

## Article

# Common Factors in Neurodegeneration:

## A Meta-Study revealing Shared Patterns on a Multi-Omics Scale

Nicolas Ruffini <sup>1§</sup>, Susanne Klingenberg<sup>1§</sup> Susann Schweiger<sup>1</sup> and Susanne Gerber <sup>1,\*</sup>

<sup>1</sup> Institute for Human Genetics, University Medical Center, Johannes Gutenberg University, Mainz, Germany

\* Correspondence: sugerber@uni-mainz.de;

§ Both authors contributed equally

**Abstract:** Neurodegenerative diseases such as Alzheimer's disease, Parkinson's disease, Huntington's disease, and Amyotrophic Lateral Sclerosis are heterogeneous, progressive diseases with frequently overlapping symptoms characterized by a loss of neurons. Studies suggested relations between neurodegenerative diseases for many years, e.g., regarding the aggregation of toxic proteins or triggering endogenous cell death pathways. Within this study, publicly available genomic, transcriptomic and proteomic data were gathered from 188 studies and more than one million patients to detect shared genetic patterns between the neurodegenerative diseases and the analyzed omics-layers within conditions. The results show a remarkably high number of shared genes between the transcriptomic and proteomic levels for all diseases while showing a significant relation between genomic and proteomic data only in some cases. A set of 139 genes was found to be differentially expressed in several transcriptomic experiments of all four diseases. These 139 genes showed overrepresented GO-Terms and pathways mainly involved in stress response, cell development, cell adhesion, and the cytoskeleton. Furthermore, the overlap of two and three omics-layers per disease were used to search for overrepresented pathways and GO-Terms. Taken together, we could confirm the existence of many relations between Alzheimer's disease, Parkinson's disease, Huntington's disease, and Amyotrophic Lateral Sclerosis on the transcriptomic and proteomic level by analyzing the pathways and GO-Terms arising in these intersections. The significance of the connection between the transcriptomic and proteomic data for all four analyzed neurodegenerative diseases showed that exploring these omics-layers simultaneously holds new insights that do not emerge from analyzing these omics-layers separately. Our data therefore suggests addressing human patients with neurodegenerative diseases as complex biological systems by integrating multiple underlying data sources.

**Keywords:** Multi-Omics, Alzheimer's Disease, Parkinson's disease, Huntington's disease, Amyotrophic Lateral Sclerosis, Neurodegeneration)

### 1. Introduction

Neurodegenerative diseases (NDD), including Alzheimer's disease, Parkinson's disease, Amyotrophic Lateral Sclerosis, and Huntington's disease, are heterogeneous, progressive

diseases characterized by a loss of neurons, accumulation of aggregated and misfolded proteins (Rubinsztein, 2006; Serrano-Pozo *et al.*, 2011; Hussain *et al.*, 2018; Esteves and Cardoso, 2020), cognitive decline and locomotive dysfunction (Xie *et al.*, 2014; Dugger and Dickson, 2017; Gan *et al.*, 2018). Despite decades of research and considerable progress in identifying risk genes, potent biomarkers, and environmental risk factors, this progression cannot be impeded. As details regarding the various (patho)physiological processes associated with Neurodegenerative Diseases (NDD) remain unclear, the diseases are still incurable.

It became generally accepted that the underlying mechanisms are polyfactorial and depend on the complex interplay of multiple (partly unknown) genetic and non-genetic variables (Mahalingam and Levy, 2014; Hinz and Geschwind, 2017; Ghasemi and Brown, 2018; Reed *et al.*, 2019; Bellenguez, Grenier-Boley and Lambert, 2020). Excessive immune response and inflammation (Kim, Seo and Suh, 2004; Heneka, Kummer and Latz, 2014; Doty, Guillot-Sestier and Town, 2015; Heckmann, Tummers and Green, 2019), misguided apoptosis and autophagy (Bredesen, Rao, and Mehlen, 2006; Guo *et al.*, 2018), dysfunction in mitochondria (Silva *et al.*, 2017; Briston and Hicks, 2018; Esteves *et al.*, 2018) and ion channels (Li and Lester, 2001; Kumar *et al.*, 2016) and various forms of cell stress have been recognized to play a major role in neurodegenerative processes (Jomova *et al.*, 2010; Ramanan and Saykin, 2013; Cabral-Miranda and Hetz, 2018).

By far the most prevalent of these diseases, **Alzheimer's disease (AD)**, is an inexorably progressive brain disorder that affects higher cognitive functions. Memory loss is the typical sign of the disease. Still, there is also a significant decline in other cognition domains like language, visual-spatial skills, practical skills, reasoning, and judgment capability. (Ballard *et al.*, 2011; Scheltens *et al.*, 2016; Joe and Ringman, 2019). The accumulation of abnormally folded extracellular  $\beta$ -amyloid (senile plaques) and intracellular phosphorylated tau (neurofibrillary tangles) proteins are the distinctive pathological hallmarks of the disease (Braak and Braak, 1991; Mandelkow and Mandelkow, 1998; Iqbal and Grundke-Iqbal, 2002). Plaques and tangles have been shown to interfere with calcium signaling and synaptic transmission (Li and Lester, 2001; Sheng, Sabatini and Südhof, 2012; Berridge, 2014; Perdigão *et al.*, 2020), to induce a persistent inflammatory response, and to lead to synapse loss and ultimately neuronal degeneration (Hashimoto *et al.*, 2003; Wyss-Coray, 2006; Kinney *et al.*, 2018). Inflammation caused by a central nervous system infection could be an additive factor in the pathogenesis (Itzhaki *et al.*, 2004; Maheshwari and Eslick, 2015). Changes primarily occur in the entorhinal cortex and hippocampus, and then spread in the frontal cortex, amygdala, basal forebrain, and brainstem (Rademakers and Rovelet-Lecrux, 2009; Halliday, 2017).

Parkinson's disease (PD) is the second most common neurodegenerative disorder, mainly affecting the motor system (Dauer and Przedborski, 2003; Wright Willis *et al.*, 2010). The aggregation of  $\alpha$ -synuclein into Lewy bodies and Lewy neurites, mainly in the substantia nigra pars compacta and the resulting loss of dopaminergic neurons leads to distinctive symptoms including resting tremors, bradykinesia, stooped posture, and in some cases, dementia (Berardelli *et al.*, 2001; Alexander, 2004; Jankovic, 2008). This classical view of disease origin has now been broadened to other non-motor related brain regions. There is evidence that areas like the autonomic and enteric nervous system (Knudsen and Borghammer, 2018), the olfactory bulb (Niu *et al.*, 2018), the medulla oblongata

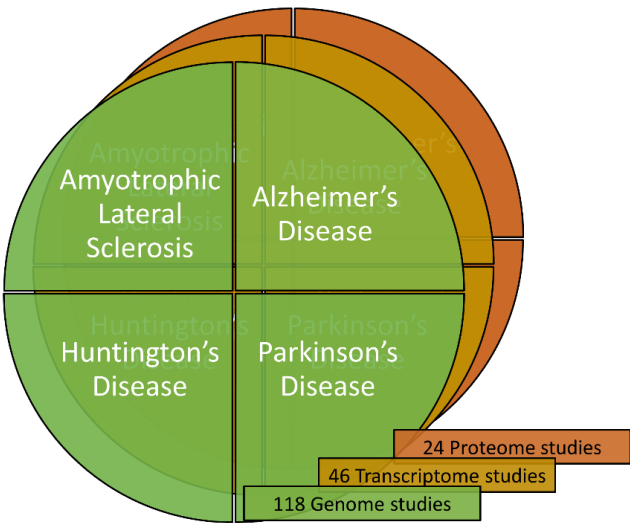
(Pyatigorskaya *et al.*, 2016), and pontine tegmentum may be affected by neurodegeneration even before the involvement of the substantia nigra. In contrast, the neocortex might be involved later throughout the disease (Novellino *et al.*, 2020). This observation is supported by some non-motor symptoms observed in patients with Morbus Parkinson, including olfactory loss, sleep disturbance, depression, autonomic dysfunction, and cognitive impairment (Braak *et al.*, 2003; Cerasa, Novellino, and Quattrone, 2016).

Huntington's disease (HD) is a progressive neurodegenerative disease that manifests pathologically with the significant loss of the striatum's GABAergic medium-sized spiny neurons (Ross and Tabrizi, 2011; Brandt, 2018). Symptoms appear in midlife and include chorea, cognitive decline, psychiatric disorders, and depression. The disease is fatal within 15 - 20 years after onset (Damiano *et al.*, 2010; McQuade *et al.*, 2014). While both, familial and sporadic forms of AD and PD exist, HD is an autosomal dominant neurodegenerative disease caused by the expansion of a CAG repeat in exon 1 of the huntingtin gene translating into a polyglutamine (polyQ) expansion in the N-terminus of the Huntingtin protein (Walker, 2007; Johnson and Davidson, 2010; Roos, 2010). The length of the repeat negatively correlates with the age of onset. More than 40 CAG repeats being fully penetrant with adult-onset while more than 70 repetitions generally result in juvenile-onset (Roos, 2010; Ross and Tabrizi, 2011; Pringsheim *et al.*, 2012). Physiologically, huntingtin seems to have multiple biological functions, including axonal and vesicular transport, endocytosis, post-synaptic signaling, and cell survival pathways (Harjes and Wanker, 2003; Cattaneo, Zuccato and Tartari, 2005). The mutant huntingtin gene is prone to cleavage. The resulting short fragments containing the N-terminal polyglutamine expansion oligomerize and form aggregates that have been implicated in neurotoxicity (Wanker *et al.*, 2019). Furthermore, dysregulation of several functions occur, including gene transcription (Cha, 2000; Roze *et al.*, 2008), axonal transport of critical factors (Gunawardena *et al.*, 2003), calcium and potassium signaling (Mackay, Nassrallah and Raymond, 2018; Zhang, Wan and Tong, 2018), protein-protein interactions, autophagy (Ochaba *et al.*, 2014; Martin *et al.*, 2015), and proteasomal and mitochondrial capacities (Damiano *et al.*, 2010; Reddy and Shirendeb, 2012; McQuade *et al.*, 2014). Even though the *huntingtin* gene is expressed ubiquitously, the striatum and cortical areas are the most affected regions as shown in postmortem histological evaluation (Ferrante *et al.*, 1987; Tang *et al.*, 2007; Ross and Tabrizi, 2011) or non-invasive brain magnetic resonance imaging (Aylward *et al.*, 2004; Kassubek *et al.*, 2004). The severity of striatal alterations is correlated with the degree of motor, cognitive, and psychiatric perturbations, suggesting that striatal degeneration is an essential aspect of HD physiopathology (Myers *et al.*, 1988). Pre-symptomatic patients (i.e., carrying the mutation but are still asymptomatic) already show significant atrophy of the caudate and putamen (Aylward *et al.*, 2004; Kassubek *et al.*, 2004). This suggests that degenerative events like cell shrinkage or loss begin years before the occurrence of clinical symptoms.

Amyotrophic Lateral Sclerosis (ALS) is a neurodegenerative disease affecting both the upper and lower motor neurons (Mitchell and Borasio, 2007; Wijesekera and Leigh, 2009; Kiernan *et al.*, 2011). It is characterized by progressive muscular paralysis reflecting the degeneration of motor neurons in the primary motor cortex, corticospinal tracts, brainstem, and spinal cord (Kang *et al.*, 2013; Philips and Rothstein, 2014; Novellino *et al.*, 2020). The mean age of onset for sporadic ALS is about 60 years. Approximately two-thirds of patients with typical ALS have a spinal form of the disease (limb onset) where muscle weakness starts either distally or proximally in the upper and lower limbs (Turner *et al.*, 2011;

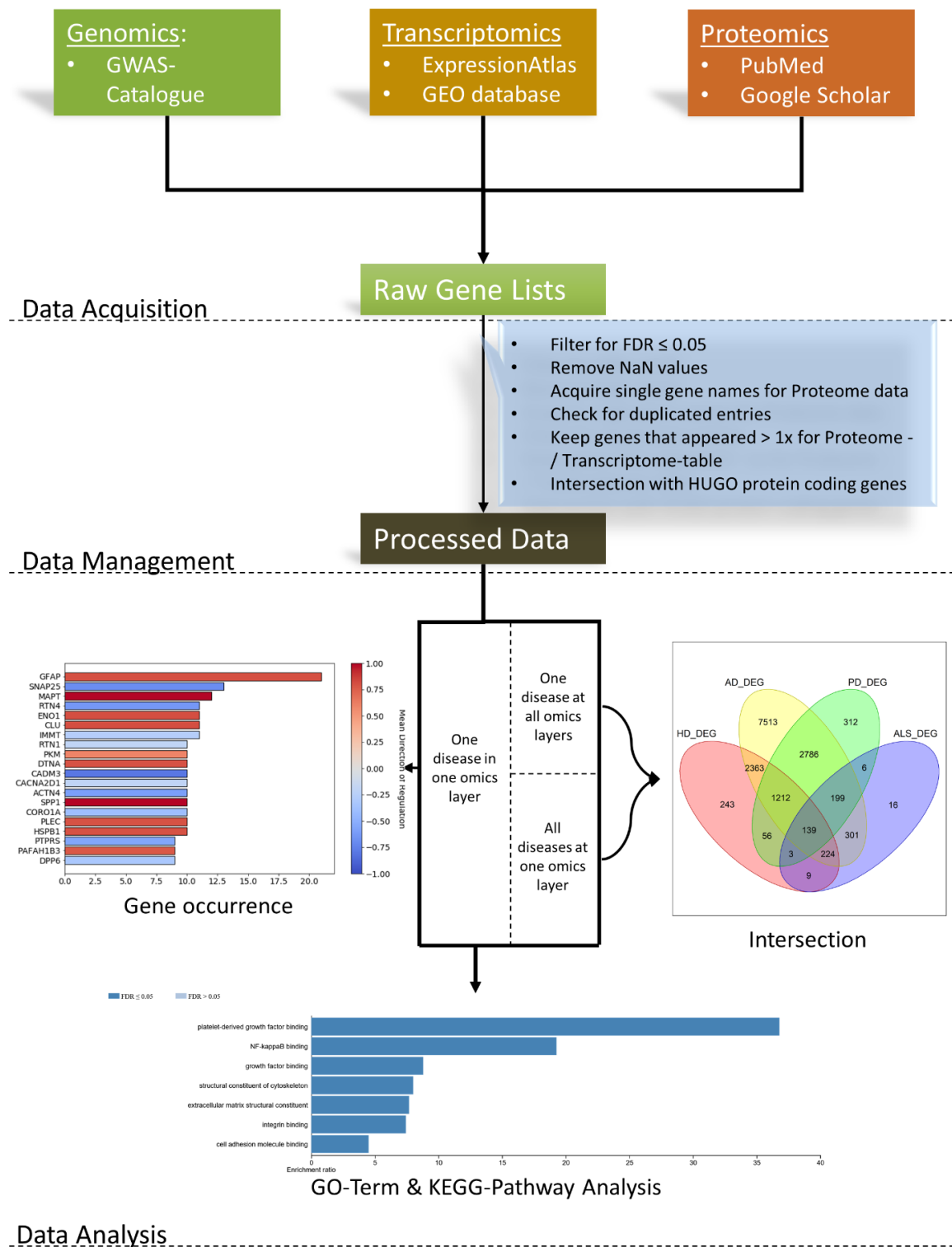
Robberecht and Philips, 2013). Patients with bulbar onset ALS usually suffer from dysarthria and dysphagia, and limb symptoms can develop almost simultaneously with bulbar symptoms (Kiernan *et al.*, 2011; Cistaro *et al.*, 2012; Guareschi *et al.*, 2012). Paralysis is progressive and leads to death due to respiratory failure within 2–5 years (Mitchell and Borasio, 2007; Kiernan *et al.*, 2011). Most ALS cases are sporadic, but 5–10% of cases are familial with mutations of the *SOD1* gene and the *TARDBP* gene (TDP-43) (Wijesekera and Leigh, 2009; Renton, Chiò and Traynor, 2014). The antioxidant enzyme SOD1 protects cells from the damaging effects of superoxide radicals. Deleterious mutations of *SOD1* lead to the accumulation of highly toxic hydroxyl radicals, which causes degradation of both nuclear and mitochondrial DNA and protein misfolding (Guareschi *et al.*, 2012; Kaur, McKeown and Rashid, 2016). The transactivating response DNA binding protein of 43 kD (TDP-43) is a highly conserved and ubiquitously expressed protein localized primarily within the nucleus. It is important in gene transcription and RNA splicing regulation (Lagier-Tourenne, Polymenidou, and Cleveland, 2010; Prasad *et al.*, 2019), but also regulates the activity of retrotransposons (Savage *et al.*, 2018; Tam *et al.*, 2019). It is predominantly nuclear, but shuttles in and out of the cytoplasm and along axons, where it can aggregate to hyperphosphorylated and ubiquitinated TDP-43 deposits both in the familial and the sporadic form of ALS (Prasad *et al.*, 2019).

Overlap of phenotypic traits of the NDDs described above suggests common pathogenic mechanisms underlying distinct NDD. Compared to studying individual diseases separately, identifying and analyzing the common dysfunctional proteins and dysregulated pathways of the diseases might elucidate the fundamental insights into their pathogenic process (Li, Nie and Yu, 2015). It was previously shown that there is no overlap between AD, PD, and ALS on genomic data and some shared pathways for AD, PD, ALS, and HD transcriptomic data (Arneson *et al.*, 2018), but proteomic data and the latest entries in the databases were not considered. Besides looking for overlapping genes between the different NDD or omics layers, we also analyzed whether this number is sufficiently high to claim a significant relationship between NDD or omics layers. An overview of the methodologic procedure is given in Figure 2. By investigating 188 studies (see Figure 1) in total, this study can detect a stable signal that arises in mainly late-stage NDD across tissues, methods, and omics-layers and, therefore, can help unravel underlying patterns across neurodegenerative diseases.



**Figure 1** Overview of the analyzed NDD and omics-layers. Data from 188 studies, including more than 1 million human samples, were gathered to find common patterns of genes, pathways, and GO-terms connecting AD, PD, ALS, and HD. The area of each disease's fraction in the pie chart is just visualization and does not represent the correct number of studies per disease and omics-layer. For the absolute numbers per disease and omics-layer see

and **Table 2**.



**Figure 2** Workflow Overview: Data Acquisition was performed using the GWAS-catalog for genomic data, the EMBL-EBI Expression Atlas, the GEO database for transcriptomic data, and manual PubMed and Google Scholar research for proteomic data. After filtering these raw data tables and applying some data transformation, the processed data were used for the data analysis. For every omics-layer in each disease, Gene occurrence bar plots were created for the 20 most abundant genes. For detecting overlapping gene names on multiple omics-layers, stacked bar plots were created for every disease, showing the average direction of regulation for the transcriptomic and proteomic layer. The intersections were visualized as Venn diagrams of all combinations on the disease and the omics-layer level. Finally, each set of genes after the intersections was used for KEGG pathway and GO-Term analyses.

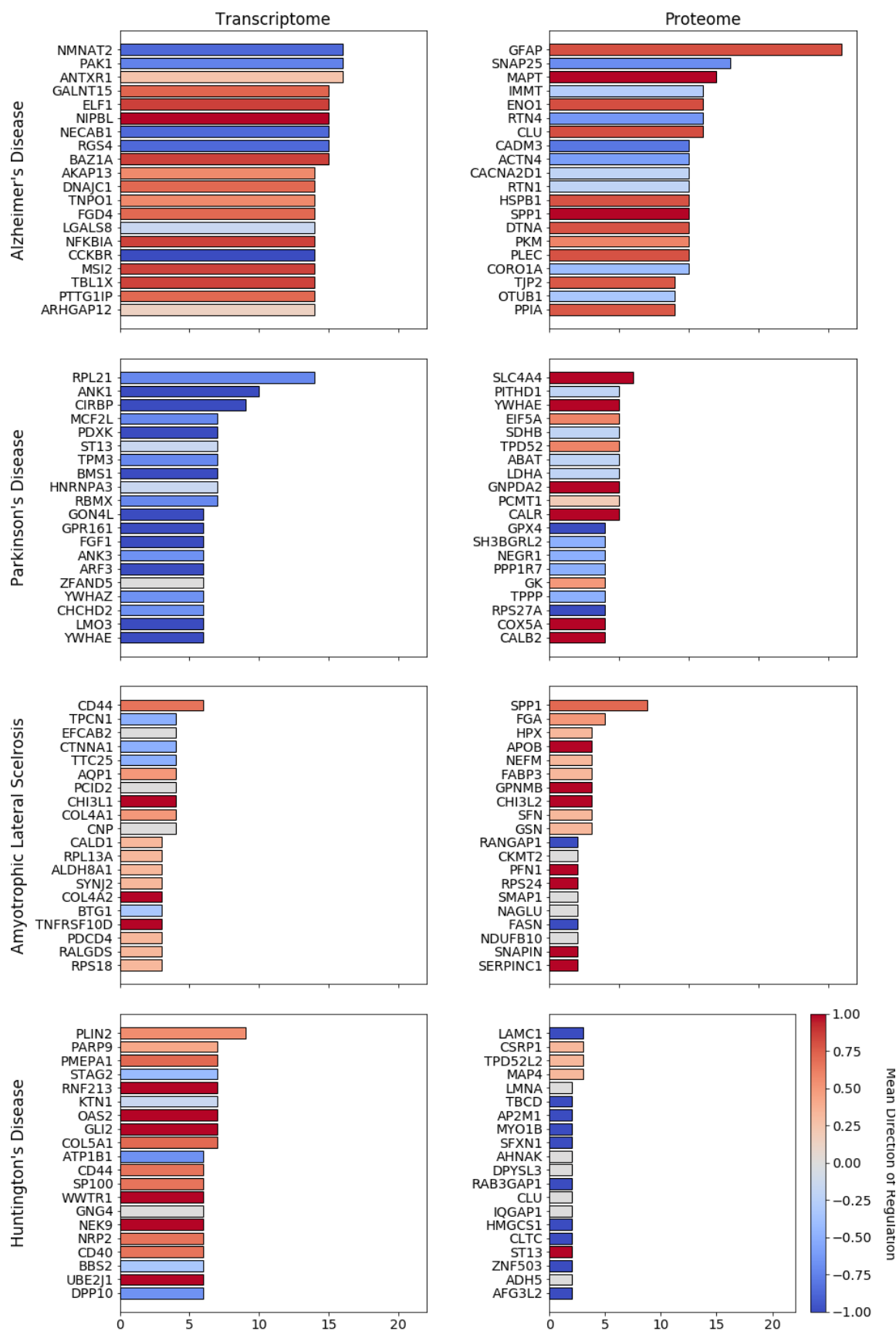


## Results

**Gene Occurrence:** The 20 most abundant genes per disease for the transcriptomic and proteomic data are shown in colored bar plots representing the abundance and mean direction of regulation (see **Equation 1**) of each gene (see Figure 3). The overall most abundant gene was *GFAP* in the proteomic AD data. Most of the twenty most abundant AD genes appeared more than ten times, whereas none of HD and ALS genes did appear at least ten times. PD data showed some highly abundant genes in the transcriptomic data – *ANK1* appeared ten times, *RPL21* appeared 14 times. Interestingly, none of the twenty most abundant genes in the PD transcriptomic data showed a positive mean direction of regulation. The most abundant genes for the other diseases and omics-layers showed more heterogeneous mean directions of regulation.

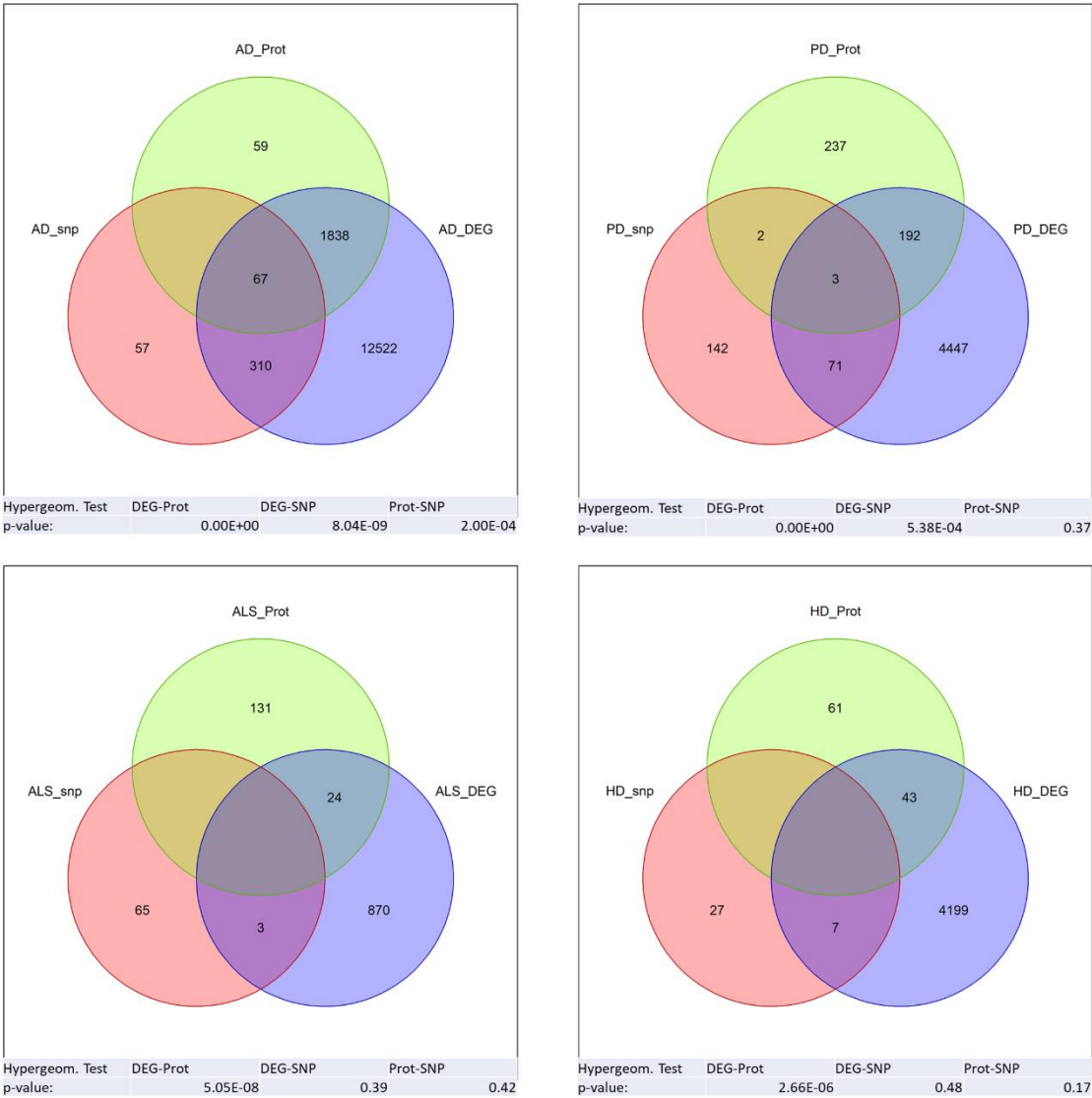
**Intersection:** To quantify, if the number of overlapping genes is high between omics-layers for one disease and between diseases for one omics-layer, a hypergeometric test was performed. Analyzing the number of overlapping genes between transcriptomic and proteomic data showed a significant number of overlapping genes for all four diseases (see Figure 4**Error! Reference source not found.****Error! Reference source not found.**). The number of overlapping genes between the genomic level and the other two omics-layers was not significantly enriched for ALS and HD, and no single gene occurred in the intersection between the genomic and the proteomic level for these diseases. For AD, all pairwise intersections between these three omics-layers showed a significantly enriched number of overlapping genes. For PD, the number of overlapping genes between genomic and transcriptomic data was significantly enriched, but no significant overlap could be seen for the proteomic – genomic overlap. AD showed the most considerable overlap between all three omics-layers with 67 genes. For PD, three genes were found in the intersection between all three analyzed omics-layers. The number of genes found in the GWAS catalog was the highest for AD with 434 single nucleotide polymorphisms (SNPs). For PD, 218 SNPs were found; 68 for ALS and 34 SNPs for HD. The number of overlapping SNPs between each pair of diseases ranged from zero to eleven and was significantly high for the pairwise overlaps between AD and PD as well as AD and ALS in a hypergeometric test (see Figure 5).

For the transcriptomic data, AD again showed the highest number of genes with a total of 14,737 genes that were differentially expressed in at least two experiments. PD showed 4,713 differentially expressed genes, ALS showed 897 and 4,249 differentially expressed genes could be found for HD. All pairwise comparisons of diseases on the transcriptomic level showed highly significant enrichment in the number of overlapping genes. One thousand nine hundred sixty-four gene names could be related to differentially expressed proteins for AD. For PD, we found 434 gene names, 155 for ALS, and 104 for HD. All pairwise overlaps between the four diseases were highly significantly enriched for the proteome data.

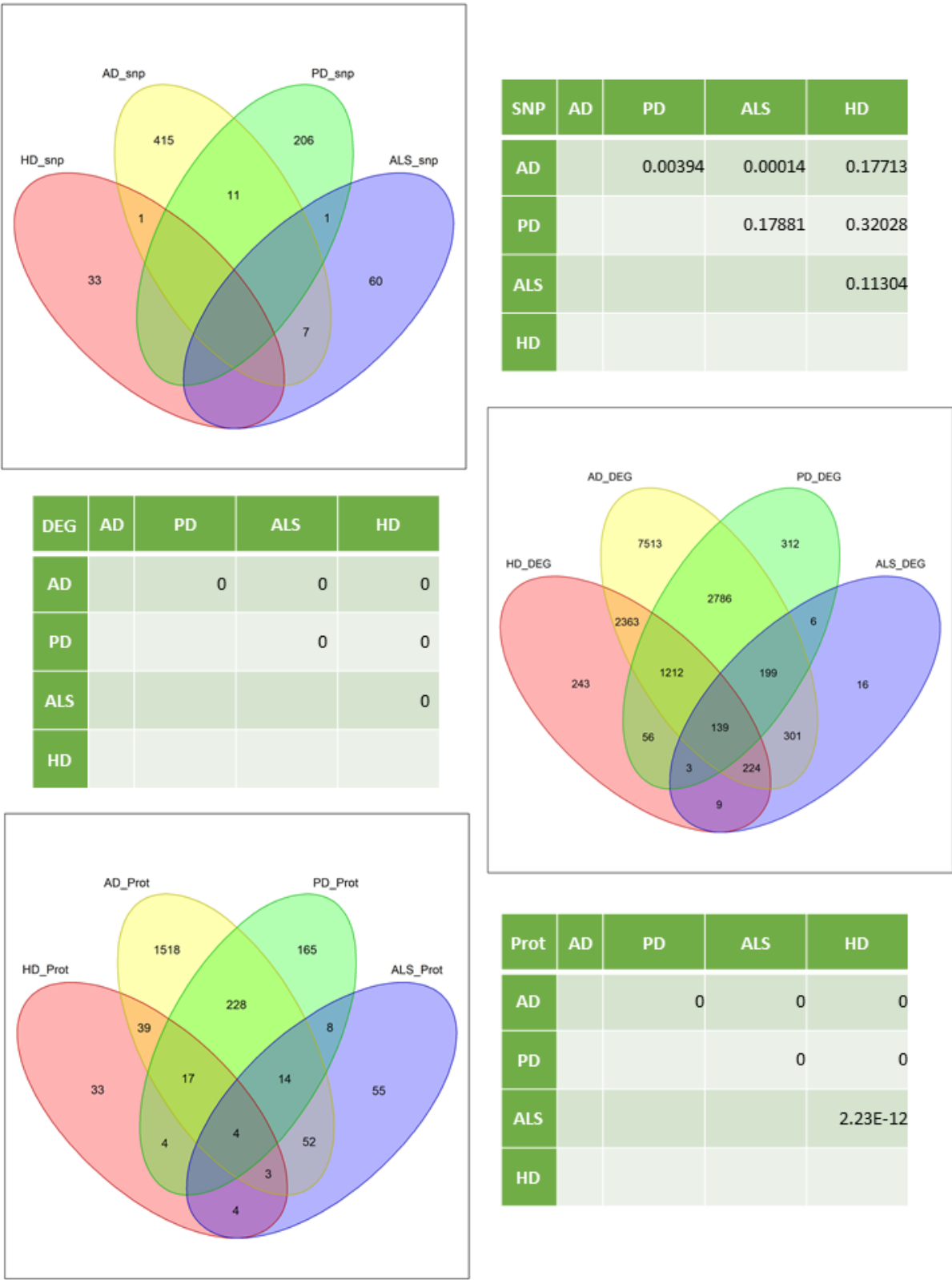


**Figure 3** Gene Occurrence Bar Plot showing the occurrences of the twenty most abundant genes in terms of experiments they appeared in as significantly up- or downregulated on the transcriptomic (left column) and proteomic layer (right column) for the four analyzed NDDs (from top: AD, PD, ALS, HD). The color of each bar gives the mean direction of regulation.





**Figure 4** Venn Diagrams and tables are showing intersections and the hypergeometric test results for the overlap between transcriptome and proteome data (column 1), transcriptome and SNP data (column 2), and proteome and SNP data (column 3) for AD, PD, ALS, and HD. All overlaps between the transcriptomic and proteomic levels were significantly higher than expected. AD also showed a significant overlap between the genomic and transcriptomic/proteomic level, PD for genomic-transcriptomic. The other intersections with genomic data did not show significantly more genes than expected by chance.



**Figure 5** Venn diagrams and hypergeometric test results for the overlap between significantly differentially expressed genes on the genomic level (top), transcriptomic level (middle), and proteomic level (bottom) for AD, PD, ALS, and HD. All tested intersections show highly significant enriched numbers of overlapping genes for the transcriptomic and proteomic data. The genomic data showed significantly enriched numbers of overlapping genes for the AD-PD and AD-ALS intersections.

**GO-Term- & Pathway-Analyses:** As the number of possible combinations and analyses was very high this study concentrated on describing the analyses of the genes appearing in the intersection of all diseases per omics layer and on those, appearing in the transcriptomic and proteomic data per disease (+ genomic data for AD). GO-Terms and genes were classified according to the conceptual model of candidate pathways contributing to neurodegeneration shown in **Figure 6** (modified version according to (Ramanan and Saykin, 2013)). NDD can be classified into broader functional groups based on their primary site or mode of action (intracellular mechanisms, local tissue environment influences, and systemic influences). Candidate pathways influencing the balance of neuronal survival, and degeneration are shown below. This model's pathways and mechanisms are highly related and can have overlapping or interacting components that can collectively modulate neurodegenerative processes.

Intracellular mechanisms	Local tissue environment	Systemic environment
<ul style="list-style-type: none"><li>•mitochondrial functions</li><li>•cell cycle (mitosis, meiosis)</li><li>•expression regulation (transcriptomic, (post)translational, epigenetic)</li><li>•endocytosis (vesicles, lysosomes, secretion)</li><li>•cytoskeleton (actin filaments, microtubuli, kinesin)</li><li>•stress response (oxidative, heat, cold, DNA damage)</li><li>•transport</li><li>•homeostasis</li><li>•autophagy (ubiquitin/proteasome)</li><li>•Apoptosis</li></ul>	<ul style="list-style-type: none"><li>•cell adhesion (attachment, tight junctions, gap junctions, Blood-Brain-Barrier)</li><li>•cell migration</li><li>•cell differentiation</li><li>•cell proliferation</li><li>•extracellular matrix</li><li>•organ development (brain, bone, muscle)</li><li>•signal transduction (synapsis, ionchannels, membrane shuttle, receptor)</li></ul>	<ul style="list-style-type: none"><li>•immune system (inflammation, infection response, innate immune response)</li><li>•metabolism (energy, maintenance)</li><li>•endocrine factors</li><li>•vascular factors</li><li>•neurotrophic factors</li><li>•aging</li></ul>

**Figure 6** Classification of candidate pathways contributing to neurodegeneration into three groups according to their cellular mechanisms or their primary site of action (modified and extended version) (Ramanan et al., 2014). All of our found GO-Terms Biological Processes (BP) were involved in at least one of the given categories' intracellular mechanisms, local tissue environment, or systemic environment.

**The intersection of genomic, transcriptomic, and proteomic data**

**AD:** The over-representation analysis (ORA) of the gene set emerging from the interaction of the genomic, transcriptomic, and proteomic AD data showed 18 GO-Terms that are significantly overrepresented (FDR ≤0.05) in the category Biological Process (BP). By affinity propagation, they are categorized into altered autophagy and immune response (β-amyloid formation), stress response/neuronal death, and extracellular matrix/cell communication. All eight Cellular Components (CC)-Terms are related to extracellular matrix and cell migration. Both Molecular Function (MF)-Terms belong to metabolism (lipoprotein binding).

## Intersection of transcriptomic and proteomic data

**AD:** There are many more genes overlapping if only transcriptomic and proteomic data are intersected, and thus much more BP-Terms (about 1350) can be found. By affinity propagation, BP was reduced to about 400 terms. The majority of these terms deal with cell adhesion and cell differentiation and metabolism, especially sugar- and lipid metabolism. Signal transduction (synapsis-related processes) was also frequently overrepresented, followed by stress response, autophagy and apoptosis, mitochondrial functions, signal transduction (ion channels), and immune system. The circa 270 overrepresented CC-Terms (75 after affinity propagation) were consequentially high in endocytosis (vesicles and membranes), signal transduction (neurons and synapses), mitochondrial functions, and other organelles and cell adhesion. More than half of the 48 MF-Terms deal with metabolism (protein binding and lipid transport), whereas the remaining terms are distributed between cell adhesion, signal transduction, and stress and immune response.

**PD:** The approx. 200 overrepresented BP-Terms found for the intersection of transcriptomic and proteomic PD data were classified by affinity propagation to cytoskeleton and cell development (10/36), metabolism (9/36), almost half of it concerning sugar metabolism, stress, and immune response (6/36) and transcription regulation (4/36). The rest dealt with apoptosis, signal transduction (synapses, ion channels), and organs' development (muscle cells and neurons of the substantia nigra). The 75 significantly overrepresented CC-Terms were reduced to 11 categories by affinity propagation, i.e., endocytosis (vesicles, especially in synapses) (5/11), mitochondrial functions (3/11), and cell adhesion (3/11). From 49 molecular functions, 11 terms with very diverse processes like autophagy (protein tagging for proteasome) and cytoskeleton (especially tubulin-binding) and immune system, translation regulation, and metabolism emerge affinity propagation.

**ALS:** Most of the overrepresented BP-Terms deal with cell adhesion and cytoskeleton (7/8). Only one process is immune response (*Staphylococcus aureus* infection). All five overrepresented CC-Terms are connected to cell differentiation (focal adhesion, extracellular matrix), and all 3 MF-Terms refer to the cytoskeleton.

**HD:** The 21 overrepresented GO-Terms for BP can be classified by affinity propagation into the categories: cell development, signal transduction, and metabolism, especially lipid transport; all three CC-Terms are associated with cell adhesion, and both MF-Terms concern metabolism, especially lipid transport.

## Intersection of AD, PD, ALS, and HD

**Transcriptome:** The 28 overrepresented BP-Terms for the intersection of all four diseases' transcriptomic data can be classified by affinity propagation, mainly to stress response (5/8). The remaining processes are concerning transcriptional regulation and extracellular matrix. All of the six detected CC-Terms are related to focal adhesion. The six overrepresented MF-Terms are mainly about cell adhesion and cytoskeleton (4/6), vascular factors (growth factor binding) (2/6), and stress response (1/6).

**Proteome:** For the intersection of proteomic data for all four analyzed NDD, three significantly overrepresented GO-Terms were found. They are all relevant to metabolism (maintenance of protein stability) and the immune system (humoral immune response).

#### 4. Discussion

**AD – Transcriptome:** Among the 20 top hits we found for the differentially expressed genes in AD are *ELF1*, *MSI2*, and *TBL1X*, which regulate transcription, *BAZ1A* for chromatin remodeling, *DNAJC1* as a response to heat stress, *PAK1*, and *ANTRX1* involved in cell adhesion and cell migration and *GDAP1L1* for neuronal development. The most intriguing finding is the transcript of the nicotinamide nucleotide adenylyltransferase 2 (*NMNAT2*) being downregulated in 16 studies, according to the literature (Ali et al., 2016; Bennett and Keeney, 2018). *NMNAT2* is an essential enzyme for energy metabolism in the brain, catalyzing an essential step in NAD/NADP biosynthesis. Its expression levels correlate positively with cognitive function and negatively with AD pathology. *NMNAT2* appears to be required for axon survival and functions as a chaperone to aid in the refolding of misfolded proteins to reduce proteotoxic stress (Zhai et al., 2008; Lukacs et al., 2019).

**AD - Proteome:** The top 20 hits for the genes related to the differentially expressed proteins in AD are mainly involved in cytoskeleton organization, in cell adhesion and neuronal activity. The microtubule-associated protein tau (*MAPT*) is expressed at its highest levels in neurons throughout the central nervous system. It is involved in assembling and stabilizing the cytoskeleton (Strang, Golde and Giasson, 2019). Microtubules help cells maintain their shape, assist in cell division, and are essential for transporting materials within cells. One of tau's primary functions is to bind to and promote microtubules' assembly and stability; this binding activity can be negatively regulated by phosphorylation at select sites (Arendt, Stieler and Holzer, 2016). An imbalanced regulation in protein kinases and protein phosphatases is the direct cause of AD-like tau hyperphosphorylation (Wang and Liu, 2008) and hyperphosphorylated tau accumulation induces synaptic toxicity and cognitive impairments (Ma et al., 2017). Clusterin (*CLU*) or *APOJ* is a multifunctional glycoprotein that has been implicated in several biological processes, including lipid transport, membrane recycling, cell adhesion, programmed cell death, and the complement cascade. Although Clusterin usually is a secreted protein, it has also been found intracellularly under certain stress conditions (Koltai, 2014). Clusterin's ability to interact and bind to A $\beta$  appears to alter aggregation and promote A $\beta$  clearance, suggesting a neuroprotective role (DeMattos *et al.*, 2004). *CLU* is considered the third most significant genetic risk factor for the non genetic form of AD, after *APOE* and *BIN1* with reduced secreted Clusterin as a mechanism for Alzheimer-associated *CLU* mutations (Bettens *et al.*, 2015). Interestingly, in most of our studies, *CLU* was upregulated.

**PD - Transcriptome:** Most of the differentially expressed genes found in PD were involved in gene expression regulation and stress response/autophagy processes. Among the top 20 differentially expressed genes in PD, downregulated *ANK1* was found. Ankyrins are a family of proteins that link the integral membrane proteins to the underlying spectrin-actin cytoskeleton and play critical roles in cell motility, activation, proliferation, contact, and specialized membrane maintenance domains. They attach integral membrane proteins to cytoskeletal elements. *ANK1* shows altered methylation and expression in Alzheimer's disease (De Jager *et al.*, 2014; Lunnon *et al.*, 2014) and Parkinson's disease (Liscovitch and French, 2014). Another interesting finding is the enzyme pyridoxal kinase (*PDXK*) that catalyzes the conversion of vitamin B6 (pyridoxine, pyridoxal, and pyridoxamine) to pyridoxal 5'-phosphate (PLP), which acts as a cofactor for over 140 different enzymatic



reactions. Of direct relevance to PD, the second step in the biosynthesis of dopamine by the enzyme dopa decarboxylase is dependent on PLP as a cofactor and becomes rate-limiting in patients receiving L-DOPA therapy (Tan *et al.*, 2005). An adaptive mechanism may explain the upregulation of PDXK in dopaminergic neurons to increased dopamine metabolism in the remaining functional dopaminergic neurons of the SN or L-DOPA therapy (Elstner *et al.*, 2009). Commonly, however, as in our studies, *PDXK* is downregulated in PD patients.

**PD - Proteome:** Like the differentially expressed genes most of the differentially expressed proteins were involved in gene expression regulation and stress response/autophagy processes but also in tubulin polymerization. Microtubules are essential for the mitotic spindle, the extracellular matrix, cell migration, and the growth of the myelin sheath protecting neurons.

One of the enzymatic reactions where the above mentioned PLP acts as a cofactor is the catabolism of gamma-aminobutyric acid (GABA) into succinic semialdehyde. In this reaction, PLP is bound to the 4-aminobutyrate aminotransferase (ABAT), which belongs to the top 20 genes related to the differentially expressed proteins found in our study. GABA is an important, mostly inhibitory neurotransmitter in the central nervous system, estimated to be present in nearly one-third of human synapses. Parkinson patients with ABAT deficiency showed increased GABA levels in the plasma and cerebrospinal fluid and associated symptoms such as encephalopathy, psychomotor retardation, seizures, hypotonia, hyperreflexia, lethargy, and abnormal electroencephalogram (Kim *et al.*, 2018; O'Gorman Tuura, Baumann and Baumann-Vogel, 2018). Another interesting differentially expressed protein in PD is the tyrosine 3-monooxygenase/tryptophan 5-monooxygenase activation protein epsilon YWHAE. This protein belongs to the 14-3-3 family, it is especially abundant in brain tissue (Boston, Jackson and Thompson, 1982), comprises several isoforms and interacts mainly with phosphorylated protein partners. Members of the protein family have been associated with the development of neurodegenerative diseases (Berg, Holzmann and Riess, 2003), among them Parkinson's disease and Alzheimer's disease (Umahara *et al.*, 2004), where they are found in neurofibrillary tangles (NFTs). NFTs are composed of hyperphosphorylated Tau protein displaying paired helical filaments and substantial amounts of 14-3-3 proteins. It could be shown that the functional and direct interaction of overexpressed 14-3-3 with Tau proteins leads to Tauopathies, such as Tau accumulation and Tau detachment from microtubules (Joo *et al.*, 2015).

**HD – Transcriptome:** The top 20 differentially expressed genes in HD are mainly involved in the mitotic cell cycle, epigenetic modifications, cell differentiation and proliferation, apoptosis, and immune responses to inflammation and infection. In most of our studies, Perilipin 2 (*PLIN2*) was found to be upregulated. *PLIN2* is a perilipin family member, which coat intracellular lipid storage droplets and develops and maintains adipose tissue. However, it is not restricted to adipocytes but is also found in astrocytes in the brain. The lipid imbalance plays a significant role in neurodegeneration, such as in Alzheimer's disease (Dalhaimer, 2019). The E4 allele of *APOE*, gene product of which is a major structural component of low-density lipoproteins, is a decisive genetic risk factor for developing late-onset Alzheimer's disease (Liu *et al.*, 2013). Astrocytes expressing E4 accumulate significantly more and smaller lipid droplets compared to E3 astrocytes. Accordingly, the expression of *perilipin-2* was higher in E4 astrocytes (Farmer, Kluemper, and Johnson, 2019). Another exciting gene found in HD is Kinectin 1 (*KTN1*), also known



to be related to PD. *KTN1* encodes an integral membrane protein, primarily localized in the endoplasmic reticulum membrane. This protein binds kinesin, a motor protein that is involved in intracellular organelle motility. Powered by the hydrolysis of ATP, kinesins move along microtubule filaments. The active movement of kinesins supports several cellular functions, including mitosis, meiosis, and cellular transport, such as in axonal transport. Most kinesins move towards the plus end of a microtubule, transporting cargo such as protein and membrane components from the inner cell towards the periphery. *KTN1* facilitates vesicle binding to kinesin, regulating crucial developmental processes including axonal guidance, vesicular transport of molecules, and apoptosis (Kumar, Yu, and Sheetz, 1995; Hibar *et al.*, 2015), as well as neuronal cell shape and neuronal migration through kinectin–kinesin interactions (Zhang *et al.*, 2010). Neurons with more kinectin1 have larger cell bodies (Toyoshima and Sheetz, 1996). A positive relationship among PD risk and *KTN1* mRNA expression in the putamen and putamen volumes, and a modest relation between PD risk and *KTN1* mRNA expression in SNc could be shown, suggesting that *KTN1* may play a functional role in the development of PD (Mao *et al.*, 2020).

**HD – Proteome:** The genes related to the differentially expressed proteins could be linked to mitochondrial and lysosomal functions, and the actin-, dystrophin- and kinesin filaments of the cytoskeleton and muscle cells. Among the proteins that were upregulated in HD, we found the Cysteine and Glycine Rich Protein 1 (*CSRP1*), which may be involved in regulatory processes essential for development and cellular differentiation. *CSRP1*'s LIM/double zinc-finger motif occurs in proteins with critical functions in gene regulation, cell growth, and somatic differentiation. Members of the CRP family are involved in a late stage in muscle differentiation (Pomiès, Louis and Beckerle, 1997), and it could play a role in neuronal development (Knierim *et al.*, 2016). Laminin Subunit Gamma 1 (*LAMC1*), a high-molecular-weight glycoprotein of the extracellular matrix, which belongs to the family of laminins, was found to be downregulated in all our HD studies. Laminins are the principal non-collagenous constituent of the basal lamina, a protein network foundation for most cells and organs, influencing cell differentiation, migration, and adhesion (Durbeej, 2010). They are also an integral part of the structural scaffolding in almost every organism's tissue and are incorporated and secreted into cell-associated extracellular matrices. Defective laminins can cause improperly formed muscles, thus leading to a form of muscular dystrophy (Yurchenco and Patton, 2009).

**ALS – Transcriptome:** The main pathways for the top 20 genes of the differentially expressed genes for ALS are cell cycle and gene expression regulation, stress response, apoptosis, the maintenance of the cytoskeleton, and the extracellular matrix neuronal development. CD44, also present among the top 20 differentially expressed HD genes, was found to be upregulated in ALS, too. Its protein is a transmembrane, highly glycosylated protein involved in cell-cell interactions, cell adhesion, and migration, helping cells sense and respond to tissue microenvironment (Dzwonek and Wilczyński, 2015). In the nervous system, CD44 expression occurs in both neuronal and glial cells (McKenzie, Dalchau, and Fabre, 1982). The role of CD44 in the physiology and pathology of the nervous system is not entirely understood. However, there exists evidence suggesting it might be involved in the axon guidance, cytoplasmic Ca<sup>2+</sup> clearance, dendritic arborization, synaptic transmission, epileptogenesis, oligodendrocyte and astrocyte differentiation, and post-traumatic brain repair (Jones *et al.*, 2000; Matzke *et al.*, 2007; Vedunova *et al.*, 2013; Skupien *et al.*, 2014). Aquaporin 1 is another crucial ubiquitous protein that was among the top 20

differentially expressed genes for ALS. Aquaporins are a family of widely distributed membrane-inserted water channel proteins providing a pathway for osmotically driven water, glycerol, urea, or ions transport through cell membranes and thus control particular aspects of homeostasis. Aquaporins, especially *AQP4*, are abnormally expressed in the Central Nervous System under some pathological conditions like neurodegenerative diseases in which preservation of brain homeostasis is at risk (Foglio and Luigi Fabrizio, 2010). *AQP1* encodes a small integral membrane protein, which is mostly found in the plasma membranes of red cells and kidney proximal tubules with high permeability to water. Herewith, water can be held even against the direction of an osmotic gradient (Badaut *et al.*, 2014).

**ALS – Proteome:** Among the genes related to ALS's differentially expressed proteins were secreted phosphoprotein 1 (*SPP1*) or osteopontin and hemopexin (*HPX*) genes. *SPP1* is a matrix phosphoprotein expressed by various tissues and cells, including the immune system and the nervous system. Immune cells, such as macrophages and T lymphocytes, are essential sources of *SPP1* during inflammatory processes (Mousavi, Agah, and Tafakhori, 2020). *SPP1* may have a role in neurodegenerative diseases, including multiple sclerosis, PD, AD, and frontotemporal dementia (Carecchio and Comi, 2011; Agah *et al.*, 2018). It was reported that *SPP1* was selectively expressed in alpha motor neurons, which are the most vulnerable in ALS (Misawa *et al.*, 2012; Yamamoto *et al.*, 2017). Therefore, it may also play a role in the pathogenesis of ALS. Hemopexin encodes a plasma glycoprotein that binds heme with high affinity. It is an acute-phase protein that transports heme from the plasma to the liver for breakdown and iron recovery. It may be involved in protecting cells from oxidative stress (Ashraf *et al.*, 2020), because Hb oxidation liberates free heme, which is a source of redox-active iron-producing reactive oxygen species, inducing lipid peroxidation (Papanikolaou and Pantopoulos, 2005). This toxicity is modulated by hemopexin. Proteomics studies demonstrated increased plasma and cerebrospinal fluid hemopexin in AD patients compared to cognitively normal controls, suggesting impaired compensation in neurodegeneration (Castaño *et al.*, 2006). It appears that surplus contribution to heme by the breakdown of Hb may overwhelm the capacity of the heme scavenging system in AD (Smith and McCulloh, 2015; Ma *et al.*, 2016).

The analysis of the twenty most abundant genes in each NDD's transcriptome and proteome data interestingly showed many highly abundant genes in single NDD that were also described for other NDD, e.g., *SPP1*, that was highly abundant in the ALS proteomic data but already described for PD, AD and other NDD like multiple sclerosis. Taken together, many of the upcoming genes were related to cytoskeleton, cell adhesion, extracellular matrix formation, apoptosis, and stress response. The results show the general conformity of the most abundant genes with the current study situation. Many genes are described as playing essential roles in more than one of the investigated NDD. Additionally, looking at the mean direction of regulation among all studies involving a particular gene on the respective omics-layer showed that in some cases, the current study situation seems to be contradictory even for some of the most abundant significantly differentially expressed genes, such as *ANTXR1* and *ARHGAP12* for transcriptomic AD data and *ZFAND5* and *PCMT1* for PD transcriptomic and proteomic data. However, apart from these contradictory regulated genes, most of the twenty top abundant genes were consistent among most studies.

**Intra-disease relations:** The intersection of genes found in genomic, transcriptomic, and proteomic data per disease showed more surprising results. HD and ALS did not show any genes that were shared between the genomic and proteomic data. The number of five genes shared between these two omics-layers for PD is not higher than expected by chance. Only in AD a significant overrepresentation of genes shared between the genomic and proteomic layer was found. On the other hand, both AD and PD showed a significantly high number of genes in the genomic and transcriptomic data (p-value<sub>AD</sub>: 8.04E-9, p-value<sub>PD</sub>: 5.38E-4). The number of shared genes between the transcriptomic and proteomic level was highly significantly enriched in all four investigated NDD.

**Inter-disease relations:** Interestingly, the number of shared genes between the four NDD on the genomic level was significantly enriched only for the AD-PD and AD-ALS comparisons. For the proteomic and transcriptomic data, all numbers of pairwise overlapping genes were significantly enriched. However, the overlap between all four analyzed NDD was large enough to be further analyzed only at the transcriptomic level. The genomic and proteomic data of all four NDD showed no (genomic) and four (proteomic) overlapping genes. Nevertheless, the overlap between all four NDD on the transcriptomic level showed 139 genes. The overrepresented GO-Terms for biological process in the genes appearing in the intersection of all four NDD on the transcriptomic level showed results that are partially concordant to a meta-study of AD, PD and ALS from 2019 (Bayraktar *et al.*, 2019), that analyzed raw data of 259 individuals. They found pathways and biological processes associated to heat shock proteins, cellular response to heat, stress response, GABA synthesis and Protein folding as overrepresented in the four used datasets and stated the importance of heat shock proteins (HSP) as a general target of NDD (Kampinga, Harm H., 2016) and the importance of HSP associated pathways in HD (Kaltenbach *et al.*, 2007).

Our GO-Term analysis revealed biological processes that are associated to cellular response to heat, stress response and protein folding. However, no significant overrepresentation was found for HSP or GABA synthesis in our analysis of AD, PD, ALS and HD. We furthermore found biological pathways associated to response to hypoxia and cell ageing as overrepresented in the gene set that was common to all four NDD on the transcriptomic level. Consequently, we could confirm the findings regarding heat response, protein folding and stress response, although our analysis did neither explicitly show pathways or biological processes associated to HSP nor GABA synthesis.

## Conclusion:

GWA studies made a big contribution towards understanding NDD in the last 15 years, with several hundred disease-associated risk loci. However, until now for most NDD no targeted therapies have emerged from these GWA studies (Diaz-Ortiz and Chen-Plotkin, 2020). As NDD are causing vast transcriptomic changes in the ageing brain, it is crucial to take transcriptomic data analysis into account as well, when analyzing NDD (De Jager, Yang and Bennett, 2018). The further translation from transcriptomic changes to proteomic occurs only indirectly and shows just a limited correlation between mRNA and protein expression (Becker *et al.*, 2018). Consequently, even the combination of genetic and transcriptomic data is not adequate to give a complete picture of changes taking place due to NDD. Each additional level of information can contribute to a better understanding of the complex interrelationships of these interacting omics-layers.

For addressing the challenges of such complicated diseases, the whole field of biomedicine is changing towards creating and facilitating a variety of databases and analysis pipelines for separate omics-layers as well as multi-omics integration (Manzoni, Lewis and Ferrari, 2020). Many of these pipelines are mainly data-driven and facilitate clustering and supervised machine learning techniques to find important patterns of features contributing to the identification of e.g. proteins that are associated to NDD (Yu *et al.*, 2020) or to reveal cross-talk patterns in multi-omics data (Nguyen and Wang, 2020).

According to the necessity of approaching complex diseases with the use of multiple omics-layers and data-driven methods utilizing large amounts of data, we combined data of three omics-layers from databases and literature mining of more than one million subjects and 188 studies to show shared genes within and between the four analyzed NDD and extract the pathways and processes in which they are overrepresented.

To classify the gained information in this study, it is important to keep in mind that the transcriptomic and proteomic data were gathered from various tissues, partly different severities of diseases and using different methods. Consequently, the signals that have been found in this meta-study rather represent stable signals across tissues emerging in the late stage of NDD than subtle effects that might only be present in specific brain regions, or earlier disease states as those subtle effects would probably be canceled out when doing a GO-Term or Pathway-Analysis by the number of other signals and the fact that the majority of the used data was based on late stage NDD. The inequality between the four different diseases at the given time point concerning the study situation and the amount of insight should also be considered and was visible in the differing number of samples, studies and differentially expressed genes between AD and the other three NDD. The highly significant overrepresentation of genes in the intersection of proteomic and transcriptomic data in all investigated NDD shows the importance of simultaneously analyzing multiple omics-layers. On the other hand, despite the given overlap, many differences between the transcriptomic and proteomic layers remain, which was also shown in the differences between pathways and GO-Terms being significantly enriched in the transcriptomic layer compared to the proteomic layer. While the intersection of all NDD on the transcriptomic layer showed terms related to stress response, cell development, cell adhesion, and cytoskeleton, the genes appearing in the intersection of the proteomic data were related to protein stability and immune response. Accordingly, analyzing transcriptomic and proteomic data simultaneously in a multi-omics approach while still considering those effects exclusive to one omics-layer is necessary to investigate NDD to the full extend. Regarding future research on this topic, it might be helpful to expand the repertoire of omics-layers by epigenomics and / or adding further NDD. However, analyzing and interpreting all possible intersections was already not manageable for the current set of diseases and omics-layers with the used approaches due to the large amount of combinatorically possible intersections. Consequently, different analysis approaches might be an option to deal with the further rising degrees of freedom regarding the analysis. Concentrating further on the differences and coherences in the direction of regulation of interesting genes also holds new insight regarding the clustering of diseases or finding similarly regulated gene sets.

## Materials & Methods

**Genome:** The GWAS Catalog data for Alzheimer's disease (AD), Parkinson's disease (PD), Amyotrophic Lateral Sclerosis (ALS), and Huntington's disease (HD) were downloaded on the 28<sup>th</sup> of April 2020. The GWAS Catalog contains SNP data of GWAS studies for SNPs showing a statistical significance of SNP-trait p-value < 1.0E-5 in the overall population. For every SNP, data such as p-value, upstream gene(s), mapped gene, reported gene(s), and many more are stored. We focused on the genes given as "Reported Gene(s)" in the four examined diseases' full data tables for our analysis. The EFO numbers for the exact search pattern were EFO\_0000249 (Alzheimer's disease), EFO\_0002508 (Parkinson's disease), Orphanet\_399 (Huntington's disease), and EFO\_0000253 (Amyotrophic Lateral Sclerosis). A table containing all studies' names and the number of investigated samples for every disease is appended in the supplement (A1: GWAS Catalog study information)**Error! Reference source not found.**

**Transcriptome:** We browsed the databases Gene Expression Omnibus (GEO) (Barrett *et al.*, 2013) and the Expression Atlas (Papatheodorou *et al.*, 2018). The GEO is a public data repository, in which microarray and RNA-seq datasets can be found. The keywords for the GEO database were <name of disease> AND ("microarray" OR "RNAseq") AND "human." The latest literature research was done in July 2020. The Expression Atlas is a service of EMBL-EBI and provides re-analyzed and manually curated data of more than 3000 experiments. It was used in release 35 (May 2020, <https://www.ebi.ac.uk/gxa/home>) and scanned for Alzheimer, Parkinson, Huntington, and Amyotrophic Lateral Sclerosis setting the filter to "Homo sapiens" in the section Differential Experiments. Whenever experiments with increasing severities of an NDD were conducted, we used the differentially expressed genes appearing in the comparison between the group of the most severe disease state and the control.

An overview of all used studies for gathering the transcriptomic data is given in

. A table showing each study's information is given in the supplement (A2: Transcriptome study information).

<i>Transcriptome</i>	Case	Control	$\Sigma$ of samples	studies
<i>AD</i>	1029	756	1785	14
<i>PD</i>	154	167	321	12
<i>ALS</i>	170	131	301	8
<i>HD</i>	39	29	68	12
$\Sigma$	1392	1083	2475	46

**Table 1** Overview of the number of cases, controls, and the total number of studies per disease throughout all analyzed transcriptome studies. In total, data of 2475 samples were gathered from 46 studies analyzing transcriptomic data.



**Proteome** We browsed publications of the last ten years in PubMed and Google Scholar with the keywords: ("neurodegenerative diseases" OR "Alzheimer\* disease" OR "Parkinson\* disease" OR "Huntington\* disease" OR "Amyotrophic Lateral Sclerosis") AND (proteomics OR "quantitative proteomics" OR "differentially expressed proteins" OR biomarkers) AND human NOT mice. An overview of all used studies for gathering the transcriptomic data is given in **Table 2**. A table showing each study's information is given in the supplement (**Error! Reference source not found.**). As well as with the transcriptomic data, we used the results emerging from the comparison of the group with the most severe disease state compared to the control, when experiments of increasing severity were available.

	<i>Proteome</i>	Case	Control	$\Sigma$ of samples	studies
AD		293	274	567	9
PD		302	279	581	7
ALS		471	691	1162	5
HD		360	181	541	3
$\Sigma$		1426	1425	2851	24

**Table 2** Overview of the number of cases, controls, and the total number of studies per disease throughout all analyzed proteome studies. In total, data of 2851 samples were gathered from 24 studies analyzing proteomic data.

**Data Management:** The raw genomic, transcriptomic, and proteomic data tables from 188 different studies were transformed into standardized tables for each disease on every omics-layer. Different conversion was applied within this data management process, such as converting fold change to log<sub>2</sub>-fold change (log<sub>2</sub>FC), log<sub>10</sub>-p-value to p-value, removal of entries with missing Gene Name or separating rows that contained several Gene Names (proteomic data). Differences in multiple testing corrections were accepted, such as differences in the exact calculation of the fold change (log<sub>2</sub>FC, G-fold Change). Only those genes with a false-discovery-ratio (FDR)  $\leq 0.05$  were selected after applying those conversions where necessary. Besides, all genes that appeared as differentially expressed in only one experiment on the transcriptomics or proteomics level were discarded, to further reduce the number of genes that appeared randomly. Finally, all remaining genes from the genomic, transcriptomic, and proteomic data sources were intersected with the latest list of protein-coding gene symbols (04.08.20) from the HUGO Gene Nomenclature Committee (HUGO) to exclude non-standard gene names.

**Gene Occurrence:** For every final table of processed data, the top 20 genes were visualized in bar plots, showing the abundance and mean direction of every gene's regulation. The mean direction of regulation for each gene is represented by the bar plots' corresponding



bar's color. The color represents the ratio of studies in which a gene was significantly up- or downregulated. The mean direction of regulation was computed as follows:

Equation 1: Calculation of mean regulation of direction

$$MeanRegDir(gene) = \frac{1}{n} \sum_{i=1}^n \left( sig \left( gene_{foldChange_i} \right) \right)$$

with:

$$n = \text{number of appearances for gene with } FDR \leq 0.05$$

$$sig(x) = \begin{cases} 1, & \text{if } x > 0 \\ -1, & \text{if } x < 0 \end{cases}$$

**Intersection:** By intersecting the three analyzed omics-layers per disease and the four diseases per omics-layer, it is possible to test if the number of shared genes between some omics-layer or diseases is significantly increased. We used a hypergeometric test to test the overlapping sets, having the total amount of 19,324 Gene Symbols of protein-coding genes (HUGO Gene Nomenclature Committee 04.08.20) (Braschi *et al.*, 2019) as the total population. Intersections were performed and visualized using the R package Venn (Dusa, 2016).

**GO-Term- & Pathway-Analyses:** Independent of the test results, these sets of overlapping genes were also used for KEGG-pathway analyses (M Kanehisa - Novartis Foundation Symposium, 2002) and GO-Term (Gene Ontology Consortium, 2015) analysis. We used the R API WebGestaltR 0.4.4 of the online tool WebGestalt 2020 (Liao *et al.*, 2019) for performing Overrepresentation Analyses (ORA) for all possible intersections per disease and per omics-layer. For performing the ORA, the command **WebGestaltR** was used with the options:

```
enrichDatabase=c("pathway_KEGG","geneontology_Biological_Process","geneontology_Cellular_Component", "geneontology_Molecular_Function"),
```

```
interestGeneType="gene symbol
```

```
referenceSet="genome"
```

```
topThr = 10000
```

```
reportNum = 10000
```

The organism was set to "hsapiens" by default.

**Author Contributions:** "Conceptualization, S.G.; methodology, N. R.; S. K. and S.G. ; software, N.R. and S.K.; validation, N.R. and S.K.; formal analysis, N.R. and S.K.; writing—original draft preparation, N.R, S.K. and S.G. writing—review and editing, S. G., N.R. and S.K. ; visualization, N.R. supervision, S. G.; funding acquisition, S. G. All authors have read and agreed to the published version of the manuscript."

**Funding:** This research was partially funded by the Emergent AI Center funded by the Carl-Zeiss-Stiftung and the Leibniz Institute for Resilience Research.

**Conflicts of Interest:** The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results

## References

- Agah, E. *et al.* (2018) 'Osteopontin (OPN) as a CSF and blood biomarker for multiple sclerosis: A systematic review and meta-analysis', *PLOS ONE*. Edited by H. Wiendl. Public Library of Science, 13(1), p. e0190252. doi: 10.1371/journal.pone.0190252.
- Alexander, G. E. (2004) 'Biology of Parkinson's disease: Pathogenesis and pathophysiology of a multisystem neurodegenerative disorder', *Dialogues in Clinical Neuroscience*. Les Laboratoires Servier, pp. 259–280. Available at: [www.dialogues-cns.org](http://www.dialogues-cns.org) (Accessed: 31 August 2020).
- Ali, Y. O. *et al.* (2016) 'NMNAT2:HSP90 Complex Mediates Proteostasis in Proteinopathies', *PLoS Biology*. Public Library of Science, 14(6), p. 1002472. doi: 10.1371/journal.pbio.1002472.
- Arendt, T., Stieler, J. T. and Holzer, M. (2016) 'Tau and tauopathies', *Brain Research Bulletin*. Elsevier Inc., pp. 238–292. doi: 10.1016/j.brainresbull.2016.08.018.
- Arneson, D. *et al.* (2018) 'Shared mechanisms among neurodegenerative diseases: from genetic factors to gene networks', *Journal of Genetics*. Springer, pp. 795–806. doi: 10.1007/s12041-018-0963-3.
- Ashraf, A. *et al.* (2020) 'Plasma transferrin and hemopexin are associated with altered A $\beta$  uptake and cognitive decline in Alzheimer's disease pathology', *Alzheimer's research & therapy*. NLM (Medline), 12(1), p. 72. doi: 10.1186/s13195-020-00634-1.
- Aylward, E. H. *et al.* (2004) 'Onset and rate of striatal atrophy in preclinical Huntington disease', *Neurology*. Lippincott Williams and Wilkins, 63(1), pp. 66–72. doi: 10.1212/01.WNL.0000132965.14653.D1.
- Badaut, J. *et al.* (2014) 'Aquaporin and brain diseases', *Biochimica et Biophysica Acta - General Subjects*. Elsevier, pp. 1554–1565. doi: 10.1016/j.bbagen.2013.10.032.
- Ballard, C. *et al.* (2011) 'Alzheimer's disease', *The Lancet*. Lancet Publishing Group, pp. 1019–1031. doi: 10.1016/S0140-6736(10)61349-9.
- Barrett, T. *et al.* (2013) 'NCBI GEO: Archive for functional genomics data sets - Update', *Nucleic Acids Research*. Oxford Academic, 41(D1), pp. D991–D995. doi: 10.1093/nar/gks1193.
- Bayraktar, A. *et al.* (2019) 'Meta-analysis of Gene Expression in Neurodegenerative Diseases Reveals Patterns in GABA Synthesis and Heat Stress Pathways'. Available at: <http://arxiv.org/abs/1909.07469> (Accessed: 7 October 2020).
- Becker, K. *et al.* (2018) 'Quantifying post-transcriptional regulation in the development of *Drosophila melanogaster*', *Nature Communications*. Nature Research, 9(1), pp. 1–14. doi:

10.1038/s41467-018-07455-9.

Bellenguez, C., Grenier-Boley, B. and Lambert, J. C. (2020) 'Genetics of Alzheimer's disease: where we are, and where we are going', *Current Opinion in Neurobiology*. Elsevier Ltd, pp. 40–48. doi: 10.1016/j.conb.2019.11.024.

Bennett, J. P. and Keeney, P. M. (2018) 'RNA-Sequencing Reveals Similarities and Differences in Gene Expression in Vulnerable Brain Tissues of Alzheimer's and Parkinson's Diseases', *Journal of Alzheimer's Disease Reports*. IOS Press, 2(1), pp. 129–137. doi: 10.3233/adr-180072.

Berardelli, A. *et al.* (2001) 'Pathophysiology of bradykinesia in parkinson's disease', *Brain*. Oxford University Press, pp. 2131–2146. doi: 10.1093/brain/124.11.2131.

Berg, D., Holzmann, C. and Riess, O. (2003) '14-3-3 proteins in the nervous system', *Nature Reviews Neuroscience*. Nature Publishing Group, 4(9), pp. 752–762. doi: 10.1038/nrn1197.

Berridge, M. J. (2014) 'Calcium regulation of neural rhythms, memory and alzheimer's disease', *Journal of Physiology*. Wiley-Blackwell, 592(2), pp. 281–293. doi: 10.1113/jphysiol.2013.257527.

Bettens, K. *et al.* (2015) 'Reduced secreted clusterin as a mechanism for Alzheimer-associated CLU mutations', *Molecular Neurodegeneration*. BioMed Central Ltd., 10(1), p. 30. doi: 10.1186/s13024-015-0024-9.

Boston, P. F., Jackson, P. and Thompson, R. J. (1982) 'Human 14-3-3 Protein: Radioimmunoassay, Tissue Distribution, and Cerebrospinal Fluid Levels in Patients with Neurological Disorders', *Journal of Neurochemistry*. John Wiley & Sons, Ltd, 38(5), pp. 1475–1482. doi: 10.1111/j.1471-4159.1982.tb07928.x.

Braak, H. *et al.* (2003) 'Staging of brain pathology related to sporadic Parkinson's disease', *Neurobiology of Aging*. Elsevier, 24(2), pp. 197–211. doi: 10.1016/S0197-4580(02)00065-9.

Braak, H. and Braak, E. (1991) 'Neuropathological stageing of Alzheimer-related changes', *Acta Neuropathologica*. Springer-Verlag, pp. 239–259. doi: 10.1007/BF00308809.

Brandt, J. (2018) 'Behavioral Changes in Huntington Disease', *Cognitive and Behavioral Neurology*. Lippincott Williams and Wilkins, pp. 26–27. doi: 10.1097/WNN.0000000000000147.

Braschi, B. *et al.* (2019) 'Genenames.org: The HGNC and VGNC resources in 2019', *Nucleic Acids Research*. Oxford University Press, 47(D1), pp. D786–D792. doi: 10.1093/nar/gky930.

Bredesen, D. E., Rao, R. V. and Mehlen, P. (2006) 'Cell death in the nervous system', *Nature*. Nature Publishing Group, pp. 796–802. doi: 10.1038/nature05293.

Briston, T. and Hicks, A. R. (2018) 'Mitochondrial dysfunction and neurodegenerative

proteinopathies: Mechanisms and prospects for therapeutic intervention', *Biochemical Society Transactions*. Portland Press Ltd, pp. 829–842. doi: 10.1042/BST20180025.

Cabral-Miranda, F. and Hetz, C. (2018) 'ER stress and neurodegenerative disease: A cause or effect relationship?', in *Current Topics in Microbiology and Immunology*. Springer Verlag, pp. 131–157. doi: 10.1007/82\_2017\_52.

Carecchio, M. and Comi, C. (2011) 'The role of osteopontin in neurodegenerative diseases', *Journal of Alzheimer's Disease*. IOS Press, pp. 179–185. doi: 10.3233/JAD-2011-102151.

Castaño, E. M. *et al.* (2006) 'Comparative proteomics of cerebrospinal fluid in neuropathologically- confirmed Alzheimer's disease and non-demented elderly subjects', *Neurological Research*. Neurol Res, 28(2), pp. 155–163. doi: 10.1179/016164106X98035.

Cattaneo, E., Zuccato, C. and Tartari, M. (2005) 'Normal huntingtin function: An alternative approach to Huntington's disease', *Nature Reviews Neuroscience*. Nature Publishing Group, pp. 919–930. doi: 10.1038/nrn1806.

Cerasa, A., Novellino, F. and Quattrone, A. (2016) 'Connectivity Changes in Parkinson's Disease', *Current Neurology and Neuroscience Reports*. Current Medicine Group LLC 1, pp. 1–11. doi: 10.1007/s11910-016-0687-9.

Cha, J. H. J. (2000) 'Transcriptional dysregulation in Huntington's disease', *Trends in Neurosciences*. Elsevier Ltd, pp. 387–392. doi: 10.1016/S0166-2236(00)01609-X.

Cistaro, A. *et al.* (2012) 'Brain hypermetabolism in amyotrophic lateral sclerosis: A FDG PET study in ALS of spinal and bulbar onset', *European Journal of Nuclear Medicine and Molecular Imaging*. Springer Verlag, 39(2), pp. 251–259. doi: 10.1007/s00259-011-1979-6.

Dalhaimer, P. (2019) 'Lipid Droplets in Disease', *Cells*. NLM (Medline), p. 974. doi: 10.3390/cells8090974.

Damiano, M. *et al.* (2010) 'Mitochondria in Huntington's disease', *Biochimica et Biophysica Acta - Molecular Basis of Disease*. Elsevier, pp. 52–61. doi: 10.1016/j.bbdis.2009.07.012.

Dauer, W. and Przedborski, S. (2003) 'Parkinson's disease: Mechanisms and models', *Neuron*. Cell Press, pp. 889–909. doi: 10.1016/S0896-6273(03)00568-3.

DeMattos, R. B. *et al.* (2004) 'ApoE and Clusterin Cooperatively Suppress A $\beta$  Levels and Deposition: Evidence that ApoE Regulates Extracellular A $\beta$  Metabolism In Vivo', *Neuron*. Cell Press, 41(2), pp. 193–202. doi: 10.1016/S0896-6273(03)00850-X.

Diaz-Ortiz, M. E. and Chen-Plotkin, A. S. (2020) 'Omics in Neurodegenerative Disease: Hope or Hype?', *Trends in Genetics*. Elsevier Ltd, pp. 152–159. doi: 10.1016/j.tig.2019.12.002.

Doty, K. R., Guillot-Sestier, M. V. and Town, T. (2015) 'The role of the immune system in neurodegenerative disorders: Adaptive or maladaptive?', *Brain Research*. Elsevier B.V., pp.

155–173. doi: 10.1016/j.brainres.2014.09.008.

Dugger, B. N. and Dickson, D. W. (2017) 'Pathology of neurodegenerative diseases', *Cold Spring Harbor Perspectives in Biology*. Cold Spring Harbor Laboratory Press. doi: 10.1101/cshperspect.a028035.

Durbeej, M. (2010) 'Laminins', *Cell and Tissue Research*. Springer, pp. 259–268. doi: 10.1007/s00441-009-0838-2.

Dusa, A. (2016) *Package 'venn'*. Available at: <https://en.wikipedia.org/wiki/Centroid> (Accessed: 14 August 2020).

Dzwonek, J. and Wilczyński, G. M. (2015) 'CD44: Molecular interactions, signaling and functions in the nervous system', *Frontiers in Cellular Neuroscience*. Frontiers Research Foundation, 9(MAY). doi: 10.3389/fncel.2015.00175.

Elstner, M. *et al.* (2009) 'Single-cell expression profiling of dopaminergic neurons combined with association analysis identifies pyridoxal kinase as Parkinson's disease gene', *Annals of Neurology*. John Wiley and Sons Inc., 66(6), pp. 792–798. doi: 10.1002/ana.21780.

Esteves, A. R. *et al.* (2018) 'Mitochondrial Metabolism Regulates Microtubule Acetylation and Autophagy Through Sirtuin-2: Impact for Parkinson's Disease', *Molecular Neurobiology*. Humana Press Inc., 55(2), pp. 1440–1462. doi: 10.1007/s12035-017-0420-y.

Esteves, A. R. and Cardoso, S. M. (2020) 'Differential protein expression in diverse brain areas of Parkinson's and Alzheimer's disease patients', *Scientific Reports*. Nature Research, 10(1). doi: 10.1038/s41598-020-70174-z.

Farmer, B., Kluemper, J. and Johnson, L. (2019) 'Apolipoprotein E4 Alters Astrocyte Fatty Acid Metabolism and Lipid Droplet Formation', *Cells*. MDPI AG, 8(2), p. 182. doi: 10.3390/cells8020182.

Ferrante, R. J. *et al.* (1987) 'Morphologic and Histochemical Characteristics of a Spared Subset of Striatal Neurons in Huntington's Disease', *Journal of Neuropathology & Experimental Neurology*. Oxford Academic, 46(1), pp. 12–27. doi: 10.1097/00005072-198701000-00002.

Foglio, E. and Luigi Fabrizio, R. (2010) 'Aquaporins and Neurodegenerative Diseases', *Current Neuropharmacology*. Bentham Science Publishers Ltd., 8(2), pp. 112–121. doi: 10.2174/157015910791233150.

Gan, L. *et al.* (2018) 'Converging pathways in neurodegeneration, from genetics to mechanisms', *Nature Neuroscience*. Nature Publishing Group, pp. 1300–1309. doi: 10.1038/s41593-018-0237-7.

Gene Ontology Consortium (2015) 'Gene ontology consortium: going forward', *Nucleic acids research*. Available at: <https://academic.oup.com/nar/article->

abstract/43/D1/D1049/2439067 (Accessed: 14 August 2020).

Ghasemi, M. and Brown, R. H. (2018) 'Genetics of amyotrophic lateral sclerosis', *Cold Spring Harbor Perspectives in Medicine*. Cold Spring Harbor Laboratory Press, 8(5), p. a024125. doi: 10.1101/cshperspect.a024125.

Guareschi, S. *et al.* (2012) 'An over-oxidized form of superoxide dismutase found in sporadic amyotrophic lateral sclerosis with bulbar onset shares a toxic mechanism with mutant SOD1', *Proceedings of the National Academy of Sciences of the United States of America*. National Academy of Sciences, 109(13), pp. 5074–5079. doi: 10.1073/pnas.1115402109.

Gunawardena, S. *et al.* (2003) 'Disruption of axonal transport by loss of huntingtin or expression of pathogenic polyQ proteins in *Drosophila*', *Neuron*. Cell Press, 40(1), pp. 25–40. doi: 10.1016/S0896-6273(03)00594-4.

Guo, F. *et al.* (2018) 'Autophagy in neurodegenerative diseases: pathogenesis and therapy', *Brain Pathology*. Blackwell Publishing Ltd, pp. 3–13. doi: 10.1111/bpa.12545.

Halliday, G. (2017) 'Pathology and hippocampal atrophy in Alzheimer's disease', *The Lancet Neurology*. Lancet Publishing Group, pp. 862–864. doi: 10.1016/S1474-4422(17)30343-5.

Harjes, P. and Wanker, E. E. (2003) 'The hunt for huntingtin function: Interaction partners tell many different stories', *Trends in Biochemical Sciences*. Elsevier Ltd, pp. 425–433. doi: 10.1016/S0968-0004(03)00168-3.

Hashimoto, M. *et al.* (2003) 'Role of Protein Aggregation in Mitochondrial Dysfunction and Neurodegeneration in Alzheimer's and Parkinson's Diseases', *NeuroMolecular Medicine*. Springer, 4(1–2), pp. 21–35. doi: 10.1385/NMM:4:1-2:21.

Heckmann, B. L., Tummers, B. and Green, D. R. (2019) 'Crashing the computer: apoptosis vs. necroptosis in neuroinflammation', *Cell Death and Differentiation*. Nature Publishing Group, pp. 41–52. doi: 10.1038/s41418-018-0195-3.

Heneka, M. T., Kummer, M. P. and Latz, E. (2014) 'Innate immune activation in neurodegenerative disease', *Nature Reviews Immunology*. Nature Publishing Group, pp. 463–477. doi: 10.1038/nri3705.

Hibar, D. P. *et al.* (2015) 'Common genetic variants influence human subcortical brain structures', *Nature*. Nature Publishing Group, 520(7546), pp. 224–229. doi: 10.1038/nature14101.

Hinz, F. I. and Geschwind, D. H. (2017) 'Molecular genetics of neurodegenerative dementias', *Cold Spring Harbor Perspectives in Biology*. Cold Spring Harbor Laboratory Press, 9(4). doi: 10.1101/cshperspect.a023705.

Hussain, R. *et al.* (2018) 'Neurodegenerative diseases: Regenerative mechanisms and novel



therapeutic approaches', *Brain Sciences*. MDPI AG. doi: 10.3390/brainsci8090177.

Iqbal, K. and Grundke-Iqbal, I. (2002) 'Neurofibrillary pathology leads to synaptic loss and not the other way around in Alzheimer disease', *Journal of Alzheimer's Disease*. IOS Press, pp. 235–238. doi: 10.3233/JAD-2002-4313.

Itzhaki, R. F. *et al.* (2004) 'Infiltration of the brain by pathogens causes Alzheimer's disease', *Neurobiology of Aging*. Elsevier Inc., 25(5), pp. 619–627. doi: 10.1016/j.neurobiolaging.2003.12.021.

De Jager, P. L. *et al.* (2014) 'Alzheimer's disease: Early alterations in brain DNA methylation at ANK1, BIN1, RHBDF2 and other loci', *Nature Neuroscience*. Nature Publishing Group, 17(9), pp. 1156–1163. doi: 10.1038/nn.3786.

De Jager, P. L., Yang, H. S. and Bennett, D. A. (2018) 'Deconstructing and targeting the genomic architecture of human neurodegeneration', *Nature Neuroscience*. Nature Publishing Group, pp. 1310–1317. doi: 10.1038/s41593-018-0240-z.

Jankovic, J. (2008) 'Parkinson's disease: Clinical features and diagnosis', *Journal of Neurology, Neurosurgery and Psychiatry*. BMJ Publishing Group, pp. 368–376. doi: 10.1136/jnnp.2007.131045.

Joe, E. and Ringman, J. M. (2019) 'Cognitive symptoms of Alzheimer's disease: Clinical management and prevention', *The BMJ*. BMJ Publishing Group. doi: 10.1136/bmj.l6217.

Johnson, C. D. and Davidson, B. L. (2010) 'Huntington's disease: Progress toward effective disease-modifying treatments and a cure', *Human Molecular Genetics*. Oxford Academic, 19(R1), pp. R98–R102. doi: 10.1093/hmg/ddq148.

Jomova, K. *et al.* (2010) 'Metals, oxidative stress and neurodegenerative disorders', *Molecular and Cellular Biochemistry*. Springer, pp. 91–104. doi: 10.1007/s11010-010-0563-x.

Jones, L. L. *et al.* (2000) 'Regulation of the cell adhesion molecule CD44 after nerve transection and direct trauma to the mouse brain', *Journal of Comparative Neurology*. J Comp Neurol, 426(3), pp. 468–492. doi: 10.1002/1096-9861(20001023)426:3<468::AID-CNE9>3.0.CO;2-I.

Joo, Y. *et al.* (2015) 'Involvement of 14-3-3 in tubulin instability and impaired axon development is mediated by Tau', *The FASEB Journal*. FASEB, 29(10), pp. 4133–4144. doi: 10.1096/fj.14-265009.

Kaltenbach, L. S. *et al.* (2007) 'Huntingtin Interacting Proteins Are Genetic Modifiers of Neurodegeneration', *PLoS Genetics*. Edited by H. Orr. Public Library of Science, 3(5), p. e82. doi: 10.1371/journal.pgen.0030082.

Kampinga, Harm H., S. B. (2016) 'Heat shock proteins as potential targets for protective strategies in neurodegeneration', *The Lancet Neurology*. Available at:

<https://www.sciencedirect.com/science/article/pii/S1474442216000995> (Accessed: 7 October 2020).

Kang, S. H. *et al.* (2013) 'Degeneration and impaired regeneration of gray matter oligodendrocytes in amyotrophic lateral sclerosis', *Nature Neuroscience*. Nature Publishing Group, 16(5), pp. 571–579. doi: 10.1038/nn.3357.

Kassubek, J. *et al.* (2004) 'Topography of cerebral atrophy in early Huntington's disease: A voxel based morphometric MRI study', *Journal of Neurology, Neurosurgery and Psychiatry*. BMJ Publishing Group Ltd, 75(2), pp. 213–220. doi: 10.1136/jnnp.2002.009019.

Kaur, S. J., McKeown, S. R. and Rashid, S. (2016) 'Mutant SOD1 mediated pathogenesis of Amyotrophic Lateral Sclerosis', *Gene*. Elsevier B.V., pp. 109–118. doi: 10.1016/j.gene.2015.11.049.

Kiernan, M. C. *et al.* (2011) 'Amyotrophic lateral sclerosis', in *The Lancet*. Elsevier, pp. 942–955. doi: 10.1016/S0140-6736(10)61156-7.

Kim, D. *et al.* (2018) 'Proteomic change by Korean Red Ginseng in the substantia nigra of a Parkinson's disease mouse model', *Journal of Ginseng Research*. Elsevier B.V., 42(4), pp. 429–435. doi: 10.1016/j.jgr.2017.04.008.

Kim, S., Seo, J. H. and Suh, Y. H. (2004) ' $\alpha$ -Synuclein, Parkinson's disease, and Alzheimer's disease', in *Parkinsonism and Related Disorders*. Elsevier BV, p. S9. doi: 10.1016/j.parkreldis.2003.11.005.

Kinney, J. W. *et al.* (2018) 'Inflammation as a central mechanism in Alzheimer's disease', *Alzheimer's and Dementia: Translational Research and Clinical Interventions*. Elsevier Inc, pp. 575–590. doi: 10.1016/j.trci.2018.06.014.

Knierim, E. *et al.* (2016) 'Mutations in Subunits of the Activating Signal Cointegrator 1 Complex Are Associated with Prenatal Spinal Muscular Atrophy and Congenital Bone Fractures', *American Journal of Human Genetics*. Cell Press, 98(3), pp. 473–489. doi: 10.1016/j.ajhg.2016.01.006.

Knudsen, K. and Borghammer, P. (2018) 'Imaging the Autonomic Nervous System in Parkinson's Disease', *Current Neurology and Neuroscience Reports*. Current Medicine Group LLC 1, pp. 1–13. doi: 10.1007/s11910-018-0889-4.

Koltai, T. (2014) 'Clusterin: a key player in cancer chemoresistance and its inhibition', *OncoTargets and Therapy*. DOVE Medical Press Ltd., 7, p. 447. doi: 10.2147/OTT.S58622.

Kumar, J., Yu, H. and Sheetz, M. P. (1995) 'Kinesin, an essential anchor for kinesin-driven vesicle motility', *Science*. Oxford Univ. Press, 267(5205), pp. 1834–1837. doi: 10.1126/science.7892610.

Kumar, P. *et al.* (2016) 'Ion Channels in Neurological Disorders', in *Advances in Protein*

*Chemistry and Structural Biology*. Academic Press Inc., pp. 97–136. doi: 10.1016/bs.apcsb.2015.10.006.

Lagier-Tourenne, C., Polymenidou, M. and Cleveland, D. W. (2010) 'TDP-43 and FUS/TLS: Emerging roles in RNA processing and neurodegeneration', *Human Molecular Genetics*. Oxford University Press, 19(R1), p. R46. doi: 10.1093/hmg/ddq137.

Li, M. and Lester, H. A. (2001) 'Ion channel diseases of the central nervous system', *CNS Drug Reviews*. Neva Press Inc., pp. 214–240. doi: 10.1111/j.1527-3458.2001.tb00196.x.

Li, P., Nie, Y. and Yu, J. (2015) 'An Effective Method to Identify Shared Pathways and Common Factors among Neurodegenerative Diseases', *PLOS ONE*. Edited by M. R. Cookson. Public Library of Science, 10(11), p. e0143045. doi: 10.1371/journal.pone.0143045.

Liao, Y. *et al.* (2019) 'WebGestalt 2019: gene set analysis toolkit with revamped UIs and APIs', *Nucleic Acids Research*. Oxford University Press, 47(W1), pp. W199–W205. doi: 10.1093/nar/gkz401.

Liscovitch, N. and French, L. (2014) 'Differential co-expression between  $\alpha$ -synuclein and IFN- $\gamma$  signaling genes across development and in parkinson's disease', *PLoS ONE*. Public Library of Science, 9(12). doi: 10.1371/journal.pone.0115029.

Liu, C. C. *et al.* (2013) 'Apolipoprotein e and Alzheimer disease: Risk, mechanisms and therapy', *Nature Reviews Neurology*. Nat Rev Neurol, pp. 106–118. doi: 10.1038/nrneurol.2012.263.

Lukacs, M. *et al.* (2019) 'Severe biallelic loss-of-function mutations in nicotinamide mononucleotide adenylyltransferase 2 (NMNAT2) in two fetuses with fetal akinesia deformation sequence', *Experimental Neurology*. Academic Press Inc., 320. doi: 10.1016/j.expneurol.2019.112961.

Lunnon, K. *et al.* (2014) 'Methylomic profiling implicates cortical deregulation of ANK1 in Alzheimer's disease', *Nature Neuroscience*. Nature Publishing Group, 17(9), pp. 1164–1170. doi: 10.1038/nn.3782.

M Kanehisa - Novartis Foundation Symposium (2002) 'The KEGG database', *Wiley Online Library*. Available at: <https://onlinelibrary.wiley.com/doi/pdf/10.1002/0470857897#page=99> (Accessed: 14 August 2020).

Ma, B. *et al.* (2016) 'Deletion of the hemopexin or heme oxygenase-2 gene aggravates brain injury following stroma-free hemoglobin-induced intracerebral hemorrhage', *Journal of Neuroinflammation*. BioMed Central Ltd., 13(1), p. 26. doi: 10.1186/s12974-016-0490-1.

Ma, R. hong *et al.* (2017) 'Role of microtubule-associated protein tau phosphorylation in Alzheimer's disease', *Journal of Huazhong University of Science and Technology - Medical Science*. Tongji Medical University, pp. 307–312. doi: 10.1007/s11596-017-1732-x.

Mackay, J., Nassrallah, W. and Raymond, L. (2018) 'Cause or compensation?-Altered neuronal Ca<sup>2+</sup> handling in Huntington's disease', *CNS Neuroscience & Therapeutics*, 24(4), pp. 301–310. doi: 10.1111/cns.2018.24.issue-4.

Mahalingam, S. and Levy, L. M. (2014) 'Genetics of Huntington disease', *American Journal of Neuroradiology*. American Society of Neuroradiology, pp. 1070–1072. doi: 10.3174/ajnr.A3772.

Maheshwari, P. and Eslick, G. D. (2015) 'Bacterial infection and Alzheimer's disease: A meta-analysis', *Journal of Alzheimer's Disease*. IOS Press, 43(3), pp. 957–966. doi: 10.3233/JAD-140621.

Mandelkow, E. M. and Mandelkow, E. (1998) 'Tau in Alzheimer's disease', *Trends in Cell Biology*. Elsevier Ltd, 8(11), pp. 425–427. doi: 10.1016/S0962-8924(98)01368-3.

Manzoni, C., Lewis, P. A. and Ferrari, R. (2020) 'Network Analysis for Complex Neurodegenerative Diseases', *Current Genetic Medicine Reports*. Springer Science and Business Media LLC, 8(1), pp. 17–25. doi: 10.1007/s40142-020-00181-z.

Mao, Q. *et al.* (2020) 'KTN1 Variants Underlying Putamen Gray Matter Volumes and Parkinson's Disease', *Frontiers in Neuroscience*. Frontiers Media S.A., 14, p. 651. doi: 10.3389/fnins.2020.00651.

Martin, D. D. O. *et al.* (2015) 'Autophagy in Huntington disease and huntingtin in autophagy', *Trends in Neurosciences*. Elsevier Ltd, pp. 26–35. doi: 10.1016/j.tins.2014.09.003.

Matzke, A. *et al.* (2007) 'Haploinsufficiency of c-Met in cd44<sup>-/-</sup> Mice Identifies a Collaboration of CD44 and c-Met In Vivo', *Molecular and Cellular Biology*. American Society for Microbiology, 27(24), pp. 8797–8806. doi: 10.1128/mcb.01355-07.

McKenzie, J. L., Dalchau, R. and Fabre, J. W. (1982) 'Biochemical Characterisation and Localization in Brain of a Human Brain-Leucocyte Membrane Glycoprotein Recognised by a Monoclonal Antibody', *Journal of Neurochemistry*. John Wiley & Sons, Ltd, 39(5), pp. 1461–1466. doi: 10.1111/j.1471-4159.1982.tb12592.x.

McQuade, L. R. *et al.* (2014) 'Proteomics of Huntington's disease-affected human embryonic stem cells reveals an evolving pathology involving mitochondrial dysfunction and metabolic disturbances', *Journal of Proteome Research*. American Chemical Society, 13(12), pp. 5648–5659. doi: 10.1021/pr500649m.

Misawa, H. *et al.* (2012) 'Osteopontin is an alpha motor neuron marker in the mouse spinal cord', *Journal of Neuroscience Research*. John Wiley & Sons, Ltd, 90(4), pp. 732–742. doi: 10.1002/jnr.22813.

Mitchell, J. and Borasio, G. (2007) 'Amyotrophic lateral sclerosis', *Lancet*. Elsevier, pp. 2031–2041. doi: 10.1016/S0140-6736(07)60944-1.

- Mousavi, S. V., Agah, E. and Tafakhori, A. (2020) 'The Role of Osteopontin in Amyotrophic Lateral Sclerosis: A Systematic Review', *Archives of Neuroscience*. Kowsar Medical Institute, In Press(In Press). doi: 10.5812/ans.94205.
- Myers, R. H. *et al.* (1988) 'Clinical and neuropathologic assessment of severity in huntington's disease', *Neurology*. Wolters Kluwer Health, Inc. on behalf of the American Academy of Neurology, 38(3), pp. 341–347. doi: 10.1212/wnl.38.3.341.
- Nguyen, N. D. and Wang, D. (2020) 'Multiview learning for understanding functional multiomics', *PLoS Computational Biology*. Public Library of Science, p. e1007677. doi: 10.1371/journal.pcbi.1007677.
- Niu, H. *et al.* (2018) 'Alpha-synuclein overexpression in the olfactory bulb initiates prodromal symptoms and pathology of Parkinson's disease', *Translational Neurodegeneration*. BioMed Central Ltd., 7(1), p. 25. doi: 10.1186/s40035-018-0128-6.
- Novellino, F. *et al.* (2020) 'Innate Immunity: A Common Denominator between Neurodegenerative and Neuropsychiatric Diseases', *International Journal of Molecular Sciences*. MDPI AG, 21(3), p. 1115. doi: 10.3390/ijms21031115.
- O'Gorman Tuura, R. L., Baumann, C. R. and Baumann-Vogel, H. (2018) 'Beyond Dopamine: GABA, Glutamate, and the Axial Symptoms of Parkinson Disease', *Frontiers in Neurology*. Frontiers Media S.A., 9(SEP), p. 806. doi: 10.3389/fneur.2018.00806.
- Ochaba, J. *et al.* (2014) 'Potential function for the Huntingtin protein as a scaffold for selective autophagy', *Proceedings of the National Academy of Sciences of the United States of America*. National Academy of Sciences, 111(47), pp. 16889–16894. doi: 10.1073/pnas.1420103111.
- Papanikolaou, G. and Pantopoulos, K. (2005) 'Iron metabolism and toxicity', *Toxicology and Applied Pharmacology*. Toxicol Appl Pharmacol, pp. 199–211. doi: 10.1016/j.taap.2004.06.021.
- Papatheodorou, I. *et al.* (2018) 'Expression Atlas: Gene and protein expression across multiple studies and organisms', *Nucleic Acids Research*. Oxford University Press, 46(D1), pp. D246–D251. doi: 10.1093/nar/gkx1158.
- Perdigão, C. *et al.* (2020) 'Intracellular Trafficking Mechanisms of Synaptic Dysfunction in Alzheimer's Disease', *Frontiers in Cellular Neuroscience*. Frontiers Media S.A. doi: 10.3389/fncel.2020.00072.
- Philips, T. and Rothstein, J. D. (2014) 'Glial cells in amyotrophic lateral sclerosis', *Experimental Neurology*. Academic Press Inc., pp. 111–120. doi: 10.1016/j.expneurol.2014.05.015.
- Pomiès, P., Louis, H. A. and Beckerle, M. C. (1997) 'CRP1, a LIM domain protein implicated in muscle differentiation, interacts with  $\alpha$ -actinin', *Journal of Cell Biology*. The



Rockefeller University Press, 139(1), pp. 157–168. doi: 10.1083/jcb.139.1.157.

Prasad, A. *et al.* (2019) 'Molecular mechanisms of TDP-43 misfolding and pathology in amyotrophic lateral sclerosis', *Frontiers in Molecular Neuroscience*. Frontiers Media S.A., p. 25. doi: 10.3389/fnmol.2019.00025.

Pringsheim, T. *et al.* (2012) 'The incidence and prevalence of Huntington's disease: A systematic review and meta-analysis', *Movement Disorders*. John Wiley & Sons, Ltd, pp. 1083–1091. doi: 10.1002/mds.25075.

Pyatigorskaya, N. *et al.* (2016) 'Medulla oblongata damage and cardiac autonomic dysfunction in Parkinson disease', *Neurology*. Lippincott Williams and Wilkins, 87(24), pp. 2540–2545. doi: 10.1212/WNL.0000000000003426.

Rademakers, R. and Rovelet-Lecrux, A. (2009) 'Recent insights into the molecular genetics of dementia', *Trends in Neurosciences*. NIH Public Access, pp. 451–461. doi: 10.1016/j.tins.2009.05.005.

Ramanan, V. K. *et al.* (2014) 'APOE and BCHE as modulators of cerebral amyloid deposition: A florbetapir PET genome-wide association study', *Molecular Psychiatry*. NIH Public Access, 19(3), pp. 351–357. doi: 10.1038/mp.2013.19.

Ramanan, V. K. and Saykin, A. J. (2013) 'Pathways to neurodegeneration: mechanistic insights from GWAS in Alzheimer's disease, Parkinson's disease, and related disorders', *Am J Neurodegener Dis*, 2(3), pp. 145–175. Available at: www.AJND.us.

Reddy, P. H. and Shirendeb, U. P. (2012) 'Mutant huntingtin, abnormal mitochondrial dynamics, defective axonal transport of mitochondria, and selective synaptic degeneration in Huntington's disease', *Biochimica et Biophysica Acta - Molecular Basis of Disease*. Elsevier, pp. 101–110. doi: 10.1016/j.bbdis.2011.10.016.

Reed, X. *et al.* (2019) 'The role of monogenic genes in idiopathic Parkinson's disease', *Neurobiology of Disease*. Academic Press Inc., pp. 230–239. doi: 10.1016/j.nbd.2018.11.012.

Renton, A. E., Chiò, A. and Traynor, B. J. (2014) 'State of play in amyotrophic lateral sclerosis genetics', *Nature Neuroscience*. Nature Publishing Group, pp. 17–23. doi: 10.1038/nn.3584.

Robberecht, W. and Philips, T. (2013) 'The changing scene of amyotrophic lateral sclerosis', *Nature Reviews Neuroscience*. Nature Publishing Group, pp. 248–264. doi: 10.1038/nrn3430.

Roos, R. A. C. (2010) 'Huntington's disease: A clinical review', *Orphanet Journal of Rare Diseases*. BioMed Central, p. 40. doi: 10.1186/1750-1172-5-40.

Ross, C. A. and Tabrizi, S. J. (2011) 'Huntington's disease: From molecular pathogenesis to clinical treatment', *The Lancet Neurology*. Elsevier, pp. 83–98. doi: 10.1016/S1474-4422(10)70245-3.



Roze, E. *et al.* (2008) 'Mitogen- and stress-activated protein kinase-1 deficiency is involved in expanded-huntingtin-induced transcriptional dysregulation and striatal death', *The FASEB Journal*. Wiley, 22(4), pp. 1083–1093. doi: 10.1096/fj.07-9814.

Rubinsztein, D. C. (2006) 'The roles of intracellular protein-degradation pathways in neurodegeneration', *Nature*. Nature Publishing Group, pp. 780–786. doi: 10.1038/nature05291.

Savage, A. L. *et al.* (2018) 'Retrotransposons in the development and progression of amyotrophic lateral sclerosis', *Journal of Neurology, Neurosurgery and Psychiatry*. BMJ Publishing Group, 90(3), pp. 284–293. doi: 10.1136/jnnp-2018-319210.

Scheltens, P. *et al.* (2016) 'Alzheimer's disease', *The Lancet*. Lancet Publishing Group, pp. 505–517. doi: 10.1016/S0140-6736(15)01124-1.

Serrano-Pozo, A. *et al.* (2011) 'Neuropathological alterations in Alzheimer disease', *Cold Spring Harbor Perspectives in Medicine*. Cold Spring Harb Perspect Med, 1(1). doi: 10.1101/cshperspect.a006189.

Sheng, M., Sabatini, B. L. and Südhof, T. C. (2012) 'Synapses and Alzheimer's disease', *Cold Spring Harbor Perspectives in Biology*. Cold Spring Harbor Laboratory Press, 4(5), p. 10. doi: 10.1101/cshperspect.a005777.

Silva, D. F. *et al.* (2017) 'Mitochondrial Metabolism Power SIRT2-Dependent Deficient Traffic Causing Alzheimer's-Disease Related Pathology', *Molecular Neurobiology*. Humana Press Inc., 54(6), pp. 4021–4040. doi: 10.1007/s12035-016-9951-x.

Skupien, A. *et al.* (2014) 'CD44 regulates dendrite morphogenesis through Src tyrosine kinase-dependent positioning of the Golgi', *Journal of cell science*. J Cell Sci, 127(23), pp. 5038–5051. doi: 10.1242/jcs.154542.

Smith, A. and McCulloh, R. J. (2015) 'Hemopexin and haptoglobin: Allies against heme toxicity from hemoglobin not contenders', *Frontiers in Physiology*. Frontiers Media S.A., p. 187. doi: 10.3389/fphys.2015.00187.

Strang, K. H., Golde, T. E. and Giasson, B. I. (2019) 'MAPT mutations, tauopathy, and mechanisms of neurodegeneration', *Laboratory Investigation*. Nature Publishing Group, pp. 912–928. doi: 10.1038/s41374-019-0197-x.

Tam, O. H. *et al.* (2019) 'Postmortem Cortex Samples Identify Distinct Molecular Subtypes of ALS: Retrotransposon Activation, Oxidative Stress, and Activated Glia', *Cell Reports*. Elsevier B.V., 29(5), pp. 1164–1177.e5. doi: 10.1016/j.celrep.2019.09.066.

Tan, E.-K. *et al.* (2005) 'Functional COMT variant predicts response to high dose pyridoxine in Parkinson's disease', *American Journal of Medical Genetics Part B: Neuropsychiatric Genetics*. John Wiley & Sons, Ltd, 137B(1), pp. 1–4. doi:

10.1002/ajmg.b.30198.

Tang, T. S. *et al.* (2007) 'Dopaminergic signaling and striatal neurodegeneration in Huntington's disease', *Journal of Neuroscience*. Society for Neuroscience, 27(30), pp. 7899–7910. doi: 10.1523/JNEUROSCI.1396-07.2007.

Toyoshima, I. and Sheetz, M. P. (1996) 'Kinectin distribution in chicken nervous system', *Neuroscience Letters*. Elsevier Ireland Ltd, 211(3), pp. 171–174. doi: 10.1016/0304-3940(96)12752-X.

Turner, M. R. *et al.* (2011) 'Concordance between site of onset and limb dominance in amyotrophic lateral sclerosis', *Journal of Neurology, Neurosurgery and Psychiatry*. BMJ Publishing Group Ltd, 82(8), pp. 853–854. doi: 10.1136/jnnp.2010.208413.

Umahara, T. *et al.* (2004) '14-3-3 proteins and zeta isoform containing neurofibrillary tangles in patients with Alzheimer's disease', *Acta Neuropathologica*. Springer, 108(4), pp. 279–286. doi: 10.1007/s00401-004-0885-4.

Vedunova, M. *et al.* (2013) 'Seizure-like activity in hyaluronidase-treated dissociated hippocampal cultures', *Frontiers in Cellular Neuroscience*. Front Cell Neurosci, 7(SEP). doi: 10.3389/fncel.2013.00149.

Walker, F. O. (2007) 'Huntington's disease', *Lancet*. Elsevier, pp. 218–228. doi: 10.1016/S0140-6736(07)60111-1.

Wang, J. Z. and Liu, F. (2008) 'Microtubule-associated protein tau in development, degeneration and protection of neurons', *Progress in Neurobiology*. Pergamon, pp. 148–175. doi: 10.1016/j.pneurobio.2008.03.002.

Wanker, E. E. *et al.* (2019) 'The pathobiology of perturbed mutant huntingtin protein–protein interactions in Huntington's disease', *Journal of Neurochemistry*. Blackwell Publishing Ltd, 151(4), pp. 507–519. doi: 10.1111/jnc.14853.

Wijesekera, L. C. and Leigh, P. N. (2009) 'Amyotrophic lateral sclerosis', *Orphanet Journal of Rare Diseases*. BioMed Central, 4(1), p. 3. doi: 10.1186/1750-1172-4-3.

Wright Willis, A. *et al.* (2010) 'Geographic and ethnic variation in Parkinson disease: A population-based study of us medicare beneficiaries', *Neuroepidemiology*. Neuroepidemiology, 34(3), pp. 143–151. doi: 10.1159/000275491.

Wyss-Coray, T. (2006) 'Inflammation in Alzheimer disease: Driving force, bystander or beneficial response?', *Nature Medicine*. Nature Publishing Group, pp. 1005–1015. doi: 10.1038/nm1484.

Xie, T. *et al.* (2014) 'A genome-wide association study combining pathway analysis for typical sporadic amyotrophic lateral sclerosis in Chinese Han populations', *Neurobiology of Aging*. Elsevier Inc., 35(7), pp. 1778.e9–1778.e23. doi: 10.1016/j.neurobiolaging.2014.01.014.

Yamamoto, T. *et al.* (2017) 'Expression of secreted phosphoprotein 1 (osteopontin) in human sensorimotor cortex and spinal cord: Changes in patients with amyotrophic lateral sclerosis', *Brain Research*. Elsevier B.V., 1655, pp. 168–175. doi: 10.1016/j.brainres.2016.10.030.

Yu, X. *et al.* (2020) 'Protein-protein interaction network with machine learning models and multiomics data reveal potential neurodegenerative disease-related proteins', *Human Molecular Genetics*. Oxford University Press, 29(8), pp. 1378–1387. doi: 10.1093/hmg/ddaa065.

Yurchenco, P. and Patton, B. (2009) 'Developmental and Pathogenic Mechanisms of Basement Membrane Assembly', *Current Pharmaceutical Design*. Bentham Science Publishers Ltd., 15(12), pp. 1277–1294. doi: 10.2174/138161209787846766.

Zhai, R. G. *et al.* (2008) 'NAD synthase NMNAT acts as a chaperone to protect against neurodegeneration', *Nature*. Nature Publishing Group, 452(7189), pp. 887–891. doi: 10.1038/nature06721.

Zhang, X. *et al.* (2010) 'Kinectin-mediated endoplasmic reticulum dynamics supports focal adhesion growth in the cellular lamella', *Journal of Cell Science*. The Company of Biologists Ltd, 123(22), pp. 3901–3912. doi: 10.1242/jcs.069153.

Zhang, X., Wan, J.-Q. and Tong, X.-P. (2018) 'Potassium channel dysfunction in neurons and astrocytes in Huntington's disease', *CNS Neuroscience & Therapeutics*. Blackwell Publishing Ltd, 24(4), pp. 311–318. doi: 10.1111/cns.12804.

Agah, E. *et al.* (2018) 'Osteopontin (OPN) as a CSF and blood biomarker for multiple sclerosis: A systematic review and meta-analysis', *PLOS ONE*. Edited by H. Wiendl. Public Library of Science, 13(1), p. e0190252. doi: 10.1371/journal.pone.0190252.

Alexander, G. E. (2004) 'Biology of Parkinson's disease: Pathogenesis and pathophysiology of a multisystem neurodegenerative disorder', *Dialogues in Clinical Neuroscience*. Les Laboratoires Servier, pp. 259–280. Available at: [www.dialogues-cns.org](http://www.dialogues-cns.org) (Accessed: 31 August 2020).

Ali, Y. O. *et al.* (2016) 'NMNAT2:HSP90 Complex Mediates Proteostasis in Proteinopathies', *PLoS Biology*. Public Library of Science, 14(6), p. 1002472. doi: 10.1371/journal.pbio.1002472.

Arendt, T., Stieler, J. T. and Holzer, M. (2016) 'Tau and tauopathies', *Brain Research Bulletin*. Elsevier Inc., pp. 238–292. doi: 10.1016/j.brainresbull.2016.08.018.

Arneson, D. *et al.* (2018) 'Shared mechanisms among neurodegenerative diseases: from genetic factors to gene networks', *Journal of Genetics*. Springer, pp. 795–806. doi: 10.1007/s12041-018-0963-3.

- Ashraf, A. *et al.* (2020) 'Plasma transferrin and hemopexin are associated with altered A $\beta$  uptake and cognitive decline in Alzheimer's disease pathology', *Alzheimer's research & therapy*. NLM (Medline), 12(1), p. 72. doi: 10.1186/s13195-020-00634-1.
- Aylward, E. H. *et al.* (2004) 'Onset and rate of striatal atrophy in preclinical Huntington disease', *Neurology*. Lippincott Williams and Wilkins, 63(1), pp. 66–72. doi: 10.1212/01.WNL.0000132965.14653.D1.
- Badaut, J. *et al.* (2014) 'Aquaporin and brain diseases', *Biochimica et Biophysica Acta - General Subjects*. Elsevier, pp. 1554–1565. doi: 10.1016/j.bbagen.2013.10.032.
- Ballard, C. *et al.* (2011) 'Alzheimer's disease', *The Lancet*. Lancet Publishing Group, pp. 1019–1031. doi: 10.1016/S0140-6736(10)61349-9.
- Barrett, T. *et al.* (2013) 'NCBI GEO: Archive for functional genomics data sets - Update', *Nucleic Acids Research*. Oxford Academic, 41(D1), pp. D991–D995. doi: 10.1093/nar/gks1193.
- Bayraktar, A. *et al.* (2019) 'Meta-analysis of Gene Expression in Neurodegenerative Diseases Reveals Patterns in GABA Synthesis and Heat Stress Pathways'. Available at: <http://arxiv.org/abs/1909.07469> (Accessed: 7 October 2020).
- Becker, K. *et al.* (2018) 'Quantifying post-transcriptional regulation in the development of *Drosophila melanogaster*', *Nature Communications*. Nature Research, 9(1), pp. 1–14. doi: 10.1038/s41467-018-07455-9.
- Bellenguez, C., Grenier-Boley, B. and Lambert, J. C. (2020) 'Genetics of Alzheimer's disease: where we are, and where we are going', *Current Opinion in Neurobiology*. Elsevier Ltd, pp. 40–48. doi: 10.1016/j.conb.2019.11.024.
- Bennett, J. P. and Keeney, P. M. (2018) 'RNA-Sequencing Reveals Similarities and Differences in Gene Expression in Vulnerable Brain Tissues of Alzheimer's and Parkinson's Diseases', *Journal of Alzheimer's Disease Reports*. IOS Press, 2(1), pp. 129–137. doi: 10.3233/adr-180072.
- Berardelli, A. *et al.* (2001) 'Pathophysiology of bradykinesia in parkinson's disease', *Brain*. Oxford University Press, pp. 2131–2146. doi: 10.1093/brain/124.11.2131.
- Berg, D., Holzmann, C. and Riess, O. (2003) '14-3-3 proteins in the nervous system', *Nature Reviews Neuroscience*. Nature Publishing Group, 4(9), pp. 752–762. doi: 10.1038/nrn1197.
- Berridge, M. J. (2014) 'Calcium regulation of neural rhythms, memory and alzheimer's disease', *Journal of Physiology*. Wiley-Blackwell, 592(2), pp. 281–293. doi: 10.1113/jphysiol.2013.257527.
- Bettens, K. *et al.* (2015) 'Reduced secreted clusterin as a mechanism for Alzheimer-associated CLU mutations', *Molecular Neurodegeneration*. BioMed Central Ltd., 10(1), p. 30. doi: 10.1186/s13024-015-0024-9.

- Boston, P. F., Jackson, P. and Thompson, R. J. (1982) 'Human 14-3-3 Protein: Radioimmunoassay, Tissue Distribution, and Cerebrospinal Fluid Levels in Patients with Neurological Disorders', *Journal of Neurochemistry*. John Wiley & Sons, Ltd, 38(5), pp. 1475–1482. doi: 10.1111/j.1471-4159.1982.tb07928.x.
- Braak, H. *et al.* (2003) 'Staging of brain pathology related to sporadic Parkinson's disease', *Neurobiology of Aging*. Elsevier, 24(2), pp. 197–211. doi: 10.1016/S0197-4580(02)00065-9.
- Braak, H. and Braak, E. (1991) 'Neuropathological staging of Alzheimer-related changes', *Acta Neuropathologica*. Springer-Verlag, pp. 239–259. doi: 10.1007/BF00308809.
- Brandt, J. (2018) 'Behavioral Changes in Huntington Disease', *Cognitive and Behavioral Neurology*. Lippincott Williams and Wilkins, pp. 26–27. doi: 10.1097/WNN.0000000000000147.
- Braschi, B. *et al.* (2019) 'Genenames.org: The HGNC and VGNC resources in 2019', *Nucleic Acids Research*. Oxford University Press, 47(D1), pp. D786–D792. doi: 10.1093/nar/gky930.
- Bredesen, D. E., Rao, R. V. and Mehlen, P. (2006) 'Cell death in the nervous system', *Nature*. Nature Publishing Group, pp. 796–802. doi: 10.1038/nature05293.
- Briston, T. and Hicks, A. R. (2018) 'Mitochondrial dysfunction and neurodegenerative proteinopathies: Mechanisms and prospects for therapeutic intervention', *Biochemical Society Transactions*. Portland Press Ltd, pp. 829–842. doi: 10.1042/BST20180025.
- Cabral-Miranda, F. and Hetz, C. (2018) 'ER stress and neurodegenerative disease: A cause or effect relationship?', in *Current Topics in Microbiology and Immunology*. Springer Verlag, pp. 131–157. doi: 10.1007/82\_2017\_52.
- Carecchio, M. and Comi, C. (2011) 'The role of osteopontin in neurodegenerative diseases', *Journal of Alzheimer's Disease*. IOS Press, pp. 179–185. doi: 10.3233/JAD-2011-102151.
- Castaño, E. M. *et al.* (2006) 'Comparative proteomics of cerebrospinal fluid in neuropathologically- confirmed Alzheimer's disease and non-demented elderly subjects', *Neurological Research*. Neurol Res, 28(2), pp. 155–163. doi: 10.1179/016164106X98035.
- Cattaneo, E., Zuccato, C. and Tartari, M. (2005) 'Normal huntingtin function: An alternative approach to Huntington's disease', *Nature Reviews Neuroscience*. Nature Publishing Group, pp. 919–930. doi: 10.1038/nrn1806.
- Cerasa, A., Novellino, F. and Quattrone, A. (2016) 'Connectivity Changes in Parkinson's Disease', *Current Neurology and Neuroscience Reports*. Current Medicine Group LLC 1, pp. 1–11. doi: 10.1007/s11910-016-0687-9.
- Cha, J. H. J. (2000) 'Transcriptional dysregulation in Huntington's disease', *Trends in Neurosciences*. Elsevier Ltd, pp. 387–392. doi: 10.1016/S0166-2236(00)01609-X.



Cistaro, A. *et al.* (2012) 'Brain hypermetabolism in amyotrophic lateral sclerosis: A FDG PET study in ALS of spinal and bulbar onset', *European Journal of Nuclear Medicine and Molecular Imaging*. Springer Verlag, 39(2), pp. 251–259. doi: 10.1007/s00259-011-1979-6.

Dalhaimer, P. (2019) 'Lipid Droplets in Disease', *Cells*. NLM (Medline), p. 974. doi: 10.3390/cells8090974.

Damiano, M. *et al.* (2010) 'Mitochondria in Huntington's disease', *Biochimica et Biophysica Acta - Molecular Basis of Disease*. Elsevier, pp. 52–61. doi: 10.1016/j.bbadis.2009.07.012.

Dauer, W. and Przedborski, S. (2003) 'Parkinson's disease: Mechanisms and models', *Neuron*. Cell Press, pp. 889–909. doi: 10.1016/S0896-6273(03)00568-3.

DeMattos, R. B. *et al.* (2004) 'ApoE and Clusterin Cooperatively Suppress A $\beta$  Levels and Deposition: Evidence that ApoE Regulates Extracellular A $\beta$  Metabolism In Vivo', *Neuron*. Cell Press, 41(2), pp. 193–202. doi: 10.1016/S0896-6273(03)00850-X.

Diaz-Ortiz, M. E. and Chen-Plotkin, A. S. (2020) 'Omics in Neurodegenerative Disease: Hope or Hype?', *Trends in Genetics*. Elsevier Ltd, pp. 152–159. doi: 10.1016/j.tig.2019.12.002.

Doty, K. R., Guillot-Sestier, M. V. and Town, T. (2015) 'The role of the immune system in neurodegenerative disorders: Adaptive or maladaptive?', *Brain Research*. Elsevier B.V., pp. 155–173. doi: 10.1016/j.brainres.2014.09.008.

Dugger, B. N. and Dickson, D. W. (2017) 'Pathology of neurodegenerative diseases', *Cold Spring Harbor Perspectives in Biology*. Cold Spring Harbor Laboratory Press. doi: 10.1101/cshperspect.a028035.

Durbeej, M. (2010) 'Laminins', *Cell and Tissue Research*. Springer, pp. 259–268. doi: 10.1007/s00441-009-0838-2.

Dusa, A. (2016) *Package 'venn'*. Available at: <https://en.wikipedia.org/wiki/Centroid> (Accessed: 14 August 2020).

Dzwonek, J. and Wilczyński, G. M. (2015) 'CD44: Molecular interactions, signaling and functions in the nervous system', *Frontiers in Cellular Neuroscience*. Frontiers Research Foundation, 9(MAY). doi: 10.3389/fncel.2015.00175.

Elstner, M. *et al.* (2009) 'Single-cell expression profiling of dopaminergic neurons combined with association analysis identifies pyridoxal kinase as Parkinson's disease gene', *Annals of Neurology*. John Wiley and Sons Inc., 66(6), pp. 792–798. doi: 10.1002/ana.21780.

Esteves, A. R. *et al.* (2018) 'Mitochondrial Metabolism Regulates Microtubule Acetylation and Autophagy Through Sirtuin-2: Impact for Parkinson's Disease', *Molecular Neurobiology*. Humana Press Inc., 55(2), pp. 1440–1462. doi: 10.1007/s12035-017-0420-y.

Esteves, A. R. and Cardoso, S. M. (2020) 'Differential protein expression in diverse brain

areas of Parkinson's and Alzheimer's disease patients', *Scientific Reports*. Nature Research, 10(1). doi: 10.1038/s41598-020-70174-z.

Farmer, B., Kluemper, J. and Johnson, L. (2019) 'Apolipoprotein E4 Alters Astrocyte Fatty Acid Metabolism and Lipid Droplet Formation', *Cells*. MDPI AG, 8(2), p. 182. doi: 10.3390/cells8020182.

Ferrante, R. J. *et al.* (1987) 'Morphologic and Histochemical Characteristics of a Spared Subset of Striatal Neurons in Huntington's Disease', *Journal of Neuropathology & Experimental Neurology*. Oxford Academic, 46(1), pp. 12–27. doi: 10.1097/00005072-198701000-00002.

Foglio, E. and Luigi Fabrizio, R. (2010) 'Aquaporins and Neurodegenerative Diseases', *Current Neuroparmacology*. Bentham Science Publishers Ltd., 8(2), pp. 112–121. doi: 10.2174/157015910791233150.

Gan, L. *et al.* (2018) 'Converging pathways in neurodegeneration, from genetics to mechanisms', *Nature Neuroscience*. Nature Publishing Group, pp. 1300–1309. doi: 10.1038/s41593-018-0237-7.

Gene Ontology Consortium (2015) 'Gene ontology consortium: going forward', *Nucleic acids research*. Available at: <https://academic.oup.com/nar/article-abstract/43/D1/D1049/2439067> (Accessed: 14 August 2020).

Ghasemi, M. and Brown, R. H. (2018) 'Genetics of amyotrophic lateral sclerosis', *Cold Spring Harbor Perspectives in Medicine*. Cold Spring Harbor Laboratory Press, 8(5), p. a024125. doi: 10.1101/cshperspect.a024125.

Guareschi, S. *et al.* (2012) 'An over-oxidized form of superoxide dismutase found in sporadic amyotrophic lateral sclerosis with bulbar onset shares a toxic mechanism with mutant SOD1', *Proceedings of the National Academy of Sciences of the United States of America*. National Academy of Sciences, 109(13), pp. 5074–5079. doi: 10.1073/pnas.1115402109.

Gunawardena, S. *et al.* (2003) 'Disruption of axonal transport by loss of huntingtin or expression of pathogenic polyQ proteins in *Drosophila*', *Neuron*. Cell Press, 40(1), pp. 25–40. doi: 10.1016/S0896-6273(03)00594-4.

Guo, F. *et al.* (2018) 'Autophagy in neurodegenerative diseases: pathogenesis and therapy', *Brain Pathology*. Blackwell Publishing Ltd, pp. 3–13. doi: 10.1111/bpa.12545.

Halliday, G. (2017) 'Pathology and hippocampal atrophy in Alzheimer's disease', *The Lancet Neurology*. Lancet Publishing Group, pp. 862–864. doi: 10.1016/S1474-4422(17)30343-5.

Harjes, P. and Wanker, E. E. (2003) 'The hunt for huntingtin function: Interaction partners tell many different stories', *Trends in Biochemical Sciences*. Elsevier Ltd, pp. 425–433. doi:

10.1016/S0968-0004(03)00168-3.

Hashimoto, M. *et al.* (2003) 'Role of Protein Aggregation in Mitochondrial Dysfunction and Neurodegeneration in Alzheimer's and Parkinson's Diseases', *NeuroMolecular Medicine*. Springer, 4(1–2), pp. 21–35. doi: 10.1385/NMM:4:1-2:21.

Heckmann, B. L., Tummers, B. and Green, D. R. (2019) 'Crashing the computer: apoptosis vs. necroptosis in neuroinflammation', *Cell Death and Differentiation*. Nature Publishing Group, pp. 41–52. doi: 10.1038/s41418-018-0195-3.

Heneka, M. T., Kummer, M. P. and Latz, E. (2014) 'Innate immune activation in neurodegenerative disease', *Nature Reviews Immunology*. Nature Publishing Group, pp. 463–477. doi: 10.1038/nri3705.

Hibar, D. P. *et al.* (2015) 'Common genetic variants influence human subcortical brain structures', *Nature*. Nature Publishing Group, 520(7546), pp. 224–229. doi: 10.1038/nature14101.

Hinz, F. I. and Geschwind, D. H. (2017) 'Molecular genetics of neurodegenerative dementias', *Cold Spring Harbor Perspectives in Biology*. Cold Spring Harbor Laboratory Press, 9(4). doi: 10.1101/cshperspect.a023705.

Hussain, R. *et al.* (2018) 'Neurodegenerative diseases: Regenerative mechanisms and novel therapeutic approaches', *Brain Sciences*. MDPI AG. doi: 10.3390/brainsci8090177.

Iqbal, K. and Grundke-Iqbal, I. (2002) 'Neurofibrillary pathology leads to synaptic loss and not the other way around in Alzheimer disease', *Journal of Alzheimer's Disease*. IOS Press, pp. 235–238. doi: 10.3233/JAD-2002-4313.

Itzhaki, R. F. *et al.* (2004) 'Infiltration of the brain by pathogens causes Alzheimer's disease', *Neurobiology of Aging*. Elsevier Inc., 25(5), pp. 619–627. doi: 10.1016/j.neurobiolaging.2003.12.021.

De Jager, P. L. *et al.* (2014) 'Alzheimer's disease: Early alterations in brain DNA methylation at ANK1, BIN1, RHBDF2 and other loci', *Nature Neuroscience*. Nature Publishing Group, 17(9), pp. 1156–1163. doi: 10.1038/nn.3786.

De Jager, P. L., Yang, H. S. and Bennett, D. A. (2018) 'Deconstructing and targeting the genomic architecture of human neurodegeneration', *Nature Neuroscience*. Nature Publishing Group, pp. 1310–1317. doi: 10.1038/s41593-018-0240-z.

Jankovic, J. (2008) 'Parkinson's disease: Clinical features and diagnosis', *Journal of Neurology, Neurosurgery and Psychiatry*. BMJ Publishing Group, pp. 368–376. doi: 10.1136/jnnp.2007.131045.

Joe, E. and Ringman, J. M. (2019) 'Cognitive symptoms of Alzheimer's disease: Clinical management and prevention', *The BMJ*. BMJ Publishing Group. doi: 10.1136/bmj.l6217.

Johnson, C. D. and Davidson, B. L. (2010) 'Huntington's disease: Progress toward effective disease-modifying treatments and a cure', *Human Molecular Genetics*. Oxford Academic, 19(R1), pp. R98–R102. doi: 10.1093/hmg/ddq148.

Jomova, K. *et al.* (2010) 'Metals, oxidative stress and neurodegenerative disorders', *Molecular and Cellular Biochemistry*. Springer, pp. 91–104. doi: 10.1007/s11010-010-0563-x.

Jones, L. L. *et al.* (2000) 'Regulation of the cell adhesion molecule CD44 after nerve transection and direct trauma to the mouse brain', *Journal of Comparative Neurology*. J Comp Neurol, 426(3), pp. 468–492. doi: 10.1002/1096-9861(20001023)426:3<468::AID-CNE9>3.0.CO;2-I.

Joo, Y. *et al.* (2015) 'Involvement of 14-3-3 in tubulin instability and impaired axon development is mediated by Tau', *The FASEB Journal*. FASEB, 29(10), pp. 4133–4144. doi: 10.1096/fj.14-265009.

Kaltenbach, L. S. *et al.* (2007) 'Huntingtin Interacting Proteins Are Genetic Modifiers of Neurodegeneration', *PLoS Genetics*. Edited by H. Orr. Public Library of Science, 3(5), p. e82. doi: 10.1371/journal.pgen.0030082.

Kampinga, Harm H., S. B. (2016) 'Heat shock proteins as potential targets for protective strategies in neurodegeneration', *The Lancet Neurology*. Available at: <https://www.sciencedirect.com/science/article/pii/S1474442216000995> (Accessed: 7 October 2020).

Kang, S. H. *et al.* (2013) 'Degeneration and impaired regeneration of gray matter oligodendrocytes in amyotrophic lateral sclerosis', *Nature Neuroscience*. Nature Publishing Group, 16(5), pp. 571–579. doi: 10.1038/nn.3357.

Kassubek, J. *et al.* (2004) 'Topography of cerebral atrophy in early Huntington's disease: A voxel based morphometric MRI study', *Journal of Neurology, Neurosurgery and Psychiatry*. BMJ Publishing Group Ltd, 75(2), pp. 213–220. doi: 10.1136/jnnp.2002.009019.

Kaur, S. J., McKeown, S. R. and Rashid, S. (2016) 'Mutant SOD1 mediated pathogenesis of Amyotrophic Lateral Sclerosis', *Gene*. Elsevier B.V., pp. 109–118. doi: 10.1016/j.gene.2015.11.049.

Kiernan, M. C. *et al.* (2011) 'Amyotrophic lateral sclerosis', in *The Lancet*. Elsevier, pp. 942–955. doi: 10.1016/S0140-6736(10)61156-7.

Kim, D. *et al.* (2018) 'Proteomic change by Korean Red Ginseng in the substantia nigra of a Parkinson's disease mouse model', *Journal of Ginseng Research*. Elsevier B.V., 42(4), pp. 429–435. doi: 10.1016/j.jgr.2017.04.008.

Kim, S., Seo, J. H. and Suh, Y. H. (2004) ' $\alpha$ -Synuclein, Parkinson's disease, and Alzheimer's disease', in *Parkinsonism and Related Disorders*. Elsevier BV, p. S9. doi:

10.1016/j.parkreldis.2003.11.005.

Kinney, J. W. *et al.* (2018) 'Inflammation as a central mechanism in Alzheimer's disease', *Alzheimer's and Dementia: Translational Research and Clinical Interventions*. Elsevier Inc, pp. 575–590. doi: 10.1016/j.trci.2018.06.014.

Knierim, E. *et al.* (2016) 'Mutations in Subunits of the Activating Signal Cointegrator 1 Complex Are Associated with Prenatal Spinal Muscular Atrophy and Congenital Bone Fractures', *American Journal of Human Genetics*. Cell Press, 98(3), pp. 473–489. doi: 10.1016/j.ajhg.2016.01.006.

Knudsen, K. and Borghammer, P. (2018) 'Imaging the Autonomic Nervous System in Parkinson's Disease', *Current Neurology and Neuroscience Reports*. Current Medicine Group LLC 1, pp. 1–13. doi: 10.1007/s11910-018-0889-4.

Koltai, T. (2014) 'Clusterin: a key player in cancer chemoresistance and its inhibition', *OncoTargets and Therapy*. DOVE Medical Press Ltd., 7, p. 447. doi: 10.2147/OTT.S58622.

Kumar, J., Yu, H. and Sheetz, M. P. (1995) 'Kinetin, an essential anchor for kinesin-driven vesicle motility', *Science*. Oxford Univ. Press, 67(5205), pp. 1834–1837. doi: 10.1126/science.7892610.

Kumar, P. *et al.* (2016) 'Ion Channels in Neurological Disorders', in *Advances in Protein Chemistry and Structural Biology*. Academic Press Inc., pp. 97–136. doi: 10.1016/bs.apcsb.2015.10.006.

Lagier-Tourenne, C., Polymenidou, M. and Cleveland, D. W. (2010) 'TDP-43 and FUS/TLS: Emerging roles in RNA processing and neurodegeneration', *Human Molecular Genetics*. Oxford University Press, 19(R1), p. R46. doi: 10.1093/hmg/ddq137.

Li, M. and Lester, H. A. (2001) 'Ion channel diseases of the central nervous system', *CNS Drug Reviews*. Neva Press Inc., pp. 214–240. doi: 10.1111/j.1527-3458.2001.tb00196.x.

Li, P., Nie, Y. and Yu, J. (2015) 'An Effective Method to Identify Shared Pathways and Common Factors among Neurodegenerative Diseases', *PLOS ONE*. Edited by M. R. Cookson. Public Library of Science, 10(11), p. e0143045. doi: 10.1371/journal.pone.0143045.

Liao, Y. *et al.* (2019) 'WebGestalt 2019: gene set analysis toolkit with revamped UIs and APIs', *Nucleic Acids Research*. Oxford University Press, 47(W1), pp. W199–W205. doi: 10.1093/nar/gkz401.

Liscovitch, N. and French, L. (2014) 'Differential co-expression between  $\alpha$ -synuclein and IFN- $\gamma$  signaling genes across development and in parkinson's disease', *PLoS ONE*. Public Library of Science, 9(12). doi: 10.1371/journal.pone.0115029.

Liu, C. C. *et al.* (2013) 'Apolipoprotein e and Alzheimer disease: Risk, mechanisms and therapy', *Nature Reviews Neurology*. Nat Rev Neurol, pp. 106–118. doi:



10.1038/nrneurol.2012.263.

Lukacs, M. *et al.* (2019) 'Severe biallelic loss-of-function mutations in nicotinamide mononucleotide adenylyltransferase 2 (NMNAT2) in two fetuses with fetal akinesia deformation sequence', *Experimental Neurology*. Academic Press Inc., 320. doi: 10.1016/j.expneurol.2019.112961.

Lunnon, K. *et al.* (2014) 'Methylomic profiling implicates cortical deregulation of ANK1 in Alzheimer's disease', *Nature Neuroscience*. Nature Publishing Group, 17(9), pp. 1164–1170. doi: 10.1038/nn.3782.

M Kanehisa - Novartis Foundation Symposium (2002) 'The KEGG database', *Wiley Online Library*. Available at: <https://onlinelibrary.wiley.com/doi/pdf/10.1002/0470857897#page=99> (Accessed: 14 August 2020).

Ma, B. *et al.* (2016) 'Deletion of the hemopexin or heme oxygenase-2 gene aggravates brain injury following stroma-free hemoglobin-induced intracerebral hemorrhage', *Journal of Neuroinflammation*. BioMed Central Ltd., 13(1), p. 26. doi: 10.1186/s12974-016-0490-1.

Ma, R. hong *et al.* (2017) 'Role of microtubule-associated protein tau phosphorylation in Alzheimer's disease', *Journal of Huazhong University of Science and Technology - Medical Science*. Tongji Medical University, pp. 307–312. doi: 10.1007/s11596-017-1732-x.

Mackay, J., Nassrallah, W. and Raymond, L. (2018) 'Cause or compensation?-Altered neuronal Ca<sup>2+</sup> handling in Huntington's disease', *CNS Neuroscience & Therapeutics*, 24(4), pp. 301–310. doi: 10.1111/cns.2018.24.issue-4.

Mahalingam, S. and Levy, L. M. (2014) 'Genetics of Huntington disease', *American Journal of Neuroradiology*. American Society of Neuroradiology, pp. 1070–1072. doi: 10.3174/ajnr.A3772.

Maheshwari, P. and Eslick, G. D. (2015) 'Bacterial infection and Alzheimer's disease: A meta-analysis', *Journal of Alzheimer's Disease*. IOS Press, 43(3), pp. 957–966. doi: 10.3233/JAD-140621.

Mandelkow, E. M. and Mandelkow, E. (1998) 'Tau in Alzheimer's disease', *Trends in Cell Biology*. Elsevier Ltd, 8(11), pp. 425–427. doi: 10.1016/S0962-8924(98)01368-3.

Manzoni, C., Lewis, P. A. and Ferrari, R. (2020) 'Network Analysis for Complex Neurodegenerative Diseases', *Current Genetic Medicine Reports*. Springer Science and Business Media LLC, 8(1), pp. 17–25. doi: 10.1007/s40142-020-00181-z.

Mao, Q. *et al.* (2020) 'KTN1 Variants Underlying Putamen Gray Matter Volumes and Parkinson's Disease', *Frontiers in Neuroscience*. Frontiers Media S.A., 14, p. 651. doi: 10.3389/fnins.2020.00651.

Martin, D. D. O. *et al.* (2015) 'Autophagy in Huntington disease and huntingtin in

autophagy', *Trends in Neurosciences*. Elsevier Ltd, pp. 26–35. doi: 10.1016/j.tins.2014.09.003.

Matzke, A. *et al.* (2007) 'Haploinsufficiency of c-Met in cd44<sup>-/-</sup> Mice Identifies a Collaboration of CD44 and c-Met In Vivo', *Molecular and Cellular Biology*. American Society for Microbiology, 27(24), pp. 8797–8806. doi: 10.1128/mcb.01355-07.

McKenzie, J. L., Dalchau, R. and Fabre, J. W. (1982) 'Biochemical Characterisation and Localization in Brain of a Human Brain-Leucocyte Membrane Glycoprotein Recognised by a Monoclonal Antibody', *Journal of Neurochemistry*. John Wiley & Sons, Ltd, 39(5), pp. 1461–1466. doi: 10.1111/j.1471-4159.1982.tb12592.x.

McQuade, L. R. *et al.* (2014) 'Proteomics of Huntington's disease-affected human embryonic stem cells reveals an evolving pathology involving mitochondrial dysfunction and metabolic disturbances', *Journal of Proteome Research*. American Chemical Society, 13(12), pp. 5648–5659. doi: 10.1021/pr500649m.

Misawa, H. *et al.* (2012) 'Osteopontin is an alpha motor neuron marker in the mouse spinal cord', *Journal of Neuroscience Research*. John Wiley & Sons, Ltd, 90(4), pp. 732–742. doi: 10.1002/jnr.22813.

Mitchell, J. and Borasio, G. (2007) 'Amyotrophic lateral sclerosis', *Lancet*. Elsevier, pp. 2031–2041. doi: 10.1016/S0140-6736(07)60944-1.

Mousavi, S. V., Agah, E. and Tafakhori, A. (2020) 'The Role of Osteopontin in Amyotrophic Lateral Sclerosis: A Systematic Review', *Archives of Neuroscience*. Kowsar Medical Institute, In Press(In Press). doi: 10.5812/ans.94205.

Myers, R. H. *et al.* (1988) 'Clinical and neuropathologic assessment of severity in huntington's disease', *Neurology*. Wolters Kluwer Health, Inc. on behalf of the American Academy of Neurology, 38(3), pp. 341–347. doi: 10.1212/wnl.38.3.341.

Nguyen, N. D. and Wang, D. (2020) 'Multiview learning for understanding functional multiomics', *PLoS Computational Biology*. Public Library of Science, p. e1007677. doi: 10.1371/journal.pcbi.1007677.

Niu, H. *et al.* (2018) 'Alpha-synuclein overexpression in the olfactory bulb initiates prodromal symptoms and pathology of Parkinson's disease', *Translational Neurodegeneration*. BioMed Central Ltd., 7(1), p. 25. doi: 10.1186/s40035-018-0128-6.

Novellino, F. *et al.* (2020) 'Innate Immunity: A Common Denominator between Neurodegenerative and Neuropsychiatric Diseases', *International Journal of Molecular Sciences*. MDPI AG, 21(3), p. 1115. doi: 10.3390/ijms21031115.

O'Gorman Tuura, R. L., Baumann, C. R. and Baumann-Vogel, H. (2018) 'Beyond Dopamine: GABA, Glutamate, and the Axial Symptoms of Parkinson Disease', *Frontiers in Neurology*. Frontiers Media S.A., 9(SEP), p. 806. doi: 10.3389/fneur.2018.00806.

Ochaba, J. *et al.* (2014) 'Potential function for the Huntingtin protein as a scaffold for selective autophagy', *Proceedings of the National Academy of Sciences of the United States of America*. National Academy of Sciences, 111(47), pp. 16889–16894. doi: 10.1073/pnas.1420103111.

Papanikolaou, G. and Pantopoulos, K. (2005) 'Iron metabolism and toxicity', *Toxicology and Applied Pharmacology*. Toxicol Appl Pharmacol, pp. 199–211. doi: 10.1016/j.taap.2004.06.021.

Papatheodorou, I. *et al.* (2018) 'Expression Atlas: Gene and protein expression across multiple studies and organisms', *Nucleic Acids Research*. Oxford University Press, 46(D1), pp. D246–D251. doi: 10.1093/nar/gkx1158.

Perdigão, C. *et al.* (2020) 'Intracellular Trafficking Mechanisms of Synaptic Dysfunction in Alzheimer's Disease', *Frontiers in Cellular Neuroscience*. Frontiers Media S.A. doi: 10.3389/fncel.2020.00072.

Philips, T. and Rothstein, J. D. (2014) 'Glial cells in amyotrophic lateral sclerosis', *Experimental Neurology*. Academic Press Inc., pp. 111–120. doi: 10.1016/j.expneurol.2014.05.015.

Pomès, P., Louis, H. A. and Beckerle, M. C. (1997) 'CRP1, a LIM domain protein implicated in muscle differentiation, interacts with  $\alpha$ -actinin', *Journal of Cell Biology*. The Rockefeller University Press, 139(1), pp. 157–168. doi: 10.1083/jcb.139.1.157.

Prasad, A. *et al.* (2019) 'Molecular mechanisms of TDP-43 misfolding and pathology in amyotrophic lateral sclerosis', *Frontiers in Molecular Neuroscience*. Frontiers Media S.A., p. 25. doi: 10.3389/fnmol.2019.00025.

Pringsheim, T. *et al.* (2012) 'The incidence and prevalence of Huntington's disease: A systematic review and meta-analysis', *Movement Disorders*. John Wiley & Sons, Ltd, pp. 1083–1091. doi: 10.1002/mds.25075.

Pyatigorskaya, N. *et al.* (2016) 'Medulla oblongata damage and cardiac autonomic dysfunction in Parkinson disease', *Neurology*. Lippincott Williams and Wilkins, 87(24), pp. 2540–2545. doi: 10.1212/WNL.0000000000003426.

Rademakers, R. and Rovelet-Lecrux, A. (2009) 'Recent insights into the molecular genetics of dementia', *Trends in Neurosciences*. NIH Public Access, pp. 451–461. doi: 10.1016/j.tins.2009.05.005.

Ramanan, V. K. *et al.* (2014) 'APOE and BCHE as modulators of cerebral amyloid deposition: A florbetapir PET genome-wide association study', *Molecular Psychiatry*. NIH Public Access, 19(3), pp. 351–357. doi: 10.1038/mp.2013.19.

Ramanan, V. K. and Saykin, A. J. (2013) 'Pathways to neurodegeneration: mechanistic insights from GWAS in Alzheimer's disease, Parkinson's disease, and related disorders',

*Am J Neurodegener Dis*, 2(3), pp. 145–175. Available at: [www.AJND.us](http://www.AJND.us).

Reddy, P. H. and Shirendeb, U. P. (2012) 'Mutant huntingtin, abnormal mitochondrial dynamics, defective axonal transport of mitochondria, and selective synaptic degeneration in Huntington's disease', *Biochimica et Biophysica Acta - Molecular Basis of Disease*. Elsevier, pp. 101–110. doi: 10.1016/j.bbadis.2011.10.016.

Reed, X. *et al.* (2019) 'The role of monogenic genes in idiopathic Parkinson's disease', *Neurobiology of Disease*. Academic Press Inc., pp. 230–239. doi: 10.1016/j.nbd.2018.11.012.

Renton, A. E., Chiò, A. and Traynor, B. J. (2014) 'State of play in amyotrophic lateral sclerosis genetics', *Nature Neuroscience*. Nature Publishing Group, pp. 17–23. doi: 10.1038/nn.3584.

Robberecht, W. and Philips, T. (2013) 'The changing scene of amyotrophic lateral sclerosis', *Nature Reviews Neuroscience*. Nature Publishing Group, pp. 248–264. doi: 10.1038/nrn3430.

Roos, R. A. C. (2010) 'Huntington's disease: A clinical review', *Orphanet Journal of Rare Diseases*. BioMed Central, p. 40. doi: 10.1186/1750-1172-5-40.

Ross, C. A. and Tabrizi, S. J. (2011) 'Huntington's disease: From molecular pathogenesis to clinical treatment', *The Lancet Neurology*. Elsevier, pp. 83–98. doi: 10.1016/S1474-4422(10)70245-3.

Roze, E. *et al.* (2008) 'Mitogen- and stress-activated protein kinase-1 deficiency is involved in expanded-huntingtin-induced transcriptional dysregulation and striatal death', *The FASEB Journal*. Wiley, 22(4), pp. 1083–1093. doi: 10.1096/fj.07-9814.

Rubinsztein, D. C. (2006) 'The roles of intracellular protein-degradation pathways in neurodegeneration', *Nature*. Nature Publishing Group, pp. 780–786. doi: 10.1038/nature05291.

Savage, A. L. *et al.* (2018) 'Retrotransposons in the development and progression of amyotrophic lateral sclerosis', *Journal of Neurology, Neurosurgery and Psychiatry*. BMJ Publishing Group, 90(3), pp. 284–293. doi: 10.1136/jnnp-2018-319210.

Scheltens, P. *et al.* (2016) 'Alzheimer's disease', *The Lancet*. Lancet Publishing Group, pp. 505–517. doi: 10.1016/S0140-6736(15)01124-1.

Serrano-Pozo, A. *et al.* (2011) 'Neuropathological alterations in Alzheimer disease', *Cold Spring Harbor Perspectives in Medicine*. Cold Spring Harb Perspect Med, 1(1). doi: 10.1101/cshperspect.a006189.

Sheng, M., Sabatini, B. L. and Südhof, T. C. (2012) 'Synapses and Alzheimer's disease', *Cold Spring Harbor Perspectives in Biology*. Cold Spring Harbor Laboratory Press, 4(5), p. 10. doi: 10.1101/cshperspect.a005777.

Silva, D. F. *et al.* (2017) 'Mitochondrial Metabolism Power SIRT2-Dependent Deficient Traffic Causing Alzheimer's-Disease Related Pathology', *Molecular Neurobiology*. Humana Press Inc., 54(6), pp. 4021–4040. doi: 10.1007/s12035-016-9951-x.

Skupien, A. *et al.* (2014) 'CD44 regulates dendrite morphogenesis through Src tyrosine kinase-dependent positioning of the Golgi', *Journal of cell science*. J Cell Sci, 127(23), pp. 5038–5051. doi: 10.1242/jcs.154542.

Smith, A. and McCulloh, R. J. (2015) 'Hemopexin and haptoglobin: Allies against heme toxicity from hemoglobin not contenders', *Frontiers in Physiology*. Frontiers Media S.A., p. 187. doi: 10.3389/fphys.2015.00187.

Strang, K. H., Golde, T. E. and Giasson, B. I. (2019) 'MAPT mutations, tauopathy, and mechanisms of neurodegeneration', *Laboratory Investigation*. Nature Publishing Group, pp. 912–928. doi: 10.1038/s41374-019-0197-x.

Tam, O. H. *et al.* (2019) 'Postmortem Cortex Samples Identify Distinct Molecular Subtypes of ALS: Retrotransposon Activation, Oxidative Stress, and Activated Glia', *Cell Reports*. Elsevier B.V., 29(5), pp. 1164–1177.e5. doi: 10.1016/j.celrep.2019.09.066.

Tan, E.-K. *et al.* (2005) 'Functional COMT variant predicts response to high dose pyridoxine in Parkinson's disease', *American Journal of Medical Genetics Part B: Neuropsychiatric Genetics*. John Wiley & Sons, Ltd, 137B(1), pp. 1–4. doi: 10.1002/ajmg.b.30198.

Tang, T. S. *et al.* (2007) 'Dopaminergic signaling and striatal neurodegeneration in Huntington's disease', *Journal of Neuroscience*. Society for Neuroscience, 27(30), pp. 7899–7910. doi: 10.1523/JNEUROSCI.1396-07.2007.

Toyoshima, I. and Sheetz, M. P. (1996) 'Kinectin distribution in chicken nervous system', *Neuroscience Letters*. Elsevier Ireland Ltd, 211(3), pp. 171–174. doi: 10.1016/0304-3940(96)12752-X.

Turner, M. R. *et al.* (2011) 'Concordance between site of onset and limb dominance in amyotrophic lateral sclerosis', *Journal of Neurology, Neurosurgery and Psychiatry*. BMJ Publishing Group Ltd, 82(8), pp. 853–854. doi: 10.1136/jnnp.2010.208413.

Umahara, T. *et al.* (2004) '14-3-3 proteins and zeta isoform containing neurofibrillary tangles in patients with Alzheimer's disease', *Acta Neuropathologica*. Springer, 108(4), pp. 279–286. doi: 10.1007/s00401-004-0885-4.

Vedunova, M. *et al.* (2013) 'Seizure-like activity in hyaluronidase-treated dissociated hippocampal cultures', *Frontiers in Cellular Neuroscience*. Front Cell Neurosci, 7(SEP). doi: 10.3389/fncel.2013.00149.

Walker, F. O. (2007) 'Huntington's disease', *Lancet*. Elsevier, pp. 218–228. doi:



10.1016/S0140-6736(07)60111-1.

Wang, J. Z. and Liu, F. (2008) 'Microtubule-associated protein tau in development, degeneration and protection of neurons', *Progress in Neurobiology*. Pergamon, pp. 148–175. doi: 10.1016/j.pneurobio.2008.03.002.

Wanker, E. E. *et al.* (2019) 'The pathobiology of perturbed mutant huntingtin protein–protein interactions in Huntington's disease', *Journal of Neurochemistry*. Blackwell Publishing Ltd, 151(4), pp. 507–519. doi: 10.1111/jnc.14853.

Wijesekera, L. C. and Leigh, P. N. (2009) 'Amyotrophic lateral sclerosis', *Orphanet Journal of Rare Diseases*. BioMed Central, 4(1), p. 3. doi: 10.1186/1750-1172-4-3.

Wright Willis, A. *et al.* (2010) 'Geographic and ethnic variation in Parkinson disease: A population-based study of us medicare beneficiaries', *Neuroepidemiology*. Neuroepidemiology, 34(3), pp. 143–151. doi: 10.1159/000275491.

Wyss-Coray, T. (2006) 'Inflammation in Alzheimer disease: Driving force, bystander or beneficial response?', *Nature Medicine*. Nature Publishing Group, pp. 1005–1015. doi: 10.1038/nm1484.

Xie, T. *et al.* (2014) 'A genome-wide association study combining pathway analysis for typical sporadic amyotrophic lateral sclerosis in Chinese Han populations', *Neurobiology of Aging*. Elsevier Inc., 35(7), pp. 1778.e9–1778.e23. doi: 10.1016/j.neurobiolaging.2014.01.014.

Yamamoto, T. *et al.* (2017) 'Expression of secreted phosphoprotein 1 (osteopontin) in human sensorimotor cortex and spinal cord: Changes in patients with amyotrophic lateral sclerosis', *Brain Research*. Elsevier B.V., 1655, pp. 168–175. doi: 10.1016/j.brainres.2016.10.030.

Yu, X. *et al.* (2020) 'Protein-protein interaction network with machine learning models and multiomics data reveal potential neurodegenerative disease-related proteins', *Human Molecular Genetics*. Oxford University Press, 29(8), pp. 1378–1387. doi: 10.1093/hmg/ddaa065.

Yurchenco, P. and Patton, B. (2009) 'Developmental and Pathogenic Mechanisms of Basement Membrane Assembly', *Current Pharmaceutical Design*. Bentham Science Publishers Ltd., 15(12), pp. 1277–1294. doi: 10.2174/138161209787846766.

Zhai, R. G. *et al.* (2008) 'NAD synthase NMNAT acts as a chaperone to protect against neurodegeneration', *Nature*. Nature Publishing Group, 452(7189), pp. 887–891. doi: 10.1038/nature06721.

Zhang, X. *et al.* (2010) 'Kinectin-mediated endoplasmic reticulum dynamics supports focal adhesion growth in the cellular lamella', *Journal of Cell Science*. The Company of Biologists Ltd, 123(22), pp. 3901–3912. doi: 10.1242/jcs.069153.

Zhang, X., Wan, J.-Q. and Tong, X.-P. (2018) 'Potassium channel dysfunction in neurons and astrocytes in Huntington's disease', *CNS Neuroscience & Therapeutics*. Blackwell Publishing Ltd, 24(4), pp. 311–318. doi: 10.1111/cns.12804.