winzeler , Lars Steinmetz, Andrew Conway , Lisa Wodicka , Tyra G Wolfsberg , Andrei E Gabrielian , David Landsman, David J Lockhart , Ronald W Davis. "A genome-wide transcriptional analysis of the mitotic cell cycle" *Molecular cell* . 1998;2
123. James L Hargrove, Frederick Schmidt H. "The role of mRNA and protein stability in gene expression" *The FASEB Journal*. 1989;3 :2360–2370.
124. Bjorn Schwanhaussner, Dorothea Busse, Na Li, Gunnar Dittmar, Johannes Schuchhardt, Jana Wolf, Wei Chen, Matthias Selbach. "Global quantification of mammalian gene expression control" *Nature* . 2011;473 :337–342.
125. Andreas Bachmair, Daniel Finley, Alexander Varshavsky. "In vivo half-life of a protein is a function of its amino-terminal residue" *Science* . 1986;234 :179–186.
126. Pawar, S., & Stanam, A. (2018). Software effort prediction with algorithm based frameworks. *International Journal of Engineering and Computer Science*, 7(09), 24206–24213. Retrieved from <http://103.53.42.157/index.php/ijecs/article/view/4174>
127. Tobias Maier, Marc Güell, Luis Serrano. "Correlation of mrna and protein in complex biological samples" *FEBS Letters* . 2009;583 (24):3966–3973.
128. Susan B Altenbach , William H Vensel , Frances M DuPont. "Integration of transcriptomic and proteomic data from a single wheat cultivar provides new tools for understanding the roles of individual alpha gliadin proteins in flour quality and celiac disease" *Journal of Cereal Science* . 2010;52 (2):143–151.
129. shrikant pawar, Aditya Stanam. Predicting the prognosis for cancer patients with interleukins gene expression level [abstract]. In: *Proceedings of the American Association for Cancer Research Annual Meeting 2019*; 2019 Mar 29-Apr 3; Atlanta, GA. Philadelphia (PA): AACR; *Cancer Res* 2019;79(13 Suppl):Abstract nr 4247.
130. J P McRedmond , S D Park , D F Reilly, J A Coppinger , P B Maguire , D C Shields, D J Fitzgerald. "Integration of proteomics and genomics in platelets" *Molecular and Cellular Proteomics* . 2004; 3 (2):133–144.
131. Paul Perco, Irmgard Muhlberger, Gert Mayer, Rainer Oberbauer, Arno Lukas, Bernd Mayer. "Linking transcriptomic and proteomic data on the level of protein interaction network" *Electrophoresis* . 2010;31 :1780–789.
132. Marcin Imielinski, Sangwon Cha, Tomas Rejtar, Elizabeth A Richardson, Barry L Karger , Dennis C Sgroi. "Integrated proteomic, transcriptomic, and biological network analysis of breast carcinoma reveals molecular features of tumorigenesis and clinical relapse" *Molecular and Cellular Proteomics*. 2012
133. Wandaliz Torres-García, Weiwen Zhang, George C Runger , Roger H Johnson, Deirdre R Meldrum. "Integrative analysis of transcriptomic and proteomic data of *desulfovibrio vulgaris*: a non-linear model to predict abundance of undetected proteins" *Bioinformatics* . 2009;25 (15):1905–1914.
134. Jerome H Friedman. "Stochastic gradient boosting" *Computational Statistics and Data Analysis*. 2002;38 (4):367–378.
135. Wandaliz Torres-Garcia, Steven D Brown , Roger H Johnson , Weiwen Zhang, George C Runger, Deirdre R Meldrum. "Integrative analysis of transcriptomic and proteomic data of *shewanella oneidensis*: missing value imputation using temporal datasets" *Mol. BioSyst* . 2011;7 (7):1093–1104.
136. Feng Li, Lei Nie, Gang Wu, Jianjun Qiao, Weiwen Zhang. "Prediction and characterization of missing proteomic data in *desulfovibrio vulgaris*" *Comparative and Functional Genomics* . 2011;2011
137. Viehweger K (2014) How plants cope with heavy metals. *Bot Stud* 55:1–12
138. Walliwalagedara C, van Keulen H, Willard B, Wei R (2012) Differential proteome analysis of *Chlamydomonas reinhardtii* response to arsenic exposure. *Am J Plant Sci* 3:764–772
139. Pawar S., Stanam A. (2019) Developing a DEVS-JAVA Model to Simulate and Pre-test Changes to Emergency Care Delivery in a Safe and Efficient Manner. In: Rojas I., Valenzuela O., Rojas F., Ortuño F. (eds) *Bioinformatics and Biomedical Engineering. IWBBIO 2019. Lecture Notes in Computer Science*, vol 11466. Springer, Cham. https://doi.org/10.1007/978-3-030-17935-9_1
140. Wang HC, Wu JS, Chia JC, Yang CC, Wu YJ, Juang RH (2009) Phytochelatin synthase is regulated by protein phosphorylation at a threonine residue near its catalytic site. *J Agric Food Chem* 57:7348–7355. <https://doi.org/10.1021/jf9020152>
141. Wang Y, Hu H, Zhu LY, Li XX (2012) Response to nickel in the proteome of the metal accumulator plant *Brassica juncea*. *J Plant Interact* 7:230–237
142. Wang R, Gao F, Guo BG, Huang JC, Wang L, Zhou YJ (2013) Short-term chromium-stress-induced alterations in the maize leaf proteome. *Int J Mol Sci* 14:11125–11144
143. Wang Y, Xu L, Shen H, Wang J, Liu W, Zhu X, Wang R, Sun X, Liu L (2015) Metabolomic analysis with GC-MS to reveal potential metabolites and biological pathways involved in Pb and Cd stress response of radish roots. *Sci Rep* 5:18296
144. Witters N, Van Slycken S, Meers E, Adriaenssens K, Meiresonne L, Tack FMG, Vangronsveld J, Thewys T (2009) Short-rotation coppice of willow for phytoremediation of a metal-contaminated agricultural area: a sustainability assessment. *Bioenergy Res* 2:144–152

145. Wray GA, Hahn MW, Abouheif E, Balhoff JP, Pizer M, Rockman MV, Romano LA (2003) The evolution of transcriptional regulation in eukaryotes. *Mol Biol Evol* 20:1377–1419
146. Pawar, S., & Batra, H. (2018). Online electronic laboratory notebook: A secured cloud storage system scripted in Hypertext Pre-processor (PHP) programming language. *Journal of Engineering and Technology Research*, 10(1), 1-6.
147. Wu H, Chen C, Du J, Liu H, Cui Y, Zhang Y, He Y, Wang Y, Chu C, Feng Z, Li J, Ling HQ (2012) Co-overexpression FIT with AtbHLH38 or AtbHLH39 in Arabidopsis-enhanced cadmium tolerance via increased cadmium sequestration in roots and improved iron homeostasis of shoots. *Plant Physiol* 158:790–800. <https://doi.org/10.1104/pp.111.190983>
148. Wu CS, Chen DY, Chang CF, Li MJ, Hung KY, Chen LJ, Chen PW (2014) The promoter and the 50-untranslated region of rice metallothionein OsMT2b gene are capable of directing high-level gene expression in germinated rice embryos. *Plant Cell Rep* 33:793–806
149. Xu J, Zhu Y, Ge Q, Li Y, Sun J, Zhang Y, Liu X (2012) Comparative physiological responses of *Solanum nigrum* and *Solanum torvum* to cadmium stress. *New Phytol* 196:125–138
150. Xu L, Wang Y, Liu W, Wang J, Zhu X, Zhang K, Yu R, Wang R, Xie Y, Zhang W, Gong Y, Liu L (2015) De novo sequencing of root transcriptome reveals complex cadmium-responsive regulatory networks in radish (*Raphanus sativus* L.). *Plant Sci* 236:313–323
151. Yoshihara T, Hodoshima H, Miyano Y, Shoji K, Shimada H, Goto F (2006) Cadmium inducible Fe deficiency responses observed from macro and molecular views in tobacco plants. *Plant Cell Rep* 25:365–373. <https://doi.org/10.1007/s00299-005-0092-3>
152. Yu L-J, Luo Y-F, Liao B, Xie L-J, Chen L, Xiao S, Li JT, Hu S, Shu W-S (2012) Comparative transcriptome analysis of transporters, phytohormone and lipid metabolism pathways in response to arsenic stress in rice (*Oryza sativa*). *New Phytol* 195:97–112