FABRICATION AND CHARACTERIZATION OF LIGNIN/DENDRIMER ELECTROSPUN BLEND MATS

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Abstract: Blending lignin as the second most abundant polymer in nature with nanostructured compounds such as dendritic polymers will not only add value to lignin, but also increase its application in various fields. In this study, softwood Kraft lignin/polyamidoamine dendritic polymer (PAMAM) blends were fabricated by solution electrospinning method to produce bead-free nanofiber mats. The mats were characterized by scanning electron microscopy (SEM), contact angle measurement, Fourier transform infrared (FTIR), zeta potential, and thermogravimetry analysis (TGA). The chemical intermolecular interactions between lignin functional groups and abundant amino groups in PAMAM were investigated by FTIR and viscosity measurement. These interactions enhanced the mechanical and thermal characteristics of lignin/PAMAM mats, providing further potential applications at industry level.
1. Introduction

Lignin is one of the complex natural polymers that mostly exists in the by-product of plants, woods, pulp, and paper [1, 2]. It is a biopolymer with an irregular network which has made up three main units linked together with carbon-carbon bonds [3]. Lignin contains an aromatic structure with a phenolic ring in its precursors’ monomer that mainly classified into three monomer units of coniferyl alcohol (G), p-coumaryl alcohol (H), and sinapyl alcohol (S) [4-7]. Softwood lignin existed in gymnosperm plants commonly consists of 24-33% of the cell wall [1], of which nearly 95% is G-lignin [8]. The various structure of functional groups in lignin comes from the type of plant and extraction methods [9]. The number-average molecular weight (Mn) between 3000 and 10000 and weight-average molecular weight (Mw) of 8000 to 80000 are very common for lignin depending on its type and other effective parameters [10]. Besides various structure and type of lignin products, they reveal some characteristics, such as antioxidant, antiseptic, bactericide, bacteriostatic, and disinfectant agent; whilst, its non-cytotoxicity toward human cells [11, 12]. These characteristics increase the productions and demands on lignin based products. For instance, in 1998, only 1% of lignin was isolated and sold [13]; whilst, in 2017, there were almost 70 million tons of wood pulping operation in worldwide of which 30 wt.% was extracted lignin [14].

One of the advantages of using lignin products is its renewable resource of carbon due to phenolic structure [15]. Thus, a wide range of lignin based products in the form of powders and/or fibers utilized as additives in composite products [1, 16]. The mechanical and thermal properties of the composite structures can improve when lignin is grafted or blended with other polymers, such as polypropylene (PP) as a cross-linker [17], polybutadiene (PB) and PP as an antioxidant [18], polyaniline as a conductivity modifier [19] and starch materials as a hydrophobic co-plasticizer [20]. However, much remains to research to find out the higher ability and new applications for lignin [21]. Lignin has also used as an anti-UV Stabilizer due to its phenolic and other organic structure and three-dimensional network of hydroxyl groups [22]. Besides on its novel characteristics and structure, lignin has also disadvantages, such as hydrophobic behavior [23], small particle size and variation in adhesion properties for coating composite structures, and extensive thermal behavior and Tg range because of its complicated structures [24], and content condensation during extraction and production [25]. Due to these disadvantages, researchers have aimed to improve lignin characteristic by surface modifications or bulk modifications [26]. One of the novel techniques for modification of lignin-based products is blending with other polymers, such as dendritic structure polymers.

Dendritic structures have high potentials due to their special design, novel characteristics particularly host-guest properties, and nano dimension structures. They have formed from small repeating units, named generation numbers, onto the a central core to which other branches link and make a macromolecular structure with novel structure and characteristics [27]. The most outstanding characteristic, host-guest properties, comes from large numbers of functional reactive end-groups as well as hollow interior cavities between branches. The unique
characteristic of dendrimers is their branched topology and multifunctional reactive groups [28]. One of the well-known dendrimers is polyamidoamine (PAMAM) dendrimer which provides amine-terminated functional groups with various kinds of applications. In this paper, PAMAM was blended with softwood kraft lignin and electrospun fibers to achieve new improvements for the first time.

Electrospinning is an efficient method for fabricating fibers in nano and micro-scale [29]. Thus, it is believed that electrospinning of lignin with other polymers can provide opportunities to achieve new characterizations resulting new applications [30]. For instances, electrospun lignin can be used for carbon fiber production [31-34], filtration membranes [35], biorefinery processes [36], biomedical and drug delivery applications [37], and wood panel products [38]. Various parameters, such as surface tension, viscosity, and electrical conductivity are affected on to the lignin nanofiber production [39]. Due to the chemical structure and properties of lignin, it has been blended with other polymers, such as polyethylene oxide (PEO) [40], polyvinyl alcohol (PVA) [41], polyacrylonitrile (PAN) [32], and polyethylene terephthalate (PET) [42]. Blended polymers help lignin to become spinnable fiber due to their good miscibility.

In order to improve the mechanical properties of electrospun lignin fiber particularly due to swelling in water, the fiber is heat-set or thermo stabilized [43]. In heat-setting process physical property changes like the color of lignin mats, which transforms from light yellow to dark brown, and the chemical reactions such depolymerization, recondensation would occur after increasing the temperature [44]. Lignin has also potential to participate in condensation reaction with fragments and side chain cleavage of ether linkage occur [43]. Besides, decreasing phenolic bond and forming new reactions would produce higher molecular weight in heat-setting processes; While, in stabilization process there are higher temperatures than heat-set process in order to achieve a sample with high strength and modulus [44].

The aim of this paper is to evaluate the fabrication and characterization of a novel blending between PAMAM dendritic structure and softwood kraft lignin (SKL). Furthermore, the heat-setting and stabilization process of SKL nanofibers are analyzed in the presence of PAMAM polymers. The interaction between lignin and PAMAM dendrimers are studied by FTIR, TGA, mechanical properties, surface charge, and viscosity measurement. In addition, the morphology of all samples is determined by SEM analysis. Overall, the results to follow confirmed the novel characteristics of lignin/PAMAM electrospun fibers which would make it a good candidate for further applications.

2. Results and Discussion

2.1 Morphology analyses

The SEM images of electrospun lignin, lignin/PAMAM (1wt%) and lignin/PAMAM (2wt%) showed in Fig. 1. It must be noted that after increasing the concentration of dendrimer from 1% to 2wt%, it is not possible to continue the process of electrospinning. The optimum concentration
of PAMAM in this blending was 1% and other experiments applied on lignin/PAMAM with 1wt% of PAMAM. As a result, the abbreviation of lignin/PAMAM in all text means 1wt% of PAMAM based on the lignin weight. By increasing the PAMAM concentration, the viscosity of the main solution reduced. Thus, the fiber fabrication is impossible. This behavior for other polymers has been also elucidated by other investigators [45].

(a) (b) (c)

Figure 1. SEM of (a) lignin; (b) lignin/PAMAM 1wt%; (c) lignin/PAMAM 2wt%

The fiber diameters of lignin and lignin/PAMAM were measured before and after thermal-treatment (Heat-set) using SEM, as illustrated in Fig. 2. It is observed that the average diameter of the lignin/PAMAM nanofibers in both heat-set and not heat-set samples was less than the pristine lignin nanofibers due to the reduction of surface tension in electrospinning procedure by cationic charges of PAMAM polymers. PAMAM could act as a poly-electrolyte and reduce the diameter of electrospun fiber [46]. In addition, chemical interaction between cationic functional groups of PAMAM and anionic hydroxyl groups in lignin could be another reason for decreasing at about 10 percentage of the fiber diameter. On the other hand, as shown in Table 1, the total average diameter of both lignin and lignin/PAMAM nanofibers has been reduced after heat-setting process due to dehydration and possible reaction of the hydroxyl group in lignin and lignin/PAMAM. This diameter reduction after heat-setting processes can be as a result of the depolymerization, recondensation processes [44].
Figure 2. SEM of (a) lignin, (b) lignin/PAMAM, (c) heat-set lignin, (d) heat-set lignin/PAMAM, along with the corresponding fibre diameter distributions.

Fig. 3 reveals the SEM images of lignin and lignin/PAMAM after thermostabilization processes (TSP). The TSP commonly used to prepare the carbon nanofibers (CNF) [47]. In this processes, higher temperature comparing to heat-setting processes causes more diameter reduction. The condensation procedure between amino terminated groups of PAMAM (−NH₂) and hydroxyl groups lignin (−OH) not only reduce the fiber diameter but also make a strong interaction between dendritic groups and lignin structure.

Figure 3. SEM of (a)Lignin Stabilized, (b)Lignin/PAMAM Stabilized

Table 1. The average fiber diameter of electrospun lignin and lignin/PAMAM at the state of untreated, heat-setting, and stabilized in nanometer (nm).

<table>
<thead>
<tr>
<th>Sample</th>
<th>Untreated (nm)</th>
<th>Heat-set (nm)</th>
<th>Stabilized (nm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lignin</td>
<td>1432±62</td>
<td>940±39</td>
<td>622±21</td>
</tr>
<tr>
<td>Lignin/PAMAM</td>
<td>1278±26</td>
<td>909±41</td>
<td>866±29</td>
</tr>
</tbody>
</table>
2.2 Mechanical properties

Mechanical properties of electrospun mats with and without PAMAM before and after heat-setting processes were evaluated. According to the heat treatment condition in each part of the process, as shown in Table 2, the maximum stress at yield point (σ_y) and elastic moduli (E) of the both lignin heat-set and lignin/PAMAM heat-set nanofibers reduced. While some researchers revealed tenacity enhancement of lignin electrospun fiber after heat-setting procedure due to the more cross-linking and chemical interaction between molecular chains in lignin based polymers [43, 44], other researchers elucidated tenacity reduction because of becoming more brittle [48]. It seems the process conditions, such as heating temperature, time of heating, and applied pressure have direct effect on the mechanical properties. However, almost all investigators have agreed that the heat-setting improve durability, biological behavior, and the dimensional stability [49].

Although there was an approximately sharp drop in the stress-strain curve in heat-setting processes, the effect of PAMAM on both heat-set and untreated mats was significant. As shown in Table 2, both (σ_y) and (E) increased. As a result, the fibrous mats of lignin/dendrimer were stronger the pristine samples and elastic recovery in both blended sample improved completely. The phenolic compounds and hydroxyl groups in lignin chemical structure react with functional groups of PAMAM; thus, the blended mats were much stronger and could tolerate more stress at high tension. The strong interaction between amino terminated groups of PAMAM (−NH₂) in dendritic groups and hydroxyl groups in lignin(−OH) is the main reason for increasing the stress at yield and elastic moduli for lignin/PAMAM comparing to lignin electrospun fiber mats.

<table>
<thead>
<tr>
<th></th>
<th>untreated lignin nanofiber mats</th>
<th>heat-set lignin nanofiber mats</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Lignin</td>
<td>Lignin/PAMAM</td>
</tr>
<tr>
<td>Stress at yield (σ_y) (MPa)</td>
<td>4.52±0.97</td>
<td>6.30±1.70</td>
</tr>
<tr>
<td>Strain(%)</td>
<td>5.68±1.54</td>
<td>6.73±2.67</td>
</tr>
<tr>
<td>Elastic moduli (E) (MPa)</td>
<td>79.66±1.01</td>
<td>93.61±1.73</td>
</tr>
</tbody>
</table>

2.3 Surface charge

Zeta potential can provide important information about surface chemistry and surface charge of electrospun mats, which is useful for various applications, such as water treatment, pharmaceutical, environmental protection, paint industry, and other applications [50]. Zeta potential describes the electrical charge and interactions of small particles at the surface which are dispersed in a material and can indicate the physical stability of the system in different states.
In addition, zeta potential can assist to reduce surface energy and optimize many related processes [51, 52].

In this research, lignin nanofiber mats were blended with PAMAM and the surface electrical charge was measured with the specific device for measuring surface charge of electrospun mats. The surface electrical charge of lignin is completely negative due to abundant hydroxyl and phenolic compounds in its structure as well as hydrogenation and decarbonylation alkyl groups [53]. While, the NH₃⁺ groups in PAMAM reveals the positive zeta potential. The zeta potential of lignin was measured in the range of -23.28 mV, while it increases to -20.72 mV after adding 1% PAMAM based on the lignin weight. This increasing data confirms that only 1wt% of PAMAM lead to reacts between amine groups in PAMAM with some negative hydroxyl groups of lignin by more electrostatic interaction between lignin and PAMAM leading to high elastic moduli in tensile test for lignin/PAMAM mats comparing to lignin ones. Dallmeyer et. al [34] also revealed that inter-fiber bonding significantly enhanced the mechanical properties of the electrospun fabrics.

2.4 Thermogravimetric analysis (TGA)

The Thermogravimetry (TG) or Thermogravimetric Analysis (TGA) and Derivative thermogravimetry (DTG) curves of lignin and lignin/PAMAM before and after heat treatment (heat-set) are shown in Fig. 4a and b, respectively. The main characteristic and important peaks and information from these analyses are illustrated in Table 4. Lignin's main mass loss peak (according to its type and extraction) commonly appears at about 370-395 °C. While, the first peak appear in range of 150-270 °C corresponding to phenolic compounds of lignin [54]. The weight loss before 150 °C is due to the evaporation of water and small amount of dehydration reaction. At 380°C, there is a peak related to hydrogenation of methoxy groups on the aromatic ring and consequently the evolution of methanol. Methane is another product that can release which starts at 350 °C and ends in 420-430 °C. Furthermore, carbon dioxide (CO₂) could release from 200 °C to 640-690 °C. Carbon monoxide (CO) also releases from 220 °C to 790 °C due to decarbonylation react of alkyl side-chain carbonyl terminate and the reaction of formaldehyde with free-radical coupling reactors in lignin [55, 56].

The results revealed that the weight of residue left by the lignin/PAMAM blended fiber mats was higher than that of pristine lignin in both untreated and heat-set (HS), as shown in Table 3 and also Fig. 4. The interaction between amine groups in PAMAM and hydroxyl groups in lignin caused more resistant to high temperature. This indicates that lignin/PAMAM is more thermally stable than pristine lignin. In Table 4, the comparison between the first and second notable peaks in DTG has given indicates the positive effect of dendrimer and its rich functional group on thermal stability. This effect confirmed that using dendrimer in lignin mats could be a sufficient method for the production of carbon fiber from lignin and also high mechanical properties.
Fig. 4. The TG (a) and DTG (b) curves of the lignin and lignin/PAMAM untreated and heat-set state (HS).
Table 3. The characteristic points of TG and DTG curve at two main peaks.

<table>
<thead>
<tr>
<th>Component</th>
<th>$T_d^a$ (°C)</th>
<th>%weight</th>
<th>$\alpha_1^b$</th>
<th>$\alpha_2^c$</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lignin</td>
<td>595.53</td>
<td>41.12</td>
<td>0.374</td>
<td>0.029</td>
</tr>
<tr>
<td>Lignin/PAMAM</td>
<td>594.32</td>
<td>43.2</td>
<td>0.345</td>
<td>0.038</td>
</tr>
<tr>
<td>Lignin (HS)</td>
<td>596.20</td>
<td>41.69</td>
<td>0.369</td>
<td>0.024</td>
</tr>
<tr>
<td>Lignin/PAMAM (HS)</td>
<td>589.22</td>
<td>45.54</td>
<td>0.332</td>
<td>0.036</td>
</tr>
</tbody>
</table>

a The decomposition temperature.

b The derivative weight (%wt./C) at 66% weight.

c The derivative weight (%wt./C) at 94% weight.

2.5 Fourier transform infrared spectroscopy (FTIR)

Lignin has lots of functional groups due to its many different types of extraction and utilization. Therefore, it is often complicated to characterize all lignin and lignin based samples. As shown in Fig. 5, the important characteristic bands of lignin are revealed in Table 4. The peak at 1429-1509 and 1601 cm$^{-1}$ is related to C-C aromatic rings [57]. The lignin’s alcohol picks appeared different bands (at 1260-1270 and 1330-1375 cm$^{-1}$) related to the guaiacyl and syringyl, respectively. In addition, the carbonyl group is seen at 1700-1715 cm$^{-1}$ [58]. The absorption bands of water appear in a range of 1300-1800 cm$^{-1}$ and 3500-3964 cm$^{-1}$. The existence of CO is elucidated at 2112 and 2180 cm$^{-1}$, while CO$_2$ appeared at 2217-2391 cm$^{-1}$ [56]. The most notable band in lignin is O–H due to the presence of the alcoholic and phenolic structure of lignin at 1300-1400 cm$^{-1}$. The absorption bands at 2850-3200 cm$^{-1}$ indicate the hydrocarbons, such as methane [54]. On the other hand, PAMAM, which is rich in amine terminated functional group, produce pick bands at 1550-1640 cm$^{-1}$. The band at 3100-3500 cm$^{-1}$ is elucidated the amino groups of PAMAM. At 1630-1680 cm$^{-1}$ and these absorptions appeared in FTIR spectroscopy of lignin/PAMAM (Fig. 5) which is a proof to the reaction between lignin and PAMAM. As it has shown in Fig.5 (C), a new peak illustrated by No.4 confirm the reaction of amine group of PAMAM and hydroxyl group of lignin at 1485 cm$^{-1}$[59]. Moreover, there are characteristic peaks of lignin in Fig.5: (No.1) which indicates hydroxyl group at 1650 cm$^{-1}$, (No.2) that appear at 1600 cm$^{-1}$ according to aromatic rings, (No.3) which reveals at 1520 cm$^{-1}$ indicates C=C bonds and derivatives [58].
Table 4. Main contents of different lignin samples and their range of evolution detected by FTIR.

<table>
<thead>
<tr>
<th>Wavenumber (cm⁻¹)</th>
<th>Functional group</th>
<th>Vibration</th>
<th>Compounds</th>
<th>Refs.</th>
</tr>
</thead>
<tbody>
<tr>
<td>645-690</td>
<td>C=O</td>
<td>Dactyl-zone</td>
<td></td>
<td>[55, 58]</td>
</tr>
<tr>
<td>974-1058</td>
<td>C–O</td>
<td>Stretching</td>
<td>R–OH</td>
<td>[55, 57]</td>
</tr>
<tr>
<td>1000-1300</td>
<td>C–O</td>
<td>Stretching</td>
<td></td>
<td>[54, 58]</td>
</tr>
<tr>
<td>1093-1188</td>
<td>C–C</td>
<td>Skeleton</td>
<td></td>
<td>[55]</td>
</tr>
<tr>
<td>1310-1365</td>
<td>C–CH₃</td>
<td>Bending</td>
<td>Alkyls</td>
<td>[55, 58]</td>
</tr>
<tr>
<td>1300-1400</td>
<td>O–H</td>
<td>Bending</td>
<td></td>
<td>[56, 57]</td>
</tr>
<tr>
<td>1513</td>
<td>C=C–OH</td>
<td>Stretching</td>
<td></td>
<td>[58]</td>
</tr>
<tr>
<td>1613</td>
<td>C=C</td>
<td>Stretching</td>
<td>Aromatics</td>
<td>[58]</td>
</tr>
<tr>
<td>1645-1750</td>
<td>O–H</td>
<td>Bending</td>
<td>H₂O</td>
<td>[55]</td>
</tr>
<tr>
<td>2020-2220</td>
<td>C–O</td>
<td>Stretching</td>
<td>CO</td>
<td>[55]</td>
</tr>
<tr>
<td>2210-2390</td>
<td>C=O</td>
<td>Stretching</td>
<td>CO₂</td>
<td>[55, 58]</td>
</tr>
<tr>
<td>2750-2990</td>
<td>C–H₂</td>
<td>Asymmetric Stretching</td>
<td></td>
<td>[54, 55]</td>
</tr>
<tr>
<td>2904-2979</td>
<td>C–H</td>
<td>Stretching</td>
<td></td>
<td>[57]</td>
</tr>
<tr>
<td>2990-3010</td>
<td>C–H</td>
<td>Stretching</td>
<td></td>
<td>[54, 57]</td>
</tr>
<tr>
<td>3020-3190</td>
<td>C–H</td>
<td>Stretching</td>
<td>CH₄</td>
<td>[56, 57]</td>
</tr>
<tr>
<td>3500-3600</td>
<td>O–H</td>
<td>Stretching</td>
<td></td>
<td>[54, 57]</td>
</tr>
<tr>
<td>3823-3870</td>
<td>O–H</td>
<td>Stretching</td>
<td></td>
<td>[55, 57]</td>
</tr>
</tbody>
</table>

Fig. 5. FTIR spectra of (a) lignin (b) lignin/PAMAM 1wt% (c) lignin/PAMAM 2wt%.
2.6. Viscosity

To reveal the interaction of lignin with PAMAM, lignin and lignin/PAMAM solutions were prepared and their viscosity measured in corresponding to time. Furthermore, due to the relationships between viscosity measurement and fiber diameter, the SEM images of the electrospun fibers after 5 days were also investigated. As general rules in polymeric solution, the viscosity would be increased subsequently by increasing more entanglement between polymeric chains, while it is reduced due to the less entanglement [60]. The viscosity of lignin and lignin/PAMAM in DMF solvent in 5 consecutive days were elucidated in Fig. 6. The result reveals that the viscosity for both lignin and lignin/PAMAM show increasing trend and then decreased. It seems that the unique structure of lignin, which has rich in phenolic rings and hydroxyl groups, cause the depolymerization of lignin in DMF. The structure causes higher viscosity. The viscosity of lignin just like other polymer reduced finally due to solution transition mechanism and reduction rate in radical polymerization in DMF due to the transfer reaction to solvent [61]. Also, the rate of free radical polymerization mechanism increases due to the radicals which react with lignin functional group and observed the increasing viscosity of lignin at first three days. After three days, the viscosity reduced due to the chain transfer to solvent. The viscosity measurement results in lignin/PAMAM solution is interesting due to the increase of solution viscosity. The interaction between PAMAM and lignin made the solution denser due to more molecular chain entanglement. The highest viscosities were 72 and 76.6 mPa.s for lignin and lignin/PAMAM, respectively.

![Fig. 6. The viscosity of lignin and lignin/PAMAM through 5 days.](image-url)
2.7 Reaction mechanism between lignin and amine-terminated dendrimer

Despite of attempts made to define the various structural elements of lignin, the clear chemical structure of softwood kraft lignin (SKL) is still under investigation due to the complexity of various type [62]. Even though the most quantity of define structure belongs to the methoxy groups, SKL has various inter-unit bonds, such as Arylglycerol-β-aryl ethers, Phenyl coumaran, Pinoresinols, Lignin carbohydrate α-benzyl ethers, Stilbenes, Aryl enol ethers, Secoisolariciresinols, and end groups, such as Cinnamyl alcohols, Arvlacetic acid, Aryl-hydroxy-acetic acid, Aryl ethyl ketones, Aryl propanols, Aryl hydroxyethyl ketone, Aromatic aldehydes, Cinnamyl aldehydes, and functional groups, such as Aromatic C–H, Methoxy, Quinones, and also hydroxyl groups, such as Carboxylic OH, Aliphatic OH, o-Disubstituted phenols (including o-substituted catechols), o-Monosubstituted phenols, and phenolic OH were figured out in SKL with different percentages [62]. It is well-known that, the amination reaction of lignin is regularly gone through Mannich reaction with amino group and formaldehyde [63]. In the absent of formaldehyde in lignin/PAMAM solutions, the reaction between PAMAM and main segment of lignin (methoxy groups) is carried out with hydrogen bonding and ionic interaction between hydroxyl group in lignin and amino terminated group in PAMAM. The regular reaction has been occurred between R-OH and R’-NH₂ which R and R’ coming from the substitutions of lignin and PAMAM, respectively. The results of these interactions confirm by new peaks in FTIR, strength enhancement, and particularly viscosity after one week results in previous section.
Since it is very complicated to demonstrate the reaction between wood kraft lignin and PAMAM, the chemical interaction between Guaiacyl (G) lignin and lower generation, 1st generation, of PAMAM have been illustrated in Fig. 8.

![Diagram of possible ionic and hydrogen bonding between lignin and PAMAM](image)

**Fig. 8.** The possible ionic and hydrogen bonding between lignin and PAMAM.

3. Materials and Methods

3.1. Materials

Softwood Kraft Lignin (SKL) was supplied by FP Innovations (Vancouver, BC) and used with no further purification. N, N- Dimethylformamide (DMF, 99.9%), Polyethylene Oxide (PEO) (M_w = 900 kDalton) was purchased from Aldrich. polyamidoamine (PAMAM) dendritic polymers was donated by Delta Innovative Company Poland.

3.2 Preparation and Properties of Electrospinning Solutions
Electrospinning solutions were prepared by dissolving of exact amount of SKL into the DMF solvent in the presence of 1 wt. % PEO based on the lignin powder. For lignin/PAMAM samples, first 1 and 2 wt. % of PAMAM based on the lignin powder were dissolved in DMF solvent, while lignin and PEO separately dissolved in DMF solvent. After 2 hours mixing, these two solution mixing together and continue for another 1 hour before electrospinning. The solution concentration for all samples was 30 wt%. The electrospinning procedure has been applied in electrospinning distance of 15 cm at a point of an applied voltage range of 17kV and the 0.01 mL/min pump speed.

3.2.1. Heat-setting and stabilization procedure

After drying the mats at room temperature for 10 hours, for preparing the heat-set samples, the samples were heated in oven at air atmosphere to 150°C for 120 minutes. While, for stabilization procedure, the samples were heated at 4°C/min heating rate to 250°C for 90 minutes in a gas chromatography oven (Hewlett Packard 5890 Series II).

3.3 Characterization of Composite Membranes

3.3.1 Morphology measurement

The morphology of the electrospun nanofibrous mats was observed by a scanning electron microscope (SEM) (Hitachi S-2300) with an acceleration voltage of 5-20 kV. All of the samples were gold coated before SEM observation. The average diameter of nanofibers was determined by analyzing the SEM images via Image J software.

3.3.2. Mechanical Properties

The mechanical properties of the electrospun mats were evaluated using a multipurpose micro-tensile tester (KES-G1, Kato-Tech Co. Ltd) with a 5 kg capacity force transducer. Strip-specimens (0.5 cm W×5 cm L) were tested with 5 replications for each sample at a cross-head speed of 0.2 cm/s. The strain was calculated by dividing the displacement by the gauge length. Stress was obtained using the following formula:

\[
\sigma_{\text{specific}} = \text{Specific Stress (N/Tex)} = \frac{\text{Force(N)}}{\text{Width of specimen (mm)}} \times \frac{\text{Areal density of specimen (g/sq.M)}}{
\]

1N/tex = 145,000 psi = 1 GPa

Load cell setting was g/10v, deformation rate was set at 0.02 cm/s, the specimen length was 5 cm, and the mass of specimen was calculated separately for each sample. The engineering stress \( \sigma_{\text{Eng}} \) was achieved from the equations below (where \( V \) represent the signal voltage read through the machine):

\[
\text{Deformation(cm)} = \text{Deformation rate} \times \Delta t
\]

\[
\text{Strain} \left( \frac{\text{cm}}{\text{cm}} \right) = \frac{\text{deformation}}{\text{specimen length}}
\]
Load(g) = Load cell setting × ΔV

Areal density was calculated as:

\[
\text{Areal Density} \left( \frac{g}{m^2} \right) = \frac{\text{Sample Mass}(g)}{\text{length}(m) \times \text{width}(m)},
\]

\[
\sigma_{\text{specific}} \left( \frac{g}{te\text{x}} \right) = \frac{\text{load}(g)}{\text{Areal Density} \left( \frac{g}{m^2} \right) \times \text{Width(mm)}}
\]

Converting the specific stress to engineering stress can be achieved by:

\[
\sigma_{\text{Eng}} (\text{MPa}) = \sigma_{\text{specific}} \left( \frac{g}{te\text{x}} \right) \times 9.81 \times \rho_{\text{polymer}} \left( g/cm^3 \right)
\]

3.3.3. Surface charge

The Surface Zeta Potential (SZP) electrode was used for analyzing the charge on a fibrous mats surface by NanoBrook instrument Brookhaven Instruments Corporation 750 Blue Point Road, Holtsville, NY 11742. Measurements were accomplished by attaching the sample to a horizontal place situated perpendicularly between two electrodes. The sample and electrodes were placed into a diluent containing probe particles for which the charge is well-known.

3.3.4. Thermogravimetric analysis (TGA)

Thermogravimetry is a method to recognize the thermal resistance of the sample and its components to obtain useful properties like oxidative degradation, dissolution, decomposition [64]. Thermogravimetric analysis (TGA) was carried out on a thermogravimetric analyzer (Q500, TA Instruments, USA). Samples were heated at 20 °C/min from room temperature to 600 °C in a dynamic nitrogen atmosphere (flow rate = 60 ml/min).

3.3.5 Fourier transform infrared spectroscopy (FTIR)

Fourier transform infrared spectroscopy (FTIR) can provide important information on the molecular structure of components and has been used mainly for chemical characterization of geological samples recently [65-67]. The mechanism of the FTIR technique is related to transitions between quantized vibrational energy states[65, 68]. The FTIR spectra were recorded over the range 400-4000cm\(^{-1}\) with 64 scans and the resolution of 4 cm\(^{-1}\).

3.3.6 Viscosity measurement

The solution viscosity of lignin for electrospinning is an essential factor that should be considered and controlled. The concentration of lignin in the solution and time are the main two elements that affect the viscosit [69, 70]. In this study, dimethylformamide (DMF) was the solvent, and all experiments were applied on lignin/DMF and lignin/PAMAM/DMF solution. The effect of time
the mixing on the viscosity measurement through 5 days was determined. The viscosity reported by the torsional oscillation-type viscometer, SEKONIK corporation, VM 10A series Nerima-ku, Tokyo, 178-8686 Japan.

4. Conclusions

The blending of softwood kraft lignin and PAMAM dendritic polymers, and electrospinning of the resulting blends were demonstrated in this paper. Heat-setting and stabilization procedure were applied to both lignin and lignin/PAMAM mats. The chemical interaction between hydroxyl groups in lignin-based mats and amino functional groups in PAMAM dendritic structures were systematically confirmed by FTIR and viscosity measurement. Increasing surface charge of lignin/PAMAM was another evidence for the demonstrating interaction between cationic amino groups in PAMAM and lignin products. There was more than 45% enhancement in breaking stress for lignin/PAMAM mats comparing to lignin mats, while an approximately 20% improvement was obtained for untreated mats. The reduction in fiber diameter for lignin/PAMAM fiber was found as a result of higher surface tension of the solution in PAMAM lignin. Such enhancements in mechanical and chemical properties of lignin/PAMAM fibers could provide an encouraging strategy for strength improvement during the future development of low cost precursors for multifunctional lignin fiber-based products, such as those for filtration.

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