Review

# **Current Advances in Allosteric Modulation of Muscarinic Receptors**

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**Abstract:** Allosteric modulators are ligands that bind to a site on the receptor that is spatially separated from the orthosteric binding site for the endogenous neurotransmitter. Allosteric modulators modulate the binding affinity, potency and efficacy of orthosteric ligands. Muscarinic acetylcholine receptors are prototypical allosterically-modulated G-protein-coupled receptors. They are a potential therapeutic target for the treatment of psychiatric, neurologic and internal diseases like schizophrenia, Alzheimer's disease, Huntington disease, type 2 diabetes or chronic pulmonary obstruction. Here we review progress made during the last decade in our understanding of their mechanisms of binding, allosteric modulation and in vivo actions of in order to understand the translational impact of studying this important class of pharmacological agents. We overview newly developed allosteric modulators of muscarinic receptors as well as new spin-off ideas like bitopic ligands combining allosteric and orthosteric moieties and photo-switchable ligands based on bitopic agents.

Keywords: acetylcholine; muscarinic receptors; allosteric modulation

## 1. Introduction

Slow metabotropic responses to acetylcholine are mediated by muscarinic receptors. Five distinct subtypes of muscarinic acetylcholine receptors ( $M_1$ - $M_5$ ) have been identified in the human genome[1]. The structure of all five receptor subtypes was resolved by X-ray crystallography[2–6]. Muscarinic receptors are members of class A of G-protein-coupled receptor (GPCR).  $M_1$ ,  $M_3$  and  $M_5$  subtypes preferentially activate phospholipase C and calcium mobilization through  $G_{q/11}$ , whereas  $M_2$  and  $M_4$  receptors inhibit the activity of adenylyl cyclase by activation of the  $\alpha$ -subunit of the  $G_{i/0}$  family of G-proteins. The latter two receptors also modulate the conductance of ion channels (e.g. inward rectifying potassium ion channels) by  $\beta\gamma$ -dimers of the  $G_{i/0}$  G-proteins[7]. Muscarinic receptors mediate a wide range of physiological functions in the central and peripheral nervous system and innervated tissues. Muscarinic receptors thus represent a potential therapeutic target for the treatment of psychiatric and neurologic conditions (e.g., schizophrenia, Alzheimer's disease, Huntington disease)[8,9] as well as internal diseases (e.g. type 2 diabetes, asthma, chronic pulmonary obstruction, incontinence)[10–12].

The concept of allosterism was formally introduced into the field of enzymology by Monod et al.[13] and Koshland et al.[14] in 1965 and 1966, respectively. The former model was termed concerted the latter one sequential. Allosteric modulation of GPCR is much simpler than that of enzymes. GPCR allosteric modulators bind to a site on the receptor that is spatially distinct from that of the endogenous transmitter, acetylcholine in case of muscarinic receptors. Consequently, binding of an allosteric modulator and an orthosteric ligand is not mutually exclusive, i.e., both

ligands may bind to the receptor simultaneously to form a ternary complex. Binding of allosteric modulators induces a change in the conformation of the receptor that results in changes in affinity (eventually potency and efficacy) of the orthosteric ligand[15].

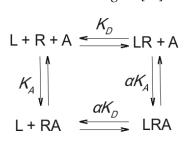


Figure 1. An orthosteric ligand L binds to the receptor R with equilibrium dissociation constant  $K_D$  and an allosteric modulator A binds to the receptor R with equilibrium dissociation constant  $K_A$ . The orthosteric ligand L and the allosteric modulator A can bind concurrently to the receptor R to form a ternary complex LRA. Binding of one ligand to the receptor changes the equilibrium dissociation constant of the other ligand by factor of cooperativity  $\alpha[15]$ .

Based on the effects of an allosteric modulator on the affinity of an orthosteric ligand, allosteric modulators may be classified into 3 categories: 1. Positive allosteric modulators (PAM) that increase the affinity of orthosteric ligands; 2. Negative allosteric modulators (NAM) that decrease the affinity of orthosteric ligands; and 3. Neutral allosteric modulators that do not affect the affinity of the orthosteric ligand. When potency and efficacy are taken into account, these 3 categories expand to 5: 1. Pure PAMs; 2. PAM-agonists that possess intrinsic agonistic propensity in the absence of the orthosteric agonists they modulate; 3. PAM-antagonists that lower the efficacy of the agonists they modulate; 4. Pure NAMs; 5. NAM-agonists that possess own agonistic propensity in the absence of the agonists they modulate; 6. Silent allosteric modulators (SAMs) that, although they bind to the receptor, do not affect the affinity, potency or efficacy of the orthosteric ligand and do not have agonistic propensity on their own.

As early as in 1969, Lüllmann et al. have shown in their pioneering work—that alkane-bis-ammonium compounds inhibit the functional response to the conventional muscarinic agonist carbachol non-competitively[16]. Later, Clark and Mitchelson discovered that gallamine similarly inhibits the action of acetylcholine and carbachol on the function of heart atria in a non-competitive manner[17]. The concentration-response curves to the agonists were shifted to the right but the magnitude of the progressive shifts diminished with increasing concentrations of gallamine. When the action of acetylcholine on the heart was evaluated in the combined presence of gallamine and the antagonist atropine, the inhibition of functional response to carbachol was smaller than expected for the effects of two competitive antagonists. These observations led to the conclusion that the action of gallamine takes place at an allosteric site on the receptor, resulting in negative cooperativity with binding of both orthosteric agonists and antagonists. Since then a wide variety of allosteric modulators has been discovered. These include inhibitors of acetylcholinesterase, ion channel blockers, various alkaloids, small peptides, etc. For review see[18]. Thanks to early intensive research, muscarinic receptors became a useful prototype of allosterically-modulated GPCRs.

### 2. Advantages of allosteric modulators as therapeutics

2.1. Selectivity by targeting less conserved domains on the receptor

Muscarinic receptor subtypes share high structural homology in the transmembrane domains where the orthosteric binding site is located. On the other hand, domains out of membrane are less conserved. Targeting allosteric domains allows achieving binding selectivity for certain receptor subtypes to an extent that is not possible with orthosteric ligands.

2.2. Conservation of space and time pattern of signaling

Theoretically, a pure PAM of acetylcholine would only induce an action when endogenous acetylcholine is released. Consequently, its action would be restricted in space and time to those synapses where signaling is currently happening. Thus, enhanced space and time pattern of signaling could be restored under diminished acetylcholine release that is typical in neurodegenerative disorders, e.g., Alzheimer's disease.

#### 2.3. Absolute selectivity

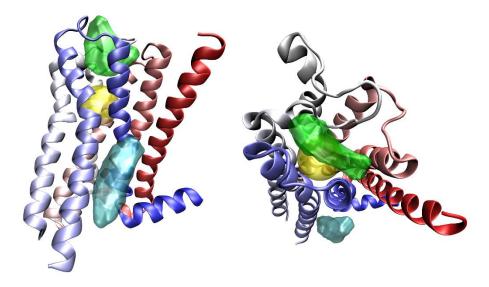
Theoretically, absolute selectivity of an allosteric agent can be achieved by having a ligand with the desired cooperativity at a given receptor subtype and neutral (silent) cooperativity at the rest of the subtypes. Selectivity may be derived from binding cooperativity as well as from effects on potency and efficacy.

#### 2.4. Selective blocking of activated receptors

PAM-antagonists that exhibit partial or absolute selectivity for a given receptor subtype may be used to selectively target receptors activated by the endogenous neurotransmitter located in specific tissues or organs[19]. This feature can be used to selectively reverse persistent excessive agonism under certain pathological conditions (e.g. bronchospasm in asthma[20]) or overstimulation of salivary and lacrimal glands after organophosphate poisoning in case of M<sub>3</sub>-selective PAM-antagonists).

# 3. Location of the allosteric binding sites on muscarinic receptors

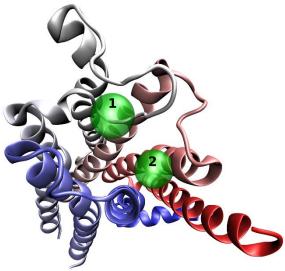
The binding site of classical allosteric modulators like gallamine, alcuronium or alkane-bis-ammonium compounds has been located between the second (o2) and third extracellular (o3) loops (Figure 2, green) [21,22]. The charged EDGE motif in the o2 loop plays a critical role in the binding of these ligands[23]. In the crystal structure of the  $M_2$  receptor[4], the allosteric modulator LY2119620 (that binds to the same site between o2 and o3) does not form a hydrogen bond to E172 or E175 of the EDGE motif but makes  $\pi$  - $\pi$  interactions to the adjacent Y177 in the o2 and Y426 in the extracellular edge of TM7.



**Figure 2.** Side view with TM6 front (left) and extracellular view (right) of the orthosteric (yellow), common allosteric (green) and cholesterol (cyan) binding sites at the M<sub>2</sub> receptor (colored in redwhite-blue gradient).

Computer modelling studies of allosteric ligand binding to M<sub>2</sub> receptors revealed two centers for binding of the electropositive part of allosteric ligands (Figure 3)[24]. The first center consists of Y177, N410, N419 and W422 and the second consists of Y80, Y83, T84 and T423. Further, this study shows that alkane-bis-ammonium compounds, gallamine, alcuronium and strychnine bind to the fist center. M<sub>1</sub>-selective PAM benzyl quinolone carboxylic acid (BQCA) binds to Y179 and F182 in the o2 loop, and E397 and W400 in TM7[25]. Thus, the binding site of BQCA overlaps with the common

allosteric binding site. In contrast, M<sub>4</sub>-selective PAM LY2033298 interacts with the o1, o2 and o3 loops[26]. Key binding residues of LY2033298 are K95 in the o1, F186 in o2 and D432 in o3 (M<sub>4</sub> numbering). A cryptic allosteric binding pocket in the extracellular domain that is absent in existing crystal structures was predicted by simulation of molecular dynamics and confirmed by mutagenesis experiments[27]. This cryptic pocket is dynamically formed in the vicinity of the common allosteric site center 1 by rearrangement of conserved E<sup>7,36</sup> in TM7, Y<sup>2,64</sup> in the o2 loop close to TM2 and C<sup>45,50</sup> in the middle of the o3 loop (Ballesteros-Weinstein numbering[28]). The site is preferentially formed at the M<sub>1</sub> receptor and was identified as a binding site of highly M<sub>1</sub>-selective PAM BQZ12 (3-((1S,2S)-2-hydroxycyclohexyl)-6-((6-(1-methyl-1H-pyrazol-4-yl)pyridin-3-yl)methyl)benzo[h]quinazolin-4(3H)-one)[29,30].



**Figure 3.** Extracellular view of two binding centers (green) in the common allosteric binding site at the M<sub>2</sub> receptor (colored in red-white-blue gradient).

While the majority of known muscarinic allosteric ligands bind to the site between the o2 and o3 loops, sterol-based WIN-compounds were found to interact with gallamine and strychnine in a non-competitive manner[31]. Thus, the binding site for WIN-compounds is not between the o2 and o3 loops but somewhere else. However, the precise location of the WIN-compound binding site has not been determined yet. Interestingly, cholesterol also allosterically modulates binding and function of muscarinic receptors[32–35]. Using site-directed mutagenesis, the binding site for membrane cholesterol was located to the groove between TM6 and TM7 in the intracellular leaflet of the membrane[35]. Membrane cholesterol allosterically modulates many GPCRs[36]. Cholesterol co-crystalized with GPCRs at various sites, both in the extracellular and intracellular leaflet of the membrane[36]. Computer modeling of the interaction between membrane cholesterol and GPCR suggests the possibility of several cholesterol binding sites per one molecule of GPCR[37]. How many cholesterol binding sites muscarinic receptors do have and whether WIN-compounds bind to the cholesterol binding site remains to be elucidated.

#### 4. Molecular mechanisms of action of allosteric modulators

The maximum magnitude of the effects of an allosteric modulator at a given receptor (cooperativity) varies from one orthosteric ligand to another. Comparison of the cooperativity of a given allosteric ligand with several orthosteric ligands suggested that cooperativity is dependent on the distance between the electropositive and electronegative part of the orthosteric ligand[38]. Thus, data suggested that allosteric modulators change the distance between TM3 interacting with electropositive and TM6 interacting with an electronegative part of the orthosteric ligand. This hypothesis was confirmed later by crystal structures and molecular modeling[4,39]. Numerous studies identified differential key amino acids to govern allosteric action of various allosteric modulators, suggesting the existence of multiple allosteric switches on muscarinic receptors. For

example, M<sub>4</sub>-selective PAM LY2033298 gains its efficacy from interaction with K95 in the o1 loop and its binding cooperativity results from interaction with F186 in the o2 loop (M<sub>4</sub> numbering)[26]. In contrast, tyrosine in the o2 loop (Y179 in M<sub>1</sub>, Y177 in M<sub>2</sub>) and tryptophan in TM7 (W<sup>7,35</sup>, Ballesteros-Weinstein numbering[40]) were identified as key residues defining efficacy and cooperativity of M<sub>1</sub>-selective PAM of acetylcholine (BQCA) as well as M<sub>2</sub>-selective PAM of iperoxo (LY2119620) [4,25,29]. The common feature found for all PAMs is shrinkage of the vestibule to the orthosteric binding site that is accompanied by closure of the binding pocket[41]. Divergent mechanisms underlying how shrinkage of the vestibule is achieved probably represent the molecular basis of PAM receptor subtype selectivity.

Many GPCRs including muscarinic receptors activate several signaling pathways upon activation by agonist[42-44]. It is generally accepted that structurally different agonists induce specific changes in the conformation of GPCRs that can lead to non-uniform modulation of signaling pathways. This preferential orientation of the signaling of a GPCR towards a subset of its signal transducers is termed signaling bias[45]. In principle, structure of the receptor in a ternary complex with agonist and allosteric modulator is distinct from the one in a binary complex with agonist only and may result in biased signaling too. Attaining signaling bias via allosteric modulation could bring a new plethora of possibilities to modulate receptor function. It has been shown at M1 receptors that the allosteric modulator VU0029767 ((E)-2-(4-ethoxyphenylamino)-N'-((2-hydroxynaphthalen-1yl)methylene)acetohydrazide) acts as a PAM of acetylcholine-induced intracellular calcium mobilization while being a NAM of acetylcholine-induced activation of phospholipase D [46]. Based on this observation it has been suggested that differential modulation of receptor coupling to downstream signaling pathways by allosteric modulators may result in signaling bias. However, structurally divergent M<sub>1</sub> PAMs BQCA and MIPS1674 have shown only small differences in the modulation of the inositol phosphate, β-arrestin and ERK<sub>1/2</sub> pathways[47]. Residues near the common allosteric binding site have been implicated in mediating biased signaling even of orthosteric ligands at several GPCRs[48]. Thus, divergent mechanisms of shrinkage of the vestibule to the binding pocket may not only stand behind subtype-selective effects of allosteric modulators but also drive ligand bias that is common to the entire A class of GPCRs[49].

#### 5. Role of the common allosteric binding site in the binding of orthosteric ligands

The common allosteric binding site located between the o2 and o3 loops represents the vestibule to the orthosteric binding site. The model in which an orthosteric ligand binds transiently to a secondary site before moving to the orthosteric binding site (so-called tandem two-site model[50]) was proposed to explain apparent receptor isomerization upon antagonist binding[51,52]. Interaction of orthosteric ligands with the allosteric domain between the o2 and o3 loops was confirmed and studied in detail by computer modeling[3,53,54]. These studies showed that two-step binding is common for all orthosteric ligands studied, that orthosteric ligands interact primarily with Y177 (common allosteric center 1), N410, N419 and W422 (common allosteric center 2) (M2 numbering). Subtype variations of affinity and kinetics of orthosteric ligands may be attributed to variation in their interaction with the allosteric site[6].

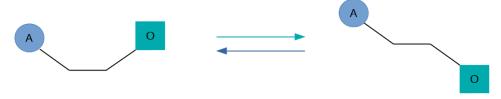
The strong interaction of tiotropium with the allosteric binding site also explains the discrepancy between extremely slow binding kinetics and immediate inhibitory action of this bronchodilator[55]. Tiotropium binding to the allosteric site is fast and results in antagonism of the functional effects of M<sub>3</sub> receptor.

The M<sub>2</sub>-selective antagonist methoctramine (N,N'-bis[6-[[(2-methoxyphenyl)-methyl]hexyl]-1,8-octane] diamine) competitively inhibits binding of orthosteric ligands. At high concentrations slows down dissociation of [<sup>3</sup>H]N-methylcopolamine. Detailed analysis of methoctramine binding revealed two modes of interaction with the receptor. Transient binding to the common allosteric site center 1 (namely E175 in the o2 loop) is followed by stable binding that occurs concurrently to both the allosteric and orthosteric sites[56]. Methoctramine is thus a bitopic dualsteric antagonist.

# 6. Bitopic ligands

The idea of utilizing high efficacy of orthosteric ligands and subtype diversity of the allosteric binding sites led to the development of bitopic ligands that span both sites and interact with them concurrently. In principle, a bitopic ligand consists of two molecules, one targeting the orthosteric site, and the other interacting with the allosteric site. The two parts are connected by a linker of a proper length. Combination of the nonselective muscarinic agonist iperoxo and its M<sub>1</sub>-selective PAM BQCA led to a set of M<sub>1</sub>-selective agonists with graded efficacy that was dependent on the chemical structure of the linker[57]. The orientation of the orthosteric moiety within the orthosteric binding site is crucial for subtype selectivity[58]. Therefore, the structure of the linker affects not only efficacy but also selectivity of bitopic ligands. Other examples of bitopic agonists that gain their selectivity from interaction with the common allosteric binding site are 77-LH-28-1 (1-[3- (4-butyl-1piperidinyl)propyl]-3,4-dihydro-2(1H)-quinolinone) and **TBPB** (1-(1'-(2-methylbenzyl)-1,4'bipiperidin-4-yl)-1H-benzo[d]imidazol-2(3H)-one)[59,60]. Mutations of the allosteric and orthosteric binding sites revealed that NDMC (N-desmethylclozapine) that was considered an allosteric agonist and McN-A-343 (4-(m-chlorophenyl-carbamoyloxy)-2-butynyltri-methylammonium) that was considered an orthosteric agonist are in fact bitopic ligands[61,62]. Allosteric effects of the bitopic agonists McN-A-343 and NDMC and the allosteric agonists AC-42 and 77-LH-28-1 are mediated via Y177 in the o2 loop of the M2 receptor[61]. Taken together, these bitopic ligands also interact with the center 1 (Figure 3) of the common allosteric binding site (Figure 2).

Further development of bitopic ligands led to the discovery of the photo-switchable ligands[63]. In case of a photo-switchable ligand, the conformation of the linker is sensitive to exposure to light of a specific wavelength, which leads to isomerization of the liker and change in the mutual orientation of orthosteric and allosteric moieties (Figure 4). In one linker conformation, the orthosteric and allosteric moieties are in an orientation that does not allow simultaneous interaction of individual moieties with their respective binding sites that makes a ligand inactive. After linker isomerization, the orientation of individual moieties allows simultaneous interaction of individual moieties with their respective binding sites and the ligand becomes active. Photo-switchable ligands are an invaluable tool in basic research since they allow an experimenter to apply ligand at the desired place and desired time in the blink of a laser.



**Figure 4.** Scheme of isomerization of photo-switchable ligand consisting of the orthosteric (O) and allosteric (A) moiety connected by the linker.

#### 7. Novel allosteric modulators

Although positive allosteric modulation of acetylcholine was presented as proof of concept in the '90s[64,65], it took more than 10 years to develop first PAMs with physiologically active properties suitable for further drug development[66–69]. However, progress of research on muscarinic allosteric modulators accelerated and selective PAMs for each subtype were discovered during the last decade.

Namely, two new classes of M<sub>1</sub>-selective PAMs were developed. Derivates of heterocyclic carboxamides display various degrees of intrinsic activity and various modulatory profiles of affinity and efficacy of acetylcholine[70]. Similarly, derivates of 4-phenylpyridin-2-one display a diverse range of activities ranging from pure PAMs to pure allosteric agonists[71].

The availability of the M<sub>2</sub> receptor crystal structure in an inactive[2], as well as active conformation[4] and advances in computer modeling, allowed for acceleration of structure-based design of chemically diverse allosteric modulators of this receptor[72]. The combination of *in silico* simulation of molecular dynamics and virtual screening led to the identification of allosteric

modulators of the M<sub>2</sub> receptor of chemically novel structures that were verified as NAMs and PAMs of super agonist iperoxo in binding experiments.

Positive allosteric modulators of antagonists may turn otherwise a non-selective orthosteric antagonist to the subtype-selective antagonist with therapeutic potential. Recently, as a proof of concept M<sub>2</sub>-selective positive modulators of orthosteric antagonists were developed[73]. In this study, triazolo-quinazolinone analogues were identified as M<sub>2</sub> selective positive modulators of NMS by docking a large library of molecules to the allosteric binding site of the M<sub>2</sub> receptor in an inactive conformation. Resulting allosteric modulators increased NMS affinity about 5-times and slowed NMS dissociation from the M<sub>2</sub> receptor about 50-fold while having no such effects at the other subtypes of muscarinic receptors.

Testing of novel muscarinic allosteric modulators in animal models of human diseases showed their good efficacy. Cholinergic neurons regulate glutamatergic neurons in the striatum that regulate selection and decision-making behavior[74]. The M<sub>1</sub>-selective PAM of acetylcholine BQCA improved efficacy of the antipsychotics haloperidol, clozapine and aripiprazole in the glutamatergic deficit mouse model of behavior[75]. M<sub>1</sub> receptors play a critical role in cognitive processes[7]. Thus, positive allosteric modulation of M<sub>1</sub> receptors appears to be the way to treat cognitive deficits in Alzheimer's disease or schizophrenia[76]. However, a subgroup of patients with schizophrenia showed decreased responsiveness to M<sub>1</sub>-selective PAM of acetylcholine BQCA that may explain why some individuals are resistant to the treatment[77].

The striatal M<sub>4</sub> receptors attenuate dopaminergic and glutamatergic neurotransmission[78]. Thus, selective positive modulation of M<sub>4</sub> receptors may provide a novel treatment strategy of psychotic symptoms in schizophrenia that are considered to result from dopaminergic hyperactivity[76]. A series of M<sub>4</sub> PAMs was found to be centrally active and efficient in an animal model of schizophrenia[79]. Huntington's disease results from increased transmission at glutamatergic corticostriatal synapses that may be attenuated by the cholinergic system. In accordance, M<sub>4</sub> PAMs improved behavioral symptoms in the mouse model of Huntington disease[69].

M₅ muscarinic receptors are expressed solely in the substantia nigra and ventral tegmental area (VTA)[7]. Cholinergic neurons in the nucleus accumbens (NAcc) project to VTA where they stimulate dopaminergic neurons. The VTA dopaminergic neurons project back to NAcc where they stimulate cholinergic neurons. This positive feedback works as the reward circuit[80]. A blockade or negative allosteric modulation of M₅ receptors could break the reward cycle and prevent addiction. The M₅-selective NAM of acetylcholine (ML375) ((S)-9b-(4-chlorophenyl)-1-(3,4-difluorobenzoyl)-2,3-dihydro-1H-imidazo[2,1-a]isoindol-5(9bH)-one)[81] effectively prevented cocaine self-administration and development of addiction in rats[82].

In type 2 diabetes, pancreatic  $\beta$  cells are unable to release sufficient amounts of insulin to maintain physiological blood glucose levels. Parasympathetic activation of pancreatic  $\beta$  cells by acetylcholine stimulates the release of insulin[83]. Effects of acetylcholine on insulin secretion are mediated by the  $M_3$  receptor[84]. In accordance,  $M_3$  PAM of acetylcholine VU0119498 (1-(4-bromobenzyl)indole-2,3-dione) enhanced glucose-stimulated insulin secretion and greatly improved glucose tolerance in lean and obese glucose-intolerant mice[85]. These effects were absent from mice lacking  $M_3$  receptors in their  $\beta$  cells.

# 7. Perspectives

Recent discoveries of selective positive allosteric modulators of acetylcholine with therapeutic potential in the treatment of psychiatric and neurologic disorders like Alzheimer's or schizophrenia are encouraging. Research on muscarinic allosteric modulators has surpassed the stage of optimization of selectivity, efficacy and bioavailability and advanced to a translational stage of medicinal research. In the field of basic research, further progress is mainly achieved by the introduction of novel concepts and techniques. Novel photo-switchable ligands may greatly increase accuracy of timely delivery as well as allow varied permutations of experimental design. Thus, focus on the design of novel photo-switchable ligands is currently gaining momentum.

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**Conflicts of Interest:** The authors declare no conflict of interest

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