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2 **Different responses of bacterial and archaeal** 3 **communities in river sediments to water transfer and** 4 **seasonal changes**

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15 **Abstract:** Bacteria and archaea participate in and are influenced by processes of substance
16 circulation and energy exchanges in natural environment. Generally, the community changes of
17 bacteria and archaea in sediment are mainly driven by seasonality in mid-latitude regions. But in
18 our study, water diversion to Fen river played a more important role on OTU number, diversity
19 and community structure of bacteria and archaea in sediment than seasonal variation, which was
20 found by 16S rRNA high-throughput sequencing technology. This phenomenon might be caused
21 by external transferred water on the physicochemical water environment and accelerated release of
22 positive nitrogen from sediment caused by rise of water level. Changes of carbon-nitrogen cycle
23 and increase of electrical conductivity (EC) value induced more diversion-responders than
24 season-responders both for bacteria and archaea. Seasonal changes have been influencing bacteria
25 and archaea mildly throughout the whole study reach. After water diversion, the environment
26 indicators relating to bacteria community obviously changed from nutrients to salinity while that
27 for archaea almost disappeared. Our research showed the effects of human activities on the
28 communities of bacteria and archaea outweigh the forcing from natural seasonal changes in
29 mid-latitude regions and revealed the mechanism, highlighting different responses of bacteria and
30 archaea to environmental changes.

31 **Keywords:** archaea; bacteria; 16SrRNA high-throughput sequencing; water transfer; seasonal
32 changes; river sediments

33

34 **1. Introduction**

35 Bacteria and archaea are important components of microorganism, which participate in and
36 were influenced by processes of substance circulation and energy exchanges in natural environment
37 [1-3]. Bacteria and archaea belong to two distinct evolutionary branches. Bacteria are widely
38 distributed in all corners of the earth [2], while the abundance and diversity of bacteria goes down
39 under extreme conditions with archaea becoming main microorganisms [4,5]. In the early days,
40 archaea were thought to be all extremophiles that thrived under high temperature, high salinity, low
41 or high pH, absolutely anaerobic or combinations. Scientists speculated archaea's physiological
42 properties may make them weaker competitors to bacteria in a more general niche.

43 What turned the tide is the appearance of PCR amplification and Sanger-based sequencing of
44 16S rRNA genes, which is the beginning of finding archaea in non-extreme environments. Many

45 new archaeal and bacterial lineages (including phyla) have been discovered since then. Recently,
46 studies of comparison between archaea and bacteria in non-extreme environments, including
47 wetlands [6,7], coastal zones [8,9], lakes [10,11] and soil [12,13] attract more and more scientific
48 interest. These results indicate that bacteria and archaea show significant seasonal pattern [14]. For
49 example, the bacteria and archaea in the Alps show obvious seasonal variation, and the abundance
50 of archaea reaches peak in cold spring and winter [15]. Temperature is considered as the main factor
51 driving the seasonal changes of bacterial community in a coastal wetland [16]. Both bacteria and
52 archaea display different community structure and composition in different habitats, and the
53 environmental driving factors are usually different [17].

54 In river sediments, bacteria and archaea dominate the nitrogen cycle [18-20]. Water diversion
55 projects, especially inter-basin water diversion projects, have a profound impact on the bacterial and
56 archaeal communities, and may lead to a strong heterogeneity of the environment in the water
57 receiving area [21,22]. Most water diversion projects are located in the mid-latitude region with
58 dense population and arid/semi-arid climates. The heterogeneity can be further amplified by
59 seasonal changes. It is expected that bacterial and archaeal communities can sensitively respond to
60 the strong spatial-temporal heterogeneity. Previous studies showed that bacteria in water bodies
61 and sediments affected by water diversion have significant spatial-temporal changes [23]. However,
62 the different responses of bacterial and archaeal communities in river sediments to water diversion
63 and seasonality are currently unknown.

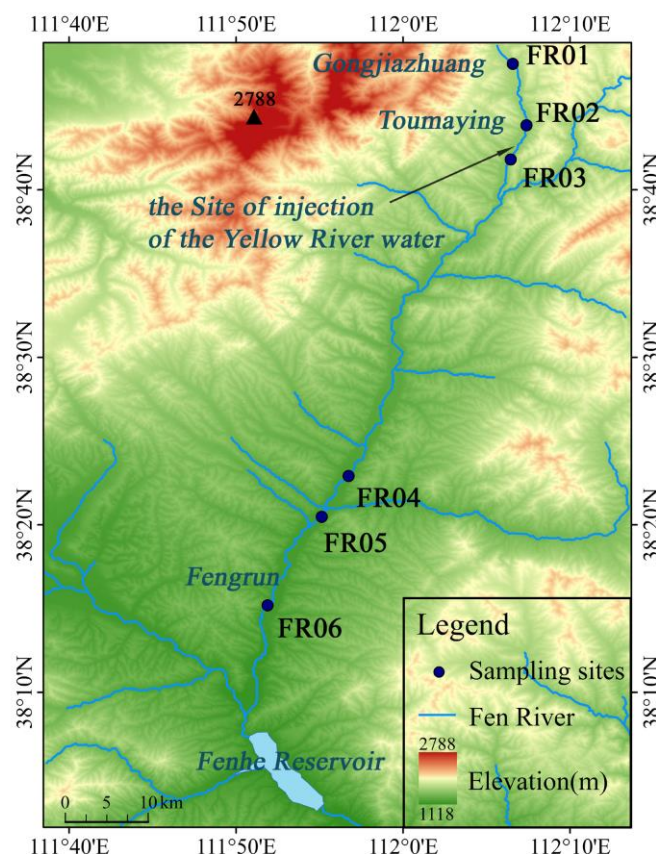
64 Based on 16S rRNA high-throughput sequencing technology, we determined bacterial and
65 archaeal community compositions in sediments influenced by inter-basin water transfer in the
66 mid-latitude river, and discussed the responses of bacterial and archaeal communities to
67 anthropogenic water transfer and the natural seasonality. We aimed to deepen the understanding of
68 bacteria and archaea communities in a changing environment.

69 2. Materials and Methods

70 2.1 Study area and sampling sites

71 The Fen River, 710 km in length, is the second largest tributary of the Yellow River (Fig.1). The
72 study area located in 38° 48' ~ 38° 15' N, 111° 52' ~ 112° 07' E, covering the reach from the river head
73 (Gongjiazhuang) till Fengrun, with a length around 80 km. The research area has a temperate
74 monsoon climate with dry cold winters and wet hot summers. The annual average temperature is
75 about 7°C, with an average temperature of -9°C in January and 21°C in June. The annual rainfall is
76 380 mm to 500 mm, about 70% of which occurs during June to September. The study river reach has
77 been receiving an injection of 320 million m³/a from the Yellow River water in Toumayang since 2003.
78 The water diversion is usually suspended in February, March, August, and September.

79 The sampling site of FR01 (Gongjiazhuang) is about 1 km away from the source of the Fen
80 River. The site of FR02 is 100 m upstream of the Yellow River water injection point (Toumayang). The
81 sampling site of FR03 is 4.5 km downstream of Toumayang, with no tributary. The site of FR04 is
82 located about 50 km downstream of FR03. Multiple tributaries are fed in the river reach between
83 FR03 to FR04, although their total runoff is relatively limited. The sampling site of FR05 is located
84 about 6 km downstream of FR04, followed by FR06 about 15 km downstream.



85
86 **Figure.1** Map of study area and sampling sites

87 **2.2 Sample collection and measurements**

88 Samples were collected from the 6 sampling sites in winter (January) and summer (June) of
89 2017, respectively. Before sampling, physicochemical parameters, such as water temperature, pH,
90 electrical conductivity (EC), oxidation-reduction potential (ORP) and total dissolved solid (TDS) of
91 the river water, were measured (HORIBA U-51). Then, water samples below 20 cm from the water
92 surface in the center of the river were collected and loaded into three 60 mL high density
93 polyethylene bottles. River sediment samples of 1-2cm below the surface of sediments were collected
94 at three different locations of each site, and then loaded into a 10 mL aseptic centrifugal tube. About
95 1 kg sediment samples were collected and put into sterile zip-lock bags. All sediment samples were
96 frozen with dry ice immediately after collection.

97 The contents of Ca^{2+} , Mg^{2+} , Na^+ and K^+ in water samples were determined using ICP-OES
98 (PerkinElmer 5300DV), and the contents of Cl^- , SO_4^{2-} and NO_3^- were determined using Ion
99 Chromatograph (Dionex ICS-900). The contents of HCO_3^- were measured by titration on the
100 sampling day. The particle size (PS) of river sediment samples was analyzed by Laser particle size
101 analyzer (Mastersizer 2000). In addition, total nitrogen (TN), alkaline-nitrogen (AN), total
102 phosphorus (TP), total potassium (TK), olsen-phosphorus (OP), available potassium (AK) and soil
103 organic matter (SOM) were measured according to standard methods (Kjeldahl method for TN,
104 Alkali solution diffusion method for AN, Sodium hydroxide melting method for TP and TK, Sodium
105 bicarbonate extraction method (Olsen) for OP, Ammonium acetate extraction measurement for AK
106 and $\text{H}_2\text{SO}_4 - \text{K}_2\text{Cr}_2\text{O}_7$ external heating oxidation method for SOM). The instruments used include
107 Atomic Absorption Spectrometer (Zeenit of jena, Germany 700 p), Spectrophotometer and Kjeldahl
108 Apparatus.

109 The V3-V4 hypervariable regions of the bacteria and archaea 16S rRNA gene were amplified
110 with primers 338F/806R and 524F10extF/Arch958RmodR by thermocycler PCR system (GeneAmp
111 9700, ABI, USA). The V3-V4 hypervariable regions of the bacteria and archaea 16S rRNA gene were

112 amplified with primers
 113 338F(ACTCCTACGGGAGGCAGCAG)/806R(GGACTACHVGGGTWTCTAAT) and 524F10extF
 114 (5'-TGYCAGCCGCGCGGTAA-3')/Arch958RmodR(5'-YCCGGCGTTGAVTCCAATT-3') using
 115 thermocycler PCR system (GeneAmp 9700, ABI, USA) following the program: 3 min of denaturation
 116 of template DNA at 95°C, 27 cycles of 30s at 95°C, 30s for annealing at 55°C, and 45s for elongation at
 117 72°C, and a final extension at 72 °C for 10 min. Three replicates were used for each sample. Sequence
 118 determination was completed using MiSeq PE300 sequencer (Illumina). The original paired Illumina
 119 MiSeq reads were deposited in the NCBI sequence read archive (accession number: SRP158824). The
 120 sequencing results of the sampling points FR01-FR06 in winter and summer were labeled as W1-W6
 121 and S1-S6, respectively.

122 2.3 Statistical analysis

123 We used T test to analyze the physicochemical characteristics of river water and sediment by
 124 IBM SPSS Statistics 20. To analyze the bioinformation of the high-throughput data, we used QIIME
 125 (Quantitative Insights into Microbial Ecology) to classify the sequences into operational taxonomic
 126 units (OTUs) using a 97% identity threshold. The α -diversity, Coverage, Chao and Shannon indexes,
 127 were calculated using the MOTHUR. Principal component analysis (PCA) was done both for
 128 environmental indicators and microbial community by origin2018. Wilcoxon rank-sum test was
 129 used to identify species that differ significantly from this group. The 'pheatmap' package in R was
 130 employed to visually display the correlation between environmental factors and species.

131 3. Results

132 3.1 Environmental changes induced by water diversion and seasonality

133 A part of environmental factors changed significantly in the water-receiving reach compared to
 134 the regular river reach (Table 1). The average increase of EC and TDS in the water-receiving reach
 135 was about 2.5 times higher than that in the regular river reach. Especially, the concentrations of Cl⁻
 136 and Na⁺ in the water-receiving reach were 1985% and 1725% higher than those in the regular river
 137 reach. In January, the contents of most nutrients in the water-receiving reach sediments increased
 138 obviously compared to the regular river reach, with an increase of 156% for AN, 125% for AK, 89%
 139 for OP, 88% for TN, and 58% for SOM. However, in June, TN, AN, and OP decreased by 44%, 54%
 140 and 55% respectively in the water-receiving reach sediments compared to the regular river reach. On
 141 average, PS in the sediments of the water-receiving reach decreased by 51% than that in the regular
 142 river reach (Table 2).

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Table.1 Main water environmental factors affected by Yellow River diversion and seasonal changes

	January		June	
	Regular reach ¹	Water-receiving reach ²	Regular reach ¹	Water-receiving reach ²
T(°C)	1.44±1.01	1.15±0.52	19.59±0.07*	21.90±0.56*
EC(μs/cm)	535.50±28.50**	1257.50±27.50**	436.00±34.00**	1205.00±32.50**
ORP(mv)	410.00±55.00	366.50±15.25	209.50±26.50	199.75±22.25
TDS(g/L)	0.34±0.02**	0.81±0.02**	0.28±0.02**	0.77±0.02**
NO ₃ (mg/L)	11.95±1.65*	19.65±0.45*	12.35±2.55	11.38±1.69
SO ₄ ²⁻ (mg/L)	70.05±6.65**	205.53±3.66**	121.60±0.90*	400.85±116.28*
Cl(mg/L)	6.55±0.75**	132.25±3.78**	7.00±0.40**	150.85±67.63**
Na ⁺ (mg/L)	6.65±0.55**	121.10±3.80**	8.40±1.20**	152.20±3.90**
K ⁺ (mg/L)	1.80±0.00	3.88±0.04	2.39±0.28	4.88±0.57

Mg ²⁺ (mg/L)	15.45±0.65**	36.95±0.58**	16.84±1.50*	40.94±0.57*
Ca ²⁺ (mg/L)	63.70±0.20	67.60±1.60	76.31±6.94	65.19±3.73

145 Notes:

146 ¹ Regular reach: FR01~FR02; ² Water-receiving reach: FR03~FR06.

147 * represents the value of non-water-receiving reach and water-receiving reach's significant difference were
 148 achieved at the 0.05 level; ** represents the value of non-water-receiving reach and water-receiving reach's
 149 significant difference were achieved at the 0.01 level.

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Table.2 Main soil environmental factors affected by Yellow River diversion and seasonal changes

	January		June	
	Regular reach ¹	Water-receiving reach ²	Regular reach ¹	Water-receiving reach ²
PS(um)	105±33	53±18	247±147	116±61
pH	7.80±0.05	8.05±0.16	8.35±0.44	8.91±0.13
SOM(g/kg)	14.65±1.85	23.18±8.28	11.85±3.65	16.03±4.18
TN(mg/kg)	4.40 ±0.00	8.25 ±0.03	11.30 ±0.00*	6.38 ±0.01*
AN(mg/kg)	33.00±2.00*	84.25±23.13*	160.00±29.00*	73.00±8.50*
TP(mg/kg)	464.50±36.50	578.50±45.25	530.00±50.00	589.25±44.38
OP(mg/kg)	5.50±0.50	10.25±3.88	17.00±4.00	7.75±1.75
TK(mg/kg)	2.83±0.01	2.70±0.19	2.82±0.20	2.35±0.14
AK(mg/kg)	48.50±6.50*	109.75±24.25*	69.00±1.00	78.00±15.50

152 Notes:

153 ¹ Regular reach: FR01~FR02; ² Water-receiving reach: FR03~FR06.

154 ³ PS: Particle size; ⁴ SOM: Soil organic matter; ⁵ TN: Total nitrogen; ⁶ AN: Alkali-nitrogen; ⁷ TP: Total
 155 phosphorus; ⁸ OP: Olsen-phosphorus; ⁹ TK: Total kalium; ¹⁰ AK: Alkali-kalium.

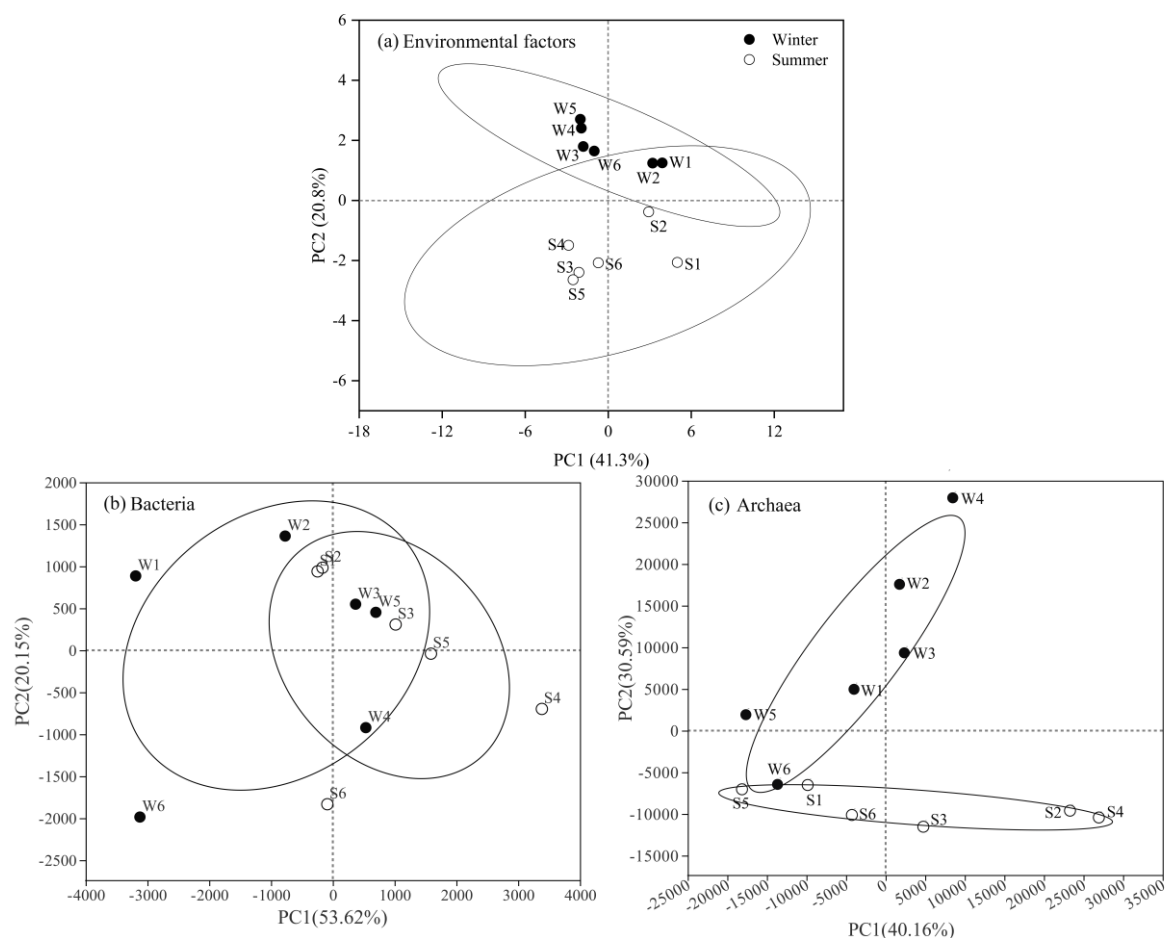
156 * represents the value of non-water-receiving reach and water-receiving reach's significant difference were
 157 achieved at the 0.05 level; ** represents the value of non-water-receiving reach and water-receiving reach's
 158 significant difference were achieved at the 0.01 level.

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160 A part of environmental factors showed seasonal variation obviously. From winter (January) to
 161 summer (June), the river water temperature, Na⁺, Cl⁻ and SO₄²⁻ in the river water increased 1260.4%,
 162 26.3%, 6.9%, 74.2% and 1804.3%, 25.7%, 14.1%, 94.7%, while ORP decreased 48.9% and 45.5% in the
 163 regular reach and water-receiving reach, respectively (Table 1). From winter to summer, PS and pH
 164 of the river sediment increased, while SOM decreased 19.0% and 30.6% in the regular reach and
 165 water-receiving reach, respectively (Table 2).

166 The water diversion and seasonality cause significant alteration in environmental factors, which
 167 induced microbial community changes. Based on the result of the principal components analysis
 168 (PCA) of environmental factors, samples in the regular river reach and the water-receiving reach
 169 were separated by the first principal component. The environmental factors, including TDS, EC, Na⁺,
 170 K⁺, Mg²⁺, Cl⁻, and SO₄²⁻, all responded actively to the water transfer constituting the first principal
 171 component. Therefore, the first principal component could recognize the alteration in microbial
 172 community induced by the water diversion. At the same time, the second principal component
 173 distinguished samples from summer and winter (Fig. 2a). Water temperature and ORP significantly
 174 responded to seasons forming the majority of the second principal component. It is considered that
 175 the second principal component can identify the alteration in microbial community induced by
 176 seasonality. Furthermore, the first principal component explained 41.3% of the total variance of the

177 environment factors, while 20.8% of the total variance was explained by the second principal
 178 component. It is indicated that the influence of water diversion on the physicochemical conditions of
 179 water and sediments is beyond the influence of seasonality.



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181 **Figure.2** PCA (Principle component analysis) of environmental factors(a), bacteria(b) and archaea(c) in winter
 182 and summer. 95% confidence ellipses of sampling groups were showed.

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184 3.2 OTU number and diversity of bacteria and archaea

185 Under the influence of water transfer, OTU number and diversity of bacteria increased while
 186 that of archaea decreased. The average OTU numbers of bacteria were about 2750 and 3329 in the
 187 regular reach and the water-receiving reach, respectively (Table 3). The average OTU numbers of
 188 archaea were 538 and 335 in the regular reach and the water-receiving reach. The average Shannon
 189 diversity index of bacteria were about 6.2 and 6.5 in the regular reach and the water-receiving reach,
 190 respectively. The average Shannon diversity index of archaea were about 3.3 and 3.0 in the regular
 191 reach and the water-receiving reach.

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Table 3 OTU number and α -diversity index of bacteria and archaea

Sample\Estimators	Season	OTUs		Shannon Index		Chao Index		Coverage Index	
		Bacteria	Archaea	Bacteria	Archaea	Bacteria	Archaea	Bacteria	Archaea
FR01	Winter	2822	590	6.06	3.56	3768.44	733.13	0.98	0.99
	Summer	2149	504	5.94	3.32	2859.82	602.89	0.99	0.99
FR02	Winter	3146	512	6.51	3.10	4103.01	610.25	0.98	0.99
	Summer	2884	456	6.42	3.17	3857.02	619.43	0.98	0.99

FR03	Winter	3631	582	6.76	3.26	4722.66	789.59	0.97	0.99
	Summer	3549	392	6.86	2.95	4470.78	553.25	0.98	0.99
FR04	Winter	3659	363	6.77	3.08	4717.44	462.13	0.98	0.99
	Summer	3602	237	6.92	3.03	4479.15	326.44	0.98	0.99
FR05	Winter	3441	288	6.51	2.42	4513.04	380.57	0.98	0.99
	Summer	3737	298	7.01	2.94	4746.85	385.15	0.98	0.99
FR06	Winter	2708	335	5.79	2.89	4044.36	450.93	0.98	0.99
	Summer	2303	348	5.76	3.06	3261.68	461.56	0.98	0.99
Average	Winter	3234.5	445	6.40	3.05	4311.49	580.51	0.98	0.99
	Summer	3037.3	372.5	6.48	3.08	3945.88	491.45	0.98	0.99

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The average OTU number of bacteria and archaea was higher in winter than that in summer, which is the same as the result of Chao index, indicating the abundance of bacteria and archaea was higher in winter than in summer. However, Shannon diversity of bacteria and archaea increased from summer to winter in different degrees (1.23% for bacteria, and 0.97% for archaea). The sequencing data are sufficiently representative because the coverage indexes of all the twelve samples were greater than 0.974.

In our case, the water diversion induced greater impacts on the OTU number and diversity of the microbial community than seasonality. Compared to the regular reach, the average of OTU number decreased by 37.7% for archaea, while the average of OTU number increased by 21.1% for bacteria. Moreover, the average of Shannon diversity index increased by 5.1% for bacteria and decreased by 11.3% for archaea because of water diversion. From summer to winter, the average OTU number increased by 6.5% for bacteria and 19.5% for archaea, respectively. Moreover, the average of Shannon diversity index decreased by 2.6% for bacteria and 1.0% for archaea.

The sites of FR02 and FR03 are located close to each other and are respectively distributed in the upstream and downstream of Toumayang where the Yellow River water is injected into the Fen River. The number of bacteria OTU and Shannon diversity index increased in the site FR03 as the result of the water diversion.

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3.3 Bacteria and archaea community structure and classification

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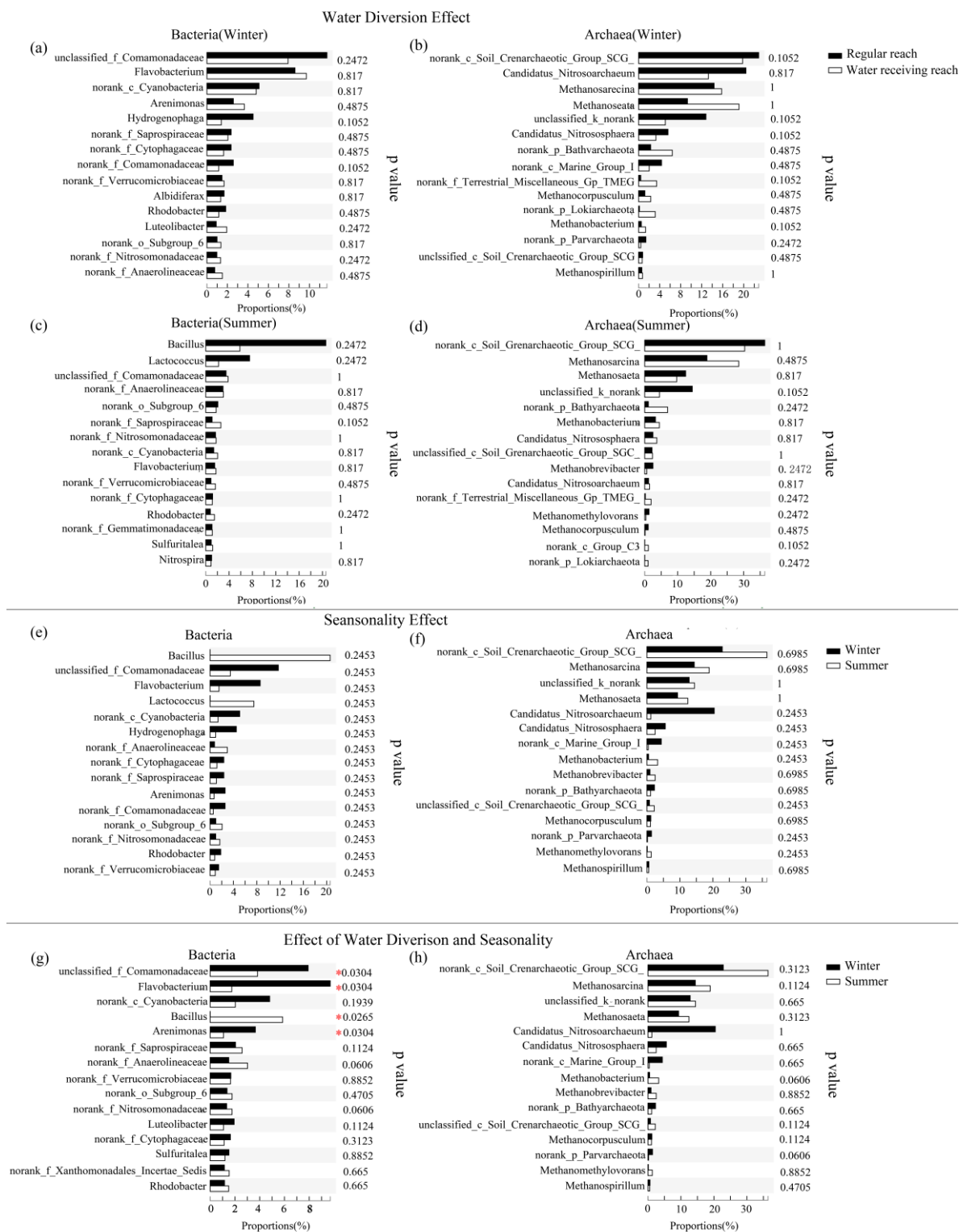
PCA used sample points and 95% confidence ellipse to represent the similarity and difference of community composition between sample groups. The influence of water diversion on sediment bacteria was beyond the influence of seasonality in the Fen river. The first and second principal coordinates of the weighted analysis accounted for 53.62% and 20.15% of the variation in the data, respectively (Fig.2b). PC1 clearly separated the samples in FR01, FR02 and FR06 from other samples. FR01 and FR02 located at the upstream of the place the Yellow River water injected into Fen river and FR06 after the long-distance self-purification of water bodies so they were not affected by the water diversion project and the bacterial community structures of them were quite different from other sampling sites. PC1 axis represented the influence of water diversion project on bacterial community. Meanwhile, the distribution of bacteria samples was compact in winter and dispersed in summer, indicating that the spatial heterogeneity of bacterial community structure was greater in summer than in winter.

PC1 axis (accounted for 40.16%) separated the samples in FR01, FR05 and FR06 from other samples which indicated PC1 axis represented the influence of water diversion project on archaea community and water diversion project on archaea of Fen river sediments had implications on FR02-FR04 (Fig.2c). The spatial distance of the effect on archaea was less than that on bacteria. The PC2 axis (accounted for 30.5%) failed to separate the samples of the archaea community in different seasons.

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3.4 Identification of diversion-responders and season-responders

231 Responses of bacteria and archaea with 15 highest abundance to the water diversion or seasonal
232 variation were identified by Wilcoxon rank-sum test (Fig. 3). The bacteria and archaea that have p
233 value less than 0.11 were selected as responders [24,25]. The bacteria and archaea that have p value
234 less than 0.25 but more than 0.15 were thought as responded environmental change obviously. For
235 bacteria in the genera level, *Hydrogenophage* and *Cyanobacteria* ($p = 0.1052$), *Comamonadaceae*,
236 *Luteolibacter* and *Nitrosomonadaceae* ($p = 0.2472$, Fig.3a) obviously responded to the water diversion in
237 winter, while *Saprospiraceae* ($p = 0.1052$), *Bacillus*, *Lactococcus*, *Rhodobacter* ($p = 0.2472$ Fig. 3b) can
238 respond distinctly in summer. Diversion-responders are more in winter than in summer. The results
239 of Qv also showed that the bacteria were more responsive to water diversion in winter [23]. The
240 possible reason was that the flow of rivers was more dependent on diversion in winter due to the
241 monsoon climate in the mid-latitudes. No bacteria were selected as season-responders but all
242 bacteria with 15 highest abundance responded seasonality obviously (Fig.3e). *Bacillus*,
243 *Comamonadaceae*, *Flavobacterium* and *Arenimonas* responded significantly to both water diversion and
244 seasonality ($p > 0.05$, Fig.3g). *Bacillus* decreased while *Comamonadaceae*, *Flavobacterium* and
245 *Arenimonas* increased from winter to summer. In our case, the diversion-responders are more than
246 the season-responders.



247
 248 **Figure.3** The strength of bacteria(a) and archaea(b) in winter, bacteria(c) and archaea(d) in summer responded
 249 to water diversion. Strength of bacteria(e) and archaea(f) in regular reach responded to seasonality. Strength of
 250 bacteria(g) and archaea(h) in water receiving reach responded to water diversion and seasonality by
 251 Kruskal-Wallis H test. The significant level was symbolled as *: $p \leq 0.05$.

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 253 The archaea genera responded obviously to the water diversion included *SCG*, *Nitrososphaera*,
 254 *Terrestrial Miscellaneous Euryarchaeal Group (TMGE)*, *Methanobacterium*, *unclassified_k_norank* ($p =$
 255 0.1052) and *Parvarchaeota* in winter ($p = 0.2472$, Fig. 3b). In summer, archaea genera response

256 obviously to the water diversion included *Unclassified_k_norank archaea*, *Methanocorpusculum* ($p =$
257 0.1052) and *Bathyarchaeota Methanobrevilbacter*, *TMEG*, *Methanomethylovorans*, *Lokiarchaeota* ($p =$
258 0.2472, Fig. 3d). There were more archaea obviously responding to the water diversion in winter.
259 There were no season-responder archaea in water-receiving reach (Fig. 3f). And no archaea with 15
260 highest abundance responded significantly to both water diversion and seasonality (Fig. 3h).

261 The responses of the bacteria and archaea with 15 highest abundance groups to the water
262 diversion were more obvious than that to the seasonal variation. The effect of the water diversion on
263 bacteria and archaea was more obvious in winter. In the regular river reach where seasonal variation
264 factors impacted alone, the response of bacteria with 15 highest abundance was extensive and
265 uniform, while that of archaea with 15 highest abundance was weak. In the water-receiving river
266 reach where seasonal changes and the water diversion impacted simultaneously, the response of 4
267 genera bacteria was significant, while only one species of archaea showed a relatively obvious
268 response.

269 4. Discussion

270 4.1 Water transfer is a more significant driver of microbial community changes than seasonality

271 Seasonal variation is often the most important factor affecting water system bacteria and
272 archaea in mid-latitudes. Seasonal dynamics had a greater impact on the formation of bacterial
273 communities than the effects of anthropogenic pollution on rivers [26]. The seasonal variation of
274 bacterial and archaeal structures didn't change after wetland drainage [27]. Significant seasonal
275 changes and non-significant spatial changes can be simultaneously observed in the beta diversity
276 and community structure of sediment bacteria in a mid-latitude lake [28]. The seasonal variation of
277 rainfall was considered as a key factor affecting the abundance and diversity of bacteria and archaea
278 [29]. However, the water diversion is a more significant driver of microbial community OTU
279 number, diversity index, and community structure variation than seasonality in our case.

280 In this study, the average Shannon index of bacteria and archaea in summer was higher than
281 that in winter (Table 3). Generally, diversity of bacteria and archaea is high in warm seasons and low
282 in cold seasons [30-32]. It was reported that the bacterial diversity in river sediments from five
283 different pollution sites was higher in summer than in winter [33]. Microorganisms always show the
284 best temperature characteristics [34,35]. The composition of bacterial community and archaea
285 community changed with the fluctuation of water temperature. Increased nutrient levels also
286 contribute to the growth of bacterial communities in summer [36]. Seasonality of water temperature
287 and nutrition could play an important role in seasonal variance of bacteria and archaea. It was
288 considered as the reason that the densely growing phytoplankton and benthic plants in summer
289 could stimulate the specific activity, growth efficiency and proliferation of bacterial cells [38,39]. The
290 assimilation material released by these organisms is used by bacteria as a food or energy source,
291 increasing their abundance. Furthermore, farmland irrigation in summer can lead to the increase of
292 nitrogen and phosphorus content in natural channels which could explain the increase of total
293 nitrogen and phosphorus in Fen River (Table 2) [37]. Therefore, the content of nitrifying and
294 desulphurizing bacteria increased [38,39].

295 Our data also showed the average OTU numbers of bacteria and archaea in summer were lower
296 than that in winter in water-receiving reach (Table 3). It is notable that OTU numbers of bacteria and
297 archaea decline in summer. In previous study, OTU number of bacteria and archaea were also high
298 in summer and low in winter [30-32]. Salinity changes should be responsible for the result. The
299 salinity of water is determined by soil leaching, rock weathering, and pollution [40,41]. The OTU
300 numbers of bacteria and archaea in freshwater ecosystems were significantly positively correlated
301 with EC value of water (as an indicator of salinity) [42]. In our case, water transfer from the Yellow
302 River brought not only nutrients (caused an increase of 43.53% for TN, and an increase of 17.86% for
303 TP) but also salinity (increased by 2.5 times) to the Fen River. The receiving reach of the Fen River
304 was more affected by the water diversion in winter. According to these reports and our results, we

305 speculated this abnormal phenomenon in our case was because water diversion project was a more
306 significant diver of microbial community changes than seasonality.

307 Responses of bacteria and archaea's community structures to water diversion were more
308 significant than that to seasonality. *P* value represents the possibility of rejecting hypothesis that
309 water diversion or seasonality has no effect on microbial community changes [25]. When water
310 diversion impacted alone, the *p* value of the sediment bacterial and archaeal community was as low
311 as 0.1052. When the seasonality impacted alone, the *p* values of that were 0.2472 (Fig.3). The results
312 indicate that the water diversion is more possible factor than the seasonal variation of environment
313 in promoting changes in a microbial community structure.

314 Under the strong influences of the water diversion, the OTU number and diversity of bacteria
315 and archaea responded more strongly to seasonal variation (Table 3). We could offer two
316 explanations relating with the water transfer for the strong response of bacteria and archaea. One
317 was high concentration of nutrients, EC value and pH value in water body of the Yellow River went
318 into Fen river by water diversion project. It was reported that the TN concentration of the Yellow
319 River sediments (35.3 ~ 1142.5 mg kg⁻¹) was much higher than that of the Fen river sediments. EC
320 value and pH value in the transferred Yellow River was stable above 1000 μS/cm and 8, respectively
321 [43]. Therefore, water diversion obviously increased the TN concentration by 88% in winter (Table 2)
322 and the EC value from 535 μS/cm to 1257 μS/cm in winter and from 436 μS/cm to 1205 μS/cm in
323 summer, pH value of the water-receiving river from 7.8 to 8.05 in winter and from 8.35 to 8.91 in
324 summer (Table 1). Nitrogen is important for nitrifying and denitrifying bacteria and archaea. Most
325 archaea with 15 highest abundance were methanogens (according to Berger's handbook of
326 systematic bacteriology, Fig.3). The number of methanogens obviously decreases with salinity [44].
327 The increased pH value can obviously inhibit the OTU number of archaea [45]. As a seasonal river,
328 Fen river had low water level and velocity in winter naturally, so water from the Yellow River
329 played a more important role. The other was, in Fen reaches, the influence of water diversion on
330 river water volume and velocity exceeds seasonality. Water from the Yellow River was injected at a
331 rate nearly 15 m³/s, which was roughly 30 times higher than the dry-season flow of the Fen river at
332 this site [46], bringing high water velocity, which strongly dilutes the bacteria in the sediment, thus
333 producing positive nitrogen [47,48]. TN was one of the key factors relating to bacterial community
334 structure and composition which would affect bacteria and archaea obviously [49].

335 4.2 Diversion-responders more than season-responders

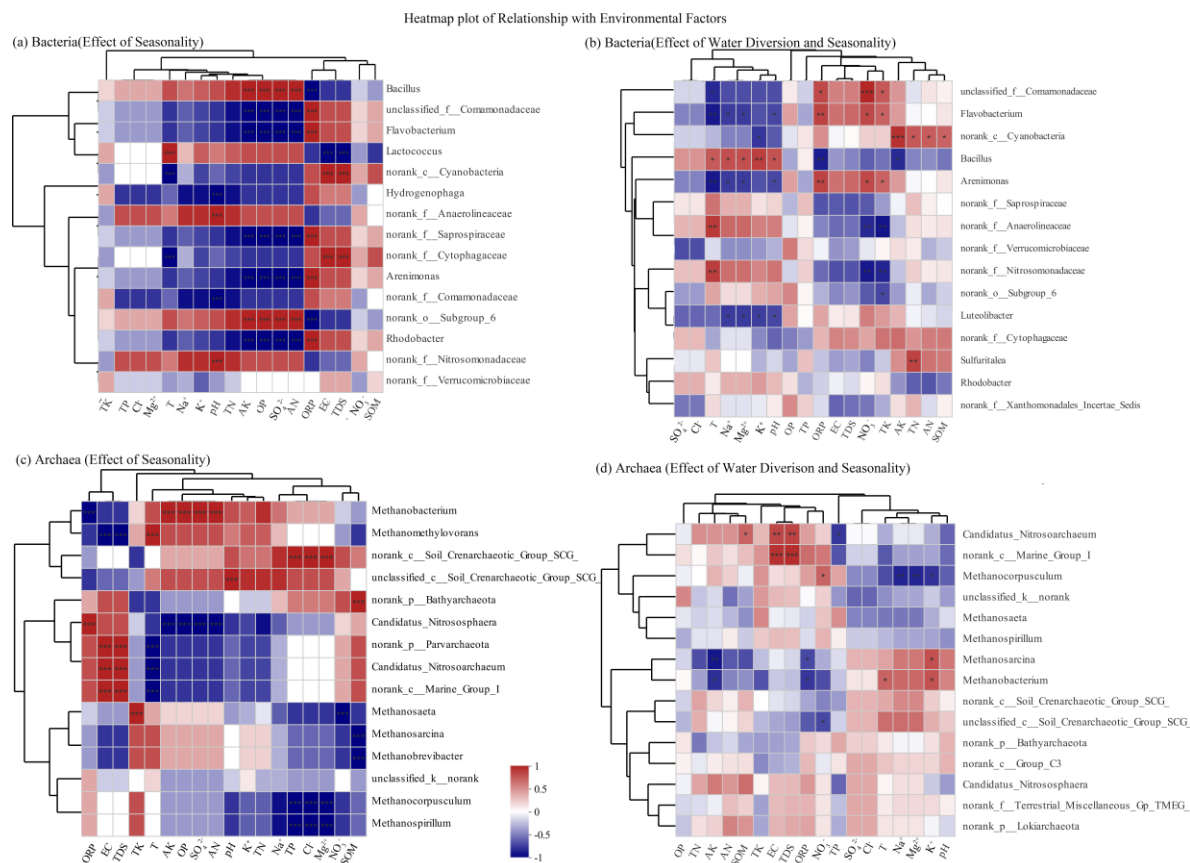
336 In our case, diversion-responders were more than season-responders in bacteria (Fig.3). Almost
337 all diversion-responders are carbon - nitrogen cycle related. When total nitrogen concentration
338 increasing, these bacteria would respond obviously. *Hydrogenophaga* is a kind of denitrifying bacteria
339 and had positive interactions with other denitrifying bacteria [50]. Cyanobacteria is positively
340 correlated to nitrogen concentration [51]. *Comamonadaceae* is a common denitrifying bacterium in
341 water [52]. *Luteolibacter* is independent of the nitrogen cycle and mainly uses iso-C_{16:0} and iso-C_{15:0},
342 which play an important role in the carbon cycle in the environment [53]. *Nitrosomonadaceae* is a
343 typical nitrite oxidizing bacterium [54]. *Saprospiraceae* have the ability to hydrolyze and utilize
344 complex carbon sources, and this bacterium may also play an important role in the decomposition of
345 complex organisms [55]. Compared with conventional denitrification bacteria, the *Flavobacterium*
346 grow well under aerobic conditions and show good denitrification activities [56-59]. *Bacillus* are
347 often used to indicate good quality water bodies [60]. *Bacillus* remained one of the dominant
348 bacteria, although its abundance declined with the onset of winter or water degradation after
349 diversion. Some of the bacteria in this genus were heterotrophic nitrifying bacteria and vulcanized
350 bacteria, which had obvious response to nitrate nitrogen [61]. *Lactococcus* is a pathogenic bacterium
351 capable of causing sepsis in fish and humans [62,63], and is also a common heterotrophic
352 denitrifying bacterium in the environment [64]. *Rhodobacter* contain autotrophic bacteria and
353 heterotrophic bacteria that can utilize carbon sources [65]. Xing [66] used a stable-isotope probing
354 technique and reported that *Arenimonas* is capable of removing nitrate. Denitrification bacteria were
355 dominant in the sediments of the water-receiving reach of Fen River, and the nitrogen in this reach

356 was fractionated [67]. The main reason for the rise in nitrogen concentration in river water is
357 diversion (88 % for water diversion, 39% for seasonal variation, Table 2). Due to that, more
358 responders were sensitive to water diversion.

359 We had more diversion-responders than season-responders in archaea (Fig.3). We believed the
360 reason diversion-responders were more than season-responders in archaea is that EC value
361 increasing of the water-receiving river. *Methanobacterium*, *Methanocorpusculum*, *Methanobrevibacter*,
362 *Methanomelovorans* are all typical methanogens [44,68]. Methanogens occupy a unique niche in the
363 anaerobic environment and are the ultimate acceptor of electrons [69]. We speculated that the
364 dominant position of methanogens increased after water transfer, because the increase of water EC
365 provided electrons for methanogens. Possibly, this characteristic of the dominant methanogenic
366 archaea caused more archaea species with a *p* value of less than 0.2472 than bacteria in the
367 water-receiving reach. *Nitrososphaera* belongs to ammonia-oxidizing archaea, and its response to
368 most inhibitors was weaker than that of ammonia-oxidizing archaea [70]. TMEG has been detected
369 many times under extremely acidic conditions, and scientists speculated that this genus of archaea
370 had acidophilic members [71,72]. Some studies have shown that *Parvarchaeota* was only found in
371 acid mine drainage and hot spring habitat, and belongs to microaerobic/anaerobic archaea, which
372 was involved in carbon, nitrogen and iron cycling [73]. However, in the Fen reaches, *Parvarchaeota*
373 emerged as the dominant archaea, indicating that this genus of archaea had a wider habitat
374 distribution than expected. *Bathyarchaeota*, with complex organic proteins, lipids and benzoic acid as
375 the main metabolites, is widely found in the environment [74]. *Lokiarchaeota* is a genus of archaea
376 with a series of pathways involving the carbon, nitrogen, sulfur and phosphorus cycles in its
377 macrogenes [75,76].

378 4.3 Responses of high-abundance bacteria and archaea to environmental factors reduced by water diversion

379 When seasonal factors impact alone, ORP, EC, TDS, pH, temperature, SO_4^{2-} , AK, OP and AN
380 significantly affected bacteria. The bacteria with 15 highest abundance showed the opposite
381 response to ORP, EC, TDS and pH, temperature, AK, OP, SO_4^{2-} , AN which was named as niche
382 complementation effect (Fig. 4a). *Bacillus* and *Lactococcus*, *Anaerolineaceae*, *Supgroup_6*,
383 *Nitrosomonadaceae* showed obvious negative correlation to ORP, EC, TDS, and positive correlation to
384 pH, temperature, AK, OP, SO_4^{2-} , AN. *Comamonadaceae*, *Flavobacterium*, *Cyanobacteria*, *Hydrogenophaga*,
385 *Saprosiraceae*, *Cytophagaceae*, *Arenimonas*, *Comamonadaceae*, *Rhodocater* had obvious positive correlation
386 with ORP, EC, TDS, and significant negative correlation to pH, temperature, AK, OP, SO_4^{2-} , AN. In
387 previous studies, temperature was often considered to be one of the key environmental factors that
388 caused seasonal changes in bacterial communities in aquatic ecosystems. Inter-seasonal temperature
389 fluctuations could partly explain seasonal changes in bacterial community structure and diversity
390 [33]. During the rainy season, the warm environment was conducive to the growth of bacteria [77].
391 Temperature and salinity were the key environmental factors leading to the seasonal dynamics of
392 bacterial communities [78]. Our results demonstrated that the seasonally-driven mechanisms
393 leading to changes of bacterial communities in the river sediments were actually more complex.
394 Nutrients also played an important role on affecting bacteria in seasonal change process.
395 Interestingly, *Verrucomicrobiaceae* exhibited weak or no correlation with the above environmental
396 factors but it was often the dominant bacterium after water transfer [43,79].



397
398 **Figure.4** Spearman coefficients between bacteria(a) and archaea(b) in regular reach, bacteria(c) and archaea(d)
399 in water-receiving reach and environmental parameters. The significance level was symbolled as: *: $p \leq 0.05$.

400

401 When factors of water diversion and seasonal variation impacted collectively, the
402 environmental factors that had a significant impact on the main bacterial communities in
403 water-receiving reach were significantly responsive to water diversion, including temperature, Na⁺,
404 Mg²⁺, K⁺, pH, ORP, NO₃⁻, and TK (Fig.4b), which indicating water diversion had an important effect
405 on the dominant bacteria. The bacteria with 15 highest abundance in water-receiving reach could be
406 divided into two types that were complementary to the response to environmental factors. The
407 bacteria in the top 15 abundance of water-receiving reach showed opposite responses to
408 temperature, Na⁺, Mg²⁺, K⁺, pH and ORP, NO₃⁻, TK. Among them, *Bacillus*, *Anaerolineaceae*,
409 *Supgroup_6*, *Nitrosomonadaceae* showed obvious positive correlation to temperature, Na⁺, Mg²⁺, K⁺,
410 pH, while presented obvious negative correlation relationship to ORP, NO₃⁻, TK. Among them, *Bacillus*,
411 *Flavobacterium*, *Arenimonas*, *Luteolibacter* showed significant negative correlation relationship to
412 temperature, Na⁺, Mg²⁺, K⁺, pH, presented obvious positive correlation relationship to ORP, NO₃⁻,
413 TK. In particular, Cyanobacteria showed obvious negative correlation with temperature, Na⁺, Mg²⁺,
414 K⁺ and pH, little correlation with ORP, NO₃⁻ and TK, and a significant positive correlation with
415 indicators indicating nutrient concentration such as AK, TN, AN and SOM.

416 Temperature, pH and ORP have been the key environmental factors both in the regular reach
417 and water-receiving reach. Temperature and ORP had a significant response to seasonal change
418 (Table 1), indicating that the influence of seasonal change has been impacting on the whole study
419 reach, which was the same as previous studies [80,81]. Therefore, the species in the top 15 abundance
420 showed obvious response to seasonal changes ($p \geq 0.2472$, Fig.3g). In contrast, EC, TDS, AK, OP, SO₄²⁻,
421 and AN were no longer the key environmental factors in water-receiving reach. They were
422 transformed into Na⁺, Mg²⁺, K⁺, NO₃⁻, and TK. Water transfer project made nutrients impact on
423 sediment bacteria was abate, river sediment bacteria mainly controlled by salinity.

424 In previous studies, *Cyanobacteria* tended to be positively correlated with temperature [82,83].
425 However, Haakonsson's study [84] proved that this phenomenon only existed in the static water

426 system. The growth promotion effect of temperature rise on the *Cyanobacteria* was greatly weakened
427 in the flowing river [85,80]. The study of Amazon river reach with many tributaries showed that
428 discharge and water salinity were the principal variables for constructing a series of environmental
429 changes [86]. Our results supported both theories. In this study, the abundance of *Cyanobacteria* in
430 winter was much higher than that in summer. We speculated water body was influenced more in
431 winter by the water diversion because of the increase of water flows velocity, a good water
432 circulation and increased the nutrient level in winter, which resulting in weakening of positive
433 correlation between *Cyanobacteria* and temperature. Due to that, *Cyanobacteria* increased in winter
434 and showed negative correlation to temperature. This mechanism also affected *Flavobacterium*, a
435 kind of aerobic denitrifying bacteria, was provided aerobic conditions and food sources by higher
436 water flow velocity after the water transfer, which inhibited the growth of other denitrifying
437 bacteria. Therefore, the *Flavobacterium* had become the dominant bacteria in the water receiving
438 reach and showed the same relationship with environmental indicators as *Cyanobacteria*.

439 When seasonal factors impact alone, almost all the environmental factors measured could cause
440 significant responses in archaea in the top 15 of abundance (Fig.4c). These archaea could be divided
441 into three categories according to their response to environmental factors. (1) *Methanobacterium*,
442 *Methanomethylovorans* and SCG showed obvious positive correlation to temperature, AK, OP, SO_4^{2-} ,
443 AN, pH, K^+ , TN, Na^+ , TP, Cl^- , Mg^{2+} and NO_3^- , while showed obvious negative correlation to ORP, EC
444 and TDS. Among them, the correlation index between *Methanomethylovorans* and Cl^- , Mg^{2+} , NO_3^- was
445 0. (2) The correlation index of *Norank_c_Soil_Crenarchaeotic_Group_SCG_* to EC, TDS and
446 temperature was 0. *Bathyarchaeota*, *Nitrososphaera*, *Parvarchaeera*, *Nitrosoarchaeum*, *Marine_Group_I*
447 presented obvious positive correlation to ORP, EC, TDS, and showed significant negative correlation
448 relationship to the temperature, AK, OP, SO_4^{2-} , AN, pH, K^+ , TN; (3) *Methanosaeta*, *Methanosarcina*,
449 *Methanobrevibacter*, *Methanocorpusculum* and *Methanospirillum* showed significant negative correlation
450 to AK, OP, SO_4^{2-} , AN, pH, K^+ , TN, Na^+ , TP, Cl^- , Mg^{2+} , NO_3^- , SOM, for TK was significantly positive
451 correlation.

452 When factors of water diversion and seasonal variation impacted at the same time, the response
453 mechanism of archaea community to environmental factors in Fen sediments in water-receiving
454 reach became complex (Fig. 4d). There was only a significant correlation between a few
455 environmental factors and a few archaea. After water diversion, the number of environmental
456 factors that could affect the archaea in top 15 abundance decreased at the significance level.

457 In regular river reach only affected by seasonal factors, the bacteria and archaea in top 15
458 abundance had obvious responses to environmental factors. In water-receiving reach affected by
459 seasonal factors and water diversion factors, the correlation of microorganism and environmental
460 factors reached significance level decline. At the same time, abundance of microorganism was not
461 sensitive to environmental factors changed of increase. Resulting in microorganism of Fen river
462 sediment with 15 highest abundance response to environmental factors of was weak totally, the
463 response mechanism became more complex. According to Qu [23], after water diversion, the
464 ecological niche range of microorganism (OMI value) in the water-receiving reach increased,
465 competition among dominant species became fiercer, and species' ability to utilize a variety of
466 environmental resources was enhanced. So, the response of a single species to a particular
467 environmental factor was weaker. At the same time, water diversion and seasonal factors impacted
468 together, the species with no obvious response to environmental factors were more likely to become
469 the dominant bacteria.

470 5. Conclusions

471 The influence of the water diversion on environmental factors and OTU number, diversity
472 index and community structure of bacteria and archaea in the water-receiving river beyond seasonal
473 variation. We had two explanation of this phenomenon. One was high concentration of nutrients, EC
474 and pH value in water body of the Yellow River went into Fen river by water diversion project. As a
475 seasonal river, Fen river had low water level and velocity in winter naturally, so water from the
476 Yellow River played a more important role. The other was, in Fen reaches, the influence of water

477 diversion on river water volume and velocity exceeds seasonality to producing positive nitrogen.
478 Therefore, water diversion had a greater impact on bacteria and archaea in winter.

479 According to specific diversion-responders and season-responders were identified, we had
480 more diversion-responders than season-responders both in bacteria and archaea. We speculated the
481 reason diversion-responders were more than season-responders in bacteria is that almost all
482 responders are carbon - nitrogen cycle related. When total nitrogen concentration increasing, these
483 bacteria would respond to them. For archaea, the reason is that EC value increasing of the
484 water-receiving reach. Methanogens dominated archaeal community in Fen river and could be
485 inhibited by high EC value.

486 Temperature, pH and ORP have been the key environmental factors both in the regular reach
487 and water-receiving reach. After water diversion, the environment indicators could affect bacteria
488 community changed from nutrients to salinity. While the environment indicators could affect
489 archaea community almost disappeared. Interestingly, the correlation between bacteria and archaea
490 and environmental factors was obviously weakened when water diversion and seasonality impact
491 collectively due to the ecological niche range of microorganism (OMI value) increased.

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500

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