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A flexible genome-scale resource of SARS-CoV-2 coding sequence clones

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Abstract

The world is facing a major health crisis, the global pandemic of COVID-19 caused by the SARS-CoV-2 coronavirus, for which no approved antiviral agents or vaccines are currently available. Here we describe a collection of codon-optimized coding sequences for SARS-CoV-2 cloned into Gateway-compatible entry vectors, which enable rapid transfer into a variety of expression and tagging vectors. The collection is freely available via Addgene. We hope that widespread availability of this SARS-CoV-2 resource will enable many subsequent molecular studies to better understand the viral life cycle and how to block it.

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Introduction

The world is facing a major health crisis: A global pandemic of the coronavirus disease COVID-19, a severe respiratory illness caused by a novel virus from the family *Coronaviridae* (SARS-CoV-2), has infected millions and caused over 100,000 deaths (1). COVID-19 manifestation in patients can range from asymptomatic (no symptoms) to severe pneumonia and death (2). Early analysis of the outbreak in China outlines symptoms that commonly include fever, dry cough, shortness of breath and myalgia (3). Person-to-person spread through respiratory droplets has been identified as a major source of transmission of the virus (4). To limit contagion, various measures from social distancing to nationwide lockdowns, have been imposed to contain and control the transmission of SARS-CoV-2 (5). Despite these measures, the number of confirmed COVID-19 cases has continued to rise (1), highlighting the need for an effective vaccine and antiviral agents. Furthermore, the extrapolations concerning the evolution of the pandemic are particularly alarming (6). It is therefore of intense and pressing interest to better understand this virus and its interaction with host cells on a molecular level.

Shortly after the outbreak, the complete genome of one SARS-CoV-2 strain was published (7). Using the genome sequence as a reference, Chan *et al*. identified 12 viral open reading frames (ORFs) (8), including ORF1ab, a large polyprotein which is post-translationally processed into 16 proteins. More recently, Wu *et al.* discovered two additional viral ORFs (ORF9Bwu and ORF10wu) with unclear functions (7). Progress on molecular characterization has been made on several viral proteins (9, 10), providing valuable insights into host-virus interaction. However, more research is necessary. The Gateway system offers efficient and high-throughput transfer of the CDSs into a large selection of Gateway-compatible destination vectors used for protein expression in many biological systems, e.g. *Escherichia coli*, *Saccharomyces cerevisiae*, insect, or mammalian cells (11). Broad availability of a collection of SARS-CoV-2 coding sequences (CDSs) has the potential to enable many downstream biochemical and structural studies and thus a better understanding of processes within the viral life cycle, possibly yielding scalable assays for screening drug candidates that could disrupt these processes.

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Methods and Results

Based on the published annotation of the genome sequence of the HKU-SZ-005b (8) and Wuhan-Hu-1 (7) isolates of SARS-CoV-2(8), we requested the synthesis of ORF sequences (GenScript, IDT), including termination codons and *attB* recombination sequences, with optimization of codon usage to reduce GC content and optimize expression in human and insect cells. ORF9Bwu, an alternative ORF within the N gene from the SARS-COV-2 (7), was subsequently amplified by Polymerase Chain Reaction (PCR) from the viral N gene (see Supplementary Table 1). These sequences were then incorporated into Gateway Entry plasmids: either pDONR207 (Invitrogen) or pDONR223 (12).

To enable C-terminal fusion constructs, we also generated an equivalent set of Gateway-compatible clones without termination codons. These clones were made by either PCR-amplifying the whole plasmid with primers that eliminated the stop codon, or by amplifying CDS regions from the first collection (see Supplementary Table 1 for primers), using downstream primers with complementarity regions that were internal to each stop codon, and which simultaneously incorporated the flanking sequences necessary for incorporation into a Gateway Entry plasmid (pDONR207 and 223).

Each SARS-CoV-2 CDS bacterial clone was isolated from a single colony, and its inserted CDS was confirmed by full-length Sanger sequencing. All clones with a pDONR223 backbone were sequenced with M13F and M13R primers (TCAG DNA sequencing facility, Toronto, Canada). Clones with a pDONR207 backbone were sequenced with customized forward and reverse primers. Primer sequences are available in Supplementary Table 1.

A total of 54 clones are currently included in the Gateway-compatible collection (Table 1), covering 28 out of 29 total annotated CDSs in the SARS-CoV-2 genome. (NSP11 was omitted because of its 36 base pair length which makes it incompatible with the Gateway cloning system (13).) All 28 of these CDS regions are available in clones with termination codons, while 26 of 28 are currently available without termination codons.

To promote open-access dissemination of the collection, all clones have been deposited to Addgene (14) [Addgene deposit numbers 77998 and 78047]. Supplementary Table 2

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summarizes all CDSs in the collection, together with their nucleotide and amino acid lengths, annotated functions and direct links to Addgene.

We hope that this SARS-CoV-2 CDS-clone collection will be a valuable resource for many applications, including study of how coronaviruses can exploit host cellular processes for the viral replication cycle, e.g., (15), and understanding virus-host protein-protein interactions (16, 17), production of recombinant virus proteins for structural studies (18), mapping of protein subcellular localization using N-terminal fluorescent reporters (19), or development of vaccines or other therapeutics (20, 21).

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Conflict of Interest

The authors declare that there is no conflict of interest.

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Table 1. The genome-scale SARS-CoV-2 coding sequence clone collection. Production status for clones with (Term) or without termination codon (No-term) is included. Check marks (✔) indicates that clones are currently available on Addgene.

Gene Symbol	CDS Name	Putative Function/Domain	Protein Length	Status Term|No-term
ORF1AB	NSP1	Suppress antiviral host response	180	✔|✔
	NSP2	Unknown	639	✔|✔
	PLPRO (NSP3)	Putative PL-pro domain	1,946	✔|✔
	NSP4	Complex with NSP3 & 6 for DMV (double-membrane vesicle) formation	501	✔|✔
	NSP5	3CL-pro domain	307	✔|✔
	NSP6	Complex with NSP 3 & 4 for DMV formation	291	✔|✔
	NSP7	DNA primase subunits	84	✔|✔
	NSP8		199	✔|✔
	NSP9	RNA/DNA binding activity	114	✔|✔
	NSP10	Complex with NSP14: Replication fidelity	140	✔|✔
	RNA-pol (NSP12)	RNA-dependent RNA polymerase	919	✔|✔
	Heli (NSP13)	Helicase	602	✔|✔
	NSP14	ExoN: 3'-5' exonuclease	528	✔|✔
	NSP15	XendoU: poly(U)-specific endoribonuclease	347	✔|✔
	NSP16	2'-O'-MT: 2'-O-ribo methyltransferase	299	✔|✔
S	S	Spike glycoprotein trimer that binds to host cell receptors (e.g. ACE2)	1,273	✔|*In production*
ORF3A	3A	Induce inflammatory response and apoptosis	275	✔|✔
ORF3B	3B	Induce inflammatory response and inhibit the expression of IFNβ	58	✔|✔

E	E	Envelope protein pentamer	75	✔|✔
M	M	Membrane protein	222	✔|✔
ORF6	6	Antagonize STAT1 function and IFN signalling, and induce DNA synthesis	61	✔|✔
ORF7A	7A	Induce inflammatory response and apoptosis	121	✔|✔
ORF7B	7B	Induce inflammatory response	43	✔|✔
ORF8	8	Induce apoptosis and DNA synthesis	121	✔|✔
N	N	Facilitate viral RNA packaging	419	✔|*In production*
ORF9B	9B	Induce apoptosis	98	✔|✔
ORF9Bwu	9Bwu	Unknown	73	✔|✔
ORF10wu	10wu	Unknown	38	✔|✔

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