

1 **Purinergic Signaling in neutrophils during inflammatory diseases**

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8

9 **Abstract**

10 Purinergic signaling is that nucleotides (especially ATP) and adenosine are utilized as transmitter
11 molecules, which play an important role in the immune system. In the extracellular ventricle, ATP
12 plays a significant role of pro-inflammatory molecules mainly through activating P2 receptors,
13 while adenosine plays the role as anti-inflammatory molecule mainly through activating P1
14 receptors. As we know, neutrophils are the most abundant immune cells in our circulation and have
15 become an essential part of coordinating a series of complex events during inflammatory diseases.
16 However, due to the destruction of inflammatory substances from neutrophils, the activation of
17 neutrophils is fine-tuned, and purinergic signaling is associated with this process. As a matter of fact,
18 altering the balance between P2 and P1 signals is of great importance for neutrophils to exert
19 immune activities properly. Here, we review the role of purinergic signaling in regulatory function
20 of neutrophils during inflammatory disease, and then discuss the potential contribution of targeted
21 purinergic signals in the treatment of the neutrophil during inflammatory diseases.

22 **Keywords: purinergic signaling, regulatory role, neutrophils, inflammation**

23

1. Introduction

Purinergic signaling is a heterocyclic aromatic molecule, which is one of the oldest and most influential biochemical compounds in the history of evolution(1, 2). Purine nucleotide adenosine triphosphate (ATP) is a universal energy currency for intracellular biological reactions on which mammalian life depends(3). Here, we study the role of purine as an extracellular signal molecule, with special attention to ATP, adenosine diphosphate (ADP) and adenosine. In recent decades, further studies have found various biological effects of ATP, ADP and adenosine signals(4-6). These mediators have a role by activating G-protein coupling or ligand-gated ion channel receptors(2). Adenosine is generated by ATP and ADP through the action of cell surface enzymes, which also express receptors for these mediators(7). Because ATP and ADP receptors and adenosine receptors often play the opposite role, the cellular response is not simply related to the ratio of adenosine concentration, but also to the relative expression level and signal intensity of adenosine receptors(1, 8). Purinergic signaling system is a revolutionary selected system that can fine-tune the function of immune cells(1). In fact, purinergic signals mediated by P2R and P1R often play the opposite role in regulating the function of immune cells(9). Specifically, ATP-mediated P2 receptor signal generally promotes the activation of immune cells, while ADO-mediated P1R signal mainly limits the activation of immune cells(2, 10). The transfer of balance from the pro-inflammatory P2R signal to the anti-inflammatory P1R signal, or vice versa, may have a significant effect on the outcome of the immune response(8).

Neutrophils are the most abundant immune cells in human blood and have been made an important part of coordinating a series of complex events in the process of inflammation(11). However, due to the short lifespan of neutrophils, it is difficult to maintain in vitro culture and genetic manipulation, so it has been largely ignored in the field of purinergic signaling in previous studies(12). Recently, a large amount of pharmacological and genetic evidence has changed this paradigm by expanding the role of purinergic signals in neutrophils(13-15). The coordinated interaction between P2 and P1 purinergic signals can effectively activate the immune activity of neutrophils and restore homeostasis in the tissue(8). Here, we review the evidence of neutrophil purinergic signaling and their role in the mechanism of inflammatory diseases, and emphasize their therapeutic potential in the treatment of neutrophil-related diseases, such as inflammatory bowel disease (IBD), pulmonary inflammation and ischemia-reperfusion injury.

2. Classification of Purinergic receptors

The first report of purinergic signaling dates back to 1929, when scientists injected heart tissue extracts intravenously into intact animals and observed a temporary slowdown in heart rate; they later confirmed that the biological agent was an "adenine compound"(16). The ATP (P2 receptor) and ADO (P1 receptor) receptor families were first discovered in 1978(17, 18). When nucleotides and nucleosides bind to specific plasma membrane receptors needed for cell-to-cell communication, they are called purinergic receptors, which are divided into P2 and P1 receptors, respectively(1). According to their sequence and signal characteristics, G-protein-coupled P1 receptors are further divided into four subtypes (A1, A2A, A2B and A3), while P2 receptors form two groups, including metabolic G-protein-coupled P2Y receptors and ionic P2X receptors. Nucleotides (ATP, ADP, UTP and UDP) formed or released outside the cell bind and activate P2 receptor, while adenosine, which exists in the extracellular environment, is the agonist of P1 receptor(2).

2.1 P1Rs

1 P1Rs is a classic G-protein-coupled metabolic receptor, which is a single polypeptide consisted of
 2 seven α -helices perpendicular to the plasma membrane(19). ADO is the endogenous ligand of all
 3 four ADO receptors. A1R and A3R inhibit adenylate cyclase activity through G_i-protein and
 4 stimulate phospholipase C / inositol triphosphate / diacylglycerol pathway through G- β γ G-
 5 protein(20). A2AR and A2BR are mainly coupled to G_s G-protein to enhance AC activity. In
 6 addition, all four ADO receptors are coupled to mitogen-activated protein kinase (MAPK), which
 7 makes them have more complex biological functions(19).

8 **2.2 P2Rs**

9 As mentioned previously, P2 receptors form two groups, including metabolic G-protein-coupled
 10 P2X receptors and P2Y receptors. P2XR is an ATP-gated trimer ion channel. P2XRs has a tertiary
 11 topology similar to intracellular NH₂ and longer COOH ends, extracellular macroscopic responsible
 12 for ligand binding, and two transmembrane regions (TM1 and TM2)(21). In addition, activation of
 13 P2X₇R forms a macropore that allows molecules as long as 900kDa to pass through, which is
 14 involved in the release of pro-inflammatory cytokines(2). P2YRs belong to the delta branch of class
 15 A G-protein-coupled receptor family, which contains seven hydrophobic transmembrane
 16 regions(22). Based on phylogenetic and structural differences, two different P2YR subgroups have
 17 been determined(21). P2Y_{1/2/4/6/11}Rs is mainly coupled with G_q/G₁₁, activating phospholipase
 18 C/inositol triphosphate/diacylglycerol pathway to increase intracellular calcium, while
 19 P2Y_{12/13/14}Rs is mainly coupled with G_{i/o} to inhibit adenylate cyclase (AC), and reduce
 20 intracellular cyclic adenosine monophosphate (cAMP). More evidence suggests that P2Y₁₁R uses
 21 G_s to stimulate AC, to increase intracellular cAMP(22).

22 **3. Release\Signal transduction and Termination of Nucleotide**

23 **3.1 Extracellular nucleotide release**

24 In the physiological state, mammalian cells contain a high concentration of ATP (5-8mM), and
 25 extracellular ATP exists as a very small amount of ~10nM(23). However, pathological conditions
 26 such as inflammation or ischemia can lead to the release of ATP. Cell necrosis is linked to the
 27 outflow of ATP from intracellular storage, and cell necrosis releases a large amount of ATP from the
 28 intracellular storage pool. In most cases, the release of extracellular ATP is finely controlled by
 29 diffusion through plasma membrane channels or exocytosis of ATP-rich vesicles(24). Connexin and
 30 Pannexin half channel are widely reflected in diverse cell types, including neutrophils, endothelial
 31 cells and epithelial cells(24).

32 In extracellular space, ATP and ADP are rapidly metabolized into adenosine monophosphate (AMP),
 33 and AMP is metabolized into adenosine. This nucleotide phosphate hydrolysis includes a two-step
 34 enzyme process regulated by extracellular enzymes(25). In the first step, both ATP and ADP are
 35 transformed into AMP by extracellular nucleoside triphosphate diphosphate hydrolase 1
 36 (CD39)(Figure 1)(1, 26). The subsequent step in the extracellular synthesis of adenosine is the
 37 conversion of extracellular AMP to adenosine by extracellular 5-nucleotidase (CD73)(26). In the
 38 extracellular environment, the levels of ATP and ADP are strictly controlled by plasma membrane
 39 extracellular nucleotidase, such as nucleoside triphosphate diphosphate hydrolase 1 (NTPDase1,
 40 also known as CD39) and extracellular-5-phosphate-nucleotidase (also known as CD73), which
 41 convert ATP/ADP to AMP, and then AMP to ADO, respectively(26). The CD39/CD73 pathway is a
 42 key checkpoint that promotes the transition from ATP-induced pro-inflammatory environment to
 43 ADO-induced anti-inflammatory environment(27, 28). Both CD39 and CD73 are expressed in

1 neutrophils and seem to play a key role in regulating neutrophil activation(29). In addition to CD39
2 and CD73, nucleotide pyrophosphatase and phosphodiesterase (NPPs), alkaline phosphatase (ALP),
3 acid phosphatase (ACP) and extracellular kinases can also degrade extracellular nucleotides and
4 nucleosides(2).

5 **3.2 Extracellular adenosine signal transduction**

6 Adenosine can signal through four distinct G protein-coupled receptors: adenosine A1 receptor
7 (ADORA1), adenosine A2A receptor (ADORA2A), adenosine A2B receptor (ADORA2B) and
8 adenosine A3 receptor (ADORA3)(30-32). Each adenosine receptor subtype is distributed
9 differently on each adenosine target cell. For example, ADORA2B is highly reflected in vascular
10 endothelial cells, while ADORA2A is highly expressed in immune cells, such as neutrophils and
11 lymphocytes(33, 34). The specific human disease state associated with defects or mutations in any
12 of the four defined adenosine receptors is unclear. Adenosine receptor gene knockout mice are alive,
13 indicating that the system is redundant under physiological conditions(35). More than twenty years
14 ago, Burnstock described the existence of the ATP receptor, which he called the "P2" receptor(36).
15 Then, according to the chemical properties, it is split into a P2X receptor (ligand gated ion channel)
16 and P2Y receptor (G protein coupled receptor)(37). Targeted mice with P2 receptor deletion usually
17 survive, indicating that the signal system is redundant under physiological conditions(38). However,
18 P2 receptor gene knockout mice can be free from inflammatory diseases under pathological
19 conditions such as asthma, vascular inflammation or graft-versus-host disease(39). Genomic studies
20 have directly shown that ATP signal is related to human inflammatory or tumor diseases(40). For
21 instance, in chromosomal studies involving humans, mutations in the P2X7 receptor gene are
22 associated with susceptibility to tuberculosis and clinical outcomes of chronic lymphoblastic
23 leukemia(41), and genetic abnormalities of T-cell-dependent P2Y11 signals are associated with
24 paroxysmal narcolepsy(42, 43). On the contrary, under pathological conditions, many specific
25 biological and cell-specific functions of each receptor have been determined. The chronotropic
26 effect of adenosine, which plays a crucial role in the treatment of supraventricular tachycardia,
27 depends on ADORA. Pharmacological studies have shown that ADORA2A signal plays an anti-
28 inflammatory function in human neutrophils, ADORA2A plays a key role in reducing the activation
29 of inflammatory cells in many tissue sites, ADORA2A antagonists have a therapeutic effect on
30 Parkinson's disease(44), and ADORA2B plays a role in tissue adaptation to hypoxia, inflammation
31 or ischemia(45-47).

32 **3.3 Termination of adenosine signal**

33 In the process of signal termination, adenosine is transported from the extracellular space to the
34 intracellular space. This transport involves balancing nucleoside transporters-diffusion-limited
35 channels, allowing adenosine to pass freely through the cell membrane depending on its
36 concentration gradient(48). Mice with balanced nucleoside transporter gene deletion survived, but
37 their adenosine levels increased during disease, which assists in the protection of them during organ
38 ischemia. Dipyridamole can be utilized to induce coronary artery vasodilation by inhibiting
39 balanced nucleoside transporter and increasing the volume of extracellular adenosine. In cells,
40 adenosine is quickly metabolized to inosine by adenosine deaminase (ADA), or to AMP by
41 adenosine kinase(Figure 1)(1, 49).

42 **4. Regulation of purinergic receptors on neutrophil function**

43 **Expression of purinergic receptors in neutrophils**

1 The expression of all purinergic receptors except P2Y12R has been analyzed(50). Convincing data
 2 from mRNA, protein and functional analysis show that P2X1R, P2X7R, P2Y2R, P2Y14R and all
 3 four ADO receptors are taken into account in neutrophils(4). The evidence for the expression of
 4 other purinergic receptors is relatively weak, and further studies are needed to confirm whether
 5 neutrophils express P2X2-6RS, P2Y1R, P2Y4R, P2Y6R, P2Y11R, P2Y12R and P2Y13R(4, 14, 51).

6 **4.1 Purinergic signaling can regulate neutrophil phagocytosis and neutrophil extracellular 7 trap (NET)**

8 Neutrophils are professional phagocytes with a unique phagocytic ability to remove pathogens and
 9 cell fragments(12). The discovery of NET expands the understanding of neutrophil antimicrobial
 10 strategies(52). The net is a large extracellular reticular structure composed of discontended
 11 chromatin and neutrophil antimicrobial factors. The network can not only collect and kill various
 12 microorganisms, but also activate and regulate innate immunity and acquired immunity(53). Both
 13 phagocytosis and Net are essential for neutrophils to effectively eliminate invasive pathogens.
 14 However, when P2X1R antagonist was introduced, the promoting effect of LPS was removed(14).
 15 These results suggest that autocrine activation of P2 signal may be the key to enhance the phagocytic
 16 function of neutrophils due to the release of extracellular ATP, by neutrophils induced by LPS(4).
 17 UDP is the natural ligand of P2Y6R(54). Although UDP itself can not initiate the network formation
 18 of human neutrophils, the P2Y6 signal mediated by UDP is involved in the formation of Net induced
 19 by sodium urate crystals(55). Neutrophil A3R enhances bacterial clearance and activates human
 20 neutrophils with A3R agonists to promote the formation of filamentous processes of neutrophils,
 21 which are recognized as cell lines(55, 56).

22 **4.2 Purinergic signaling can regulate neutrophil chemotaxis**

23 Chemotaxis is the capacity of cells to sense gradient, polarization and directional migration in the
 24 chemotactic gradient field(4). Recent studies have shown that autocrine purinergic signals play a
 25 key role in guiding neutrophil chemotaxis(4, 57). In the process of chemotaxis, neutrophils need to
 26 be polarized, with an anterior pseudopod at the anterior edge of sense the gradient of chemical
 27 attraction and a posterior pseudopod at the posterior edge to maintain directional migration(57).
 28 Once neutrophils sense the chemical inducer Panx1 half channel quickly shifts to the anterior edge
 29 and releases mitochondrial-derived ATP from the pseudopodia(58, 59). The extracellular release of
 30 ATP acts as an autocrine messenger, amplifying chemotactic signals by activating P2Y2R-mediated
 31 frontal mTOR signals(60). The mechanism of neutrophil gradient sensing is constituted by
 32 extracellular ATP and positive feedback of P2Y2R receptor(61). Then, the released ATP is
 33 hydrolyzed into ADO, in the CD73 associated with the neutrophil membrane in situ, and then the
 34 frontal neutrophil A3R is activated to drive the second round of signal amplification. The subsequent
 35 step amplification is also essential for promoting the initial expansion of neutrophil chemotaxis
 36 because it controls the rate of migration. At the trailing edge, diffuse or locally produced ADO
 37 activates A2AR and triggers cAMP/PKA signals to suppress P2Y2R-mediated mTOR signals(61).
 38 Activation of A2AR maintains cellular polarization and promotes the contraction of the tail pods.

39 **4.3 Purinergic signaling can regulate the rolling, adhesion and migration of neutrophils**

40 Neutrophils are usually the primary immune cells recruited to the inflammatory site(12). In most
 41 tissues, the neutrophil recruitment cascade includes the following recognized steps: lineage, rolling,
 42 adhesion, crawling, and subsequent outcome(62). A recent in vivo study showed that there was not
 43 any difference in lipopolysaccharide-induced neutrophil rolling and adhesion between WT and
 44 *P2rx1*^{-/-} mice. However, neutrophil migration in *P2rx1*^{-/-} mice was inhibited, suggesting that P2

1 signaling may be associated with neutrophil recruitment cascades by promoting neutrophil
2 migration rather than rolling and adhesion(63). The adoptive transfer of neutrophils from WT and
3 *P2rx1*^{-/-} mice to WT mice showed that the loss of P2X1R in neutrophils rather than the loss of
4 P2X1R in vascular endothelial cells or other immune cells caused neutrophils to migrate from
5 venules. In contrast to P2X1R, A2AR expressed by neutrophils appears to inhibit the recruitment
6 cascade of neutrophils. Using A2AR agonist to activate A2AR signal, tumor necrosis factor-2
7 integrin-mediated neutrophil rolling and adhesion were significantly inhibited in β - α -attacked
8 mouse epidermal muscle retroactively venule and in vitro flow chamber model(63).

9 **4.4 Purinergic signaling can regulate neutrophil degranulation**

10 Neutrophil granules, including primary granule (PGs), secondary granule (SGS), tertiary granule
11 (TGS) and SVS, are formed successively during granulogenesis(64). Neutrophil granules contain a
12 large number of antimicrobial peptides and proteolytic enzymes(64, 65). These proteins enable
13 neutrophils to exert bactericidal and immune functions, but if they are not released properly, they
14 are potentially harmful to the host. Purinergic signals play a bi-directional role in regulating
15 neutrophil degranulation. An in vitro study showed that ATP γ S, a unhydrolyzable adenosine
16 triphosphate analogue, could further enhance fMLP-induced neutrophil degranulation, but
17 hydrolyzable adenosine triphosphate could not further promote fMLP-induced neutrophil
18 degranulation. On the contrary, ATP can be hydrolyzed to inhibit fMLP-induced neutrophil
19 degranulation(66). Considering the strong hydrolytic activity of extracellular nucleotidase on
20 neutrophil membrane to convert ATP into ADO, further studies have demonstrated that this
21 inhibitory effect on neutrophil degranulation is induced by the hydrolysate of ATP and ADO(66). In
22 addition, with the use of selective agonists and antagonists, a recent study has demonstrated that
23 LPS-induced ATP autocrine promotes neutrophil exocytosis to SVS, TGS and SGS by activating
24 P2X1R. The described two-way effect of purinergic signals on neutrophil degranulation may be
25 necessary for neutrophils to properly release their granule contents to regulate their antibacterial
26 activity during infection and to avoid damage to healthy tissue(66).

27 **4.5 Purinergic signaling can regulate the oxidative burst of neutrophils**

28 Superoxide is only an effective antimicrobial agent, which can kill microbial pathogens and regulate
29 a variety of signal pathways(67). Owing to the destructiveness of superoxide, the oxidation burst is
30 fine-tuned, and the purine energy signal is closely connected with this process. P2Y2R is the
31 activator of neutrophil oxidative burst. The down-regulation of P2Y2R expression in differentiated
32 neutrophil-like HL-60 cells (DHL-60) can dramatically inhibit the oxidative burst induced by fMLP.
33 Sodium urate crystals can induce oxidative burst of neutrophils, but the antagonist of P2Y6R can
34 inhibit the oxidative burst of neutrophils induced by sodium urate crystals(54). It is useful to noting
35 that the absence of autocrine activation of P2Y2R and P2Y6R in exogenous ATP, neutrophils in
36 these two studies may amplify oxidative bursts. Compared with P2Y2R and P2Y6R, A2BR and
37 A3R inhibited the oxidative burst of neutrophils(68).

38 **4.6 Purinergic signaling can regulate neutrophil apoptosis**

39 The circulating neutrophils is under a short lifespan of only 20 hours and will not proliferate.
40 However, the lifespan of neutrophils is markedly prolonged under inflammation and other
41 pathological conditions(69). Extracellular ATP is a major regulator of neutrophil apoptosis. Even
42 exposure to ATP for 10 minutes is sufficient to cause a persistent delay in human neutrophil
43 apoptosis. Utilizing various selective purine receptor antagonists, researchers have verified that
44 ATP-mediated delay in neutrophil apoptosis is P2Y11R-dependent. P2Y11R mediates the anti-

1 apoptosis effect of ATP by elevating cAMP in neutrophils and activating subsequent cAMP-
2 dependent protein kinases(70).

3 **5. Neutrophil-related Infectious and Inflammatory Diseases**

4 **5.1 Purinergic signaling of neutrophils in Inflammatory Bowel Disease**

5 Inflammatory bowel disease is linked to excessive intestinal inflammation, and purine signal is
6 related to inflammatory bowel disease. Intestinal inflammation is related to the profound changes
7 of metabolic supply and demand, which leads to the deep hypoxia of inflammatory mucosa(71).
8 Effective immune response depends on the effective activation of neutrophils(72). Neutrophils
9 release extracellular ATP under the action of inflammatory mediators. Neutrophils may reduce key
10 immune responses, such as cell adhesion and chemotaxis, partly due to changes in the expression
11 of purinergic signal-related genes mRNA(4). Studies have shown that hypoxia signals are
12 transcribed to induce CD39 and CD73(73). During intestinal inflammation, thus transferring the
13 balance from ATP signals to adenosine signals. In addition, pharmacological studies in mice have
14 shown that HIF activator can reduce intestinal inflammation(73). The ATP signal of neutrophils is
15 associated with long-term duodenal motility disturbance and intestinal nerve injury in IBD, which
16 is consistent with the pro-inflammatory effect of P2 receptor signal in this disease(74). P2X7,
17 pAnnexin-1 channel, ASC adaptor protein and caspases are all involved in ATP-induced signaling
18 pathways that drive intestinal nerve death during enteritis. In addition, a subset of CD4+T
19 lymphocytes of regulatory T cells needs extracellular adenosine production dependent on CD39 and
20 CD73 to inhibit experimental enteritis in order to comply with its inhibitory function in mouse
21 models, and neutrophils strengthen the inhibitory function in this process. Generally speaking, in
22 mouse models, CD39 or CD73(75, 76). Adenosine receptor signaling appears to be anti-
23 inflammatory and barrier protective effects in experimentally induced colonic inflammation through
24 adenosine signaling events involving Adora2a or Adora2b receptors(77). In addition, sulfadiazine
25 and methotrexate-two commonly used drugs-are utilized to treat the anti-inflammatory effects of
26 IBD involving the release of extracellular adenosine. Taken together, these findings highlight the
27 therapeutic potential of anti-inflammatory drug strategies that balance the pro-inflammatory
28 activation of P2 receptors to adenosine receptors, especially Adora2a and Adora2b, in the treatment
29 of IBD(78, 79). For instance, this conversion can be done through the use of HIF to achieve
30 adenosine receptor agonists, soluble forms of apyrase (converting ATP and ADP to AMP) or
31 extracellular nucleotidase (converting AMP to adenosine) or adenosine receptor agonists.

32 **5.2 Purinergic signaling of neutrophils in Acute Lung injury**

33 Acute lung injury is part of the main causes of morbidity and mortality associated with acute
34 diseases(80). Massive neutrophil infiltration is a sign of acute lung injury and exposure to lung
35 injury increases the level of ATP in the lung, indicating that activation of P2 receptors such as P2Y6
36 or P2X7 leads to increased inflammation and vascular leakage(81, 82). Some genetic and
37 pharmacological studies have confirmed that inappropriate activation of P2R signal is related to
38 neutrophil-induced inflammation and tissue damage. Knockout or antagonism of mouse P2X7R can
39 inhibit neutrophil recruitment into the lung and protect ALI(83). LPS provides dual signals for
40 alveolar macrophages, induces the production of cytokines through TLR4/MYD88 signals, induces
41 cytokine necrosis through P2X7R/CD14 signals, promotes the release of pre-IL-1 α , then activates
42 IL-1 receptors on endothelial cells, induces tight junctions to open, and makes neutrophils infiltrate
43 into the lungs(84, 85). In addition, P2X7R-induced soluble VCAM-1 exfoliated from type I alveolar

1 epithelial cells and used as a chemoattractant to recruit neutrophils during ALI(86). Intratracheal
2 LPS-induced pulmonary neutrophil recruitment was also inhibited in mice treated with P2Y1R or
3 P2Y14R antagonists. In the ALI model induced by severe pulmonary infection, P2X7R is associated
4 with the aggravation of inflammatory injury and neutrophil infiltration. When mice are infected with
5 the highly virulent *Mycobacterium bovis*, it can cause severe tuberculosis and lung damage. The
6 lung injury of chimeric mice with P2X7R deletion in bone marrow-derived cells was substantially
7 alleviated, indicating that P2X7R in bone marrow-derived cells played an important role in the
8 occurrence and development of severe tuberculosis(87). Increased immune response is one of the
9 principal causes of lung injury during influenza virus infection. P2X7R gene knockout led to better
10 results for influenza virus infection, characterized by reduced lung pathology and neutrophil
11 infiltration. In addition, deletion of P2X7R or inhibition of P2X7R activation by selective
12 antagonists or apyrase can inhibit pulmonary inflammation and neutrophil infiltration in the early
13 stage of acute adenovirus infection(88). Cecal ligation and puncture (CLP) is another method to
14 induce SIRS-related ALI. CLP, in *P2ry2-/-* and *Adora3-/-* mice the researchers observed a decrease
15 in neutrophil recruitment in the lungs and a decrease in ALI progress compared with WT mice.
16 P2Y12R antagonists can successfully reverse the pathological changes of WT mice. Similar results
17 have been observed in *P2y12-/-* mice, and P2Y12R antagonists can successfully reverse the
18 pathological changes of WT mice. These studies upheld the role of P2Y2R, A3R and P2Y12R in
19 promoting neutrophil infiltration during ALI(89, 90).

20 **5.3 Purinergic signaling of neutrophils in Ischemia-reperfusion injury**

21 Ischemia-reperfusion injury occurs after initial ischemia or hypoxia, with the return of blood and
22 accompanying oxygen to tissue, which is a common complication of myocardial infarction,
23 transplantation, stroke and trauma(91, 92). Neutrophil infiltration is a sign of IR injury and is an
24 important part of protracted inflammatory response and its severity. It has been confirmed that
25 purinergic signal has a profound effect on neutrophils during IR injury (93). In a variety of
26 experimental animals, the application of A2AR agonists has achieved beneficial effects in inhibiting
27 neutrophil infiltration and concomitant neutrophil infiltration in kidney, lung and myocardial IR
28 injury(94). Further through-hole analysis in vitro showed that A3R agonists could inhibit the
29 migration of neutrophils, suggesting that the protective effect of A3R may be explained by the direct
30 inhibition of neutrophil activation(95). In addition, A3R activation decreased neutrophil infiltration
31 in IR-injured myocardium in WT mice, but not in global *A3R* deficient or chimeric mice lacking
32 A3R in BM-derived cells. Subsequent experimental results are consistent with previous studies,
33 indicating that A3R expressed by neutrophils is essential for the inhibition of neutrophil
34 infiltration(95).

35 Distinct from the inhibitory effect of A1R, A2AR and A3R on neutrophil activation during IR injury,
36 A2BR plays a bi-directional role in myocardial and lung IR injury(96). However, according to data
37 obtained from BM chimeric mice, the pro-inflammatory effect of A2BR may be specific to resident
38 lung cells, but not to BM-derived neutrophils(96). A recent report showed that the antagonistic effect
39 of P2X7R could improve renal neutrophil infiltration and tissue damage induced by IR, while
40 P2X7R gene knockout achieved comparable results. Chimeric mice with bone marrow-derived cell
41 P2X7R deficiency further confirmed that the activation of P2X7R in bone marrow-derived cells is
42 required for renal neutrophil infiltration during IR injury(97, 98). The effects of other P2 receptors
43 on neutrophils during IR injury have yet to be studied.

44 **6. Conclusion**

1 Purinergic signal transduction is a major regulatory mechanism in various inflammatory diseases.
 2 Significant progress has been made in identifying various types of neutrophil purinergic receptors
 3 and understanding their function in coordinating the appropriate immune response to invasive
 4 pathogens or diseased tissues. The fine-tuning balance between P2R and P1R signals seems to be
 5 indispensable for the formation of neutrophil plasticity and heterogeneity to coordinate a series of
 6 complex events during inflammation. Several drugs that target purinergic signals, such as adenosine
 7 and cuspidate, have been used in patients. In the near future, the research progress in the field of
 8 neutrophil purinergic signal transduction may be further used in the treatment of inflammatory or
 9 infectious diseases, and the continuous development of this field will open up several new ways for
 10 the treatment of patients with inflammatory diseases.

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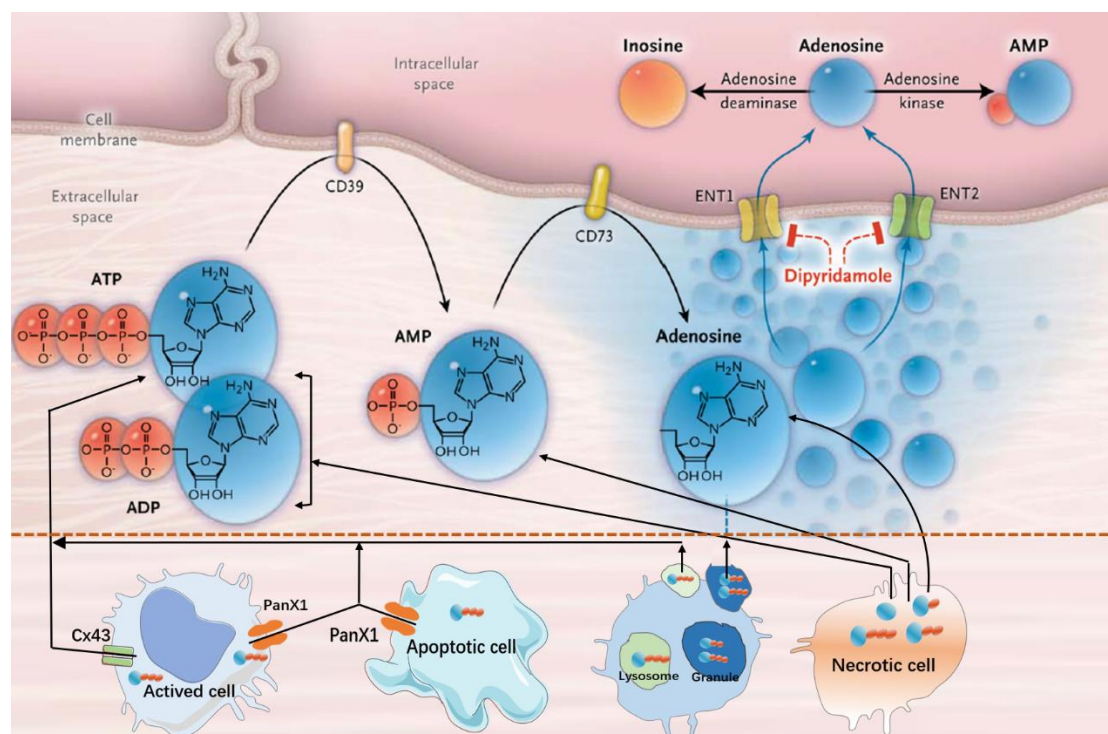
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8 **Figure 1 Purinergic signal and release of extracellular ATP and ADO.**

9 Under inflammatory conditions, purinergic signal is mainly derived from precursor nucleotides ATP
10 and ADP are converted to AMP, through extracellular nucleoside diphosphate hydrolase 1 (CD39)
11 enzyme activity, and then AMP is converted to adenosine through extracellular-5'- nucleotidase
12 (CD73). In the process of apoptosis, Panx1 is cleaved by apoptotic executive enzymes (caspase3
13 and 7) to produce a truncated activation subunit, which regulates the ATP release of apoptotic cells.
14 In some pathological conditions, the release of extracellular ATP is finely regulated by the diffusion
15 of semi-tubules of connexin 43 (Cx43) or connexin 1 (Panx1) and the exocytosis of ATP-rich
16 vesicles such as granules and lysosomes. Under extreme conditions, cell necrosis releases a large
17 amount of ATP from the intracellular storage pool.