

# 1 Genetic variation of traits in natural enemies 2 relevant for biological control: a systematic review

## 3 Authors

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12

## 13 Abstract

14 The concept of genetic improvement in relation to biological control involves the  
15 exploitation of natural genetic variation for the benefit of existing biological control  
16 agents (BCAs). Despite recent calls for this process to be adopted in biological control  
17 research, there is no clear overview of the current state of research into genetic  
18 variation within a biological control context, including quantifiable estimates such as  
19 narrow-sense heritability ( $h^2$ ). In this systematic review, we aim to determine the  
20 current state of research on the genetic variation of biological control traits in natural  
21 enemies. After the searching process, screening for papers that can deliver on our  
22 research question reduced the initial 2,927 search hits to only a mere 69 papers for  
23 data extraction. Of these, the majority (73.6%) did not report quantitative values for  
24 genetic variation. Extracting the traits measured in these papers, we categorized  
25 them according to two approaches; the first related to fitness components, and the  
26 second related to biological control importance. This systematic review highlights the  
27 need for more rigorous reporting of the quantitative values of genetic variation to  
28 enable the successful genetic improvement of biological control agents.

## 29 Keywords

30 Genetic improvement, genetic variation, heritability, systematic review, biocontrol  
31 agent, life history traits

32

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## 35 1. Introduction

36 With a rising global population, as well as a changing global climate, food production  
37 is increasingly under pressure (IPCC, 2007; Misra, 2014). Rising arthropod pest pressure  
38 (due to modern monoculture practice as well as expanding pest ranges related to  
39 global change (Das et al., 2011)), coupled with a desire for more environmentally  
40 sustainable agriculture (i.e. using less pesticides) (Rigby and Cáceres, 2001), has led  
41 to an increased interest in biological control for pest management (van Lenteren et  
42 al., 2018). The use of arthropod natural enemies as biological control agents (BCAs) is  
43 not new, with records of first releases dating back over 100 years (Barratt et al., 2018).  
44 For several decades, imported exotic natural enemies were relied upon to control  
45 non-native pests, which are becoming increasingly more prevalent due to expanding  
46 or shifting ranges linked with changing climates (Aguilar-Fenollosa and Jacas, 2014;  
47 van Lenteren et al., 2006). By taking the natural enemy from the native range of the  
48 pest, it was considered to be better adapted to tackling the pest due to their shared  
49 evolutionary past. Due to knowledge gained on the risks of releasing non-native  
50 species, along with stricter regulations on the import and export of species (such as  
51 the Nagoya Protocol on Access and Benefit Sharing, 2010), this practice has become  
52 less desirable (CBD Secretariat, 2011; Lommen et al., 2017). These recent  
53 developments in combination with an expansion of pest ranges due to global  
54 warming and the anthropogenic movement of produce across the globe, make it  
55 more important than ever to improve the efficacy of those native natural enemies  
56 that are already at our disposal (Leung et al., 2019; Lommen et al., 2017).

57 Selection of effective BCAs is usually determined at species level, by looking at  
58 interspecific variation (Lommen et al., 2017), i.e. finding the best species to tackle a  
59 certain pest. More recently, there is increased attention on exploiting the large

60 amount of intraspecific variation between, and even within, populations (Kruitwagen  
61 et al., 2018; Lommen et al., 2017). Recent studies have also suggested the use of  
62 genetic modification for increased effectiveness of BCAs, particularly in fungal and  
63 pathogen-based agents (Karabörklü et al., 2018; Lovett and St. Leger, 2018; St. Leger  
64 and Wang, 2010), as well as insects (Poppy and Powell, 2009; Routray et al., 2016).  
65 However, as with using genetically modified crop plants, the release of genetically  
66 modified natural enemies would come under strict regulation (Barratt et al., 2018;  
67 Reeves et al., 2012) and does not seem compatible with the eco-friendly mission of  
68 biological control. On the other hand, artificial selection of traits is an accepted  
69 method already commonly used across agriculture, such as in livestock and plant  
70 breeding (Brotherstone and Goddard, 2005), which can be exploited to effectively  
71 improve BCAs (Kruitwagen et al., 2018; Lommen et al., 2017).

72 A recent review on the use of experimental evolution in biological control supports  
73 the potential of artificial selection for improving BCAs (Lirakis and Magalhães, 2019).  
74 The key prerequisite for artificial selection is the presence of genetic variation for the  
75 trait of interest. The prime estimate of genetic variation is heritability, which is defined  
76 as the proportion of the total variation between individuals in a defined population  
77 that is due to genetic variation (Lommen et al., 2017). Heritability or evolvability is  
78 regularly studied within the realm of basic and evolutionary research, and often so  
79 with insects due to the ease of their rearing and workability in a controlled laboratory  
80 setting. However, it has been studied to a lesser extent within an applied context. This  
81 can be to the detriment of BCAs and their effective use, both in regards to deleterious  
82 impacts of genetic drift and subsequent loss of efficacy (Paspati et al., 2019; Szűcs et  
83 al., 2019), and that without variation data for traits of interest, there is no way of  
84 knowing if the trait in question can be improved through genetic selection (Houle,  
85 1992). As for the latter, full reporting of values such as variance and determining

86 evolvability in place of heritability values was proposed more than twenty years ago  
87 (Hansen et al., 2011; Houle, 1992) as a more appropriate measure, yet the uptake has  
88 been slow and inaccurate calculations remain a concern (Garcia-Gonzalez et al.,  
89 2012).

90 With the increased interest in the role of experimental evolution and artificial selection  
91 in the improvement of biological control (Kruitwagen et al., 2018; Leung et al., 2019;  
92 Lirakis and Magalhães, 2019; Lommen et al., 2017; Routray et al., 2016), it is necessary  
93 to determine what has already been assessed with regards to genetic variation and  
94 heritable traits of BCAs, and where there are gaps in our knowledge.

95 In this systematic review, our goal is to compile and summarize findings from across  
96 literature regarding genetic variation in BCAs for traits that are potentially useful for  
97 biological control, regardless of whether these studies have an applied focus or not.  
98 This will identify the advances already made with regards to identifying suitable  
99 heritable traits within arthropod BCAs, and where more work is necessary.

## 100 2. Methods

### 101 2.1 Search Criteria

102 As the steps of a meta-analysis and systematic review are similar up until the data  
103 extraction phase (Côté and Jennions, 2013) for searching, screening, and the  
104 extraction protocol, we followed the steps outlined in Chapter 2 of the Handbook of  
105 Meta-Analysis in Ecology and Evolution (Koricheva et al., 2013). An overview of the  
106 process is provided in Figure 1. Our question was refined to a single search term that  
107 focused on two aspects: BCAs and genetic variation or heritability. We did not wish to  
108 limit our initial search results by *a priori* defining traits of interest.

109 The databases used were CAB Abstracts (1910 to 2019), the Biological Abstracts  
110 database (1969 to 2016), and the Agricola database (1970 to April 2019), which were  
111 all accessed via the CAB Abstracts search platform. Additional search parameters  
112 were chosen, including using the "field= keyword" search field (searching for terms in  
113 the title, abstract, or keywords), as well as limiting document type to 'Article' and  
114 'Review.' No language restrictions were imposed at the time of the search.

115 To validate our search term, a positive control group was set up of 15 papers that  
116 would suit the purpose of initial screening for our research question. This set of controls  
117 can be found in the Appendix (Table S1).

118 As we were interested in finding publications that assessed genetic variation or  
119 heritability of different traits of BCAs, the search term contained various terms and  
120 wildcard symbols (\*) to aid in the search, as well as the AND/OR operator to tie the  
121 two ideas together. To limit the search to arthropod BCAs, the NOT operator was used  
122 in conjunction to avoid the inclusion of several undesired returns.

123 Consequently, the final search term was as follows:

124 ("genetic varia\*" OR "genotypic varia\*" OR heritab\*)

125 AND (enem\* OR predat\* OR parasitoid\* OR "parasitic wasp\*" OR  
126 biological control OR biocontrol)

127 NOT (virus\* OR nematode\* OR weed\* OR fung\*)

128 Initial search hits were deduplicated using the duplication removal tool found  
129 within the CAB Abstracts search tool (based on *Abstract*). The total bibliographic  
130 information for these hits were downloaded and arranged into a table for further  
131 screening. Search queries were performed on September 17, 2018 and May 16, 2019.

132 Duplicate entries overlapping both search periods were removed manually, while  
133 some duplicates remained in the dataset until screening. Importantly, all positive  
134 controls were returned in the search results.

## 135 2.2 Screening search results

136 Unique entries were subject to a preliminary screening that sought to determine,  
137 based on title, abstract, and any keywords whether the entries were appropriate for  
138 the systematic review. This initial screening was performed by a team of 16 readers,  
139 and was based solely on content related to study species: first, if the species of study  
140 (as determined in the title or abstract) was truly an arthropod; and second, if the  
141 species of study was classified as a BCA by checking whether it appeared on either  
142 the EPPO list of BCAs available for use in Europe (PM 6/003, both 2016 and 2019  
143 versions, <https://gd.eppo.int/standards/PM6/>), or on the list of BCA species compiled  
144 in previous reviews (van Lenteren, 2012; van Lenteren et al., 2018). This list contains 361  
145 species names, some of which are former scientific names that are no longer in use,  
146 but allow for scanning older papers.

147 Two additional screening measures occurred: on language and availability. First,  
148 language was either listed as a variable from the database bibliographical  
149 information or was assessed by viewing the full text of the entry. Languages that were  
150 understood by our team of readers were assessed, while languages not understood  
151 by the team remained unassessed. Second, unavailable entries were listed as  
152 'missing', that is, while the entry existed in one of the three databases used for the  
153 initial search, neither electronic nor physical records in the Wageningen University  
154 Library network were available and additional online queries yielded no records.

155 Remaining duplicates (based on entry title), as well as conference proceedings, were  
156 removed manually.

157 At this point, each entry in the dataset corresponds to a single peer-reviewed article.  
158 The final screening category was 'relevance'. Relevance to the review terms was  
159 evaluated based on the article's abstract, title, or full-text in some cases, and was  
160 determined based on two parameters: 'methods' and 'trait of interest'. If the methods  
161 allowed for determining of genetic variation, the article was deemed relevant. Such  
162 methods would include the use of isofemale lines, tracking over multiple generations,  
163 common garden set-ups, parent-offspring regressions, or a type of sibling or crossing  
164 strategy. For the second parameter, 'traits of interest', it was the presence of any kind  
165 of phenotypic trait being studied that was important. For instance, articles that solely  
166 focused on population structure via neutral molecular markers, but were not looking  
167 at phenotypic traits, were deemed unrelated to this systematic review. All articles  
168 determined to be relevant were then subjected to full-text assessment and data  
169 extraction.

### 170 2.3 Data extraction and categorization

171 In-depth data extraction was performed by six readers, with a follow-up by two  
172 readers to double-check the values of the extractions. Each article was viewed in print  
173 and assessed for the following aspects: 1) estimation method, 2) trait of interest, 3)  
174 species, and 4) population values (such as size and number of populations). When  
175 applicable, a wide range of variables related to each trait and population that were  
176 assessed in the study were extracted as well. Interest was paid to values directly  
177 informative of genetic variation, such as narrow-sense heritability ( $h^2$ ), broad-sense  
178 heritability ( $H^2$ ), or evolvability ( $CV_A$ ) estimates. For a single article, several unique data

179 extraction outputs (termed 'observations') are possible, and all received a unique  
180 identification number in order to track the total data extracted. This granularity allows  
181 for assessing how often a trait is measured for different populations or species in various  
182 studies, as multiple unique values related to genetic variation of traits of interest are  
183 possible for a single article.

184 For each trait of interest, categories were assigned to each observation to refine the  
185 extraction data to clear themes. Two categorical systems were used. The first system  
186 uses the categories of phenotypic traits based on biological function as defined by  
187 Mousseau and Roff (1987): physiology (P), morphology (M), life history (L), and  
188 behavior (B) (Mousseau and Roff, 1987). These categories are explicit, and each trait  
189 can only be attributed to a single category. The second system is based on the  
190 application of biological control and uses categories as defined in Leung et al 2019,  
191 which posits four aspects of biological control that could be improved using genetic  
192 knowledge: pest suppression ability (1), adaptation to abiotic factors (2), reducing  
193 ecological risk (3), and improving mass rearing conditions (4). Three additional  
194 categories were created to address the outcomes of this extraction, and include:  
195 insecticide resistance (5), adaptation to biotic factors (6), and a null category named  
196 'unrelated' (7), as to avoid forcing a trait into one of the other six categories. This  
197 second classification is not explicit, and one trait may be attributed to multiple  
198 categories. Simple calculations (counts and means) were performed manually, while  
199 figure generation was performed using R (version 3.5.0), R Studio (version 1.2.1335),  
200 and ggplot2 (version 3.2.0)(R Core Team, 2015; RStudio Team, 2015; Wickham, 2016).

201



## 202 3. Results

### 203 3.1 Search results

204 The first search using the full search term yielded 3,612 hits but was reduced to 2,880  
205 following deduplication (Figure 1). The second search, carried out eight months later,  
206 yielded 3,471 hits, and deduplication reduced it to 2,891. This includes new articles  
207 published as well as any entries added to any of the three databases within the  
208 intervening period. From the second search, 87 unique hits were found that cover the  
209 period between the first and second searches (September 2018 to April 2019).  
210 Overlapping hits were removed by hand, resulting in a total of 2,927 hits for the  
211 systematic review. It is worth noting that some hits that were in the first search were no  
212 longer present in the database for the second search. The complete dataset of  
213 search returns at this point are available in the Appendix (Table S2) in the format  
214 delivered by the CAB Abstracts Database.

215 Results of this combined search return, according to year of publication, can be found  
216 in Figure 2. Year of publication ranges from 1914 to 2019. Several years in the earlier  
217 time of the search returned no hits, representing the true minimum. Fifteen different  
218 years are represented by only a single hit, while the maximum belongs to 2014 with  
219 181 entries. Values skew closer to the present-day, likely attributed to both the well-  
220 documented increase in scientific publishing (Larsen and von Ins, 2010) and a general  
221 increase in biological control studies. Similar results occurred in a recent review on  
222 zoophytophagous predators (Puentes, Stephan, and Björkman, 2018). The majority of  
223 returns are from the CAB Abstracts database (1304), which was chosen as the  
224 preferred database for the search and deduplication, followed by Biological  
225 Abstracts (969), and Agricola (654).

## 226 3.2 Screening

227 Preliminary screening was performed to remove all the entries that were non-  
228 arthropods, not on the EPPO or van Lenteren lists, or those that were problematic  
229 (duplicates, missing, or languages unavailable to readers). From the 2,927 entries, only  
230 1,606 were entries containing arthropod research (1,321 non-arthropods were  
231 removed). From these 1,606, only 393 were determined to be species that were on  
232 the combined EPPO and van Lenteren 2012 list of BCAs. All remaining duplicates were  
233 removed manually (94) along with conference proceedings (3), resulting in a total of  
234 296 articles.

235 At this point, several articles were identified as problematic, either for availability or  
236 language. Four papers were identified as missing and were removed from the  
237 dataset. Eighteen articles were not in English. Most of these articles were in Chinese  
238 (9), followed by Portuguese (3), Russian (3), Czech (1), German (1), Persian (1), Spanish  
239 (1), and French (1). Where possible, articles were assessed by native speakers with a  
240 biological background to determine relevance, followed by non-native but fluent  
241 speakers with a biological background. All foreign language papers could be  
242 assessed with the exception of all three Portuguese papers and one Russian paper.  
243 These four unassessed papers were deemed unrelated, followed by an additional  
244 eight papers that were assessed. Of the 20 non-English papers, seven were assessed  
245 and kept for further screening. This final step reduced the articles in this dataset to 279,  
246 all of which are listed in the Appendix (Table S3).

## 247 3.3 Extraction

248 Of the 279 screened articles, only 69 were ultimately deemed relevant to this  
249 systematic review, based on the estimation methods and presence of measured traits,

250 and are listed in Table 1. Results from the extraction process, with reduced variable  
251 extractions for each extraction output, can be found in Table 2. The amount of  
252 observations depends on the species studied, the number of populations assessed,  
253 and the trait of interest. For example, one paper may look at a single trait in two  
254 species using isofemale lines, which would result in two observations. Within our results,  
255 there are 302 observations in total: 137 for parasitoids, and 165 for predators. Per  
256 paper, the average number of observations is 4.4, however, a small number of papers  
257 exceed 10 observations, and can therefore account for any skewing of popularity in  
258 estimation methods or species interest. For example, the most observations per  
259 species belong to *Harmonia axyridis* with 50 observations, while only being found in  
260 five papers. While the full dataset is informative for traits of interest, other parameters  
261 such as year of publication or estimation method will be better observed by reducing  
262 to unique observations.

263 Focusing on publication year, the spread of articles that reach the extraction phase  
264 is relatively well-distributed from 1979 to 2018 with no clear bias towards the present  
265 day (Table 1). Moving on to the BCAs themselves, there are 29 different species  
266 studied in this dataset, 13 of which only appeared once (Table 2). The most studied  
267 species in our dataset is the parasitoid *Aphidius ervi*, which was covered in eight  
268 articles. The parasitoid *Trichogramma* is the most studied genus of the extraction with  
269 12 articles, followed by *Aphidius*, with eight articles. Considering the genetic variation  
270 estimation methods (Figure 3), just over a quarter of observations used isofemale  
271 populations to estimate inheritance (27.1%), followed by artificial selection (17.1%).

272 Despite the use of these estimation methods, the majority of papers did not provide  
273 heritability estimates. We limited this to values of broad-sense heritability ( $H^2$ ) and  
274 narrow-sense heritability ( $h^2$ ), as well as evolvability ( $CV_A$ ). In some cases, these

275 estimates were not reported directly, but we were able to convert the reported  
276 genetic or dominance variance into these estimates using other data reported in the  
277 paper. Of our 302 observations, only 105 were supported by measures of genetic  
278 variation. When reduced to unique species per paper (72, as a few papers cover two  
279 species), 19 provided a heritability estimate for the trait assessed as opposed to the  
280 53 (73.6%) that did not (Figure 4). The most commonly used were either  $h^2$  (9.7%) or  $H^2$   
281 (8.3%), followed by either in combination with  $CV_A$  ( $h^2$  and  $CV_A$  at 4.2%, and  $H^2$  and  
282  $CV_A$  at 2.8%),  $CV_A$  on its own (1.4%), or both broad and narrow sense heritability at the  
283 same time (1.4%). These amounts include papers where we were able to calculate  
284 estimates using reported data in the text. Papers that reported on variations of  $CV_A$   
285 nearly always reported  $CV_G$ , but in the cases referenced here these were found to be  
286 referring to the same measure ( $CV_A$ ). Meanwhile, the majority of papers did not report  
287 any heritability estimates or include values necessary for their calculation.

288 An important goal of our analysis is the categorization of traits according to fitness  
289 component, or importance to biological control research. Given the variety of traits  
290 being measured, both in nomenclature and formulation, categories were assigned  
291 and pooled by observation count in two ways: the Mousseau and Roff assessment  
292 based on biological function used in their review of the heritability of fitness  
293 components, and the Leung et al. 2019 assessment based on biological control  
294 application (Mousseau and Roff 1987; Leung et al. 2019). The outcome of the  
295 Mousseau and Roff assessment indicates that the majority of the 302 observations  
296 were of life history traits (53%), followed by behavior (21%), morphology (19%), and  
297 physiology (7%; Figure 5). Also within the different BCA types, parasitoid or predator,  
298 life history is still the most accounted for trait for both types, while behavior and  
299 morphology switch middling positions depending on BCA type.

300 The second assessment method, based on biological control application (Leung et al  
301 2019), allowed for multiple classifications per observation (364 total). As shown in  
302 Figure 6, the majority of the traits measured were related to pest suppression ability  
303 (53%), followed by improving mass rearing conditions (23%). The third most common  
304 classification of the observations was the 'unrelated' category with 13% of the  
305 observations and the traits associated with them were most likely not useful for  
306 improving biological control directly.

307

## 308 4. Discussion

309 This systematic review sought to identify studies on BCAs that look at the genetic  
310 variation and heritability of traits and ordered these traits by two classification systems.  
311 According to the Mousseau and Roff classification, a majority of studies dealing with  
312 trait heritability focused on life history traits, regardless of BCA type. The other  
313 classifications of Mousseau and Roff (morphology, behavior, and physiology) were  
314 also approached in various heritability studies, but to a much lesser extent than those  
315 focusing on life history traits. This is most probably due to the importance of life history  
316 traits for biological control, which resulted in a large body of descriptive literature for  
317 BCAs (Mayhew, 2016). Furthermore, life history traits are generally considered to be an  
318 established measure of an organism's reproductive strategy and, ultimately, fitness.  
319 As such, they are considered important in the establishment and success of BCAs  
320 (Plouvier and Wajnberg, 2018). They are also wide ranging and can vary from obvious  
321 characteristics such as longevity and number of offspring, to more specific attributes  
322 such as conversion efficiency, certain of which are proven to be very useful in

323 particular applied settings. Of the papers in the final selection, 30.6% were found to  
324 include usable heritability estimates or the means to calculate them.

325 The functionality of traits in a biological control setting was further explored using the  
326 second classification. This classification gives a more targeted approach to labelling  
327 traits as to their potential utility in a biological control setting. The majority of traits  
328 analyzed within this set were attributed to 'pest suppression ability'. This attribute  
329 would include traits classed as life history traits in the Mousseau and Roff classification  
330 (i.e. number of offspring or sex ratio), but could also include certain behavioral traits  
331 (i.e. patch residence time). In terms of number of observations, the next group of traits  
332 are those 'useful in improving mass rearing conditions', which would include life history  
333 traits such as sex ratio. These are potentially two of the more important classes within  
334 the classification for biological control application, indicating that the traits outlined  
335 in Leung et al. are in line with the current trends in biological control research (Leung  
336 et al., 2019). It is also important to note that certain traits could fit into several of the  
337 biological control trait classifications, whereas the Mousseau and Roff classifications  
338 are clear-cut. In this way, the classification for biological control application could be  
339 deemed more relevant or concise when narrowing down traits for their importance in  
340 biological control, and by doing so in such a study as ours, pinpoints those that are  
341 also heritable. Our results further show that traits related to 'adaptation to biotic  
342 factors,' and 'reducing ecological risk' are relatively understudied compared to the  
343 others, which could be seen as an incentive for more studies.

344 The lack of data on genetic variation in our results does not necessarily represent a  
345 lack of heritability studies - indeed in our original search we found 69 papers  
346 concerning heritability of life history traits in BCAs with appropriate estimation  
347 methods. Of these, however, only 19 papers contained heritability values, either in the

348 form of broad-sense heritability, narrow-sense heritability, evolvability, or the genetic  
349 and environmental variance estimates required for calculation. As for estimation  
350 methods, nearly 9 out of 10 papers were removed from the analysis during our  
351 screening due to opaque reporting of methods and results, yet the papers still  
352 referenced heritability or genetic variation. This opacity has been documented  
353 previously, such as in a recent review on the use of experimental evolution to improve  
354 biological control, where the majority of studies found did not carry out replicates or  
355 maintain control populations (Lirakis and Magalhães, 2019). Discrepancies such as  
356 these are indications of the flexibility of terms and experimental design between  
357 research disciplines; the full reporting of values is simply a recommendation for  
358 improving research on genetic variation that has been made before (see Houle,  
359 1992). Reporting sample size is necessary for variance calculations, whereas  
360 calculating heritability values is encouraged in order to add quantitative data to a  
361 discussion on heritability. With this in mind, it became clear during the extraction of  
362 data that further analysis, such as a meta-analysis of the heritability results, would be  
363 intractable. This is due to the wide spread of species and traits of interests juxtaposed  
364 to a lack of heritability values and other values necessary for such an analysis. This lack  
365 of reporting could be due to oversight or publication restrictions (such as limitation on  
366 manuscript length or perceived triviality of some data), but another possibility is that  
367 the perceived importance of reporting these values is not high. We hope that recent  
368 calls on the importance of genetic variation for improving BCAs, such as Leung *et al.*  
369 (2019), and the need for these values will shift towards studies publishing more basic  
370 information and additional data in the future.

371 Finally, there are fairly similar number of papers on parasitoids and predators, both  
372 before and after extraction. While initially there looked to be a slight bias towards  
373 parasitoids, there is a larger bias in the registered and published BCA types in use, with

374 far more parasitoids than predators (van Lenteren, 2012), making this more-or-less  
375 equal outcome more surprising.

376 This systematic review was carried out across three different reference databases with  
377 an intentional search term that was tested against positive and negative controls.  
378 Once the search results returned, assessment of the study for its inclusion was based  
379 on both objective and subjective measures. In this respect, certain papers that would  
380 fit the desired outcome of the search may have been absent due to database or  
381 search term or dismissed prematurely due to human error. Taxonomic changes could  
382 also play a part in missing papers that would otherwise be suitable for this analysis.  
383 Finally, refining the species studied to the combined list of the 2016 and 2019 EPPO lists  
384 of commercial BCAs (PM 6/003) as well as using a commercial list from 2012 that was  
385 updated in 2018 (van Lenteren, 2012; van Lenteren et al., 2018) may remove any  
386 species that are currently being investigated as a potential BCA.

## 387 5. Conclusion

388 The results of this systematic review indicate a variety of research and interest in  
389 studying the heritability of traits in BCAs and can act as a starting point for improving  
390 natural enemies. The presence of genetic variation for traits related to biological  
391 control supports the calls for improvement of biological control agents via natural  
392 genetic variation made previously (Kruitwagen et al., 2018; Leung et al., 2019; Lirakis  
393 and Magalhães, 2019; Lommen et al., 2017). Indeed, a large variety of groundwork  
394 has been established in these fields related to species, estimation methods, and traits  
395 of interest. However, studies that report on genetic variation cannot necessarily add  
396 to the advancement of biological control research if heritability estimates and  
397 associated values are not reported. With this in mind, this aspect of biological control



398 research is ripe with opportunities, and several of the studies we have profiled here  
399 are an excellent starting point for the genetic improvement of the efficacy of  
400 biological control agents.

401

## 402 CRediT Author Statement

403 **Kim Ferguson:** Methodology, Investigation, Visualisation, Writing - Original Draft

404 **Sophie Chattington:** Methodology, Resources, Investigation, Writing - Original Draft

405 **Wouter Plouvier:** Methodology, Investigation, Data Curation, Writing - Review & Editing

406 **Bart Pannebakker:** Conceptualisation, Supervision, Writing - Review & Editing

407

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## 418 Declaration of interest

419 The authors declare there is no conflict of interest.

## 420 Appendix

421 Supplementary material indicated in this manuscript (Tables S1, S2, and S3) are

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## 423 References

- 424 Abbas, N., Mansoor, M.M., Shad, S.A., Pathan, A.K., Waheed, A., Ejaz, M., Razaq, M.,  
425 Zulfiqar, M.A., 2014. Fitness cost and realized heritability of resistance to spinosad  
426 in *Chrysoperla carnea* (Neuroptera: Chrysopidae). *Bull. Entomol. Res.* 104, 707–  
427 715. <https://doi.org/10.1017/S0007485314000522>
- 428 Aguilar-Fenollosa, E., Jacas, J.A., 2014. Can we forecast the effects of climate  
429 change on entomophagous biological control agents? *Pest Manag. Sci.* 70,  
430 853–859. <https://doi.org/10.1002/ps.3678>
- 431 Al-Khateeb, N., Asslan, L., Ibrahim, G., El-Heneidy, A., 2018. A selection index of  
432 multiple quantitative traits of the predator *Cryptolaemus montrouzieri* (Mulsant)  
433 (Coleoptera: Coccinellidae) through 12 inbreeding generations. *Egypt. J. Biol.*  
434 *Pest Control* 28. <https://doi.org/10.1186/s41938-018-0049-9>
- 435 Antolin, M.F., 1992. Sex Ratio Variation in a Parasitic Wasp II. Diallel Cross. *Evolution*  
436 (N. Y). 46, 1511. <https://doi.org/10.2307/2409954>
- 437 Astles, P.A., Moore, A.J., Preziosi, R.F., 2005. Genetic variation in response to an  
438 indirect ecological effect. *Proc. R. Soc. B Biol. Sci.* 272, 2577–2581.  
439 <https://doi.org/10.1098/rspb.2005.3174>
- 440 Barratt, B.I.P., Moran, V.C., Bigler, F., van Lenteren, J.C., 2018. The status of biological  
441 control and recommendations for improving uptake for the future. *BioControl*  
442 63, 155–167. <https://doi.org/10.1007/s10526-017-9831-y>
- 443 Bennett, D.M., Hoffmann, A.A., 1998. Effects of size and fluctuating asymmetry on  
444 field fitness of the parasitoid *Trichogramma carverae* (Hymenoptera:  
445 Trichogrammatidae). *J. Anim. Ecol.* 67, 580–591. <https://doi.org/10.1046/j.1365-2656.1998.00218.x>
- 447 Bilodeau, E., Simon, J.C., Guay, J.F., Turgeon, J., Cloutier, C., 2013. Does variation in  
448 host plant association and symbiont infection of pea aphid populations induce  
449 genetic and behaviour differentiation of its main parasitoid, *Aphidius ervi*? *Evol.*  
450 *Ecol.* 27, 165–184. <https://doi.org/10.1007/s10682-012-9577-z>
- 451 Boulétreau, M., Wajnberg, E., 1986. Comparative responses of two sympatric  
452 parasitoid cynipids to the genetic and epigenetic variations of the larvae of  
453 their host, *Drosophila melanogaster*. *Entomol. Exp. Appl.* 41, 107–114.  
454 <https://doi.org/10.1111/j.1570-7458.1986.tb00516.x>
- 455 Brotherstone, S., Goddard, M., 2005. Artificial selection and maintenance of genetic  
456 variance in the global dairy cow population. *Philos. Trans. R. Soc. B Biol. Sci.* 360,  
457 1479–1488. <https://doi.org/10.1098/rstb.2005.1668>
- 458 Bruins, E.B.A.W., Wajnberg, E., Pak, G.A., 1994. Genetic variability in the reactive  
459 distance in *Trichogramma brassicae* after automatic tracking of the walking  
460 path. *Entomol. Exp. Appl.* 72, 297–303. <https://doi.org/10.1111/j.1570-7458.1994.tb01830.x>
- 462 CBD Secretariat, 2011. About the Nagoya Protocol [WWW Document]. URL  
463 <https://www.cbd.int/abs/about/> (accessed 11.14.19).
- 464 Côté, I.M., Jennions, M.D., 2013. The procedure of meta-analysis in a nutshell, in:  
465 Koricheva, J., Gurevitch, J., Mengersen, K. (Eds.), *Handbook of Meta-Analysis in*

- 466 Ecology and Evolution. Princeton University Press, Oxford, pp. 14–24.  
467 <https://doi.org/10.1515/9781400846184-004>
- 468 Das, D.K., Singh, J., Vennila, S., 2011. Emerging Crop Pest Scenario under the Impact  
469 of Climate Change – A Brief Review. *J. Agric. Phys.* 11, 13–20.  
470 <https://doi.org/ISSN 0973-032X>
- 471 El-Heneidy, A.H., Shoeb, M.A., 2007. Comparative biological aspects of two strains  
472 from the egg parasitoid, *Trichogramma evanescens* Westwood (Hymenoptera:  
473 Trichogrammatidae) in Egypt. *Egypt. J. Biol. Pest Control* 17, 99–106.
- 474 Facon, B., Crespin, L., Loiseau, A., Lombaert, E., Magro, A., Estoup, A., 2011. Can  
475 things get worse when an invasive species hybridizes? The harlequin ladybird  
476 *Harmonia axyridis* in France as a case study. *Evol. Appl.* 4, 71–88.  
477 <https://doi.org/10.1111/j.1752-4571.2010.00134.x>
- 478 Fleury, F., Allemand, R., Fouillet, P., Boulétreau, M., 1995. Genetic variation in  
479 locomotor activity rhythm among populations of *Leptopilina heterotoma*  
480 (Hymenoptera: Eucoilidae), a larval parasitoid of *Drosophila* species. *Behav.*  
481 *Genet.* 25, 81–89. <https://doi.org/10.1007/BF02197245>
- 482 Fukunaga, Y., Akimoto, S.I., 2007. Toxicity of the aphid *Aulacorthum magnoliae* to  
483 the predator *Harmonia axyridis* (Coleoptera: Coccinellidae) and genetic  
484 variance in the assimilation of the toxic aphids in *H. axyridis* larvae: Original  
485 article. *Entomol. Sci.* 10, 45–53. <https://doi.org/10.1111/j.1479-8298.2006.00197.x>
- 486 Garcia-Gonzalez, F., Simmons, L.W., Tomkins, J.L., Kotiaho, J.S., Evans, J.P., 2012.  
487 Comparing evolvabilities: Common errors surrounding the calculation and use  
488 of coefficients of additive genetic variation. *Evolution* (N. Y). 66, 2341–2349.  
489 <https://doi.org/10.1111/j.1558-5646.2011.01565.x>
- 490 Geden, C.J., Smith, L., Long, S.J., Rutz, D.A., 1992. Rapid Deterioration of Searching  
491 Behavior, Host Destruction, and Fecundity of the Parasitoid *Muscidifurax raptor*  
492 (Hymenoptera: Pteromalidae) in Culture. *Ann. Entomol. Soc. Am.* 85, 179–187.  
493 <https://doi.org/10.1093/aesa/85.2.179>
- 494 Gilchrist, G.W., 1996. A Quantitative Genetic Analysis of Thermal Sensitivity in the  
495 Locomotor Performance Curve of *Aphidius ervi*. *Evolution* (N. Y). 50, 1560.  
496 <https://doi.org/10.2307/2410892>
- 497 Grill, C.P., Moore, A.J., Brodie, E.D., 1997. The genetics of phenotypic plasticity in a  
498 colonizing population of the ladybird beetle, *Harmonia axyridis*. *Heredity*  
499 (Edinb). 78, 261–269. <https://doi.org/10.1038/sj.hdy.6881030>
- 500 Guzmán-Larralde, A., Cerna-Chávez, E., Rodríguez-Campos, E., Loyola-Licea, J.C.,  
501 Stouthamer, R., 2014. Genetic variation and the performance of a mass-reared  
502 parasitoid, *Trichogramma pretiosum* (Hymenoptera: Trichogrammatidae), in  
503 laboratory trials. *J. Appl. Entomol.* 138, 346–354.  
504 <https://doi.org/10.1111/jen.12073>
- 505 Hansen, T.F., Pélabon, C., Houle, D., 2011. Heritability is not Evolvability. *Evol. Biol.* 38,  
506 258–277. <https://doi.org/10.1007/s11692-011-9127-6>
- 507 Henry, L.M., May, N., Acheampong, S., Gillespie, D.R., Roitberg, B.D., 2010. Host-  
508 adapted parasitoids in biological control: Does source matter? *Ecol. Appl.* 20,  
509 242–250. <https://doi.org/10.1890/08-1869.1>

- 510 Henter, H.J., 1995. The Potential for Coevolution in a Host-Parasitoid System. II.  
511 Genetic Variation within a Population of Wasps in the Ability to Parasitize an  
512 Aphid Host. *Evolution* (N. Y). 49, 439. <https://doi.org/10.2307/2410268>
- 513 Henter, H.J., Brasch, K., Lenteren, J.C., 1996. Variation between laboratory  
514 populations of *Encarsia formosa* in their parasitization behavior on the host  
515 *Bemisia tabaci*. *Entomol. Exp. Appl.* 80, 435–441. <https://doi.org/10.1111/j.1570-7458.1996.tb00957.x>
- 517 Holloway, G.J., Brakefield, P.M., de Jong, P., Ottenheim, M.M., De Vos, H., Kesbeke,  
518 F., Peynenburg, L., 1995. A quantitative genetic analysis of an aposematic  
519 colour pattern and its ecological implications. *Philos. Trans. R. Soc. B Biol. Sci.*  
520 348, 373–379. <https://doi.org/10.1098/rstb.1995.0075>
- 521 Holloway, G.J., de Jong, P.W., Ottenheim, M., 1993. The Genetics and Cost of  
522 Chemical Defense in the Two-Spot Ladybird (*Adalia bipunctata* L.). *Evolution* (N.  
523 Y). 47, 1229. <https://doi.org/10.2307/2409988>
- 524 Houle, D., 1992. Comparing evolvability and variability of quantitative traits. *Genetics*  
525 130, 195–204.
- 526 Hufbauer, R.A., 2001. Pea aphid-parasitoid interactions: Have parasitoids adapted to  
527 differential resistance? *Ecology* 82, 717–725. [https://doi.org/10.1890/0012-9658\(2001\)082\[0717:PAPIHP\]2.0.CO;2](https://doi.org/10.1890/0012-9658(2001)082[0717:PAPIHP]2.0.CO;2)
- 529 Hufbauer, R.A., Via, S., 1999. Evolution of an Aphid-Parasitoid Interaction: Variation in  
530 Resistance to Parasitism among Aphid Populations Specialized on Different  
531 Plants. *Evolution* (N. Y). 53, 1435. <https://doi.org/10.2307/2640890>
- 532 IPCC, 2007. *Climate Change 2007: The Physical Science Basis. Contribution of*  
533 *Working Group I to the Fourth Assessment Report of the Intergovernmental*  
534 *Panel on Climate Change, Climate Change 2013 - The Physical Science Basis.*  
535 *Cambridge University Press, Cambridge.*  
536 <https://doi.org/10.1017/CBO9781107415324.015>
- 537 Jia, F., Margolies, D.C., Boyer, J.E., Charlton, R.E., 2002. Genetic variation in foraging  
538 traits among inbred lines of a predatory mite. *Heredity* (Edinb). 89, 371–379.  
539 <https://doi.org/10.1038/sj.hdy.6800145>
- 540 Kamala Jayanthi, P.D., Sangeetha, P., Verghese, A., 2014. Study of inheritance of  
541 feeding potential in natural populations of predatory coccinellid *Cryptolaemus*  
542 *montrouzieri* Mulsant using isofemale strains. *J. Genet.* 93, 113–122.  
543 <https://doi.org/10.1007/s12041-014-0350-7>
- 544 Kamping, A., Katju, V., Beukeboom, L.W., Werren, J.H., 2007. Inheritance of  
545 gynandromorphism in the parasitic wasp *Nasonia vitripennis*. *Genetics* 175,  
546 1321–1333. <https://doi.org/10.1534/genetics.106.067082>
- 547 Karabörklü, S., Azizoglu, U., Azizoglu, Z.B., 2018. Recombinant entomopathogenic  
548 agents: a review of biotechnological approaches to pest insect control. *World*  
549 *J. Microbiol. Biotechnol.* 34, 14. <https://doi.org/10.1007/s11274-017-2397-0>
- 550 Koricheva, J., Gurevitch, J., Mengersen, K., 2013. *Handbook of meta-analysis in*  
551 *ecology and evolution, Handbook of Meta-analysis in Ecology and Evolution.*  
552 *Princeton University Press, Oxford.*  
553 <https://doi.org/10.23943/princeton/9780691137285.001.0001>

- 554 Kruitwagen, A., Beukeboom, L.W., Wertheim, B., 2018. Optimization of native  
555 biocontrol agents, with parasitoids of the invasive pest *Drosophila suzukii* as an  
556 example. *Evol. Appl.* 11, 1473–1497. <https://doi.org/10.1111/eva.12648>
- 557 Legner, E.F., 1993. Theory for quantitative inheritance of behavior in a protelean  
558 parasitoid, *Muscidifurax raptorellus* (Hymenoptera: Pteromalidae). *Eur. J.*  
559 *Entomol.* 90, 11–21.
- 560 Legner, E.F., 1991a. Estimations of number of active loci, dominance and heritability  
561 in polygenic inheritance of gregarious behavior in *Muscidifurax raptorellus*  
562 [Hymenoptera: Pteromalidae]. *Entomophaga* 36, 1–18.  
563 <https://doi.org/10.1007/BF02374633>
- 564 Legner, E.F., 1991b. Recombinant males in the parasitic wasp *Muscidifurax*  
565 *raptorellus* [Hymenoptera: pteromalidae]. *Entomophaga* 36, 173–181.  
566 <https://doi.org/10.1007/BF02374553>
- 567 Legner, E.F., 1988. *Muscidifurax raptorellus* (Hymenoptera: Pteromalidae) Females  
568 Exhibit Postmating Oviposition Behavior Typical of the Male Genome. *Ann.*  
569 *Entomol. Soc. Am.* 81, 522–527. <https://doi.org/10.1093/aesa/81.3.522>
- 570 Legner, E.F., 1987. Inheritance of gregarious and solitary oviposition in *Muscidifurax*  
571 *raptorellus kogan* and *legner* (Hymenoptera: Pteromalidae). *Can. Entomol.* 119,  
572 791–808. <https://doi.org/10.4039/Ent119791-9>
- 573 Lesna, I., Sabelis, M.W., 1999. Diet-dependent female choice for males with 'good  
574 genes' in a soil predatory mite. *Nature* 401, 581–584.  
575 <https://doi.org/10.1038/44125>
- 576 Leung, K., Ras, E., Ferguson, K.B., Ariëns, S., Babendreier, D.B., Bijma, P., Bourtzis, K.,  
577 Brodeur, J., Bruins, M., Centurión, A., Chattington, S., Chinchilla-Ramírez, M.,  
578 Dicke, M., Fatouros, N., González Cabrera, J., Groot, T., Haye, T., Knapp, M.,  
579 Koskinioti, P., Le Hesran, S., Lirakis, M., Paspatis, A., Pérez-Hedo, M., Plouvier, W.,  
580 Schlötterer, C., Stahl, J., Thiel, A., Urbaneja, A., van de Zande, L., Verhulst, E., Vet,  
581 L., Visser, S., Werren, J., Xia, S., Zwaan, B., Magalhães, S., Beukeboom, L.,  
582 Pannebakker, B., 2019. Next Generation Biological Control: the Need for  
583 Integrating Genetics and Evolution. *Preprints* 1–34.  
584 <https://doi.org/10.20944/preprints201911.0300.v1>
- 585 Lirakis, M., Magalhães, S., 2019. Does experimental evolution produce better  
586 biological control agents? A critical review of the evidence. *Entomol. Exp. Appl.*  
587 584–597. <https://doi.org/10.1111/eea.12815>
- 588 Liu, F.H., Smith, S.M., 2000. Measurement and selection of parasitoid quality for mass-  
589 reared *Trichogramma minutum riley* used in inundative release. *Biocontrol Sci.*  
590 *Technol.* 10, 3–13. <https://doi.org/10.1080/09583150029332>
- 591 Lommen, S.T.E., de Jong, P.W., Pannebakker, B.A., 2017. It is time to bridge the gap  
592 between exploring and exploiting: prospects for utilizing intraspecific genetic  
593 variation to optimize arthropods for augmentative pest control – a review.  
594 *Entomol. Exp. Appl.* 162, 108–123. <https://doi.org/10.1111/eea.12510>
- 595 Lovett, B., St. Leger, R.J., 2018. Genetically engineering better fungal biopesticides.  
596 *Pest Manag. Sci.* 74, 781–789. <https://doi.org/10.1002/ps.4734>
- 597 Maeda, T., Takabayashi, J., Yano, S., Takafuji, A., 1999. Response of the predatory

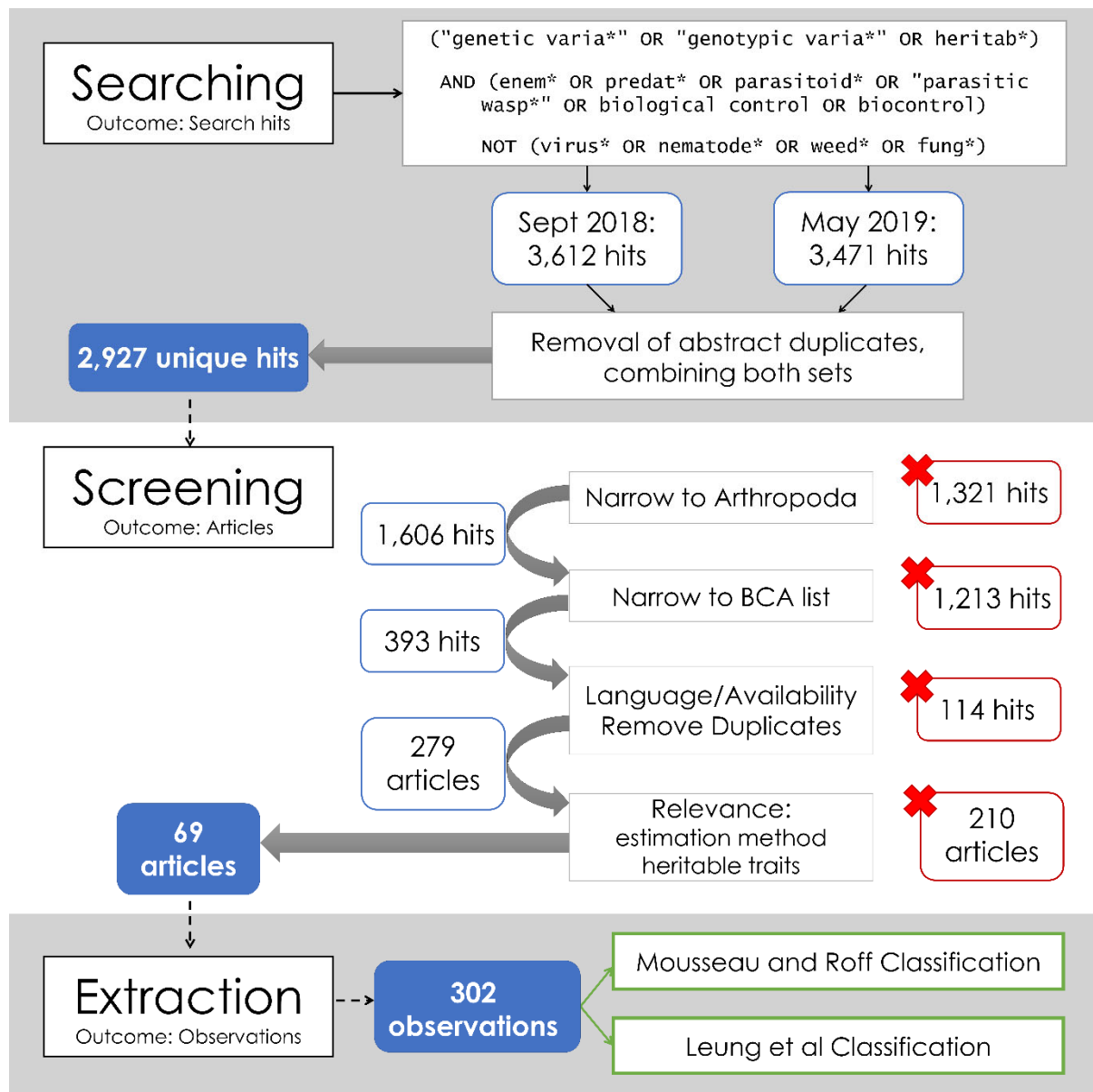
- 598 mite, *Amblyseius womersleyi* (Acari: Phytoseiidae), toward herbivore-induced  
599 plant volatiles: Variation in response between two local populations. *Appl.*  
600 *Entomol. Zool.* 34, 449–454. <https://doi.org/10.1303/aez.34.449>
- 601 Mansoor, M.M., Abbas, N., Shad, S.A., Pathan, A.K., Razaq, M., 2013. Increased  
602 fitness and realized heritability in emamectin benzoate-resistant *Chrysoperla*  
603 *carnea* (Neuroptera: Chrysopidae). *Ecotoxicology* 22, 1232–1240.  
604 <https://doi.org/10.1007/s10646-013-1111-8>
- 605 Mansoor, M.M., Raza, A.B.M., Abbas, N., Aqueel, M.A., Afzal, M., 2017. Resistance of  
606 green lacewing, *Chrysoperla carnea* Stephens to nitenpyram: Cross-resistance  
607 patterns, mechanism, stability, and realized heritability. *Pestic. Biochem. Physiol.*  
608 135, 59–63. <https://doi.org/10.1016/j.pestbp.2016.06.004>
- 609 Margolies, D.C., Sabelis, M.W., Boyer, J.E., 1997. Response of a phytoseiid predator to  
610 herbivore-induced plant volatiles: Selection on attraction and effect on prey  
611 exploitation. *J. Insect Behav.* 10, 695–709. <https://doi.org/10.1007/BF02765387>
- 612 Marples, N.M., de Jong, P.W., Ottenheim, M.M., Verhoog, M.D., Brakefield, P.M.,  
613 1993. The inheritance of a wingless character in the 2spot ladybird (*Adalia*  
614 *bipunctata*). *Entomol. Exp. Appl.* 69, 69–73. <https://doi.org/10.1111/j.1570-7458.1993.tb01729.x>
- 616 Mason, P.G., Hopper, K.R., 1997. Temperature Dependence in Locomotion of the  
617 Parasitoid *Aphelinus asychis* (Hymenoptera: Aphelinidae) from Geographical  
618 Regions with Different Climates. *Environ. Entomol.* 26, 1416–1423.  
619 <https://doi.org/10.1093/ee/26.6.1416>
- 620 Mayhew, P.J., 2016. Comparing parasitoid life histories. *Entomol. Exp. Appl.* 159, 147–  
621 162. <https://doi.org/10.1111/eea.12411>
- 622 Misra, A.K., 2014. Climate change and challenges of water and food security. *Int. J.*  
623 *Sustain. Built Environ.* 3, 153–165. <https://doi.org/10.1016/j.ijsbe.2014.04.006>
- 624 Mousseau, T.A., Roff, D.A., 1987. Natural selection and the heritability of fitness  
625 components. *Heredity (Edinb)*. 59, 181–197. <https://doi.org/10.1038/hdy.1987.113>
- 626 Nachappa, P., Margolies, D.C., Nechols, J.R., Morgan, T.J., 2010. Response of a  
627 complex foraging phenotype to artificial selection on its component traits. *Evol.*  
628 *Ecol.* 24, 631–655. <https://doi.org/10.1007/s10682-009-9318-0>
- 629 Navia, D., Domingos, C.A., Mendonça, R.S., Ferragut, F., Rodrigues, M.A.N., de  
630 Morais, E.G.F., Tixier, M.S., Gondim, M.G.C., 2014. Reproductive compatibility  
631 and genetic and morphometric variability among populations of the predatory  
632 mite, *Amblyseius largoensis* (Acari: Phytoseiidae), from Indian Ocean Islands  
633 and the Americas. *Biol. Control* 72, 17–29.  
634 <https://doi.org/10.1016/j.biocontrol.2014.01.011>
- 635 Obrycki, J.J., Krawfur, E.S., Bogran, C.E., Gomez, L.E., Cave, R.E., 2001. Comparative  
636 Studies of Three Populations of the Lady Beetle Predator *Hippodamia*  
637 *convergens* (Coleoptera: Coccinellidae). *Florida Entomol.* 84, 55.  
638 <https://doi.org/10.2307/3496663>
- 639 Orzack, S.H., Gladstone, J., 1994. Quantitative genetics of sex ratio traits in the  
640 parasitic wasp, *Nasonia vitripennis*. *Genetics* 137, 211–220.
- 641 Orzack, S.H., Parker, E.D., Gladstone, J., 1991. The comparative biology of genetic

- 642 variation for conditional sex ratio behavior in a parasitic wasp, *Nasonia*  
643 *vitripennis*. *Genetics* 127, 583–599.
- 644 Pannebakker, B.A., Halligan, D.L., Reynolds, K.T., Ballantyne, G.A., Shuker, D.M.,  
645 Barton, N.H., West, S.A., 2008. Effects of spontaneous mutation accumulation on  
646 sex ratio traits in a parasitoid wasp. *Evolution* (N. Y). 62, 1921–1935.  
647 <https://doi.org/10.1111/j.1558-5646.2008.00434.x>
- 648 Paolucci, S., Van de Zande, L., Beukeboom, L.W., 2013. Adaptive latitudinal cline of  
649 photoperiodic diapause induction in the parasitoid *Nasonia vitripennis* in  
650 Europe. *J. Evol. Biol.* 26, 705–718. <https://doi.org/10.1111/jeb.12113>
- 651 Parker, E.D., Orzack, S.H., 1985. Genetic variation for the sex ratio in *Nasonia*  
652 *vitripennis*. *Genetics* 110, 93–105.
- 653 Parreño, M.A., Ivanovic, A., Petrovic, A., Zikic, V., Tomanovic, Z., Vorburger, C., 2017.  
654 Wing shape as a taxonomic trait: Separating genetic variation from host-  
655 induced plasticity in aphid parasitoids. *Zool. J. Linn. Soc.* 180, 288–297.  
656 <https://doi.org/10.1111/zoj.12490>
- 657 Paspati, A., Ferguson, K.B., Verhulst, E.C., Urbaneja, A., González-Cabrera, J.,  
658 Pannebakker, B.A., 2019. Effect of mass rearing on the genetic diversity of the  
659 predatory mite *Amblyseius swirskii* Athias-Henriot (Acari: Phytoseiidae). *Entomol.*  
660 *Exp. Appl.* 167, 670–681. <https://doi.org/10.1111/eea.12811>
- 661 Pérez-Maluf, R., Rafalimanana, H., Campan, E., Fleury, F., Kaiser, L., 2008.  
662 Differentiation of innate but not learnt responses to host-habitat odours  
663 contributes to rapid host finding in a parasitoid genotype. *Physiol. Entomol.* 33,  
664 226–232. <https://doi.org/10.1111/j.1365-3032.2008.00636.x>
- 665 Plouvier, W.N., Wajnberg, E., 2018. Improving the efficiency of augmentative  
666 biological control with arthropod natural enemies: A modeling approach. *Biol.*  
667 *Control* 125, 121–130. <https://doi.org/10.1016/j.biocontrol.2018.05.010>
- 668 Pompanon, F., Fouillet, P., Bouletreau, M., 1994. Locomotor behaviour in females of  
669 two *Trichogramma* species: description and genetic variability. *Nor. J. Agric. Sci.*  
670 185–190.
- 671 Poppy, G.M., Powell, W., 2009. Genetic manipulation of natural enemies: can we  
672 improve biological control by manipulating the parasitoid and/or the plant?, in:  
673 *Genetics, Evolution and Biological Control*. CABI, Wallingford, pp. 219–233.  
674 <https://doi.org/10.1079/9780851997353.0219>
- 675 R Core Team, 2015. R Development Core Team. *R A Lang. Environ. Stat. Comput.* 55,  
676 275–286.
- 677 Rasmussen, L.B., Jensen, K., Sørensen, J.G., Sverrisdóttir, E., Nielsen, K.L., Overgaard,  
678 J., Holmstrup, M., Kristensen, T.N., 2018. Are commercial stocks of biological  
679 control agents genetically depauperate? – A case study on the pirate bug  
680 *Orius majusculus* Reuter. *Biol. Control* 127, 31–38.  
681 <https://doi.org/10.1016/j.biocontrol.2018.08.016>
- 682 Reeves, R.G., Denton, J.A., Santucci, F., Bryk, J., Reed, F.A., 2012. Scientific standards  
683 and the regulation of genetically modified insects. *PLoS Negl. Trop. Dis.* 6.  
684 <https://doi.org/10.1371/journal.pntd.0001502>
- 685 Rigby, D., Cáceres, D., 2001. Organic farming and the sustainability of agricultural



- 686 systems. *Agric. Syst.* 68, 21–40. [https://doi.org/10.1016/S0308-521X\(00\)00060-3](https://doi.org/10.1016/S0308-521X(00)00060-3)
- 687 Rodríguez, J.R., Pintureau, B., Galán, M., 1994. Déterminisme de la couleur des hôtes  
688 parasités par *Trichogramma fuentesi*. *Entomol. Exp. Appl.* 70, 121–128.  
689 <https://doi.org/10.1111/j.1570-7458.1994.tb00740.x>
- 690 Routray, S., Dey, D., Baral, S., Das, A.P., Mahantheshwara, B., 2016. Genetic  
691 improvement of natural enemies: A review. *Agric. Rev.* 37.  
692 <https://doi.org/10.18805/ag.v37i4.6463>
- 693 RStudio Team, 2015. RStudio: Integrated Development for R.
- 694 Sandrock, C., Gouskov, A., Vorburger, C., 2010. Ample genetic variation but no  
695 evidence for genotype specificity in an all-parthenogenetic host-parasitoid  
696 interaction. *J. Evol. Biol.* 23, 578–585. <https://doi.org/10.1111/j.1420->  
697 [9101.2009.01925.x](https://doi.org/10.1111/j.1420-9101.2009.01925.x)
- 698 Sayyed, A.H., Pathan, A.K., Faheem, U., 2010. Cross-resistance, genetics and stability  
699 of resistance to deltamethrin in a population of *Chrysoperla carnea* from  
700 Multan, Pakistan. *Pestic. Biochem. Physiol.* 98, 325–332.  
701 <https://doi.org/10.1016/j.pestbp.2010.07.004>
- 702 Sepúlveda, D.A., Zepeda-Paulo, F., Ramírez, C.C., Lavandero, B., Figueroa, C.C.,  
703 2017. Loss of host fidelity in highly inbred populations of the parasitoid wasp  
704 *Aphidius ervi* (Hymenoptera: Braconidae). *J. Pest Sci.* (2004). 90, 649–658.  
705 <https://doi.org/10.1007/s10340-016-0798-8>
- 706 Sequeira, R., Mackauer, M., 1992. Quantitative genetics of body size and  
707 development time in the parasitoid wasp *Aphidius ervi* (Hymenoptera:  
708 Aphidiidae). *Can. J. Zool.* 70, 1102–1108. <https://doi.org/10.1139/z92-154>
- 709 Shuker, D.M., Phillimore, A.J., Burton-Chellew, M.N., Hodge, S.E., West, S.A., 2007. The  
710 quantitative genetic basis of polyandry in the parasitoid wasp, *Nasonia*  
711 *vitripennis*. *Heredity* (Edinb). 98, 69–73. <https://doi.org/10.1038/sj.hdy.6800897>
- 712 Sorati, M., Newman, M., Hoffmann, A.A., 1996. Inbreeding and incompatibility in  
713 *Trichogramma nr. brassicae*: Evidence and implications for quality control.  
714 *Entomol. Exp. Appl.* 78, 283–290. <https://doi.org/10.1111/j.1570->  
715 [7458.1996.tb00792.x](https://doi.org/10.1111/j.1570-7458.1996.tb00792.x)
- 716 St. Leger, R.J., Wang, C., 2010. Genetic engineering of fungal biocontrol agents to  
717 achieve greater efficacy against insect pests. *Appl. Microbiol. Biotechnol.* 85,  
718 901–907. <https://doi.org/10.1007/s00253-009-2306-z>
- 719 Szűcs, M., Vercken, E., Bitume, E. V., Hufbauer, R.A., 2019. The implications of rapid  
720 eco-evolutionary processes for biological control - a review. *Entomol. Exp. Appl.*  
721 *eea.12807*. <https://doi.org/10.1111/eea.12807>
- 722 Tauber, C.A., Tauber, M.J., 1992. Phenotypic Plasticity in *Chrysoperla*: Genetic  
723 Variation in the Sensory Mechanism and in Correlated Reproductive Traits.  
724 *Evolution* (N. Y). 46, 1754. <https://doi.org/10.2307/2410029>
- 725 Tauber, C.A., Tauber, M.J., 1986. Genetic variation in all-or-none life-history traits of  
726 the lacewing *Chrysoperla carnea*. *Can. J. Zool.* 64, 1542–1544.  
727 <https://doi.org/10.1139/z86-229>
- 728 Thomson, L.J., Hoffmann, A.A., 2009. Laboratory Fecundity as Predictor of Field

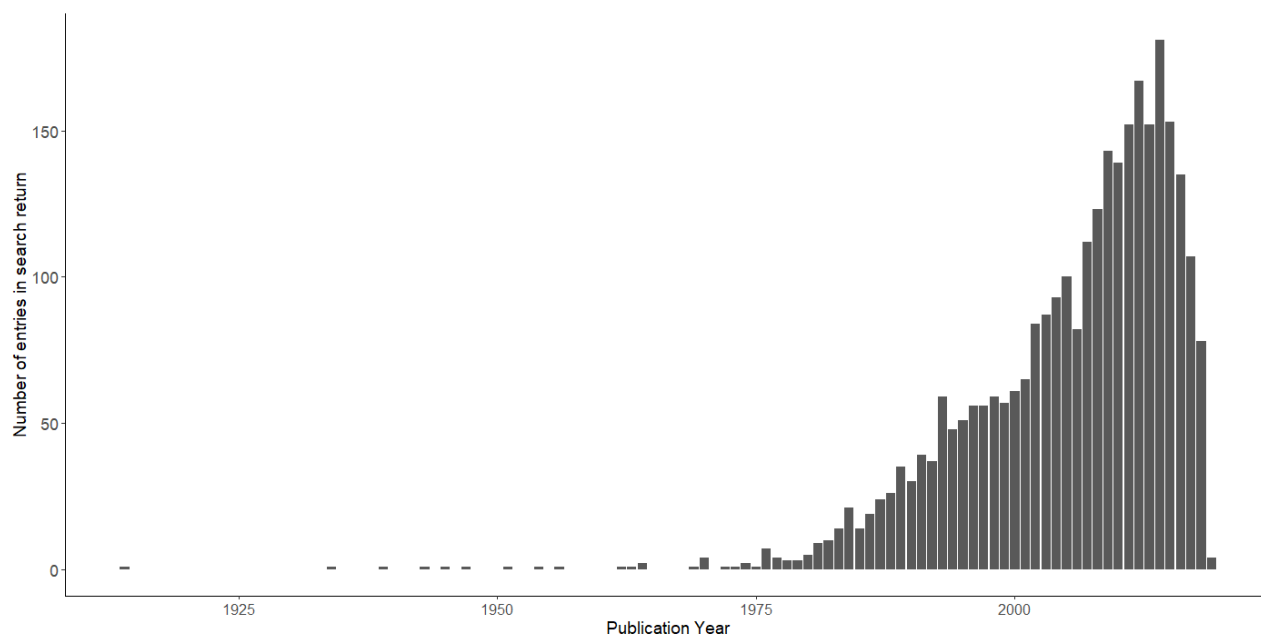
- 729 Success in *Trichogramma carverae* (Hymenoptera: Trichogrammatidae).  
730 J. Econ. Entomol. 95, 912–917. <https://doi.org/10.1603/0022-0493-95.5.912>
- 731 van Houten, Y.M., van Stratum, P., Bruin, J., Veerman, A., 1995. Selection for non-  
732 diapause in *Amblyseius cucumeris* and *Amblyseius barkeri* and exploration of  
733 the effectiveness of selected strains for thrips control. Entomol. Exp. Appl. 77,  
734 289–295. <https://doi.org/10.1111/j.1570-7458.1995.tb02326.x>
- 735 van Lenteren, J.C., 2012. The state of commercial augmentative biological control:  
736 plenty of natural enemies, but a frustrating lack of uptake. BioControl 57, 1–20.  
737 <https://doi.org/10.1007/s10526-011-9395-1>
- 738 van Lenteren, J.C., Bale, J., Bigler, F., Hokkanen, H.M.T., Loomans, A.J.M., 2006.  
739 Assessing Risks of Releasing Exotic Biological Control Agents of Arthropod Pests.  
740 Annu. Rev. Entomol. 51, 609–634.  
741 <https://doi.org/10.1146/annurev.ento.51.110104.151129>
- 742 van Lenteren, J.C., Bolckmans, K., Köhl, J., Ravensberg, W.J., Urbaneja, A., 2018.  
743 Biological control using invertebrates and microorganisms: plenty of new  
744 opportunities. BioControl 63, 39–59. <https://doi.org/10.1007/s10526-017-9801-4>
- 745 Vianen, A. van, Lenteren, J.C. van, 1986. The parasite-host relationship between  
746 *Encarsia formosa* Gahan (Hym., Aphelinidae) and *Trialeurodes vaporariorum*  
747 (Westwood) (Hom., Aleyrodidae). XIV. Genetic and environmental factors  
748 influencing body-size and number of ovarioles of *Encarsia formosa*. J. Appl.  
749 Entomol. 101, 321–331. <https://doi.org/http://dx.doi.org/10.1111/j.1439-0418.1986.tb00866.x>
- 751 Wagner, J.D., Glover, M.D., Moseley, J.B., Moore, A.J., 1999. Heritability and fitness  
752 consequences of cannibalism in *Harmonia axyridis*. Evol. Ecol. Res. 1, 375–388.
- 753 Wajnberg, E., Colazza, S., 1998. Genetic variability in the area searched by a  
754 parasitic wasp: Analysis from automatic video tracking of the walking path. J.  
755 Insect Physiol. 44, 437–444. [https://doi.org/10.1016/S0022-1910\(98\)00032-8](https://doi.org/10.1016/S0022-1910(98)00032-8)
- 756 Wajnberg, E., Curty, C., Colazza, S., 2004. Genetic variation in the mechanisms of  
757 direct mutual interference in a parasitic wasp: Consequences in terms of patch-  
758 time allocation. J. Anim. Ecol. 73, 1179–1189. <https://doi.org/10.1111/j.0021-8790.2004.00892.x>
- 760 Wajnberg, E., Curty, C., Jervis, M., 2012. Intra-Population Genetic Variation in the  
761 Temporal Pattern of Egg Maturation in a Parasitoid Wasp. PLoS One 7, e45915.  
762 <https://doi.org/10.1371/journal.pone.0045915>
- 763 Wang, Q., Gu, H., Dorn, S., 2004. Genetic relationship between olfactory response  
764 and fitness in *Cotesia glomerata* (L.). Heredity (Edinb). 92, 579–584.  
765 <https://doi.org/10.1038/sj.hdy.6800464>
- 766 Wang, Q., Gu, H., Dorn, S., 2003. Selection on olfactory response to semiochemicals  
767 from a plant-host complex in a parasitic wasp. Heredity (Edinb). 91, 430–435.  
768 <https://doi.org/10.1038/sj.hdy.6800340>
- 769 Wickham, H., 2016. ggplot2: Elegant graphics for data analysis. Springer-Verlag, New  
770 York.



772

773 Figure 1. Flow diagram of the searching, screening, and extraction process used in this  
 774 systematic review. Classification step is based on fitness components from Mousseau  
 775 and Roff (Mousseau and Roff, 1987) and aspects of biological control research from  
 776 Leung et al (Leung et al., 2019)

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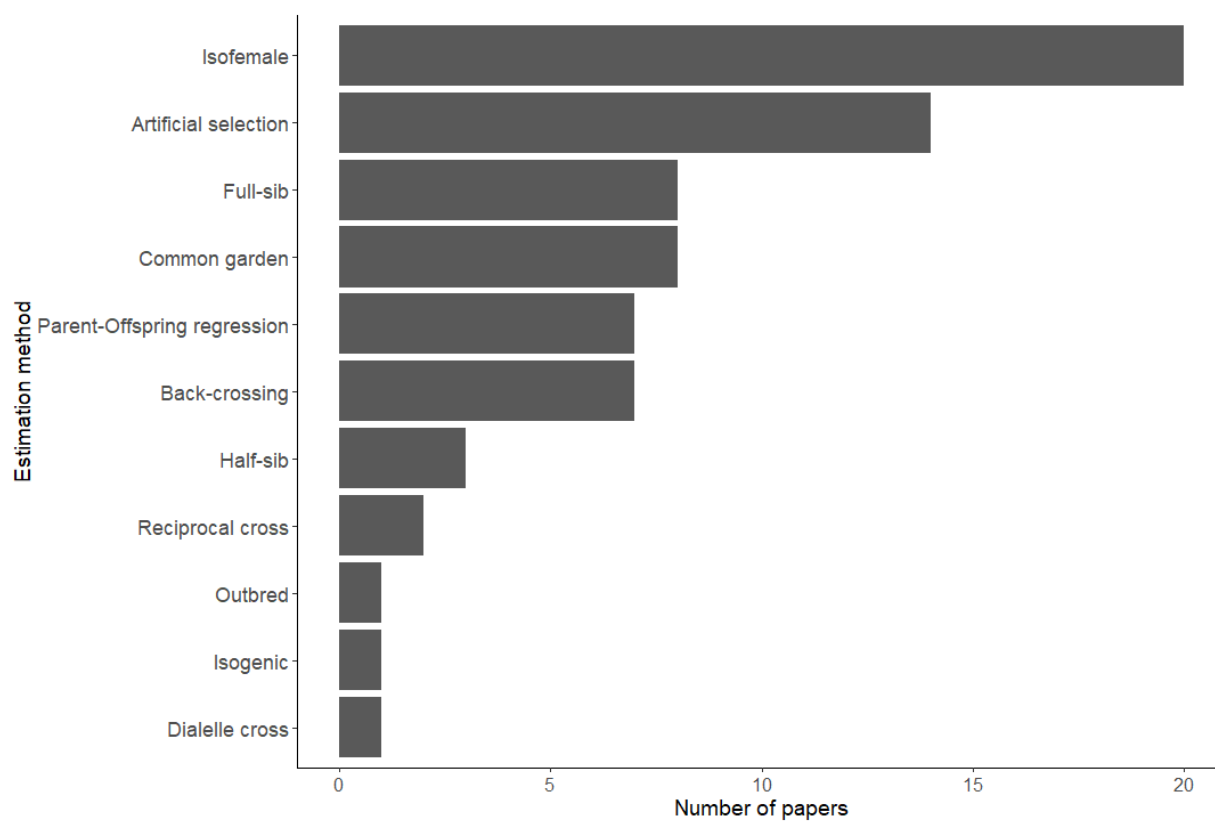


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779 Figure 2. Initial search returns displayed according to publication year (n = 2, 927).

780 Absent bar indicates 0 search hits for that year.

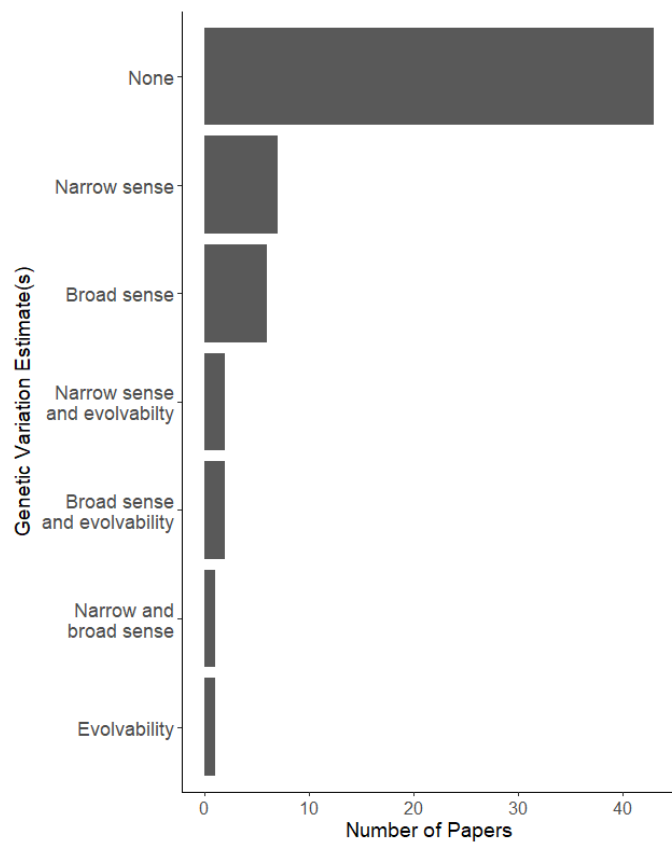
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783 Figure 3. Estimation methods of extracted papers, reduced to unique species per  
784 paper (n = 72).

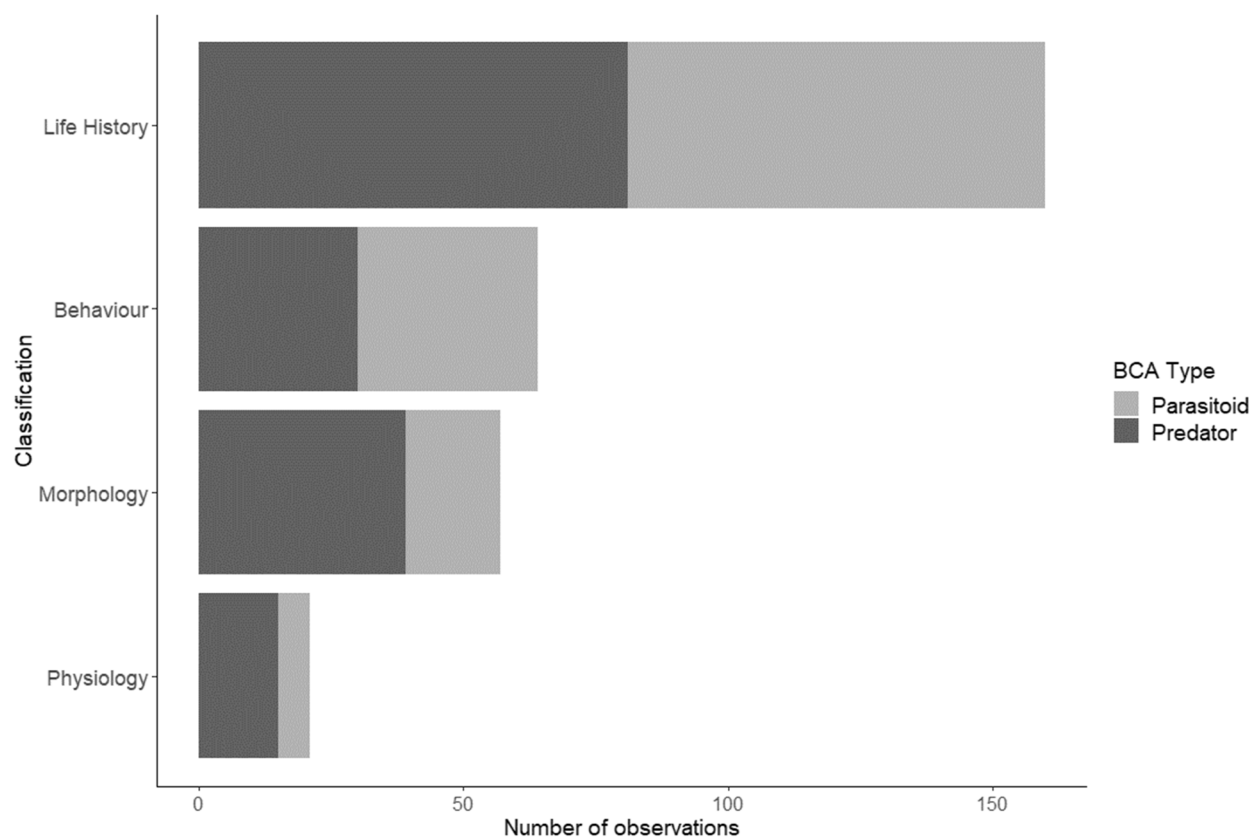
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787 Figure 4. Heritability estimates found in extracted papers (n = 69).

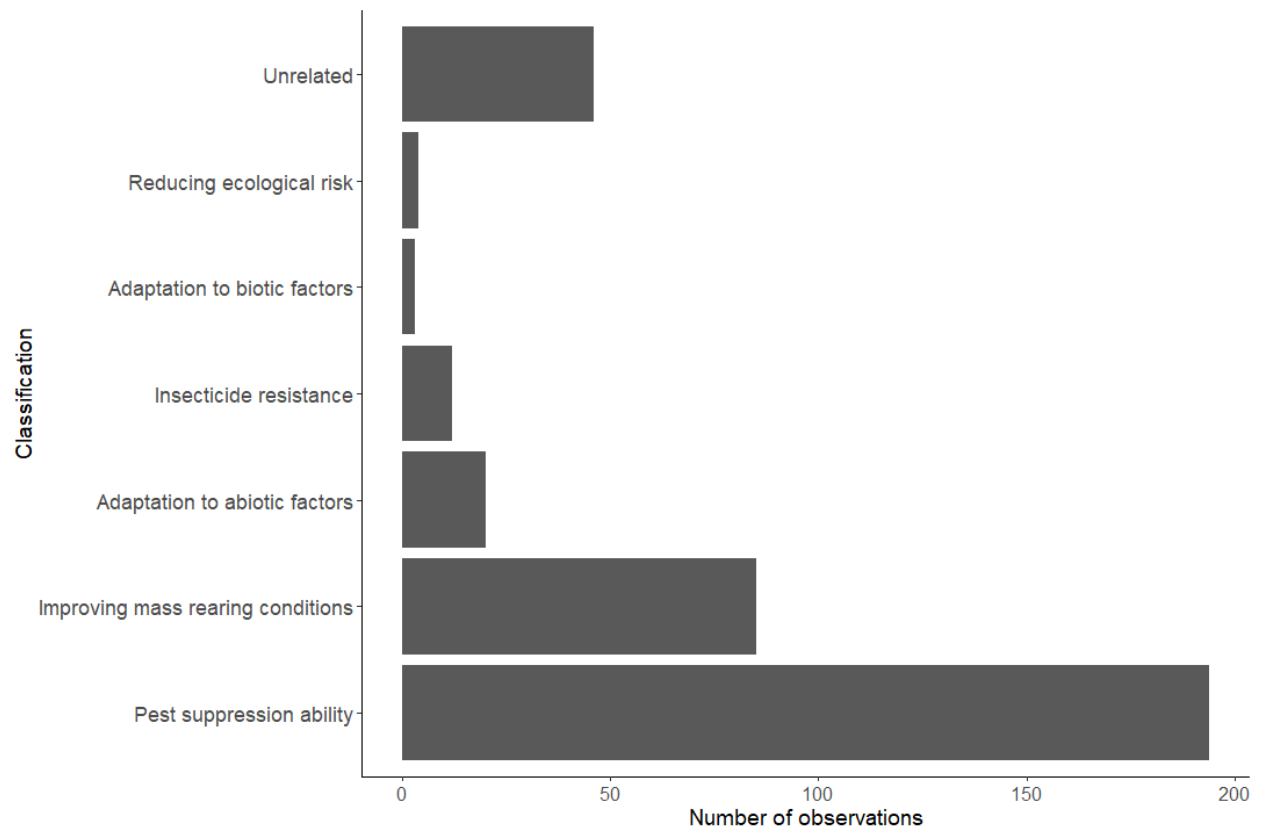
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790 Figure 5. Classification of heritable traits related to fitness, based on Mousseau and  
791 Roff (1976), in extracted papers according to BCA type (n = 302).

792



793

794 Figure 6. Classification of heritable traits related to importance to biological control  
795 research, based on Leung et al (2019), in extracted papers (n = 302).

796



797 Table 1. List of 69 studies examining the genetic heritability of traits in biological control agents  
798 (BCAs) that were extracted for this systematic review.

Paper ID <sup>a</sup>	In-text reference	Focal BCA	BCA Type
88	(Hufbauer, 2001)	<i>Aphidius ervi</i>	Parasitoid
299	(Guzmán-Larralde et al., 2014)	<i>Trichogramma pretiosum</i>	Parasitoid
340	(Bilodeau et al., 2013)	<i>Aphidius ervi</i>	Parasitoid
471	(Astles et al., 2005)	<i>Harmonia axyridis</i>	Predator
515	(Obrycki et al., 2001)	<i>Hippodamia convergens</i>	Predator
525	(Liu and Smith, 2000)	<i>Trichogramma minutum</i>	Parasitoid
543	(Grill et al., 1997)	<i>Harmonia axyridis</i>	Predator
564	(Rodríguez et al., 1994)	<i>Trichogramma fuentesi</i>	Parasitoid
571	(Marples et al., 1993)	<i>Adalia bipunctata</i>	Predator
577	(Sequeira and Mackauer, 1992)	<i>Aphidius ervi</i>	Parasitoid
768	(Navia et al., 2014)	<i>Amblyseius largoensis</i>	Predator
947	(Facon et al., 2011)	<i>Harmonia axyridis</i>	Predator
962	(Sayyed et al., 2010)	<i>Chrysoperla carnea</i>	Predator
1092	(Fukunaga and Akimoto, 2007)	<i>Harmonia axyridis</i>	Predator
1174	(Wang et al., 2004)	<i>Cotesia glomerata</i>	Parasitoid
1226	(Thomson and Hoffmann, 2009)	<i>Trichogramma carverae</i>	Parasitoid
1286	(Hufbauer and Via, 1999)	<i>Aphidius ervi</i>	Parasitoid
1311	(Bennett and Hoffmann, 1998)	<i>Trichogramma carverae</i>	Parasitoid
1379	(van Houten et al., 1995)	<i>Amblyseius barkeri</i> , <i>A. cucumeris</i>	Predator
1401	(Bruins et al., 1994)	<i>Trichogramma brassicae</i>	Parasitoid
1439	(Antolin, 1992)	<i>Muscidifurax raptor</i>	Parasitoid
1466	(Legner, 1991a)	<i>Muscidifurax raptorellus</i>	Parasitoid
1514	(Legner, 1988)	<i>Muscidifurax raptorellus</i>	Parasitoid
1520	(Legner, 1987)	<i>Muscidifurax raptorellus</i>	Parasitoid
1528	(Vianen and Lenteren, 1986)	<i>Encarsia formosa</i>	Parasitoid
1536	(Parker and Orzack, 1985)	<i>Nasonia vitripennis</i>	Parasitoid
1642	(Parreño et al., 2017)	<i>Lysiphlebus fabarum</i>	Parasitoid
1675	(Sepúlveda et al., 2017)	<i>Aphidius ervi</i>	Parasitoid
1684	(Mansoor et al., 2017)	<i>Chrysoperla carnea</i>	Predator
1851	(Kamala Jayanthi et al., 2014)	<i>Cryptolaemus montrouzieri</i>	Predator
1868	(Abbas et al., 2014)	<i>Chrysoperla carnea</i>	Predator
1899	(Paolucci et al., 2013)	<i>Nasonia vitripennis</i>	Parasitoid
1937	(Mansoor et al., 2013)	<i>Chrysoperla carnea</i>	Predator
2009	(Wajnberg et al., 2012)	<i>Trichogramma brassicae</i>	Parasitoid
2106	(Sandrock et al., 2010)	<i>Lysiphlebus fabarum</i>	Parasitoid
2108	(Henry et al., 2010)	<i>Aphidius ervi</i>	Parasitoid
2141	(Nachappa et al., 2010)	<i>Phytoseiulus persimillis</i>	Predator
2236	(El-Heneidy and Shoeb, 2007)	<i>Trichogramma evanescens</i>	Parasitoid
2278	(Pérez-Maluf et al., 2008)	<i>Leptopilina heterotoma</i>	Parasitoid
2280	(Pannebakker et al., 2008)	<i>Nasonia vitripennis</i>	Parasitoid
2307	(Shuker et al., 2007)	<i>Nasonia vitripennis</i>	Parasitoid
2312	(Kamping et al., 2007)	<i>Nasonia vitripennis</i>	Parasitoid

Paper ID <sup>a</sup>	In-text reference	Focal BCA	BCA Type
2453	(Wang et al., 2003)	<i>Cotesia glomerata</i>	Parasitoid
2465	(Wajnberg et al., 2004)	<i>Trissolcus basalıs</i>	Parasitoid
2470	(Jia et al., 2002)	<i>Phytoseiulus persimillis</i>	Predator
2595	(Maeda et al., 1999)	<i>Amblyseius womersleyi</i>	Predator
2601	(Wagner et al., 1999)	<i>Harmonia axyridis</i>	Predator
2633	(Lesna and Sabelis, 1999)	<i>Hypoaspis aculeifer</i>	Predator
2645	(Mason and Hopper, 1997)	<i>Aphelinus asychis</i>	Parasitoid
2657	(Wajnberg and Colazza, 1998)	<i>Trichogramma brassicae</i>	Parasitoid
2675	(Gilchrist, 1996)	<i>Aphidius ervi</i>	Parasitoid
2689	(Margolies et al., 1997)	<i>Phytoseiulus persimillis</i>	Predator
2696	(Sorati et al., 1996)	<i>Trichogramma brassicae</i>	Parasitoid
2707	(Henter et al., 1996)	<i>Encarsia formosa</i>	Parasitoid
2736	(Holloway et al., 1995)	<i>Adalia bipunctata</i>	Predator
2738	(Fleury et al., 1995)	<i>Leptopilina heterotoma</i>	Parasitoid
2741	(Henter, 1995)	<i>Aphidius ervi</i>	Parasitoid
2755	(Pompanon et al., 1994)	<i>Trichogramma brassicae, T. cacaoeciae</i>	Parasitoid
2759	(Geden et al., 1992)	<i>Muscidifurax raptor</i>	Parasitoid
2763	(Orzack and Gladstone, 1994)	<i>Nasonia vitripennis</i>	Parasitoid
2765	(Holloway et al., 1993)	<i>Adalia bipunctata</i>	Predator
2781	(Tauber and Tauber, 1992)	<i>Chrysoperla carnea</i>	Predator
2782	(Legner, 1993)	<i>Muscidifurax raptorellus</i>	Parasitoid
2791	(Legner, 1991b)	<i>Muscidifurax raptorellus</i>	Parasitoid
2799	(Orzack et al., 1991)	<i>Nasonia vitripennis</i>	Parasitoid
2845	(Boulétreau and Wajnberg, 1986)	<i>Leptopilina heterotoma</i>	Parasitoid
2846	(Tauber and Tauber, 1986)	<i>Chrysoperla carnea</i>	Predator
3001	(Al-Khateeb et al., 2018)	<i>Cryptolaemus montrouzieri</i>	Predator
3007	(Rasmussen et al., 2018)	<i>Orius majusculus</i>	Predator

<sup>a</sup> Paper identification number is a unique identifier for each paper and is cross-referenced in Table 2.

Table 2. Extracted observations from 69 studies examining the genetic heritability of life history traits in biological control agents (BCAs), listed according to type of BCA and then by species (n=302).

Type of BCA	Species	Trait of interest as reported	Estimation method	Genetic variation measure	Fitness component <sup>a</sup>	Biological control relevance <sup>b</sup>	Paper ID <sup>c</sup>	
Parasitoid	<i>Aphelinus asychis</i>	walking speed	common	none	B	7	2645	
		<i>Aphidius ervi</i>	body size, female offspring	full-sibling	$h^2$	L	7	577
			body size, male offspring	full-sibling	$h^2$	L	7	577
		body size, offspring mean	full-sibling	$h^2$	L	7	577	
		emergence	parent-offspring reg	$h^2$	L	1	2108	
		emergence	parent-offspring reg	$h^2$	L	1	2108	
		emergence	common	none	L	1	340	
		female sex ratio	parent-offspring reg	$h^2$	L	1;4	2108	
		female sex ratio	parent-offspring reg	$h^2$	L	1;4	2108	
		fitness	artificial selection	none	L	1	1675	
		handling time	artificial selection	none	B	1	1675	
		host adaptation	parent-offspring reg	$h^2$	L	1;3	2108	
		host adaptation	parent-offspring reg	$h^2$	L	1;3	2108	
		mass	full-sibling	none	M	1	2675	
		ovipositor contact	common	none	B	1	340	
	parasitism	half-sibling	$h^2, CV_A$	L	1	2741		
	parasitism	common	none	L	1	1286		
	performance breadth	full-sibling	none	P	2	2675		
	prey recognition	artificial selection	none	B	1;3	1675		
	time to oviposition	artificial selection	none	B	1	1675		
	total parasitism	parent-offspring reg	$h^2$	L	1	2108		
	total parasitism	parent-offspring reg	$h^2$	L	1	2108		
	virulence	common	none	P	1	88		
	<i>Cotesia glomerata</i>	walking speed	full-sibling	none	B	7	2675	
		development time	half-sibling	none	L	7	1174	
		efficiency of parasitism	half-sibling	none	L	1	1174	
		females per brood	half-sibling	none	L	1;4	1174	
		flight orientation	artificial selection	none	B	7	2453	

<u>Parasitoid</u>	<i>Cotesia glomerata</i>	landing success	artificial selection	none	B	7	2453	
		tibia length	half-sibling	none	M	1	1174	
	<i>Encarsia formosa</i>	contacts with host	common	none	B	7	2707	
		drumming behaviour	common	none	B	7	2707	
		encounters	common	none	B	7	2707	
		head width	parent-offspring reg	none	M	1;4	1528	
		host acceptance	common	none	B	1;3	2707	
		host feeding	common	none	B	1	2707	
		host rejection	common	none	B	1	2707	
		ovariole count	parent-offspring reg	none	L	7	1528	
		oviposition acceptance	common	none	B	1	2707	
		oviposition posture	common	none	B	7	2707	
		successful parasitization	common	none	L	1	2707	
		<i>Leptopilina heterotoma</i>	cumulative duration of oviposition	common	none	B	1	2278
			locomotor activity	isofemale	none	B	7	2738
			parasitism rate	common	none	L	1	2845
			probing duration	common	none	B	1	2278
	probing latency		common	none	B	1	2278	
	response to olfactory stimuli		common	none	B	1	2278	
	survival rate		common	none	L	1	2845	
	time to find larvae		common	none	B	1	2278	
	time to oviposition	common	none	B	1	2278		
	<i>Lysiphlebus fabarum</i>	parasitism	isofemale	$H^2$	L	1	2106	
		wing shape	isofemale	none	M	7	1642	
		wing size	isofemale	none	M	7	1642	
	<i>Muscidifurax raptor</i>	development time egg to adult	dialelle cross	$H^2$	L	7	1439	
		fecundity	dialelle cross	$H^2$	L	1;4	1439	
		fecundity	parent-offspring reg	$h^2$	L	1;4	2579	
		reproductive lifespan	dialelle cross	$H^2$	L	1;4	1439	
		sex ratio	dialelle cross	$H^2$	L	1;4	1439	
		wing length	parent-offspring reg	$h^2$	M	7	2579	
		wing length	parent-offspring reg	$h^2$	M	7	2579	

	<i>Muscidifurax raptorellus</i>	eggs laid	backcrossing	none	L	1	1514
		eggs per gregarious oviposition	backcrossing	none	L	1	1514
<u>Parasitoid</u>	<i>Muscidifurax raptorellus</i>	eggs per gregarious oviposition	backcrossing	none	L	1	1520
		eggs per gregarious oviposition	backcrossing	none	L	1	2791
		gregarious oviposition	backcrossing	none	L	1	1514
		gregarious oviposition	backcrossing	none	L	1	2782
		gregarious oviposition	backcrossing	none	L	1	1520
		gregarious oviposition	backcrossing	none	L	1	2791
		gregarious oviposition, total	backcrossing	none	L	1	2791
		hosts killed	backcrossing	none	P	1	1520
		hosts parasitized	backcrossing	none	L	1	1520
		longevity females	backcrossing	none	L	1	1520
		parasitism	isofemale	$h^2, H^2$	L	1	1466
		parasitism	isofemale	$h^2, H^2$	L	1	1466
		parasitism	backcrossing	none	L	1	2782
		parasitism	backcrossing	none	L	1	2791
		parasitism rate	backcrossing	none	L	1	1514
		sex ratio	backcrossing	none	L	1;4	1520
		sex ratio	backcrossing	none	L	1;4	2791
		total eggs laid	backcrossing	none	L	1	1514
		total eggs laid	backcrossing	none	L	1	2791
		total progeny	backcrossing	none	L	1	1520
	<i>Nasonia vitripennis</i>	copulation duration	half-sibling	$h^2, CV_A$	B	7	2307
		courtship duration	half-sibling	$h^2, CV_A$	B	7	2307
		critical photoperiod	isofemale	none	P	1;4	1899
		diapause response	isofemale	none	L	4	1899
		gynandromorphism	isofemale	none	L	7	2312
		lifespan	isofemale	none	L	1;4	1899
		receptivity at 1st courtship	half-sibling	$h^2, CV_A$	B	7	2307
		receptivity within 10 min	half-sibling	$h^2, CV_A$	B	7	2307
		second foundress sex ratio	isofemale	$H^2$	L	1	2280
		sex ratio	isofemale	none	L	1;4	1536

<u>Parasitoid</u>	<i>Trichogramma brassicae</i>	sex ratio	isofemale	none	L	1;4	2763
		sex ratio	isofemale	none	L	1;4	2799
		single foundress sex raio	isofemale	$H^2$	L	1;4	2280
		area searched	isofemale	none	B	1	2657
		body length	isofemale	none	M	1	2696
		body length	outbred	none	M	1	2696
		fecundity	isofemale	none	L	1;4	2696
		fecundity	outbred	none	L	1;4	2696
		head width	isofemale	none	M	1;4	2696
		head width	outbred	none	M	1;4	2696
		hind tibia length	isofemale	none	M	1;4	2696
		hind tibia length	outbred	none	M	1;4	2696
		locomotor activity	isofemale	none	B	7	2755
		male mating	isofemale	none	L	7	2696
		male mating	outbred	none	L	7	2696
		ovigeny index	parent-offspring reg	none	L	1	2009
		reactive distance	full-sibling	none	B	7	1401
		reproductive concentration index	parent-offspring reg	none	L	1	2009
	walking speed	isofemale	none	B	7	2657	
	locomotor activity	isofemale	none	B	7	2755	
	fecundity	isofemale	$H^2$	L	1;4	1226	
	forewing length	parent-offspring reg	none	M	7	1311	
	forewing width	parent-offspring reg	none	M	7	1311	
	head width	parent-offspring reg	none	M	1;4	1311	
	hind tibia length	parent-offspring reg	none	M	1;4	1311	
	hindwing length	parent-offspring reg	none	M	7	1311	
	<i>Trichogramma evanescens</i>	developmental period	isofemale	none	L	4	2236
		emergence rate	isofemale	none	L	1;4	2236
		parasitism	isofemale	none	L	1	2236
		sex ratio	isofemale	none	L	1;4	2236
		thermal requirements	isofemale	none	P	2	2236
		host egg color	isofemale	none	P	7	564

<u>Parasitoid</u>	<i>Trichogramma evanescens</i>	emergence	isogenic	$H^2$	L	1	525	
		fecundity	isogenic	$H^2$	L	1;4	525	
		longevity, females	isogenic	$H^2$	L	1	525	
		number female offspring	isogenic	$H^2$	L	1	525	
		number male offspring	isogenic	$H^2$	L	1	525	
	<u>Predator</u>	<i>Trissolcus basalidis</i>	sex ratio	isogenic	$H^2$	L	1;4	525
			fecundity	isofemale	none	L	1;4	299
			female offspring	isofemale	none	L	1	299
			mean generation time	isofemale	none	L	1;4	299
			reproductive rate	isofemale	none	L	1	299
<i>Adalia bipunctata</i>		sex ratio	isofemale	none	L	1;4	299	
		patch residency	isofemale	none	B	1	2465	
		patch-leaving tendency	isofemale	none	B	1	2465	
		adaline concentration	parent-offspring reg	none	P	6	2765	
		body weight	parent-offspring reg	none	M	1	2765	
	growth rate	parent-offspring reg	none	M	4	2765		
	reflex fluid	parent-offspring reg	none	P	2	2765		
	spot 1 female	parent-offspring reg	none	M	2	2736		
	spot 1 male	parent-offspring reg	none	M	2	2736		
	spot 2 female	parent-offspring reg	none	M	2	2736		
	spot 2 male	parent-offspring reg	none	M	2	2736		
	spot 3 female	parent-offspring reg	none	M	2	2736		
	spot 3 male	parent-offspring reg	none	M	2	2736		
	spot 4 female	parent-offspring reg	none	M	2	2736		
	spot 4 male	parent-offspring reg	none	M	2	2736		
<i>Amblyseius barkeri</i>	wingless	outbred	none	M	1	571		
	oviposition rate	artificial selection	none	B	1	1379		
<i>Amblyseius cucumeris</i>	predation rate	artificial selection	none	L	1	1379		
	eggs laid per female per day	artificial selection	none	L	1	1379		
	eggs laid per female per day	artificial selection	none	L	1	1379		
	eggs laid per female per day	artificial selection	none	L	1	1379		

<u>Predator</u>	<i>Amblyseius cucumeris</i>	eggs laid per female per day	artificial selection	none	L	1	1379
		larvae killed per female per day	artificial selection	none	B	1	1379
		larvae killed per female per day	artificial selection	none	B	1	1379
		larvae killed per female per day	artificial selection	none	B	1	1379
		larvae killed per female per day	artificial selection	none	B	1	1379
	<i>Amblyseius largoensis</i>	larvae killed per female per day	artificial selection	none	B	1	1379
		oviposition rate	artificial selection	none	B	1	1379
		predation rate	artificial selection	none	L	1	1379
		egg viability	backcrossing	none	L	1;4	768
		eggs laid per female per day	backcrossing	none	L	1	768
	<i>Amblyseius womersleyi</i>	number of eggs	backcrossing	none	L	1	768
		sex ratio	backcrossing	none	L	1;4	768
		dispersal ratio over time	common	none	B	1	2595
		patch choice	common	none	B	1	2595
		<i>Chrysoperla carnea</i>	benzoate resistance	artificial selection	none	P	2
	development time egg to adult		artificial selection	none	L	4	1868
	development time egg to adult		artificial selection	none	L	4	1937
	diapause		full-sibling	none	L	4	2781
	diapause		full-sibling	none	L	4	2846
	eggs laid per female		artificial selection	none	L	1	1868
	eggs laid per female		artificial selection	none	L	1	1937
	emergence rate of healthy adults		artificial selection	none	L	1;4	1868
	emergence rate of healthy adults		artificial selection	none	L	1;4	1937
	fecundity		full-sibling	none	L	1;4	2781
	hatchability		artificial selection	none	L	1;4	1868
	hatchability		artificial selection	none	L	1;4	1937
	larval development time		artificial selection	none	L	4	1868
	larval development time	artificial selection	none	L	4	1937	
	larval survival rate	artificial selection	none	L	5	1868	
	larval survival rate	artificial selection	none	L	5	1937	
	LD50	artificial selection	none	P	5	1684	
	LD50	artificial selection	none	P	5	1684	
	LD50	artificial selection	none	P	5	1684	



<u>Predator</u>	<i>Chrysoperla carnea</i>	LD50	artificial selection	none	P	5	1684
		net replacement rate	artificial selection	none	L	1;4	1868
		net replacement rate	artificial selection	none	L	1;4	1937
		nondiapause preoviposition period	full-sibling	none	L	1;4	2781
		number of nextg gen larvae	artificial selection	none	L	1;4	1868
		number of nextg gen larvae	artificial selection	none	L	1;4	1937
		pupal duration	artificial selection	none	L	4	1868
		pupal duration	artificial selection	none	L	4	1937
		pupal weight	artificial selection	none	M	7	1868
		pupal weight	artificial selection	none	M	7	1937
		pupation rate	artificial selection	none	L	7	1868
		pupation rate	artificial selection	none	L	7	1937
		relative fitness	artificial selection	none	L	1	1868
		relative fitness	artificial selection	none	L	1	1937
		resistance alphamethrin	artificial selection	$h^2$	P	5	962
	resistance chlorpyrifos2	artificial selection	$h^2$	P	5	962	
	resistance deltamethrin2	artificial selection	$h^2$	P	5	962	
	resistance lambdacyhalothrin2	artificial selection	$h^2$	P	5	962	
	resistance profenofos2	artificial selection	$h^2$	P	5	962	
	spinosad resistance	artificial selection	none	P	5	1868	
	<i>Cryptolaemus montrouzieri</i>	developmental period	artificial selection	none	L	4	3001
		fecundity	artificial selection	none	L	1;4	3001
		female length	artificial selection	none	M	1	3001
		larval developmental time	artificial selection	none	L	4	3001
		prey eaten per day	artificial selection	none	B	1	1851
prey potential of 3rd instar		artificial selection	none	B	1	3001	
prey potential of female		artificial selection	none	B	1	3001	
reproduction		artificial selection	none	L	1	3001	
survival rate	artificial selection	none	L	1;4	3001		
<i>Harmonia axyridis</i>	abdomen length	full-sibling	$H^2, CV_A$	M	1	471	
	adult development time	artificial selection	$h^2, CV_A$	L	1	2601	
	adult development time	artificial selection	$h^2, CV_A$	L	1	2601	

Predator

*Harmonia axyridis*

adult mass	full-sibling	$H^2, CV_A$	M	1	471
adult size	artificial selection	$h^2, CV_A$	M	1	2601
adult size	artificial selection	$h^2, CV_A$	M	1	2601
body length	reciprocal cross	CV	M	1	947
body length	reciprocal cross	CV	M	1	947
body length	reciprocal cross	CV	M	1	947
body length	reciprocal cross	CV	M	1	947
cannibalism rate	artificial selection	$h^2, CV_A$	B	4	2601
cannibalism rate	artificial selection	$h^2, CV_A$	B	4	2601
development	full-sibling	$H^2, CV_A$	L	4	543
development	full-sibling	$H^2, CV_A$	L	4	543
development time	full-sibling	$H^2, CV_A$	L	4	471
development time egg to adult	reciprocal cross	$CV_A$	L	4	947
development time egg to adult	reciprocal cross	$CV_A$	L	4	947
development time egg to adult	reciprocal cross	$CV_A$	L	4	947
development time egg to adult	reciprocal cross	$CV_A$	L	4	947
female reproductive investment	reciprocal cross	$CV_A$	L	1	947
female reproductive investment	reciprocal cross	$CV_A$	L	1	947
female reproductive investment	reciprocal cross	$CV_A$	L	1	947
female reproductive investment	reciprocal cross	$CV_A$	L	1	947
hatching time	full-sibling	$H^2, CV_A$	L	4	471
larval development time	artificial selection	$h^2, CV_A$	L	4	2601
larval development time	artificial selection	$h^2, CV_A$	L	4	2601
larval size	artificial selection	$h^2, CV_A$	M	1	2601
larval size	artificial selection	$h^2, CV_A$	M	1	2601
larval survival to adulthood	reciprocal cross	$CV_A$	L	7	947
larval survival to adulthood	reciprocal cross	$CV_A$	L	7	947
larval survival to adulthood	reciprocal cross	$CV_A$	L	7	947
larval survival to adulthood	reciprocal cross	$CV_A$	L	7	947
larval weight	full-sibling	$H^2$	M	1	1092
larval weight	full-sibling	$H^2$	M	1	1092
orange wing reflectance	full-sibling	$H^2, CV_A$	M	2	543
orange wing reflectance	full-sibling	$H^2, CV_A$	M	2	543

<u>Predator</u>	<i>Harmonia axyridis</i>	pronotum width	full-sibling	$H^2, CV_A$	M	7	471	
		pronotum width	full-sibling	$H^2, CV_A$	M	7	543	
		pronotum width	full-sibling	$H^2, CV_A$	M	7	543	
		red wing reflectance	full-sibling	$H^2, CV_A$	M	2	543	
		red wing reflectance	full-sibling	$H^2, CV_A$	M	2	543	
		starvation resistance	reciprocal cross	$CV_A$	L	1;4	947	
		starvation resistance	reciprocal cross	$CV_A$	L	1;4	947	
		starvation resistance	reciprocal cross	$CV_A$	L	1;4	947	
		starvation resistance	reciprocal cross	$CV_A$	L	1;4	947	
		survival rate in quiescent conditions	reciprocal cross	$CV_A$	L	1;4	947	
		survival rate in quiescent conditions	reciprocal cross	$CV_A$	L	1;4	947	
		survival rate in quiescent conditions	reciprocal cross	$CV_A$	L	1;4	947	
		survival rate in quiescent conditions	reciprocal cross	$CV_A$	L	1;4	947	
		survival rate in quiescent conditions	reciprocal cross	$CV_A$	L	1;4	947	
		thorax length	full-sibling	$H^2, CV_A$	M	1	471	
		<i>Hippodamia convergens</i>	development time egg to adult	backcrossing	none	L	4	515
			elytra size	backcrossing	none	M	2	515
			fecundity	backcrossing	none	L	1;4	515
			fertility	backcrossing	none	L	1;4	515
pronotum size	backcrossing		none	M	7	515		
sex ratio	backcrossing		none	L	1	515		
survival	backcrossing		none	L	1;4	515		
weight, female	backcrossing		none	M	1	515		
<i>Hypoaspis aculeifer</i>	weight, male	backcrossing	none	M	1	515		
	mate choice	backcrossing	none	B	4	2633		
<i>Orius majusculus</i>	basal activity	reciprocal cross	none	B	2	3007		
	body size	reciprocal cross	none	M	1	3007		
	critical thermal maximum	reciprocal cross	none	P	2	3007		
	predation rate	reciprocal cross	none	B	1	3007		
	starvation tolerance	reciprocal cross	none	L	1;4	3007		
	temperature of maximal activity	reciprocal cross	none	P	2	3007		
<i>Phytoseiulus persimillis</i>	consumption rate	artificial selection	$h^2$	B	1	2141		
	consumption rate	artificial selection	$h^2$	B	1	2141		

<u>Predator</u>	<i>Phytoseiulus persimilis</i>	consumption rate	artificial selection	$h^2$	B	1	2141	
		consumption rate	isofemale	$h^2$	B	1	2470	
		conversion efficiency	artificial selection	$h^2$	L	1	2141	
		conversion efficiency	artificial selection	$h^2$	L	1	2141	
		conversion efficiency	artificial selection	$h^2$	L	1	2141	
		dispersal response to prey density	artificial selection	$h^2$	B	1	2141	
		dispersal response to prey density	artificial selection	$h^2$	B	1	2141	
		dispersal response to prey density	artificial selection	$h^2$	B	1	2141	
		eggs laid per female per day avg	artificial selection	none	L	1	2689	
		olfactory attraction to <i>T. urticae</i>						
		induced plant volatiles	artificial selection	$h^2$	B	6	2141	
		olfactory attraction to <i>T. urticae</i>						
		induced plant volatiles	artificial selection	$h^2$	B	6	2141	
		oviposition rate	isofemale	$h^2$	L	1	2470	
		patch residency	isofemale	$h^2$	B	1	2470	
		prey consumption	artificial selection	none	B	1	2689	
prey location	isofemale	$h^2$	B	1	2470			
residence time	artificial selection	none	B	1	2689			

<sup>a</sup> Classification based on Mousseau and Roff (Mousseau and Roff, 1987)

<sup>b</sup> Classification based on Leung et al (Leung et al., 2019)

<sup>c</sup> Paper identification numbers are unique to each paper, and are cross-referenced in Table 1.