INCREASED ALDEHYDE FORMATION AS A KEY COMPONENT OF THE METABOLIC SYNDROME IN ADOLESCENTS: A MACHINE LEARNING STUDY

Kamila Landucci BONIFACIO, Ph.D¹; Michael MAES, Ph.D²,³,⁴, Carine Coneglian de FARIAS, Ph.D ¹; Andressa Keiko MATSUMOTO, MD¹; Crisieli Maria TOMERELI, Ph.D⁵, Danilo Rodrigues Pereira da SILVA, Ph.D⁵; Edilson Serpeloni CYRINO, Ph.D⁵; Suzana Lucy NIXDORF, Ph.D⁶; Danielle VENTURINI, Ph.D ¹,⁷; Décio Sabbatini BARBOSA, Ph.D ¹,⁷

¹Laboratory of Graduation Research, State University of Londrina, University Hospital, Londrina, Paraná, Brazil
²Department of Psychiatry, Faculty of Medicine, King Chulalongkorn Memorial Hospital, Bangkok, Thailand
³Department of Psychiatry, Medical University of Plovdiv, Plovdiv, Bulgaria
⁴IMPACT Strategic Research Centre, Deakin University, Geelong, VIC, Australia
⁵Metabolism, Nutrition, and Exercise Research Group, Sport and Physical Education Center, State University of Londrina
⁶Department of Chemistry, State University of Londrina, Londrina, PR, Brazil
⁷Department of Clinical Analysis and Toxicological, State University of Londrina

Corresponding author:
Kamila Landucci Bonifacio, Ph.D
Laboratory of Graduation Research, State University of Londrina, University Hospital, Londrina, Paraná, Brazil
E-mail address: kamilalondrina@hotmail.com
ABSTRACT

**Purpose:** To investigate the alterations in nitro-oxidative stress (OS) and antioxidant status in adolescents with metabolic syndrome (MetS) and whether these alterations occur independently from effects of overweight or obesity.

**Methods:** Blood was collected in 47 adolescents with MetS and 94 adolescents without MetS as assessed with the International Diabetes Federation criteria. The International Obesity Task Force (IOTF) criteria were used to classify the subjects into those with overweight or obesity. We measured nitro-oxidative biomarkers including nitric oxide metabolites (NOx), lipid hydroperoxides (LOOH), and malondialdehyde (MDA), and antioxidant biomarkers, i.e. total radical-trapping antioxidant parameter (TRAP), paraoxonase (PON)-1 activity, thiol (SH-) groups, as well as tumor necrosis factor-α, glucose, insulin, triglycerides, uric acid and high-density lipoprotein cholesterol (HDL-C).

**Results:** Logistic regression analysis showed that increased MDA and NOx and a lowered TRAP/uric acid ratio were associated with MetS. Machine learning including soft independent modeling of class analogy (SIMCA) showed that the top-3 most important features of MetS were increased glucose and MDA and lowered HDL-C. Support vector machine using MDA, glucose, insulin, HDL-C, triglycerides and body mass index as input variables yielded a 10-fold cross-validated accuracy of 89.8% when discriminating MetS from controls. The association between MetS and increased MDA was independent from the effects of overweight-obesity, glucose, insulin, triglycerides and HDL-C.

**Conclusion:** In adolescents, increased MDA formation is a key component of MetS, indicating that increased production of reactive oxygen species with consequent lipid peroxidation and aldehyde formation participate in the development of MetS.
**Keywords:** Metabolic Syndrome; Obesity; inflammation; Oxidative Stress; nitrosative stress; biomarkers

Emails:

kamilalondrina@hotmail.com
carine_coneglian@yahoo.com.br
dr.michaelmaes@hotmail.com
dessamatsu@hotmail.com
danilorpsilva@gmail.com
crisieli@uol.com.br
emcyrino@uel.com.br
s.nixdorf@hotmail.com
danielle.venturini@bol.com.br
sabatini2011@hotmail.com
**List of abbreviations:** oxidative stress (OS); Metabolic syndrome (MetS); International Diabetes Federation (IDF); International Obesity Task Force (IOTF); tobacco use disorder (TUD); body mass index (BMI); waist circumference (WC); high-density lipoprotein cholesterol (HDL-C); low-density lipoprotein cholesterol (LDL-C); reactive oxygen species (ROS), reactive nitrogen species (RNS), oxidative and nitrosative stress (O&NS); malondialdehyde (MDA); lipid hydroperoxides (LOOH); nitric oxide metabolites (NOx); total radical-trapping antioxidant parameter (TRAP); tumor necrosis factor α (TNFα); paraoxonase (PON)-1 activity; thiol groups (SH-); protein (PT); statistical isolinear multiple component analysis (SIMCA); support vector machine (SVM); principal component analysis (PCA); neutral network (NN).
INTRODUCTION

Key features of metabolic syndrome (MetS) are central obesity, insulin resistance and dyslipidemia as evidenced by increased levels of insulin, glucose and triglycerides, in conjunction with lowered high-density lipoprotein cholesterol (HDL-C) [1]. Central obesity is accompanied by a chronic mild inflammatory state mediated by an increased production of pro-inflammatory cytokines by immune cells and adipocytes, including tumor necrosis factor alpha (TNF-α). This pro-inflammatory response is regarded to function as a homeostatic process that attenuates excess fat accumulation [2].

MetS is accompanied by increased reactive oxygen (ROS) and nitrogen (RNS) species and decreases in antioxidant defenses creating an environment that promotes oxidative and nitrosative stress (O&NS) [3,4]. Decreased levels of antioxidant enzymes including catalase, superoxide dismutase and glutathione peroxidase, and increased levels of malondialdehyde (MDA, indicating lipid peroxidation) and protein carbonyls (indicating protein oxidation) were observed in MetS [5]. Furthermore, concentrations of nitric oxide metabolites (NOx) are associated with MetS [6], while other studies reported lower concentrations of nitric oxide (NO) formation biomarkers in adult MetS [7,8]. Another study found lower levels of thiol (-SH) groups in patients with MetS features as compared to controls, although there are also contradictory results [9]. The assay of -SH groups may be used as a marker of the redox status of albumin reflecting non-enzymatic antioxidant defenses.

Nevertheless, obesity may impact the relationship between oxidative stress biomarkers and MetS. For example, obesity is associated with low systemic antioxidant defenses and enhanced lipid peroxidation, as indicated by elevated MDA levels [10–12]. Lowered NO formation is not only reported in MetS but also in obese juveniles [7,8], while other authors reported higher NOx levels in obese children [6]. The total radical-trapping antioxidant parameter (TRAP) is decreased in obese children especially when MetS features
are present [1,13]. Likewise, the activity of paraoxonase 1 (PON), a strong antioxidant enzyme, is decreased in adult obese patients and in patients with MetS [14]. On the other hand, increased uric acid levels (which displays antioxidant and pro-oxidant effects) is a component of the MetS in adults [15] as well as in adolescents and children [16] and is positively associated with waist circumference, fasting glucose and insulin, and insulin resistance, and inversely with HDL-cholesterol [17].

Therefore, it is not always clear whether changes in these redox biomarkers in MetS are a feature of MetS or overweight/obesity or both. In addition, only few studies in children and adolescents focused on O&NS in MetS as compared with a much larger number of studies in adults [18]. Hence, the aims of the present study are to examine whether a) increased O&NS biomarkers, including MDA and lipid hydroperoxides (LOOH), NOx, lowered antioxidant biomarkers, i.e. TRAP, SH- groups and PON-1 activity, and increased TNF-α are hallmarks of MetS in adolescents, b) these biomarkers contribute to an accurate classification of MetS; and c) these O&NS measurements are biomarkers of MetS or overweight-obesity.

2 SUBJECTS AND METHODS

2.1 Subjects

We included 47 adolescents with the MetS and 94 adolescents without the MetS. All subjects were 11-17 years old and both genders were included. They were recruited from public schools located in the city of Londrina, Paraná, Brazil. Exclusion criteria were clinical or laboratory signs of medical disease, being under treatment for any disease, use of illicit drugs, and not being regularly enrolled in schools. All participants underwent a physical examination, anthropometric measurements and a structured interview. The adolescents were always accompanied by their adult relatives (parents or first-degree) when they completed the
structured interview. Both parents and adolescents gave their signed informed consent to participate in the study. The protocol was approved by the Ethics Committee on Research Involving Human Subjects of the State University of Londrina.

The MetS was diagnosed according to the International Diabetes Federation (IDF) consensus criteria [19]. Most of our controls had no or less than 2 features of the MetS. The International Obesity Task Force (IOTF) [20] criteria were used to divide the subjects into those with a normal BMI, overweight and obesity (BMI cut off values are adjusted for age and gender). The diagnosis tobacco use disorder (TUD) was made using DSM IV criteria.

Body weight and height were measured using standardized methods [21]. Body weight was assessed using a digital scale with an accuracy of 0.1 kg, and body height using a portable wooden stadiometer with an accuracy of 0.1 cm. Waist circumference (WC) was measured at the level of the umbilicus. Hip circumference was measured over light clothing at the widest girth of the hip using an unstretched tape meter, without any pressure to body surface. Systolic and diastolic blood pressure were assessed using the digital OMRON Model HEM-742 [22].

2.2. Methods

Fasting blood was collected from the antecubital vein between 8 a.m. and 10.00. a.m. Uric acid, total cholesterol and HDL-C, triglycerides, glucose, insulin were determined by automated methods using the Dimension®RxL, (Deerfield, IL, USA) and i2000SR Architect (Abbott, IL, USA). LDL-cholesterol was calculated using the Friedewald equation. Nitric oxide metabolites (NOx) levels were assessed by an adaptation of the technique described Navarro-Gonzálvez et al.[23] Lipid hydroperoxides (LOOH) were determined using the ferrous oxidation xylene orange technique [24]. Total radical-trapping antioxidant parameter (TRAP) was evaluated according to the method described by Repetto et al. [25]. The
TRAP/MDA ratio was computed as a biomarker of antioxidant defenses versus oxidative stress. Thiol (SH-) groups were assessed as described by Hu [26]. Total plasma paraoxonase 1 (PON)-1 activity was determined as described by Richter et al. The activity was expressed in U/mL based on the phenyl acetate molar extinction coefficient of 1.31 mMol/L cm⁻¹ [27]. Malondialdehyde (MDA) was quantified according to the technique described by Bastos et al. [28]. TNFα was quantified by ELISA (Human TNF alpha ELISA Ready-Set-Go, catalog number: 88-7346, eBioscience). The inter-assay coefficients of variability for all analytes were less than 10%.

2.3 Statistical analyses

Analyses of contingency Tables (χ²-test) or Fisher's exact probability test were employed to examine associations between grouping data. We assessed the differences in socio-demographic, clinical and biomarker data between groups using analyses of variance (ANOVAs) followed by the Tukey test to examine multiple comparisons among group means. Automatic stepwise binary logistic regression analysis with the MetS as dependent variable (and no MetS as reference group) and the O&NS biomarkers as explanatory variables was used to delineate the significant explanatory variables. Automatic, stepwise, multinomial logistic regression analysis was used to examine the significant predictors of groups divided according to the BMI into individuals with overweight and obesity. We employed univariate and multivariate general linear model (GLM) analyses to examine the relationships between the O&NS biomarkers, on the one hand and the metabolic and central obesity markers and the MetS, overweight and obesity, on the other. Exploratory factor analysis (principal component method with varimax rotation) was employed to interpret the associations between the O&NS biomarkers, the MetS, obesity / overweight and metabolic data. Normality of distribution was checked by means of the Kolmogorov–Smirnov goodness-of-fit test.
of-fit test. The Levene test was used to test for homogeneity of variance. We used logarithmic (Ln) transformations in case variables were not normally distributed or when there was a heterogeneity of variance between study groups. We used the IBM SPSS version 25 for windows to perform all statistical analyses. All tests were 2-tailed and a p-value of 0.05 was used for statistical significance.

To classify MetS patients and controls based on the biomarkers we used Support Vector Machine with linear kernel (linear SVM) or radial basis function (RBF SVM) (CAMO 2019). Input variables (biomarkers) were normalized with a standard deviation weighting process and the model was cross-validated using a 10-fold scheme. The classification data are summarized in the confusion matrix and using the cross-validated accuracy. Multilayer perceptron (MLP) neural network (NN) was used to delineate the associations between biomarkers (input variables) and MetS versus controls (output variables) using an automated feedforward architecture. Using SPSS25, we trained models and used two hidden layers with up to 8 nodes and 200 epochs, minibatch training with gradient descent, and with one consecutive step with no further decrease in the error term as stopping rule. We allocated cases to a training set (46.7%) to estimate the network parameters, a testing set (20%) to prevent overtraining, and a holdout sample (33.3%) to evaluate the predictive value of the network. Error, relative error, the importance of the input variables and area under the ROC curve with sensitivity and specificity were computed.

Soft Independent Modeling of Class Analogy or Statistical Isolinear Multiple Component Analysis (SIMCA) was employed as a class modeling technique (CAMO 2019). Principal component analysis (PCA) models were used to construct PCA models of both MetS patients and controls in a training set (50% of MetS patients and controls) and the number of PCs used to construct the models was determined by cross-validation. Outliers were eliminated based on Hotelling’s T2, influence and stability plots and evaluation of
residual values and leverages. In order to compute figures of merit (including the model-to-
model distance, the discriminatory power of the input variables, and the classification
accuracy) we projected cases allocated to the testing set (the remaining 50% of patients and
controls) into the PCA models.

RESULTS

Features of MetS

Table 1 shows the socio-demographic and clinical data of the subjects with and
without MetS. There were no significant differences in age, gender or TUD between both
groups. Body weight, BMI, waist circumference, systolic and diastolic blood pressure,
glucose, insulin and triglyceride levels were significantly higher in individuals with the MetS
as compared with those without. HDL-C levels were significantly lower in those with MetS.
There were no significant differences in total cholesterol or LDL-C between both groups.
MetS was associated with significantly higher uric acid and MDA levels. Table 1 shows that
there was a trend towards significantly higher NOx in MetS. The TRAP / uric acid ratio was
significantly lower in the MetS.

Results of logistic regression analyses predicting MetS.

Table 2 shows the results of automatic logistic regression analysis with MetS as
dependent variable (and controls as reference group). Regression #1 shows that MDA, NOx
(positively) and TRAP / uric acid ratio (inversely) were significantly associated with MetS
($\chi^2=23.81$, df=3, p<0.001, Nagelkerke=0.164; sensitivity=58.6% and specificity=66.0%).
Forced entry of age, gender and TUD showed that those factors were not associated with
MetS and that they did not change the significance levels of the three explanatory
biomarkers. After entering HDL-C in the regression (as an antioxidant with the other
oxidative and antioxidant biomarkers) we found that (regression #2) increased MDA and NOx levels combined with lowered HDL-C best predicted MetS with a huge effect size ($\chi^2=136.87$, df=3, p<0.001, Nagelkerke=0.780; sensitivity=83.9% and specificity=85.1%). Finally, regression #3 evaluates the combined effects of all oxidative and antioxidant biomarkers, BMI, and lipid and IR features of MetS. This regression shows that MetS was best predicted by MDA, BMI, glucose, triglycerides (all positively associated), HDL-C (inversely associated) while insulin showed a trend towards a positive association ($\chi^2=184.59$, df=6, p<0.001, Nagelkerke=0.853; sensitivity=91.5% and specificity=89.7%, overall accuracy=90.6%).

Results of SVM, SIMCA, PCA and NN

SVM with radial basis function and 10-fold cross-validation was employed for classification purposes. We found an accurate discrimination of MetS from controls with the biomarkers shown in table 2, regression #2 (except insulin) yielding a training accuracy of 93.82% and a validation accuracy of 89.89%. Figure 1 shows the statistic table as well as a 2D scatter plot of the discrimination of MetS and controls using MDA and HDL-C only, whereby patients and controls are color and shape-coded according to their predefined classes). Consequently, we have examined the separation of both treatment groups employing MDA, HDL-C, triglycerides, insulin, glucose and BMI as modelling and discriminatory variables in a SIMCA. We made a training set with 50% of all participants and a test set with the remaining participants. Two controls were omitted as statistical outliers, while all patients with MetS could be included. We modelled both classes of MetS patients and controls using 5 PCs. The model-to-model distance was highly significant, namely 158.68. All MetS cases in the test set were correctly authenticated except 3, while 12 controls intruded the MetS group limits (and thus are aliens). As such, cross-validation showed a sensitivity of 89.29%
Figure 2 shows the discrimination power of all input variables. The top-3 discriminatory variables were in descending order: glucose (14.1752), HDL-C (14.1664), MDA (14.1620), followed by BMI, and again at a distance by triglycerides and insulin.

Figure 3 shows the outcome of the best neural network discriminating MetS patients from controls using all immune and oxidative stress biomarkers as well as BMI and waist circumference. The feedforward network was trained with 2 hidden layers, with 6 units in layer 1 and 5 units in layer 2. As activation functions we used hyperbolic tangent in the hidden layers and identity in the output layer. The sum of squares error term was much lower in the testing (4.810) than in the training (10.628) set, while the percentage of incorrect classifications was even lower in the test (11.4%) than in the training (15.8%) set, indicating that the model learned to generalize from the trend. The AUC ROC was 0.932 and the holdout sample showed a sensitivity of 84.2% and specificity of 73.9%. Figure 3 shows the importance chart indicating that waist circumference, MDA, -SH groups and BMI have the highest predictive power of the model, followed at a distance by NOx, TRAP and PON1. The other input variables comprising gender, uric acid, LOOH and TNF-α have no or less predictive value.

**MetS versus IOTF classification**

Table 3 shows that there was a highly significant association between groups divided according to the IOTF and the MetS ($\chi^2=33.30$, df=2, $p<0.001$). All paired comparisons were significantly different from each other at $p<0.05$. More importantly, this table shows that some MetS patients have obesity (51.06%), overweight (34.04), or a normal BMI (14.89%), and that some subjects without MetS were classified as having obesity (11.70%) or overweight (27.78%).
In order to decipher the biomarkers which independently from IOTF diagnoses are associated with MetS, we conducted a multivariate GLM analysis examining the association between the classification into MetS and IOTF and the biomarkers, namely MDA, uric acid, SH groups, TRAP, glucose, insulin, HDL-C, and triglycerides (the other variables yielded non-significant results and are not shown). Table 4 shows that there are significant associations between the residualized biomarker levels and MetS. Figure 4 shows the residualized biomarker values after adjusting for IOTF, age, sex, and TUD and that MDA, SH, triglycerides, glucose, and insulin were higher in MetS, while HDL-C was significantly lower in MetS as compared with adolescents without MetS. Table 4 and Figure 5 shows that there was a significant association between the residualized biomarker data (after adjusting for MetS, age, sex and TUD) and the IOTF diagnosis whereby subjects with overweight and obesity show higher uric acid and triglyceride levels and lower HDL-C levels as compared with those without overweight or obesity.

4. DISCUSSION

The major findings of this study are that a) MetS in adolescents is characterized by increased MDA and -SH groups while increased uric acid is more specifically associated with overweight-obesity. Most importantly, machine learning techniques, including SIMCA, showed that MDA has a discriminatory power in separating MetS from controls, which is as strong as that of glucose, HDL-C, and BMI. Finally, our classification models including neural networks showed that MetS was significantly separated from controls with an AUC ROC curve of 0.932 using waist circumference, MDA, -SH groups, BMI and NOx as top-5 discriminatory variables.

Our findings extend those of previous studies showing a significantly increased aldehyde production in adults and adolescents with MetS. Venturini et al. (2012) observed
increased MDA in MetS but not in obese individuals, suggesting that MDA is associated with a cluster of cardiometabolic risks rather than with obesity per se [29]. On the other hand, Abdilla et al. (2007) found no relationship between MDA and MetS components and a weak association with BMI [30,31]. Other studies reported increased MDA levels in adolescents with obesity as compared with adolescents without obesity [32]. Several other studies found strong associations of MDA or TBARS with overweight/obesity or waist circumference (WC) [5,33,34]. In obese and overweight men and women, lipid peroxidation increases significantly with increasing BMI, while elevated MDA also occurs in obese adolescents and children between 8 and 18 years old [35]. In addition, obese children show MDA levels that are twice as high as those in non-obese children [35,36].

Nevertheless, using machine learning techniques, including SIMCA and neural networks, we detected that increased MDA was a key feature of MetS in adolescents along with glucose, HDL-C and BMI. It is interesting to note that in our study, MDA levels were not associated with obesity or overweight after considering the effects of MetS and that the association between MDA and MetS was not affected by overweight or obesity. Data in adolescents are most important because the results allow to estimate the pathways that are involved in the development of this condition. Consequently, our results show that increased aldehyde formation is a key phenomenon in adolescents with MetS and, therefore, that aldehyde formation starts at an early age and may participate in the development of the condition. Increased MDA formation indicates damage and toxicity to lipids as a consequence of increased ROS production, which play a key in endothelial dysfunction, oxidation of LDL-C, and the development of atherosclerosis, as well as contributing to insulin resistance and diabetes type 2 [37–41]. Moreover, serum MDA levels significantly correlate with advanced glycation and glycoxidation in diabetic children [42] leading to early occurrence of atherosclerotic changes [34]. Previous studies showed that the earliest signs of
cardiovascular heart disease, such as coronary artery fatty streaks, are already present in childhood and rapidly increase during adolescence, particularly in adolescents with elevated BMI [43].

Another characteristic of the MetS, albeit less significant than MDA, is the increase in -SH groups. These findings are not in agreement with our a priori hypothesis and with previous findings on lowered SH-groups in adult MetS [44]. Increased ROS may cause oxidation of -SH groups in albumin and consequently a depletion in their blood levels which, in turn, may exacerbate functional and structural disturbances of the damaged macromolecules [45]. It is possible that the increase in -SH groups in our adolescents with MetS may be explained by dysfunctions in the folate-dependent methionine cycle, including increased homocysteine levels in MetS [46]. It is also possible that the increases in -SH groups are a compensatory mechanism whereby -SH groups are being exposed due to protein unfolding (as a consequence of oxidative stress) followed by increases in -SH groups via thioredoxin-related mechanisms. Such compensatory mechanisms could be prominent in the early phase of disease as observed in our study, whereas a depletion of -SH groups may be detected in later phases of MetS [44].

Logistic regression and neural networks showed that increased NOx (when coupled with other data including MDA, -SH groups and HDL-C) contributes to the prediction of MetS. Chedraui et al (2012) reported that NO levels were higher in women with MetS in association with altered serum HDL-C, triglycerides and glucose levels. Serum NOx levels are also increased in adults with MetS in association with BMI, waist-to-hip ratio, and fasting plasma glucose levels [47]. Correia-Costa et al (2016) reported that NOx levels are increased in obese children and correlate with cardiometabolic risk, renal function and are positively associated with uric acid, suggesting that both iNOS and xanthine oxidase are activated in parallel and exacerbate the systemic proinflammatory and pro-oxidant status [48]. Codoner-
Franch et al. (2011) reported an increase in plasma nitrite, nitrate, and nitrotyrosine levels associated with markers of oxidative stress in obese children [49]. Gruber et al. (2008) found a decrease in NOx in obese juveniles compared to normal weight juveniles [8]. The measurement of serum NOx levels estimate basal NO generation by endothelial cells. Therefore, it may be speculated that increased serum NOx in the MetS may be due to endothelial NO synthase (eNOS) inhibition and inducible NOS (iNOS) overexpression [6]. Moreover, high glucose induces activity of endothelial NOS and NOx leading to overproduction of NO and ROS, respectively, which may cause nitrosative stress-mediated vascular endothelial cell dysfunctions [50]. NOx is believed to be a major player in endothelial dysfunction that influences vascular homeostasis and contributes towards development of vascular complications such as atherosclerosis [51]. The higher NOx levels found in our adolescents may well reflect a compensatory feedback mechanism present in the earlier stage of the syndrome [52].

We found that uric acid was significantly associated with overweight or obesity rather than with the MetS. Our findings extend these of previous authors showing that serum uric acid levels are increased in subjects with obesity, dyslipidemia, hypertension and impaired glucose tolerance, and are associated with the MetS and cardiovascular disease [53,54]. Most importantly, in a longitudinal study, Wang et al. [53] found that serum uric acid levels were significantly correlated with future increases in waist circumference, systolic blood pressure, and triglyceride and HDL-C plasma levels and, thus, with an increased risk of MetS. These results suggest that in adolescents, increased levels of uric acid are associated with obesity and consequently also with the MetS.

The interpretation of our TRAP findings is more difficult because uric acid (which is strongly related to obesity and BMI) is responsible for ~60% of plasma TRAP, showing that the increased uric acid levels in obesity strongly contribute to TRAP levels. Thus, when
TRAP values were normalized to uric acid or when the results are controlled for BMI we found that lowered TRAP concentrations were associated with MetS. These findings extend those of Molnár et al. (2004) who showed that TRAP was decreased in the multimetabolic syndrome group in spite of the slightly elevated concentrations of uric acid [13].

In the present study we found no significant association between lowered PON-1 activity and MetS or obesity, which contrasts our a priori hypothesis. PON-1 is a HDL-C associated enzyme, which plays a significant role in inhibiting the oxidation of HDL-C particles thereby contributing to most of the antioxidant activity of HDL-C [55]. Ferre et al (2013) reported that PON1 may play a role in the onset and development of metabolic alterations in childhood obesity [56]. In a juvenile population, decreased PON-1 activity is potentially relevant for the development of atherosclerosis and this may occur independently from effects on HDL-C [57]. However, some studies found contradictory results, hypothesizing that a regulatory increase of the antioxidant system could be present to compensate for the higher oxidative stress levels [48,58]. For example, in obese subjects, lowered PON-1 activity in HDL-C particles could be due to the presence of circulating inhibitors such as lipid peroxidation products [59].

This study has some strengths and limitations that must be considered in the interpretation of the results. This is a cross sectional study and, therefore, we cannot draw firm conclusions on causal associations. Strengths are that the results are analyzed with machine learning including deep learning which allow for a better prediction as well as feature description.

In conclusion, adolescents with MetS have a redox imbalance, characterized by increased aldehyde formation following lipid oxidation, increased NOx production, increased expression of -SH-groups, but lowered TRAP/uric acid levels, while overweight-obesity is characterized by increased uric acid. Moreover, increased MDA appears to be a key
phenomenon of MetS with a discriminatory power comparable to that of atherogenic and glucotoxicity biomarkers. The results suggest that early increases in lipid peroxidation and aldehyde formation are involved in the development of MetS.

Acknowledgements

The authors acknowledge the Health Sciences Postgraduate Program of the State University of Londrina, Brazil, and the Ministry for Sciences and Technology of Brazil (CNPq), Brazilian Federal Agency for Support and Evaluation of Graduate Education (CAPES).

Author Disclosure Statement

The authors declare no conflict of interest

Funding source

Ministry for Science and Technology of Brazil (CNPq).

REFERENCES


https://doi.org/10.1161/CIRCGENETICS.108.811638.


https://doi.org/10.1016/j.clinbiochem.2013.08.020.


https://doi.org/10.1172/JCI1649.
Table 1. Clinical and socio-demographic data, and biochemical, inflammatory and oxidative stress biomarkers in adolescents with and without metabolic syndrome (MetS).

<table>
<thead>
<tr>
<th>Variables</th>
<th>No MetS (n=94)</th>
<th>MetS (n=47)</th>
<th>F</th>
<th>df</th>
<th>p-values</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>13.02±1.53</td>
<td>13.04±1.71</td>
<td>0.006</td>
<td>1/139</td>
<td>0.940</td>
</tr>
<tr>
<td>Gender (M/F)</td>
<td>53 / 41</td>
<td>27 / 20</td>
<td>0.01</td>
<td>1</td>
<td>0.904</td>
</tr>
<tr>
<td>TUD (no / yes)</td>
<td>88 / 6</td>
<td>42 / 5</td>
<td>0.79</td>
<td>1</td>
<td>0.374</td>
</tr>
<tr>
<td>Body weight (kg)</td>
<td>54.73±11.23</td>
<td>68.63±17.08</td>
<td>33.47</td>
<td>1/139</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>22.03±3.59</td>
<td>26.97±4.80</td>
<td>46.98</td>
<td>1/139</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>WC (cm)</td>
<td>71.52±7.41</td>
<td>82.56±9.89</td>
<td>55.25</td>
<td>1/139</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Systolic blood pressure (mm Hg)</td>
<td>112.49±11.08</td>
<td>124.38±10.08</td>
<td>38.29</td>
<td>1/139</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Diastolic blood pressure (mm Hg)</td>
<td>64.54±6.99</td>
<td>70.65±7.29</td>
<td>23.24</td>
<td>1/139</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Glucose (mg/dL)</td>
<td>89.47±4.97</td>
<td>94.43±7.30</td>
<td>22.55</td>
<td>1/139</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Insulin* (µU/mL)</td>
<td>7.04±4.52</td>
<td>14.90±9.31</td>
<td>50.41</td>
<td>1/130</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Total cholesterol (mg/dL)</td>
<td>162.76±31.86</td>
<td>165.55±29.54</td>
<td>0.25</td>
<td>1/139</td>
<td>0.615</td>
</tr>
<tr>
<td>HDL-C (mg/dL)</td>
<td>49.19±11.23</td>
<td>34.15±5.71</td>
<td>74.43</td>
<td>1/139</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>LDL-C (mg/dL)</td>
<td>100.26±29.58</td>
<td>105.03±30.04</td>
<td>0.807</td>
<td>1/139</td>
<td>0.371</td>
</tr>
<tr>
<td></td>
<td>Mean ± SD</td>
<td>ANOVA p-value</td>
<td>df</td>
<td>Significance</td>
<td></td>
</tr>
<tr>
<td>----------------------</td>
<td>-----------------</td>
<td>---------------</td>
<td>------</td>
<td>--------------</td>
<td></td>
</tr>
<tr>
<td><strong>Triglycerides</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>(mg/dL)</td>
<td>66.53(±29.07)</td>
<td>131.87(±114.32)</td>
<td>43.77</td>
<td>1/139</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td><strong>NOx (uM/L)</strong></td>
<td>4.13(±1.57)</td>
<td>4.75(±2.15)</td>
<td>3.73</td>
<td>1/139</td>
<td>0.06</td>
</tr>
<tr>
<td><strong>LOOH (mM/L)</strong></td>
<td>0.75(±0.22)</td>
<td>0.77(±0.22)</td>
<td>0.15</td>
<td>1/139</td>
<td>0.69</td>
</tr>
<tr>
<td><strong>TRAP (uM Trolox)</strong></td>
<td>867.46(±118.13)</td>
<td>899.47(±131.70)</td>
<td>2.19</td>
<td>1/139</td>
<td>0.15</td>
</tr>
<tr>
<td><strong>Uric acid (mg/dL)</strong></td>
<td>4.66(±1.15)</td>
<td>5.48(±1.32)</td>
<td>12.75</td>
<td>1/122</td>
<td>0.001</td>
</tr>
<tr>
<td><strong>TRAP/Uric acid</strong></td>
<td>193.92(±40.03)</td>
<td>171.21(±33.40)</td>
<td>9.95</td>
<td>1/122</td>
<td>0.002</td>
</tr>
<tr>
<td><em><em>TNF-α</em>(pg/mL)</em>*</td>
<td>17.41(±27.87)</td>
<td>24.52(±67.49)</td>
<td>0.58</td>
<td>1/139</td>
<td>0.45</td>
</tr>
<tr>
<td><strong>PON-1 (U/mL)</strong></td>
<td>206.70(±52.11)</td>
<td>205.32(±45.50)</td>
<td>0.02</td>
<td>1/139</td>
<td>0.88</td>
</tr>
<tr>
<td><strong>SH-group (uM/mg PT)</strong></td>
<td>41.97(±6.19)</td>
<td>43.92(±7.37)</td>
<td>2.71</td>
<td>1/137</td>
<td>0.10</td>
</tr>
<tr>
<td><strong>MDA (uM/mg PT)</strong></td>
<td>99.50(±19.07)</td>
<td>109.61(±19.48)</td>
<td>8.61</td>
<td>1/137</td>
<td>0.004</td>
</tr>
</tbody>
</table>

All results are shown as mean (±SD). All results of ANOVAs.

*These variables are processed in Ln transformation.

TUD: tobacco use disorder; BMI: body mass index; WC: waist circumference; HDL-C: high-density lipoprotein cholesterol; LDL-C: low-density lipoprotein cholesterol.

Table 2. Results of binary logistic regression analysis with metabolic syndrome as dependent variable (healthy controls as reference group) and the inflammatory, oxidative stress and antioxidant biomarkers as explanatory variables.

<table>
<thead>
<tr>
<th>Explanatory Variables</th>
<th>B</th>
<th>S.E</th>
<th>Wald</th>
<th>df</th>
<th>p</th>
<th>Odds ratio</th>
<th>Lower CI 95%</th>
<th>Upper CI 95%</th>
</tr>
</thead>
<tbody>
<tr>
<td>#1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>NOx</td>
<td>0.331</td>
<td>0.165</td>
<td>4.03</td>
<td>1</td>
<td>0.046</td>
<td>1.39</td>
<td>1.01</td>
<td>1.93</td>
</tr>
<tr>
<td>TRAP/Uric acid</td>
<td>-0.514</td>
<td>0.183</td>
<td>7.85</td>
<td>1</td>
<td>0.005</td>
<td>0.60</td>
<td>0.42</td>
<td>0.86</td>
</tr>
<tr>
<td>MDA</td>
<td>0.471</td>
<td>0.168</td>
<td>7.89</td>
<td>1</td>
<td>0.005</td>
<td>1.60</td>
<td>1.15</td>
<td>2.22</td>
</tr>
<tr>
<td>#2</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>MDA</td>
<td>1.127</td>
<td>0.283</td>
<td>15.87</td>
<td>1</td>
<td>&lt;0.001</td>
<td>3.09</td>
<td>1.77</td>
<td>5.37</td>
</tr>
<tr>
<td>HDL-cholesterol</td>
<td>-3.720</td>
<td>0.595</td>
<td>39.10</td>
<td>1</td>
<td>&lt;0.001</td>
<td>0.02</td>
<td>0.01</td>
<td>0.08</td>
</tr>
<tr>
<td>NOx</td>
<td>0.505</td>
<td>0.233</td>
<td>4.69</td>
<td>1</td>
<td>0.030</td>
<td>1.66</td>
<td>1.05</td>
<td>2.62</td>
</tr>
<tr>
<td>#3</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>MDA</td>
<td>1.098</td>
<td>0.409</td>
<td>7.20</td>
<td>1</td>
<td>0.007</td>
<td>3.00</td>
<td>1.35</td>
<td>6.69</td>
</tr>
<tr>
<td>BMI</td>
<td>1.474</td>
<td>0.448</td>
<td>10.83</td>
<td>1</td>
<td>0.001</td>
<td>4.37</td>
<td>1.82</td>
<td>10.51</td>
</tr>
<tr>
<td>Glucose</td>
<td>1.574</td>
<td>0.441</td>
<td>12.72</td>
<td>1</td>
<td>&lt;0.001</td>
<td>4.83</td>
<td>2.03</td>
<td>11.47</td>
</tr>
<tr>
<td>Insulin</td>
<td>1.041</td>
<td>0.586</td>
<td>3.16</td>
<td>1</td>
<td>0.075</td>
<td>2.83</td>
<td>0.90</td>
<td>8.93</td>
</tr>
<tr>
<td>Triglycerids</td>
<td>1.315</td>
<td>0.478</td>
<td>7.56</td>
<td>1</td>
<td>0.006</td>
<td>3.72</td>
<td>1.46</td>
<td>9.51</td>
</tr>
<tr>
<td>HDL-cholesterol</td>
<td>-3.584</td>
<td>0.849</td>
<td>17.82</td>
<td>&lt;0.001</td>
<td>0.03</td>
<td>0.01</td>
<td>0.147</td>
<td></td>
</tr>
</tbody>
</table>

NOx: nitric oxide metabolites; TRAP: Total radical-trapping antioxidant parameter; MDA: malondialdehyde; BMI: body mass index.

All explanatory variables are entered in z scores

CI: confidence intervals.
Table 3. Association between the metabolic syndrome (MetS) and the International Obesity Task Force classification into subjects with a normal body mass index (BMI), overweight and obesity.

<table>
<thead>
<tr>
<th>Groups</th>
<th>Normal BMI</th>
<th>Overweight</th>
<th>Obesity</th>
</tr>
</thead>
<tbody>
<tr>
<td>No MetS</td>
<td>55 (58.51%)</td>
<td>28 (29.78%)</td>
<td>11 (11.70%)</td>
</tr>
<tr>
<td>MetS</td>
<td>7 (14.89%)</td>
<td>16 (34.04%)</td>
<td>24 (51.06%)</td>
</tr>
</tbody>
</table>
Table 4. Results of multivariate GLM analysis which examine the association between nitro-oxidative biomarkers and diagnoses.

<table>
<thead>
<tr>
<th>Tests</th>
<th>Dependent variables</th>
<th>Explanatory variables</th>
<th>F</th>
<th>df</th>
<th>p</th>
<th>Partial η²</th>
</tr>
</thead>
<tbody>
<tr>
<td>Multivariate*</td>
<td>MDA, uric acid, SH groups, TRAP, glucose, insulin, HDL-C, and triglycerides.</td>
<td>MetS</td>
<td>13.39</td>
<td>8/127</td>
<td>&lt;0.001</td>
<td>0.459</td>
</tr>
<tr>
<td></td>
<td></td>
<td>IOTF</td>
<td>2.75</td>
<td>16/256</td>
<td>&lt;0.001</td>
<td>0.147</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Sex</td>
<td>3.88</td>
<td>8/127</td>
<td>&lt;0.001</td>
<td>0.197</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Age</td>
<td>3.15</td>
<td>8/127</td>
<td>0.003</td>
<td>0.166</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Smoking</td>
<td>1.00</td>
<td>8/127</td>
<td>0.436</td>
<td>0.059</td>
</tr>
<tr>
<td>Univariate*</td>
<td>Glucose</td>
<td>MetS</td>
<td>34.26</td>
<td>1/134</td>
<td>&lt;0.001</td>
<td>0.204</td>
</tr>
<tr>
<td></td>
<td>Insulin</td>
<td>MetS</td>
<td>26.87</td>
<td>1/134</td>
<td>&lt;0.001</td>
<td>0.167</td>
</tr>
<tr>
<td></td>
<td>HDL-C</td>
<td>MetS</td>
<td>39.01</td>
<td>1/134</td>
<td>&lt;0.001</td>
<td>0.225</td>
</tr>
<tr>
<td></td>
<td>Triglycerides</td>
<td>MetS</td>
<td>22.67</td>
<td>1/134</td>
<td>&lt;0.001</td>
<td>0.145</td>
</tr>
<tr>
<td></td>
<td>MDA</td>
<td>MetS</td>
<td>10.03</td>
<td>1/134</td>
<td>0.002</td>
<td>0.070</td>
</tr>
<tr>
<td></td>
<td>-SH</td>
<td>MetS</td>
<td>5.08</td>
<td>1/134</td>
<td>0.026</td>
<td>0.037</td>
</tr>
<tr>
<td>Univariate**</td>
<td>HDL-C</td>
<td>IOTF</td>
<td>9.76</td>
<td>1/135</td>
<td>0.002</td>
<td>0.067</td>
</tr>
<tr>
<td></td>
<td>IOTF</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>----------------</td>
<td>------</td>
<td>-----</td>
<td>-----</td>
<td>-----</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Triglycerides</td>
<td>5.99</td>
<td>1/135</td>
<td>0.016</td>
<td>0.042</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Uric acid</td>
<td>13.25</td>
<td>1/135</td>
<td>&lt;0.001</td>
<td>0.089</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

All results of GLM analysis with age, sex, and smoking as covariates. *with MetS (no MetS versus MetS) and IOTF (3 groups: no obesity/overweight versus overweight versus obesity) as classes, ** with MetS (no MetS versus MetS) and IOTF (2 groups: no overweight versus overweight/obesity) as
Figure 1. Results of Support Vector Machine and 2D scatter plot of the discrimination of metabolic syndrome (red squares) and controls (blue squares) using MDA and HDL-C.
Figure 2. Results of Soft Independent Modeling of Class Analogy (SIMCA) showing the discrimination power of the input variables. The top-4 discriminatory variables are in descending order: glucose, HDL-C, MDA, and body mass index (BMI) followed at a distance by triglycerides and insulin.
Figure 3. Results of a neural network (importance chart) discriminating metabolic syndrome from controls. WC (waist circumference), MDA (malondialdehyde), -SH (thiol) groups and body mass index (BMI) have the highest predictive power of the model followed at a distance
by NOx (nitric oxide metabolites), TRAP (total radical-trapping potential of plasma), and paraoxonase-1 (PON1). Sex, uric acid, LOOH (lipid hydroperoxides), and TNF-α (tumor necrosis factor-α) have less predictive value.
Figure 4. Malondialdehyde (MDA), thiol (-SH) groups, triglycerides, glucose, and insulin are significantly higher and high density lipoprotein (HDL) cholesterol significantly lower in metabolic syndrome (MetS) as compared with healthy controls (HC) after adjusting for
obesity/overweight, age, sex, and tobacco use disorder (TUD). No differences in the other biomarkers were detected including TRAP (total radical trapping parameter of plasma).
Figure 5. Uric acid and triglycerides are significantly higher and high density lipoprotein (HDL) cholesterol significantly lower in overweight and obesity as compared with healthy controls (HC) after adjusting for metabolic syndrome, age, sex, and tobacco use disorder (TUD). No differences in the other biomarkers were detected including MDA (malondialdehyde), -SH (thiol) groups, glucose and insulin.