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# The application of catalytic processes on the production of algae-based biofuels: a review

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**Abstract:** Over the last decades, microalgal biomass has gained a significant role in the development of different high-end (nutraceuticals, colorants, food supplements, and pharmaceuticals) and low-end products (biodiesel, bioethanol, and biogas) due to rapid growth and high carbon fixing efficiency. Therefore, microalgae are considered a useful and sustainable resource to attain energy security while reducing our current reliance on fossil fuels. From the technologies available for obtaining biofuels using microalgae biomass, thermochemical processes (pyrolysis, HTL, gasification) have proven to be processed with higher viability, because they use all biomass. However, because of the complexity of the biomass (lipids, carbohydrates, and proteins), the obtained biofuels from direct thermochemical conversion have large amounts of heteroatoms (oxygen, nitrogen, and sulfur). As a solution, catalyst-based processes have emerged as a sustainable solution for the increase in biocrude production. This paper's objective is to present a comprehensive review of recent developments on catalyst mediated conversion of algal biomass. Special attention will be given to operating conditions, strains evaluated, and challenges for the optimal yield of algal-based biofuels through pyrolysis and HTL.

**Keywords:** microalgal biomass; Thermochemical conversion; Catalytic upgrading; liquid fuels; Hydrothermal liquefaction; pyrolysis; Gasification.

## 1. Introduction

Fossil fuels have been a critical commodity for the economic and social development of the modern world. However, their consumption has inevitably increased the levels of anthropogenic carbon dioxide (CO<sub>2</sub>) emissions to concentrations that exceed the earth's absorption capacity through the natural carbon cycle [1]. The production of biomass-based fuels (or biofuels) are considered as one of the prospective replacements to the conventional fossil fuels [2] for both developed and non-developed countries due to its abundance and distribution [3].

Over the last years, several biomass resources such as grass, wood, crops and residues, animal waste, municipal solid waste, and even aquatic plants have been studied to produce biofuels [4]. However, up to date, microalgae is considered one of the most attractive sources of renewable energy and raw materials. It diversifies the scope of different industries in the elaboration of food and feed, pharmaceuticals, pigments, and colorants, bioplastics, and protein hydrolysates [5].

Microalgae and cyanobacteria are a diverse group of photosynthetic microorganisms that naturally grow in lakes, rivers, and oceans. Microalgae offer several advantages over plant-based biofuels such as (i), high growth rate, (ii) use of non-arable lands, (iii) can be grown in wastewater, (iv) high consumption of CO<sub>2</sub>, and (v) their production can be directed toward the synthesis of several compounds of commercial interest [6].

To obtain biomass with a high concentration of specific metabolites is one the cornerstones of microalgae biotechnology. Several authors have proved that specific culturing conditions such as nutrient concentration [7], photobioreactor configuration [8], environmental conditions (temperature and illuminance), agitation and pH [9] directly influence the cellular composition, resulting in the final concentration and productivity of the strain, as well as the variation in the content of specific metabolites (lipids, carbohydrates, proteins and of other components) [10].

The transformation of algal biomass into biofuels is not new. Several studies have covered different areas on the strain selection, culture method, and transformation into biofuel, which is the critical link in the production chain towards obtaining sustainable biofuels from microalgae.

The algal biomass produced under specific conditions can be transformed into energy by applying thermochemical and biochemical methods. Biofuel such as Bio-oil, biochar, synthesis gas (syngas), and heat are obtained through thermochemical conversion. On the other side, biodiesel, biohydrogen, biomethane (or biogas), and bioethanol can be produced via the biochemical conversion of algal biomass [1]. Although different forms of cultivation and production have been developed in recent years, it is still necessary to find an effective and sustainable production mechanism to reach the full potential of microalgae-based biofuels, especially in large scale industrial applications.

One possible solution to achieve the potential of algae as a feedstock for biofuels is the use of reactions that employ whole biomass such as Anaerobic digestion (AD) and thermochemical conversion. Biogas is the main product of AD and is considered one of the most promising biofuels that can address rising concerns about fossil fuels [11]. Another alternative is the application of catalytic-based processes such as Hydrothermal Liquefaction (HTL) and pyrolysis. Through thermochemical conversion, the biomass is decomposed under oxygen/air or steam under deficient conditions to produce synthetic gas or syngas which primarily consist of hydrogen ( $H_2$ ), carbon monoxide (CO), and carbon dioxide ( $CO_2$ ) [12], the quantity and quality of the final product depends upon the process, reaction temperature, heating rate, and oxygen supply [13]. In comparison to the biochemical conversion of algal biomass, the thermochemical approach is a more straightforward route to produce biofuels due to several factors: (i) employ the entire biomass as feedstock, (ii) the process times is shorter, and (iii) their final yield can be improved by the addition of chemical catalyst [14]. The present study is intended to give a comprehensive overview of the state-of-the-art usage of catalysts on the thermochemical conversion of algal biomass into solids, liquids, and gas biofuels. Special attention will be given to operating conditions, strains evaluated, and challenges for the optimal yield of algal-based biofuels through pyrolysis and HTL.

## 2. Algae-based biofuels

Biofuels are broadly classified by generations. First-generation (1st gen) is produced from food feedstock (corn, sugarcane, soybean, potato, beet, soybeans, coconut, sunflower, rapeseed, palm oil, switchgrass, Jatropha, Camelina, Cassava). Although 1st gen is considered a sustainable source of energy due to the reduction on greenhouse gas (GHG) emissions, specific details such as their competition with food supply, high requirement of government subsidies, large amounts of non-sustainable fertilizers, and environmental concerns due to the loss biodiversity linked to the promotion of deforestation for large monoculture areas [15], hinder their true impact as a cleaner and more sustainable option over fossil fuels.

Second-generation (2nd gen) was conceived as a partial solution of several drawbacks of 1st gen biofuels. This generation relies on nonfood items such as cellulosic biomass, straw, manure, used cooking oil and other non-conventional sources, which usually finish in landfills once their useful portion has been removed [12]. However, 2nd gen is still not industrially profitable due to biomass complexity and problems associated with its production, storage, and transportation [2].

Third-generation (3rd gen) focuses on the upgrade of aquatic feedstock, such as microalgal and cyanobacterial biomass, into different fuels. Microalgae have been praised as a better solution for the energy problem due to specific qualities of algal production: (i) do not compete with human and animal food stock, (ii) its harvesting can be done through the year, (iii) can employ saline and

wastewater, (iv) have better growth rate than higher plants, (v) can convert 183 G tons of CO<sub>2</sub> to produce 100 G tons of biomass [16], and (vi) the concentration of transformable metabolites (lipids and carbohydrates) is stable on the biomass. First, the selected strain had to be cultured until it reaches the largest possible biomass concentration in the photobioreactor; once reached, the biomass is removed from the culture media (centrifugation, flocculation, filtration, and other techniques) and dried. The dried biomass is then ready to be used as feedstock for several biofuels (biodiesel, bioethanol, biogas, and so on). These different sections have been the main topic of research over the last 20 years, attracting the attention of different universities, research centers, and energy companies worldwide like Ecopetrol (Colombia), Exxon Mobile, Shell (US), Petrobras (Brazil), Total (France).

2.1. How algae-based biofuels changed over time

Companies like Solix biofuels, Corbion (previously known as Terravia or Solazyme), Cellana, Sapphire Energy, Seambiotic, Oil Fox, Synthetic genomics, Euglena, and others attracted over 200 million dollars from private and public sectors. However, after years of research, none of the companies reach the economic balance for algal-based biofuels. The latter can be due to several problems identified through the last decade. First, the microalgal biodiversity is so vast that after ten years of research, we are still far from identifying the total diversity of algae and cyanobacteria (Table 1). Another problem related to the strains is the stability of their growth on industrial photobioreactors and the synthesis of the target metabolites.

Table 1. Different strains studied for biodiesel production.

Strain	Lipids	Carbohydrates	Proteins	Reference
<i>Arthrospira platensis</i>	30.23 wt%	31.89 wt%	16.81 wt%	[17]
<i>Auxenochlorella protothecoides</i>	20.58 wt%	--	--	[18]
<i>Botryococcus braunii</i>	17.85 wt%	--	12.54 wt%	[19]
	60 wt%	20wt%	--	[20]
<i>Chlamydomonas reinhardtii</i>	25.25 wt%	--	--	[21]
<i>Ch. reinhardtii</i> CC-400	28.5 wt%	--	--	[22]
<i>Ch. Reinhardtii</i> CC-4349	64.25 wt%	--	--	[23]
<i>Chlorella</i> sp G-9	32.6 – 34.2 (mg/L/day)	--	--	[24]
<i>C. kessleri</i>	14.42 – 24.19% TVSS	--	--	[25]
<i>C. protothecoides</i>	12.94 – 19.48% TVSS	--	--	
<i>C. pyrenoidosa</i>	20.9 – 25.48 wt%	--	--	[26]
<i>C. vulgaris</i>	18.75 wt%	--	--	[27]
	32 – 38 wt%	--	--	[28]
<i>C. vulgaris</i> LBL3-M	10 wt%			[29]
<i>C. vulgaris</i> UTEX 1803	9 wt%	--	--	[30]
<i>C. vulgaris</i> Mutant (UV715)	41 wt%	--	--	[31]
<i>Chlorococcum oleofaciens</i>	34 wt%	--	--	[32]
<i>Coccomyxa</i> sp strain Obi (AG125)	43 wt%	1 wt%	--	[33]
<i>Dunaliella tertiolecta</i>	15 wt%	--	--	[34]
	23.4 wt%	--	--	[35]
<i>Nannochloropsis gaditana</i>	17.6 wt%	--	24.1 wt%	[36]
<i>Phaeodactylum tricornutum</i>	55.7 wt%	9 wt%	22 wt%	[37]
<i>Scenedesmus almeriensis</i>	24.6 wt%	--	--	[38]
<i>S. dimorphus</i>	15.15 – 24.4 wt%	--	--	[26]
<i>S. obliquus</i>	32.5 wt%	--	--	[39]
	24.9 wt%	--	--	[40]

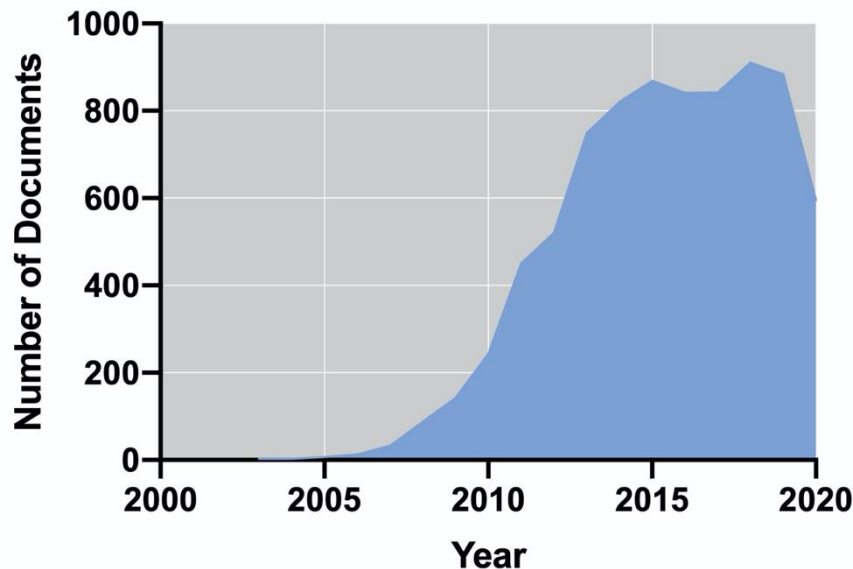
	25.2 wt%	--	--	[41]
	60 wt%	--	--	[42]
<i>S. quadricauda</i>	22.5 wt%	--	--	[43]
	26 wt%	18 wt%	15 wt%	[44]
<i>Tetraselmis suecica</i>	53.8 wt%	--	--	[45]

Microalgae can be produced under autotrophic, mixotrophic, or heterotrophic conditions. Different systems for the production of algae are available for their culture under the three conditions, as mentioned earlier [46]. Autotrophic systems are the most common since the algae only require light as an energy source and dissolved CO<sub>2</sub> as a source of carbon. Usually, algae growth under autotrophic systems can be produced in open or closed photobioreactors. Open ponds are the simplest of all systems for algal production, and it requires low energy inputs. It has easy maintenance; however, it is severely affected by seasonal variations and is prone to contamination by other microbes [47]. Mixotrophic and heterotrophic production of algae requires the addition of organic carbon sources (glucose, acetate, and others), which can lead to contamination by the presence of bacteria and fungi; therefore, these systems require closed photobioreactors (PBR). Closed PBR offers several advantages over open systems: (i) aseptic growth conditions, (ii) increased cell concentration due to better light distribution, (iii) improved pH control, and (iv) reduced water loss due to evaporation. However, their operation cost, maintenance, and energy inputs are considerably higher than in open ponds [47].

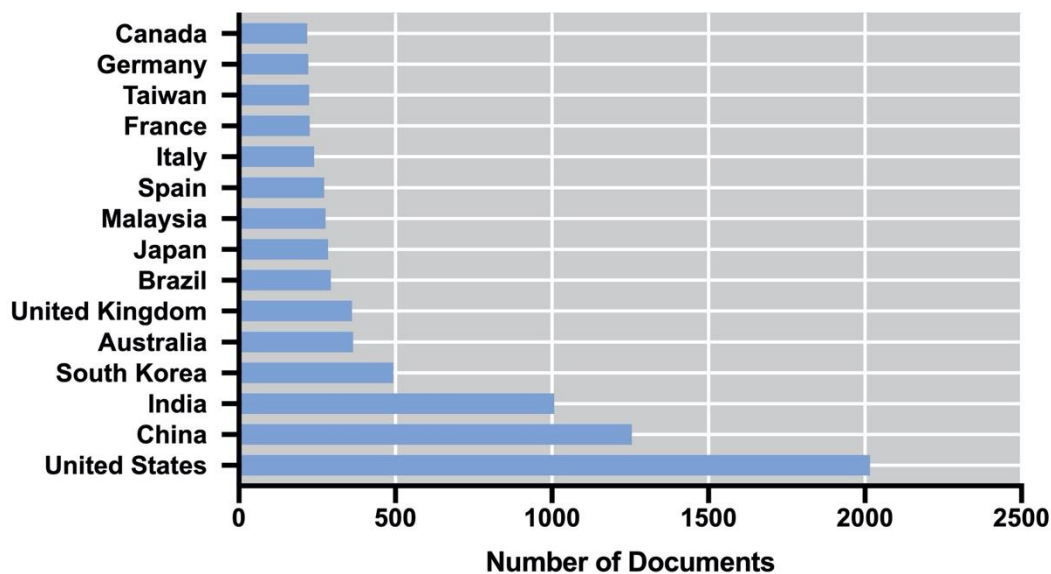
Once the biomass within the reactor reached the expected concentration comes the harvesting; due to their nature, microalgal cells have a small size, low specific gravity, and are highly diluted on the culture media; therefore, their concentration is labor, energy, and time-intensive step [48]. Several techniques are available at industrial scale such as centrifugation, filtration, flocculation, flotation, electroflotation, and so on [10]. However, the method's selection and application lie on the technical and economic analysis since some of them can be extremely expensive and energy-intensive for the production of algal-based biofuels [49]. Once the biomass is removed from the media, most of cell water content must be removed via spray drying, drum drying, freeze-drying, or solar drying to avoid any interference with the extraction [46]. Following drying comes the extraction of lipids and carbohydrates, which is considered as the crucial step that inhibits the industrial-scale production of algae-based biofuels [49]. The Microalgal cell wall is made of polysaccharides and cellulose synthesized from silicic acid [50], and must be broken in order to release, both lipids and carbohydrates; in consequence, only a fraction of the biomass is used in biofuel process production. Therefore, biodiesel and bioethanol production are still not economically feasible due to the high cost and energy inputs in almost all stages [51]. Other biofuels such as biogas and biohydrogen have gained attention as sustainable alternatives for energy production using microalgal biomass.

Biogas is produced via a sequence of biochemical processes converting the organic material: hydrolysis, fermentation, acetogenesis, and methanogenesis, also known as Anaerobic Digestion (AD) [52]. In this process, the whole biomass is used for the production of methane (55–75%) and carbon dioxide (25–45%) [53]; therefore, the energy performance is higher in comparison to biodiesel and bioethanol [54]. Additionally, nutrients like organic nitrogen or phosphorus may be mineralized and subsequently recycled for algae cultivation [55]. Unlike biogas, biohydrogen is produced via their metabolic pathways along with the cell growth, therefore it does not require further processing of the biomass (i.e harvesting, dewatering, drying, and extraction), and is considered clean and renewable, with higher energy production (142 MJ/Kg<sup>-1</sup>) [56]. Biohydrogen can be obtained by photofermentation, dark fermentation, direct and indirect biophotolysis [57], however Hydrogen production cannot be achieved amidst effective photosynthesis as oxygen inactivates hydrogenase [58]. The Research and Development on algal-based biofuels is a field that, in recent years, has been maintained with a considerable number of publications. Figure 1 Shows the evolution of the number of publications per year along the last 18 years period. According to the data obtained from the Scopus database (Elsevier). It is possible to observe an exponential increase in the number of

publications between 2006 to 2015. Since 2016, the number of documents has remained almost constant up to a final number of 8022 (including accepted documents for 2021), where United States, China, India, South Korea, and United Kingdom dominate the scientific publication on algal-based biofuels.



(a)



(b)

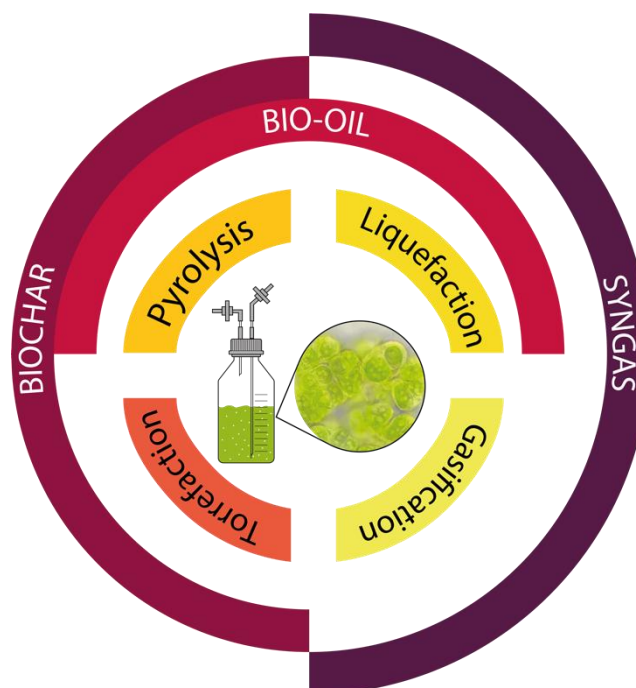
**Figure 1.** Evolution of the number of publications from 2003 to 2020 on algal biofuels (a) and their country of origin (b).

### 3. Thermochemical conversion of algal biomass

Thermochemical methods (figure 2) can be grouped into four classes, hydrothermal liquefaction, pyrolysis, gasification, and torrefaction [59]. In thermochemical process, the algal biomass is thermally decomposed into usable biofuels such as syngas, bio-oil and biochar (figure 2). Unlike biochemical production of biofuels, thermochemical processes does not require the extraction of lipids nor carbohydrates; therefore, the entire biomass can be used. Finally, the reaction time is short; providing a simpler route for the biofuel production [10].



180



181 **Figure 2.** Different thermochemical conversion methods of microalgal biomass and their main  
 182 products.

### 183 3.1. Pyrolysis of microalgae

184 Pyrolysis is the thermal decomposition of biomass at high temperature (400-600°C), in an  
 185 atmospheric-pressure inert environment. Compared to other conversion technologies, pyrolysis of  
 186 algal biomass has achieved reliable and promising outcomes that could lead to commercial  
 187 exploitation [60]. The bio-oils obtained from the pyrolytic reaction of algal biomass, due to the lipid  
 188 and protein content, had a higher heating value, higher aromatics, and lower acidity compared to  
 189 lignocellulosic biomass. [61]; while bio-oils derived from wood holds solids with chemically  
 190 dissolved oxygen concentrations [8]; implying that better quality algae-based bio-oil could be  
 191 produced from catalytic pyrolysis.

192 Pyrolysis can be categorized in five modes (i) slow, (ii) intermediate (iii) fast, (iv) flash, and (v)  
 193 microwave pyrolysis; each one possesses a differential heating rate, the presence and/or heating route  
 194 [9]. Slow pyrolysis is characterized by the heating of biomass under a “slow” heating rate (0.1-  
 195 0.8°C/s), with a moderate temperature (550–900°C) and long retention times (> 5 min) [62]. Their main  
 196 product is biochar with by-products such as bio-oil and syngas [63]. Under slow pyrolysis different  
 197 particle sizes can be processed, therefore both macro and microalgae can be used without mechanical  
 198 pre-treatment. Intermediate pyrolysis is carried out using the intermediate conditions between slow  
 199 and fast pyrolysis [64]. Normally Intermediate pyrolysis occurs at moderate temperatures of reaction  
 200 (up to 500°C), 0.5-25 min residence times for feedstocks, and 2-4 s moderate residence times for  
 201 vapour. [65]. The main product from intermediate pyrolysis is bio-oil (40– 60%) followed by non-  
 202 condensable syngas (20-30%) and biochar (15-25%) [66]. The bio-oil obtained is characterized by  
 203 reduced viscosity and low tar content with small concentration of tar [67]. The preferred method for  
 204 optimizing bio-oil production is fast pyrolysis; this method is carried out at elevated temperatures  
 205 (850-1100°C), fast heating rate (> 1°C/s) and short pyrolysis time (0.5-10s) [68], this conditions reduce  
 206 secondary reactions (secondary cracking, condensation, and polymerization of intermediates) which  
 207 contribute to the production of a bio-oil with enhanced qualities.

208 Flash pyrolysis use high temperatures (950–1250 °C), high heating rates (1.000-10.000 °C/s) and  
 209 short pyrolysis time (0.5–10s). due to the fast reaction, bio-oil is the main product of the reaction (90  
 210 wt%) [69]. Finally, Microwave-Assisted Pyrolysis (MAP) is a gentle and medium speed process, with  
 211 a heating rate between conventional pyrolysis and fast pyrolysis [70]. This process has gained large

attention in the recent years because is considered as a more energy-efficient method in comparison with other pyrolysis-related systems [71] and there is no requirement of mechanical pretreatment of biomass, resulting in substantial energy savings.

From pyrolysis it can be obtained bio-oils, chars, and non-condensable gases; however, the final content and amounts will depend directly from the operation conditions and microalgae properties and reaction type [70]. Under lower temperatures chars are the major product; at moderate temperatures (400 – 550°C) with short residence times (2–3s) liquid production is favored. Finally, the gas product increases when the temperature is increased [9]. Over the last years, several studies have been conducted to increase the efficiency of pyrolysis process using microalgal genera such as *Arthrospira* sp [72], *Chaetoceros* sp [67], *Chlamydomonas* sp [4,73], *Chlorella* sp [74-79], *Desmodesmus* sp [80], *Dunaliella* sp [67], *Haematococcus* sp [67,81], *Isochrysis* sp [82,84], *Microcystis* sp [69], *Nannochloropsis* sp [85-88], *Oscillatoria* sp [89], *Pavlova* sp [90,91], *Schizochytrium* sp [92], *Tetraselmis* sp [67, 82], *Spirulina* sp [76,93,94], and *Synechococcus* [67]. A detailed list of species studied can be found on table 2.

In order to exploit the potential of pyrolysis in microalgal conversion, the process has to be improved towards a higher bio-oil yield [63] with less oxygenic compounds to prevent polymerization and condensation. Suitable catalyst could lead to in situ upgrading of generated bio-oil [74,95]. Another advantage of catalytic pyrolysis is that catalysts used for pyrolysis can be recycled to the reactor [74]. The most common catalysts used for microalgae pyrolysis include Na<sub>2</sub>CO<sub>3</sub>, metallic based catalyst such as Ni, Mo and ceria-based catalysts (NiCe/Al<sub>2</sub>O<sub>3</sub> and NiCe/ZrO<sub>2</sub>) [90] have shown great catalytic efficiency. On the other hand, other metal catalysts including Ce, Ti, Co, Mg, and Al did not show obvious catalytic effect [70]. ZSM-5-based zeolites such as H-ZSM-5, Fe-ZSM-5 Cu-ZSM-5 and Ni-ZSM-5 are considered as the most effective catalyst for the pyrolysis of algal biomass. Other zeolites such as ITQ-2 and MCM-22 had a similar but less effective function [96].

In a study on the catalytic pyrolysis of *Nannochloropsis* sp, [85] were able to significantly reduce the oxygen content (from 30 to 19wt%) and a higher calorific value (from 24.6 to 32.5MJ/kg). Other studies such as [97-99] proved the ability of catalytic-mediated pyrolysis to increase the yield of bio-oil. In another study, [99] used HZSM-5 and found that an increase in catalyst-to-biomass ratio from 1:1 to 5:1 significantly improved the aromatic yields. On the other hand, [100] studied pyrolysis of cyanobacteria over Mg-Al layered double oxide/ZSM-5 composites, and the pyrolytic bio-oil contained less nitrogenated compounds. On another study, [82] improved the yield and quality of bio-oil from *Tetraselmis* sp. and *Isochrysis* sp in a fixed bed reactor with the addition of NiCe/Al<sub>2</sub>O<sub>3</sub> and NiCe/ZrO<sub>2</sub>. In another study, [75] investigated the efficiency of five different zeolite-based catalyst (H-, Fe-, Cu-, and Ni-ZSM-5) in the bio-oil production from *Chlorella* biomass, they found that HZSM-5 increased the yield of the hydrocarbon fraction in the organic phase from 21 to 43 wt%. Finally, [93] evaluated the efficiency of MgO and ZSM-5 under environment enriched with N<sub>2</sub> and CO<sub>2</sub>, where maximum bio-oil (46.2 wt%) was obtained with basic metal MgO.

Table 2. Strains studied on catalytic pyrolysis and their catalyst.

Strain	Catalyst	Bio-oil wt%	Bio-char wt%	Syngas wt%	Reference
<i>Arthrospira plantensis</i>	Ni/HMS-ZSM5	33.44	32.52	27.67	[72]
	Fe/HMS-ZSM5	38.15	30.01	58.94	
	Ce/HMS-ZSM5	36.41	31.80	28.58	
<i>Chlamydomonas reinhardtii</i>	hydrotalcite	54.84	37.59	7.57	[73]
<i>Ch. debaryana</i>	β-zeolite	23.5	--	--	[4]
	Activated charcoal	43.8	--	--	
<i>Chlorella</i> sp	Na <sub>2</sub> CO <sub>3</sub>	41.0	54.4	34.1	[74]
	Fe-ZSM-5	43.1	29.7	27.1	
	Cu-ZSM-5	46.9	27.9	24.6	[75]
	Ni-ZSM-5	45.1	30.1	25.4	

	Magnetite	53.8	27.4	22.8	
	Activated carbon	49.4	37.3	13.3	[76]
<i>Chlorella vulgaris</i>	H <sup>+</sup> ZSM-5	25	24	--	[77]
	Ni-ZSM-5	18.97	--	--	[78]
	H <sup>+</sup> ZSM-5	52.7	25.7	21.6	[79]
<i>Desmodesmus communis</i>	HZSM-5	8	42	--	[80]
<i>Haematococcus pluvialis</i>	KCl	12	60	28	
	KOH	11	65	76	
	K <sub>2</sub> CO <sub>3</sub>	13	64.8	22.2	
	MgO	12.5	62	25.5	[81]
	Al <sub>2</sub> O <sub>3</sub>	15	61	24	
	CaO	13	63	24	
	Microalgae Residue	15	60	25	
	CeO <sub>3</sub>	23	30	47	
	Ce/Al <sub>2</sub> O <sub>3</sub>	25	32	42	
	NiCe/Al <sub>2</sub> O <sub>3</sub>	24	32	43	
<i>Isochrysis</i> sp	MgCe/Al <sub>2</sub> O <sub>3</sub>	23	31	46	[82]
	Ce/ZnO <sub>2</sub>	25	29	54	
	NiCe/ZnO <sub>2</sub>	23	27	50	
	MgCe/ZnO <sub>2</sub>	23	28	49	
	Li-LSX-zeolite	29	35	36	[83]
		42.5	33	24.5	[84]
<i>Nannochloropsis</i> sp	HZSM-5	25	38	--	[85]
	Ni-Ce/Al <sub>2</sub> O <sub>3</sub>	23.3	30.9	--	[86]
	HZSM-5	49	40	10	[87]
<i>N. oculata</i>	Co-Mo/γ-Al <sub>2</sub> O <sub>3</sub>	26	42	--	[88]
<i>Oscillatoria</i> sp	TiO <sub>2</sub> , ZnO	33.33	43.05	26.25	[89]
	CeO <sub>3</sub>	21.07	47.96	45.92	
	TiO <sub>3</sub>	20.04	48.18	45.10	
	Ce/TiO <sub>3</sub>	21.67	47.44	46.26	[90]
	Ni/TiO <sub>3</sub>	22.55	47.66	45.39	
<i>Pavlova</i> sp	Co/TiO <sub>3</sub>	20.4	48.28	44.61	
	CeO <sub>2</sub>	21.07	37.86	41.07	
	TiO <sub>2</sub>	20.04	39.49	40.47	
	Ce/TiO <sub>2</sub>	21.67	37.46	40.87	[91]
	Ni/TiO <sub>2</sub>	22.55	37.16	40.29	
	Co/TiO <sub>2</sub>	20.41	38.85	40.74	
<i>Schizochytrium limacinum</i>	ZYNa	26	9	--	[92]
	CeO <sub>3</sub>	23	19	58	
	Ce/Al <sub>2</sub> O <sub>3</sub>	25	17	58	
	NiCe/Al <sub>2</sub> O <sub>3</sub>	25	17	58	
<i>Tetraselmis</i> sp	MgCe/Al <sub>2</sub> O <sub>3</sub>	23	16	51	
	Mg/ZnO <sub>2</sub>	23	18	59	[82]
	Ce/ZnO <sub>2</sub>	23	17	58	
	NiCe/ZnO <sub>2</sub>	23	16	51	
	MgCe/ZnO <sub>2</sub>	23	17	58	
<i>Spirulina</i> sp	Magnetite	49.4	25.4	25.2	
	Activated carbon	46.4	33.2	20.4	[76]
	ZSM-5	44.8	21.1	34.1	[93]
<i>Spirulina platensis</i>	MgO	46.2	29.5	24.3	
	Ce(II)/HZSM-5	49.7	20	30.3	[94]

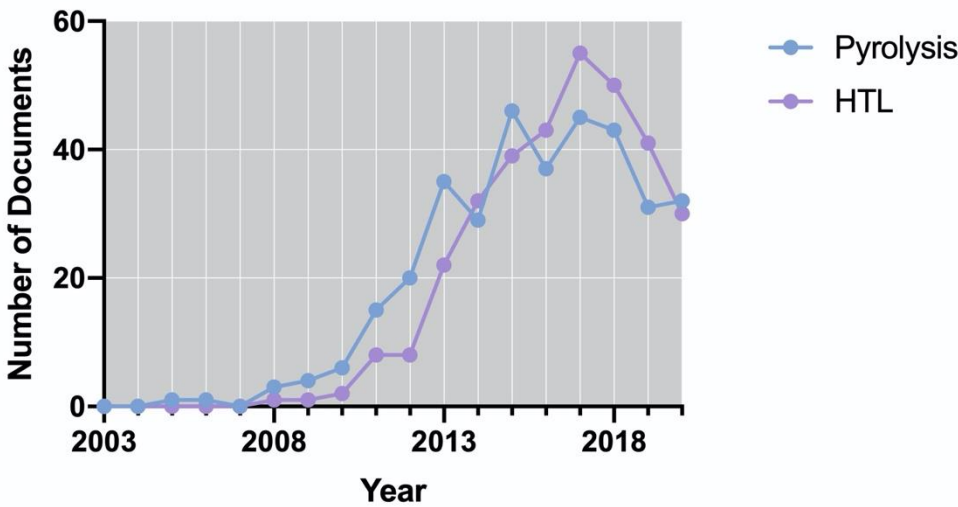


3.2. Hydrothermal liquefaction of algal biomass

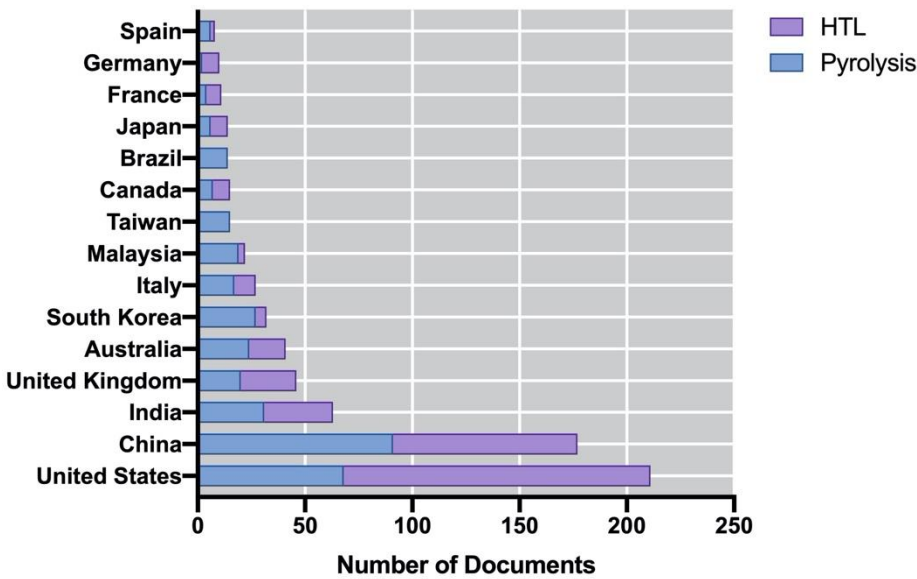
One of the problems with algal biomass is the necessity to remove the high-water content prior to the production of biofuels. In this case Hydrothermal liquefaction (HTL) stands out as a promising technology for the thermochemical conversion of biomass into more useful liquid fuels [101]. Unlike pyrolysis, HTL can convert high-moisture biomass to biocrude in water medium and thus does not require preliminary drying processes [102].

Hydrothermal liquefaction is performed in the presence of water under high pressure (5-25 MPa), subcritical water temperature (280-370°C). Under this conditions, all the macromolecules found within algal biomass (including lipid, protein, and carbohydrate) undergoes depolymerization reactions (fragmentation, hydrolysis, dehydration, deoxygenation, aromatization and repolymerization) [103] for the production of several products such as bio-oil, gas, solid residue and aqueous phase-by-products [104]. HTL is considered a more robust thermochemical technology, not only for the usage of wet biomass, but also due to their high biocrude yield (24 -64 wt%) [105], some essential nutrients (N, P, Mg, and K) can be recycled for microalgal culture [106]. Additionally, up to 50% of oxygen can be removed, resulting in a biocrude with a Higher Heating Value (HHV) ranging from 30 to 40 MJ/kg [107,108]. However, the algae-derived biocrude possess some disadvantages such as a high-water content, high viscosity, and high heteroatom content, which impede its upgrade into usable fuels [105]. Several studies underline that biomass load/ratio, reaction temperature, residence time, pressure, catalyst (including homogenous and heterogenous catalyst), reaction medium, influence the yield, composition and physico-chemical properties of biocrude obtained under HTL [109].

The application of catalysts on HTL reaction is an interesting opportunity to improve the process in several aspects such as the yield and quality of biocrude [110,111], inhibition of side reactions, decrease of reaction temperature, pressure, reduce its viscosity, and reduction in the processing time [112]. The catalysts employed can be separated into homogeneous (water soluble) and heterogeneous (non-water soluble) [101], Table 3 presents a list of homogeneous and heterogeneous catalyst employed on the conversion of algal biomass into biofuels. Figure 3 Shows the evolution of the number of publications per year along the last 16 years period. According to the data obtained from the Scopus database (Elsevier). It is possible to observe an exponential increase in the number of publications between 2008 to 2017. Finally, where United States, China, India, South Korea, and United Kingdom dominate the scientific publication on the application of the pyrolysis and HTL.



(a)



(b)

**Figure 3.** Evolution of the number of publications from 2003 to 2020 on pyrolysis and HTL using algal biomass (a) and their country of origin (b).

3.2.1. Homogeneous catalysis.

Homogeneous catalysts are water-soluble at room temperature. During its reaction the formation of char/tar is inhibited while enhancing product yield by expediting water-gas shift reaction [112].

The most common forms include alkali salts ( $\text{Na}_2\text{CO}_3$  and  $\text{KOH}$ ), mineral and organic acids ( $\text{CH}_3\text{COOH}$  and  $\text{HCOOH}$ ), metallic cations ( $\text{Zn}^{2+}$  and  $\text{Co}^{3+}$ ) [111,112].  $\text{Na}_2\text{CO}_3$  is the most common catalyst employed, and can enhance the production of BTEX (benzene, toluene, ethylbenzenes and xylenes) and C5 to C18 aliphatic hydrocarbons, which are critical elements of gasoline and diesel fuels [101]. In their work, [113] observed that  $\text{Na}_2\text{CO}_3$  enhanced the yield of bio-crude from *Nannochloropsis* sp at 250 °C. However, at higher temperatures (300–350 °C) other species studied such as *Pavlova* and *Isochrysis* sp, have higher bio-oil yields (50–60%). The difference between results can be explained by the difference on biomass composition, since *Pavlova* and *Isochrysis* sp has high lipid and carbohydrate contents. These results are consistent with those reported by [114], who observed that algae with high carbohydrate content was efficiently liquefied. In other study, [115] found that  $\text{Na}_2\text{CO}_3$  increased the bio-oil yield up to 52% (29% higher than for the uncatalysed process) on *Spirulina platensis*, and  $\text{Ca}_3(\text{PO}_4)_2$  and  $\text{NiO}$  produced a negative effect on bio-oil yield. On the other hand, [116] found that  $\text{Na}_2\text{CO}_3$  does not improved the formation of bio-oil on a strain of *C. vulgaris*.  $\text{KOH}$  has been reported as an interesting catalyst; according to [117], in the catalytic HTL of *Cyanidioschyzon merolae*,  $\text{KOH}$  can increase the bio-oil yield in range of 5–10% of bio-oil (from 16.9 to 22.7%) than for the non-catalytic process under similar reaction conditions. The performance of alkali catalyst is significantly affected by the temperature of the process, irrespective of the species evaluated [116,118]. For example, the formation of aliphatics and cyclics are directly affected with an increment of temperature (300 °C); however, at higher temperatures their concentrations declined due to subsequent cracking [101].

Apart from alkaline catalysts, both organic ( $\text{HCOOH}$  and  $\text{CH}_3\text{COOH}$ ) and inorganic acid ( $\text{H}_2\text{SO}_4$ ) catalysts have been used [116,119]. According to [120], a concentration of 6% of  $\text{H}_2\text{SO}_4$  increased up to 70% the bio-oil production from macroalga *Ulva prolifera* sp, with oxygenates as the dominant products in the bio-oil. In another research, [119] found that 2.4%  $\text{H}_2\text{SO}_4$  had a positive effect on the bio-crude oil production from *Dunaliella tertiolecta*; it can be highlighted that the bio-oil obtained is composed mainly of esters, carboxylic acids and ketones. In the application of  $\text{HCOOH}$

and CH<sub>3</sub>COOH in a reaction with *C. vulgaris* (300-350 °C for 1 h) [120] demonstrate that acid catalyst produced a higher bio-crude oil yield with a better flowability of oil product. [121] obtained a maximum yield of 28% of bio-oil using H<sub>2</sub>SO<sub>4</sub> and CH<sub>3</sub>COOH in the catalytic HTL of *Enteromorpha* sp.

There are certain challenges that hinder the prospect of industrial application of homogeneous catalyst on HTL. Alkali and acidic catalyst negative affect the pH of the reaction, leading to corrosion on the reaction equipment [122]. Catalysts based on carbonates (hydroxides or simple carboxylic acids) have a low efficiency on the decarboxylation, isomeration and aromatization of fatty acids [102]. Formic acid and acetic acid can induce the formation of gas fractions (30wt% and 16–22wt%, respectively) [118] and are consumed through the reaction stage; therefore, this type of catalyst must be removed and disposed [112].

Table 3. Strains evaluated and their catalyst.

Strain	Catalyst	Catalyst type	Bio-oil Yield (wt%)	Reference
<i>Chlorella</i> sp	Na <sub>2</sub> CO <sub>3</sub>	homogeneous catalysts	28	[114]
	Formic acid		28	
	CH <sub>3</sub> COOH		15.7	
<i>C. pyrenoidosa</i>	KOH		13.6	[119]
	Na <sub>2</sub> CO <sub>3</sub>		41.78	
	CH <sub>3</sub> COOH		21.23	
<i>Cyanidioschyzon merolae</i>	NaOH		21.78	[117]
	KOH		22.67	
	Na <sub>2</sub> CO <sub>3</sub>		42.0	
<i>Dunaliella tertiolecta</i>	KOH		49.09	[125]
	Na <sub>2</sub> CO <sub>3</sub>		23.0	
<i>Enteromorpha prolifera</i>	Na <sub>2</sub> CO <sub>3</sub>	homogeneous catalysts	23.0	[126]
<i>Isochrysis</i> sp	Na <sub>2</sub> CO <sub>3</sub>		50	[113]
<i>Laminaria saccharina</i>	KOH		67.0	
<i>Microcystis viridis</i>	Na <sub>2</sub> CO <sub>3</sub>		33.0	[127]
	Na <sub>2</sub> CO <sub>3</sub>		40	
	Formic acid		28	
<i>Nannochloropsis</i> sp	Na <sub>2</sub> CO <sub>3</sub>		28	[114]
	Na <sub>2</sub> CO <sub>3</sub>		24.2	
	Na <sub>2</sub> CO <sub>3</sub>		50	
<i>Pavlova</i> sp	Na <sub>2</sub> CO <sub>3</sub>		27.1	[113]
<i>Porphyridium</i>	Na <sub>2</sub> CO <sub>3</sub>		27.1	
<i>Spirulina</i> sp	KOH	heterogeneous catalysts	9.0	[118]
	CH <sub>3</sub> COOH		19.5	
	Na <sub>2</sub> CO <sub>3</sub>		51.6	
<i>S. platensis</i>	Na <sub>2</sub> CO <sub>3</sub>		35	[129]
	Na <sub>2</sub> CO <sub>3</sub>		40	
	Na <sub>2</sub> CO <sub>3</sub>		40	
<i>Tetraselmis</i> sp.	KOH		26.7	[130]
<i>Ulva prolifera</i>	Na <sub>2</sub> CO <sub>3</sub>		20.1	
<i>Green macroalgal blooms</i>	Na <sub>2</sub> CO <sub>3</sub>		20.1	[131]
<i>Chlorella</i> sp	CuO/Al-SBA-15		65.7	
	Pt/C	heterogeneous catalysts	37.9	[133]
	Raney nickel		50	
<i>C. pyrenoidosa</i>	H-ZSM-5		34.2	[134]
	Ce/H-ZSM-5		49.87	
	Pd/C		4	
	Pd/Al <sub>2</sub> O <sub>3</sub>		8	

	H-ZSM-5	73	[134]
	Pt/Al <sub>2</sub> O <sub>3</sub>	38.9	
<i>C. vulgaris</i>	Ni/ Al <sub>2</sub> O <sub>3</sub>	30.0	[135]
	Co/Mo/Al <sub>2</sub> O <sub>3</sub>	38.7	
<i>D. tertiolecta</i>	KtB	49.09	[125]
	Pt/Al <sub>2</sub> O <sub>3</sub>	30.2	
<i>N. oculata</i>	Ni/Al <sub>2</sub> O <sub>3</sub>	18.1	[142]
	Co/Mo/Al <sub>2</sub> O <sub>3</sub>	25.5	
	Pd/C	57	
	Pt/C	49	
	Ru/C	50	
	Ni/SiO <sub>2</sub> -Al <sub>2</sub> O <sub>3</sub>	50	[143]
<i>Nannochloropsis</i>	CoMo/Al <sub>2</sub> O <sub>3</sub>	55	
<i>sp.</i>	Zeolite	48	
	Nano-Ni/SiO <sub>2</sub>	30	[128]
	Pd/C	40	
	Pd/C	38	[135]
	Fe/HZSM-5	38.1	[136]
<i>Spirulina sp</i>	Pd/HZSM-5@MS catalyzed	37.3	[137]
	CeO <sub>2</sub>	34	[138]
<i>S. platensis</i>	Fe <sub>3</sub> O <sub>4</sub>	27.6	[139]
<i>Ulva prolifera</i>	ZSM-5	29.3	[140]
<i>Microalgae</i>			
<i>consortium</i>	H-ZSM-5	16.0	[141]

3.2.1. Heterogeneous catalysis.

Heterogenous catalyst, or water-insoluble catalyst, exists in the different phases with liquefaction medium, therefore they can be recovered and recycled [109]. Another major advantage over homogeneous catalyst is their low corrosion rate and high catalytic activity under severe reaction conditions that often damage the homogeneous catalysts [101].

Several catalysts have been studied for HTL of algal biomass (table 3) including supported metal catalysts (such as Pd, Pt, Ni, Ru), metal oxide catalyst, and metals supported on Al<sub>2</sub>O<sub>3</sub>, SiO<sub>2</sub> and zeolites. However, the influence of metal catalysts in the biocrude yield is complex, and not all of the evaluated metals can positively improve the yield, even some of them can significantly reduce the overall performance of HTL. According to the results obtained by [142], the catalytic activity of Pt, Ni and CoMo supported in Al<sub>2</sub>SO<sub>3</sub> had a positive influence on the yield in the HTL obtained from *C. vulgaris* and *Nannochloropsis occulata*. Results proved that the intrinsic characteristics of each strain (carbohydrates, lipids, protein and ash content) and the catalyst composition play a crucial role on the yield of bio-oil. In this scenario, bio-oil from *C. vulgaris* was positively affected by Pt/Al<sub>2</sub>O<sub>3</sub> and CoMo/Al<sub>2</sub>O<sub>3</sub> (from 34 to 39 wt%); on the other hand, the biocrude yield of *N. occulata* was reduced with each of the three heterogeneous catalysts. Results from strains of the same genera can be completely different. [143] evaluated the performance of several metal catalyst (Pd/C, Pt/C, Ru/C, Ni/SiO<sub>2</sub>-Al<sub>2</sub>O<sub>3</sub>, CoMo/ $\gamma$ -Al<sub>2</sub>O<sub>3</sub> (sulfided), and zeolite) for the conversion of *Nannochloropsis sp* biomass. Their results show a similar of those obtained by [142], since the yield of biocrude obtained with Ni/SiO<sub>2</sub>-Al<sub>2</sub>O<sub>3</sub>, was lower than without catalyst, the reduced oil yield could be due to the promoted gas formation by gasification reactions [109]. However, Pd/C effectively increased the bio-oil yield (from 35 to 57 wt%). In another study, [122] evaluated the efficiency of REHY and Ni/REHY in the conversion of *Dunalliella salina*. Results shown an interesting increase of bio-oil yield from 35%, up to 52 and 72% for REHY and Ni/REHY respectively. The authors found that Ni/REHY catalyst favored the deoxygenation and desulfurization, since the bio-oils were composed mainly of hydrocarbons whereas the content O- and S bearing compounds was negligible; therefore Ni-based catalyst may improve the overall biomass conversion by catalyzing bond cleavages and

depolymerization process. Raney-Ni and HZSM-5 type zeolite (ethanol rather than water as solvent) were evaluated on the catalytic efficiency over *Chlorella pyrenoidosa* biomass [144]. Results show that either catalysts do not improve the yield of bio-oil for the different conditions evaluated. However, the catalyst employed did improved the concentration of different reaction products such as light fuel-range (gasoline range) hydrocarbons. Other zeolite-based catalyst such as H-ZSM-5 and Ce/H-ZSM-5 has been reported for the conversion of *C. pyrenoidosa* biomass [145], their results highlight the efficiency of zeolite-based catalyst, due to a raise in the yield of bio-oil from 32% to 38% and 52% for H-ZSM-5 and Ce/H-ZSM-5 respectively.

Even after all the different research highlighted in the present review, there is no clarity on the underlying mechanism of heterogenous catalyst in the liquefaction process of algal biomass. According to literature, heterogeneous catalyst is considered superior to their counterpart; however, there are some conditions that hamper their efficiency. [146] found that biomass impurities such ash and excess of media nutrients can produce catalyst deactivation after a certain period in a continuous operation. [143,147] found that high concentration of S, N and O derivatives can accelerate the deactivation of heterogeneous catalyst.

## 5. Conclusions

This paper critically reviews the experimental aspects of conventional and catalytic thermochemical conversion of microalgal biomass and their product distribution, yields, and quality. Thermochemical conversion of algal biomass is a promising route to obtain alternative fuels for energy generation; however, several challenges must be overcome to increase the sustainability of algal-based biofuels. Pyrolysis is a well-established technology that shows the right concentration of bio-oil, char and syngas, decent quality, and macroalgal biomass. It can be more interesting for this technology due to the necessity of dried biomass. On the other hand, Hydrothermal liquefaction can convert high-moisture biomass to biocrude in water medium and thus does not require preliminary drying processes, which makes HTL the most promising process an energetic point of view for the conversion of algal-based biofuels. The application of catalyst (both homogeneous and heterogeneous) has increased the overall efficiency of conversion of algal biomass in bio-oil and syngas; however, particular challenges hinder the prospect of industrial application of catalyst, such as possible corrosion on the reaction equipment, low recycling capacity, catalyst deactivation after a certain period in a continuous operation. Therefore, designing novel catalysts for the selective conversion of microalgae into biofuels is a mandatory step to increase the efficiency of the process.

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