

1 **Practical implications of different phenotypic and molecular responses of evergreen**
2 **conifer and broadleaf deciduous forest tree species to regulated water deficit in a**
3 **container nursery**

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5 ^{1*}Piotr Robakowski, ²Tomasz Wyka, ¹Wojciech Kowalkowski, ¹Władysław Barzdajn, ³Emilia
6 Pers-Kamczyc, ^{2,3}Artur Jankowski, ⁴Barbara Politycka

7

8

9 ^{1*}Piotr Robakowski, Poznan University of Life Sciences, Faculty of Forestry, Wojska
10 Polskiego 71E, PL 60-625 Poznań, Poland, pierrot@up.poznan.pl, ORCID: 0000-0001-5564-
11 7360

12 ²Tomasz Wyka, Adam Mickiewicz University, Faculty of Biology, General Botany
13 Laboratory, Umultowska 89, PL 61-614 Poznań, twyka@amu.edu.pl; ORCID: 0000-0003-
14 0569-8009

15 ¹Wojciech Kowalkowski, Poznan University of Life Sciences, Faculty of Forestry, Wojska
16 Polskiego 71E, PL 60-625 Poznań, Poland, wojciech.kowalkowski@up.poznan.pl, ORCID:
17 0000-0002-5801-9818

18 ¹Władysław Barzdajn, Poznan University of Life Sciences, Faculty of Forestry, Wojska
19 Polskiego 71E, PL 60-625 Poznań, Poland, barzdajn@up.poznan.pl, ORCID: 0000-0002-
20 0018-2057

21 ³Emilia Pers-Kamczyc, Institute of Dendrology, Polish Academy of Sciences, Parkowa 5, PL
22 62-035 Kórnik, Poland, epk@man.poznan.pl, ORCID: 0000-0002-5610-2124

23 ^{2,3}Artur Jankowski, Adam Mickiewicz University, Faculty of Biology, General Botany
24 Laboratory, Umultowska 89, PL 61-614 Poznań; Institute of Dendrology, Polish Academy of
25 Sciences, Parkowa 5, PL 62-035 Kórnik, Poland, artur.jankowski@amu.edu.pl, ORCID:
26 0000-0001-6765-8008

27 ⁴Poznan University of Life Sciences, Faculty of Horticulture and Landscape Architecture,
28 Department of Plant Physiology, Wołyńska 35, PL 60-625 Poznań, Poland,
29 barbara.politycka@up.poznan.pl, ORCID: 0000-0002-5078-6596

30

31 * Author for correspondence, e-mail: piotr.robakowski@up.poznan.pl, phone: +48 61 848 77

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35 Abstract

36 Recent climatic changes have resulted in an increased frequency and prolonged periods of
37 drought and strained water resources affecting plant production. We explored the possibility of
38 reducing irrigation in a container nursery and studied the growth response of seedlings of
39 economically important forest trees: broadleaf deciduous angiosperms *Fagus sylvatica*,
40 *Quercus petraea* and evergreen conifers *Abies alba* and *Pinus sylvestris*. We also studied
41 markers of water stress including modifications of biomass allocation, leaf anatomy, proline
42 accumulation and expression of selected genes. Growth of the broadleaved deciduous species
43 was more sensitive to the reduced water supply than that of conifers. Remarkably, growth of
44 the shade tolerant *Abies* was not affected. Adjustment of biomass allocations was strongest in
45 *P. sylvestris*, with a remarkable increase in allocation to roots. In response to water deficit both
46 deciduous species accumulated proline in leaves and produced leaves with shorter palisade cells,
47 reduced vascular tissues and smaller conduit diameters, but not conifers. Relative transcript
48 abundance of a gene encoding a Zn-finger protein in *Q. petraea* and a gene encoding a pore
49 calcium channel protein 1 in *A. alba* increased as water deficit increased. These findings suggest
50 that in container nursery, the genetic selection can be initiated by water deficit. Our study shows
51 major differences between functional groups in response to irrigation, with seedlings of
52 evergreen conifers having higher tolerance than the deciduous species. This suggests that major
53 water savings could be achieved by adjusting irrigation regime to functional group or species
54 requirements.

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56 Key words: biomass allocation; drought; irrigation; leaf anatomy; mRNA level; proline

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58 **Acknowledgments:** This study was supported by The State Forests National Forest Holding.
59 We thank M. Wisniewski for help in maintaining of the experiment. The publication is co-
60 financed within the framework of Ministry of Science and Higher Education programme as
61 "Regional Initiative Excellence" in years 2019-2022, project number 005/RID/2018/19

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68 **Introduction**

69 According to predictions, global warming will lead to an increased frequency and extent of
70 summer drought periods in many regions of the Earth [1]. Although in northern and western
71 European forests global warming is expected to stimulate forest growth, the accompanying
72 drought may cause adverse effects [2]. This is because climate warming will increase
73 evapotranspiration more than precipitation and periodic drought events will enhance effects of
74 increased temperature on growth and photosynthesis of boreal and temperate tree species [3,4].
75 The understanding of drought effects on trees is thus vital for proper management and
76 conservation of forest ecosystems.

77 Water shortages affect all biological functions of trees, with the risk of drought-induced
78 mortality depending on the local geographical conditions, precipitation, soil properties and
79 anthropogenic factors [5]. Vulnerabilities of individual species vary widely [6,7,8,9]. Much
80 knowledge of the variability of tree responses to drought has been gathered from studies of
81 mature stands [10,11,12], however, the most vulnerable stage in the tree's ontogeny is the
82 seedling. In particular, it is not certain to what extent the responses and vulnerabilities of adult
83 trees vs. seedlings to drought are consistent in between-species comparisons [13,14].

84 Trees acclimate to restriction in water supply through modifications of growth, biomass
85 allocation, organ structure, and through functional adjustments encompassing processes from
86 gene expression to enzymatic activity and metabolic regulation [15,7,16,17]. These adaptive
87 changes may lead to improved plant survival through an increased water uptake and a more
88 conservative water use [18]. Water deficit typically leads to an increase in relative biomass
89 allocation to roots, a decrease in allocation to leaves and reduced leaf area ratio and leaf mass
90 fraction, resulting in greater water uptake potential and reduced transpiration [19,20,21].
91 Individual leaves produced under the influence of drought are usually smaller and thicker than
92 those from well hydrated plants [22,23,24] and show reduced amount of vascular tissues and
93 smaller diameters of conductive elements [25,26,27]. Plants subjected to drought may
94 accumulate metabolites responsible for osmotic adjustment and oxidative stress protection as
95 well as change hormone and regulatory molecule concentrations [28,29,30]. Profiles of gene
96 expression and enzymatic as well as transport activities may become greatly altered [31,32].
97 Adjustments occur at different time scales, but ultimately may result in drought resistant or
98 drought tolerant phenotypes. The ability to produce such plastic modification may be the key
99 to plant survival under natural settings. However, traits that enhance drought survival,
100 especially those related to water conservation, frequently trade off with growth potential [33].

101 For example, increased allocation to roots may lead to a decreased ratio of assimilatory to non-
102 assimilatory biomass, restricting whole-plant carbon supply even after the cessation of drought
103 [34]. Moreover, long term carry-over effects (so called drought memory) may also result from
104 metabolic adjustment to drought [35,36].

105 Water requirements and responses to drought differ significantly among taxa and
106 functional groups. For major forest tree species the grouping into drought tolerance categories
107 or rankings has been based on characteristics of their habitat and responses of growth and
108 physiological indices to environmental drought [6,7]. In general, tree species adapted to drier
109 environments are more tolerant to drought compared with those from humid habitats [37,38].
110 At the functional group level, evergreen conifers are more tolerant to drought than deciduous
111 broadleaved tree species [6] and late successional, shade tolerant tree species are often more
112 sensitive to water deficit than pioneer and shade intolerant species [39]. Many insights have
113 been provided by physiological studies, especially those focusing on vulnerability to cavitation
114 and responses of stomatal conductance [7,40]. Such results may provide general information
115 on species requirements and drought vulnerability ranking, yet, since they focus on saplings or
116 adult trees, their relevance for seedling stages may be limited.

117 The species selected for this study were two deciduous broadleaf angiosperms (*Fagus*
118 *sylvatica* L. and *Quercus petraea* (Matt.) Liebl. and two evergreen conifers (*Abies alba* Mill.
119 and *Pinus sylvestris* L.). They differ in ecological requirements with respect to moisture and
120 their overall sensitivity to water deficit has been investigated under various settings. Briefly,
121 *Fagus sylvatica* is a dominant European forest tree species of high economic and ecological
122 importance. It is a shade-tolerant, late successional and the most water-demanding tree among
123 our selected species [41,7]. It requires high air humidity and grows preferentially on fertile,
124 well-drained and moist, but not waterlogged soils. *Quercus petraea* is a moderately shade
125 tolerant, late successional tree. It maintains higher leaf conductance [42] and higher net CO₂
126 assimilation rates than *F. sylvatica* under the same experimental conditions [43,44]. *Abies alba*
127 is a strongly shade tolerant, late successional and slow-growing tree [45]. It is considered rather
128 sensitive to drought [46,47]. *Pinus sylvestris* is the most widely distributed Eurasian conifer. It
129 is a pioneer, early-successional, light-demanding and drought-tolerant tree. Severe drought,
130 however, has a limiting effect on *P. sylvestris* growth and may triggered Scots pine decline [48].
131 It may be expected that species' growth sensitivity to low water supply should follow ecological
132 species ranking [41]. Our study species can be classified according to their decreasing drought
133 tolerance in the following order: *P. sylvestris* > *A. alba* > *Q. petraea* > *F. sylvatica*. However,

134 this ranking cannot be generalized to seedlings and to the whole geographical ranges of these
135 species. In addition, it is not clear whether seedlings of our study species growing in a container
136 nursery will show similar drought tolerance as in natural conditions.

137 Raising seedlings under polyethylene tunnel has become an important method of
138 containerized forest tree production [49]. Usually, seedlings are grown in containers placed
139 under an unheated tunnel. Under Central European conditions, seed planting takes place in May
140 and seedlings may be grown till the end of July, August or September, depending on the species,
141 and then the seedlings are transferred to an outdoor nursery. Nutrients are provided with water
142 used for irrigation or as slow-released fertilizers added to the substrate. Irrigation in container
143 forest nurseries is usually carried out by programmable automatic systems delivering water
144 according to established norms [50]. The rising cost of water provides incentive to reduce water
145 consumption, however decreasing irrigation doses below physiological optimum may result in
146 water stress and decrease the development rate of seedlings. On the other hand, reduced
147 substrate water content may improve substrate aeration which is beneficial for growth and the
148 quality of seedlings [51] and reduce nutrient leaching from the substrate [52]. Lower substrate
149 water content, especially at the end of the growing season, accelerates tissue maturation and
150 bud set, resulting in improved frost tolerance [53]. These additional benefits of reduced
151 irrigation thus occur in addition to water savings.

152 In this study we evaluated the effects of reduced irrigation on growth and on several
153 morphological, anatomical, metabolic and molecular variables that are known to predictably
154 respond to water deficit. We studied whole plant allocation of biomass among major organ
155 types (leaves, roots and stems), examined some key anatomical dimensions of foliar cross-
156 section, and determined the concentration of proline – a compound involved in the process of
157 osmotic adjustment [54,55]. Additionally, we studied the expression of genes known to respond
158 to water deficit in our focal species. Although the molecular basis of phenotypic responses of
159 forest trees to water deficit is not well understood, and the identification of water deficit related
160 genes in trees, especially in conifers, has been difficult, partly because of their large genomes
161 [56], sets of such reference drought-responsive genes have been identified first in *Arabidopsis*
162 and some crops [57], and later in other plants, including trees [58,59]. These genes may be
163 considered as indicators of drought stress [60].

164 We tested the hypotheses that under decreased irrigation water supply: (1) seedling
165 growth will be reduced and the reduction of biomass accumulation will be relatively larger in
166 broadleaved deciduous and shade-tolerant species than in conifers and light-demanding species;

167 (2) allocation to roots will increase and allocation to leaves will decrease; (3) leaves will show
168 xeromorphic anatomical adjustments and (4) proline accumulation will be enhanced; (5)
169 relative levels of mRNA of stress-related genes in leaves will be elevated. The additional,
170 applied facet of our study was to test whether, and in which species, irrigation doses can be
171 reduced below the established norm without compromising growth and affecting morphological,
172 metabolic and molecular characteristics of the seedlings.

173

174 **Material and methods**

175 Study site

176 The experiment was conducted in the Rogoziniec forest nursery (N52°18' E15°46') located
177 in Babimost Forest Division, SW Poland. The experiment was set up in an unheated
178 polyethylene tunnel (8 x 50 m).

179

180 Plant material

181 *Fagus sylvatica* seeds and *Quercus petraea* acorns originated from the seed stands located
182 close to Swiebodzin (52°15' N, 15°32' E), *Pinus sylvestris* seeds were from the local
183 'Rogoziniec' provenance (52°10' N, 15°50' E) and *Abies alba* seeds originated from the
184 Sudety Foothills (51°7' N, 15°55' E), Poland.

185

186 Experimental design

187 All seeds were subjected to stratification. At the end of May 2015 they were sown into
188 individual compartments of 54-compartment polystyrene containers (Robin, France). Each
189 compartment was filled with 430 ml of substrate consisting of peat and perlite (3:1 v/v, pH =
190 6.5) with the addition of 2 g of the 'Osmocote Exact Standard' slow-release fertilizer
191 (N:P:K:Mg 15:9:12:2, with micronutrients). During a ten-day germination period containers
192 were watered every day up to the field capacity. Containers were placed on benches 0.30 m
193 above the ground and arranged in a full-factorial completely randomized block design with
194 species and irrigation levels as the fixed factors and blocks as the random factor. In our
195 experiment, the irrigation norms recommended by Pierzgałski et al. [61] for forest nurseries
196 in Poland were used as the benchmark. In these guidelines, the species of trees are classified
197 into one of three groups: (1) conifers including *Pinus sylvestris* and *Abies alba* with one
198 deciduous species *Tilia cordata*, (2) *Quercus*, *Fagus sylvatica* and *Tilia platyphyllos*, (3) other
199 broadleaved deciduous species. From April to June, according to these norms, at the initial

200 phase of growth, seedlings are irrigated every two days with 5 mm for *P. sylvestris* and *A.*
201 *alba* and with 7 mm every three days for the broadleaved deciduous species. In our
202 experiment, we had regard for the norms and nurseryman' experience. Four daily irrigation
203 levels were established: 6 mm (6 l/m², 100%), 4.5 mm (4.5 l/m², 75%), 3 mm (3 l/m², 50%)
204 and 1.5 mm (1.5 l/m², 25% of the maximal dose). When recalculated per area of one cavity of
205 container, 12 (100%), 9 (75%), 6 (50%) or 3 ml (25%) of water were delivered every day.
206 After leaf expansion, water amount reaching the substrate in cavity was reduced when
207 compared to these values due to interception. To compare species-specific responses to the
208 irrigation doses, we applied the same irrigation regime for each species. Each irrigation
209 regime was replicated four times with four containers per species and irrigation treatment per
210 block (4 blocks x 4 species x 4 treatments x 4 containers x 54 cavities). At the end of the
211 experiment, the total number of seedlings was 8008. Seedlings were randomly selected, and
212 the numbers of replicates per each analysis (*n*) are given in the Results. Additional containers
213 with seedlings of Scots pine were placed around the experimental blocks to eliminate the edge
214 effect.

215

216 Microclimate

217 Air temperature and relative humidity (*RH*) in the tunnel were monitored with two HOBO Pro v2
218 probes (*OnSet Computers*, Pocasset, MA, USA). The probes were attached at the height of 90 cm
219 above the ground (close to the apical parts of seedlings) and logged data every twenty min.
220 During the active growing phase from mid-June to the beginning of August mean air
221 temperature ($T_{mean} \pm SD$, where SD is standard deviation) was 23.4 ± 7.4 °C and the average
222 relative humidity ($RH_{mean} \pm SD$) was 61 ± 22 %. In July $T_{mean} = 20.7 \pm 6.0$ °C, maximal
223 temperature (T_{max}) was 38.8 °C, minimal temperature (T_{min}) was 10.8 °C, and $RH_{mean} =$
224 65 ± 19 %, $RH_{max} = 96$ %, and $RH_{min} = 28$ %. In August $T_{mean} = 24.2 \pm 7.6$ °C, $T_{max} = 42.5$ °C,
225 $T_{min} = 7.1$ °C, and $RH_{mean} = 60 \pm 23$ %, $RH_{max} = 96$ %, and $RH_{min} = 19$ %. Extremely high
226 temperatures exceeding 40 °C occurred on 6 and 8 August between 2:00 and 4:30 p.m.

227

228 Regulated water deficit

229 The terms water deficit and drought have often been considered synonyms although water
230 deficit is more commonly used when referring to water availability below field capacity,
231 especially in relation to crop cultivation whereas drought is generally related to low
232 precipitation over a period of time. In our study, seedlings were cultivated under controlled

233 conditions in an unheated tunnel, therefore we use the term ‘water deficit’ for irrigation levels
234 below the established norm.

235 Plants were watered by an overhead sprinkler system. The different irrigation levels
236 were obtained by regulating the duration of water delivery over particular blocks. The water
237 pressure (0.25 MPa in each sprinkler) and irrigation time were monitored using an automated
238 system (Rathmakers, Germany). A pluviometer installed below the overhead sprinklers were
239 used to measure water amount in millimeters in each irrigation treatment.

240

241 Growth, biomass allocation and water contents

242 The height and root collar diameter of seedling were measured after 60 days of growth under
243 experimental conditions ($n = 10$, n – number of seedlings per block, species and per
244 treatment). Seedlings were subsequently removed from containers and their roots were gently
245 washed ($n = 4$). Plants were dissected into roots, stem and leaves and fresh mass (FM) of each
246 fraction was determined. Biomass fractions were then dried at 65°C for 7 days until the weight
247 was unchanged using a climate cabinet (Pol-Eko, Poland), and weighed for dry mass
248 determination. Water amount in each type of organs was calculated by subtracting organ dry
249 mass from its fresh mass and water content per unit of dry mass was obtained by dividing the
250 water amount by dry mass of the organ fraction or by total seedling dry mass.

251

252 Microscopic observations

253 On 13 August 2015 one mature, undamaged leaf per seedling was sampled from eight
254 seedlings (two from each block) in 25%, 50% and 100% irrigation regimes. We expected that
255 anatomical acclimation in needles could be observed among extremely different irrigation
256 treatments, therefore the 75% irrigation treatment was not considered in the microscopic leaf
257 variables. A 2 mm wide segment from the central part of the needle or the broadleaf lamina
258 (including midrib) was excised and fixed by vacuum infiltration for 2 hours in a mixture of
259 paraformaldehyde (2%) and glutaraldehyde (2%) followed by overnight incubation under
260 atmospheric pressure. Samples were dehydrated in graded ethanol series. *Abies*, *Fagus* and
261 *Quercus* samples were embedded in Technovit resin (Heraeus Kulzer GmbH, Germany),
262 sectioned to 5 μm with a microtome (Leica RM 2265, Leica Biosystems, Germany) and
263 stained with 0.01% solution of toluidine blue in 1% sodium borate. Sections of *Pinus* needles
264 (35 μm thick) were obtained using a VT 1200 S vibratome (Leica, Germany). Micrographs
265 were taken using a light microscope (Axioscope A1, Zeiss, Germany) with an attached digital

266 camera (AxioCam MRC5, Zeiss). Micrometric traits were measured with Axiovision 4.9.1
267 (Zeiss, Germany). In each leaf we determined the thickness of lamina, the length of palisade
268 cells (mean of 3-5 cells per leaf, except in *Pinus* where palisade mesophyll does not occur),
269 transverse area of xylem and phloem in the central vein, and mean and maximal diameter of
270 conduits (based on 10 widest vessels in the angiosperms and tracheids in the conifer species).

271

272 Determination of leaf proline

273 Proline concentration was determined with the method of Bates et al. [62]. Fully expanded
274 leaves from the apical portion of the seedlings were sampled on 7 August 2015. Purified
275 proline was used to standardize the procedure for quantifying sample values. Acid-ninhydrin
276 was prepared by warming 1.25 g ninhydrin in 30 ml glacial acetic acid and 20 ml 6 M
277 phosphoric acid at the temperature 75°C until dissolved. A 0.3 g sample of plant material was
278 homogenized in 3 ml of 5% aqueous sulfosalicylic acid and the homogenate filtered through
279 Whatman # 2 filter paper. A 2 ml aliquot of filtrate was incubated with 2 ml of acid-ninhydrin
280 and 2 ml of glacial acetic acid in a test tube for 1 hour at 100°C, and the reaction was
281 terminated in an ice bath. The reaction mixture was extracted with 4 ml toluene by mixing
282 with a test tube stirrer for 15-20 sec. The chromophore containing toluene was aspirated from
283 the aqueous phase, warmed to room temperature and the absorbance read at $\lambda = 515$ nm
284 (Spekol, CarlZeiss, Jena) using toluene as blank. The proline concentration was determined
285 from a standard curve and calculated on a dry mass basis using the formula $S_k = K \times A \times T$,
286 where: K is coefficient calculated with the standard curve, A is absorbance and T is
287 conversion factor accounting for the volume of toluene. Proline leaf concentration was
288 expressed in $\mu\text{g g}^{-1}$ DM.

289

290 Expression of stress-related genes

291 Leaf samples for RNA extraction were collected on 7 August 2015. Leaves without
292 symptoms of necrosis or other visible damage were collected. In each species five seedlings per
293 block and treatment were sampled and three samples were used for analyses. Leaf material was
294 immediately frozen in liquid nitrogen and stored at -80°C until RNA extraction.

295 Plant material was ground in liquid nitrogen and total RNA was extracted with 2% B-
296 mercaptoethanol in the extraction buffer as described by Chang et al. [63]. Plant material was
297 ground in liquid nitrogen and total RNA was extracted with 2% B-mercaptoethanol in the
298 extraction buffer as described by Chang et al. [63]. The isolated RNA was dissolved in 30 μl

299 ddH₂O. The extracted RNA was quantified using the Qubit[®]RNA BR Assay kit (Life
300 Technologies, Carlsbad, CA, USA). The quality of the RNA was checked on a 1% agarose
301 gel. One microgram of RNA per sample was treated with DNase RQ (Life Technologies) and
302 used for cDNA synthesis (Transcriptor First Strand cDNA Synthesis Kit, Roche). The
303 resulting cDNA was diluted 1:10 and used for quantitative real-time PCR in a LightCycler 96
304 (Roche, Mannheim, Germany). To each well 2.5 µl of diluted cDNA, 10 µl of SYBR Green I
305 Master (Roche), 1.25 µl of 5 µM mixture of the forward and reverse primers and 5 µl of
306 ddH₂O were added for the amplification reaction. Each biological sample for each gene was
307 run twice. The PCR programs were as follows: 5 min at 95°C, 45 cycles of amplification for
308 10 s at 95°C, 10 s at primer annealing temperature (Supplementary Table 1), and 20 s at 72°C.
309 Specificity of PCR amplification was analyzed by melting curve and peaks (60 – 95°C, with
310 temperature ramp 2.2°C per s) and verified on agarose gel.

311 After the RT-qPCR procedure was complete, the quantification cycles were determined
312 using the second derivative maximum method implemented in the Roche LightCycler 96. The
313 primer efficiency was determined by a dilution series and ranged between 1.8 and 2.0 for the
314 genes of this study. The relative expression was analyzed with $\Delta\Delta C_T$ [64]. The potential
315 reference genes were tested for their expression stability among the all treatment groups using
316 Normfinder [65] implemented in the RefFinder [66]. Gene expression was normalized by the
317 gene expression derived from housekeeping genes: Ubiquitin (*UBQ*, GenBank: AF461687) for
318 *P. sylvestris*, Cyclophilin 2 (*CYC*, GenBank: ERP001867) for *Q. robur*, actin (*ACT*, GenBank:
319 AM063027) for *F. sylvatica* and translation initiation factor IF-2 subunit alpha (*TIF2A*,) for *A.*
320 *alba* (Supplementary Table 1). Real-time quantitative RT-PCR (qRT-PCR) was used to
321 describe plants responses to water deficit. On the base of published data, we selected drought-
322 related genes which were previously found by others to be related to drought stress response
323 (up-regulated or down-regulated) or to abiotic stress in analyzed species. In *F. sylvatica*,
324 abscisic acid (*ABA*)-related drought signaling genes were analyzed: *NCED1* - e9-cis-epoxy-
325 dioxygenase - required for *ABA* biosynthesis (GenBank: DQ787262); *PP2C* - protein phosphate
326 2C - involved in *ABA* signal transduction (GenBank: AJ277743); *ERD10* - early responsive to
327 dehydration, an *ABA*-responsive transcription factor that attenuates *ABA* responses (GenBank:
328 FR775803); stress protection *APX1* - ascorbate peroxidase genes encoding enzymes required
329 for the scavenging of superoxide radicals, hydrogen peroxide and toxic aldehydes (GenBank:
330 FR774767) [67]. In *Q. petraea* the following genes expression was investigated: α -tubulin
331 (*TUB*) related with actively dividing tissues; Zn-finger protein of *Arabidopsis thaliana* (*ZAT11*)

332 – a gene involved in oxidative stress-induced programmed cell death; glutathione-s-transferase
333 (*GST*) and used primers were previously published by Makela et al. [68]. In *P. sylvestris*: genes
334 involved in biosynthesis (*SPDS* - spermidine synthase, GenBank: HM236827) and metabolism
335 of polyamines (*PAO* – polyamine oxidase, GenBank: HM236830); catalase (*CAT*, GenBank:
336 EU513163), involved in reactive oxygen species homeostasis regulation; pyruvate
337 decarboxylase (*PDC*) (GenBank: CO161777.1) marker of alcohol fermentation in tissues under
338 hypoxia; late-embryogenesis abundant protein (*LEA*), (GenBank: AAX68990.1) the well-
339 known osmoprotectors; glutamate-cysteine ligase (*GCL*), (GenBank: AJ132540.1);
340 glyceraldehyde-3-phosphate dehydrogenase (*GAPDH*) (GenBank: L07501) [69]. In *A. alba*:
341 genes classified as early responsive to dehydration (*ERD10*) (GenBank: P42759);
342 chloroplastic beta-amylase 1, b isoform (*BAMI.b*) - (GenBank: Q9LIR6); two pore calcium
343 channel protein 1 *TPC1* (GenBank: AT4G03560) involved in stomatal closure and abiotic stress
344 response; xyloglucan endotransglucosylase/hydrolase (*XTH7*, GenBank: Q8LER3) related with
345 cell growth; one of putative uncharacterized protein (*PUP1.a10.13.9*) (GenBank: A9NLY4)
346 belongs to dehydrin family [58].

347

348 Statistical analyses

349 Prior to analyses, the data were tested for normality and homogeneity of variance in groups
350 with Shapiro-Wilk's and Levene's test, respectively. Data were \log_{10} transformed to fulfill the
351 ANOVA conditions. One-way ANOVA with irrigation regimes as the fixed factor and blocks
352 as the random factor was applied to compare the irrigation treatments within each species for
353 water contents in organs, whole plant dry mass, biomass allocation and gene expression or two-
354 way ANOVA with species and irrigation regimes as fixed factors for leaf proline concentration
355 and anatomical leaf traits at $P < 0.05$. When ANOVA showed significant differences, the mean
356 values were compared with the analysis of contrasts at $P < 0.05$.

357

358 Results

359 Seedling water status

360 At time of harvest both gymnosperm species (*Abies* and *Pinus*) had higher LWC (leaf water
361 content) and lower RWC (root water content) than the angiosperm species (*Fagus* and *Quercus*)
362 whereas their SWC (stem water content) were similar (Fig. 1). Lower irrigation resulted in
363 slightly reduced LWC in *Quercus* and *Pinus* although most pairwise contrasts were non-
364 significant (Fig. 1 a). There were also slight decreases of RWC in *Fagus* and SWC in *Quercus*

365 and *Abies* due to reduced irrigation (Fig. 1 b, c). Thus, the measurement of water contents
366 indicated only moderate organ-level water stress.

367

368 Growth

369 Reduction in water availability resulted in decreased height and root collar diameter of *Fagus*,
370 *Quercus* and *Pinus* seedlings while in *Abies* the growth response to the different irrigation
371 regimes was not significant (Table 1). The magnitude of response was largest in *Fagus* (2.2 and
372 1.8-fold reduction in respectively, height and diameter), followed by *Quercus* (1.8 and 1.5-fold
373 reduction and *Pinus* (1.5 and 1.2 fold). Significant height reductions occurred as irrigation was
374 decreased from 100% to 75% (*Fagus* and *Quercus*) and from 75% to 50% and from 50% to 25%
375 of the recommended dose (all three species). Mean slenderness indices (h/d) decreased with
376 reduced irrigation in *Fagus*, *Quercus* and *Pinus* but not in *Abies* (Table 1).

377 Biomass accumulation differed significantly among species, with seedlings of the two
378 angiosperms *Fagus* and *Quercus* reaching over 2.5 g dry mass under full irrigation, in contrast
379 to the gymnosperms *Abies* and *Pinus* that reached only, respectively, 0.1 and 0.4 g (Table 2;
380 Fig. 2a, b). Irrigation regime affected seedling growth in a dose-response manner in three out
381 the four species (Fig. 2). Under 25% irrigation, dry mass of *Fagus* seedlings was over 5 times
382 smaller than under the 100% dose, whereas in *Quercus* the difference was about three-fold (Fig.
383 2a). There was, however no effect of varied irrigation on the biomass of *Abies* and the biomass
384 decrease between 100% and 25% doses in *Pinus* was approximately by 1/3 (Fig. 2b).

385

386 Allocation

387 Alteration of biomass allocation in response to reduced irrigation was species-specific (Fig. 3
388 a-d). The strongest response was observed in *Pinus*, in which ABMR (aboveground to
389 belowground mass ratio) strongly declined with irrigation reduction from 75 to 50%, resulting
390 from an increased RMF (root mass fraction) and decreases in both LMF (leaf mass fraction)
391 and SMF (stem mass fraction). In contrast, allocation in *Abies* was not affected by irrigation. In
392 *Fagus*, reduced irrigation resulted in increased LMF and decreased SMF without effect at the
393 root or ABMR level. Finally, in *Quercus* only a minor decrease in ABMR was noted at
394 irrigation reduction from 100 to 75%, with variability in organ-level allocation not unaccounted
395 for by irrigation.

396

397 Leaf proline concentration

398 Under 100% irrigation leaf proline concentration was similar across all species ($F_{3, 12} = 1.9$, P
399 $= 0.183$) (Fig. 4). Decline in irrigation resulted in an increase in proline concentration in the
400 two angiosperms but not in the gymnosperm species. Specifically, the increased levels of
401 proline were observed at 50 and 25% dose in *Fagus* and at 75% and less in *Quercus*. The
402 magnitude of proline increase due to irrigation reduction was about two-fold in *Fagus* and
403 somewhat less in *Quercus*.

404

405 Anatomical leaf traits

406 Anatomical leaf traits were analyzed in seedlings exposed to three irrigation regimes: 25, 50 or
407 100% (Fig. 5). We found evidence for acclimation of some anatomical traits in *Fagus* and
408 *Quercus*. Reduction of irrigation resulted in lower leaf lamina thickness in *Quercus*, and
409 shortening of the palisade cells in *Fagus* and *Quercus* (with only a single palisade layer present;
410 Fig. 5 a, b). Both angiosperms also showed reduced development of vascular tissues, with lower
411 cross-sectional areas of xylem and phloem (Fig. 5 c, d) and their vessels exhibited smaller
412 conduits (Fig. 5 e, f). The differences typically occurred between 100% and 50% but not
413 between 50 and 25% irrigation levels. In contrast, none of the above traits varied among
414 treatments in the two conifer species.

415

416 The expression levels of selected stress-response genes

417 In *F. sylvatica*, all transcripts of the analyzed genes were present in leaf samples, but their
418 relative level of gene expression was not affected by treatment ($P > 0.05$). In *Q. petraea*,
419 efficiency of primers for *TUB* was more than 2.0, therefore this transcript was not analyzed. An
420 increase in relative gene expression of *ZAT11* with a decrease in water availability was observed
421 ($r = 0.7059$, $F = 9.9341$, $P = 0.0103$), whereas the relative level of gene expression for *GST* was
422 not affected ($P > 0.05$). In *Pinus sylvestris*, similarly as in *Fagus*, all transcripts were present in
423 all samples, but the relative level of gene expression of all analyzed transcripts was not affected
424 by the irrigation treatments ($P > 0.05$). In *A. alba*, all transcripts were present in the analyzed
425 samples. Relative gene expression of all but one transcript was not affected by the irrigation
426 treatments. We observed significant increase of relative gene expression of *TPCC* in response
427 to a decrease in water dose ($r = 0.8221$, $F = 20.8453$, $P = 0.001$).

428

429 Discussion

430 *Species-specific growth responses to irrigation and alterations in gene expression*

431 In this study of containerized-seedlings, large interspecific differences were observed in
432 the response of seedling growth to reduced irrigation. These differences were associated with
433 the classification of the different species. In agreement with our first hypothesis, the broadleaf
434 deciduous genera, *Fagus* and *Quercus*, exhibited a greater magnitude of response to reduced
435 irrigation than the two evergreen, conifer genera, *Abies* and *Pinus*. Within the deciduous species,
436 *F. sylvatica* was more greatly impacted by reduced water availability than *Q. petraea*. The
437 response of the deciduous species was in agreement with previous results obtained for seedlings
438 and older trees of *Q. petraea*, indicating that it should be classified as a drought-tolerant species
439 [70,71,72] and *F. sylvatica* as a drought-intolerant species [73,74]. Unexpectedly, the shade-
440 tolerant and climax species, *A. alba*, exhibited a weak phenotypic response to reduced level of
441 irrigation, but the enhanced expression of *TPCC* observed in needles may indicate greater levels
442 of stomatal closure in response to the lowest level of irrigation [58]. On the other hand, the
443 pioneer and shade-intolerant species, *P. sylvestris*, exhibited a positive growth response and
444 high morphological plasticity. The difference between the response of the studied conifers may
445 be the result of their contrasting growth phenology as seedlings. The slow-growing species, *A.*
446 *alba*, with its small amount of leaf biomass, requires less water than the faster growing, pioneer
447 species, *P. sylvestris*. Although all seedlings were exposed to drought during their rapid-growth
448 phase, growth in *Abies* is naturally terminated sooner than in *Pinus*; thus the ability of *Abies* to
449 respond to the decreased water availability may be restricted by its phenological cycle. This
450 conservative growth pattern of *A. alba* may represent a portion of a drought-avoidance strategy,
451 providing another example of a trade-off between survival and growth [75]. Seeds of *A. alba*
452 are also heavier than seeds of *P. sylvestris* and thus provide more resources to germinating
453 seedlings during their initial growth than seeds of *P. sylvestris*. This would provide an
454 advantage to *A. alba* seedlings, especially under water deficit conditions [76]. The interspecific
455 differences observed in our experiment resulted to some extent from a different species-specific
456 interception, which reduced amount of water reaching roots of seedlings growing in containers.
457 In mature *P. sylvestris* stands in Germany, mean precipitation interception was 32% [77], and
458 in leafed period in *F. sylvatica* forest in Belgium, the level of interception was 28% [78]. These
459 data cannot be directly compared with the interception of our study seedlings, but they allow to
460 estimate the importance of interception in the water balance of trees under natural conditions.
461 In our study, however, the interception effect on water amount reaching the substrate and roots
462 of seedlings in containers was substantially reduced: (1) Water was delivered at high pressure:
463 at the sprinklers the pressure was 4 bars, and at crowns of seedlings around 3.0 – 3.2 bars and

464 never lower than 2.5 bars. Due to high pressure, even after full leaf expansion, water reached
465 the substrate in cavities; (2) We cultivated our plants from seeds, thus at the beginning of the
466 experiment, the value of interception was 0 and during seeds' germination and initial growth,
467 it did not substantially affect the amount of water irrigating the substrate. An effect of
468 interception was important at the end of our experiment, after full leaf expansion. However,
469 this effect of different interception might be at least partially compensated by the high pressure
470 of irrigation water, high air humidity in the tunnel and foliar water absorption of seedlings
471 which can be significant under water deficit [79].

472 The select genes allowed us to compare tolerance to reduced irrigation within each
473 species. The relative abundance of most of the analyzed drought-response-related transcripts
474 was not affected by the different levels of irrigation except for *ZAT11* in *Q. petraea* and *TPCI*
475 in *A. alba* seedlings (Fig. 6). *ZAT11* was involved in oxidative stress-induced programmed cell
476 death [80] (Qureshi et al. 2013) and positive regulation of primary root growth in *A. thaliana*
477 [81]. Higher *ZAT11* could be associated with a higher production of reactive oxygen species
478 due to drought stress. An increase in *TPCI* was involved in stomatal closure and abiotic stress
479 responses in *Triticum aestivum* [82]. Although *TPCI* was proposed as a reference gene with
480 stable expression for drought stress [58], we found the opposite result. In our study, higher
481 abundance of this transcript can be related with a greater stomatal closure in *A. alba* under
482 drought stress. Importantly, this suggests that the intraspecific selection may be initiated by
483 regulated water deficit. Since only leaf mRNAs were analyzed in our study, it is plausible that
484 the expression of these drought-response-related genes may have been higher in roots. Under a
485 higher water deficit than that in our study, *Populus alba* × *Populus tremula* var. *glandulosa*
486 increased transcript levels of several drought markers in leaves and roots [83]. In agreement
487 with our results, Muilu-Mäkelä et al. [69] reported that the expression of genes involved in
488 biosynthesis was not affected by drought stress in *P. sylvestris* seedlings. They did find,
489 however, that the expression of the catabolism-related genes encoding diamine oxidase and
490 polyamine oxidase were down-regulated in stems in response to drought. The increased level
491 of *LEA* expression observed in that study in all parts of the seedlings indicates that the level of
492 drought stress imposed on *P. sylvestris* seedlings in that study was higher than the level of
493 drought stress imposed in the present study. A lack of significant expression or low expression
494 of the selected stress related, potential marker genes in the studied species suggests that the
495 water deficit stress was efficiently moderated by stomatal closure.

496

497 *Seedling water status*

498 Relative tissue water content was used as a measure of plant water status. In our study,
499 organ-level water content decreases caused by water deficit were small especially in the
500 conifers, suggesting an efficient control of transpirational water loss. This was especially true
501 in *Abies*. In addition, seedlings' water status could be substantially improved by foliar
502 absorption of intercepted water, especially under water deficit. In an earlier study, the
503 substantial improvement of water status, exceeding 1.0 MPa water potential for drought-
504 stressed *Juniperus monosperma* plants was observed, following precipitation on an
505 experimental plot that excluded soil water infiltration [79].

506 Conifers at the same time showed much larger hydration of leaves across irrigation
507 levels as compared to the two angiosperms. This may be attributed to the low amount of
508 mechanical tissues and the large mesophyll content in juvenile needles. Interestingly, conifer
509 root systems were much less hydrated than angiosperms, however since conifer roots
510 constituted lower fraction of biomass than leaves, conifer seedlings held more water per gram
511 total biomass than the angiosperm seedlings.

512 Our results suggest that the ability to accumulate and conserve foliar water may
513 constitute a fundamental difference between drought-survival strategies of broadleaved
514 deciduous and evergreen conifer species. This is supported e.g., by a tighter stomatal control,
515 substantial decrease in sap flow rates and transpiration in *P. sylvestris* compared with more
516 drought-tolerant *Quercus pubescens* [84,85]. Our suggestion is consistent with the higher
517 growth rates of deciduous angiosperm seedlings achieved through a more intensive gas
518 exchange, especially when expressed on the leaf mass basis. In the general scheme of ecological
519 strategies, conifers appear to emphasize water conservation in contrast to specialization towards
520 water acquisition and spending in seedlings of broadleaf angiosperms.

521 522 *Biomass allocation*

523 Evidence from individual and multi-species studies shows that plants typically respond
524 to water deficit by increasing RMF and decreasing ABMR, LMF and SMF [15,21,7,75]. This
525 pattern was most closely represented by the response of *P. sylvestris*. In contrast, we did not
526 observe modification of RMF in any other species while, surprisingly, *Fagus* responded to
527 water deficit by increasing LMF. While the lack of modification of biomass allocation in *Abies*
528 is understandable given its lack of growth response, the unexpected increase in LMF with
529 drought observed in *Fagus* may reflect the opposite trend of SMF. Such result may be explained

530 by the intrinsic influence of plant size on allocation ratios: small plants tend to have low fraction
531 of structural tissue (stem) and a large fraction of leaves; an effect often confounding the results
532 of experimental treatments or environmental factors [21].

533 Increased RMR as seen in droughted *Pinus*, is an important morphological adaptation
534 to low availability of underground resources [86,21]. Under prolonged drought, moderate
535 increases in root mass ratio may increase survival and ensure sufficient water supply for
536 photosynthesis, whereas increasing this ratio beyond a level required to ensure survival and
537 favourable water relations may impose construction and maintenance costs resulting in net
538 inhibition of plant growth. The role of biomass partitioning can be more important than either
539 gas exchange or phenology as a mechanism of drought response which has been suggested by
540 the results regarding *Brassica rapa* [75].

541 A decrease in growth under water deficit may be advantageous for plants if it leads to a
542 reduction of leaf transpiration area. However, under natural conditions, lower allocation to
543 leaves and overall plant size reduction reduce the odds for individual's success in competition
544 for light with other plants. This problem does not occur in container cultivation as long as
545 seedlings are spaced to prevent significant competition. Additionally, higher RMR of seedlings
546 acclimated to lower irrigation is an advantageous trait when they are planted in dry sites. Given
547 the prevalence of drought episodes, high seedling survival rates may be more important than
548 their initial growth rates and biomass production [52,87]. Thus, morphological acclimation of
549 seedlings to lower water doses in a nursery, especially stimulation of increase in RMR, may
550 benefit their survival in the planting sites. However, this expectation should be verified by
551 results of further experiments with seedlings of various species produced under the different
552 irrigation treatments in the container nursery and then planted and grown in natural conditions.

553

554 *Osmoregulation in response to regulated water deficit*

555 Plants synthesize saccharides, alcohols, and amino-acids including proline that
556 accumulate within the protoplast, especially in the vacuole, leading to a decrease in cell osmotic
557 potential [88,54,30]. Besides facilitating soil water extraction, some of these compounds,
558 including proline, perform protective roles against oxidative damage associated with reduced
559 gas exchange [89]. Although proline accumulates in response to water deficit in both
560 angiosperm trees [90] and conifers [91], in our experiment such increase in proline
561 concentration occurred only in angiosperm species but not in the conifers. In our two conifers
562 this result, together with the relative homeostasis of needle water content, suggests that needle-

563 level signal of water deficit was not sufficient to induce this biochemical response. Alternatively,
564 conifers might accumulate some alternative osmolytes [92]. The utility of proline as an indicator
565 of water deficit stress, although widely accepted [91], certainly depends on the species, ecotype
566 and stress intensity as well as tissue type [92,90].

567

568 *Leaf anatomy*

569 Modifications of anatomical leaf traits in response to reduced irrigation were rather
570 minor and involved small reduction in lamina thickness in *Quercus*, decrease in palisade cell
571 length in *Quercus* and *Fagus* and decreases in the amounts of xylem and phloem tissues as well
572 as in vessel diameters in these two angiosperms. In contrast, none of the conifer needles showed
573 significant modifications. The character of changes in the two broadleaved species was only
574 partly consistent with the previously described modifications induced by drought. Leaves that
575 express constitutive or induced xeromorphism are often thicker than mesomorphic leaves and
576 show greater development of palisade tissue [93,23,94]. Reduction, rather than increase, in leaf
577 thickness and palisade cell length observed in our study suggests occurrence of a leaf-level
578 water deficit during leaf expansion and may be classified as symptom of water stress rather than
579 adaptive modifications. On the other hand, the decrease in transverse areas of xylem and phloem
580 are consistent with acclimation to lower gas exchange. Such plastic changes have been reported
581 in both drought and salinity stressed plants [95,96]. The change in leaf conduit diameter have
582 rarely been examined in studies of drought acclimation, although the few studies reporting such
583 data also show that vessels or tracheids become narrower in leaves developing under drought
584 [97,98]. Such modification may reflect reduced demand for water transport capacity under
585 decreased stomatal conductance and, as an additional benefit, reduce the vulnerability of narrow
586 vessels to embolization by decreasing the probability of 'rare leaky pits' [99,100].

587 The quantitatively small responses of angiosperm leaves and lack of response of
588 gymnosperm needles to drought suggest that water deficit in meristematic and growing shoot
589 tissues at the time of leaf development was not severe enough to induce significant plasticity of
590 these traits (see Ivancich et al. [101] and Binks et al. [24] for similarly conservative responses).
591 It is likely that the adjustment of growth rate and/or allocation of biomass, coupled with
592 physiological adjustment, helped to at least partially offset the effect of lower availability of
593 water and preserve the plant water status at least during the period of leaf expansion. Moreover,
594 leaves of young seedlings are known to be anatomically less plastic than those of later
595 ontogenetic stages [102].

596

597 *Conclusions*

598 The divergent responses of tree species to regulated water deficit indicate that the
599 irrigation regimes in container nurseries would benefit from species-specific adjustment
600 according to water requirements. Our results indicate that in case of gymnosperm evergreen
601 conifer species the quantity of irrigation water could be reduced by 25% (for *P. sylvestris*) to
602 75% (*A. alba*), and in case of angiosperm deciduous broadleaf (*F. sylvatica* and *Q. petraea*) by
603 25% relative to the full dose of 6 mm without significantly affecting growth and inducing
604 morphological, biochemical and molecular mechanisms of defense against water deficit.
605 Although in both angiosperms growth was sensitive to water deficit, only very modest induction
606 of stress indicators was observed suggesting reduction of water dose could be considered when
607 hardening to field conditions is required.

608

609 **Author contributions statement:** Study conception and design: WB, PR, WK, acquisition of
610 data: PR, WK, TW, EPK, BP, AJ; analysis and interpretation of data PR, TW, EPK; drafting
611 of first version of the manuscript: PR and final version: PR, TW, EPK, BP, WB, WK and AJ.

612

613 **Conflict of Interest:** The authors declare that they have no conflict of interest.

614

615 **Bibliography**

- 616 1. Hoegh-Guldberg, O.; Jacob, D.; Taylor, M.; Bindi, M.; Brown, S.; Camilloni, I.;
617 Diedhiou, A.; Djalante, R.; Ebi, K.; Engelbrecht, F.; Guiot, J.; Hijioka, Y.; Mehrotra,
618 S.; Payne, A.; Seneviratne, S.I.; Thomas, A.; Warren, R.; Zhou, G. Impacts of 1.5°C
619 Global Warming on Natural and Human Systems. In: Global warming of 1.5°C. An
620 IPCC Special Report on the impacts of global warming of 1.5°C above pre-industrial
621 levels and related global greenhouse gas emission pathways, in the context of
622 strengthening the global response to the threat of climate change, sustainable
623 development, and efforts to eradicate poverty [V. Masson-Delmotte, P. Zhai, H. O.
624 Pörtner, D. Roberts, J. Skea, P.R. Shukla, A. Pirani, W. Moufouma-Okia, C. Péan, R.
625 Pidcock, S. Connors, J. B. R. Matthews, Y. Chen, X. Zhou, M. I. Gomis, E. Lonnoy,
626 T. Maycock, M. Tignor, T. Waterfield (eds.)], 2018.
- 627 2. Lindner, M.; Maroschek, M.; Netherer, S.; Kremer, A; Barbati, A; Garcia-Gonzalo, J;
628 Seidl, R.; Delzon, S.; Corona, P.; Kolstro, M.; Lexer, M.J.; Marchetti, M. Climate

- 629 change impacts, adaptive capacity, and vulnerability of European forest ecosystems.
630 *For. Ecol. Manage.* **2010**, 259, 698–709.
- 631 3. Wang, Y.; Hogg, E.H.; Price, D.T.; Edward, J.; Williamson, T. Past and projected
632 future changes in moisture conditions in the Canadian boreal forest. *Forest. Chron.*
633 **2014**, 90, 678–691.
- 634 4. Reich, P.B.; Sendall, K.M.; Stefanski, A.; Rich, R.L.; Hobbie, S.E.; Montgomery,
635 R.A. Effects of climate warming on photosynthesis in boreal tree species depend on
636 soil moisture. *Nature* **2018**, <https://doi.org/10.1038/s41586-018-0582-491>
- 637 5. Allen, C.D.; Macalady, A.K.; Chenchouni, H.; Bachelet, D.; McDowell, N. et al. A
638 global overview of drought and heat-induced tree mortality reveals emerging climate
639 change risks for forests. *For. Ecol. Manage.* **2010**, 259(4), 660–684.
640 <https://doi.org/10.1016/j.foreco.2009.09.001>
- 641 6. Maherali, H.; Pockman, W.T.; Jackson, R.B. Adaptive variation in the vulnerability of
642 woody plants to xylem cavitation. *Ecology* **2004**, 85(8), 2184–2199.
- 643 7. Bréda, N.; Huc, R.; Granier, A.; Dreyer, E. Temperate forest trees and stands under
644 severe drought: a review of ecophysiological responses, adaptation processes and
645 long-term consequences. *Ann. For. Sci.* **2006**, 63, 625–644.
- 646 8. Lévesque, M.; Saurer, M.; Siegwolf, R.; Eilmann, B.; Brang, P.; Bugmann, H.;
647 Rigling, H. Drought response of five conifer species under contrasting water
648 availability suggests high vulnerability of Norway spruce and European larch. *Global*
649 *Change Biol.* **2013**, <https://doi.org/10.1111/gcb.12268>
- 650 9. Anderegg, L.D.L.; Hillerislambers, J. Drought stress limits the geographic ranges of
651 two tree species via different physiological mechanisms. *Global Change Biol.* **2015**,
652 22(3),1029-1045.
- 653 10. Leuzinger, S.; Zotz, G.; Asshoff, R.; Körner.; Ch. Responses of deciduous forest trees
654 to severe drought in Central Europe. *Tree Physiol.* **2005**, 25, 641–650.
- 655 11. Meier, I.C.; Leuschner, C. Genotypic variation and phenotypic plasticity in the
656 drought response of the fine root system of European beech. *Tree Physiol.* **2008**, 28,
657 297–309.
- 658 12. Metz, J.; Annighöffer, P.; Schall, P.; Zimmermann, J.; Kahll, T.; Schulze, E-D.;
659 Ammer, Ch. Site-adapted admixed tree species reduce drought susceptibility of mature
660 European beech. *Global Change Biol.* **2016**, 22(2), 903-920.
661 <https://doi.org/10.1111/gcb.13113>

- 662 13. Valladares, F.; Sánchez-Gómez, D. Ecophysiological traits associated with drought in
663 mediterranean tree seedlings: individual responses versus interspecific trends in eleven
664 species. *Plant Biol.* **2006**, *8*, 688–697.
- 665 14. Choat, B.; Jansen, S.; Brodribb, T.J. et al. Global convergence in the vulnerability of
666 forests to drought. *Nature* **2012**, *491*, 752–755.
- 667 15. Abrams, M.D. Adaptations and responses to drought in *Quercus* species of North
668 America. *Tree Physiol.* **1990**, *7*, 227–238.
- 669 16. Warren, C.R.; Bleby, T.; Adams, M.A. Changes in gas exchange versus leaf solutes
670 as a means to cope with summer drought in *Eucalyptus marginata*. *Oecologia* **2007**,
671 *154*:1–10.
- 672 17. Voltaire, F. A unified framework of plant adaptive strategies to drought: crossing
673 scales and disciplines. *Global Change Biol.* **2018**, *24*, 2929–2938.
- 674 18. McDowell, N.; Pockman, W.T. .; Allen, C.D.; Breshears, D.D.; Cobb, N.; Kolb, T.;
675 Plaut, J.; Sperry, J.; West, A.; Williams, D.G.; Yezzer, E.A. Mechanisms of plant
676 survival and mortality during drought: why do some plants survive while others
677 succumb to drought? *New Phytol.* **2008**, *178*, 719–739.
- 678 19. van Hess, A.F.M. Growth and morphology of pedunculate oak (*Quercus robur* L.) and
679 beech (*Fagus sylvatica* L.) seedlings in relation to shading and drought. *Ann. Sci. For.*
680 **1996**, *54*, 9–18.
- 681 20. Tognetti, R.; Johnson, J.D.; Michelozzi, M. The response of European beech (*Fagus*
682 *sylvatica* L.) seedlings from two Italian populations to drought and recovery. *Trees*
683 **1995**, *9*, 348–354.
- 684 21. Poorter, H.; Niklas, K.J.; Reich, P.B.; Oleksyn, J.; Poot, P.; Mommer, L. Biomass
685 allocation to leaves, stems and roots: meta-analysis of interspecific variation and
686 environmental control. *New Phytol.* **2011**, *193*(1), 30–50.
- 687 22. Galmés, J.; Ochogavía, J.M.; Gago, J.; Roldán, E.J.; Cifre, J.; Conesa, M.À. Leaf
688 responses to drought stress in Mediterranean accessions of *Solanum lycopersicum*:
689 anatomical adaptations in relation to gas exchange parameters. *Plant Cell Environ.*
690 **2013**, *36*, 920–935.
- 691 23. Zang, C.; Hartl-Meier, C.; Dittmar, C.; Rothe, A.; Menzel, A. Patterns of drought
692 tolerance in major European temperate forest trees: climatic drivers and levels of
693 variability. *Global Change Biol.* **2014**, *20*, 3767–3779.

- 694 24. Binks, O.; Meir, P.; Rowland, L.; da Costa, A.C.L.; Vasconcelos, S.S.; de Oliveira,
695 A.A.R.; Ferreira, L.; Christoffersen, B.; Nardini, A.; Mencuccini, M. Plasticity in leaf-
696 level water relations of tropical rainforest trees in response to experimental drought.
697 *New Phytol.* **2016**, 211, 477–488.
- 698 25. Tyree, M.T.; Ewers, F.W. The hydraulic architecture of trees and other woody plants.
699 *New Phytol.* **1991**, 119, 345–360.
- 700 26. Tyree, M.; Davis, S.; Cochard, H. Biophysical perspectives of xylem evolution: is
701 there a tradeoff of hydraulic efficiency for vulnerability to dysfunction? *International*
702 *Association of Wood Anatomists Journal*, **1994**, 15, 335–360.
- 703 27. Sperry, J.S.; Hacke, U.G.; Pittermann, J. Size and function in conifer tracheids and
704 angiosperms vessels. *Am. J. Bot.* **2006**, 93(10), 1490–1500.
- 705 28. Chaves, M.M.; Maroco, J.P.; Pereira, J.S. Understanding plant responses to drought –
706 from genes to the whole plant. *Funct. Plant Biol.* **2003**, 30, 239–264.
- 707 29. Liu, Ch.; Liu, Y.; Guo, K.; Fan, D.; Li, G.; Zheng, Y.; Yu, L.; Yang, R. Effect of
708 drought on pigments, osmotic adjustment and antioxidant enzymes in six woody plant
709 species in karst habitats of southwestern China. *Environ. Exp. Botany* **2011**, 71, 174–
710 183.
- 711 30. Reguera, M.; Peleg, Z.; Abdel-Tawab, Y.M.; Tumimbang, E.B.; Delatorre, C.A.;
712 Blumwald, E. Stress-induced cytokinin synthesis increases drought tolerance through
713 the coordinated regulation of carbon and nitrogen assimilation in rice. *Plant Physiol.*
714 **2013**, 163, 1609–1622.
- 715 31. Vandeleurs, R.K.; Mayo, G.; Shelden, M.C.; Gilliam, M.; Kaiser, B.N.; Tyerman,
716 S.D. The role of plasma membrane intrinsic protein aquaporins in water transport
717 through roots: diurnal and drought stress responses reveal different strategies between
718 isohydric and anisohydric cultivars of grapevine. *Plant Physiol.* **2009**, 149, 445–460.
- 719 32. Pinheiro, C.; Chaves, M.M. Photosynthesis and drought: can we make metabolic
720 connections from available data? *J. Exp. Bot.* **2011**, 62(3), 869–882.
- 721 33. Markesteijn, L.; Poorter, L. Seedling root morphology and biomass allocation of 62
722 tropical tree species in relation to drought- and shade-tolerance. *J. Ecol.* **2009**, 97(2),
723 311–325.
- 724 34. Borghetti, M.; Cinnirella, S.; Magnani, F.; Saracino, A. Impact of long-term drought
725 on xylem embolism and growth in *Pinus halepensis* Mill. *Trees* **1998**, 12, 187–195.

- 726 35. Walter, J.; Nagy, L.; Heinb, R.; Rascher, U.; Beierkuhnlein, C.; Willner, E.; Jentsch,
727 A. Do plants remember drought? Hints towards a drought-memory in grasses.
728 *Environ. Exp. Bot.* **2011**, 71, 34–40.
- 729 36. Fleta-Soriano, E.; Munné-Bosch, S. Stress memory and the inevitable effects of
730 drought: a physiological perspective. *Front. Plant Sci.* **2016**, 7, 143. [https://doi.org/](https://doi.org/10.3389/fpls.2016.00143)
731 10.3389/fpls.2016.00143
- 732 37. Grassi, G.; Magnani, F. Stomatall, mesophyll conductance and biochemical limitations
733 to photosynthesis as affected by drought and leaf ontogeny in ash and oak trees. *Plant*
734 *Cell Environ.* **2005**, 28, 834–849.
- 735 38. Helman, D.; Lensky, I.M.; Yakir, D.; Osem, Y. Forests growing under dry conditions
736 have higher hydrological resilience to drought than do more humid forests. *Global*
737 *Change Biol.* **2016**, 23, 2801–2817. <https://doi.org/10.1111/gcb.13551>.
- 738 39. Abrams, M.D. Comparative water relations of three successional hardwood species in
739 central Wisconsin. *Tree Physiol.* **1988**, 4, 263–273.
- 740 40. Arango-Velez, A.; Zwiazek, J.J.; Thomas, B.R.; Tyree, M.T. Stomatal factors and
741 vulnerability of stem xylem to cavitation in poplars. *Physiol. Plantarum* **2011**, 143,
742 154–165.
- 743 41. Brzeziecki, B.; Kienast, F. Classifying the life-history strategies of trees on the basis
744 of the Grimian model. *For. Ecol. Manage.* **1994**, 69(1-3), 167–187.
- 745 42. Backes, K.; Leuschner, Ch. Leaf water relations of competitive *Fagus sylvatica* L. and
746 *Quercus petraea* (Matt.) Liebl. trees during four years differing in soil drought. *Can.*
747 *J. For. Res.* **2000**, 30, 335–346.
- 748 43. Epron, D.; Dreyer, E. Long-term effects of drought on photosynthesis of adult oak
749 trees (*Quercus petraea* (Matt.) Liebl. and *Quercus robur* L.) in a natural stand. *New*
750 *Phytol.* **1993**, 125, 381–389.
- 751 44. Epron, D.; Godard, D.; Cornic, G.; Genty, B. Limitations of net CO₂ assimilation rates
752 by internal resistances to CO₂ transfer in leaves of two species (*Fagus sylvatica* L. and
753 *Castanea sativa* Mill.) *Plant Cell Environ.* **1995**, 18, 43–51.
- 754 45. Georgea, J-P.; Schuelera, S.; Karanitsch-Ackerlb, S.; Mayerb, K.; Klumppc, R.T.;
755 Grabnerb, M. Inter- and intra-specific variation in drought sensitivity in *Abies spec.*
756 and its relation to wood density and growth traits. *Agr. Forest Meteorol.* **2015**, 214–
757 215, 430–443.

- 758 46. Toromani, E.; Sanxhaku, M.; Pasho, E. Growth responses to climate and drought in
759 silver fir (*Abies alba*) along an altitudinal gradient in southern Kosowo. *Can. J. For.*
760 *Res.* **2011**, 41, 1795–1807.
- 761 47. Nourtier, M.; Chanzy, A.; Cailleret, M.; Yingge, X.; Huc, R.; Davi, H. Transpiration
762 of silver fir (*Abies alba* Mill.) during and after drought in relation to soil properties in
763 a Mediterranean mountain area. *Ann. For. Sci.* **2012**, 1-13. [https://doi.org/](https://doi.org/10.1007/s13595-012-0229-9)
764 10.1007/s13595-012-0229-9
- 765 48. Bigler, Ch.; Braker, O.U.; Bugmann, H.; Dobbertin, M.; Rigling, A. Drought as an
766 inciting mortality factor in Scots pine stands of the Valais, Switzerland. *Ecosystems*
767 **2006**, 9, 330–343.
- 768 49. Pabian, R. Is a container nursery cost-effective? *Las Polski [Polish Forest]* **2014**, 20.
769 http://www.laspolski.pl/Czy_szkssnkarstwo_kontenerowe_sizo_opnaca_nr_202014_s
770 [trona-2633.html](http://www.laspolski.pl/Czy_szkssnkarstwo_kontenerowe_sizo_opnaca_nr_202014_s) (in Polish, 04 November 2018)
- 771 50. Małek, S.; Banach, J.; Barszcz, J. et al. Optymalizacja produkcji sadzonek z zakrytym
772 systemem korzeniowym w wybranych szkółkach kontenerowych [Optimalization of
773 cover-root seedling production in selected container nurseries]. Final report, **2017**, pp.
774 293–318. https://tbr.lasy.gov.pl/apex/f?p=102:3:::NO::P3_TEMAT:3661 (in Polish)
- 775 51. Heiskanen, J. Favourable water and aeration for growth media in containerized tree
776 seedling production: A review. *Scand. J. For. Res.* **1993**, 8, 337–358.
- 777 52. Lamhamedi, M.S.; Lambany, G.; Margolis, H.A.; Renaud, M.; Veilleux, I.; Bernier,
778 P.Y. Growth, physiology and leachate losses in *Picea glauca* seedlings (1+0) grown in
779 air-slit container under different irrigation regimes. *Can. J. For. Res.* **2001**, 31,
780 1968–1980.
- 781 53. Stowe, D.C.; Lamhamedi, M.S.; Margolis, H.A. Water relations, cuticular
782 transpiration, and bud characteristics of air-slit containerized *Picea glauca* seedlings in
783 response to controlled irrigation regimes. *Can. J. For. Res.* **2001**, 31, 2200–2212.
- 784 54. Ramanjulu, S.; Bartels, D. Drought- and desiccation-induced modulation of gene
785 expression in plants. *Plant Cell Environ.* **2002**, 25, 141–151.
- 786 55. Sancho-Knapik, D.; Sanz, M.A.; Peguero-Pina, J.J.; Niinemets, Ü.; Gil-Pelegrín, E.
787 Changes of secondary metabolites in *Pinus sylvestris* L. needles under increasing soil
788 water deficit. *Ann. For. Sci.* **2017**, 74, 24. <https://doi.org/10.1007/s13595-017-0620-7>
- 789 56. Murray, B.G. Nuclear DNA amounts in Gymnosperms. *Ann. Bot.* **1998**, 82, 3–15.

- 790 57. Bray, E.A. Abscisic acid regulation of gene expression during water-deficit stress in
791 the era of the *Arabidopsis* genome. *Plant Cell Environ.* **2002**, 25, 153–161.
- 792 58. Behringer, D; Zimmermann, H; Ziegenhagen, B; Liepelt, S. Differential gene
793 expression reveals candidate genes for drought stress response in *Abies alba*
794 (Pinaceae). *Plos One* **2015**, <https://doi.org/10.1371/journal.pone.0124564>.
- 795 59. Bizet, F.; Bogeat-Triboulot, M-B.; Montpied, P.; Christophe, A.; Ningre, N.; Cohen,
796 D.; Hummel, I. Phenotypic plasticity toward water regime: response of leaf growth
797 and underlying candidate genes in *Populus*. *Physiol. Plantarum* **2015**, 154, 39–53.
- 798 60. Taeger, S.; Fussi, B.; Konnert, M.; Menzel, A. Large-scale genetic structure and
799 drought-induced effects on European Scots pine (*Pinus sylvestris* L.) seedlings. *Eur. J.*
800 *Forest Res.* **2013**, 132, 481. <https://doi.org/10.1007/s10342-013-0689-y>
- 801 61. Pierzgałski, E.; Tyszka, J.; Boczoń, J.; Wiśniewski, J.; Jeznach, J.; Żakowicz, S.
802 Wytyczne nawadniania szkółek leśnych na powierzchniach otwartych. [Guidelines for
803 irrigation in forest nurseries] CILP, Warsaw, **2002**, pp 5–63.
- 804 62. Bates, L.S.; Waldren, R.P.; Teare, J.D. Rapid determination of proline for water stress
805 studies. *Plant Soil* **1973**, 39, 205–207.
- 806 63. Chang, S.; Puryear, J.; Cairney, J. A simple and efficient method for isolating RNA
807 from pine trees. *Plant Mol. Biol. Rep.* **1993**, 11:113–116. doi: 10.1007/BF02670468.
- 808 64. Livak, K.J.; Schmittgen, T.D. Analysis of relative gene expression data using real-
809 time quantitative PCR and the $2^{-\Delta\Delta C_t}$ method. *Methods* **2001**, 25, 402–408.
- 810 65. Andersen, C.L.; Ledet-Jensen, J.; Ørntoft, T. Normalization of real-time quantitative
811 RT-PCR data: a model based variance estimation approach to identify genes suited for
812 normalization - applied to bladder- and colon-cancer data-sets. *Cancer Res.* **2004**, 64,
813 5245–5250.
- 814 66. Xie, F.; Xiao, P.; Chen, D.; Xu, L.; Zhang, B. miRDeepFinder: a miRNA analysis tool
815 for deep sequencing of plant small RNAs. *Plant Mol. Biol.* **2012**, 80, 75–84.
- 816 67. Carsjens, C.; Ngoc, Q.N.; Guzy, J.; Knutzen, F.; Meier, I.C.H.; Müller, M.; Finkeldey,
817 R.; Leuschner, Ch.; Polle, A. Intra-specific variations in expression of stress-related
818 genes in beech progenies are stronger than drought-induced responses. *Tree Physiol.*
819 **2014**, 34, 1348–1361.
- 820 68. Makela, M.; Michael, P.; Theriault, G.; Nkongolo, K.K. High genetic variation among
821 closely related red oak *Quercus rubra* populations in an ecosystem under metal stress:
822 analysis of gene regulation. *Genes Genom.* **2016**, 38, 967–976.

- 823 69. Muilu-Mäkelä, R.; Vuosku, J.; Läärä, E.; Saarinen, M.; Heiskanen, J.; Häggman, H.;
824 Sarjala, T. Water availability influences morphology, mycorrhizal associations, PSII
825 efficiency and polyamine metabolism at early growth phase of Scots pine seedlings.
826 *Plant Physiol. Bioch.* **2015**, *88*, 70–81.
- 827 70. Bréda, N.; Cochard, H.; Dreyer, E.; Granier, A. Field comparison of transpiration,
828 stomatal conductance and vulnerability to cavitation of *Quercus petraea* and *Quercus*
829 *robur* under water stress. *Ann. Sci. For.* **1993**, *50*, 571–582.
- 830 71. Bréda, N.; Granier, A.; Aussnac, G. La sécheresse de 2003 dans le contexte climatique
831 des 54 dernières années. Analyse écophysiological et influence sur les arbres forstiers.
832 [The 2003 drought in the climate context of the last 54 years – ecophysiological
833 analysis and impact on forest trees]. *Rev. For. Fr.* **2003**, LVI(2), 109–131. (in French)
- 834 72. Ponton, S.; Dupouey, J-L.; Bréda, N.; Dreyer, E. Comparison of water use efficiency
835 of seedlings from two sympatric oak species: genotype x environment interactions.
836 *Tree Physiol.* **2002**, *22*, 413–422.
- 837 73. Betch, P.; Bonal, D.; Bréda, N.; Montpied, P.; Peiffer, M.; Tuzet, A.; Granier, A.;
838 Drought effect on water relations in beech: The contributions of exchangeable water
839 reservoirs. *Agr. Forest Meteorol.* **2011**, *151*, 531–543.
- 840 74. Michelot, A.; Bréda, N.; Damesin, C.; Dufrière, E. Differing growth responses to
841 climatic variations and soil water deficits of *Fagus sylvatica*, *Quercus petraea*, and
842 *Pinus sylvestris* in a temperate forest. *For. Ecol. Manage.* **2012**, *265*, 161–171.
- 843 75. Edwards, Ch.E.; Ewers, B.E.; Weinig, C. Genotypic variation in biomass allocation in
844 response to field drought has a greater effect on yield than gas exchange or phenology.
845 *BMC Plant Biol.* **2016**, *16*, 185. <https://doi.org/10.1186/s12870-016-0876-3>
- 846 76. Tylkowski, T. Przedsiębiorne traktowanie nasion drzew, krzewów, pnączy i krzewinek.
847 [Pre-sowing treatment of seeds of trees, shrubs, climbers and subshrubs]. CILP
848 [Information Centre of the State Forests] **2016**, pp 35–36, 185–186, 300–301,
849 343–344 (in Polish).
- 850 77. Ahrends, B.; Penne, C. Modeling the impact of canopy structure on the spatial
851 variability of net forest precipitation and interception loss in Scots pine stands. *The*
852 *Open Geography Journal* **2010**, *3*, 115–124.
- 853 78. Staelens, J.; De Schrijver, A.; Verheyen, K.; Verhoest, N.E.C. Rainfall partitioning
854 into throughfall, stemflow, and interception within a single beech (*Fagus sylvatica* L.)

- 855 canopy: influence of foliation, rain event characteristics, and meteorology. *Hydrol.*
856 *Process* **2008**, 22, 33 – 45.
- 857 79. Breshears, D.D.; McDowell, N.G.; Goddard, K.L.; Dayem, K.E.; Martens, S.N.;
858 Meyer, C.W.; Brown, K.M. Foliar absorption of intercepted rainfall improves woody
859 plants water status most during drought. *Ecology* **2008**, 89(1), 41–47.
- 860 80. Qureshi, M.K.; Sujeeth, N.; Gechev, T.S.; Hille, J. The zinc finger protein ZAT11
861 modulates paraquat-induced programmed cell death in *Arabidopsis thaliana*. *Acta*
862 *Physiol. Plant.* **2013**, 35, 1863–1871.
- 863 81. Liu, J.; Han, H.J.; Kim, S.H.; Lim, C.O.; Yun, D.J.; Chung, W.K. ZAT11, a zinc
864 finger transcription factor, is a negative regulator of nickel ion tolerance in
865 *Arabidopsis*. *Plant Cell. Rep.* **2014**, 33, 2015–2021.
- 866 82. Wang, Y-J.; Yu, J-N.; Chen, T.; Zhang, Z-G.; Hao, Y-J.; Zhang, J-S.; Chen, S-Y.
867 Functional analysis of a putative Ca²⁺ channel gene *TaTPC1* from wheat. *J. Exp. Bot.*
868 **2005**, 56, 3051–3060.
- 869 83. Jia, J.; Li, S.; Cao, X.; Li, H.; Shi, W.; Polle, A.; Liu, T.X.; Peng, C.; Luo, Z.B.
870 Physiological and transcriptional regulation in poplar roots and leaves during
871 acclimation to high temperature and drought. *Physiol. Plantarum* **2016**, 157, 38–53.
- 872 84. Irvine, J.; Perks, M.P.; Magnani, F.; Grace, J. The response of *Pinus sylvestris* to
873 drought: stomatal control of transpiration and hydraulic conductance. *Tree Physiol.*
874 **1998**, 18, 393–402.
- 875 85. Poyatos, R.; Llorens, P.; Piñol, J.; Rubio, C. Responses of Scots pine (*Pinus sylvestris*
876 L.) and pubescent oak (*Quercus pubescens* Willd.) to soil and atmospheric water
877 deficit under Mediterranean mountain climate. *Ann. For. Sci.* **2008**, 306.
878 <https://doi.org/10.1051/forest:2008003>
- 879 86. Cregg, B.M.; Zhang, J.W. Physiology and morphology of *Pinus sylvestris* seedlings
880 from diverse sources under cyclic drought stress. *For. Ecol. Manage.* **2001**, 154(1–2),
881 131-139. [https://doi.org/10.1016/S0378-1127\(00\)00626-5](https://doi.org/10.1016/S0378-1127(00)00626-5)
- 882 87. Bergeron, O.; Lamhamedi, M.S.; Margolis, H.A.; Bernier, P.Y.; Stowe, D.C. Irrigation
883 control and physiological responses of nursery-grown black spruce seedlings (1+0)
884 cultivated in air-slit containers. *HortScience* **2004**, 39(3), 599–605.
- 885 88. Singh, T.N.; Aspinall, D.; Paleg, L.G. Proline accumulation and varietal adaptability
886 to drought in barley: a potential metabolic measure of drought resistance. *Nature-New*
887 *Biol.* **1972**, 236, 188–190.

- 888 89. Ahmed, Ch.B.; Rouina, B.B.; Sensoy, S.; Boukhriss, M.; Abdullah, F.B. Exogenous
889 proline effects on photosynthetic performance and antioxidant defense system of
890 young olive tree. *J. Agric. Food Chem.* **2010**, *58*(7), 4216–4222.
891 <https://doi.org/10.1021/jf9041479>
- 892 90. Peuke, A.D.; Schraml, C.; Hartung, W.; Rennenberg, H. Identification of drought-
893 sensitive beech ecotypes by physiological parameters. *New Phytol.* **2002**, *154*,
894 373–387.
- 895 91. Schiop, S.T.; Al Hassan, M.; Sestras, A.F.; Boscaiu, M.; Sestras, R.E.; Vicente, O.
896 Biochemical responses to drought, at the seedling stage, of several Romanian
897 Carpathian populations of Norway spruce (*Picea abies* L. Karst). *Trees* **2017**, *31*,
898 1479–1490.
- 899 92. Cyr, D.R.; Buxton, G.F.; Webb, D.P.; Dumbroff, E.B. Accumulation of free amino
900 acids in the shoots and roots of three northern conifers during drought. *Tree Physiol.*
901 **1990**, *6*, 293-303.
- 902 93. Bussotti, F.; Bettini, D.; Grossoni, P.; Mansuino, S.; Nibbi, R.; Soda, C.; Tani, C.
903 Structural and functional traits of *Quercus ilex* in response to water availability. *Environ.*
904 *Exp. Bot.* **2002**, *47*, 11–23.
- 905 94. Stojnić, S.; Orlović, S.; Miljković, D.; Galić, Z.; Kebert, M.; von Wuehlisch, G.
906 Provenance plasticity of European beech leaf traits under differing environmental
907 conditions at two Serbian common garden sites. *Eur. J. Forest Res.* **2015**, *134*, 1109.
- 908 95. Bresta, P.; Nikolopoulos, D.; Economou, G.; Vahamidis, P.; Lyra, D.; Karamanos, A.;
909 Karabourniotis, G. Modification of water entry (xylem vessels) and water exit
910 (stomata) orchestrates long term drought acclimation of wheat leaves. *Plant and Soil*
911 **2011**, *347*, 179–193.
- 912 96. Laajimi, N.O.; Boussadia, O.; Shkiri, F.H.; Teixeira da Silva, J.A.; Rezgui, S.; Hellali,
913 R. Anatomical adaptations in vegetative structures of apricot tree (*Prunus armeniaca*
914 L.) cv. ‘Amor El Euch’ grown under water stress. *Fruit Veget. Cereal Sci. Biotech.*
915 **2011**, *5*, 46–51.
- 916 97. Nardini, A.; Pedà, G.; La Rocca, N. Trade-offs between leaf hydraulic capacity and
917 drought vulnerability: morpho-anatomical bases, carbon costs and ecological
918 consequences. *New Phytol.* **2012**, *196*, 788–798.

- 919 98. Aasamaa, K.; Niinemets, Ü.; Sôber, A. Leaf hydraulic conductance in relation to
 920 anatomical and functional traits during *Populus tremula* leaf ontogeny. *Tree Physiol.*
 921 **2005**, 25, 1409–1418.
- 922 99. Hacke, U.G.; Spicer, R.; Schreiber, S.G.; Plavcová, L. An ecophysiological and
 923 developmental perspective on variation in vessel diameter. *Plant Cell Environ.* **2017**,
 924 40(6), 831–845.
- 925 100. Spicer, R.; Schreiber, S.G.; Plavcová, L. An ecophysiological and
 926 developmental perspective on variation in vessel diameter. *Plant Cell Environ.* **2017**,
 927 40, 831–845.
- 928 101. Ivancich, H.S.; Lencinas, M.V.; Pastur, G.J.M.; Esteban, R.M.S.; Hernández,
 929 L.; Lindstrom, I. Foliar anatomical and morphological variation in *Nothofagus*
 930 *pumilio* seedlings under controlled irradiance and soil moisture levels. *Tree Physiol.*
 931 **2012**, 32(5), 554–564.
- 932 102. Schramm, R. Über die anatomischen Jugendformen der Blätter einheimischer
 933 Holzpflanzen. *Flora* **1912**, 104, 225–295.

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935 Supplementary Material

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937 Table 1. Target genes and efficiency of primers used during quantitative real-time PCR for
 938 the analysis of gene expression of leaves and needles.

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940 Captions of Tables and Figures

941

942 Table 1. Effect of irrigation treatments (100%, 75%, 50% or 25 % of the recommended dose)
 943 on height (h), diameter at root collar (d) and h/d ratio in *Fagus sylvatica*, *Quercus petraea*,
 944 *Abies alba* and *Pinus sylvestris* seedlings. Means ± S.E. are given (n = 40). One-way analysis
 945 of variance *F*-values with degrees of freedom in the lower index for treatment effect and error
 946 term and the associated *P* values are shown separately for each species (****P*<0.001, **
 947 *P*<0.01, * *P*<0.05, n.s. – not significant). Shared letters in a column indicate that the pre-
 948 planned contrasts between means are not significant.

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950 Fig. 1. Water contents (determined as percentage fresh mass) in leaves (LWC; a), roots (RWC,
 951 b) and stems (SWC; c) of *Fagus sylvatica*, *Quercus petraea*, *Abies alba* and *Pinus sylvestris*

952 seedlings grown at different irrigation treatments (100%, 75%, 50% and 25% of the
953 recommended dose). Bars indicate means \pm SE ($n=16$). One-way analysis of variance F -values
954 for treatment effect and the associated P values are shown separately for each species (* $P<0.05$,
955 n.s. – not significant). Shared letters above bars indicate that the pre-planned contrasts between
956 means are not significant.

957 Fig. 2. Total plant dry mass of (a) *Fagus sylvatica* and *Quercus petraea*, and (b) *Abies alba* and
958 *Pinus sylvestris* seedlings grown at different irrigation treatments (100%, 75%, 50% and 25%
959 of the recommended dose). Bars indicate means \pm SE ($n=16$). Note the different scales. One-
960 way analysis of variance F -values for treatment effect and the associated P values are shown
961 separately for each species (*** $P<0.001$, n.s. – not significant). Shared letters above bars
962 indicate that the pre-planned contrasts between means are not significant.

963 Fig. 3. Patterns of biomass allocation in *Fagus sylvatica*, *Quercus petraea*, *Abies alba* and *Pinus*
964 *sylvestris* seedlings grown at different irrigation treatments (100%, 75%, 50% and 25% of the
965 recommended dose): aboveground to belowground mass ratio (ABMR; a), leaf mass fraction
966 (LMF; b), root mass fraction (RMF; c) and stem mass fraction (SMF, d). Bars indicate means
967 \pm SE ($n=16$). One-way analysis of variance F -values for treatment effect and the associated P
968 values are shown separately for each species (*** $P<0.001$, ** $P<0.01$, * $P<0.05$, n.s. – not
969 significant). Shared letters above bars indicate that the pre-planned contrasts between means
970 are not significant.

971 Fig. 4. Leaf proline concentration of *Fagus sylvatica*, *Quercus petraea*, *Abies alba* and *Pinus*
972 *sylvestris* seedlings grown at different irrigation treatments (100%, 75%, 50% and 25% of the
973 recommended dose). Bars indicate means \pm SE ($n=16$). Results of a two-way analysis of
974 variance with species and irrigation treatments as the fixed-factors are given (F -values for
975 treatment effect and the associated P values are shown). Shared letters above bars indicate that
976 the pre-planned contrasts between means are not significant. P symbols are: *** $P<0.001$, **
977 $P<0.01$, * $P<0.05$, n.s. – not significant.

978 Fig. 5. Anatomical parameters in leaves of *Fagus sylvatica* and *Quercus petraea*, and needles
979 of *Abies alba* and *Pinus sylvestris* leaves in seedlings grown under different levels of irrigation
980 (100%, 50% and 25% of the recommended levels). Bars represent the mean (\pm S.E.) ($n = 8$, n –
981 number of seedlings per species and irrigation treatment). Anatomical features such as leaf
982 thickness (a), palisade cell length (b), transverse midvein phloem (c) and xylem area (d), and

983 within-leaf mean (e) and maximal (f) conduit diameter are shown. Results of a two-way analysis
984 of variance with species and irrigation treatments as the fixed-factors are given (F-values for
985 treatment effect and the associated P values are shown). Shared letters above bars indicate that
986 the differences between means was not significant. P symbols are: *** P <0.001, ** P <0.01, *
987 P <0.05, n.s. – not significant.

988 Fig. 6. Relative transcript abundance of select response-to-abiotic-stress genes in leaves of
989 *Fagus sylvatica*, *Quercus petraea*, and needles of *Abies alba* and *Pinus sylvestris* seedlings
990 grown under different levels of irrigation (100%, 75%, 50% and 25% of the recommended
991 level) (means±SE, $n = 12$, number of seedlings per species and irrigation treatment). One-way
992 analysis of variance F-values for treatment effect with degrees of freedom and the associated
993 P values are shown separately for each species (*** P <0.001, ** P <0.01, * P <0.05, n.s. – not
994 significant). Shared letters above bars indicate that differences between means were not
995 significant.

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1009 **Tables and figures**

1010 Table 1

Species	Irrigation treatment (% recommended dose)	h (mm)	d (mm)	h/d
<i>Fagus sylvatica</i>	25	104±5a	1.88±0.08a	57±2a
	50	124±5b	2.13±0.09b	59±2a
	75	185±7c	2.77±0.08c	68±3b
	100	225±11d	3.37±0.11d	67±3b
$F_{3,156}$		62.5***	51.4***	6.0**
<i>Quercus petraea</i>	25	82±3a	1.90±0.07a	44±2a
	50	94±3b	1.97±0.06a	48±2ab
	75	117±7c	2.48±0.08b	48±3ab
	100	148±8d	2.78±0.08c	54±3b
$F_{3,156}$		28.6***	35.8***	3.2*
<i>Abies alba</i>	25	35±1a	1.00±0.03a	36±1a
	50	37±1a	1.05±0.03a	36±1a
	75	38±1a	1.03±0.03a	38±1a
	100	38±1a	1.00±0.02a	38±1a
$F_{3,156}$		n.s.	n.s.	n.s.
<i>Pinus sylvestris</i>	25	35±1a	1.01±0.03a	35±1a
	50	39±1b	1.06±0.03a	37±1a
	75	48±1c	1.17±0.04b	43±1b
	100	51±2c	1.18±0.04b	45±2b
$F_{3,156}$		31.9***	5.4***	11.9***

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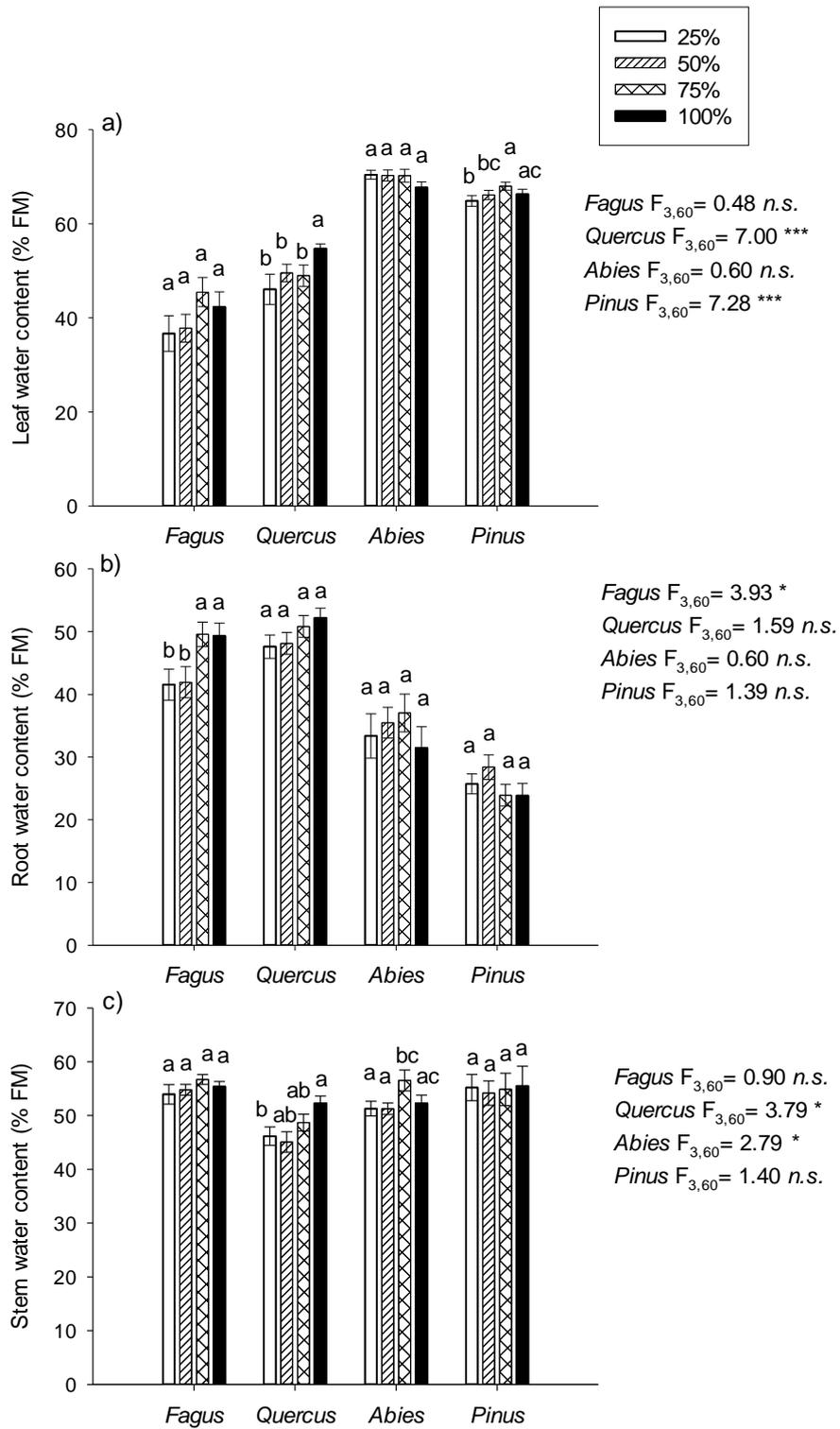
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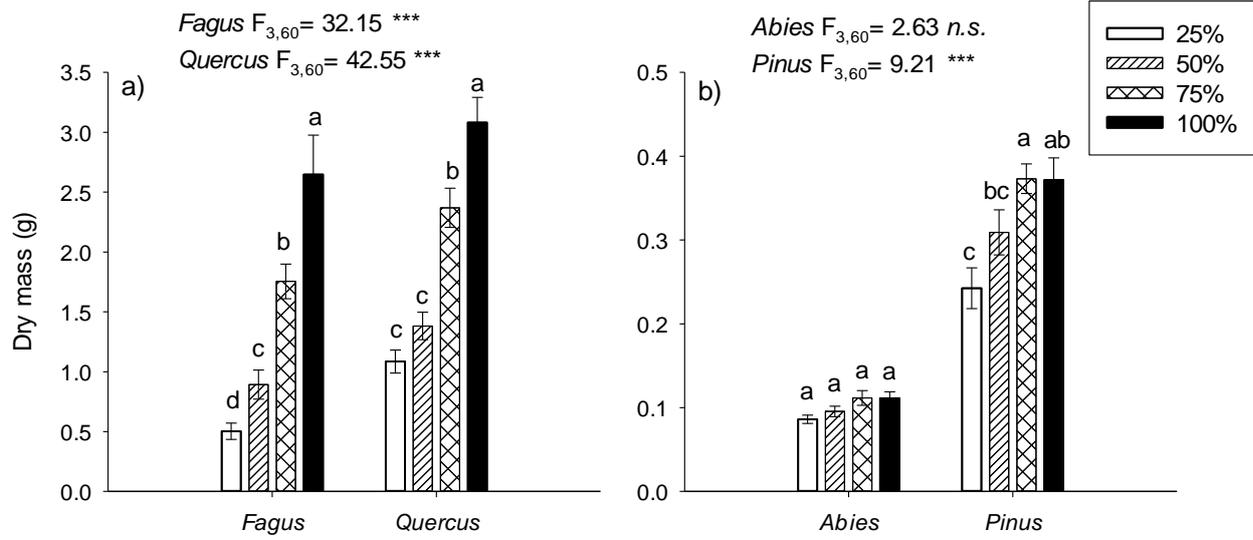


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1024 Figure 1

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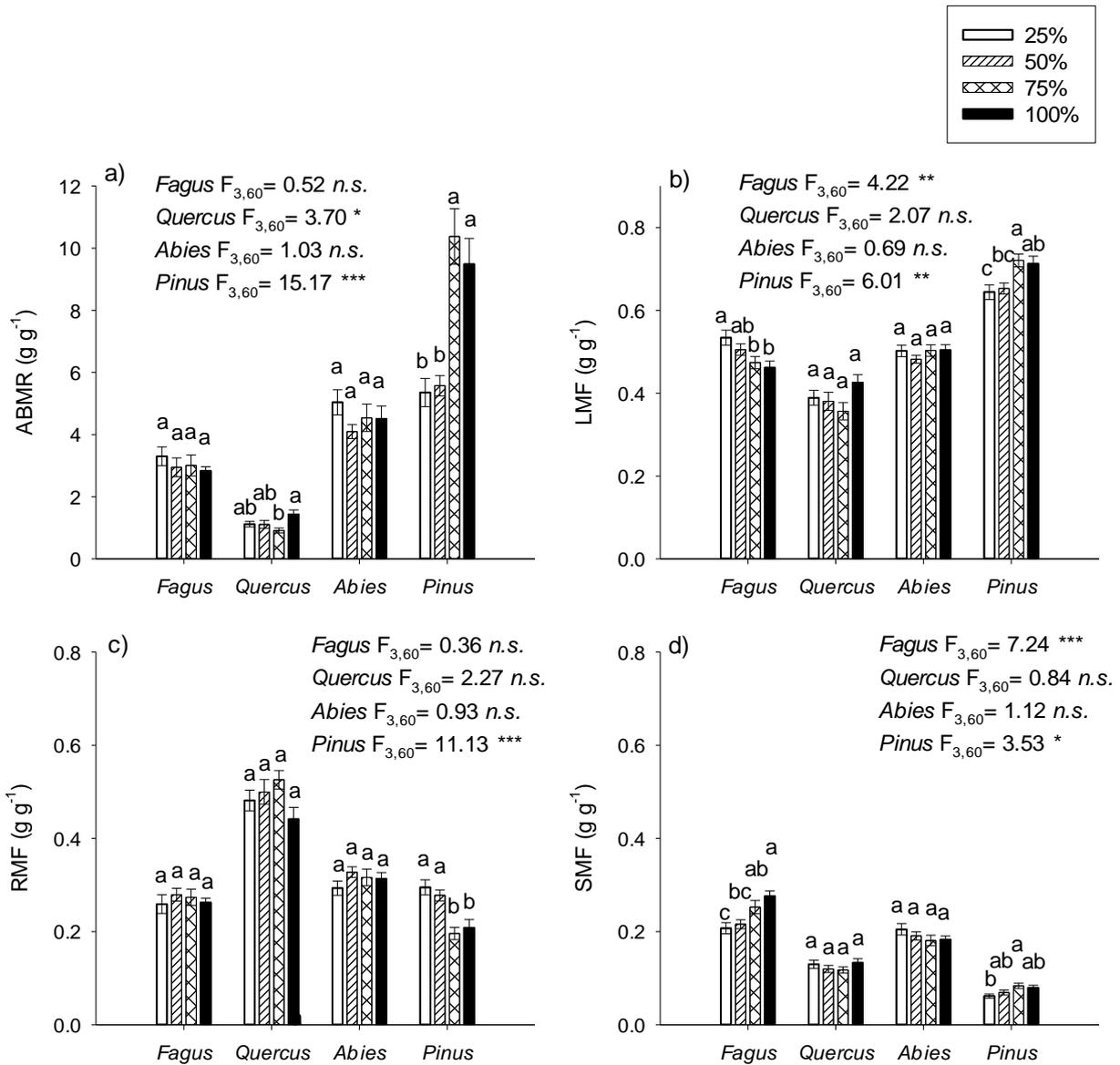


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1028 Figure 2

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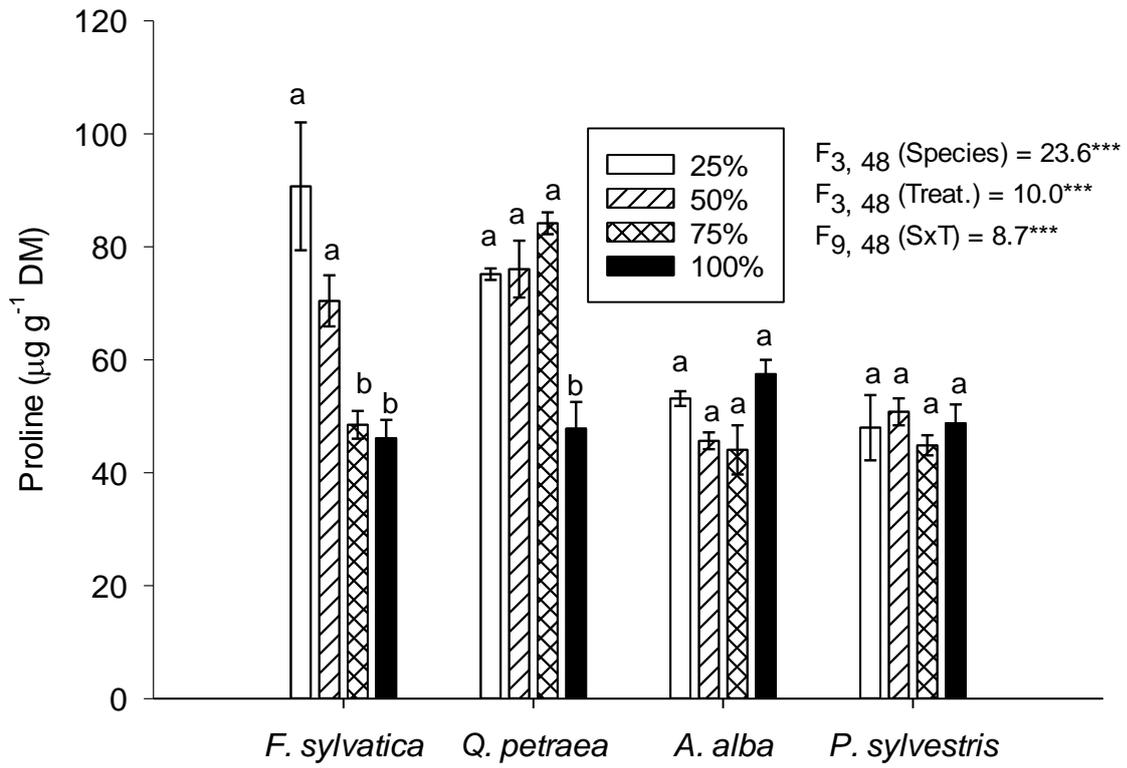
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1032 Figure 3

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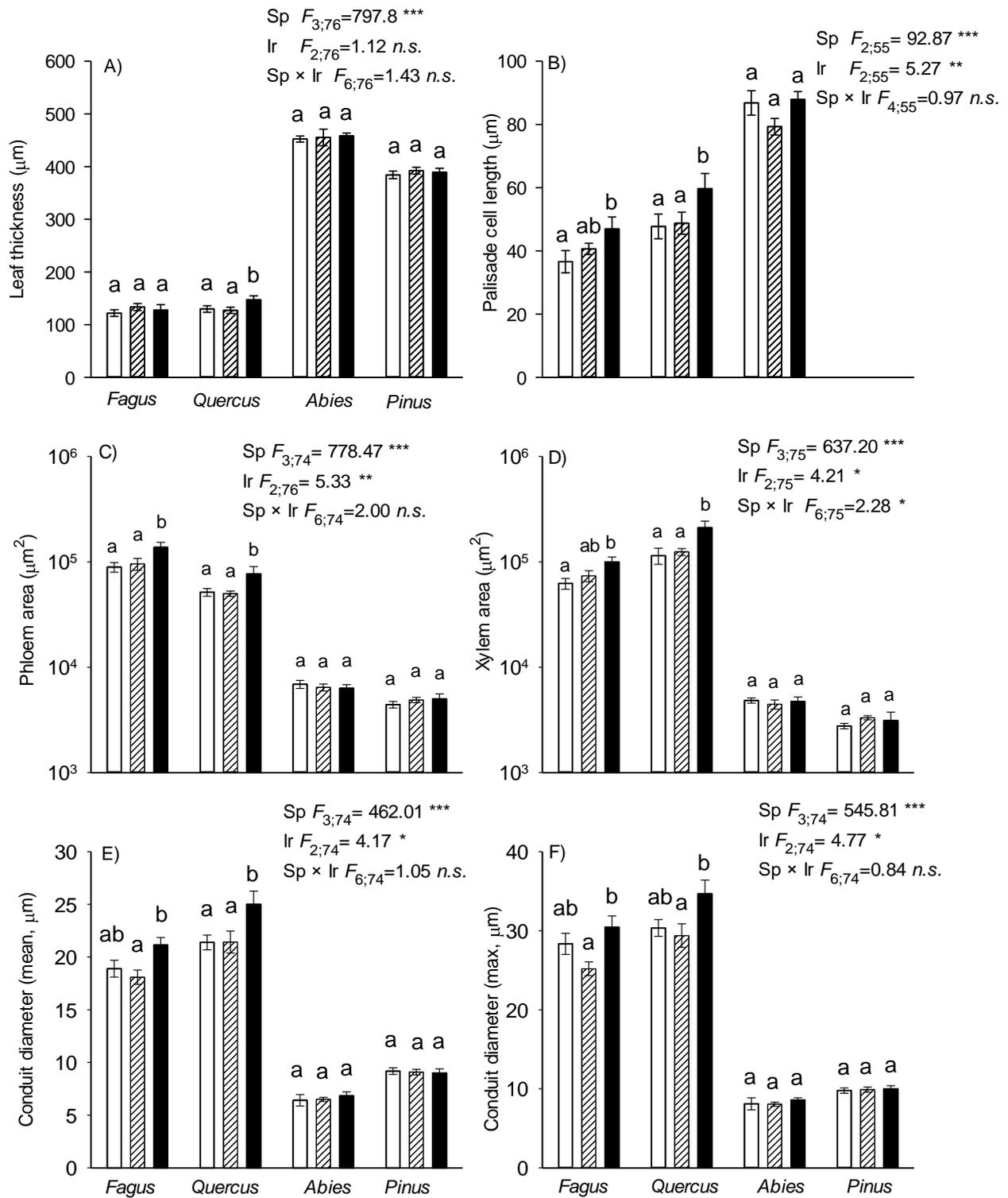
1038 Figure 4

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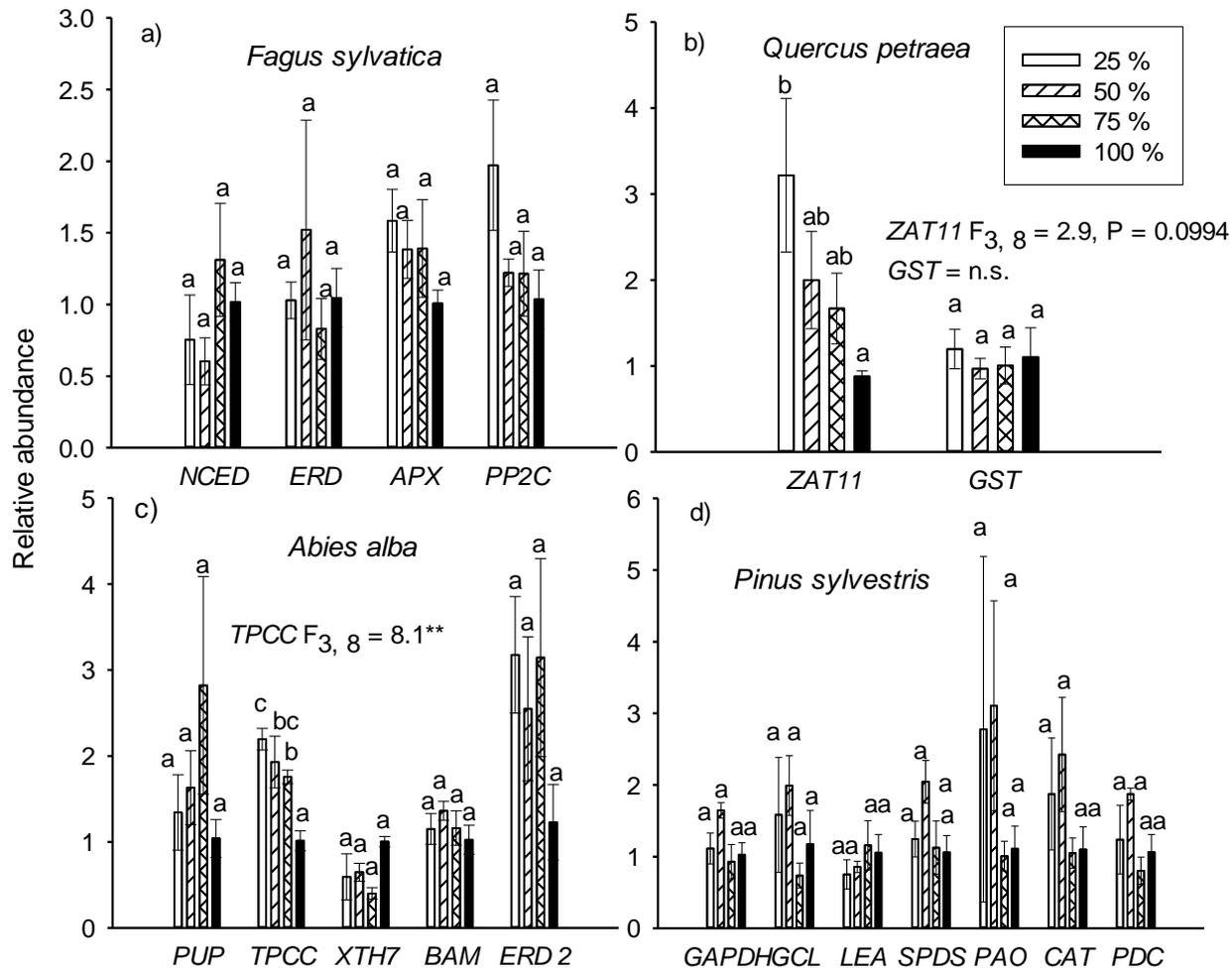
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1044 Figure 5

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1048 Figure 6