**In light of the SARS-CoV-2 pandemic: revisit of the evidence associating Zinc and anti-viral response**

Amir Noeparast[1]*, Gil Verschelden[2]*, Deborah De Geyter[3], Marco Mambretti[1], Cleo Goyvaerts[4], Maya Hites[2]

[1]Laboratory of Medical and Molecular Oncology, Vrije Universiteit Brussel, Laarbeeklaan 103, B-1090 Brussels, Belgium

[2]Department of Infectious Diseases, Erasme Hospital, Université Libre de Bruxelles (ULB), Route de Lennik 808, B-1070 Brussels

[3] Vrije Universiteit Brussel (VUB), Universitair Ziekenhuis Brussel (UZB), Department of Microbiology and Infection Control, Laarbeeklaan 101, 1090, Brussels, Belgium

[4] Laboratory for Molecular and Cellular Therapy (LMCT), Vrije Universiteit Brussel (VUB), Laarbeeklaan 103, B-1090 Brussels

* shared first-authorship

Corresponding authors: maya.hites@erasme.ulb.ac.be, cleo.goyvaerts@vub.be amir.noeparast@vub.be

**Summary**

Since the discovery of the first reported case with Zinc-deficiency in Iran[1] by Prasad et al. in 1961, the knowledge on Zinc has increased significantly. Zinc is the second most abundant common trace mineral in the human body, responsible for vital biological functions from cell growth and development to cell homeostasis and immune response [2,3]. Up to a fifth of the global population is estimated to suffer from different degrees of Zinc deficiency[4]. In the western world, Zinc deficiency is more prevalent among the geriatric population[3], vegans/vegetarians, and people with certain underlying conditions[4]such as liver cirrhosis, inflammatory bowel disease, and various auto-immune disorders[4,5]. The Zinc and Zinc deficiency has been associated with several infectious diseases [2,3].

Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) which is responsible for the ongoing pandemic belongs to the family of coronaviruses.  SARS-CoV-2 has a high genetic similarity to another family member, SARS-CoV, which caused the first major epidemic of the 21st century[6,7].  Currently there is no evidence linking the anti- SARS-CoV-2 response and the element Zinc.

Herein and in light of the SARS-CoV-2 pandemic, we marshal the evidence associating the element Zinc with the anti-viral and antibacterial immune response as well as the cytokine storm and lung injury. Such a revisit of the precedent evidence may inspire the further

investigation assessing the relationship between Zincemia status and the anti-viral response in SARS-CoV-2 patients.

*Zinc deficiency hampers anti-viral and antibacterial immune response*

Zinc deficiency is associated with increased risk of infectious diarrhea and pneumonia among children, as it is associated with an increased risk of developing *Staphylococcus aureus* pneumonia and *Streptococcus pneumoniae* tonsillitis infections among the elderly population[3,8]. Zinc correction in children might be associated with decreased incidence and prevalence of pneumonia-related with these infections [2,9,10]. The most recent meta-analysis investigating the pneumonia-preventive value of Zinc supplementation in children concludes that the Zinc correction in children is associated with 13% and 41% reduced incidence and prevalence of pneumonia, respectively[9]. Another meta-analysis, however, finds that Zinc correction does not significantly impact the mortality due to lower respiratory tract infection in children[11].

Elderly SARS-CoV-2 patients are those with the highest fatality rates, the very subgroup of patients who are at high risk for Zinc deficiency. Of note, pneumonia due to *Staphylococcus aureus* and pneumococcal infections are among the fatal co-infections of SARS-CoV-2[12], especially in the same subgroup of patients.

Toll-like receptors (TLR) are the major mediators of the innate anti-viral immune response[13]. TLR7/9-mediated virus recognition by plasmacytoid dendritic cells leads to a typical anti-viral IFN type I (IFN-β and IFN-α) response that can be applied to SARS-CoV-2 as well[14–16]. Interestingly,  Zinc correction can restore IFN type I response in Zinc-deficient patients[17,18]. In contrast, high Zinc levels can hamper IFN type III (IFN-λ3) anti-viral response[19], which also might influence  anti-SARS-COV-2 response[20]. This paradoxical effect highlights the delicacy of the possible Zinc role in the anti-viral immune response.

We question whether Zinc-deficient SARS-CoV-2 patients are at higher risk of mortality due to a weak or inefficient cellular and/or humoral immunity in response to SARS-CoV-2. Currently, there is no data available on such a correlation concerning SARS-CoV-2.

*Zinc deficiency is associated with cytokine storm and lung injury*

Zinc-deficiency not only hampers our immune system in terms of lack of efficient response when required[8], but it can also be associated with an excessive and damaging response.

Zinc deficiency increases the risk of hyperinflammatory lung damage[21,22] induced by polymicrobial sepsis[23,24], alcohol[25], hyperoxia [26], and ventilator [27]  in mice and rats. Notably, Zinc deficiency is associated with an increased risk of acute respiratory distress syndrome (ARDS) in humans[27]. One of the fatal complications of SARS-CoV-2 in a subgroup of patients is indeed the "*Cytokine Storm*" syndrome and the related acute respiratory distress syndrome (ARDS) [16,28]. The majority of intracellular Zinc is protein-bound, but a tiny fraction is unbound and labile. The free Zinc is involved in cellular signal transduction together with Nitric-Oxide (NO). In pulmonary endothelial cells, NO-induced labile Zinc signaling has an indispensable role in hypoxic vasoconstriction, cytoprotection, and inhibition of cell-apoptosis[29–32]. On the other hand, Zinc deficiency is associated with systemic oxidative stress and decreased Nitric-Oxide (NO) induced vasodilation[32,33].

Zinc concentrations among patients suffering from different auto-immune disorders appear to be lower than those observed in healthy individuals[34]. Zinc deficiency is also associated with the excessive and tissue-damaging pro-inflammatory release of cytokines, including increased TNF-$\alpha$[35,36] and IL-6[37]. Excessive TNF-$\alpha$ and IL-6 are incriminated for lung-damage and the fatal "Cytokine Storm" in a subgroup of SARS-CoV-2 patients. In contrast, Zinc correction is associated with the down-regulation of TNF-$\alpha$ [8,35,36].
However, concerning the effect of Zinc levels on IL-6 levels, there is a discrepancy in the literature. One study reports that Zinc deficiency is associated with increased IL-6 gene expression in mice[38], while another study on a human experimental model reports that Zinc deficiency does not alter IL-6 levels[18]. Nevertheless, among the geriatric female population, a gene polymorphism that leads to an increased immune response-mediated release of Zinc is associated with decreased levels of IL-6 [39]. On the other hand, Zinc supplementation can even lead to the up-regulation of IL-6 levels[40]. Despite the dilemma on the quantitative association of Zinc and IL-6, Zinc correction inversely affects the IL-6-mediated response and signaling[18,37,38,41], which is often[18,37,38,41] but not always[42] considered as a pro-inflammatory response.

Altogether, these observations merit special attention concerning SARS-CoV-2 prevention and patients management[28].

*Zinc has direct anti-viral activity*

Several *in-vitro* studies have shown that Zinc and Zinc ionophores halt the RNA replication of different RNA viruses, including the ones that can infect the human respiratory system[43–50].

Initially, in 1984, Eby et al. show that Zinc gluconate lozenges could be effective in reducing the common colds associated with viral infections of the upper respiratory tract[51]. Since then, further studies resulted in varying and conflicting conclusions on the efficacy of Zinc supplementation in the treatment of common colds[52]. Reanalysis of the reports from 1984 to 1992 by George Eby on the efficacy of zinc lozenges in the treatment of common colds revealed the importance of Zinc

lozenge formulation and type of zinc ion bioavailability in determining its efficacy[53]. Only the Zinc lozenges which were releasing positive Zinc ions at the physiological pH such as gluconate or acetate forms and not the ones with negative or neutral ion bioavailability were effective in shortening the disease period. Zinc lozenges with negative Zinc ion bioavailability even adversely increased the common cold duration[53]. The same author later put forward a hypothesis that the chemical formulation of the Zinc lozenge form determines its efficacy against common colds. Another subsequent Eby's reanalysis of twelve precedent reports suggested that his hypothesis could explain the varying results among the reports and additionally underscored the significance of a minimal dose needed for Zinc efficacy. Eby recommends an initial dose of 200mg for Zinc followed by 100-150mg daily doses for seven days during common colds treatment[54] and names the lower dose of 75mg a "candy" instead of a drug. A further meta-analysis by Eby also leads to similar conclusions[55]. A meta-analysis by Hemila further concluded no superiority of gluconate vs. acetate form and no significantly greater efficacy of doses above 100mg[52]. A recent Finnish randomized controlled trial conducted by Hemila et al. finds no benefit of zinc acetate in treating the common cold[56]. Interestingly, Hemila et al. administered the Zinc lozenge at the dose of 78 mg per day zinc for five days, a dosage, and a duration in total disagreement with Eby's recommendation[56]. As concluded by Hemila et al., further investigation is justified to determine the efficacy and optimal dose and duration of Zinc in treating common colds.

Velthuis et al. have shown that a candidate Zinc ionophore (pyrithione) and Zinc itself could inhibit SARS-CoV replication as single agents and better when combined. Notably, Velthuis et al.[57] showed that Zinc could directly disrupt the initiation of RNA synthesis by the SARS-CoV RNA-dependent RNA polymerase (RdRp) *in-vitro*, possibly via interacting with two Zinc-binding pockets present in the SARS-CoV RdRp[57]. However, likely due to the low cellular uptake of Zinc, the optimal Zinc concentration to efficiently inhibit the SARS-CoV replication was relatively high[57]. Furthermore, a Chinese group who unraveled the SARS-CoV-2 RdRp protein structure[58] observed Zinc ions being chelated at the two candidate pockets within the RdRp of SARS-CoV-2 as well.

Despite the absence of conclusive clinical data on the efficacy of two old drugs[59]; Chloroquine (CQ) and Hydroxychloroquine (HCQ), in SARS-CoV-2 patients, the off-label use of these medications has become widespread.

Zn and (H)CQ might interact at the cellular level. Xue et al. have shown that CQ increases Zinc uptake in ovarian cancer cells and mediates Zinc accumulation into the lysosomes of these cells[60]. In contrast, a study[61] by Seo et al. partially contradicts these findings reporting that CQ decreases the free Zinc levels in lysosomes in adult retinal pigment epithelial cells. However, they still observed some increased intracellular Zinc levels upon CQ treatment compared to the control group. If HCQ is a Zinc ionophore, then increasing lysosomal Zinc uptake in the epithelial cells of the respiratory system may contribute to its anti-SARS-CoV2 mechanism of action.

However, if conversely, HCQ interferes with Zinc lysosomal uptake, it would be interesting to learn whether increasing Zinc plasma concentration can counteract this adverse effect.

Whether (H)CQ are global Zinc ionophores or rather interfere with Zinc uptake into the cells or organelles should remain an open question and the subject of further investigation at this stage.

In figure 1, we summarize three lines of evidence concerning the probable link between Zinc correction and the response to anti-viral treatments that might be applied to SARS-CoV-2 (figure 1).

**The practicality of Zinc correction**

Until today, no consensus and accepted guidelines concerning Zinc deficiency diagnosis and correction has been reached. Radioisotopes can be a sensitive diagnostic tool, but they are costly and exposes patients to radioactive substances. New biomarkers such as Linoleic Acid are emerging[62–65]. However, considering the vast population involved with SARS-CoV-2 and the high economic burden of such approaches, we propose a cheap and straightforward method to evaluate Zn status. The modalities of Zn supplementation are also provided.

*Evaluation of Zinc status: Serum or plasma Zinc*

Plasma and serum Zinc concentration are the most frequently used biomarkers to establish the Zinc status of a patient [66]. Plasma Zinc concentrations respond in a dose-dependent fashion to supplementation in patients with a low or moderate baseline, independent of gender and age[67]. Consequently, Zinc deprivation results in a reduction of the plasma Zinc concentrations. Zinc deficiency is defined as serum Zinc concentrations below 70 and 74 µg/dL for females and males over ten years old, respectively[68].

Nevertheless, several other conditions also affect plasma Zinc concentrations, including hypoalbuminemia, infections, fever, contraceptives, and pregnancy. Concerning SARS-CoV-2 patients, the status of two widely used biomarkers of inflammation[69,70], C-reactive protein, and procalcitonin, are inversely associated with serum Zinc levels[71,72]. The interpretation of the concentration of plasma Zinc must, therefore, take into account all the confounding factors[67].

*Treatment:*

The proposed dose of Zinc (ZnSO4) injection for parenteral nutrition in metabolically stable adult patients is 3 mg/day [73]. The recommended oral daily dose for Zinc correction in adults is 10-30 mg/day[3,74]. The maximum level of the excipient used in FDA-approved oral Zinc tablets (excluding extended-release forms) is 25 mg[75].
Long term Zinc supplementation nevertheless should be very well-weighed[76] as the therapeutic

window of chronic Zinc supplementation is relatively narrow (reviewed here[76]). In contrast, short term Zinc supplementation may be less of a concern[376].

## Conclusion

Some evidence links the anti-viral response and Zinc deficiency.

The possible link between Zinc deficiency, clinical evolution, and response to anti-SARS-CoV2 treatments, particularly in those at risk of developing hyperinflammatory complications, deserves further investigation.

## Author Contributions

A.N and G.V developed the initial idea and co-drafted the first draft under M.H supervision. D.D critically revised the manuscript and contributed to authoring the revised version. M.M revised the manuscript and contributed to authoring the revised version. C.G critically revised the manuscript and contributed to authoring the revised version. M.H supervised the manuscript drafting and critically revised the manuscript and co-authored the final version.

## Acknowledgments

A.N deeply appreciates Dr.Erik Teugels' encouragement to the present work.

## Grant Support

A.N is the beneficiaries of a postdoctoral Fellowship from the Research Foundation-Flanders (FWO).

## Conflict of interest

The authors declare that they have no conflicts of interest.

## References

1.      PRASAD, A. S., HALSTED, J. A. & NADIMI, M. Syndrome of iron deficiency anemia, hepatosplenomegaly, hypogonadism, dwarfism and geophagia. *Am. J. Med.* **31**, 532–546 (1961).

2.      Prasad, A. S. *et al.* Zinc supplementation decreases incidence of infections in the elderly: effect of  zinc on generation of cytokines and oxidative stress. *Am. J. Clin. Nutr.* **85**, 837–844 (2007).

3.      Roohani, N., Hurrell, R., Kelishadi, R. & Schulin, R. Zinc and its importance for human health: An integrative review. *J. Res. Med. Sci.* **18**, 144–157 (2013).

4.      Read, S. A., Obeid, S., Ahlenstiel, C. & Ahlenstiel, G. The Role of Zinc in Antiviral Immunity. *Adv. Nutr.* **10**, 696–710 (2019).

5.      Vincent, M. J. *et al.* Chloroquine is a potent inhibitor of SARS coronavirus infection and spread. *Virol. J.* **2**, 69 (2005).

6.      Cherry, J. D. & Krogstad, P. SARS: the first pandemic of the 21st century. *Pediatr. Res.* **56**, 1–5 (2004).

7.      Andersen, K. G., Rambaut, A., Lipkin, W. I., Holmes, E. C. & Garry, R. F. The proximal origin of SARS-CoV-2. *Nat. Med.* (2020) doi:10.1038/s41591-020-0820-9.

8.      Prasad, A. S. Zinc in human health: effect of zinc on immune cells. *Mol. Med.* **14**, 353–357 (2008).

9.      Lassi, Z. S., Moin, A. & Bhutta, Z. A. Zinc supplementation for the prevention of pneumonia in children aged 2 months to 59  months. *Cochrane database Syst. Rev.* **12**, CD005978 (2016).

10.     Bhutta, Z. A. *et al.* Prevention of diarrhea and pneumonia by zinc supplementation in children in developing countries: pooled analysis of randomized controlled trials. Zinc Investigators' Collaborative Group. *J. Pediatr.* **135**, 689–697 (1999).

11.     Mayo-Wilson, E., Imdad, A., Junior, J., Dean, S. & Bhutta, Z. A. Preventive zinc supplementation for children, and the effect of additional iron: a systematic review and meta-analysis. *BMJ Open* **4**, e004647 (2014).

12.     Chen, Y. & Li, L. SARS-CoV-2: virus dynamics and host response. *Lancet. Infect. Dis.* (2020) doi:10.1016/S1473-3099(20)30235-8.

13.     Lester, S. N. & Li, K. Toll-like receptors in antiviral innate immunity. *J. Mol. Biol.* **426**, 1246–1264 (2014).

14.     Lokugamage, K. G., Hage, A., Schindewolf, C., Rajsbaum, R. & Menachery, V. D. SARS-CoV-2 is sensitive to type I interferon pretreatment. *bioRxiv* 2020.03.07.982264 (2020) doi:10.1101/2020.03.07.982264.

15.     Ali, S. *et al.* Sources of Type I Interferons in Infectious Immunity: Plasmacytoid Dendritic Cells  Not Always in the Driver's Seat. *Front. Immunol.* **10**, 778 (2019).

16.     Zhou, D., Dai, S.-M. & Tong, Q. COVID-19: a recommendation to examine the effect of hydroxychloroquine in preventing infection and progression. *J. Antimicrob. Chemother.* (2020) doi:10.1093/jac/dkaa114.

17.     Cakman, I., Kirchner, H. & Rink, L. Zinc supplementation reconstitutes the production of interferon-alpha by leukocytes from elderly persons. *J. Interferon Cytokine Res.* **17**, 469–472 (1997).

18.    Prasad, A. S. Effects of zinc deficiency on Th1 and Th2 cytokine shifts. *J. Infect. Dis.* **182 Suppl**, S62-8 (2000).

19.    Read, S. A. *et al.* Zinc is a potent and specific inhibitor of IFN-λ3 signalling. *Nat. Commun.* **8**, 15245 (2017).

20.    Mordstein, M. *et al.* Lambda interferon renders epithelial cells of the respiratory and gastrointestinal  tracts resistant to viral infections. *J. Virol.* **84**, 5670–5677 (2010).

21.    Wessels, I. *et al.* Zinc supplementation ameliorates lung injury by reducing neutrophil recruitment and activity. *Thorax* **75**, 253–261 (2020).

22.    Gomez, N. N., Ojeda, M. S. & Gimenez, M. S. Lung lipid composition in zinc-deficient rats. *Lipids* **37**, 291–296 (2002).

23.    Bao, S. *et al.* Zinc modulates the innate immune response in vivo to polymicrobial sepsis through  regulation of NF-kappaB. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **298**, L744-54 (2010).

24.    Knoell, D. L. *et al.* Zinc deficiency increases organ damage and mortality in a murine model of  polymicrobial sepsis. *Crit. Care Med.* **37**, 1380–1388 (2009).

25.    Joshi, P. C., Mehta, A., Jabber, W. S., Fan, X. & Guidot, D. M. Zinc deficiency mediates alcohol-induced alveolar epithelial and macrophage dysfunction in rats. *Am. J. Respir. Cell Mol. Biol.* **41**, 207–216 (2009).

26.    Taylor, C. G. & Bray, T. M. Effect of hyperoxia on oxygen free radical defense enzymes in the lung of zinc-deficient rats. *J. Nutr.* **121**, 460–466 (1991).

27.    Boudreault, F. *et al.* Zinc deficiency primes the lung for ventilator-induced injury. *JCI insight* **2**, (2017).

28.    Mehta, P. *et al.* COVID-19: consider cytokine storm syndromes and immunosuppression. *Lancet (London, England)* (2020) doi:10.1016/S0140-6736(20)30628-0.

29.    Thambiayya, K., Kaynar, A. M., St Croix, C. M. & Pitt, B. R. Functional role of intracellular labile zinc in pulmonary endothelium. *Pulm. Circ.* **2**, 443–451 (2012).

30.    Slepchenko, K. G., Lu, Q. & Li, Y. V. Zinc wave during the treatment of hypoxia is required for initial reactive oxygen  species activation in mitochondria. *Int. J. Physiol. Pathophysiol. Pharmacol.* **8**, 44–51 (2016).

31.    J., B. P. *et al.* Nitric Oxide–Mediated Zinc Release Contributes to Hypoxic Regulation of Pulmonary Vascular Tone. *Circ. Res.* **102**, 1575–1583 (2008).

32.    Choi, S., Liu, X. & Pan, Z. Zinc deficiency and cellular oxidative stress: prognostic implications in cardiovascular diseases. *Acta Pharmacol. Sin.* **39**, 1120–1132 (2018).

33.    Tomat, A. L. *et al.* Moderate Zinc Deficiency Influences Arterial Blood Pressure and Vascular Nitric Oxide Pathway in Growing Rats. *Pediatr. Res.* **58**, 672–676 (2005).

34.    Sanna, A., Firinu, D., Zavattari, P. & Valera, P. Zinc Status and Autoimmunity: A Systematic Review and Meta-Analysis. *Nutrients* **10**, (2018).

35.    von Bulow, V. *et al.* Zinc-dependent suppression of TNF-alpha production is mediated by protein kinase  A-induced inhibition of Raf-1, I kappa B kinase beta, and NF-kappa B. *J. Immunol.* **179**, 4180–4186 (2007).

36.    Suzuki, T., Katsumata, S.-I., Matsuzaki, H. & Suzuki, K. Dietary zinc supplementation increased TNFα and IL1β-induced RANKL expression,  resulting in a decrease in bone mineral density in rats. *J. Clin. Biochem. Nutr.* **58**, 48–55 (2016).

37.    Gruber, K. *et al.* Zinc deficiency adversely influences interleukin-4 and interleukin-6 signaling. *J. Biol. Regul. Homeost. Agents* **27**, 661–671 (2013).

38.    Wong, C. P., Rinaldi, N. A. & Ho, E. Zinc deficiency enhanced inflammatory response by increasing immune cell activation and inducing IL6 promoter demethylation. *Mol. Nutr. Food Res.* **59**, 991–999 (2015).

39.    Cipriano, C. *et al.* Polymorphisms in MT1a gene coding region are associated with longevity in Italian Central female population. *Biogerontology* **7**, 357–365 (2006).

40.    Mariani, E. *et al.* Effect of zinc supplementation on plasma IL-6 and MCP-1 production and NK cell function in healthy elderly: interactive influence of +647 MT1a and -174 IL-6 polymorphic alleles. *Exp. Gerontol.* **43**, 462–471 (2008).

41.    Kitabayashi, C. *et al.* Zinc suppresses Th17 development via inhibition of STAT3 activation. *Int. Immunol.* **22**, 375–386 (2010).

42.    Kimura, A. & Kishimoto, T. IL-6: regulator of Treg/Th17 balance. *Eur. J. Immunol.* **40**, 1830–1835 (2010).

43.    Krenn, B. M. *et al.* Antiviral activity of the zinc ionophores pyrithione and hinokitiol against picornavirus infections. *J. Virol.* **83**, 58–64 (2009).

44.    Lanke, K., Krenn, B. M., Melchers, W. J. G., Seipelt, J. & van Kuppeveld, F. J. M. PDTC inhibits picornavirus polyprotein processing and RNA replication by transporting zinc ions into cells. *J. Gen. Virol.* **88**, 1206–1217 (2007).

45.    Polatnick, J. & Bachrach, H. L. Effect of zinc and other chemical agents on foot-and-mouth-disease virus replication. *Antimicrob. Agents Chemother.* **13**, 731–734 (1978).

46.    Korant, B. D., Kauer, J. C. & Butterworth, B. E. Zinc ions inhibit replication of rhinoviruses. *Nature* **248**, 588–590 (1974).

47.    Si, X. *et al.* Pyrrolidine dithiocarbamate reduces coxsackievirus B3 replication through inhibition of the ubiquitin-proteasome pathway. *J. Virol.* **79**, 8014–8023 (2005).

48.    Gaudernak, E., Seipelt, J., Triendl, A., Grassauer, A. & Kuechler, E. Antiviral effects of pyrrolidine dithiocarbamate on human rhinoviruses. *J. Virol.* **76**, 6004–6015 (2002).

49.    Suara, R. O. & Crowe, J. E. J. Effect of zinc salts on respiratory syncytial virus replication.

*Antimicrob. Agents Chemother.* **48**, 783–790 (2004).

50.    Uchide, N., Ohyama, K., Bessho, T., Yuan, B. & Yamakawa, T. Effect of antioxidants on apoptosis induced by influenza virus infection: inhibition of viral gene replication and transcription with pyrrolidine dithiocarbamate. *Antiviral Res.* **56**, 207–217 (2002).

51.    Eby, G. A., Davis, D. R. & Halcomb, W. W. Reduction in duration of common colds by zinc gluconate lozenges in a double-blind study. *Antimicrob. Agents Chemother.* **25**, 20–24 (1984).

52.    Hemilä, H. Zinc lozenges and the common cold: a meta-analysis comparing zinc acetate and zinc gluconate, and the role of zinc dosage. *JRSM open* **8**, 2054270417694291 (2017).

53.    Eby, G. A. Zinc ion availability--the determinant of efficacy in zinc lozenge treatment of common colds. *J. Antimicrob. Chemother.* **40**, 483–493 (1997).

54.    Eby, G. A. Zinc lozenges: cold cure or candy? Solution chemistry determinations. *Biosci. Rep.* **24**, 23–39 (2004).

55.    Eby, G. A. 3rd. Zinc lozenges as cure for the common cold--a review and hypothesis. *Med. Hypotheses* **74**, 482–492 (2010).

56.    Hemilä, H., Haukka, J., Alho, M., Vahtera, J. & Kivimäki, M. Zinc acetate lozenges for the treatment of the common cold: a randomised controlled trial. *BMJ Open* **10**, e031662 (2020).

57.    te Velthuis, A. J. W. *et al.* Zn(2+) inhibits coronavirus and arterivirus RNA polymerase activity in vitro and zinc ionophores block the replication of these viruses in cell culture. *PLoS Pathog.* **6**, e1001176 (2010).

58.    Gao, Y. *et al.* Structure of the RNA-dependent RNA polymerase from COVID-19 virus. *Science (80-. ).* eabb7498 (2020) doi:10.1126/science.abb7498.

59.    Schrezenmeier, E. & Dorner, T. Mechanisms of action of hydroxychloroquine and chloroquine: implications for rheumatology. *Nat. Rev. Rheumatol.* **16**, 155–166 (2020).

60.    Xue, J. *et al.* Chloroquine is a zinc ionophore. *PLoS One* **9**, e109180 (2014).

61.    Seo, B.-R., Lee, S.-J., Cho, K. S., Yoon, Y. H. & Koh, J.-Y. The zinc ionophore clioquinol reverses autophagy arrest in chloroquine-treated ARPE-19 cells and in APP/mutant presenilin-1-transfected Chinese hamster ovary cells. *Neurobiol. Aging* **36**, 3228–3238 (2015).

62.    King, J. C. *et al.* Biomarkers of Nutrition for Development (BOND)-Zinc Review. *J. Nutr.* **146**, 858S-885S (2015).

63.    Knez, M., Stangoulis, J. C. R., Glibetic, M. & Tako, E. The Linoleic Acid: Dihomo-γ-Linolenic Acid Ratio (LA:DGLA)-An Emerging Biomarker of Zn Status. *Nutrients* **9**, (2017).

64.    Tran, C. D., Gopalsamy, G. L., Mortimer, E. K. & Young, G. P. The potential for zinc stable

isotope techniques and modelling to determine optimal  zinc supplementation. *Nutrients* **7**, 4271–4295 (2015).

65.  Wieringa, F. T., Dijkhuizen, M. A., Fiorentino, M., Laillou, A. & Berger, J. Determination of zinc status in humans: which indicator should we use? *Nutrients* **7**, 3252–3263 (2015).

66.  Sandstr, B. Diagnosis of zinc de fi ciency and excess in individuals. **22**, 133–137 (2001).

67.  Lowe, N. M., Fekete, K. & Decsi, T. Methods of assessment of zinc status in humans: a systematic review. *Am. J. Clin. Nutr.* **89**, 2040S-2051S (2009).

68.  Hotz, C., Peerson, J. M. & Brown, K. H. Suggested lower cutoffs of serum zinc concentrations for assessing zinc status: reanalysis of the second National Health and Nutrition Examination Survey data (1976-1980). *Am. J. Clin. Nutr.* **78**, 756–764 (2003).

69.  Zhang, J. *et al.* Therapeutic and triage strategies for 2019 novel coronavirus disease in fever clinics. *The Lancet. Respiratory medicine* vol. 8 e11–e12 (2020).

70.  Lippi, G. & Plebani, M. Procalcitonin in patients with severe coronavirus disease 2019 (COVID-19): A meta-analysis. *Clinica chimica acta; international journal of clinical chemistry* vol. 505 190–191 (2020).

71.  Bao, B. *et al.* Zinc decreases C-reactive protein, lipid peroxidation, and inflammatory cytokines in  elderly subjects: a potential implication of zinc as an atheroprotective agent. *Am. J. Clin. Nutr.* **91**, 1634–1641 (2010).

72.  Hoeger-Schäfer, J. *et al.* Persistent low serum zinc is associated with recurrent sepsis in critically ill patients - A pilot study. *PLoS One* **12**, (2017).

73.  Review, M. CENTER FOR DRUG EVALUATION AND MULTI-DISCIPLINE REVIEW Summary Review Office Director Clinical Statistical Clinical Pharmacology. *Acalabrutinib NDA* (2016).

74.  Gupta, M., Mahajan, V. K., Mehta, K. S. & Chauhan, P. S. Zinc therapy in dermatology: a review. *Dermatol. Res. Pract.* **2014**, 709152 (2014).

75.  Research, F. for D. E. and *et al.* No Title. https://www.accessdata.fda.gov/scripts/cder/iig/index.cfm?event=browseByLetter.page &Letter=Z (2020).

76.  Maret, W. & Sandstead, H. H. Zinc requirements and the risks and benefits of zinc supplementation. *J. Trace Elem. Med. Biol.* **20**, 3–18 (2006).

**Legends**

*Figure 1* Three lines of evidence associating Zinc and anti-viral response