

Integrative Roles of Gangliosides in the Nervous System: Novel Molecular Mechanisms Elucidated by Genetic Engineering

Koichi Furukawa^{1,3*}, Yuhsuke Ohmi², Yesmin Farhana^{1,3}, Orié Tajima¹, Yuji Kondo³, Pu Zhang^{1,3}, Noboru Hashimoto⁴, Yuki Ohkawa⁵, Robiul H. Bhuiyan¹, Keiko Furukawa¹

¹Department of Biomedical Sciences, and ²Department of Medical Technology, Chubu University College of Life and Health Sciences, 1200 Matsumoto, Kasugai, Aichi, 487-8501, ³Department of Biochemistry II, Nagoya University Graduate School of Medicine, 65 Tsurumai, Showa-ku, Nagoya, 466-0065, and ³Department of Tissue Regeneration, Tokushima University Graduate School of Biomedical Sciences, 3-18-5, Kuramoto-cho, Tokushima, 770-8504, Japan

*Correspondence should be addressed to: Koichi Furukawa: Department of Biomedical Sciences, Chubu University College of Life and Health Sciences, 1200 Matsumoto, Kasugai, Aichi 487-8501, Japan; Tel: +81-568-51-9512; Fax: +81-568-51-9512; E-mail: koichi@isc.chubu.ac.jp

Abstract: Acidic glycosphingolipids, gangliosides are highly and consistently expressed in nervous tissues of vertebrates at high levels. Therefore, they have been believed to be largely involved in the development and function of nervous systems. Recent studies with genetic engineering of glycosyltransferase genes have revealed novel aspects of roles of gangliosides in the regulation of nervous tissues with substantial basis. In this review, novel findings on the ganglioside functions and their modes of action elucidated mainly by studies of gene knockout mice have been summarized. In particular, roles of gangliosides in the regulation of lipid rafts to maintain the integrity of nervous systems have been reported with a focus on the roles in the regulation of neuro-inflammation and neurodegeneration via complement

systems. In addition, recent advances in the studies of congenital neurodisorders due to the genetic mutations of ganglioside synthase genes, and also in the techniques for the analysis of ganglioside functions have been introduced.

Keywords: ganglioside; knockout; neurodegeneration; glycosphingolipid; inflammation; microdomain

1. Introduction

Nervous tissues are differentiated from ectoderm, and outline of their morphology is determined until birth. During the growth stage after birth, further functional differentiation proceeds, and fundamental shapes and functions are established within several years in the case of humankind. However, formation of functional networks based on various experiences and learning activities keep proceeding continuously, leading to the maintenance of high-grade nerve functions due to the plasticity of nervous systems [1]. After reaching middle age, network function gradually decline along with aging, and physiological and pathological degeneration gradually proceed in nervous tissues [2]. Under definite pathological conditions, marked tissue degeneration is induced, leading to functionally irreversible states such as occurrence of dementia. During individual processes of natural history of nervous systems, structures of carbohydrates on proteins and lipids seem to dramatically alter [3,4] and be involved in the formation of appropriate environments for the high-grade structures and functions based on the molecular modification required for each step.

Since acidic glycosphingolipids, gangliosides are highly and consistently expressed in nervous tissues of vertebrates at high levels [3], huge contribution of them in the neurological function have been long expected [5,6]. Actually, the fact that ganglioside expression patterns dramatically change during development has suggested that they play critical roles in the evolution and differentiation of nervous systems [7,8]. On the other hand, for carbohydrates on proteins, it appears that sufficient understanding has

not been obtained in terms of integrative analysis of nervous system-specific carbohydrate functions, although there have been a number of reports on the carbohydrate functions on individual proteins.

In this review, outline of functions of gangliosides recently obtained has been summarized with focus on the findings from studies of knockout mouse of various glycosyltransferase genes.

2. Roles of gangliosides

Gangliosides are sialic acid-containing glycosphingolipids, and widely expressed in almost all tissues and cells of vertebrates. They are enriched in brain tissues, suggesting that they are involved in the evolution and regulation of nervous systems [9]. The most intriguing feature of gangliosides is that they consist of hydrophilic carbohydrates and hydrophobic lipid portions [10], being expressed on the cell membrane and present in the out layer of lipid two layers as shown in Figure 1.

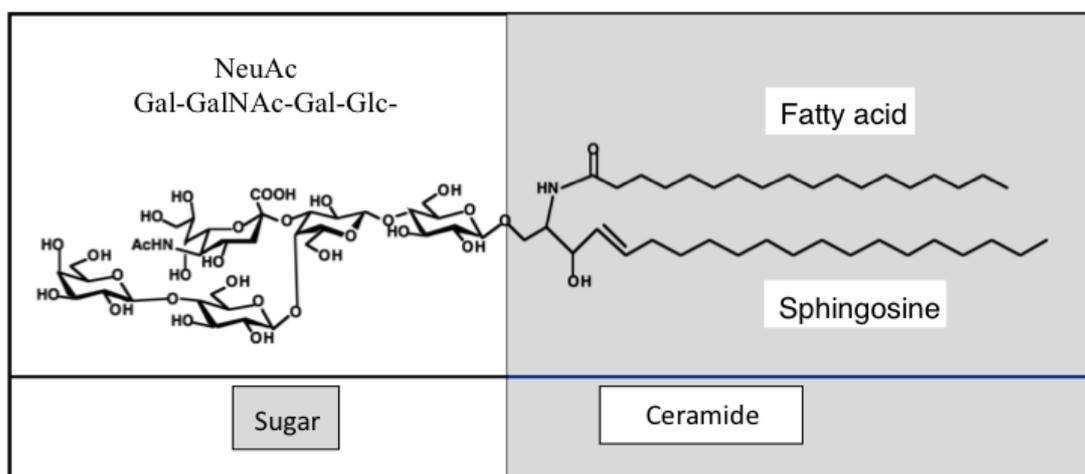


Fig. 1. Glycosphingolipids are amphipathic molecules, and expressed in the out layer of lipid two layers of cell membrane. Hydrophobic portion (ceramide) is embedded in the outer layer of the membrane, and sugar portion is protruding outside of the membrane.

Therefore, it has been a mystery how gangliosides are involved in the regulation of signals for cell differentiation, activation and malignant transformation [11]. In this review, ganglioside expression in the inflammation and neurodegeneration was summarized, and their roles in the maintenance of integrity and generation of

phenotypes of cells with focus on the inflammation and degeneration due to the altered gangliosides were summarized.

2-1. Gangliosides in development and growth

The fact that ganglioside composition in brain tissues varies along with development and growth of organism has been well known in many reports [9,12]. As well understood, simple structure gangliosides such as GM3 and GD3 mainly exist at the initial stage of developments, i.e. embryonal day 12-14 (E12-14) of mice. At the differentiation stage after E16, when extension of neuritis and synapse formation occur, mature type gangliosides such as GM1, GD1a, GD1b and GT1b increase and become main components of brain tissues [13]. Main pathway of ganglioside synthesis is shown in Figure 2.

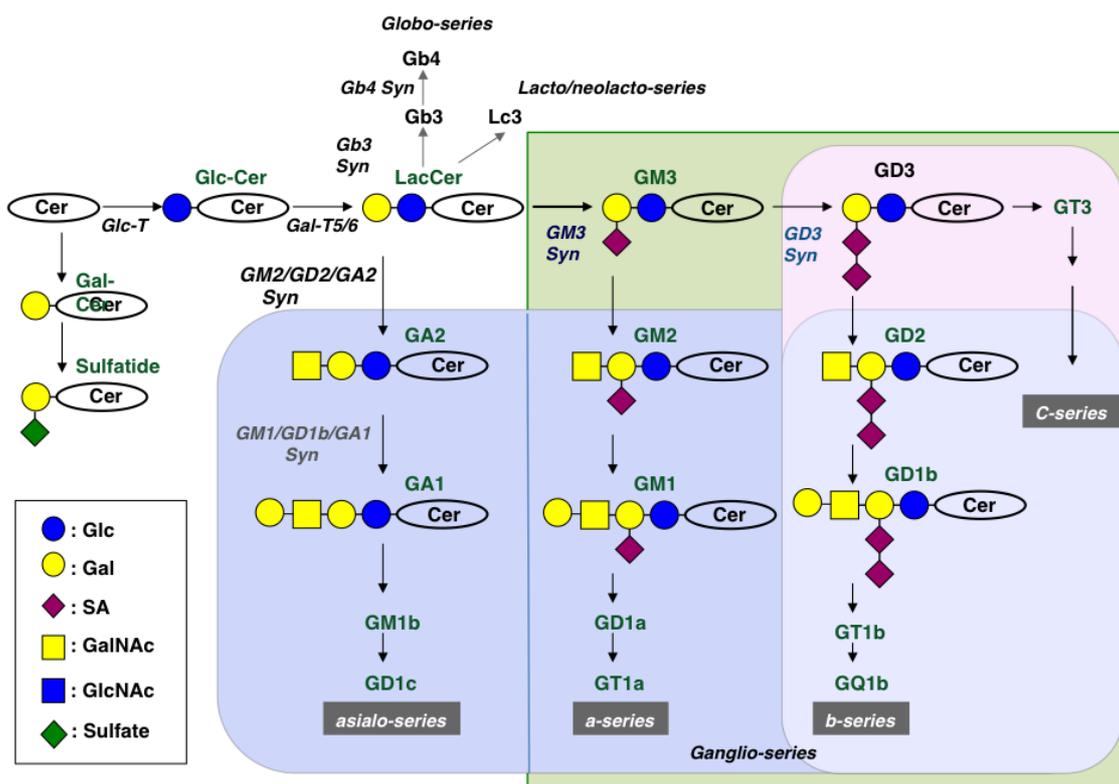


Fig. 2. Main pathway of ganglioside synthesis. Glycosyltransferases catalyzing individual steps were shown by italics, and deleted structures in KO mice were encircled by different colored squares.

On the other hand, ganglioside composition in brain tissues showed almost no change at adult stage, while total amounts gradually reduce. In particular sites in brain,

however, gradual changes emerge along with aging [9]. Eventually, no comprehensive and fine studies on ganglioside expression in nerve tissues have actually been performed to date aside from exceptions [14,15].

2-2. Function of monosialylgangliosides and disialylgangliosides

In order to analyze regulatory function of nervous systems by gangliosides, a rat pheochromocytoma cell line, PC12 has been frequently used [16,17]. PC12 cells over-expressing GM1 showed reduced sensitivity to nerve growth factor (NGF), leading to lowered neurite extension reaction, and exhibited suppressed activation of TrkA/Ras/ERK1/2 signals upon NGF stimulation [18]. On the other hand, GD3-overexpressing PC12 cells showed increased phosphorylation levels of TrkA and ERK1/2 even without NGF stimulation [19]. Similarly, contrastive effects of gene expression between GM1 synthase and GD3 synthase on phenotypes of various cancer cell lines have been observed [20,21]. From these results, it was demonstrated that monosialylgangliosides and disialylgangliosides play distinct roles in the regulation of malignant properties in cancer cells [22].

Although mechanisms for signal regulation in cancer cells and PC12 cells based on the gangliosides with different numbers of sialic acids are not clear yet, it might be a reflection of regulatory functions of gangliosides in organogenesis and differentiation of nervous tissues and cells. Dynamic changes in ganglioside expression during evolution and development of nervous tissues, and their implication have been summarized in the next chapter.

3. Aging, neurodegeneration and gangliosides

In various tissues and cells, only nervous tissues have been thought to show consistent compositions among species of mammals and birds. In fact, highly similar TLC patterns of gangliosides extracted from brain tissues were reported [23]. However, various changes in ganglioside composition under physiological and pathological conditions have been investigated [2,24,25].

3.1 Changes of gangliosides in central nervous systems with aging

There have been some reports on the changes of ganglioside expression during evolution, development and aging of mice and rats [26-28]. For human brain gangliosides, changes of ganglioside expression have been reported. Generally, contents of gangliosides gradually decrease [29], and a-series gangliosides tend to decrease mainly in frontal cortex [30]. On the other hand, b-series gangliosides decrease in cerebellum with aging. Whole amounts of lipids contained in brain tissues continuously reduce until reaching 90 years old. In particular, expression levels of gangliosides and cerebroside largely lowered at that time point. Investigation of 118 individuals with 20 ~ 100 years-old revealed that concentration of gangliosides were maintained between 20 ~ 70 years-old [31], and then, the expression patterns continuously altered with aging, leading to reduced ratios of GM1 and GD1a. Similar findings with these results have been reported based on newest analytical techniques with mass spectrometry [32]. However, there are no detailed researches on the functional relationship of altered ganglioside expression with pathological changes in brain tissues.

3.2 Gangliosides in Alzheimer disease

In a variety of neurodegenerative diseases, particularly in Alzheimer diseases, one of the most important factors involved in neuron death is local inflammation [33]. As reported previously, all components involved in the classic pathway of complement activation were detected in the nervous tissues, and this pathway have been activated in Alzheimer diseases, resulting in the detection as fibrous β amyloid [33] or neurofibrillary tangle [34]. From these findings, complement activation and subsequent inflammation seem to be main mechanisms for the induction of brain damages in Alzheimer diseases.

In these past 20 ~ 30 years, it has been demonstrated that the complement system was involved in the neurodegeneration such as Alzheimer disease [35]. C1q and other components deposit in amyloid plaques and neurofibrillary tangle [36,37] leading to

complement activation. At the lesioned areas, levels of mRNAs for complement components markedly increased [38]. Furthermore, C1q inhibitors alleviated clinical features of Alzheimer diseases, suggesting that complement activation play important roles in the neuroinflammation and subsequent neurodegeneration [39]. Interestingly, the complement activation observed in double knockout (DKO) mice (deleting GM2/GD2 synthase and GD3 synthase) containing only GM3 plays similar roles with those in Alzheimer disease. This point will be described later.

On the other hand, complement systems might work both detrimental and beneficial directions in the disease control [40]. For example, roles during developmental processes [41] or neuronal generation in adults [42] have been reported. In particular, its physiological roles in the elimination of unnecessary cellular components or in the improvement of inflammatory reaction are intriguing.

As a role of gangliosides in Alzheimer disease, GM1 has been reported to become a triggering factor to cause aggregation of A β peptide on the cell membrane [43], and mechanisms for the continuous accumulation of A β has been also proposed [44]. Murine models for Alzheimer disease generated with genetic back grounds of knockout of GD3 synthase gene or GM2/GD2 synthase gene have been generated, resulting in the milder phenotypes in the former [45], and more serious phenotypes in the latter [46]. Recently, it was reported that expression of *B4GALNT1* promoted the processing of A β [47], and complex gangliosides exacerbated clinical features of Alzheimer disease. On the other hand, different changes in ganglioside composition were found in various transgenic model mice expressing human amyloid precursor proteins. [48]. From these results, it has been indicated that altered metabolism of gangliosides is involved in the pathogenesis of Alzheimer disease.

3.3 Parkinson diseases and gangliosides

It has been reported that amounts of GM1 and GD1a reduced in the brain of patients with Parkinson disease [49]. Furthermore, it was also indicated that *B4galnt1* (GM2 synthase) KO mice lacking GM1 ganglioside [50] showed Parkinson disease-like

neurological disorders even in heterozygotes [51]. Indeed, efficiency of GM1 administration to Parkinson disease model animals was reported [52], and similar trials have been also performed in clinical cases, resulting in the improvement of clinical features [53]. On the other hand, GD3 synthase (*ST8SIA1*) KO mice lacking b-series gangliosides and having increased levels of GM1 and GD1a [54] were reported to be less susceptible to Parkinson disease [55], suggesting that complementation of GM1 and/or GD1a is also efficient for patients with other neurodegenerative diseases [49].

3.4 Inflammatory reaction and gangliosides

Changes in ganglioside expression in various inflammatory reactions have been observed to date [56]. For example, changes in gangliosides expressed on glial cells [57] or multiple sclerosis, or changes by growth factors [58] have been reported. Further more, it was reported that ganglioside GD3 was induced in inflammatory environments of brain tissues in mice and rats [59]. As for immune cells, GD3/GD2 were induced in T lymphocytes when stimulated via T cell receptor or IL-2 receptor, or various mitogenic factors [60,61]. On the other hand, ganglioside GD2 was expressed on functional T cells [62]. These results corresponded with facts that GD2 was induced on HTLV-1-infected T cells via a viral product p40tax [63]. Thus, changes in ganglioside expression patterns on immune cells are frequently observed in various inflammatory reactions [64,65].

4. Functions of glycolipids elucidated by ganglioside-deficient (knockout) mice

4.1. Abnormal phenotypes exhibited by knockout mice and inflammatory reaction

In order to investigate roles of glycosphingolipids in our bodies, cDNAs of various glycosyltransferase genes have been isolated, i.e. GM2/GD2 synthase [66], GD3 synthase [67-69], GM1/GD1b/GA1 synthase [70], GM3 synthase [71-73], Gb3 synthase [74-76] and many other glycosyltransferase cDNA, and then, knockout (KO) mouse lines of these enzyme genes have been established. For example, KO mouse lines of GM2/GD2 synthase [50], GD3 synthase [52], Gb3 synthase [77],

lactosylceramide (LacCer) synthase [78,79], and double KO mice of GM2/GD2 synthase and GD3 synthase [80,81] have been established and analyzed. DKO mice of *B6galt5* and *B4galt6* were also generated [82]. Synthetic pathway of main glycosphingolipids and glycolipid structures deleted in the individual KO mice are presented in Figure 2. Generally, abnormal phenotypes observed in these KO mouse lines were fairly milder than expected. This could be because residual glycolipids could compensate functions that deleted structures primarily exerted [50]. All these results were summarized in Table 1. However, it seems very intriguing that inflammatory reaction was found mainly in the central nervous systems of many of these KO mice of glycosyltransferase genes [83]. Biochemical and morphological changes observed in the DKO mice of GM2/GD2 synthase and GD3 synthase genes were also detected more or less in many of single gene KO mouse lines [84] (Table 1).

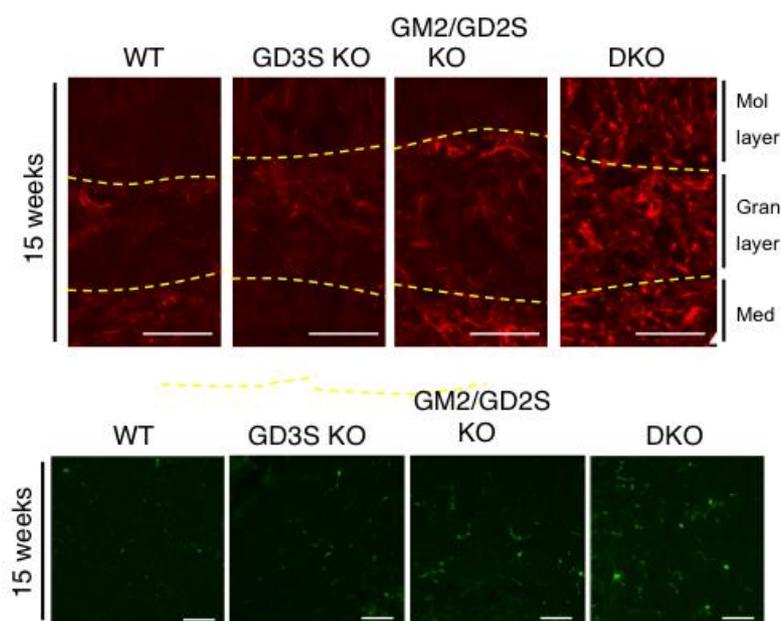


Fig. 3. Immunohistochemistry of GFAP-positive astrocytes and F4/80 antibody-reactive microglia.

Marked gliosis were found in ganglioside synthase gene KO mice. GFAP-positive cells were increased (upper), and F4/80-positive cells (lower) were assembled in cerebella, indicating astrocytes and microglia, respectively.

We reported involvement of complement systems in the neuro-inflammation as a novel aspect of ganglioside deficiency [83]. We compared degree of the complement

activation, inflammatory reaction, and destruction of lipid rafts among various KO mice of glycosyltransferase genes and wild type (WT) mice, and demonstrated extensively increased expression levels of complement-related genes. More over, we reported proliferation of astrocytes and assembly of microglia corresponding to the degree of the defects in ganglioside composition in the individual KO mice (Figure 3). It was also shown that various cytokine genes were up-regulated with aging correspondingly with the progression of neuro-inflammation as described above. The molecular mechanisms for these inflammation based on the ganglioside deficiency have been analyzed with focus on the changes in lipid rafts [84]. Details were described in the next chapter.

4.2 Neuro-inflammation corresponding to the degree of ganglioside deficiency

Gangliosides have been considered to be involved in the development, differentiation, and function of nervous systems [85]. However, gangliosides have been shown to play roles mainly in the maintenance and repair of nervous tissues based on the abnormal phenotypes detected in the genetically engineered mutant mice [86]. Generally, neurodegeneration was found in common in KO mouse lines of ganglioside synthase genes [50,87,88]. In particular, age-dependent progressive neurodegeneration was observed in KO mice of GM2/GD2 synthase, while subtle abnormal neurological signs could be detected when they were born [50]. On the other hand, DKO mice of GM2/GD2 synthase and GD3 synthase gene demonstrated apparent neurodegeneration in the early stage of life [80], or even sudden death by auditory stimulation [89]. Although KO mice lacking GlcCer synthase [90] showed embryonal lethality [91], conditional KO mice, in which GlcCer synthase was deleted after birth, also showed neurodegeneration [39]. These results indicated that ganglioside deficiency cause abnormality in the maintenance of integrity in the nervous system, leading to neurodegeneration. However, it had been not clear how ganglioside deficiency cause neurodegeneration.

Among various features indicating inflammation in the nervous tissues, abnormal

proliferation of astrocytes and assembly of microglia were markedly and characteristically found in the cerebella of ganglioside deficient mice [83]. Furthermore, those inflammatory reactions were confirmed by immunohistochemistry such as GFAP-positive astrocytes and F4/80 antibody-reactive microglia (Figure 3). GFAP+ cells apparently increased at 15 weeks after birth in DKO mice, and further increased with aging. At 50 weeks after birth, GFAP+ cells increased even in the single gene KO mice such as GD3 synthase KO or GM2/GD2 synthase KO. Microglia cells also showed increased assembly at 15 weeks after birth in DKO mice, and further increased with aging. This microglia assembly could be also found at 50 weeks after birth in the single gene KO mice [83].

As for inflammatory cytokines, increased expression levels of IL-1 β and TNF α genes were detected in RT-PCR of mRNA from cerebella of DKO mice. Expression levels of these genes tended to increase with aging in DKO mice, while no apparent changes in those gene expression could be found in WT and single gene KO mice.

4.3 Involvement of complement system in the inflammatory reaction

From results of gene expression analysis in DKO mice, it was demonstrated that complement-related genes were generally up-regulated in the cerebella of DKO mice. Therefore, it was suggested that wide range consumption of complement components was induced due to activation of complement system [83]. Actually, deposit of C1q, a complement component, could be found in the cerebella of DKO, and it was also the case in KO mice of GM2/GD2synthase. [83,92]. In order to investigate whether complement activation detected in the cerebella of DKO mice exacerbates with aging, expression levels of C1qa, C3, and C4 were examined along with aging, resulting in the increase of expression of C1q gene between 15 weeks to 50 weeks after birth, and in apparent differences from WT mice with aging. In GM2/GD2 synthase KO mice, expression of complement-related genes moderately increased. Similar inflammatory reactions were also observed in spinal cords of these KO mice [93].

To clarify the roles of complement activation in the neuro-inflammation and neurodegeneration, triple KO mice lacking complement C3 gene as well as GM2/GD2 synthase and GD3 synthase genes were established as shown in Figure 4 [83]. In this TKO mice, it was really shown that complement activation is actually involved in the complement deposition and secretion of inflammatory cytokines, and also in the neurodegeneration as demonstrated by the alleviation of brain degeneration indicated by reduction in brain weights. In the neurological disorders such as Guillan-Barre syndrome and Miller syndrome caused by anti-ganglioside antibodies, it was reported that complement systems are tightly involved [94,95]. Therefore, inhibitors for complement-related components have been tried in the therapeutic application for the control of these diseases, showing nice effects on mouse disease models [96] and

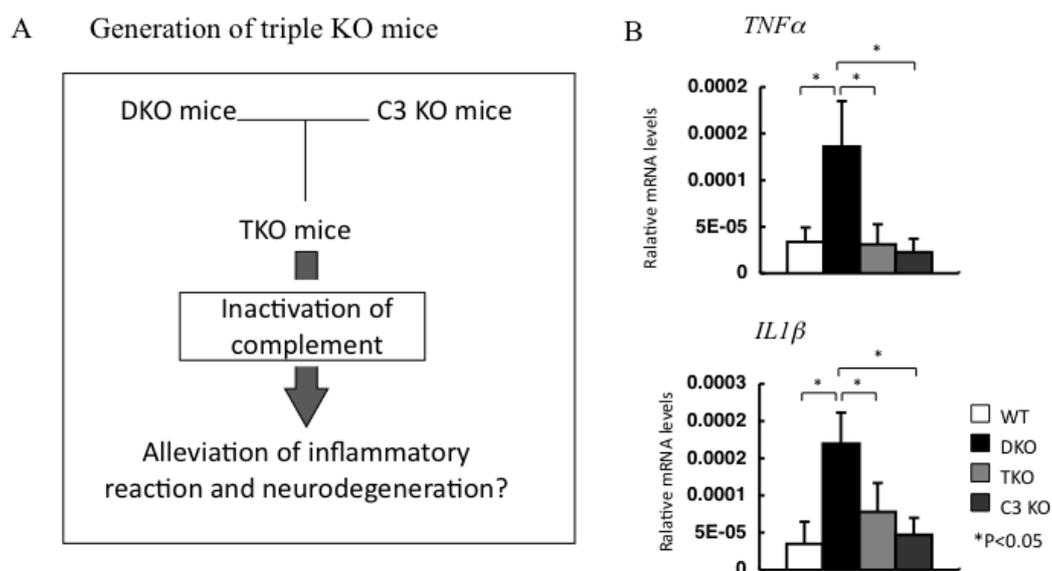


Fig. 4. Inflammatory reactions in the DKO mice were suppressed by genetic disruption of complement system. A, Triple KO (TKO) mice was generated by mating the DKO mice and C3 KO mice to clarify the roles of complement systems in the brain disorders in DKO mice. B, Expression levels of inflammatory cytokines were reduced in TKO mice.

human cases [97].

5. Microdomains on cell membrane and gangliosides

Generally speaking, membrane microdomains such as lipid rafts,

glycolipid-enriched microdomain (GEM)/rafts, or detergent-insoluble microdomains (DIM) are considered to be a platform of cell signaling, and roles of glycosphingolipids in lipid rafts have been increasingly recognized [98]. In particular, molecular composition of gangliosides consisting of polymorphic sugar chains and heterogenous lipid moiety has made us to expect that gangliosides might be one of main regulators of biological properties of the microdomains.

5.1 Gangliosides regulate cell signaling in microdomains

Various extrinsic stimulations are transduced via receptors and their adjacent molecules on the cell membrane, and these molecules often form molecular complexes in microdomains such as lipid rafts or GEM/rafts [99,100](Figure 5). With a number of experiments using cell lines and KO mice, it has been shown that changes in ganglioside expression largely affect lipid rafts, and control the cell signals and finally cellular phenotypes [98]. Therefore, integrity of lipid rafts has been investigated by analyzing changes in intracellular localization of membrane molecules depending on the conditions of glycosphingolipids and cells [101,102]. Immunocytostaining of these membrane molecules has been an efficient approach for the localization of same membrane microdomains. Further more, it seems extremely important to substantially reveal physical interaction among these molecules on the living cell membrane, and would become a prerequisite to clarify the roles of gangliosides and their associating molecules on the cell membrane [103].

5.2 Microdomain on the cell membrane of nervous systems and gangliosides

Glycosphingolipids are generally considered to be concentrated and localized in GEM/rafts [104], and intense localization is found in highly-differentiated neurons. However, they are dispersed from lipid rafts under particular environments [105 Harder and Simons 1997]. On the other hand, changes in relevant molecules such as caveolin-1 affect intracellular localization of glycosphingolipids [106]. Furthermore, fine distribution analysis of gangliosides using immunoelectron microscopy revealed that different ganglioside species (e.g. GM1 and GM3) showed distinct distribution

patterns on the cell membrane, suggesting the presence of heterogeneous microdomains, and individual gangliosides form specific microdomains [107].

Surprisingly, GPI-anchored proteins and GEM/raft markers dispersed from GEM/rafts in cerebella of ganglioside-deficient mice, and marked cell damages were induced [83,84,108]. Analysis of altered floating patterns of GPI-anchored proteins and GEM/rafts markers in various ganglioside-deficient mouse lines by immunoblotting revealed that flotillin-1 and caveolin-1 were apparently dispersed from GEM/rafts (Figure 5). DAF and NCAM showed marked dispersion from GEM/rafts in DKO mice, and DAF shifted to non-GEM/rafts fraction in GM2/GD2 synthase KO mice. Total protein amounts showed no differences among these lines. Generally, gangliosides are essential in the architecture of GEM/rafts, and it is suggested that more striking abnormalities of GEM/rafts were exhibited in DKO mice than in single gene KO mice.

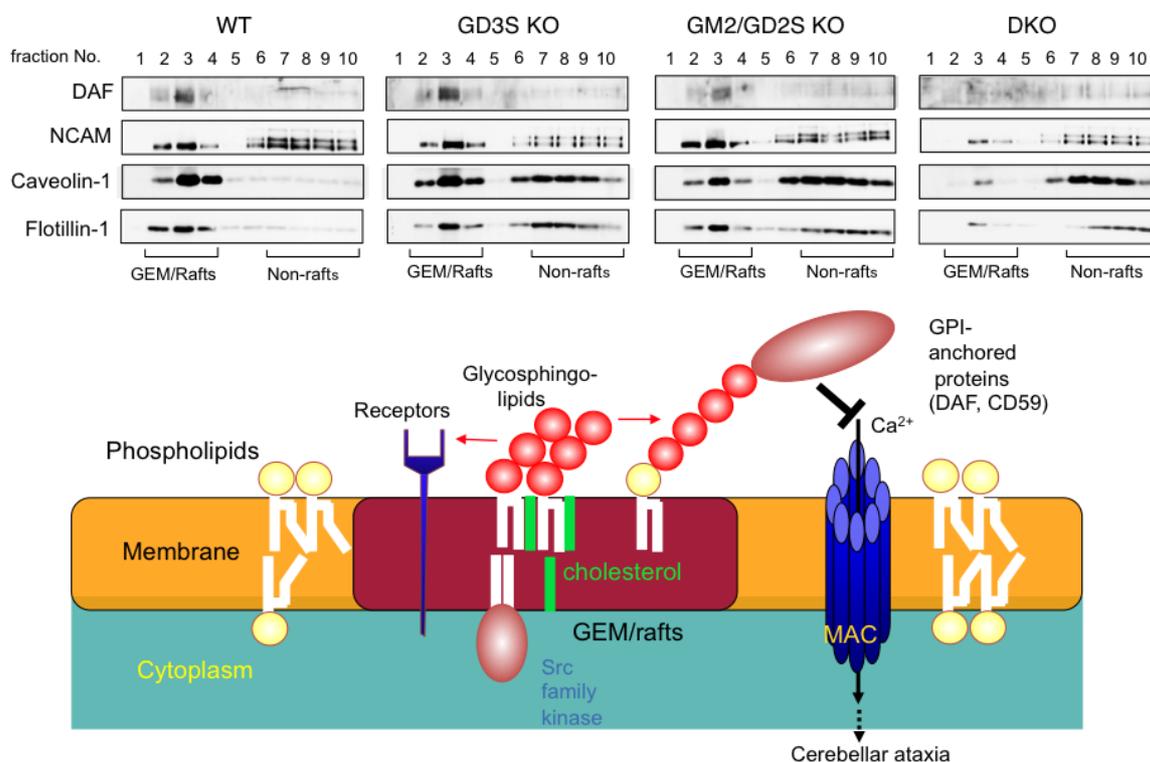


Fig. 5. Destruction of GEM/raft induced changes in the architecture and functions of raft-localizing molecules. Localization of GPI-anchored proteins including complement-regulatory molecules changed, leading to functional abnormalities and subsequent complement activation and inflammation in ganglioside-deficient mice brain.

5.3 Complement activation and destruction of lipid rafts

Results of analysis of DKO mice revealed that there were many examples of molecular dispersion of GPI-anchored proteins and GEM/raft markers even in single gene KO mice, suggesting that disordered GEM/rafts in brain tissues induce dysfunction of GPI-anchored proteins. In GM2/GD2 synthase gene KO and DKO, localization of GalCer, phospholipids, and cholesterol tended to decrease [84], suggesting that abnormal composition of gangliosides might induce abnormal distribution patterns of GPI-anchored proteins and raft marker proteins, and also of other lipids.

One of the most important factors involved in the complement activation and resulting neurological disorders should be complement-regulatory proteins. In fact, it has been well known that expression levels of CD59 largely lower at the disease site of Alzheimer disease [109]. DAF is also a crucial molecule for the maintenance of tissue integrity [110]. Many of these complement-regulatory factors belong to GPI-anchored proteins, and are concentrated in GEM/rafts fraction [111]. Therefore, it is suggested that destruction of GEM/raft induced changes in the localization of GPI-anchored proteins and their functional abnormalities, leading to complement activation and inflammation. These processes were summarized in Figure 5.

6. Human diseases caused by congenital deficiency of gangliosides

Following analysis of ganglioside functions using KO mice of glycosyltransferase genes, human cases of congenital defects of ganglioside synthase genes were reported in this century.

6.1 GM3 synthase deficiency causes severe clinical features

Although there have been a number of studies on deficiency of ganglioside catalytic enzyme genes, no reports on congenital deficiency of ganglioside synthase genes have been found until 2004. Simpson et al. reported “infantile epilepsy” families in Amish due to deficiency of GM3 synthase (*ST3GAL5*) as the first case of genetic

mutation in ganglioside synthase genes [112].

As described above, the majority of gangliosides are synthesized through GM3, and diverse carbohydrate structures are generated from a common precursor, lactosylceramide, along with several major synthetic pathways [113]. Therefore, defects of GM3 synthase gene in the Amish families actually resulted in the complete loss of ganglio-series and the patients exhibited serious infantile epileptic disorders [112] and skin abnormalities [114,115], suggesting that gangliosides are essential in the regulation of nervous tissues and other organs. Thus, patients lacking GM3 synthase activity exhibited severe neurological disorders such as infantile epilepsy, mental retardation, visual disorders, and also skin pigmentation, while no definite abnormal phenotypes were found in KO mice of GM3 synthase gene (*St3gal5*) in any sites of the body, except for the auditory system [116].

6-2 GM2/GD2 synthase gene deficiency causes hereditary spastic paraplegia

B4GALNT1 is an essential enzyme for the synthesis of complex gangliosides, lack of which resulted in progressive neurodegeneration with aging in mice [87]. Recently, 11 cases of hereditary spastic paraplegia (HSP) due to mutation in the coding region of *B4GALNT1* were reported [117-119]. We examined the enzyme activities using a cell free enzyme assay with cell extracts, and by flow cytometry of transfectant cells with mutant cDNA expression plasmids [120]. Among them, almost all mutant genes showed complete loss of B4GALNT1 activity, while two mutants showed low activity, indicating that the clinical findings of these patients derived from loss of B4GALNT1 enzyme activity, and the mutations are responsible for the clinical features of HSP. As expected from KO mice phenotypes of *B4galnt1* gene, the intensity of their neurological disorders was milder than expected. These clinical features of the patients including male hypogonadism are very similar with abnormal phenotypes detected in *B4galnt1*-deficient mice [121]. In contrast to GM3 synthase mutation, *B4GALNT1* mutations brought about much milder clinical features with slower progression.

KO gene	Glc-Cer Syn	GM3 Syn	GD3 Syn	GM2/GD2 Syn	DKO ¹⁾
Lost structures	all glycosphingo-lipids	ganglio-series (a-, b-, c-)	b-series (and c-series)	all complex gangliosides (inc. asialo-series)	all complex gangliosides (inc. asialo-series)
Remaining structures		asialo-series	a-series and asialo-series	GM3, GD3 (and GT3)	GM3
	Gal-Cer and sulfatides	Gal-Cer and sulfatides neutral glycolipids			
Phenotypes	↓ Embryonal lethal	↓ No apparent abnormalities	↓ Mild abnormalities	↓ Gradual abnormalities	↓ Neurodegeneration from early phase
Remarks		Auditory disorder	Poor repairment Low serum leptin	Male infertility Low serum testosterone	Refractory skin lesion Auditory shock
				Wallerian degen ²⁾	

Table 1. Deficient structures, remaining glycolipids and phenotypes of individual KO mice of glycosyltransferase genes. Residual glycolipids could compensate functions that deleted structures primarily exerted. ¹⁾ double KO of GD3 synthase and GM2/GD2 synthase genes. ²⁾ Wallerian degeneration.

7. Future scope of ganglioside research

Recent advances in methodologies and technologies have enabled us to further investigate modes of action for gangliosides. Fine heterogeneity in either sugar moiety or ceramide portion has been demonstrated, leading to further understanding of mechanisms by which gangliosides play their roles or interact with their recognizing molecules. For example, further analysis of derivatives of sialic acids such as deaminoneuraminic acid (KDN) [122], *O*-acetylated GD3/GD2 [123] as sugar modifications, and long fatty chain-containing glycolipids in lactosylceramide [124], saturated/unsaturated fatty acid-containing Gb4 [125], or hydroxylated ceramide-containing gangliosides [126], remain to be promoted. It is also an important issue to be solved which cell lineages more critically need gangliosides between neuron and glia [127], although few studies have been reported to date. Ultra-high-resolution imaging of gangliosides in GEM/rafts has enabled us to understand actual formation of microdomains [128] or new concept of gradual formation of GEM/rafts with different size and composition [129]. Identification of novel ligand molecules for gangliosides should be also a promoting factor for further understanding of molecular functions of gangliosides [103, 130], although not so many studies in neurology field have yet been reported to date.

8. Conclusion

It seems hard now to clearly answer the fundamental question, what are the roles of gangliosides in nerve functions. Many of “functions” presented here are results drawn from observation of abnormal situations brought about by artificially enhanced expression of particular glycosyltransferases or suppression of their functions in turn to speculate normal functions primarily exerted. Therefore, they represent a part of “functions”, but not all. The main factor to make us feel unsatisfied is the technical restriction in the manipulation of key glycosyltransferase genes, observing some phenotypes as functions of many structures belonging to one group all together. Nevertheless, clarification of roles of individual gangliosides by collecting experimental

data with various limitations should be essential for the solution of mystery in the polymorphism of carbohydrates, leading to the new pavement toward our ultimate purpose.

References

1. Ramakers, GJ. Neuronal network formation in human cerebral cortex. *Prog. Brain Res.* **2005**, 147, 1-14.
2. Ando, S. Neuronal dysfunction with aging and its amelioration. *Proc. Jpn. Acad. Ser. B Phys. Biol. Sci.* **2012**, 88, 266-282.
3. Fishman, PH.; Brady RO. Biosynthesis and function of gangliosides. *Science.* **1976**, 194(4268), 906-915.
4. Schachter, H. The role of the GlcNAc(beta)1,2Man(alpha)- moiety in mammalian development. Null mutations of the genes encoding UDP-N-acetylglucosamine: alpha-3-D-mannoside beta-1,2-N-acetylglucosaminyltransferase I and UDP-N-acetylglucosamine:alpha-D-mannoside beta-1,2-N-acetylglucosaminyltransferase I. 2 cause embryonic lethality and congenital muscular dystrophy in mice and men, respectively. *Biochim. Biophys. Acta.* **2002**, 1573, 292-300.
5. Nagai, Y. Functional roles of gangliosides in bio-signaling. *Behav. Brain Res.* **1995**, 66, 99-104.
6. Rösner, H. Developmental expression and possible roles of gangliosides in brain development. *Prog. Mol. Subcell. Biol.* **2003**, 32, 49-73.
7. Gorio, A. Ganglioside enhancement of neuronal differentiation, plasticity, and repair. *CRC Crit. Rev. Clin. Neurobiol.* **1986**, 2, 241-96.
8. Maccioni, HJ.; Quiroga, R.; Ferrari, ML. Cellular and molecular biology of glycosphingolipid glycosylation. *J. Neurochem.* **2011**, 117, 589-602.
9. Skaper, S.D.; Leon, A.; Toffano, G. Ganglioside function in the development and repair of the nervous system. From basic science to clinical application. *Mol. Neurobiol.* **1989**, 3, 173-199.
10. Wiegandt, H. Gangliosides. In *Glycolipids*, Wiegandt, H.(Ed.). Elsevier New York, pp. **1985**, 199–260.
11. Hakomori, S.I. Structure and function of glycosphingolipids and sphingolipids: recollections and future trends. *Biochim. Biophys. Acta.* **2008**, 1780, 325-346.

12. Schengrund, C.L. Gangliosides: glycosphingolipids essential for normal neural development and function. *Trends Biochem. Sci.* **2015**, *40*, 397-406.
13. Yu, R.K.; Macala, L.J.; Taki, T. et al. Weinfield HM.; Yu FS. Developmental changes in ganglioside composition and synthesis in embryonic rat brain. *J. Neurochem.* **1988**, *50*, 1825-1829.
14. Suzaki, K. The pattern of mammalian brain gangliosides. 3. Regional and developmental differences. *J. Neurochem.* **1965**, *12*, 969-979.
15. Vrbaski, S.R. Cerebellum gangliosides in postnatal rats. *J. Neurochem.* **1980**, *34*, 1032-1033.
16. Greene, L.A.; Tischler, A.S. Establishment of a noradrenergic clonal line of rat adrenal pheochromocytoma cells which respond to nerve growth factor. *Proc. Natl. Acad. Sci. USA.* **1976**, *73*, 2424-2428.
17. Mutoh, T.; Tokuda, A.; Miyadai, T.; Hamaguchi, M.; Fujiki, N. Ganglioside GM1 binds to the Trk protein and regulates receptor function. *Proc. Natl. Acad. Sci. USA.* **1995**, *92*, 5087-15091.
18. Nishio, M.; Fukumoto, S.; Furukawa, K.; et al. Overexpressed GM1 suppresses nerve growth factor (NGF) signals by modulating the intracellular localization of NGF receptors and membrane fluidity in PC12 cells. *J. Biol. Chem.* **2004**, *279*, 33368-33378.
19. Fukumoto, S.; Mutoh, T.; Hasegawa, T.; et al. GD3 synthase gene expression in PC12 cells results in the continuous activation of TrkA and ERK1/2 and enhanced proliferation. *J. Biol. Chem.* **2000**, *275*, 5832-5838.
20. Hamamura, K.; Furukawa, K.; Hayashi, T.; et al. Ganglioside GD3 promotes cell growth and invasion through p130Cas and paxillin in malignant melanoma cells. *Proc. Natl. Acad. Sci. USA.* **2005**, *102*, 11041-11046.
21. Dong, Y.; Ikeda, K.; Hamamura, K.; et al. GM1/GD1b/GA1 synthase expression results in the reduced cancer phenotypes with modulation of composition and raft-localization of gangliosides in a melanoma cell line. *Cancer Sci.* **2010**, *101*, 2039-2047.

22. Furukawa, K.; Ohkawa, Y.; Yamauchi, Y.; et al. Fine tuning of cell signals by glycosylation. *J. Biochem.* **2012**, 151, 573-578.
23. Yu, R.K. Development regulation of ganglioside metabolism. *Prog. Brain Res.* **1994**, 101, 31-44.
24. Sonnino, S.; Chiricozzi, E.; Grassi, S.; Mauri, L.; Prioni, S.; Prinetti, A. Gangliosides in membrane organization. *Prog. Mol. Biol. Transl. Sci.* **2018**, 156, 83-120.
25. Furukawa, K.; Ohmi, Y.; Ji, S.; Zhang, P.; Bhuiyan, R.H.; Ohkawa, Y.; Tajima, O.; Hashimoto, N.; Furukawa, K. Glycolipids: Essential regulator of neuro-inflammation, metabolism and gliomagenesis. *Biochim. Biophys. Acta. Gen Subj.* **2017**. 1861:2479-2484
26. Ohsawa, T. Changes of mouse brain gangliosides during aging from young adult until senescence. *Mech. Ageing Dev.* **1989**, 50, 169-177.
27. Hilbig, R.; Lauke, G.; Rahmann, H. Brain gangliosides during the life span (embryogenesis to senescence) of the rat. *Dev. Neurosci.* **1983-84**, 6, 260-270.
28. Yu, R.K.; Nakatani, Y.; Yanagisawa, M. The role of glycosphingolipid metabolism in the developing brain. *J. Lipid Res.* **2009**, 50, 440-445.
29. Segler-Stahl, K.; Webster, J.C.; Brunngraber, E.G. Changes in the concentration and composition of human brain gangliosides with aging. *Gerontology.* **1983**, 29, 161-168.
30. Svennerholm, L.; Boström, K.; Helander, C.G.; et al. Membrane lipids in the aging human brain. *J. Neurochem.* **1991**, 56, 2051-2059.
31. Svennerholm, L.; Boström, K.; Jungbjer, B.; et al. Membrane lipids of adult human brain: lipid composition of frontal and temporal lobe in subjects of age 20 to 100 years. *J. Neurochem.* **1994**, 63, 1802-1811.
32. Sarbu, M.; Dehelean, L.; Munteanu, C.V.; et al. Assessment of ganglioside age-related and topographic specificity in human brain by Orbitrap mass spectrometry. *Anal. Biochem.* **2017**, 521, 40-54.
33. Rogers, J.; Cooper, N.R.; Webster, S.; et al. Complement activation by

- beta-amyloid in Alzheimer disease. *Proc. Natl. Acad. Sci. USA.* **1992**, 89, 10016-10020.
34. Shen, Y.; Lue, L.; Yang, L.; et al. Complement activation by neurofibrillary tangles in Alzheimer's disease. *Neurosci. Lett.* **2001**, 305, 165-168.
35. Gasque, P.; Dean, Y.D.; McGreal, E.P.; et al. Complement components of the innate immune system in health and disease in the CNS. *Immuno-pharmacology.* **2000**, 49, 171-186.
36. Shen, Y.; Halperin, J.A.; Benzaquen, L.; Lee, C.M. Characterization of neuronal cell death induced by complement activation. *Brain Res. Brain Res. Protoc.* **1997**, 1, 186-94.
37. Terai, K.; Walker, D.G.; McGeer, E.G.; et al. Neurons express proteins of the classical complement pathway in Alzheimer disease. *Brain Res.* **1997**, 769, 385-390.
38. Yasojima, K.; Schwab, C.; McGeer, E.G.; et al. Up-regulated production and activation of the complement system in Alzheimer's disease brain. *Am. J. Pathol.* **1999**, 154, 927-936.
39. Jennemann, R.; Sandhoff, R.; Wang, S.; et al. Cell-specific deletion of glucosylceramide synthase in brain leads to severe neural defects after birth. *Proc. Natl. Acad. Sci. USA.* **2005**, 102, 12459-12464.
40. Tenner, A.J. Complement in Alzheimer's disease: opportunities for modulating protective and pathogenic events. *Neurobiol. Aging.* **2001**, 22, 849-861.
41. Stevens, B.; Allen, N.J.; Vazquez, L.E.; et al. The classical complement cascade mediates CNS synapse elimination. *Cell.* **2007**, 131, 1164-1178.
42. Wyss-Coray, T.; Yan, F.; Lin, A.H.; et al. Prominent neurodegeneration and increased plaque formation in complement- inhibited Alzheimer's mice. *Proc. Natl. Acad. Sci. USA.* **2002**, 99, 10837-10842.
43. Yanagisawa, K. Role of gangliosides in Alzheimer's disease. *Biochim. Biophys. Acta.* **2007**, 1768, 1943-1951.

44. Matsuzaki, K.; Kato, K.; Yanagisawa, K. A beta polymerization through interaction with membrane gangliosides. *Biochim. Biophys. Acta.* **2010**, 1801, 868-877.
45. Bernardo, A.; Harrison, F.E.; McCord, M.; et al. Elimination of GD3 synthase improves memory and reduces amyloid-beta plaque load in transgenic mice. *Neurobiol. Aging.* **2009**, 30, 1777-1791.
46. Oikawa, N.; Yamaguchi, H.; Ogino, K.; et al. Gangliosides determine the amyloid pathology of Alzheimer's disease. *Neuroreport.* **2009**, 20, 1043-1046.
47. Yamaguchi, T.; Yamauchi, Y.; Furukawa, K.; et al. Expression of B4GALNT1, an essential glycosyltransferase for the synthesis of complex gangliosides, suppresses BACE1 degradation and modulates APP processing. *Sci. Rep.* **2016**, 6, 34505. doi: 10.1038/srep34505.
48. Barrier, L.; Ingrand, S.; Damjanac, M.; et al. Genotype-related changes of ganglioside composition in brain regions of transgenic mouse models of Alzheimer's disease. *Neurobiol. Aging.* **2007**, 28, 1863-1872.
49. Schneider JS. Gangliosides and glycolipids in neurodegenerative disorders. *Adv. Neurobiol.* **2014**, 9, 449-61. doi: 10, 1007/978-1-4939-1154-7_20.
50. Takamiya, K.; Yamamoto, A.; Furukawa, K.; et al. Mice with disrupted GM2/GD2 synthase gene lack complex gangliosides but exhibit only subtle defects in their nervous system. *Proc. Natl. Acad. Sci. USA.* **1996**, 93, 10662-10667.
51. Wu, G.; Lu, ZH.; Kulkarni, N.; Ledeen, RW. Deficiency of ganglioside GM1 correlates with Parkinson's disease in mice and humans. *J. Neurosci. Res.* **2012**, 90, 1997-2008.
52. Schneider, J.S.; Pope, A.; Simpson, K.; Taggart, J.; Smith, M.G.; DiStefano, L. Recovery from experimental parkinsonism in primates with GM1 ganglioside treatment. *Science.* **1992**, 256(5058), 843-846.
53. Schneider, JS.; Gollomp, SM.; Sendek, S.; Colcher, A.; Cambi, F.; Du, W. A randomized, controlled, delayed start trial of GM1 ganglioside in treated Parkinson's disease patients. *J. Neurol. Sci.* **2013**, 324, 140-148.
54. Okada, M.; Itoh, M.; Haraguchi, M.; et al. b-series Ganglioside deficiency exhibits

- no definite changes in the neurogenesis and the sensitivity to Fas-mediated apoptosis but impairs regeneration of the lesioned hypoglossal nerve. *J. Biol. Chem.* **2002**, 277, 1633-1636
55. Akkhawattanakul, Y.; Maiti, P.; Xue, Y.; Aryal, D.; Wetsel, W.C.; Hamilton, D.; Fowler, S.C.; McDonald, M.P. Targeted deletion of GD3 synthase protects against MPTP-induced neurodegeneration. *Genes Brain Behav.* **2017**, 16, 522-536.
56. Furukawa, K.; Ohmi, Y.; Kondo, Y.; et al. Regulatory function of glycosphingolipids in the inflammation and degeneration. *Arch. Biochem. Biophys.* **2015**, 571, 58-65.
57. Sela, B.A.; Offner, H.; Konat, G.; et al. Immunological expression of gangliosides in multiple sclerosis and in a demyelinating model disease in rabbits. *Adv. Exp. Med. Biol.* **1984**, 174, 441-453.
58. Pilkington, G.J.; Dunan, J.R.; Rogers, J.P.; et al. Growth factor modulation of surface ganglioside expression in cloned neoplastic glia. *Neurosci. Lett.* **1993**, 149, 1-5.
59. Kawai, K.; Kuroda, S.; Watarai, S.; et al. Occurrence of GD3 ganglioside in reactive astrocytes--an immunocytochemical study in the rat brain. *Neurosci. Lett.* **1994**, 174, 225-227.
60. Welte, K.; Miller, G.; Chapman, P.B.; Yuasa, H.; Natoli, E.; Kunicka, J.E.; Cordon-Cardo, C.; Buhner, C.; Old, L.J.; Houghton, A.N. Stimulation of T lymphocyte proliferation by monoclonal antibodies against GD3 ganglioside. *J. Immunol.* **1987**, 139, 1763-1771.
61. Yamashiro, S.; Okada, M.; Haraguchi, M.; et al. Expression of alpha 2,8-sialyltransferase (GD3 synthase) gene in human cancer cell lines: high level expression in melanomas and up-regulation in activated T lymphocytes. *Glycoconj. J.* **1995**, 12, 894-900.
62. Yoshimura, A.; Takamiya, K.; Kato, I.; et al. GD2 ganglioside-specific monoclonal antibody reacts with murine cytotoxic T lymphocytes reactive with FBL-3N erythroleukaemia. *Scand. J. Immunol.* **1994**, 40, 557-563.

63. Furukawa, K.; Akagi, T.; Nagata, Y.; et al. GD2 ganglioside on human T-lymphotropic virus type I-infected T cells: possible activation of beta-1,4-N-acetylgalactosaminyltransferase gene by p40tax. *Proc. Natl. Acad. Sci. USA*. **1993**, 90, 1972-1976.
64. Okada, M.; Furukawa, K.; Yamashiro, S.; et al. High expression of ganglioside alpha-2,8-sialyltransferase (GD3 synthase) gene in adult T-cell leukemia cells unrelated to the gene expression of human T-lymphotropic virus type I. *Cancer Res.* **1996**, 56, 2844-2848.
65. Villanueva-Cabello, T.M.; Mollicone, R.; Cruz-Muñoz, M.E.; López-Guerrero, D.V.; Martínez-Duncker, I. Activation of human naïve Th cells increases surface expression of GD3 and induces neoexpression of GD2 that colocalize with TCR clusters. *Glycobiology*. **2015**, 25, 1454-1464.
66. Nagata, Y.; Yamashiro, S.; Yodoi, J.; et. al. Expression cloning of beta 1,4 N-acetylgalactosaminyltransferase cDNAs that determine the expression of GM2 and GD2 gangliosides. *J. Biol. Chem.* **1992**, 267, 12082-12089.
67. Haraguchi, M.; Yamashiro, S.; Yamamoto, A.; et al. Isolation of GD3 synthase gene by expression cloning of GM3 alpha-2,8-sialyltransferase cDNA using anti-GD2 monoclonal antibody. *Proc. Natl. Acad. Sci. USA*. **1994**, 91, 10455-10459.
68. Nara, K.; Watanabe, Y.; Maruyama, K.; Kasahara, K.; Nagai, Y.; Sanai Y.: Expression cloning of a CMP-NeuAc:NeuAc alpha 2-3Gal beta 1-4Glc beta 1-1'Cer alpha 2,8-sialyltransferase (GD3 synthase) from human melanoma cells. *Proc. Natl. Acad. Sci. USA*. **1994**, 91, 7952-7956.
69. Sasaki, K.; Kurata, K.; Kojima, N.; Kurosawa, N.; Ohta, S.; Hanai, N.; Tsuji, S.; Nishi, T.: Expression cloning of a GM3-specific alpha-2,8-sialyltransferase (GD3synthase). *J. Biol. Chem.* **1994**, 269, 15950-15956.
70. Miyazaki, H.; Fukumoto, S.; Okada, M.; et al. Expression cloning of rat cDNA encoding UDP-galactose: GD2 beta1,3-galactosyltransferase that determines the expression of GD1b/GM1/GA1. *J. Biol. Chem.* **1997**, 272, 24794-24799.
71. Fukumoto, S.; Miyazaki, H.; Goto, G.; et. al. Expression cloning of mouse cDNA

- of CMP-NeuAc: Lactosylceramide alpha2,3-sialyltransferase, an enzyme that initiates the synthesis of gangliosides. *J. Biol. Chem.* **1999**, 274, 9271- 9276.
72. Kono, M.; Takashima, S.; Liu, H.; Inoue, M.; Kojima, N.; Lee, Y.C.; Hamamoto, T.; Tsuji, S. Molecular cloning and functional expression of a fifth-type alpha 2,3-sialyltrans- ferase (mST3Gal V: GM3 synthase). *Biochem. Biophys. Res. Commun.* **1998**, 253, 170-175
73. Ishii, A.; Ohta, M.; Watanabe, Y.; Matsuda, K.; Ishiyama, K.; Sakoe, K.; Nakamura, M.; Inokuchi, J.; Sanai, Y.; Saito, M. Expression cloning and functional characterization of human cDNA for ganglioside GM3 synthase. *J. Biol. Chem.* **1998**, 273, 31652-31655.
74. Kojima, Y.; Fukumoto, S.; Furukawa, K.; et al. Molecular cloning of globotriaosylceramide/CD77 synthase, a glycosyltransferase that initiates the synthesis of globo series glycosphingolipids. *J. Biol. Chem.* **2000**, 275, 15152-15156.
75. Steffensen, R.; Carlier, K.; Wiels, J.; Levery, S.B.; Stroud, M.; Cedergren, B.; Nilsson Sojka, B.; Bennett, E.P.; Jersild, C.; Clausen, H. Cloning and expression of the histo-blood group Pk UDP-galactose: Ga1beta-4G1cbeta1-cer alpha1, 4-galactosyltransferase. Molecular genetic basis of the p phenotype. *J. Biol. Chem.* **2000**, 275, 16723-16729.
76. Keusch, J.J.; Manzella, S.M.; Nyame, K.A.; Cummings, R.D.; Baenziger, J.U. Cloning of Gb3 synthase, the key enzyme in globo-series glycosphingolipid synthesis, predicts a family of alpha 1, 4-glycosyltransferases conserved in plants, insects, and mammals. *J. Biol. Chem.* 2000, 275, 25315-25321.
77. Okuda, T.; Tokuda, N.; Numata, S.; et al. Targeted disruption of Gb3/CD77 synthase gene resulted in the complete deletion of globo-series glycosphingolipids and loss of sensitivity to verotoxins. *J. Biol. Chem.*, **2006**, 281, 10230-10235.
78. Nishie, T.; Hikimochi, Y.; Zama, K.; et. al. Beta4-galactosyltransferase-5 is a lactosylceramide synthase essential for mouse extra-embryonic development. *Glycobiology.* **2010**, 20, 1311-1322.
79. Tokuda, N.; Numata, S.; Li, X.; et al. β 4GalT6 is involved in the synthesis of

- lactosylceramide with less intensity than β 4GalT5. *Glycobiology*. **2014**, *23*, 1175-1183.
80. Inoue, M.; Fujii, Y.; Furukawa, K.; et al. Refractory skin injury in the complex knock-out mice expressing only GM3 ganglioside. *J. Biol. Chem.* **2002**, *277*, 29881-29888.
81. Tajima, O.; Egashira, N.; Ohmi, Y.; Fukue, Y.; Mishima, K.; Iwasaki, K.; Fujiwara, M.; Inokuchi, J.; Sugiura, Y.; Furukawa, K.; Furukawa, K. Reduced motor and sensory functions and emotional response in GM3-only mice: emergence from early stage of life and exacerbation with aging. *Behav. Brain Res.* **2009**, *198*, 74-82.
82. Yoshihara, T.; Satake, H.; Nishie, T.; Okino, N.; Hatta, T.; Otani, H.; Naruse, C.; Suzuki, H.; Sugihara, K.; Kamimura, E.; Tokuda, N.; Furukawa, K.; Furukawa, K.; Ito, M.; Asano, M. Lactosylceramide synthases encoded by B4galt5 and 6 genes are pivotal for neuronal generation and myelin formation in mice. *PLoS Genet.* **2018**, *14*, e1007545 doi: 10.1371/journal.pgen.1007545. eCollectio
83. Ohmi, Y.; Tajima, O.; Ohkawa, Y.; et al. Gangliosides play pivotal roles in the regulation of complement systems and in the maintenance of integrity in nerve tissues. *Proc. Natl. Acad. Sci. USA.* **2009**, *106*, 22405-22410.
84. Ohmi, Y.; Tajima, O.; Ohkawa, Y.; et al. Gangliosides are essential in the protection of inflammation and neurodegeneration via maintenance of lipid rafts: elucidation by a series of ganglioside-deficient mutant mice. *J. Neurochem.* **2011**, *116*, 926-935.
85. Schengrund, C.L. The roles of gangliosides in neural differentiation and repair: a perspective. *Brain Res. Bull.* **1990**, *24*, 131-141.
86. Furukawa, K.; Tajima, O.; Okuda, T.; et. al. Knockout Mice and Glycolipids. In *Comprehensive Glycoscience. From Chemistry to Systems Biology*, vol.4, Kamerling, J.P.; Boons, G.J.; Lee, Y.C.; et al. (Eds.), Oxford: Elsevier, **2007**, pp. 149-157.
87. Sugiura, Y.; Furukawa, K.; Tajima, O.; et. al. Sensory nerve-dominant nerve degeneration and remodeling in the mutant mice lacking complex gangliosides.

- Neuroscience*, **2005**, 135, 1167-1178.
88. Sheikh, K.A.; Sun, J.; Liu, Y.; et al. Mice lacking complex gangliosides develop Wallerian degeneration and myelination defects. *Proc. Natl. Acad. Sci. USA*. **1999**, 96, 7532-7537.
89. Kawai, H.; Allende, M.L.; Wada, R.; Kono, M.; Sango, K.; Deng, C.; Miyakawa, T.; Crawley, J.N.; Werth, N.; Bierfreund, U.; Sandhoff, K.; Proia, R.L. Mice expressing only monosialoganglioside GM3 exhibit lethal audiogenic seizures. *J. Biol. Chem.* **2001**, 276, 6885-6888.
90. Ichikawa, S.; Sakiyama, H.; Suzuki, G.; Hidari, K.I.; Hirabayashi, Y. Expression cloning of a cDNA for human ceramide glucosyltransferase that catalyze the first glycosylation step of glycosphingolipid synthesis. *Proc. Natl. Acad. Sci. USA*. **1996**, 93, 4638-4643.
91. Yamashita, T.; Wada, R.; Sasaki, T.; et al. A vital role for glycosphingolipid synthesis during development and differentiation. *Proc. Natl. Acad. Sci. USA*. **1999**, 96, 9142-9147.
92. Michel, V.; Bakovic, M. Lipid rafts in health and disease. *Biol. Cell*. **2007**, 99, 129-140.
93. Ohmi, Y.; Ohkawa, Y.; Tajima, O.; Sugiura, Y.; Furukawa, K.; Furukawa, K. Ganglioside deficiency causes inflammation and neurodegeneration via the activation of complement system in the spinal cord. *J. Neuroinflammation*. **2014**, 11:61. doi: 10.1186/1742-2094-11-61
94. McGonigal, R.; Rowan, E.G.; Greenshields, K.N.; et al. Anti-GD1a antibodies activate complement and calpain to injure distal motor nodes of Ranvier in mice. *Brain*. **2010**, 133, 1944-1960.
95. McGonigal, R.; Cunningham, M.E.; Yao, D.; et al. C1q-targeted inhibition of the classical complement pathway prevents injury in a novel mouse model of acute motor axonal neuropathy. *Acta Neuropathol. Commun.* **2016**, 4, 23 doi: 10.1186/s40478-016-0291-x
96. Halstead, S.K.; Zitman, F.M.; Humphreys, P.D.; et al. Eculizumab prevents

- anti-ganglioside antibody-mediated neuropathy in a murine model. *Brain*. **2008**, 131, 1197-1208.
97. Davidson, A.I.; Halstead, S.K.; Goodfellow, J.A.; et al. Inhibition of complement in Guillain-Barré syndrome: the ICA-GBS study. *J. Peripher. Nerv. Syst.* **2017**, 22, 4-12.
98. Furukawa, K.; Ohmi, Y.; Ohkawa, Y.; et al. Roles of glycosphingolipids in the regulation of the membrane organization and cell signaling in lipid rafts. In *Membrane Organization and Lipid Rafts in the Cell and Artificial Membranes*. Catala, A. (Ed.), London, Nova Science Publishers, **2016**, pp. 129-146.
99. Simons, K.; Ikonen, E. Functional rafts in cell membranes. *Nature*. **1997**, 387, 569-572.
100. Hakomori, S.; Yamamura, S.; Handa, A.K. Signal transduction through glyco(sphingo)lipids. Introduction and recent studies on glyco(sphingo)lipid-enriched microdomains. *Ann. N Y Acad. Sci.* **1998**, 845, 1-10.
101. Hamamura, K.; Tsuji, M.; Hotta, H.; et al. Functional activation of Src family kinase yes protein is essential for the enhanced malignant properties of human melanoma cells expressing ganglioside GD3. *J. Biol. Chem.* **2011**, 286, 18526-18537.
102. Ohkawa, Y.; Miyazaki, S.; Hamamura, K.; et al. Ganglioside GD3 enhances adhesion signals and augments malignant properties of melanoma cells by recruiting integrins to glycolipid-enriched microdomains. *J. Biol. Chem.* **2010**, 285, 27213-27223.
103. Kotani, N.; Gu, J.; Isaji, T.; et al. Biochemical visualization of cell surface molecular clustering in living cells. *Proc. Natl. Acad. Sci. USA*. **2008**, 105, 7405-7409.
104. Hakomori, S.I. Cell adhesion/recognition and signal transduction through glycosphingolipid microdomain. *Glycoconj. J.* **2000**, 17, 143-151.
105. Harder, T.; Simons, K. Caveolae, DIGs, and the dynamics of sphingolipid-cholesterol microdomains. *Curr. Opin. Cell Biol.* **1997**, 9, 534-542.

106. Nakashima, H.; Hamamura, K.; Houjou, T.; et al. Overexpression of caveolin-1 in a human melanoma cell line results in dispersion of ganglioside GD3 from lipid rafts and alteration of leading edges, leading to attenuation of malignant properties. *Cancer Sci.* **2007**, *98*, 512-520.
107. Fujita, A.; Cheng, J.; Hirakawa, M.; Furukawa, K.; Kusunoki, S.; Fujimoto, T. Gangliosides GM1 and GM3 in the living cell membrane form clusters susceptible to cholesterol depletion and chilling. *Mol. Biol. Cell.* **2007**, *18*, 2112-2122.
108. Ohmi, Y.; Ohkawa, Y.; Yamauchi, Y.; Tajima, O.; Furukawa, K.; Furukawa, K. Essential roles of gangliosides in the formation and maintenance of membrane microdomains in brain tissues. *Neurochem. Res.* **2012**, *37*, 1185-1191.
109. Yang, L.B.; Li, R.; Meri, S.; et al. Deficiency of complement defense protein CD59 may contribute to neurodegeneration in Alzheimer's disease. *J. Neurosci.* **2000**, *20*, 7505-7509.
110. Mason, J.C.; Yarwood, H.; Sugars, K.; et al. Induction of decay-accelerating factor by cytokines or the membrane-attack complex protects vascular endothelial cells against complement deposition. *Blood.* **1999**, *94*, 1673-1682.
111. Kinoshita, T.; Fujita, M.; Maeda, Y. Biosynthesis, remodelling and functions of mammalian GPI-anchored proteins: recent progress. *J. Biochem.*, **2008**, *144*, 287-294.
112. Simpson, M.A.; Cross, H.; Proukakis, C.; Priestman, D.A.; Neville, D.C.; Reinkensmeier, G.; Wang, H.; Wiznitzer, M.; Gurtz, K.; Verganelaki, A.; et al. Infantile-onset symptomatic epilepsy syndrome caused by a homozygous loss-of-function mutation of GM3 synthase. *Nat. Genet.* **2004**, *36*, 1225-1229.
113. Yu, R.K.; Nakatani, Y.; Yanagisawa, M. The role of glycosphingolipid metabolism in the developing brain. *J. Lipid Res.* **2009**, *50* Suppl, S440-445.
114. Boccuto, L.; Aoki, K.; Flanagan-Steet, H.; Chen, C.F.; Fan, X.; Bartel, F.; Petukh, M.; Pittman, A.; Saul, R.; Chaubey, A.; et al. A mutation in a ganglioside biosynthetic enzyme, ST3GAL5, results in salt & pepper syndrome, a neurocutaneous disorder with altered glycolipid and glycoprotein glycosylation.

- Hum. Mol. Genet.* **2014**, *23*, 418-433.
115. Wang, H.; Bright, A.; Xin, B.; Bockoven, J.R.; Paller, A.S. Cutaneous dyspigmentation in patients with ganglioside GM3 synthase deficiency. *Am. J. Med. Genet. A.* **2013**, *161A*, 875-879.
116. Yoshikawa, M.; Go, S.; Suzuki, S.; Suzuki, A.; Katori, Y.; Morlet, T.; Gottlieb, S.M.; Fujiwara, M.; Iwasaki, K.; Strauss, K.A.; Inokuchi, J. Ganglioside GM3 is essential for the structural integrity and function of cochlear hair cells. *Hum. Mol. Genet.* **2015**, *24*, 2796-2807.
117. Boukhris, A.; Schule, R.; Loureiro, J.L.; Lourenço, C.M.; Mundwiler, E.; Gonzalez, M.A.; Charles, P.; Gauthier, J.; Rekik, I.; Acosta Lebrigio, R.F.; et al. Alteration of ganglioside biosynthesis responsible for complex hereditary spastic paraplegia. *Am. J. Hum. Genet.* **2013**, *93*, 118-123.
118. Harlalka, G.V.; Lehman, A.; Chioza, B.; Baple, E.L.; Maroofian, R.; Cross, H.; Sreekantan-Nair, A.; Priestman, D.A.; Al-Turki, S.; McEntagart, M.E.; et al. Mutations in B4GALNT1 (GM2 synthase) underlie a new disorder of ganglioside biosynthesis. *Brain.* **2013**, *136*, 3618-3624.
119. Wakil, S.M.; Monies, D.M.; Ramzan, K.; Hagos, S.; Bastaki, L.; Meyer, B.F.; Bohlega, S. Novel B4GALNT1 mutations in a complicated form of hereditary spastic paraplegia. *Clin. Genet.* **2014**, *86*, 500-501.
120. Bhuiyan, R.H.; Ohmi, Y.; Ohkawa, Y.; Zhang, P.; Takano, M.; Hashimoto, N.; Okajima, T.; Furukawa, K.; Furukawa, K. Loss of enzyme activity in mutated *B4GALNT1* gene products found in patients with hereditary spastic paraplegia resulted in relatively mild neurological disorders: Similarity with phenotypes of *B4galnt1* knockout mice. *Neuroscience* **2018**, *397*:94-106.
121. Takamiya, K.; Yamamoto, A.; Furukawa, K.; Zhao, J.; Fukumoto, S.; Yamashiro, S.; Okada, M.; Haraguchi, M.; Shin, M.; Kishikawa, M.; et al. Complex gangliosides are essential in spermatogenesis of mice: possible roles in the transport of testosterone. *Proc. Natl. Acad. Sci. USA.* **1998**, *95*, 12147-12152.

122. Kitajima, K.; Varki, N.; Sato, C. Advanced Technologies in Sialic Acid and Sialoglycoconjugate Analysis. *Top. Curr. Chem.* **2015**, 367, 75-103.
123. Cavdarli, S.; Dewald, J.H.; Yamakawa, N.; Guérardel, Y.; Terme, M.; Le Doussal, J.M.; Delannoy, P.; Groux-Degroote, S. Identification of 9-O-acetyl-N-acetylneuraminic acid (Neu5,9Ac₂) as main O-acetylated sialic acid species of GD2 in breast cancer cells. *Glycoconj. J.* **2019**, 36, 79-90.
124. Nakayama, H.; Nagafuku, M.; Suzuki, A.; Iwabuchi, K.; Inokuchi, J.I. The regulatory roles of glycosphingolipid-enriched lipid rafts in immune systems. *FEBS Lett.* **2018**, 592, 3921-3942.
125. Kondo, Y.; Ikeda, K.; Tokuda, N.; Nishitani, C.; Ohto, U.; Akashi, T.S.; Ito, Y.; Uchikawa, M.; Kuroki, Y.; Taguchi, R.; et al. TLR4-MD-2 complex is negatively regulated by an endogenous ligand, globotetraosylceramide. *Proc. Natl. Acad. Sci. USA.* **2013**, 110, 4714-4719.
126. Hashimoto, N.; Ito, S.; Tsuchida, A.; Bhuiyan, R.H.; Okajima, T.; Yamamoto, A.; Furukawa, K.; Ohmi, Y.; Furukawa, K. The ceramide moiety of disialoganglioside (GD3) is essential for GD3 recognition by the sialic acid-binding lectin SIGLEC7 on the cell surface. *J. Biol. Chem.* **2019**, 294, 10833-10845.
127. Yao, D.; McGonigal, R.; Barrie, J.A.; Cappell, J.; Cunningham, M.E.; Meehan, G.R.; Fewou, S.N.; Edgar, J.M.; Rowan, E.; Ohmi, Y.; et al. Neuronal expression of GalNAc transferase is sufficient to prevent the age-related neurodegenerative phenotype of complex ganglioside deficient mice. *J. Neurosci.* **2014**, 34, 880-891.
128. Komura, N.; Suzuki, K.G.; Ando, H.; Konishi, M.; Koikeda, M.; Imamura, A.; Chadda, R.; Fujiwara, T.K.; Tsuboi, H.; Sheng, R.; et al. Ultra-high-resolution imaging of gangliosides in GEM/rafts Raft-based interactions of gangliosides with a GPI-anchored receptor. *Nat. Chem. Biol.* **2016**, 12, 402-410.
129. Lingwood, D.; Simons, K. Lipid rafts as a membrane-organizing principle. *Science.* **2010**, 327, 46-50.

130. Furukawa, K.; Ohmi, Y.; Ohkawa, Y.; Bhuiyan, RH.; Zhang, P.; Tajima, O.; Hashimoto, N.; Hamamura, K.; Furukawa, K. New era of research on cancer-associated glycosphingolipids. *Cancer Sci.* **2019**, 110, 1544-1551.
131. Furukawa, K.; Ohmi, Y.; Ohkawa, Y.; Tajima, O.; Furukawa, K. Glycosphingolipids in the regulation of the nervous system. *Adv. Neurobiol.* **2014**, 9, 307-320

Figure legends

Fig. 1. Glycosphingolipids are amphipathic molecules, and expressed in the out layer of lipid two layers of cell membrane. Hydrophobic portion (ceramide) is embedded in the outer layer of the membrane, and sugar portion is protruding outside of the membrane.

Fig. 2. Main pathway of ganglioside synthesis. Glycosyltransferases catalyzing individual steps were shown by italics, and deleted structures in KO mice were encircled by different colored squares.

Fig. 3. Immunohistochemistry of GFAP-positive astrocytes and F4/80 antibody-reactive microglia.

Marked gliosis were found in ganglioside synthase gene KO mice. GFAP-positive cells were increased (left), and F4/80-positive cells (right) were assembled in cerebella, indicating astrocytes and microglia, respectively.

Fig. 4. Inflammatory reactions in the DKO mice were suppressed by genetic disruption of complement system. A, Triple KO (TKO) mice was generated by mating the DKO mice and C3 KO mice to clarify the roles of complement systems in the brain disorders in DKO mice. B, Expression levels of inflammatory cytokines were reduced in TKO mice.

Fig. 5. Destruction of GEM/raft induced changes in the architecture and functions of raft-localizing molecules. Localization of GPI-anchored proteins including complement-regulatory molecules changed, leading to functional abnormalities and subsequent complement activation and inflammation in ganglioside-deficient mice brain.

Table 1. Deficient structures, remaining glycolipids and phenotypes of individual KO mice of glycosyltransferase genes. (attached in figures)