

Type of the Paper (Article)

Melissopalynology of Honey from Sudan: Implication for the Honey Identity and Misdesignation

Mohammed El-Nebir ¹, Mogbel El-Niweiri ^{2,3,4,*} and Seif Eldin Mohammed⁴

¹ Ministry of Agriculture, Forests and Irrigation, Forest Administration, Gadaref State, Sudan;

alnebir@gmail.com

² Department of Biology, King Khalid University, Abha, 61413, Saudi Arabia

³ Unit of Honeybees Research and Honey Production, King Khalid University, Abha, 61321, Saudi Arabia; mogbel7l@hotmail.com

⁴ Department of Bee Research, Environment and Natural Resources & Desertification Research Institute, P. O. Box 6096 – Khartoum, Sudan; seif_san@yahoo.com

* Correspondence: mogbel7@hotmail.com; Tel.:9665507594427

Abstract: Due to the great nutritional and medicinal value of honey, there has been growing consumer's preference towards honey of a known identity. However, honey now is the third food in the world subjected to adulteration. Therefore, the current study was focused on judging the identity of Sudanese honeys and checking whether there is any misdesignated from originality. Melissopalynology was used as a tool for this purpose. A number of 60 honey samples were purchased from honey sellers. Results indicated that honey bees foraged on a bio-diversified number of plant species constituted of 11 major families [Fabaceae (43.3%), being the predominant family] and 8 minor families. Respectively, 18.3% & 2% of the honey samples were found to be misdesignated by the honey sellers from their botanical and geographical identities. Some samples were predicted by melissopalynology to be originated from Ethiopia by the presence of marker pollens such as *Kniphofia foliosa*, *Guizotia abyssinica*, and *Acacia abyssinica* an indigenous Ethiopian flora. Thus these findings proved that melissopalynology is an effective tool in judging the identity of honey and pro of being inexpensive.

Keywords: melissopalynology; honey identity; forage biodiversity

1. Introduction

Honey is the natural sweet substance that produced by honey bees from the nectar of plants or from secretions of living parts of plants or excretions of plant-sucking insects on the living parts of plants which the bees collect, transform by combining with specific substances of their own, deposit, dehydrate, store and leave in the honeycomb to ripen. Honey sold as such shall not have added to it any food ingredient, including food additives, nor shall any other additions be made other than honey. No pollen or constituent particular to honey may be removed except where this is unavoidable in the removal of foreign inorganic or organic matter [1].

The adulteration of honey have increased in recent years because of the higher prices of honey compared to adulterants, and the obsolescence of official methods to detect instances of fraud. The current version of the phenomenon has a tremendous magnitude and impacts both the price of honey and the viability of the beekeeping industry [2].

According to the US Pharmacopeia's Food Fraud Database, honey is now the third "favorite" food target for adulteration, ranking only behind milk and olive oil [3]. The overall result of this dishonesty from beekeepers and honey sellers who run after short-term gains and low prices on the shelves is a big threat to food safety, food security, and ecological sustainability [4]. Different forms of honey adulteration can currently be found [5]: 1. Intentional dilution with cheap syrups (corn, rice, beet, etc.). 2. Extracting immature honey and dehumidifying it by mechanical means. 3. Use of ion exchange resins to remove residues and lighten honey color. 4. Feeding hives during a nectar flow 5. Masking the geographical and/or botanical origin of honey.

Honey is one of the best-known bee products that is in very much need to be produced properly and prepared according to high-quality parameters. Due to the growing consumer's preference towards honey of a known identity. The current study was focused on judging the identity of Sudanese honeys and checking whether there is any misdesignation from originality using melissopalynology as a tool for this purpose.

Palynological information is widely used in different disciplines. In the field of geographical and archaeological investigations, pollen fossils are used to determine vegetation and climatic features of an era in the past and oil deposits in the earth. In criminology, palynology has been used in forensic evidence in confirming the season and scene of a crime [6,7]. Also it is important in the study of pollen spectra of honeys "melissopalynology", since honey carries within itself its own certificate of identity [8,9].

Market situation of world honey indicates an increasing trend of consumer's preferences towards honey types that are originated from one or majorly one plant source (mono-floral or unifloral honey concepts). This trend could attribute to the therapeutic value of each honey type [10].

According to the Codex Alimentarius Standards and the European Standards Directive, the use of a botanical designation of honey is allowed if it originates predominately from the indicated floral source. Honey may also be designated by the name of a geographical region if it was produced strictly within the area referred to in the label [11,12].

Honey bees normally forage within a circular distance of approximately 3 km from their hives. Therefore, its characterization as an authentic and natural foodstuff can be achieved by studying its botanical and geographical origin[13]. This aspect is a requirement for the voluntary quality label of "Protected Designation of Origin" adopted by the European Union since 1992 [14].

It has been reported that identification of honey origin as well as the proof of its authenticity has become an important issue [15]. Therefore, the search for reliable chemical markers indicating its floral and/or geographical origin has been the focus of many studies". Among chemical markers reported are the volatile compounds [16], amino acids [17], and trace elements [18]. However, melissopalynology stands as "gold standard" method because it is cheap and informative. Thus the analysis of pollen in honey can give information regarding the region, season of production, patterns and variations of the local flora [19].

Higher prices are paid for certain types of honey, therefore, beekeepers and honey packers often misdesignate the botanical and geographical source of the honey[13]. In Europe, honey imported from China or Latin America has lower price than locally produced one. Even differences persist between European countries. In Saudi Arabia, honey produced locally is 10-20 folds higher priced than imported honey[20]. Thus there is a financial interest in misdesignation of the botanical and geographical source of honey[13].

In Sudan, there is consumer preference for honey from certain botanical origin as well as local honey that is produced in specific regions of the country. As for examples, consumers pay high price for Ziziphus and Acacia honey than other monofloral honeys for their ritual and therapeutic values. Also they are willing to purchase honey from Darfur region at higher price than honey produced in any other region [21]. This financial issue has led honey sellers for deliberately misdesignation of the botanical and geographical sources of honey. To address this issue we have performed a survey and melissopalynological investigation of the honey in Sudan. Aiming to prove and improve Sudanese honey identity and to check whether there is any misdesignation from originality.

94 **2. Materials and Methods**

95 Honey samples

96 Sixty honey samples were purchased from honey sellers of 10 regions in Sudan(Figure 1). Each
97 sample of honey is stirred thoroughly to assure even distribution of any botanical elements. Samples
98 were labeled (1-60) and stored under room temperature until used for the analysis. Primary data on
99 sample location and botanical source were recorded.

100 Pollen analysis

101 The analysis of pollen of honey was carried according to melissopalynology method which
102 was based on acetolysis technique [22,8]. Briefly, approximately 10 g of honey was weighted in a
103 pointed glass centrifuge tube capacity (ca. 50ml) and dissolved with with 30-35 ml of warm(20 –
104 40C°), weak sulfuric acid (3 ml of concentrated acid to 1000 ml distilled water). The solution was
105 centrifuged for 10 min and the supernatant was decanted. Distilled water (20 ml) was added to the
106 completely dissolve remaining sugar crystals and centrifuged for 5 minutes and the supernatant was
107 decanted. Taking sludge and placed in a Petri dish and stained with basic fuchsin and then distribute
108 on glass slides. Then dried with a series of alcohols 50%, 70%, and 100% where it was put a drop of
109 each focus on the slide and left for two minutes. One drop of glycerin jelly was applied to the cover
110 slip and the sample was examined through the microscope using (X 400).

111 Pollen taxa were identified using pollen reference slides deposited in the Department of Bee
112 Research, Environment and Natural Resources & Desertification Research Institute, Sudan. Other
113 references such as the study on the pollen flora of Sudan [23] , website of Australasian pollen and
114 Spore Atlas (APSA), [24] and the Atlas of pollen grains of major honey bee flora of Ethiopia [25]
115 were also used for pollen identification.

118 **3. Results**

119 3.1. Pollen analysis

120 Analysis of pollen of honey from Sudan was carried out. Data on the number and distribution
121 of the samples and sites of collection are given in Figure.1. Plant species constituting 11 families of
122 major pollen sources were identified. Another 8 families of minor pollen sources were also identified.
123 Table 1 shows sample frequencies as well as the major and minor pollen spectra of plant species
124 normally foraged by honey bees in Sudan. Families and frequencies of major plants observed were:
125 Fabaceae (43.3%), Rahmnaceae (21.6%), Boraginaceae (10%), Myrtaceae (8.3%), Meliaceae (3.3%),
126 Astraceae (3.3%), Gramineae (3.3%), Apiaceae (1.6%), Amaranthaceae (1.6%), Malvaceae (1.6%), and
127 Asphodelaceae (1.6%). On the other hand plant families like: Cucurbitaceae, Balanitaceae,
128 Combretaceae, Caesalpinaceae, Convolvulaceae, Asteraceae, Pedaliaceae, and Fabaceae represented
129 minor pollen sources in the investigated samples. Photomicrographs of some pollen morphs
130 identified are depicted in Figure 2.

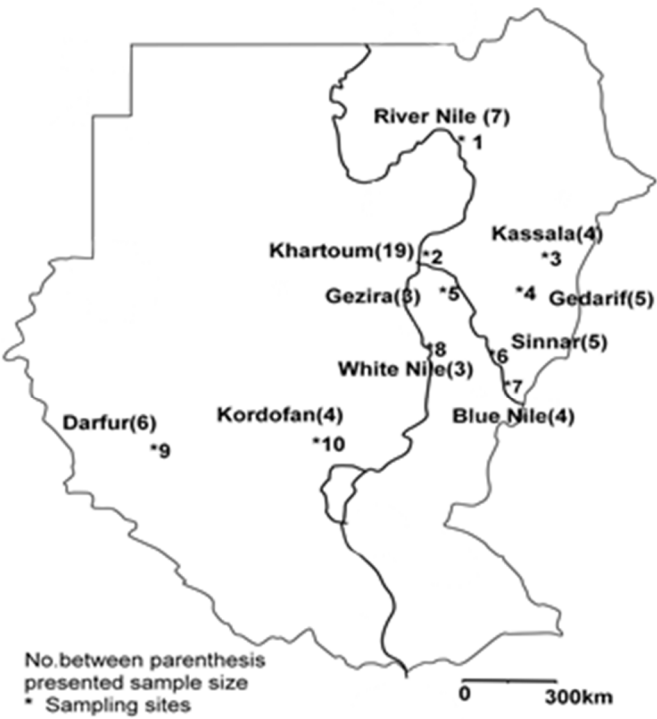


Figure 1. Map of honey samples distribution.

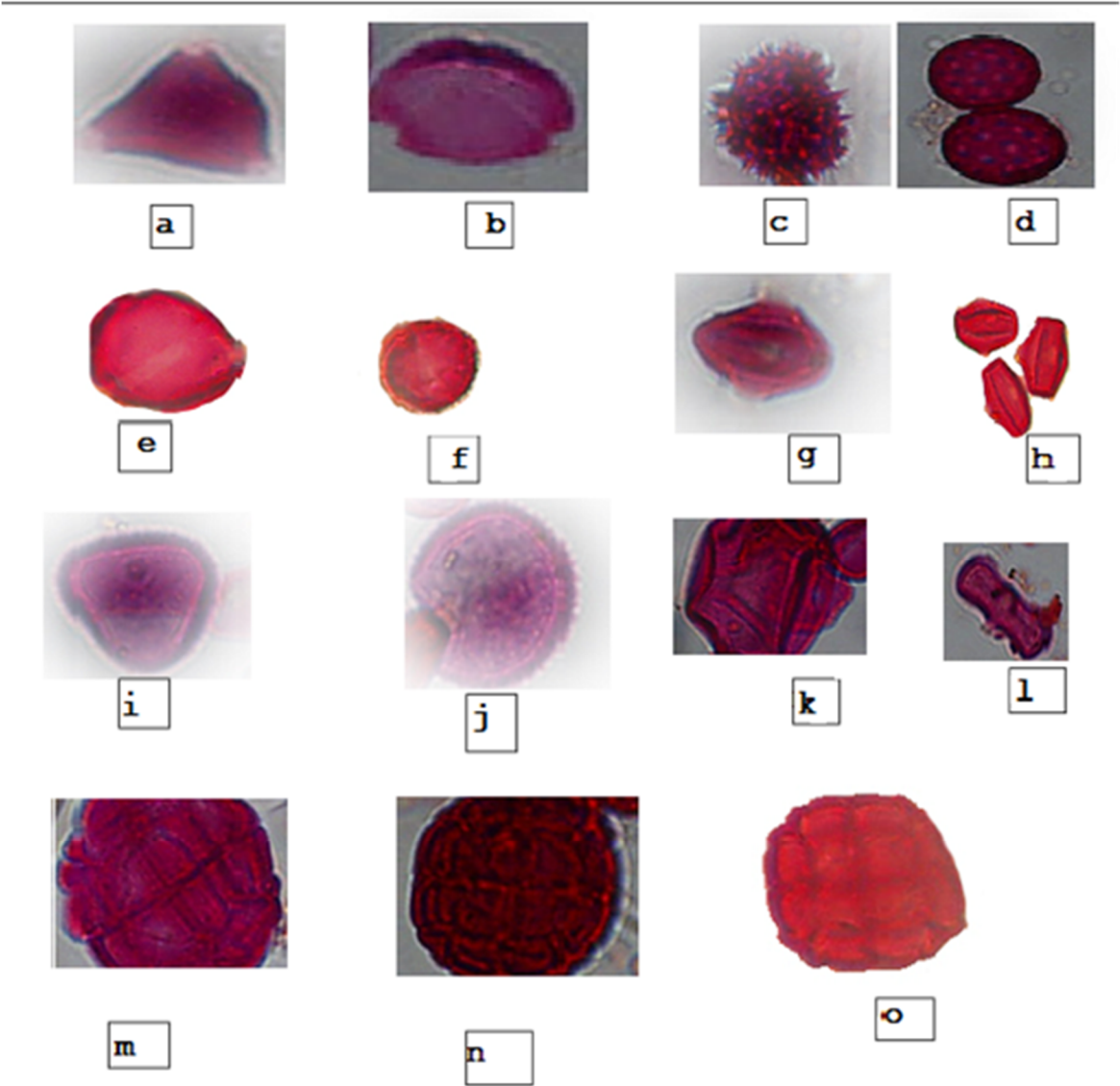


Figure 2: Photomicrographs of the identified pollen grain morphs: a - Myrtaceae (*Eucalyptus* spp.), b - Rhamnaceae (*Ziziphus* spp.) c- Astraceae (*Helianthus* spp.), d - Amaranthaceae (*Amaranthus* spp.), e- Asteraceae (*Guizotia abyssinica*), f- Pedaliaaceae (*Seasamum indicum*), g-h Anacardiaceae (*Kniphofia folios*), i-k Poaceae (*Gramineae* spp.) l- Apiaceae, (*Foeniculum* spp.) m-o Fabaceae (m- *Acacia nilotica*; n- *Faidherbia albida*; o- *Acacia abyssinica*).

Table 1. Pollen spectra of the studied honey samples.

| Families of major pollen sources | Sample frequency (%) | Plant species | Families of minor pollen sources | Plant species |
|----------------------------------|----------------------|------------------------------|----------------------------------|-----------------------------|
| Rhamnaceae | 21.6 | <i>Ziziphus</i> spp. | Cucurbitaceae | <i>Cucuribta</i> spp |
| Fabaceae | 43.3 | <i>Acacia nilotica</i> | Balanitaceae | <i>Balanites aegyptiaca</i> |
| Meliaceae | 3.3 | <i>Azadirachta indica</i> | Combretaceae | <i>Terminalia brownie</i> |
| Myrtaceae | 8.3 | <i>Eucalyptus</i> spp. | Caesalpinaceae | <i>Cassia</i> spp. |
| Astraceae | 3.3 | <i>Helianthus annulus</i> | Convolvulaceae | <i>Ipomoea</i> spp. |
| Apiaceae | 1.6 | <i>Foeniculum</i> spp. | Asteraceae | <i>Guizotia abyssinica</i> |
| Amaranthaceae | 1.6 | <i>Amaranthus</i> spp. | Fabaceae | <i>Faidherbia albida</i> |
| Boraginaceae | 10.0 | <i>Anogeissus leiocarpus</i> | Fabaceae | <i>Acacia abyssinica</i> |
| Gramineae | 3.3 | <i>Poaceae</i> spp. | Pedaliaceae | <i>Sesamum indicum</i> |
| Malvaceae | 1.6 | <i>Hibiscus</i> spp. | | |

| | | | | |
|---------------|-----|--------------------------|--|--|
| | | | | |
| Asphodelaceae | 1.6 | <i>Kinphofia foliosa</i> | | |

148 **Note:** the flora: *Kinphofia foliosa*, *Guizotia abyssinica*, and *Acacia abyssinica* are indigenous to Ethiopia and are not found in
149 Sudan.

150

151 3.2. *Misdesignaon of honey’s botanical and geographical identity*

152 The results in table 2 showed that about 2% and 18.3% of the collected honey samples were,
153 respectively geographically or botanically misdesignated by the honey sellers.

154 **Table 2.** Designaon of the honey samples (n=60) by the honey seller and by the palynological analysis.

| Code | designaon by | | | Code | designaon by | | |
|------|-------------------------|--------|-------------------------|------|----------------------|--------|----------------------|
| | seller | remark | palynology | | seller | remark | palynology |
| 1 | <i>Ziziphus</i> spp. | ✓ | <i>Ziziphus</i> sps. | 31 | <i>Ziziphus</i> spp. | ✓ | <i>Ziziphus</i> spp. |
| 2 | <i>A. seyal</i> | ✓ | <i>A. seyal</i> | 32 | <i>Ziziphus</i> spp. | ✓ | <i>Ziziphus</i> spp. |
| 3 | <i>A. nilotica</i> | ✓ | <i>A. nilotica</i> | 33 | <i>Ziziphus</i> spp. | ✓ | <i>Ziziphus</i> spp. |
| 4 | <i>A. seyal</i> | ✓ | <i>A. seyal</i> | 34 | <i>Ziziphus</i> spp. | ✓ | <i>Ziziphus</i> spp. |
| 5 | <i>Ziziphus</i> spp. | ✓ | <i>Ziziphus</i> spp. | 35 | <i>Ziziphus</i> spp. | ✓ | <i>Ziziphus</i> spp. |
| 6 | <i>A. nilotica</i> | ✓ | <i>A. nilotica</i> | 36 | <i>Ziziphus</i> spp. | ✓ | <i>Ziziphus</i> spp. |
| 7 | <i>A. seyal</i> | ✓ | <i>A. seyal</i> | 37 | <i>Ziziphus</i> spp. | ✓ | <i>Ziziphus</i> spp. |
| 8 | <i>Ziziphus</i> spp. | ✓ | <i>Ziziphus</i> spp. | 38 | <i>Ziziphus</i> spp. | ✓ | <i>Ziziphus</i> spp. |
| 9 | <i>A. nilotica</i> | ✓ | <i>A. nilotica</i> | 39 | <i>Ziziphus</i> spp. | ✓ | <i>Ziziphus</i> spp. |
| 10 | <i>Azadirachta</i> spp. | ✓ | <i>Azadirachta</i> spp. | 40 | <i>Ziziphus</i> spp. | ✓ | <i>Ziziphus</i> spp. |

| | | | | | | | |
|----|--|-----|------------------------------|----|--------------------------|---|--------------------------|
| 11 | NI | | <i>Eucalyptus</i> spp. | 41 | NI | | <i>Cordia</i> spp. |
| 12 | <i>Ziziphus</i> spp. | ✓ | <i>Ziziphus</i> spp. | 42 | NI | | <i>Gramineae</i> |
| 13 | <i>Acacia seyal</i> | ✓ | <i>Acacia seyal</i> | 43 | NI | | <i>Cordia</i> spp. |
| 14 | <i>Acacia nilotica</i> | ✓ | <i>Acacia nilotica</i> | 44 | <i>Ziziphus</i> spp. | ✓ | <i>Ziziphus</i> spp. |
| 15 | <i>Acacia seyal</i> | ✓ ✓ | <i>Acacia seyal</i> | 45 | NI | | <i>Acacia nilotica</i> |
| 16 | <i>Eucalyptus</i> spp. | ✓ | <i>Eucalyptus</i> spp. | 46 | NI | | <i>Acacia seyal</i> |
| 17 | <i>Helianthus annuus</i> | ✓ | <i>Helianthus annuus</i> | 47 | NI | | <i>Acacia nilotica</i> |
| 18 | <i>Acacia nilotica</i> | ✓ | <i>Acacia nilotica</i> | 48 | NI | | <i>Acacia seyal</i> |
| 19 | <i>Ziziphus</i> spp. | ✓ | <i>Ziziphus</i> spp. | 49 | <i>Acacia seyal</i> | ✓ | <i>Acacia seyal</i> |
| 20 | NI | | <i>Kinphofia foliosa</i> | 50 | <i>Acacia nilotica</i> | ✓ | <i>Acacia nilotica</i> |
| 21 | <i>Eucalyptus</i> spp. | ✓ | <i>Eucalyptus</i> spp. | 51 | <i>Ziziphus</i> spp. | ✓ | <i>Ziziphus</i> spp. |
| 22 | <i>Eucalyptus</i> spp. | ✓ | <i>Eucalyptus</i> sp. | 52 | <i>Acacia seyal</i> | ✓ | <i>Acacia seyal</i> |
| 23 | <i>Eucalyptus</i> spp. | ✓ | <i>Eucalyptus</i> spp. | 53 | honeydew | ✓ | honeydew |
| 24 | NI | | <i>Azadirachta</i> spp | 54 | honeydew | ✓ | honeydew |
| 25 | <i>Ziziphus</i> spp. | ☒ | <i>Amaranthus</i> spp. | 55 | <i>Helianthus annuus</i> | ✓ | <i>Helianthus annuus</i> |
| 26 | <i>Acacia seyal</i> | ✓ | <i>Acacia seyal</i> | 56 | <i>Acacia seyal</i> | ✓ | <i>Acacia seyal</i> |
| 27 | <i>Acacia seyal</i> | ✓ | <i>Acacia seyal</i> | 57 | <i>Acacia nilotica</i> | ✓ | <i>Acacia nilotica</i> |
| 28 | <i>Acacia seyal</i> | ✓ | <i>Acacia seyal</i> | 58 | <i>Ziziphus</i> spp. | ✓ | <i>Ziziphus</i> spp. |
| 29 | <i>Anogeissus</i> <i>leiocarpus</i> | ✓ | <i>Anogeissus leiocarpus</i> | 59 | <i>Ziziphus</i> spp. | ✓ | <i>Ziziphus</i> spp. |
| 30 | NI | ✓ | <i>Acacia seyal</i> | 60 | <i>Eucalyptus</i> spp. | ✓ | <i>Eucalyptus</i> spp. |

Key: Ni Not identified, ✓ the honey identity is correctly designated by the sellers, and ☒ the honey identity is incorrectly designated by the honey sellers.

4. Discussion

Our study demonstrated the photomicrographs and misdesignation of honey’s botanical and geographical identity. The obtained result indicated predominance of pollen from forest trees (80%). This finding necessitated raising awareness about the value of forests biodiversity. Hence encourages beekeepers in conscious protection, conservation and sustainable management of forest resources. Moreover, in the marketing of honey it is very important to label the true floral sources of honey in which mellisopalynology is useful tool in determining monofloral or multifloral honeys, dominant and predominant sources of honey “pollen frequencies” and detecting artificial and honeydew honeys “honey adulteration” [26].

The pollen photomicrographs depicted in figure 2 corresponded to some pollen morphs previously reported by other authors who have conducted mellisopalynological analysis of Sudanese honeys as well as honeys from other regions [27, 28, 29].

Bogdanov and Martin [13] reported that in food control, pollen analysis is very efficient for the differentiation of honeys produced in distinctly different geographical and climatic areas. In the present study, sample coded number 20, which designated as from Sudan in table 2, after mellisopalynological analysis this sample was proved to be originated from Ethiopia. Its pollen spectra revealed presence of pollens of *Guizotia abyssinica*, *Kniphofia foliosa*, and *Acacia abyssinica* an indigenous flora of Ethiopia (figure 2e, g-h, and o). The pollen morphs of these exotic flora observed in this study, were simulated by different reports of Ethiopian honeys [25, 30]. None of the three trees is reported to be grown in the Sudan. It is also necessary to note that some of the samples were purchased from Gadaref district of Sudan, which borders with Ethiopia. People from Gadaref State regularly exchange goods with traders from the borders of Ethiopia. Therefore, this sample might have originated in Ethiopia and passed into Sudan through such trade.

Sample coded 25 (table 2), was sold by the sellers as ziziphus honey but after mellisopalynological investigation it was proved to be *Amaranthus* honey (figure 2d). It is normal phenomenon in Sudan that honey sellers allege that their honey is from ziziphus tree even though if it was from any else plant source to gain high price.

Also some honey samples were sold by the honey sellers as from anonymous botanical sources. However, after mellisopalynological analysis, their corresponding botanical sources were identified e. g samples coded 11, 20, 24, 30, 41, 42, 43, 45, 46, 47, 48 (table 2) were respectively *Eucalyptus* spp., *Kniphofia foliosa*, *Azadirachta* spp., *Acacia seyal*, *Cordia* spp., *Gramineae*, *Cordia* spp., *Acacia nilotica*, *Acacia seyal*, *Acacia nilotica*, *Acacia seyal* (figure 2). Thus mellisopalynology is informative technique. It was reported, “honey is a commodity that is closely linked to a territory” [31]. Thus pollen analysis of honey is a powerful tool in detecting honey identity.

5. Conclusions

From the results obtained here by pollen analysis of honey “mellisopalynology” it can be concluded that palynology is informative for: (a) botanical and geographical identity of honey [18.3% and 2% of the studied samples showed geographical and botanical misdesignation, respectively], (b) honeybees forage biodiversity, and (c) requires inexpensive laboratory assets.

Author Contributions: All authors contributed to the study conception and design. Material preparation, data collection and analysis were performed by [Mohammed A. El-Nebir], [Mogbel Ahmed Abdalla El Niweiri] and [Seif Eldin A. Mohammed]. The first draft of the manuscript was written by [[Mohammed A. El-Nebir] and all authors commented on previous versions of the manuscript. All authors read and approved the final manuscript.

203

204 **Funding:** The present investigation was partly supported by a King Khalid University
205 Research Science Center for Advanced Materials

206 **Acknowledgments:** The authors thank the Department of Bee Research, Environment and Natural Resources &
207 Desertification Research Institute for their research facilities.

208 **Conflicts of Interest:** The authors declare no conflict of interest

209

210 References

- 211 1. Codex Alimentarius. Codex standard for honey. Codex Alimentarius **1981**, 12, 1–8.
- 212 2. García, N. A study of the causes of falling honey prices in the international market. *Am. Bee J.* **2016**, 156,
- 213 877–882.
- 214 3. United States Pharmacopeia. Food fraud database. 2018, Retrieved from: <http://www.foodfraud.org>
- 215 4. García, N. The Current Situation on the International Honey Market. *Bee World* **2018**, 2376-7618:
- 216 <https://doi.org/10.1080/0005772X.2018.1483814>
- 217 5. Dübecke, A. ; van der Meulen, J. ; Schütz, B. ; Tanner, D., Beckh, G. ; Lüllmann, C. NMR profiling a defense
- 218 against honey adulteration. **2018**, 158, 83–86.
- 219 6. Faegri, K.; Iversen, J. *Text book of pollen analysis*. John Wiley & Sons, New York, 1989
- 220 7. Barth, O.M. ; Luz, C.F.P. Mellisopalynological data from Mangrove area Rio de Janeiro, Brazil. *J. Apic. Res.*
- 221 **1998**, 37 (3), 155-163.
- 222 8. Louveaux, J. ; Maurizio, A. ; Vorwohl, G. Methods of mellisopalynology. *Bee World* 1978, 59 (4), 139-157
- 223 9. Vit, P. ; Ricciardelli D. ; Albore, G. Mellisopalynology for stingless bees (Apidae: Meliponinae) from
- 224 Venezuela. *J. Apic. Res.* **1994**, 33 (3), 145-154.
- 225 10. Waghkhle, D.M.; Nair, K.S.; Ramesh, B. Sugar composition in nectars of certain plants. *Indian Bee J.* **1981**, 43
- 226 (1), 6-8.
- 227 11. Codex . Codex Alimentarius Committee on Sugars . Codex standard 12 revised Codex Standard for Honey.
- 228 Standards and Standard Methods 2001, pp 11, 1–7.
- 229 12. Cordella, C.; Faucon, J.P.; Cabrol-Bass, D.; Sbirrazzouli, N. Application of DSC as a tool for honey floral
- 230 species characterization and adulteration detection. *J. Therm. Anal. Calorim* **2003**, 71 (1), 279-290.
- 231 13. Bogdanov, S.; Martin, P. Honey authenticity: A review. *Swiss Bee Research Centre* 2002, 1-20.
- 232 14. Serrano, S.; Jimenez-Hornero, F.J.; Gutierrez de Rave, E.; Jordal, M.L. GIS design application for “Sierra
- 233 Morena honey” designation of origin. *Comput Electron Agric* **2008**, 64 (2), 307-317.
- 234 15. Kaskoniene, V.; Vnskutonis, P.R. Floral markers in honey of various botanical and geographic origins: A
- 235 review. *Compr. Rev. Food Sci. Food Saf.* **2010**, 9 (6), 620-634.
- 236 16. Cotte, J.; Casablanca, H.; Chardon, J.; Lheritier, J.; Grenier-Loustalof, M.F. Application of carbohydrate
- 237 analysis to verify honey authenticity. *J. Chromatogr. A* **2003**, A 1021, 145-155.
- 238 17. Mohammed, S.A.; Babiker, E.E. Identification of the floral origin of honey by amino acid composition. *Aust.*
- 239 *J. Basic Appl. Sci.* **2010**, 4 (4), 552-556.
- 240 18. Tuzen, M.; Silici, S.; Mendil, D.; Soylak, M. Trace element levels in honey from different regions of
- 241 Turkey. *Food Chem.* **2007**, 103 (2), 325-330.
- 242 19. Barth, O.M. mellisopalynology in Brazil: a review of pollen analysis of honeys, propolis and pollen loads of
- 243 bees. *Sci. Agric.* **2004**, 61 (3), 342-50.
- 244 20. Al-Ghamdi, A.; Adgaba, N. Beekeeping in the Kingdom of Saudi Arabia: Opportunities and challenges. *Bee*
- 245 *world* **2013**, 90 (3), 54-57.
- 246 21. Elzaki, E.E.A.; Mohammed, S.A. Household’s honey consumption and preference due to honey’s Criteria
- 247 in Khartoum State of Sudan. APIEXPO AFRICA 2012-Addis Ababa-Ethiopia. September 26th – 30th, 2012.
- 248 <http://www.apitradeafrica.org/apiexpo-africa-2012.html>.
- 249 22. Erdtman, G. Hand book of palynology. Munksgaard. Copenhagen, Denmark 1969.
- 250 23. El- Ghazali, G.E.B. A study on the pollen flora of Sudan, with special reference to pollen identification.
- 251 1989, Ph. D. Thesis, University of Bergen Botanical Institute.
- 252 24. David, W. ; Roubik, J. ; Enrique Moreno, P. (1991). Pollen and spores of Barro Colorado Island, Published
- 253 1991 by Missouri Botanical Garden in [St. Louis] <http://www>

25. Adgoba, N. Atlas of pollen grains of major honeybee flora of Ethiopia, ed. 17 Holetta Genet, West Shoa, Ethiopia 2007, 152.
26. Aira, M.J.; Horn, H.; Seijo, M. Palynological analysis of honey from Portugal. *J. Apic. Res.* **1998**, 37 (4), 247-254.
27. Mohammed, S.A.; Kabbashi, A.S.; Koko, W.S.; Ansari, M.J.; Adgaba, N.; Al-Ghamdi, A. In vitro activity of some natural honeys against *Entamoeba histolytica* and *Giardia labilia* trophozoites. *Saudi J. Biol. Sci.* **2019**, **26** (2), 238-243.
28. Sajwani, A.; Farooq, A.S.; Patzelt, A.; Eltayeb, E.A; Dryant, V.M. Melissoplainological studies from Oman. *Palynology* **2007**, 31 (1), 63-79.
29. Santos de Novais, J.; Absy, M.L. Palynological examination of the pollen pots of native stingless bees from the lower Amazon region in Para Brazil. *Palynology* **2013**, 37(2), 218-230.
30. Adgaba, N.; Al-Ghamdi, A.; Getachew, A.; Tadesse, Y.; Belay, A.; Ansari, J.M.; Radlof, E.S.; Sharma, D. Characterization of honeys by their botanical and geographical origins based on physico-chemical properties and chemo-metrics analysis. *Food Measure* **2017**, 11, 1106-1117.
31. Bernardinelli, I.; Della Vedova, G.; Milani, N. Unapprocciometodologico per la gestione del nomadismo in apicoltura mediantei GIS: Esperienza nelle Valli del Natisone (Friuli Venecia Giulia). Ati dell' Incontro-Seminario Mappatura delle aree nettariifere. 2005, Firenze