CONVALESCENT PLASMA THERAPY FOR COVID-19.

Focosi Daniele[1, #]

Anderson Arthur O.[2]

Tang Julian W[3]

Tuccori Marco[4, 5]

[1] North-Western Tuscany Blood Bank, Pisa University Hospital, Italy.

[2] Formerly the Chief, Department of Respiratory Mucosal Immunity, US Army Medical Research Institute of Infectious Diseases, Frederick, USA

[3] Respiratory Sciences, University of Leicester, Leicester, UK.

[4] Division of Pharmacology and Pharmacovigilance, Department of Clinical and Experimental Medicine, University of Pisa, Italy

[5] Unit of Adverse Drug reaction Monitoring, Pisa University Hospital, Italy.

[#] Corresponding author: daniele.focosi@gmail.com . Via Paradisa 2, 56124 Pisa, Italy. Phone/fax : +39 050 996541.

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Abbreviations : ADE : antibody-dependent enhancement; CBP : convalescent blood product; COVID-19 : coronavirus disease 2019; CP : convalescent plasma; CWB : convalescent whole blood; ELISA : enzyme-linked immunosorbent assay; EVD : Ebolavirus disease; IVIG : intravenous immunoglobulins; MERS : Middle-East respiratory syndrome; PRNT : plaque reduction neutralization test SARS : severe acute respiratory syndrome; TRALI : transfusion-related acute lung injury ; TTI : transfusion-transmitted infection.

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Abstract

Convalescent blood product therapy has been introduced since early 1900s to treat emerging infectious disease based on the evidence that polyclonal neutralizing antibodies can reduce duration of viremia. Recent large outbreaks of viral diseases for whom effective antivirals or vaccines are still lacking has revamped the interest in convalescent plasma as life-saving treatments. This review summarizes historical settings of application, and surveys current technologies for collection, manufacturing, pathogen inactivation, and banking, with a focus on COVID-19.

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Introduction

Emerging viruses rarely provide time to develop vaccines, and prophylactic vaccines are rarely effective in therapeutic setting. Antivirals are currently available only for selected viral families, are often not affordable to developing countries, and their manufacturing is hard to scale up in short times.

Recent viruses with pandemic potential include flaviviruses (e.g. West Nile virus (WNV), dengue virus, Zika virus (1)), chikungunya virus (2), influenzaviruses A, e.g. A(H1N1), A(H5N1) (3), Ebola virus (EBOV) (4), and respiratory betacoronaviruses (SARS-CoV (5), MERS-CoV (6), and SARS-CoV2 (7)).

Transfusion of convalescent blood products (CBP), especially convalescent plasma (CP), are useful against emerging infectious agents if the latter induces neutralizing antibodies (8). CBPs are manufactured by sampling whole blood or apheresis plasma from a convalescent donor: donor selection should be based according to neutralizing antibody titer, but in resource-poor settings, ELISA or no selection at all has often been implemented. The donor should preferably live in the same area as the intended recipient(s) to consider mutations of the target viral antigens, even if in areas epidemic for other infectious diseases (e.g. malaria) this could represent a contraindication. Although the recipient is already infected, theoretically transmission of more infectious particles could worsen clinical conditions. For this reason, the right timing of collection is fundamental to ensure no transmission of the pathogen to the recipient. Nevertheless, such concern can be somewhat reduced by treatment with modern pathogen inactivation (PI) techniques.

The main accepted mechanism of action for CBP therapy is clearance of viraemia, which typically happens 10–14 after infection (9). So CBP has been typically administered after early symptoms to maximize efficacy. Concurrent treatments might synergize or antagonize CP efficacy (e.g. polyclonal intravenous immunoglobulins or steroids) (10).

In the setting of respiratory viral infections, secretory IgA, which are the main immunoglobulin isotype on mucosal surfaces, are key players. They are made of 2 IgA molecules (dimers), a joining protein (J chain), and a secretory component. IgM and IgA are actively transported across epithelia by the polymeric Ig

receptor (pIgR) or by neonatal Fc receptor (FcRn), while IgG can passively trasudate into alveolar fluids (11). The lung requires specific antiviral IgG_{2a} for protection in terminal bronchioles and alveoli (12, 13).

Given the emergency related to the COVID-19 pandemic, this review summarized historical settings of application, and surveys current technologies for collection, manufacturing, pathogen inactivation, and banking, of convalescent blood products, with a specific focus on possible applications for COVID-19.

Convalescent plasma and pathogen inactivation

Convalescent whole blood (CWB), in addition to antibodies, provides control of hemorrhagic events, as in Ebolavirus disease, if transfusion occurs within 24 hours in order to keep viable platelets and clotting factors. Nevertheless, convalescent plasma (CP) best fits developed countries standards and settings where antibodies only required. CP should be collected by apheresis in order to ensure larger volumes, more frequent donations, and do not cause unnecessary anemia in the donor.

Technologies to virally reduce plasma (pathogen inactivation)

In several settings donor screening and conventional NAT viral testing (i.e. HIV, HCV and HBV NAT) could not be enough to ensure CP safety. In those scenario, pathogen reduction technologies (PRT) should be used. Several technologies have been approved and are currently marketed.

Solvent/detergent (S/D)-filtered plasma provides quick > 4 logs inactivation of most enveloped viruses: although the technology was developed and is massively used for large plasma pools, small scale reduction have been reported. The technology relies over 1% tri (n-butyl) phosphate/1% Triton X-45, elimination of solvent and detervent via oil extraction and filtration, and finally sterile filtration (14). Filtration across 75–35 nm hollow fibers could remove large viruses while preserving IgG [48], but has not been implemented yet.

In recent years photo-inactivation in the presence of a photosensitizer has become the standard for single unit inactivation : approved technologies include combination of methylene blue + visible light (15)

(Theraflex®), amotosalen (S-59) + ultraviolet A (16) (Intercept®), and riboflavin + ultraviolet B (17) (Mirasol®). These methods do not to affect immunoglobulin activity.

Fatty acids are also an option. In 2002 it was reported that caprylic acid (18) and octanoic acid (19) were as effective as S/D at inactivating enveloped viruses.

Heat-treatment of plasma has been used in the past (20, 21) but goes with the risk of aggregation of immunoglobulins (22, 23).

Pooling

Large-pool products

Pharmaceutical-grade facilities typically pool 100/2500 donors to manufacture S/D-inactivated plasma Intravenous immunoglobulins (IVIGs) are similarly prepared from pools of 2000–4000 L of plasma (or 100-1000 L in the case of hyperimmune IVIG) (24) (25). Such size can be hardly matched from CP donors and facilities rearrangement poses hard GMP issues (25).

Mini-pool fractionation scale (MPFS) into immunoglobulins

In order to be economically sustainable contract fractionation typically requires well over 10 000 liters of plasma per year, and domestic fractionation typically over 100 000–200 000 liters per year in addition to start-up a fractionation facility. A "on the bench" MPFS process (5-10 liters of plasma, i.e. approximately 20 recovered plasma units) using disposable devices and based on caprylic acid precipitation is under development in Egypt since 2003, and has been proven effective at purifying coagulation factors (26) and immunoglobulins (6-fold enrichment) (27). The same disposable bag system has also been combined with S/D reduction (14).

Lessons from SARS

SARS-specific antibodies usually persist for 2 years, and decline in prevalence and titers occurs in the third year (28). Convalescent anti-SARS immunoglobulins were manufactured on a small scale (8, 29). Three

infected healthcare workers with SARS progression despite treatment survived after transfusion with 500 ml CP : viral load dropped to zero one day after transfusion (30). SARS-CoV RNA was found in respiratory specimens from one third of patients for up to 4 weeks following symptoms (31). Amotosalen photochemical inactivation of apheresis platelet concentrates demonstrated a >6.2 log10 mean reduction of SARS-CoV (32). Theraflex˚ reduces infectivity of SARS-CoV in plasma (33). Heating at 60°C for 15-30 minutes reduces SARS-CoV from plasma without cells (34), while 60°C for 10 hours is required for plasma products (35). In addition, SARS-CoV was found to be sensitive to S/D, (34, 36).

Lessons from MERS

Antibody responses to MERS persist for less than 1 year and magnitude correlates with the duration of viral RNA shedding in sputum (but not with viral load). Mild patients have very low titers, making CP collection challenging in MERS convalescents (37). A study reported that only 2.7% (12 out of 443) exposed cases tested positive with ELISA, and only 75% of them had reactive microneutralization assay titers (38). CP with a PRNT titre ≥1:80 provide clinical benefit in MERS (39). A case of TRALI following CP transfusion in a patient with MERS was reported (40, 41). MERS-CoV load in plasma was reduced by Theraflex® (42), Intercept® (43), Mirasol® (44), and 56°C heating for 25 minutes (45) : in all cases passaging of inactivated plasma in replication-competent cells showed no viral replication.

Convalescent plasma for COVID-19

As soon as the COVID-19 pandemic appeared (7, 46), several authors suggested CP as a potential therapeutic (47, 48). Of interest, the most critically ill patients show prolonged viremia (strongly correlated with serum IL-6 levels) (49), which leaves room for therapeutic intervention with antivirals and immunoglobulins even in late stages. Viral shedding in survivors can be as long as 37 days (46), mandating SARS-CoV2 RNA screening in CP donors. Appearance of serum IgM and IgA antibody in COVID-19 occurs since day 5 after symptom onset, while IgG is detected since day 14 (50, 51). IgG are universally detected since day 20 (52). Severe female patients generate IgG earlier and higher titers (53). Duration of anti-SARS-CoV2 antibodies in plasma remains unknown, though for other betacoronaviruses immunity typically lasts

6-12 months (54). So a suitable donor could donate 600 ml plasma (equivalent to 3 therapeutic doses) every 14 days for a minimum of 6 months. In contrast to EVD, SARS, and MERS, most COVID-19 patients exhibit few or no symptoms and do not require hospitalization, suggesting that the majority of convalescent donors are best sought after in the general population.

In a first case series from China, 5 patients under mechanical ventilation (4 of 5 with no preexisting medical conditions) received transfusion with CP with a ELISA IgG titer > 1:1000 and a neutralization titer > 40 t day 10-22 after admission. 4 patients recovered from ARDS and 3 were weaned from mechanical ventilation within 2 weeks of treatment, the remaining being stable(55).

Another Chinese pilot study (ChiCTR2000030046) on 10 critically ill patients showed that one dose of 200 mL CP with neutralizing antibody titers > 1:640 resulted in an undetectable viral load (70%), radiological and clinical improvement (56).

Table 1 lists the other ongoing CP trials in COVID-19 patients listed in World Health Organization International Clinical Trial Registry Platform (ICTRP) database. The US have developed a specific platforms for facilitating clinical trials (https://ccpp19.org/), while the International Society for Blood Transfusion created a resource library (https://isbtweb.org/coronaoutbreak/covid-19-convalescent-plasma-document-library/) . Unfortunately, most trial in Westernized countries (on the contrary of the ones ongoing in China) seem to no have no control arm, which will impair efficacy interpretation.

 Notably, several plasma manufacturers are attempting to develop SARS-CoV2-specific hyperimmune sera, (e.g. Takeda's TAK-888 merge with Biotest, BPL, LFB, Octpharma and CSL Behring into the "Convalescent Plasma Coalition" (57), or Kamada's anti-COVID19 IgG (56)), while other companies are investing on genetic engineering (e.g. CSL Behring on SAB Biotherapeutics DiversitAb™ platform).

CP donor recruitment strategies

As previously proofed, donor testing for neutralizing antibodies is mandatory in upstream donor selection. Three approaches are theoretically available to recruit CP donors, everyone having pros and cons. The least cost-effective approach is screening the general periodic donor population for presence of anti-SARS-CoV2

antibodies. In endemic areas, this strategy provides many fit donors with the additional benefit of a seroprevalence study in the general population (80% of cases being asymptomatic), but requires a high budget. On the other side of the coin, recruitment of hospital discharged patients is highly cost-effective (patients can be easily tested before discharge and tracked), but patients who have required hospitalization are highly likely to be elderlies with comorbidities, and hence unfit to donate. The intermediate approach is deploying calls to donate to positive cases under home-based quarantine: given the huge numbers, some of them are likely to be periodic donors, and home-based convalescence suggests they are fit enough to donate. Nevertheless, lessons from MERS suggest that patients with mild symptoms could have developed low-titer antibodies (38), making antibody titration even more important in the population-wide and home-based approaches.

CP banking

CP is typically used as a fresh product. Aliquots can be easily achieved with modern PI kits. Banking at temperature below -25˚C (according to EDQM guidelines for ordinary plasma for clinical use (58)) is encouraged in order to translate CP in an off-the-shelf, ready-to-use product. Most regulatory system require that CP is tracked informatically as a blood component different from ordinary plasma for clinical use. The final validation label should report that the donor has tested negative at PCR for the convalescent disorder and additional microbiological tests, and describe the inactivation method. There is no evidence that a single cycle of freezing and thawing significatively affects quantity or function of immunoglobulins.

Monitoring response to treatment

CP is considered an experimental therapy, and as such phase 3 randomized controlled trials should be encouraged. Despite this recommendation, in emergency settings phase 2 trials are usually started, hampering efficacy analysis. Response in published trials is generally measured clinically or radiologically according to target organs. Nevertheless, surrogate endpoints can include antibody titer rise in recipient's

plasma and drops in recipient's viral load. Whenever quantitative PCR is not available, cycle threshold (Ct) value increases in qualitative PCR after transfusion could be a proxy for reduced viral load.

Side benefits from CP in COVID-19

Obviously, patients with humoral immune deficiencies can benefit from polyclonal antibodies contained in CP, and patients with hemorrhagic diathesis can benefit from clotting factors. After demonstration that group 0 healthcare workers were less likely to become infected with SARS-CoV (59), a research group proved that anti-A blood group natural isoagglutinins (which can be also found in CP plasma from blood group 0 and B donors) inhibit SARS-CoV entry into competent cells (60). Such binding could opsonize virions and induce complement-mediated neutralization (61). Since SARS-CoV2 uses the same receptor as SARS-CoV, anti-A isoagglutinins are expected to have similar effects against SARS-CoV2: accordingly clusters of glycosylation sites exist proximal to the receptor-binding motif of the SARS-CoV (62) and SARS-CoV2 (63) S protein.

A recent publication showed that the odds ratio for acquiring COVID-19 is higher in blood group A than in blood group 0 (64). COVID-19 has more severe clinical presentations and outcome in elderlies and in males : intriguingly, elderly males are known to experience reductions in isoagglutinin titers (65, 66). Studies are hence ongoing to evaluate correlations between isoagglutinin titers and outcome in blood group 0 and B patients. In the meanwhile, while preserving ABO match compatibility, it could be wise to prefer blood group 0 and B donors for CP in COVID-19, and to titre their anti-A isoagglutinins.

Concerns

The main contraindications to CP therapy are allergy to plasma protein or sodium citrate, or selective IgA deficiency (< 70 mg/dl in patients 4 years old or greater). As in many other trial settings, concurrent viral or bacterial infections, thrombosis, poor compliance, short life expectancy (e.g. multiple organ failure), as well as pregnant or breastfeeding women. are also contraindications. Nevertheless, additional concerns apply.

The first concern is transfusion-transmitted infection (TTI). Modern PI technologies, combined with NAT, reduces the risk for contracting additional TTIs. Most regulatory systems require additional tests (e.g. HAV RNA, HEV RNA, parvovirus B19 DNA) to be performed on CP for additional transfusion safety. CBP obtained from donors in the UK may be problematic for a couple of reasons. Currently CBP obtained from individuals who lived for at least 6 months in the UK during 1980-1996 'mad cow disease (bovine spongiform encephalopathy – BSE)' outbreak may not be acceptable in some countries (67) – or by some individuals. In addition, there is a now a recognized risk of hepatitis E the within UK blood donor population (68), most likely due to the consumption of poorly cooked pork products (69, 70), for which screening has only relatively recently been initiated(71). Although this does not preclude such SARS-CoV-2 convalescent plasma/sera being used therapeutically within the UK, these other risks should be considered during larger clinical trial or individual patient compassionate use. As per the risk of worsening the clinical picture by delivering more viral particles of the targeted virus, it is generally unlikely to worsen the underlying scenario. Respiratory betacoronaviruses produce only a mild and transient viremia. With SARS-CoV, limited replication in lymphocytes(72) leads to significant risk only for recipients of blood products with high concentrations of donor lymphocytes (peripheral blood stem cells, bone marrow, granulocyte concentrates, etc). With SARS-CoV2, viremia has been shown persists only in critically ill patients (49).

The second concern is TRALI, which can be life-threatening in patients who already are suffering from ALI. Male donors are usually preferred in order to avoid the risk of transfusing anti-HLA antibodies from parous women. In the case of COVID-19, where female patients have been shown to have higher IgG levels, this could be detrimental, and anti-HLA antibody screening could be implemented.

Antibody-dependent enhancement (ADE) due to passive or active antibodies facilitating coated virions entry into cells via Fc receptors (73, 74) is also a theoretical concern, but its clinical relevance remains unproven (75).

Conclusions

CP manufacturing should be considered among the first responding actions during a pandemic in the meanwhile antivirals and vaccines are tested. Despite huge competition from trials employing small chemicals, multicentre randomized controlled trials should be encouraged in order to establish efficacy and provide hints about the most effective schedule (timing and dose).

Figure 1. Summary of possible convalescent blood products (CBP). Reproduced from ref (76) under STM

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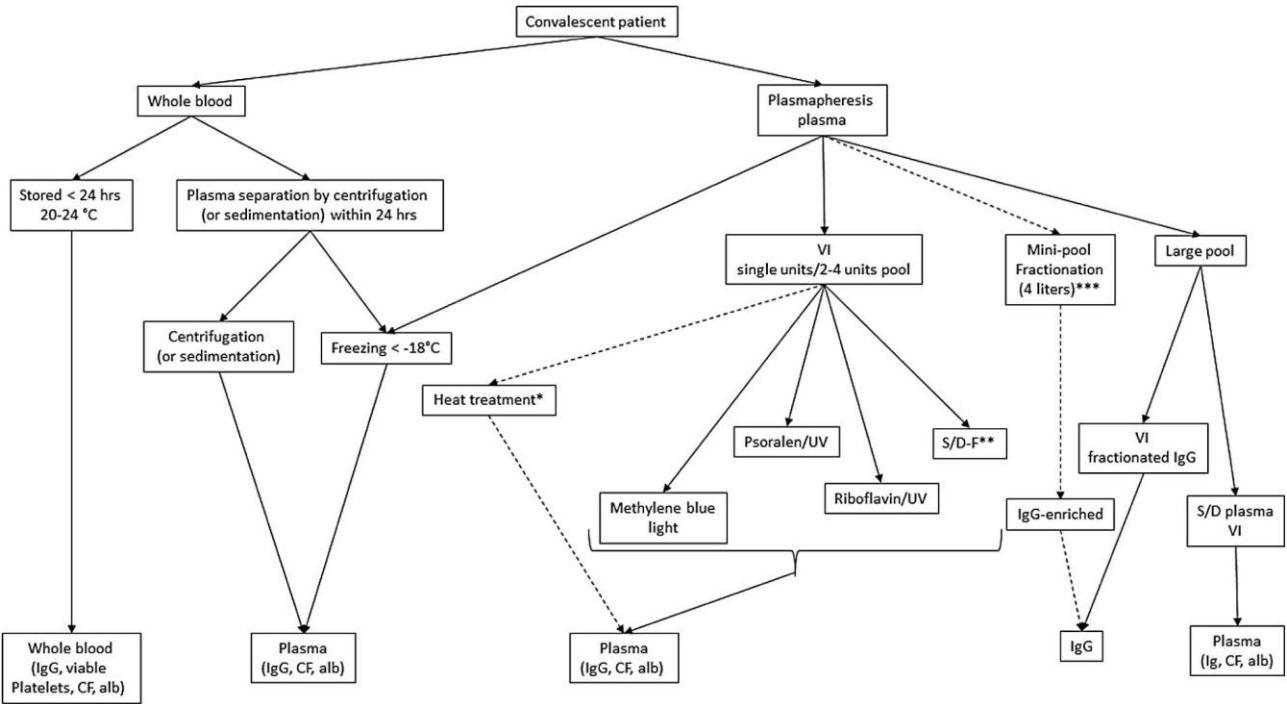
1   **Table 1.** Ongoing interventional clinical trials of convalescent plasma in COVID-19 patients listed in World Health Organization International Clinical Trial

2   Registry Platform (ICTRP) databases (accessed online at https://www.who.int/docs/default-source/coronaviruse/covid-19-trials.xls on April 6, 2020)

Trial number	Title (country)	Study population	Schedule	Donor Titer	Indication
ChiCTR2000029850	Study on convalescent plasma treatment for severe patients with novel coronavirus pneumonia (COVID-19) (China)	Exp:10 Ctr:10	NA	NA	Clinical deterioration despite conventional treatment that required intensive care
ChiCTR2000030179	Experimental study of novel coronavirus pneumonia rehabilitation plasma therapy severe novel coronavirus pneumonia (COVID-19) (China)	Exp:50 Ctr:50	NA	NA	Critically ill patients
ChiCTR2000030010	A randomized, double-blind, parallel-controlled, trial to evaluate the efficacy and safety of anti-SARS-CoV-2 virus inactivated plasma in the treatment of severe novel coronavirus pneumonia patients (COVID-19) (China)	Exp:50 Ctr:50	NA	NA	Non critically ill patients

ChiCTR2000030039	Clinical study for infusing convalescent plasma to treat patients with new coronavirus pneumonia (COVID-19) (China)	Exp:30 Ctr:60	2 units of plasma (200/500 mL/24h) *vs* BSC	NA	All patients
ChiCTR2000030627	Study for using the healed novel coronavirus pneumonia (COVID-19) patients plasma in the treatment of severe critical cases (China)	Exp:15 Ctr:15	NA	NA	Severe or critically ill patients
ChiCTR2000029757	Convalescent plasma for the treatment of severe and critical novel coronavirus pneumonia (COVID-19): a prospective randomized controlled trial (China)	Exp:100 Ctr:100	NA	NA	Severe or critically ill patients
NCT04292340	Anti-SARS-CoV-2 Inactivated Convalescent Plasma in the Treatment of COVID-19 (China)	15	NA	NA	All patients with Covid-19
ChiCTR2000030702	Plasma of the convalescent in the treatment of novel coronavirus pneumonia (COVID-19) common patient: a prospective clinical trial (China)	Exp:25 Ctr:25	NA	NA	Non critically ill patients
ChiCTR2000030929	A randomized, double-blind, parallel-	Exp:30	NA	NA	Non critically ill patients

	controlled trial to evaluate the efficacy and safety of anti-SARS-CoV-2 virus inactivated plasma in the treatment of severe novel coronavirus pneumonia (COVID-19) (China)	Ctr:30			
NCT04321421	Hyperimmune Plasma for Critical Patients With COVID-19 (COV19-PLASMA) (Italy)	49	3 units of plasma (250-300 mL/48h)	NA	Moderate to severe ARDS under mechanical ventilation
NCT04323800	Efficacy and Safety Human Coronavirus Immune Plasma (HCIP) vs. Control (SARS-CoV-2 Non-immune Plasma) Among Adults Exposed to COVID-19 (CSSC-001) (USA)	150	1 unit of plasma (200/250mL)	>1:64	Exposed to the contagion (within 96 hours of enrollment and 120 hours of receipt of plasma)
NCT04325672	Convalescent Plasma to Limit Coronavirus Associated Complications: An Open Label, Phase 2A Study of High-Titer Anti-SARS-CoV-2 Plasma in Hospitalized Patients With COVID-19 (USA)	20	1-2 units of plasma (300 mL/24h)	>1:64	Severe or critically ill patients
NCT04333251	Evaluating Convalescent Plasma to Decrease Coronavirus Associated Complications. A	115 Exp: NA	1-2 units of plasma (250 mL/24h) *vs* BSC	>1:64	All patients with COVID-19

	Phase I Study Comparing the Efficacy and Safety of High-titer Anti-Sars-CoV-2 Plasma vs Best Supportive Care in Hospitalized Patients With Interstitial Pneumonia Due to COVID-19 (USA)	Ctr: NA			
NCT04338360	Expanded Access to Convalescent Plasma for the Treatment of Patients With COVID-19 (USA)	NA	1 unit of plasma (200/250 mL)	NA	Critically ill patients
NCT04332380	Convalescent Plasma for Patients With COVID-19: A Pilot Study (CP-COVID-19) (Colombia)	10	2 units of plasma (250 mL/24h)	NA	Non critically ill patients. 250 ml day 1 + 250 ml day 2
NCT04332835	Convalescent Plasma for Patients With COVID-19: A Randomized, Open Label, Parallel, Controlled Clinical Study (Colombia)	40 Exp: NA Ctr: NA	2 units of plasma (250 mL/24h) *vs* BSC	NA	Non critically ill patients
NCT04327349	Investigating Effect of Convalescent Plasma on COVID-19 Patients Outcome: A Clinical Trial (Iran)	30	NA	NA	Non critically ill patients
NCT04333355	Phase 1 Study to Evaluate the Safety of	20	1-2 units of plasma (250	NA	Severe or critically ill patients

	Convalescent Plasma as an Adjuvant Therapy in Patients With SARS-CoV-2 Infection (Mexico)		ml/24h)		
NCT04340050	COVID-19 Convalescent Plasma (USA)	10	1 unit (300 ml)	NA	Severe or critically ill patients < 21 days from the start of illness
NCT04342182	Convalescent Plasma Therapy From Recovered Patients to Treat Severe SARS-CoV-2 Disease (CONCOVID Study) (The Netherlands)	426 Exp : NA Ctr: NA	1 unit (250 ml)	NA	All patients with COVID-19
BSC: best supportive care; NA: not available; Exp: experimental group; Ctr: control group					

References

1. **Guarner J, Hale GL.** 2019. Four human diseases with significant public health impact caused by mosquito-borne flaviviruses: West Nile, Zika, dengue and yellow fever. Semin Diagn Pathol **36:**170-176.

2. **Vairo F, Haider N, Kock R, Ntoumi F, Ippolito G, Zumla A.** 2019. Chikungunya: Epidemiology, Pathogenesis, Clinical Features, Management, and Prevention. Infect Dis Clin North Am **33:**1003-1025.

3. **Vogel OA, Manicassamy B.** 2020. Broadly Protective Strategies Against Influenza Viruses: Universal Vaccines and Therapeutics. Front Microbiol **11:**135.

4. **Hoenen T, Groseth A, Feldmann H.** 2019. Therapeutic strategies to target the Ebola virus life cycle. Nat Rev Microbiol **17:**593-606.

5. **Booth CM, Matukas LM, Tomlinson GA, Rachlis AR, Rose DB, Dwosh HA, Walmsley SL, Mazzulli T, Avendano M, Derkach P, Ephtimios IE, Kitai I, Mederski BD, Shadowitz SB, Gold WL, Hawryluck LA, Rea E, Chenkin JS, Cescon DW, Poutanen SM, Detsky AS.** 2003. Clinical features and short-term outcomes of 144 patients with SARS in the greater Toronto area. Jama **289:**2801-2809.

6. **Bermingham A, Chand MA, Brown CS, Aarons E, Tong C, Langrish C, Hoschler K, Brown K, Galiano M, Myers R, Pebody RG, Green HK, Boddington NL, Gopal R, Price N, Newsholme W, Drosten C, Fouchier RA, Zambon M.** 2012. Severe respiratory illness caused by a novel coronavirus, in a patient transferred to the United Kingdom from the Middle East, September 2012. Euro Surveill **17:**20290.

7. **Zhu N, Zhang D, Wang W, Li X, Yang B, Song J, Zhao X, Huang B, Shi W, Lu R, Niu P, Zhan F, Ma X, Wang D, Xu W, Wu G, Gao GF, Tan W.** 2020. A Novel Coronavirus from Patients with Pneumonia in China, 2019. N Engl J Med **382:**727-733.

8. **Mair-Jenkins J, Saavedra-Campos M, Baillie JK, Cleary P, Khaw FM, Lim WS, Makki S, Rooney KD, Nguyen-Van-Tam JS, Beck CR.** 2015. The effectiveness of convalescent plasma and hyperimmune immunoglobulin for the treatment of severe acute respiratory infections of viral etiology: a systematic review and exploratory meta-analysis. J Infect Dis **211:**80-90.

9. **Cheng Y, Wong R, Soo YO, Wong WS, Lee CK, Ng MH, Chan P, Wong KC, Leung CB, Cheng G.** 2005. Use of convalescent plasma therapy in SARS patients in Hong Kong. Eur J Clin Microbiol Infect Dis **24:**44-46.

10. **Luke TC, Kilbane EM, Jackson JL, Hoffman SL.** 2006. Meta-analysis: convalescent blood products for Spanish influenza pneumonia: a future H5N1 treatment? Ann Intern Med **145:**599-609.

11. **Roopenian DC, Akilesh S.** 2007. FcRn: the neonatal Fc receptor comes of age. Nat Rev Immunol **7:**715-725.

12. **Fazekas G, Rosenwirth B, Dukor P, Gergely J, Rajnavolgyi E.** 1994. IgG isotype distribution of local and systemic immune responses induced by influenza virus infection. Eur J Immunol **24:**3063-3067.

13. **Mostov KE.** 1994. Transepithelial transport of immunoglobulins. Annu Rev Immunol **12:**63-84.

14. **El-Ekiaby M, Sayed MA, Caron C, Burnouf S, El-Sharkawy N, Goubran H, Radosevich M, Goudemand J, Blum D, De Melo L, Soulié V, Adam J, Burnouf T.** 2010. Solvent-detergent filtered (S/D-F) fresh frozen plasma and cryoprecipitate minipools prepared in a newly designed integral disposable processing bag system. Transfusion Medicine **20:**48-61.

15. **Wong HK, Lee CK, Hung IF, Leung JN, Hong J, Yuen KY, Lin CK.** 2010. Practical limitations of convalescent plasma collection: a case scenario in pandemic preparation for influenza A (H1N1) infection. Transfusion **50:**1967-1971.

16. **Singh Y, Sawyer LS, Pinkoski LS, Dupuis KW, Hsu JC, Lin L, Corash L.** 2006. Photochemical treatment of plasma with amotosalen and long-wavelength ultraviolet light inactivates pathogens while retaining coagulation function. Transfusion **46:**1168-1177.

17. **Bihm DJ, Ettinger A, Buytaert-Hoefen KA, Hendrix BK, Maldonado-Codina G, Rock G, Giclas PC, Goodrich RP.** 2010. Characterization of plasma protein activity in riboflavin and UV light-treated fresh frozen plasma during 2 years of storage at −30°C. Vox Sanguinis **98:**108-115.

18. **Korneyeva M, Hotta J, Lebing W, Rosenthal RS, Franks L, Petteway SR.** 2002. Enveloped Virus Inactivation by Caprylate: A Robust Alternative to Solvent-Detergent Treatment in Plasma Derived Intermediates. Biologicals **30:**153-162.

19. **Dichtelmüller H, Rudnick D, Kloft M.** 2002. Inactivation of Lipid Enveloped Viruses by Octanoic Acid Treatment of Immunoglobulin Solution. Biologicals **30:**135-142.

20. **Goubran HA, Burnouf T, Radosevich M.** 2000. Virucidal heat-treatment of single plasma units: a potential approach for developing countries. Haemophilia **6:**597-604.

21. **Watt G, Kantipong P, Jongsakul K, de Souza M, Burnouf T.** 2001. Passive transfer of scrub typhus plasma to patients with AIDS: a descriptive clinical study. Qjm **94:**599-607.

22. **Vittecoq D, Mattlinger B, Barre-Sinoussi F, Courouce AM, Rouzioux C, Doinel C, Bary M, Viard JP, Bach JF, Rouger P, Lefrere JJ.** 1992. Passive Immunotherapy in AIDS: A Randomized Trial of Serial Human Immunodeficiency Virus-Positive Transfusions of Plasma Rich in p24 Antibodies versus Transfusions of Seronegative Plasma. The Journal of Infectious Diseases **165:**364-368.

23. **Vittecoq D, Chevret S, Morand-Joubert L, Heshmati F, Audat F, Bary M, Dusautoir T, Bismuth A, Viard JP, Barré-Sinoussi F.** 1995. Passive immunotherapy in AIDS: a double-blind randomized study based on transfusions of plasma rich in anti-human immunodeficiency virus 1 antibodies vs. transfusions of seronegative plasma. Proceedings of the National Academy of Sciences **92:**1195-1199.

24. **Radosevich M, Burnouf T.** 2010. Intravenous immunoglobulin G: trends in production methods, quality control and quality assurance. Vox Sanguinis **98:**12-28.

25. **WHO.** 2007. Annex 4. Recommendations for the collection, quality control and regulation of human plasma for fractionation. WHO Tech Rep Ser **941:**189-264.

26. **El-Ekiaby M, Radosevich M, Goubran H, El Sayed M, Burnouf T.** 2009. New methods of plasma fractionation – a presentation of the 'mini-pool' fractionation procedure developed in Egypt. ISBT Science Series **4:**99-106.

27. **El-Ekiaby M, Vargas M, Sayed M, Gorgy G, Goubran H, Radosevic M, Burnouf T.** 2015. Minipool caprylic acid fractionation of plasma using disposable equipment: a practical method to enhance immunoglobulin supply in developing countries. PLoS neglected tropical diseases **9:**e0003501-e0003501.

28. **Wu L-P, Wang N-C, Chang Y-H, Tian X-Y, Na D-Y, Zhang L-Y, Zheng L, Lan T, Wang L-F, Liang G-D.** 2007. Duration of antibody responses after severe acute respiratory syndrome. Emerging infectious diseases **13:**1562-1564.

29. **Zhang Z, Xie Y-W, Hong J, Zhang X, Kwok SY, Huang X, Wong SW, Wong B-L, Group S.** 2005. Purification of severe acute respiratory syndrome hyperimmune globulins for intravenous injection from convalescent plasma. Transfusion **45:**1160-1164.

30. **Yeh K, Chiueh T, Siu L.** 2005. Experience of using convalescent plasma for severe acute respiratory syndrome among healthcare workers in a Taiwan hospital. J Antimicrob Chemother **56:**919-922.

31. **Xu D, Zhang Z, Jin L, Chu F, Mao Y, Wang H, Liu M, Wang M, Zhang L, Gao GF, Wang FS.** 2005. Persistent shedding of viable SARS-CoV in urine and stool of SARS patients during the convalescent phase. Eur J Clin Microbiol Infect Dis **24:**165-171.

32. **Pinna D, Sampson-Johannes A, Clementi M, Poli G, Rossini S, Lin L, Vicenzi E.** 2005. Amotosalen photochemical inactivation of severe acute respiratory syndrome coronavirus in human platelet concentrates. Transfus Med **15:**269-276.

33. **Eickmann M, Gravemann U, Handke W, Tolksdorf F, Reichenberg S, Muller TH, Seltsam A.** 2020. Inactivation of three emerging viruses - severe acute respiratory syndrome coronavirus, Crimean-Congo haemorrhagic fever virus and Nipah virus - in platelet concentrates by ultraviolet C light and in plasma by methylene blue plus visible light. Vox Sang doi:10.1111/vox.12888.

34. **Darnell ME, Taylor DR.** 2006. Evaluation of inactivation methods for severe acute respiratory syndrome coronavirus in noncellular blood products. Transfusion **46:**1770-1777.

35. **Yunoki M, Urayama T, Yamamoto I, Abe S, Ikuta K.** 2004. Heat sensitivity of a SARS-associated coronavirus introduced into plasma products. Vox Sang **87:**302-303.

104 36. **Rabenau HF, Biesert L, Schmidt T, Bauer G, Cinatl J, Doerr HW.** 2005. SARS-coronavirus (SARS-CoV)
105   and the safety of a solvent/detergent (S/D) treated immunoglobulin preparation. Biologicals **33:**95-
106   99.
107 37. **Choe PG, Perera R, Park WB, Song KH, Bang JH, Kim ES, Kim HB, Ko LWR, Park SW, Kim NJ, Lau
108   EHY, Poon LLM, Peiris M, Oh MD.** 2017. MERS-CoV Antibody Responses 1 Year after Symptom
109   Onset, South Korea, 2015. Emerg Infect Dis **23:**1079-1084.
110 38. **Arabi YM, Hajeer AH, Luke T, Raviprakash K, Balkhy H, Johani S, Al-Dawood A, Al-Qahtani S, Al-
111   Omari A, Al-Hameed F, Hayden FG, Fowler R, Bouchama A, Shindo N, Al-Khairy K, Carson G, Taha
112   Y, Sadat M, Alahmadi M.** 2016. Feasibility of Using Convalescent Plasma Immunotherapy for MERS-
113   CoV Infection, Saudi Arabia. Emerg Infect Dis **22:**1554-1561.
114 39. **Ko JH, Seok H, Cho SY, Ha YE, Baek JY, Kim SH, Kim YJ, Park JK, Chung CR, Kang ES, Cho D, Muller
115   MA, Drosten C, Kang CI, Chung DR, Song JH, Peck KR.** 2018. Challenges of convalescent plasma
116   infusion therapy in Middle East respiratory coronavirus infection: a single centre experience. Antivir
117   Ther **23:**617-622.
118 40. **Chun S, Chung CR, Ha YE, Han TH, Ki CS, Kang ES, Park JK, Peck KR, Cho D.** 2016. Possible
119   Transfusion-Related Acute Lung Injury Following Convalescent Plasma Transfusion in a Patient With
120   Middle East Respiratory Syndrome. Ann Lab Med **36:**393-395.
121 41. **Arabi Y, Balkhy H, Hajeer AH, Bouchama A, Hayden FG, Al-Omari A, Al-Hameed FM, Taha Y,
122   Shindo N, Whitehead J, Merson L, AlJohani S, Al-Khairy K, Carson G, Luke TC, Hensley L, Al-
123   Dawood A, Al-Qahtani S, Modjarrad K, Sadat M, Rohde G, Leport C, Fowler R.** 2015. Feasibility,
124   safety, clinical, and laboratory effects of convalescent plasma therapy for patients with Middle East
125   respiratory syndrome coronavirus infection: a study protocol. Springerplus **4:**709.
126 42. **Eickmann M, Gravemann U, Handke W, Tolksdorf F, Reichenberg S, Muller TH, Seltsam A.** 2018.
127   Inactivation of Ebola virus and Middle East respiratory syndrome coronavirus in platelet
128   concentrates and plasma by ultraviolet C light and methylene blue plus visible light, respectively.
129   Transfusion **58:**2202-2207.
130 43. **Hindawi SI, Hashem AM, Damanhouri GA, El-Kafrawy SA, Tolah AM, Hassan AM, Azhar EI.** 2018.
131   Inactivation of Middle East respiratory syndrome-coronavirus in human plasma using amotosalen
132   and ultraviolet A light. Transfusion **58:**52-59.
133 44. **Keil SD, Bowen R, Marschner S.** 2016. Inactivation of Middle East respiratory syndrome
134   coronavirus (MERS-CoV) in plasma products using a riboflavin-based and ultraviolet light-based
135   photochemical treatment. Transfusion **56:**2948-2952.
136 45. **Leclercq I, Batejat C, Burguiere AM, Manuguerra JC.** 2014. Heat inactivation of the Middle East
137   respiratory syndrome coronavirus. Influenza Other Respir Viruses **8:**585-586.
138 46. **Zhou F, Yu T, Du R, Fan G.** 2020. Clinical course and risk factors for mortality of adult inpatients
139   with COVID-19 in Wuhan, China: a retrospective cohort study. Lancet.
140 47. **Casadevall A, Pirofski L-a.** 2020. The convalescent sera option for containing COVID-19. The Journal
141   of Clinical Investigation **130**.
142 48. **Chen L, Xiong J, Bao L, Shi Y.** 2020. Convalescent plasma as a potential therapy for COVID-19.
143   Lancet Inf Dis doi:https://doi.org/10.1016/S1473-3099(20)30141-9.
144 49. **Chen X, Zhao B, Qu Y, Chen Y, Xiong J, Feng Y, Men D, Huang Q, Liu Y, Yang B, Ding J, Li F.** 2020.
145   Detectable serum SARS-CoV-2 viral load (RNAaemia) is closely associated with drastically elevated
146   interleukin 6 (IL-6) level in critically ill COVID-19 patients. medRxiv
147   doi:10.1101/2020.02.29.20029520:2020.2002.2029.20029520.
148 50. **Guo L, Ren L, Yang S, Xiao M, Chang D, Yang F, Dela Cruz CS, Wang Y, Wu C, Xiao Y, Zhang L, Han L,
149   Dang S, Xu Y, Yang Q, Xu S, Zhu H, Xu Y, Jin Q, Sharma L, Wang L, Wang J.** 2020. Profiling Early
150   Humoral Response to Diagnose Novel Coronavirus Disease (COVID-19). Clinical Infectious Diseases
151   doi:10.1093/cid/ciaa310.
152 51. **Zhao J, Yuan Q, Wang H, Liu W, Liao X, Su Y, Wang X, Yuan J, Li T, Li J, Qian S, Hong C, Wang F, Liu
153   Y, Wang Z, He Q, Li Z, He B, Zhang T, Ge S, Liu L, Zhang J, Xia N, Zhang Z.** 2020. Antibody responses
154   to SARS-CoV-2 in patients of novel coronavirus disease 2019. medRxiv
155   doi:10.1101/2020.03.02.20030189:2020.2003.2002.20030189.

52. **Long Q-x, Deng H-j, Chen J, Hu J, Liu B-z, Liao P, Lin Y, Yu L-h, Mo Z, Xu Y-y, Gong F, Wu G-c, Zhang X-x, Chen Y-k, Li Z-j, Wang K, Zhang X-l, Tian W-g, Niu C-c, Yang Q-j, Xiang J-l, Du H-x, Liu H-w, Lang C, Luo X-h, Wu S-b, Cui X-p, Zhou Z, Wang J, Xue C-j, Li X-f, Wang L, Tang X-j, Zhang Y, Qiu J-f, Liu X-m, Li J-j, Zhang D-c, Zhang F, Cai X-f, Wang D, Hu Y, Ren J-h, Tang N, Liu P, Li Q, Huang A-l.** 2020. Antibody responses to SARS-CoV-2 in COVID-19 patients: the perspective application of serological tests in clinical practice. medRxiv doi:10.1101/2020.03.18.20038018**:**2020.2003.2018.20038018.

53. **Zeng F, Dai C, Cai P, Wang J, Xu L, Li J, Hu G, Wang L.** 2020. A comparison study of SARS-CoV-2 IgG antibody between male and female COVID-19 patients: a possible reason underlying different outcome between gender. medRxiv doi:10.1101/2020.03.26.20040709**:**2020.2003.2026.20040709.

54. **Chan KH, Cheng VC, Woo PC, Lau SK, Poon LL, Guan Y, Seto WH, Yuen KY, Peiris JS.** 2005. Serological responses in patients with severe acute respiratory syndrome coronavirus infection and cross-reactivity with human coronaviruses 229E, OC43, and NL63. Clin Diagn Lab Immunol **12:**1317-1321.

55. **Shen C, Wang Z, Zhao F, Yang Y, Li J, Yuan J, Wang F, Li D, Yang M, Xing L, Wei J, Xiao H, Yang Y, Qu J, Qing L, Chen L, Xu Z, Peng L, Li Y, Zheng H, Chen F, Huang K, Jiang Y, Liu D, Zhang Z, Liu Y, Liu L.** 2020. Treatment of 5 Critically Ill Patients With COVID-19 With Convalescent Plasma. JAMA doi:10.1001/jama.2020.4783.

56. **Duan K, Liu B, Li C, Zhang H, Yu T, Qu J, Zhou M, Chen L, Meng S, Hu Y, Peng C, Yuan M, Huang J, Wang Z, Yu J, Gao X, Wang D, Yu X, Li L, Zhang J, Wu X, Li B, Xu Y, Chen W, Peng Y, Hu Y, Lin L, Liu X, Huang S, Zhou Z, Zhang L, Wang Y, Zhang Z, Deng K, Xia Z, Gong Q, Zhang W, Zheng X, Liu Y, Yang H, Zhou D, Yu D, Hou J, Shi Z, Chen S, Chen Z, Zhang X, Yang X.** 2020. Effectiveness of convalescent plasma therapy in severe COVID-19 patients. Proceedings of the National Academy of Sciences doi:10.1073/pnas.2004168117**:**202004168.

57. **Anonymous.** 2020. Rajeev Venkayya, President, Global Vaccine Business Unit on the latest on the Coronavirus and Takeda. https://www.takeda.com/newsroom/featured-topics/rajeev-venkayya-president-global-vaccine-business-unit-on-the-latest-on-the-coronavirus-and-takeda/. Accessed

58. **Healthcare EDftQoMa.** 2017. 19th Edition of the Guide to the Preparation, Use and Quality Assurance of Blood Components.

59. **Cheng Y, Cheng G, Chui CH, Lau FY, Chan PKS, Ng MHL, Sung JJY, Wong RSM.** 2005. ABO Blood Group and Susceptibility to Severe Acute Respiratory Syndrome. JAMA **293:**1447-1451.

60. **Guillon P, Clément M, Sébille V, Rivain J-G, Chou C-F, Ruvoën-Clouet N, Le Pendu J.** 2008. Inhibition of the interaction between the SARS-CoV Spike protein and its cellular receptor by anti-histo-blood group antibodies. Glycobiology **18:**1085-1093.

61. **Neil SJ, McKnight A, Gustafsson K, Weiss RA.** 2005. HIV-1 incorporates ABO histo-blood group antigens that sensitize virions to complement-mediated inactivation. Blood **105:**4693-4699.

62. **Han DP, Lohani M, Cho MW.** 2007. Specific asparagine-linked glycosylation sites are critical for DC-SIGN- and L-SIGN-mediated severe acute respiratory syndrome coronavirus entry. J Virol **81:**12029-12039.

63. **Kumar S, Maurya VK, Prasad AK, Bhatt MLB, Saxena SK.** 2020. Structural, glycosylation and antigenic variation between 2019 novel coronavirus (2019-nCoV) and SARS coronavirus (SARS-CoV). Virusdisease **31:**13-21.

64. **Zhao J, Yang Y, Huang H, Li D, Gu D, Lu X, Zhang Z, Liu L, Liu T, Liu Y, He Y, Sun B, Wei M, Yang G, Wang X, Zhang L, Zhou X, Xing M, Wang PG.** 2020. Relationship between the ABO Blood Group and the COVID-19 Susceptibility. medRxiv doi:10.1101/2020.03.11.20031096**:**2020.2003.2011.20031096.

65. **Tendulkar AA, Jain PA, Velaye S.** 2017. Antibody titers in Group O platelet donors. Asian journal of transfusion science **11:**22-27.

66. **de França NDG, Poli MCC, Ramos PGdA, Borsoi CSdR, Colella R.** 2011. Titers of ABO antibodies in group O blood donors. Revista brasileira de hematologia e hemoterapia **33:**259-262.

67. **Anonymous.** 2019. Is it time to rethink UK restrictions on blood donation? EClinicalMedicine **15:**1-2.

208 68. **Hewitt PE, Ijaz S, Brailsford SR, Brett R, Dicks S, Haywood B, Kennedy IT, Kitchen A, Patel P, Poh J,**
209   **Russell K, Tettmar KI, Tossell J, Ushiro-Lumb I, Tedder RS.** 2014. Hepatitis E virus in blood
210   components: a prevalence and transmission study in southeast England. Lancet **384:**1766-1773.
211 69. **Said B, Usdin M, Warburton F, Ijaz S, Tedder RS, Morgan D.** 2017. Pork products associated with
212   human infection caused by an emerging phylotype of hepatitis E virus in England and Wales.
213   Epidemiol Infect **145:**2417-2423.
214 70. **Tedder RS, Ijaz S, Kitchen A, Ushiro-Lumb I, Tettmar KI, Hewitt P, Andrews N.** 2017. Hepatitis E
215   risks: pigs or blood-that is the question. Transfusion **57:**267-272.
216 71. **UK.** Sep 2017. Guidelines from the expert advisory committee on the Safety of Blood, Tissues and
217   Organs (SaBTO) on measures to protect patients from acquiring hepatitis E virus via transfusion or
218   transplantation.
219   https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/fil
220   e/680297/Hepatitis_E_Guidelines.pdf.
221 72. **Yilla M, Harcourt BH, Hickman CJ, McGrew M, Tamin A, Goldsmith CS, Bellini WJ, Anderson LJ.**
222   2005. SARS-coronavirus replication in human peripheral monocytes/macrophages. Virus Research
223   **107:**93-101.
224 73. **Takada A, Ebihara H, Feldmann H, Geisbert TW, Kawaoka Y.** 2007. Epitopes Required for Antibody-
225   Dependent Enhancement of Ebola Virus Infection. The Journal of Infectious Diseases **196:**S347-
226   S356.
227 74. **Takada A, Feldmann H, Ksiazek TG, Kawaoka Y.** 2003. Antibody-dependent enhancement of Ebola
228   virus infection. J Virol **77:**7539-7544.
229 75. **Geisbert TW, Hensley LE, Geisbert JB, Jahrling PB.** 2002. Evidence against an Important Role for
230   Infectivity-Enhancing Antibodies in Ebola Virus Infections. Virology **293:**15-19.
231 76. **Burnouf T, Seghatchian J.** 2014. Ebola virus convalescent blood products: Where we are now and
232   where we may need to go. Transfus Apher Sci **51:**120-125.

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