

COVID-19, what could sepsis, severe acute pancreatitis, gender differences and aging teach us?

Claudio G. Gallo¹, Sirio Fiorino², Giovanni Posabella³, Donato Antonacci⁴, Antonio Tropeano⁵, Emanuele Pausini⁶, Carlotta Pausini⁷, Tommaso Guarniero⁸, Marco Zancanaro⁹

¹ Infectious disease Specialist, Physician (cgallo@gmail.com)

² Internal Medicine Unit, Budrio Hospital Azienda USL, Bologna, Italy (sirio.fiorino@ausl.bologna.it)

³ Sport Medicine Specialist, Bologna, Italy (gposabe@gmail.com)

⁴ Medical Science Department, "Casa Sollievo della Sofferenza" Hospital, San Giovanni Rotondo (FG) (antonacci.donato@alice.it)

⁵ Dentist (tropeano69@gmail.com)

⁶ Biologist, Bologna, Italy (emanuele.pausini@gmail.com)

⁷ Physician (carlottapausini@gmail.com)

⁸ Bachelor of Science (tommasoguarniero@gmail.com)

⁹ Sport Physician (marcozancanaro11@gmail.com)

Corresponding Author

Claudio G. Gallo, MD

cgallo@gmail.com

Keyword: COVID-19; Sepsis; Inflammation.

Abstract

Severe COVID-19 disease is characterised by an exaggerated inflammatory response, called cytokine storm, accompanied by a condition of immune depression. Even sepsis is characterised by an exaggerated inflammatory response, called SIRS (Systemic Inflammatory Response Syndrome), accompanied by a condition of immune depression called CARS (compensatory anti-inflammatory response syndrome). Clinical studies reveal that most sepsis patients who did not die during the hyper inflammatory response (SIRS) subsequently succumbed to the condition of immune depression (CARS). Severe acute pancreatitis begins with local inflammation that induces systemic inflammatory response syndrome (SIRS), accompanied and followed by a compensatory anti-inflammatory response (CARS). In COVID-19 disease, the male response to SARS CoV-2 virus is typically characterised by a robust inflammatory response. Instead, a cell-mediated immune response is dominant in women. This means that the male sex tends to have a more robust hyper inflammatory response than the female one. Furthermore, in women the condition of immune depression is less represented, therefore they are more protected. Sepsis, severe acute pancreatitis and COVID-19 disease evolve between two fundamental aspects: hyper inflammation and immunodepression. The experience gained over years of studies of sepsis and severe acute pancreatitis suggests that therapies should be differentiated according to the evolutionary stage of the disease. The goal is to save the lives of most patients with COVID-19 disease. The identification of critical points, suitable for designing the windows of therapeutic opportunity, may allow the use of therapeutic interventions, in the COVID-19 disease, which are effective (there are no approved drugs yet), safe (without significant side effects), targeted (based on the evolutionary phase of the disease) personalized, (based on sex, co-morbidities, age, etc.) and timely (based on signs, symptoms, laboratory parameters and instrumental investigations).

Sepsis

Sepsis is a serious clinical syndrome associated with an exaggerated systemic inflammatory response induced by an infection that can lead to multiple organ failure (MOF) and death. The main causes of sepsis are bacterial infections. A lower number of sepsis is due to viruses, parasites and fungi¹².

Sepsis is a complex process characterized by an early inflammatory response (systemic inflammatory response syndrome: SIRS), followed by a subsequent anti-inflammatory response (compensatory anti-inflammatory response syndrome: CARS). Overall, the immune response is appropriate when these two phases are balanced. An exaggerated early inflammatory response can lead to death in the initial stages. An immunosuppressive response can lead to multiple infections and multi-organ failure⁹ in later stages. **Fig. 1.**

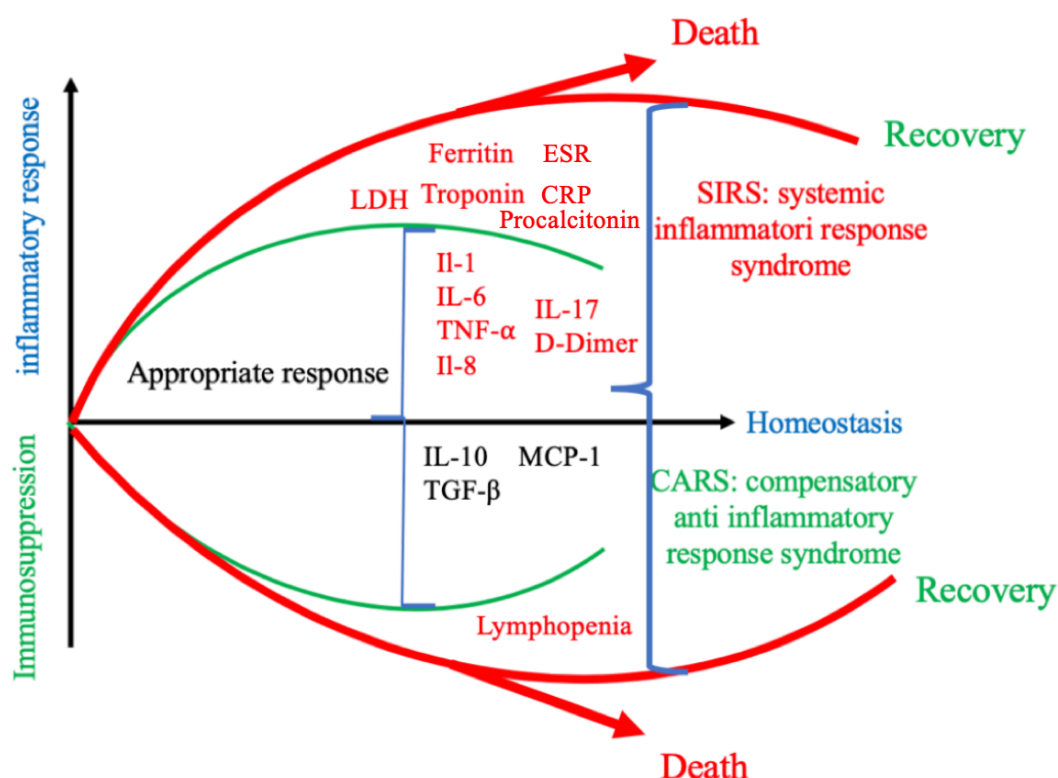


Fig. 1. SIRS and CARS in sepsis. Activation of both pro- and anti-inflammatory immune responses occurs promptly after sepsis onset.

Sepsis is a serious syndrome associated with the host's response to infection. An inappropriate sequence of early pro-inflammation (SIRS) and subsequently anti-inflammation (CARS) is responsible for the fatal outcomes in sepsis. A self-amplifying cytokine production, termed cytokine storm is the main pathogenetic mechanism by which infections can cause sepsis. In the early phase of immune response patient death is caused by SIRS. In the later phase patient death is associated with secondary infections and other complications from immunosuppression (CARS). Immune system cells and their associated molecules, inflammation mediators and coagulation factors are involved in the pathogenesis of sepsis¹¹. During the process of sepsis the two pro and anti inflammatory phases are almost superimposed. Following recognition of the pathogen, the acute phase of inflammation is triggered. This is characterized by the release of some cytokines (IL-1 β , TNF- α , IL-6, IL-8), mediators of inflammation, activation of coagulation factors and complement pathways. Pro-inflammatory cytokines, within 6 hours, induce the production of C-reactive protein (CRP) by the liver. CRP, in turn, promotes bacterial opsonization and activates complement. However, CRP is an aspecific biomarker of acute conditions and it does not allow to distinguish between infectious and non-infectious diseases. Instead, procalcitonin begins to increase already 2-4 hours after the start of the immune cascade and peaks after 6-24 hours¹³. Procalcitonin increases and reaches very high values in the course of bacterial infections. It allows to distinguish between infectious and non-infectious inflammatory diseases and between bacterial and viral pathologies^{13,14}. Only a few cytokines are constitutively expressed under non pathological conditions. On the other hand, most of these mediators are produced and secreted following the activation of several intra-cell

cascades paths. The expression of genes coding the cytokines is strongly regulated in terms of transcription and translation. The transcription factor NF κ B, for example, promotes the expression of IL-1 and IL-6¹⁵.

The goal of the inflammatory response is to fight the pathogen and restore homeostasis. Adaptive immunity is also activated immediately and anti-inflammatory cytokines are produced to reduce hyperinflammation. The body responds to the phase of hyperinflammation with an immunosuppression phase⁶. The main pro inflammatory interleukins are: TNF- α , IL-1 β , IL-6, IL-8. These interleukins are not specifically produced during sepsis, but have a well-known prognostic value. The main cytokine of the immunosuppression phase is IL-10, IL1ra, TGF- β ¹⁶. IL-10 has the function of reducing hyperinflammation^{4,5,7,8}. **Fig. 2.**

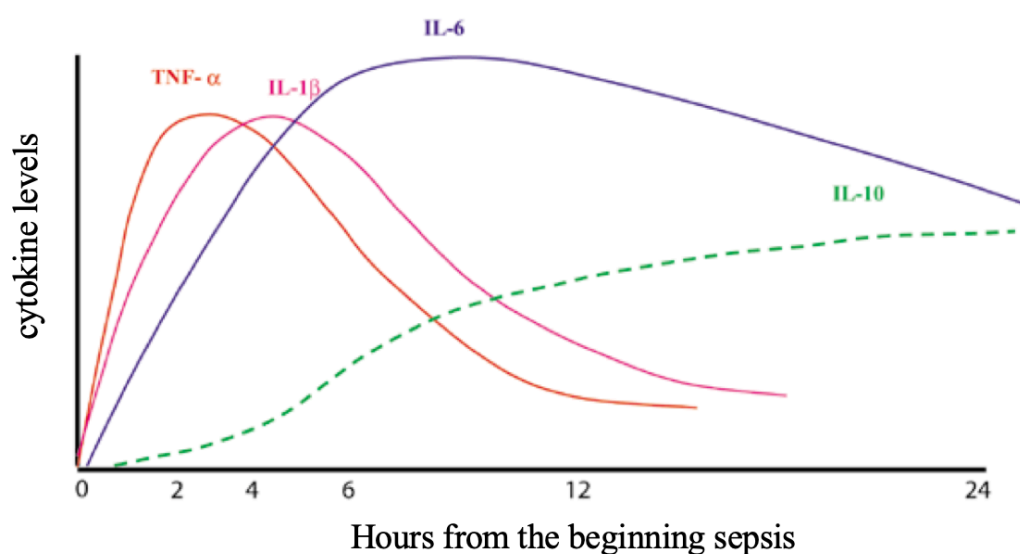


Fig. 2. Chronobiology of the inflammatory and anti-inflammatory process in sepsis (Boontham et al., 2003). IL-1 and TNF- α represent the starters in the chronobiology of inflammation. IL-6 and IL-8 peak after IL-1 β and TNF- α . IL-10 increases significantly and has the role of blocking the inflammation by modulating inflammatory cytokines.

The inflammatory response involves the release of inflammatory mediators that can trigger a self-amplifying cascade of events responsible for the "cytokine storm". Cytokine storm represents the main cause of infection severity. The local inflammatory response triggered by the infection can extend throughout the body, through the cells of the immune system and soluble mediators released into the circulation¹⁰. In the first phases of this process, the cytokines are mainly produced from macrophages, within 30-90 minutes of interaction with the pathogens¹⁶ and induces the release of other cytokines, eicosanoids and further inflammatory mediators, reactive oxygen species (ROS), adhesion molecules. IL-1 and TNF- α represent the starters in the chronobiology of inflammation. **Fig. 2.** Interleukin 1, produced by the stimulated monocytes, comprises two forms: IL-1 α and IL-1 β . Both molecules share the same receptor and, consequently, have the same biological effects. They possess a lot of common biological actions, therefore, they promote the development of inflammation with a synergistic action by binding to specific receptors and stimulate the gene expression of other inflammatory cytokines (IL-6, IL-8)¹⁸. IL-6 and IL-8 peak after IL-1 β and TNF α ¹⁷ (Fig.2). IL-6 induces the synthesis of the acute phase proteins, stimulates B and T lymphocytes and works as a pyrogen¹⁸. IL-8 works by attracting neutrophils to the inflammation site¹⁹ and it has been found in the bronchoalveolar lavage fluids (BALF) of patients with respiratory acute distress syndrome (ARDS). In addition, it's involved in multiple organ dysfunction syndrome (MOF)²⁰. In the chronobiology of the inflammatory process, IL-10 increases significantly and has the role of blocking the inflammation by modulating inflammatory cytokines (IL-6)¹⁸. IL-1Ra prevents the effects of IL-

1 by competitive binding to the IL-1R receptor ²¹. The cytokine profile in the septic patient has a prognostic value, providing useful information to establish the stage of the pathology and can help personalize the therapies.

Therefore, early mortality can be attributed to the hyperinflammatory phase. Instead, late deaths are related to the immunosuppressive phase and secondary infections⁶. Accordingly, the cytokine profile in the septic patient could provide information about the stage of the disease and the patient's prognosis, contributing to a better management. Indeed, studies have reported that IL-6 is a very promising biomarker. However, IL-1 β , TNF- α and IL-6 are not altered only in sepsis. Nevertheless, several studies have shown the importance of IL-6 in the prognosis of sepsis as it presents strong correlations with patient mortality^{2,3,4,5}. There are similarities and differences between severe COVID-19 disease and Sepsis. In fact, many COVID-19 patients have multiple organ and system involvement: lung, liver, immune system, kidneys, brain, digestive system, heart, vessels, thromboembolism^{27,42}. In addition, they develop typical clinical manifestations of septic shock: severe metabolic acidosis, cold extremities and weak peripheral pulses, regardless of evident hypotension. Therefore, according to Li's observations and hypotheses²², and on the basis of the pathogenesis of a known and already widely studied disease, such as sepsis, it is possible to draw lessons for the treatment of a new and still poorly known pathology, like COVID-19.

IL-6 is one of the main actors of the cytokine storm, contributing significantly to the increase in vascular permeability and to the impairment of multi-organ functionality²⁶. In fact, it is increased both in sepsis and in serious COVID-19 disease. It constitutes a useful biomarker and prognostic factor in both diseases. Even though IL-6 levels are significantly elevated in patients requiring ventilation, they are relatively low (hundreds of picograms/mL) compared to levels detectable in patients with septic shock (hundreds to thousands of picograms/mL)^{23,28}. IL-6 levels in COVID-19 patients increase over time with disease severity and deterioration of lung function²⁹. The kinetics of IL-6 clearly distinguishes COVID-19 patient response from septic patients²⁸. In COVID-19 patients, moderately elevated IL-6 levels are a prognostic factor associated with respiratory failure and need for mechanical ventilation (cutoff: ≥ 80 pg / ml). 92% of patients with IL-6 values ≥ 80 pg/ml require mechanical ventilation within a median time of 1.5 days (range 0-4 days)²⁴. IL-6 concentration ≥ 100 pg/mL were exclusively observed in critically ill patients and extremely high IL-6 level was closely correlated with viral load and mortality²⁵.

A useful approach for the treatment of patients with septic shock and with COVID 19 includes both in the counteracting the pathogen, but also in the suppressing the molecular response known as cytokine storm. In fact, an exaggerated cytokine response has been recognized as a possible cause of organ damage. Therefore, it was speculated that suppressing the inflammatory response by targeting IL-6, TNF-alpha, IL-1, could have a beneficial effect.

The observation of the clinical and laboratory features emerging in most studies in patients with COVID-19 underlines some analogies with the evolution of sepsis. In sepsis, the majority of patients who survive the hyperinflammatory response syndrome (SIRS) die later during the immunosuppression phase (compensatory anti-inflammatory response syndrome: CARS. In COVID-19 patients, as in septic patients, there is a condition of deep immune depression. IL-10, a cytokine with immune suppressive action, is elevated. There is a profound lymphopenia, with loss of T lymphocytes (CD4+ and CD8+), B lymphocytes, NK cells^{39,40,41}. The loss of effector cells and the lack of antibody production correlates with the development of secondary infections and mortality^{30,31}. Recent studies have highlighted the importance of cell-mediated immune response, independently from antibody production, in the defense against the SARS CoV-2 virus^{32,33,34,35}.

The autopsy results revealed significant damage in the spleen and peripheral lymphatic organs in deceased patients^{36,37,43}. Over 50% of hospitalized patients experience complications, including hospital-related infections associated with the compromise of immune system³⁰.

Therefore, the distinctive features of COVID-19 disease, against sepsis, are a) an exaggerated but more modest inflammatory response b) a deeper and progressive immunosuppression of adaptive immunity. **Fig. 3.**

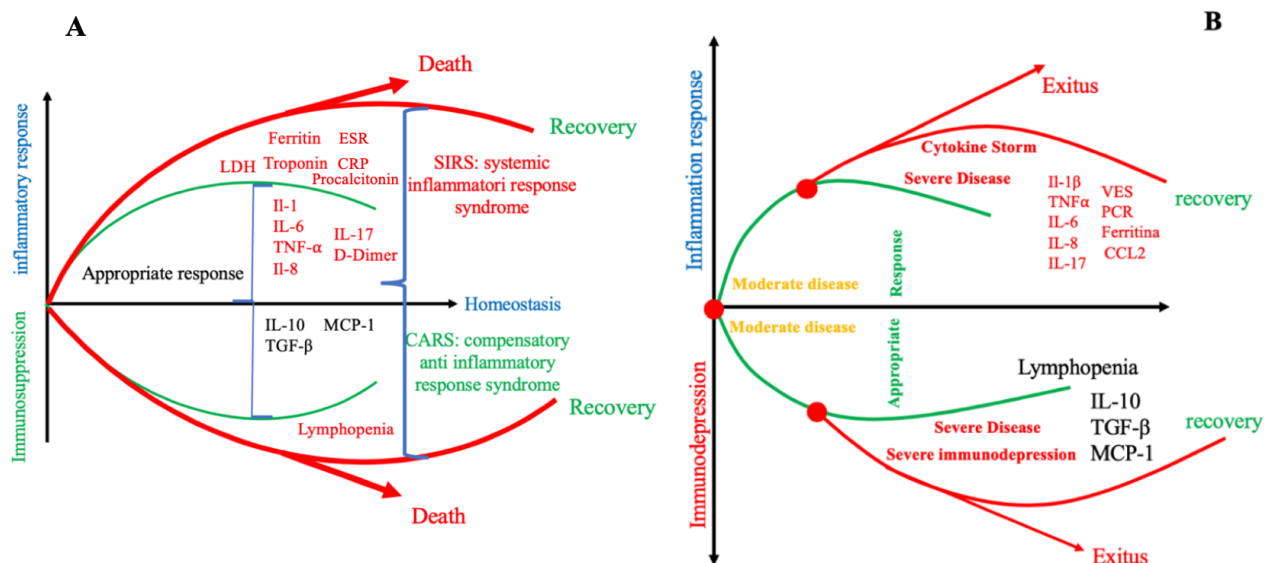


Fig. 3. Sepsis and COVID-19 in the mirror. In both pathologies the immune response includes two phases: an early pro-inflammatory condition associated with an anti-inflammatory condition. When both of these phases are balanced, an appropriate response occurs. A) An excessive early response in sepsis leads to early shock and mortality. On the other hand, an exaggerated anti-inflammatory response leads to immunosuppression or immunoparalysis with multiple infectious complications and multiorgan failure⁹. B) The immunological characteristics of COVID-19 are similar to the features detectable in sepsis, with some important differences: in sepsis the cytokine storm is more robust, while in COVID-19 there is a deeper impairment of the adaptive immune response.

Severe Acute Pancreatitis

Severe acute pancreatitis is usually fatal disease associated with a systemic inflammatory response, dominated by the production and release of pro-inflammatory cytokines and chemokines^{46,50}. It may result in ALI/ARDS (Acute Lung Injury/Acute Respiratory Distress Syndrome), MOF (Multiple Organ Failure) and culminate in patient death. Acute lung injury is the leading cause of early death in patients with severe acute pancreatitis^{47,48}. Acute pancreatitis begins with local damage at the level of the acini, due to the abnormal and early activation of pancreatic proteases⁴⁵. In response to local damage, an inflammatory reaction develops (sterile inflammation) which, thanks to self-amplification mechanisms in which the NLRP3 inflammasome is involved, causes severe disease⁴⁹. It was proposed to divide the picture into an initial hyper inflammatory phase called SIRS (systemic inflammatory response syndrome), followed by a compensatory anti-inflammatory response syndrome (CARS) phase, dominated by immunodepression and anti-inflammatory components⁴⁹. In this second phase, secondary infections mainly develop.

Sex difference and aging in COVID-19

The prevalence of COVID-19 disease is superimposable in both sexes. Male sex, however, is a risk factor for severe disease and death, regardless of age^{53,54}. Age is also an independent risk factor for severe COVID-19 disease. The immune system has different characteristics between two sexes and evolves with age. The differences are significant and affect the outcomes of COVID-19 disease. Human monocytes represent a heterogeneous population of cells of the innate immune system. At least three monocyte subpopulations have been identified, based on the expression of the CD14 and CD16 membrane receptors:

CD14⁺⁺CD16⁻ o cMonocytes (classic monocytes) (normally the prevailing subpopulation: 80%);
 CD14⁺CD16⁺ o IntMonocytes (intermediate monocytes)

CD14⁺CD16⁺⁺ nonmonocytes (non-classic monocytes).

	Men	Women
IntMonocytes	+/-	++/-
ncMonocytes	+++	+/-
Inflammatory Cytokines	+++	+/-
cell-mediated immunity	+/-	+++

Table 1. IntMonocytes: intermediate monocytes; Red: Non-classical monocytes are pathogenetic. Red: inflammatory cytokines are pathogenetic. Green: cell-mediated immunity is protective.

The transition from the classic to the intermediate and non-classic phenotype is associated with the progressive loss of the CD14 receptor and the gain of the CD16 receptor. In vitro studies have shown that non-classical monocytes are the most inflammatory in response to TLR⁵⁶. Senescence, NFkB and IL-1 β promote the phenotypic switch towards non-classic monocytes. The increase in the proportion of ncMonocytes is associated with the increase in the production of TNF α , IL-1 β and IL-8^{52,56,57,58,59,60,61,62}.

A recent study highlights an exaggerated switch towards the ncMonociti phenotype in patients with type 2 diabetes affected by COVID-19. As is known, diabetes is one of the major risk factors for severe COVID-19 disease^{63,64}. A recent study examines the different characteristics of the basic immune response to SARS Cov-2 virus in both sexes, during the early stages of the disease⁶⁵. In male subjects, a higher percentage of ncMonocytes is found which translates into high levels of cytokines and inflammatory chemokines such as IL-8 and CCL5 (Rantes).

The CCL5 chemokine (Rantes) has several effects on innate and adaptive immunity (promotes macrophage phagocytosis in the inflamed lung, contributes to the migration of dendritic cells from the lung to the draining lymph nodes, recruits the effector T lymphocytes, promoting the immune response adaptive in the lung). IL-8 is a chemokine that recruits neutrophils at the infection site level. Neutrophils are associated with a more severe prognosis for COVID-19 disease^{96,97}.

nc-Monocytes contribute to endothelial inflammation and can help to promote micro thrombosis and thromboembolism^{69,70}. Male subjects have a robust inflammatory response, with an increased production of inflammatory cytokines of the innate immune system. In female subjects there is a robust cell-mediated response, with a prevalence of CD8 T lymphocytes. **Tab. 1.**

Consistent with the effects of these peculiar immune responses, male subjects with poorer Cell-mediated response have a worse prognosis. Instead, the prognosis is worse in female subjects with a higher level of cytokines from the innate immune system. Finally, the impairment of the cell-mediated response correlates with age in male subjects but not in female subjects. **Fig. 4.**

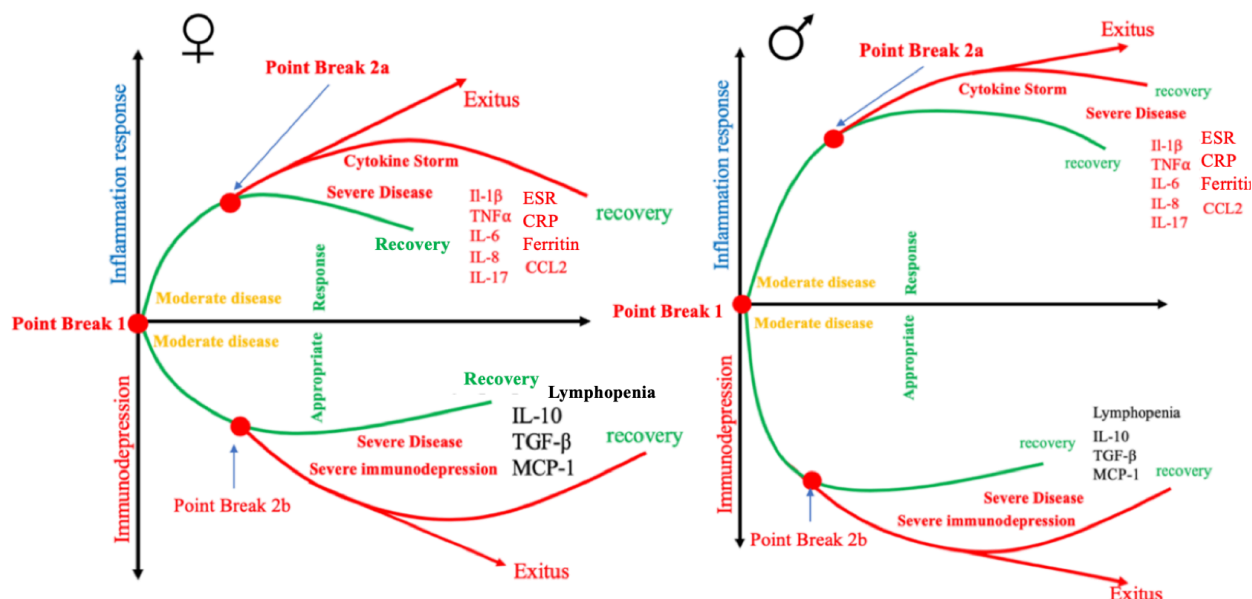


Fig. 4. comparison between the immune response to SARS CoV-2 virus of women ♀ and men ♂. Under basic conditions, the immune response to the SARS CoV-2 virus in men is more inflammatory and less immunocompetent than in women.

Conclusive remarks

Exaggerated inflammation, accompanied by immunodepression, characterises sepsis, severe acute pancreatitis and severe COVID-19 disease. Both conditions are basic for some important risk factors for severe COVID-19 disease: advanced age, male gender, type 2 diabetes. These two different conditions require a different therapeutic approach. In COVID-19 disease, in addition to modulating the exaggerated innate inflammatory response, it is essential to include a proper support for immune defenses as therapeutic strategies. The correct balance between cytokine storm contrast and adaptive immune function support must be adapted to the stage of the disease.

The windows of therapeutic opportunities should be designed to promptly identify precise reference points (symptoms, signs, laboratory parameters, instrumental investigations). Therapies should be optimised by diversifying them according to the evolutionary phase. The results of the vast amount of work in progress will provide, in the near future, a plethora of data. The correct examination of the hypotheses formulated should allow us to refine the therapies, sewn on like a tailored suit.

List of abbreviations:

ALI: Acute Lung Injury
 BALF: Bronchoalveolar Lavage Fluid
 ARDS: Acute Respiratory Distress Syndrome
 CARS: compensatory anti-inflammatory response syndrome
 CCL5: Chemokine (C-C motif) ligand 5
 cMonocytes: classical Monocytes
 CRP: C-reactive Protein
 ESR: erythrocyte sedimentation rate
 IL-1 β : Interleukin 1 β
 IL-6: Interleukin 6
 IL-8: Interleukin 8
 IL-10: Interleukin 10
 IL-17: Interleukin 17
 IL-1R: IL-1 Receptor
 IL-1RA: Interleukin-1 Receptor Antagonist
 intMonocytes: Intermediate Monocytes
 LDH: Lactate Dehydrogenase
 MCP-1: Monocyte Chemoattractant Protein-1
 MOF: Multiple organ failure
 ncMonocytes: non classical Monocytes
 Rantes: regulated on activation, normal T cell expressed and secreted
 SIRS: Systemic Inflammatory Response Syndrome
 TGF- β : transforming growth factor β
 TLRs: Toll-Like Receptors
 TNF α : Tumor Necrosis Factor α

Declaration of interests:

The authors declare that there are no conflicts of interest regarding the publication of this paper.

Reference

- ¹ Xiao W, Mindrinos MN, Seok J, et al. (2011): A genomic storm in critically injured humans. *J Exp Med* 208: 2581-2590.
- ² Pettilä V et al. Predictive value of procalcitonin and interleukin 6 in critically ill patients with suspected sepsis. *Intensive Care Medicine*. 2002;28(9):1220-1225.
- ³ Osuchowski MF et al. Stratification is the key: Inflammatory biomarkers accurately direct immunomodulatory therapy in experimental sepsis. *Critical Care Medicine*. 2009; 37 (5): 1567
- ⁴ Lusyati S et al. Cytokines patterns in newborn infants with late onset sepsis. *Journal of Neonatal-Perinatal Medicine*. 2013;6(2):153-163
- ⁵ Machado JR, Soave DF, da Silva MV, de Menezes LB, Etchebehere RM, Monteiro ML, et al. Neonatal sepsis and inflammatory mediators. *Mediators of Inflammation*. 2014; 2014: 269681
- ⁶ Biron BM, Ayala A, Lomas-Neira JL. Biomarkers for sepsis: What is and what might be? *Biomarker Insights*. 2015; 10: BMI-S29519
- ⁷ Berner R et al. Plasma levels and gene expression of granulocyte colony-stimulating factor, tumor necrosis factor- α , interleukin (IL) -1 β , IL-6, IL-8, and soluble intercellular adhesion molecule-1 in neonatal early onset sepsis. *Pediatric Research*. 1998; 44 (4): 469
- ⁸ Adib-Conquy M, Cavaillon J-M. Compensatory anti-inflammatory response syndrome. *Thrombosis and Haemostasis*. 2009; 102 (01): 36-47
- ⁹ Webster NR1, Galley HF. Immunomodulation in the critically ill. *Br J Anaesth*. 2009 Jul;103(1):70-81.
- ¹⁰ Chousterman BG1,2, Swirski FK3, Weber GF4. Cytokine storm and sepsis disease pathogenesis. *Semin Immunopathol*. 2017 Jul;39(5):517-528.
- ¹¹ Hotchkiss RS1, Monneret G, Payen D. Sepsis-induced immunosuppression: from cellular dysfunctions to immunotherapy. *Nat Rev Immunol*. 2013 Dec;13(12):862-74.

- ¹² Beale R1, Reinhart K, Brunkhorst FM, Dobb G, Levy M, Martin G, Martin C, Ramsey G, Silva E, Vallet B, Vincent JL, Janes JM, Sarwat S, Williams MD; PROGRESS Advisory Board. Promoting Global Research Excellence in Severe Sepsis (PROGRESS): lessons from an international sepsis registry. *Infection*. 2009 Jun;37(3):222-32
- ¹³ Mehanic, S. and Baljic, R. (2013) 'The Importance of Serum Procalcitonin in Diagnosis and Treatment of Serious Bacterial Infections and Sepsis', *Materia Socio Medica*, 25(4), p. 277.
- ¹⁴ Sang Tae Choi and Jung-Soo Song. Serum Procalcitonin as a Useful Serologic Marker for Differential Diagnosis between Acute Gouty Attack and Bacterial Infection. *Yonsei Med J* 2016 Sep; 57 (5): 1139-1144
- ¹⁵ Burkovskiy, I. et al., (2013) 'Cytokine release in sepsis', *Adv Bioscience Biotech*, 2013 (4), pp. 860-865. doi: 10.4236 / abb.2013.49114.
- ¹⁶ Raina Devi Ramnath, Siaw Weing, Min He, Jia Sun, Huili Zhang, Manmish Singh Bawa & Madhav Bhatia (2006) Inflammatory mediators in sepsis: Cytokines, chemokines, adhesion molecules and gases, *Journal of Organ Dysfunction*, 2: 2, 80-92.
- ¹⁷ Boontham, P. et al., (2003). 'Surgical sepsis: Dysregulation of immune function and therapeutic implications', *Surgeon*, pp. 187–206.
- ¹⁸ Blackwell TS, Christman JW. Sepsis and cytokines: current status. *Br J Anaesth*. 1996 Jul;77(1):110-7.
- ¹⁹ Gomes, A. P. et al., (2015) 'Pro-Inflammatory Cytokines in Sepsis: Biological Studies and Prospects From In Silico Research', *Biological Systems: Open Access*, 5(1), pp. 1–7.
- ²⁰ Miller, E. J., Cohen, A. B. and Matthay, M. A. (1996) 'Increased interleukin-8 concentrations in the pulmonary edema fluid of patients with acute respiratory distress syndrome from sepsis. *Critical care medicine*, 24(9), pp. 1448–54.
- ²¹ Dinarello, C. A. (1998) 'Interleukin-1, Interleukin-1 Receptors and Interleukin-1 Receptor Antagonist', *International Reviews of Immunology*, 16 (5–6), pp. 457– 499.
- ²² Hui Li, Liang Liu, Dingyu Zhang, Jiuyang Xu, Huaping Dai, Nan Tang, Xiao Su, Bin Cao. SARS-CoV-2 and viral sepsis: observations and hypotheses. *Lancet*. 2020 May 9;395(10235):1517-1520.
- ²³ Surbatovic M, Popovic N, Vojvodic D, et al. Cytokine profile in severe Gram-positive and Gram-negative abdominal sepsis. *Sci Rep*. 2015;5:11355
- ²⁴ Tobias Herold, Vindi Jurinovic, Chiara Arnreich, Johannes C Hellmuth, Michael von Bergwelt-Baildon, Matthias Klein, Tobias Weinberger. Level of IL-6 predicts respiratory failure in hospitalized symptomatic COVID-19 patients. *medRxiv*, 2020
- ²⁵ Chen X, Zhao B, Qu Y, Chen Y, Xiong J, Feng Y, Men D, Huang Q, Liu Y, Yang B, Ding J, Li F. Detectable serum SARS-CoV-2 viral load (RNAemia) is closely correlated with drastically elevated interleukin 6 (IL-6) level in critically ill COVID-19 patients. *Clin Infect Dis*. 2020 Apr 17.
- ²⁶ Eric A. Coomes MDHH, M.D., M.Sc., FRCPC. Interleukin-6 in COVID-19: A Systematic Review and Meta-Analysis. *medRxiv*. 2020.
- ²⁷ Victor G. Puelles, M.D., Ph.D.. Multiorgan and Renal Tropism of SARS-CoV-2. *n engl j med*. May 13, 2020.
- ²⁸ Kenneth E Remy, Scott C Brakenridge, Bruno Francois, Thomas Daix, Clifford S Deutschman, Guillaume Monneret, Robin Jeannet, Pierre-Francois Laterre, Richard S Hotchkiss, Lyle L Moldawer. Immunotherapies for COVID-19: lessons learned from sepsis. *Comment*. April 28, 2020.
- ²⁹ Zhang C, Wu Z, Li JW, Zhao H, Wang GQ. The cytokine release syndrome (CRS) of severe COVID-19 and interleukin-6 receptor (IL-6R) antagonist 15 tocilizumab may be the key to reduce the mortality. *Int J Antimicrob Agents* 2020; published online March 29.
- ³⁰ Wang D, Hu B, Hu C, et al. Clinical characteristics of 138 hospitalized patients with 2019 novel coronavirus-infected pneumonia in Wuhan, 17 China. *JAMA* 2020; 323: 1061–69.
- ³¹ Zhou F, Yu T, Du R, et al. Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. *Lancet* 2020; 395: 1054–62.
- ³² Annarosa Soresina Daniele Moratto Marco Chiarini Ciro Paolillo Giulia Baresi Emanuele Focà Michela Bezzi Barbara Baronio Mauro Giacomelli Raffaele Badolato. Two X-linked agammaglobulinemia patients develop pneumonia as COVID-19 manifestation but recover. *Pediatr Allergy Immunol*. 2020 Apr 22.
- ³³ Julian Braun, Lucie Loyal, Marco Frentsch, Daniel Wendisch, Philipp Georg, Florian Kurth, Stefan Hippenstiel, Manuela Dingeldey, Beate Kruse, Florent Fauchere, Emre Baysal, Maike Mangold, Larissa Henze, Roland Lauster, Marcus Mall, Kirsten Beyer, Jobst Roehmel, Juergen Schmitz, Stefan Miltenyi, Marcel A Mueller, Martin Witzernath, Norbert Suttrop, Florian Kern, Ulf Reimer, Holger Wenschuh, Christian Drosten, Victor M Corman, Claudia Giesecke-Thiel, Leif-Erik Sander, Andreas Thiel. Presence of SARS-CoV-2 reactive T cells in COVID-19 patients and healthy donors. <https://www.medrxiv.org/content/10.1101/2020.04.17.20061440v1>

- ³⁴ Grifoni, A., Weiskopf, D., Ramirez, SI, Mateus, J., Dan, JM, Moderbacher, CR, Rawlings, SA, Sutherland, A., Premkumar, L., Jadi, RS, Marrama, D., de Silva, AM, Frazier, A., Carlin, A., Greenbaum, JA, Peters, B., Krammer, F., Smith, DM, Crotty, S., Sette, A., Targets of T cell responses to SARS -CoV-2 coronavirus in humans with COVID-19 disease and unexposed individuals, *Cell* (2020).
- ³⁵ YouennJouan, Antoine Guillon, Loic Gonzalez, Yonatan Perez, Stephan Ehrmann, Marion Ferreira, Thomas Daix, Robin Jeannet, Bruno Francois, Pierre-Francois Dequin, Mustapha Si-Tahar, Thomas Baranek, Christophe Paget. Functional alteration of innate T cells in critically ill Covid-19 patients. <https://www.medrxiv.org/content/10.1101/2020.05.03.20089300v1>
- ³⁶ Xu X, Chang XN, Pan HX, Su H, Huang B, Yang M, Luo DJ, Weng MX, Ma L, Nie X. Pathological changes of the spleen in ten patients with new coronavirus infection by minimally invasive autopsies. *Zhonghua Bing Li XueZaZhi*. 2020 Apr 27;49(0):E014.
- ³⁷ Yao XH, Li TY, He ZC, Ping YF, Liu HW, Yu SC, Mou HM, Wang LH, Zhang HR, Fu WJ, Luo T, Liu F, Chen C, Xiao HL, Guo HT, Lin S, Xiang DF, Shi Y, Li QR, Huang X, Cui Y, Li XZ, Tang W, Pan PF, Huang XQ, Ding Y8, Bian XW. A pathological report of three COVID-19 cases by minimally invasive autopsies. *Zhonghua Bing Li XueZaZhi*. 2020 Mar 15;49(0):E009
- ³⁸ Zhang W, Zhao Y, Zhang F, et al. The use of anti-inflammatory drugs in the treatment of people with severe coronavirus disease 2019 (COVID-19): the perspectives of clinical immunologists from China. *Clin Immunol* 2020; 214: 108393
- ³⁹ Zheng Y, Huang Z, Guoping Y, et al. Study of the lymphocyte change between COVID-19 and non-COVID-19 pneumonia cases suggesting other 19 factors besides uncontrolled inflammation contributed to multi-organinjury. *medRxiv* 2020; published online Feb 23. DOI:10.1101/2020.02.19.20024885 (preprint)
- ⁴⁰ Zheng HY, Zhang M, Yang CX, et al. Elevated exhaustion levels and reduced functional diversity of T cells in peripheral blood may predict severe progression in COVID-19 patients. *Cell Mol Immunol* 2020; published online March 17. DOI:10.1038/s41423-020-0401-3.
- ⁴¹ Zheng M, Gao Y, Wang G, et al. Functional exhaustion of antiviral lymphocytes in COVID-19 patients. *Cell Mol Immunol* 2020; published online March 19. DOI:10.1038/s41423-020-0402-2.
- ⁴² Pavan K. Bhatraju, M.D., Bijan J. Ghassemieh, M.D., Michelle Nichols, M.D., Richard Kim, M.D., Keith R. Jerome, M.D., Arun K. Nalla, Ph.D., Alexander L. Greninger, M.D., Sudhakar Pipavath, M.D., Mark M. Wurfel, M.D., Ph.D., Laura Evans, M.D., Patricia A. Kritek, M.D., T. Eoin West, M.D., M.P.H., et al. Covid-19 in Critically Ill Patients in the Seattle Region — Case Series. March 30, 2020, at NEJM.
- ⁴³ National Health Commission & National Administration of Traditional Chinese Medicine. Diagnosis and Treatment Protocol for Novel Coronavirus Pneumonia (Trial Version 7). *Chinese Medical Journal*. March 3, 2020.
- ⁴⁴ Hirota M, Sugita H, Maeda K, Ichibara A, Ogawa M. *Nihon Rinsho*. 2004;62(11):2128-2136.
Singh VK, Wu BU, Bollen TL, et al. Early systemic inflammatory response syndrome is associated with severe acute pancreatitis. *Clin Gastroenterol Hepatol*. 2009;7(11):1247-1251.
- ⁴⁵ Sendler M, Maertin S, John D, et al. Cathepsin B activity initiates apoptosis via digestive protease activation in pancreatic acinar cells and experimental pancreatitis. *J Biol Chem* 2016;291:14717–14731.
- ⁴⁶ Mofidi R, Duff MD, Wigmore SJ, et al. Association between early systemic inflammatory response, severity of multiorgan dysfunction and death in acute pancreatitis. *Br J Surg* 2006;93:738–744
- ⁴⁷ Fu, Q., Zhai, Z., Wang, Y., Xu, L., Jia, P., Xia, P., Liu, C., Zhang, X., Qin, T., & Zhang, H. (2018). NLRP3 Deficiency Alleviates Severe Acute Pancreatitis and Pancreatitis-Associated Lung Injury in a Mouse Model. *BioMed research international*, 2018, 1294951.
- ⁴⁸ Sven M van Dijk SM, Hallensleben NDL, van Santvoort HC, et al. Acute pancreatitis: recent advances through randomised trials. *Gut* 2017;66:2024–2032.
- ⁴⁹ Sendler M, van den Brandt C, Glaubitz J, et al. NLRP3 Inflammasome Regulates Development of Systemic Inflammatory Response and Compensatory Anti-Inflammatory Response Syndromes in Mice With Acute Pancreatitis. *Gastroenterology*. 2020;158(1):253-269.e14.
- ⁵⁰ Singh VK, Wu BU, Bollen TL, et al. Early systemic inflammatory response syndrome is associated with severe acute pancreatitis. *Clin Gastroenterol Hepatol*. 2009;7(11):1247-1251.

- ⁵¹ Takahashi T, Wong P, Ellingson M, et al. Sex differences in immune responses to SARS-CoV-2 that underlie disease outcomes. medRxiv; 2020. DOI: 10.1101/2020.06.06.20123414.
- ⁵² Pence, B.D. Severe COVID-19 and aging: are monocytes the key?. *GeroScience* (2020).
- ⁵³ Jin, J. M., Bai, P., He, W., Wu, F., Liu, X. F., Han, D. M., Liu, S., & Yang, J. K. (2020). Gender Differences in Patients With COVID-19: Focus on Severity and Mortality. *Frontiers in public health*, 8, 152.
- ⁵⁴ Meng Y, Wu P, Lu W, et al. Sex-specific clinical characteristics and prognosis of coronavirus disease-19 infection in Wuhan, China: A retrospective study of 168 severe patients. *PLoS Pathog.* 2020;16(4):e1008520. Published 2020 Apr 28.
- ⁵⁵ Antonelli LR, Leoratti FM, Costa PA, et al. The CD14+CD16+ inflammatory monocyte subset displays increased mitochondrial activity and effector function during acute Plasmodium vivax malaria. *PLoS Pathog.*
- ⁵⁶ Ong, S., Hadadi, E., Dang, T. *et al.* The pro-inflammatory phenotype of the human non-classical monocyte subset is attributed to senescence. *Cell Death Dis* 9, 266 (2018).
- ⁵⁷ Fawaz ALZaid, Jean-Baptiste Julla, Marc Diedisheim, Charline Potier, Louis Potier, Gilberto Velho, Benedicte Gaborit, Philippe Manivet, Stephane Germain, Tiphaine Vidal-Trecan, Ronan Roussel, Jean-Pierre Riveline, Elise Dalmas, Nicolas Venteclef, Jean-Francois Gautier. Monocyte class switch and hyperinflammation characterise severe COVID-19 in type 2 diabetes. medRxiv 2020.06.02.20119909; doi: <https://doi.org/10.1101/2020.06.02.20119909>
- ⁵⁸ Hijdra, D., Vorselaars, A. D., Grutters, J. C., Claessen, A. M., and Rijkers, G. T. (2013) Phenotypic characterization of human intermediate monocytes. *Front Immunol* 4, 339.
- ⁵⁹ Stansfield, B. K. & Ingram, D. A. Clinical significance of monocyte heterogeneity. *Clin. Transl. Med* 4, 5 (2015).
- ⁶⁰ Belge, K. U. et al. The proinflammatory CD14+CD16+DR++ monocytes are a major source of TNF. *J. Immunol.* 168, 3536–3542 (2002).
- ⁶¹ Frankenberger, M., Sternsdorf, T., Pechumer, H., Pforte, A. & Ziegler-Heitbrock, H. W. Differential cytokine expression in human blood monocyte sub-populations: a polymerase chain reaction analysis. *Blood* 87, 373–377 (1996).
- ⁶² Mukherjee, R. et al Non-Classical monocytes display inflammatory features: validation in sepsis and systemic lupus erythematosus. *Sci. Rep.* 5, 13886 (2015).
- ⁶³ Pinto, Lana C. and BERTOLUCI, Marcello C.. Type 2 diabetes as a major risk factor for COVID-19 severity: a meta-analysis. *Arch. Endocrinol. Metab.* [online]. 2020, vol.64, n.3 [cited 2020-07-14], pp.199-200.
- ⁶⁴ Banerjee A, Pasea L, Harris S, et al. Estimating excess 1-year mortality associated with the COVID-19 pandemic according to underlying conditions and age: a population-based cohort study. *Lancet.* 2020;395(10238):1715-1725.
- ⁶⁵ Takehiro Takahashi, Patrick Wong, Mallory Ellingson, Carolina Lucas, Jon Klein, Benjamin Israelow, Julio Silva, Jieun Oh, Tianyang Mao, Maria Tokuyama, Peiwen Lu, Arvind Venkataraman, Annsea Park, Feimei Liu, Amit Meir, Jonathan Sun, Eric Wang, Anne Louise Wyllie, Chantal B.F. Vogels, Rebecca Earnest, Sarah Lapidus, Isabel Ott, Adam Moore, Arnau Casanovas, Charles Dela Cruz, John Fournier, Camila Odio, Shelli Farhadian, Nathan Grubaugh, Wade Schulz, Albert Ko, Aaron Ring, Saad Omer, Akiko Iwasaki, Yale IMPACT research team. Sex differences in immune responses to SARS-CoV-2 that underlie disease outcomes. medRxiv 2020.06.06.20123414.
- ⁶⁶ Grayson, Mitchell & Holtzman, Michael. (2006). Chemokine Signaling Regulates Apoptosis as well as Immune Cell Traffic in Host Defense. *Cell cycle* (Georgetown, Tex.). 5. 380-3.
- ⁶⁷ Ciccullo, A., Borghetti, A., Zileri Dal Verme, L., Tosoni, A., Lombardi, F., Garcovich, M., Biscetti, F., Montalto, M., Cauda, R., Di Giambenedetto, S., & GEMELLI AGAINST COVID Group (2020). Neutrophil-to-lymphocyte ratio and clinical outcome in COVID-19: a report from the Italian front line. *International journal of antimicrobial agents*, 106017.
- ⁶⁸ Liu, J., Liu, Y., Xiang, P. *et al.* Neutrophil-to-lymphocyte ratio predicts critical illness patients with 2019 coronavirus disease in the early stage. *J Transl Med* 18, 206 (2020).
- ⁶⁹ Chimen M, Yates CM, McGettrick HM, et al. Monocyte Subsets Coregulate Inflammatory Responses by Integrated Signaling through TNF and IL-6 at the Endothelial Cell Interface. *J Immunol.* 2017;198(7):2834-2843.

⁷⁰ Buscher K, Marcovecchio P, Hedrick CC, Ley K. Patrolling Mechanics of Non-Classical Monocytes in Vascular Inflammation. *Front Cardiovasc Med*. 2017;4:80.