

Review

Role of the Serotonin Receptor 7 in Brain Plasticity: From Development to Disease

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Abstract

Our knowledge on the plastic functions of the serotonin (5-HT) receptor subtype 7 (5-HT7R) in the brain physiology and pathology considerably advanced in the last few years. A wealth of data show that the 5-HT7R is a key player in the establishment and remodeling of neuronal cytoarchitecture during development and in the mature brain, and its dysfunction is linked to neuropsychiatric and neurodevelopmental diseases. The involvement of this receptor in synaptic plasticity is further demonstrated by data showing that its activation allows to rescue long term potentiation (LTP) and long term depression (LTD) deficits in various animal models of neurodevelopmental diseases. In addition, it is becoming clear that the 5-HT7R is involved in inflammatory intestinal diseases, possibly playing a role in the gut-brain axis, and modulates the function of immune cells. In this review, we will mainly focus on recent findings on this receptor's role in the structural and synaptic plasticity of the mammalian brain, although we will also illustrate novel aspects highlighted in gut and immune system.

Keywords: brain connectivity; brain development; gut-brain axis; neurodevelopmental diseases; neuronal cytoarchitecture; neuroplasticity; regulatory T cells; serotonin (5-HT).

1. Serotonin overview

1.1. Serotonin metabolism

Brain 5-HT is a neurotransmitter playing a key role in modulating neuronal circuit development and activities. The serotonergic neurons, through their extensive axonal network, are able to reach and influence nearly all Central Nervous System (CNS) areas. As a consequence, 5-HT regulates a plethora of functions such as sleep and circadian rhythms, mood, memory and reward, emotional behavior, nociception and sensory processing, autonomic responses, and motor activity [1].

Our current understanding of the development, evolution, and function of 5-HT neurotransmission derives from different model organisms, spanning from invertebrates to vertebrates [2]. It is noteworthy that in all species the serotonergic network is highly plastic, showing changes in its anatomical organization all through the life of the organisms.

5-HT metabolic pathways, reuptake, and degradation are broadly conserved among multicellular organisms [2]. 5-HT is synthesized from the amino acid tryptophan, which is an essential dietary supplement. Tryptophan is hydroxylated to 5-hydroxytryptophan (5-HTP) by the tryptophan-hydroxylase (TPH), the rate limiting enzyme for 5-HT biosynthesis. 5-HTP, in turn, is converted in 5-HT by the aromatic L-amino acid decarboxylase. The enzyme TPH has two distinct isoforms encoded by two genes: the *Tph1* is expressed in peripheral tissues and pineal gland, while the *Tph2* is selectively expressed in the CNS and in the enteric neurons of the gut [3]. Studies on TPH-knockout (KO) mice confirmed that the synthesis of 5-HT in the brain is driven by TPH2, whereas the synthesis of 5-HT in peripheral organs is driven by TPH1 [4]. Since 5-HT is unable to cross the blood-brain barrier, at least in the adult life, the central and the peripheral serotonergic systems are independently

regulated. The synaptic effects of 5-HT are mainly terminated by its reuptake into 5-HT nerve terminals by the 5-HT transporter.

1.2. Role of serotonin in morphological remodeling of CNS circuits

In the mammalian brain, 5-HT neurons are among the earliest neurons to be specified during development [5]. They are located in the hindbrain and are grouped in 9 raphe nuclei, designated as B1-B9 [6]. Although they are relatively few (about 30,000 in the mouse and 300,000 in humans), they give rise to extensive rostral and caudal axonal projections to the entire CNS, representing the most widely distributed neuronal network in the brain [7].

In addition to its well-established role as a neurotransmitter, 5-HT exerts morphogenic actions on the brain, influencing several neurodevelopmental processes such as neurogenesis, cell migration, axon guidance, dendritogenesis, synaptogenesis and brain wiring [8].

Besides the endogenous 5-HT, the brain of the fetus receives it also from the placenta of the mother. Thus, the placenta represents a crucial micro-environment during neurodevelopment, orchestrating a series of complex maternal–fetal interactions. The contribution of this interplay is essential for the correct development of the CNS and for long-term brain functions [9]. Therefore, maternal insults to placental microenvironment may alter embryonic brain development, resulting in prenatal priming of neuro-developmental disorders [10]. For instance, in mice it has been shown that maternal inflammation results in an upregulation of tryptophan conversion to 5-HT within the placenta, leading to altered serotonergic axonal growth in the fetal forebrain. These results indicate that the level of 5-HT during embryogenesis is critical for proper brain circuit wiring and open a new perspective for understanding the early origins of neurodevelopmental disorders [11–13].

The importance of a correct 5-HT level in the brain has been demonstrated by numerous studies on mice models. When the genes involved in 5-HT uptake or degradation are knocked out, the increased 5-HT levels in the brain leads to altered topographical development of the somatosensory cortex and incorrect cortical interneuron migration [14–16]. On the other hand, the transient disruption of 5-HT signaling, during a restricted period of pre- or postnatal development, using pharmacological (Selective Serotonin Reuptake Inhibitor exposure) animal models, leads to long-term behavioral abnormalities, such as increased anxiety in adulthood [17,18]. These animals do not show gross morphological alterations in the CNS suggesting that the lack of cerebral 5-HT may only affect the fine tuning of specific serotonergic circuits. This hypothesis has been recently confirmed using a mouse model in which the enhanced green fluorescent protein is knocked into the *Tph2* locus, resulting in lack of brain 5-HT, and allowing the detection of serotonergic system through enhanced fluorescence, independently of 5-HT immunoreactivity. In these mice, the serotonergic innervation was apparently normal in cortex and striatum. On the other hand, mutant adult mice showed a dramatic reduction of serotonergic axon terminal arborization in the diencephalic areas, and a marked serotonergic hyperinnervation in the nucleus accumbens and in the hippocampus [19]. These results demonstrate that brain 5-HT plays a key role in regulating the wiring of the serotonergic system during brain development. Interestingly, transient silencing of 5-HT transporter expression in neonatal thalamic neurons affects somatosensory barrel architecture by selective alteration of dendritic structure and trajectory of late postnatal interneuron development in the mouse cortex [20]. Altogether, these findings indicate that perturbing 5-HT levels during critical periods of early development influences later neuronal development through alteration of CNS connectivity that may persist into the adulthood [13,21–23]. Interestingly, recent evidence demonstrated that changes in 5-HT homeostasis affect axonal branch complexity not only during development but also in the adult life [24]. In adult *TPH2*-conditional KO mice it was shown that the administration of the serotonin precursor 5-hydroxytryptophan was able to re-establish the 5-HT signaling and to rescue defects in serotonergic system organization [24].

Interestingly, in recent elegant experiments that combined chemogenetics and fMRI, it was demonstrated that, in adult mice, endogenous stimulation of 5-HT-producing neurons does not affect global brain activity but selectively activates specific cortical and subcortical areas. By contrast,

pharmacological increase of 5-HT levels determined widespread fMRI deactivation, possibly reflecting the mixed contribution of central and perivascular constrictive effects [25].

On the whole, findings from genetic mouse models confirm that the level of 5-HT during brain ontogeny is critical for proper CNS circuit wiring and suggest that alterations in 5-HT signaling during brain development have profound implications for behavior and mental health across the life span. Indeed, a plethora of genetic and pharmacological studies have linked defects of brain 5-HT signaling with psychiatric and neurodevelopmental disorders, such as major depression, anxiety, schizophrenia, obsessive compulsive disorder and Autism Spectrum Disorders (ASD) [13,26,27]. In addition, it is becoming increasingly clear that 5-HT has a crucial role also in the maintenance of mature neuronal circuitry in the brain, opening novel perspectives in rescuing defects of CNS connectivity in the adult. For instance, the potential of 5-HT neurons to remodel their morphology during the entire life is indicated by the well-known capability of 5-HT axons of the adult to regenerate and sprout after lesions [23,28].

However, understanding the cellular and molecular mechanisms underlying the effects of 5-HT during brain development, maintenance and dysfunction is challenging, in part due to the existence of at least 14 subtypes of receptors (5-HTRs) grouped in 7 distinct classes (from 5-HT1R to 5-HT7R). All 5-HT receptors are broadly distributed in the brain where they display a highly dynamic developmental and region-selective expression pattern and trigger different signaling pathways. The 5-HT receptors are typical G-protein-coupled-receptors with seven transmembrane domains, with the exception of the 5-HT3 receptor, that is a ligand-gated ion channel [29].

2. Role of the 5-HT7R in shaping neuronal circuits

2.1. The 5-HT7R

The 5-HT7R is the last discovered member of the 5-HTR family [30-32], and its gene has been cloned in invertebrates and vertebrates. Recently, several lines of investigation on this receptor have deepened the knowledge on its crucial role in a wide range of physiological functions, both in the mammalian CNS and in peripheral organs. For instance, recently two independent research groups demonstrated that the microRNA (miR)-29a is able to modulate 5-HT7R expression in the brain, as well as in the gut [33,34].

In the mammalian CNS, the 5-HT7R is mainly expressed in spinal cord, thalamus, hypothalamus, hippocampus, prefrontal cortex, striatal complex, amygdala and in the Purkinje neurons of the cerebellum [35,36]. This wide distribution reflects the numerous functions in which the receptor is involved, such as circadian rhythms, sleep-wake cycle, thermoregulation, learning and memory processing, and nociception [37].

It is noteworthy that, in mammals, this receptor exhibits a number of functional splice variants due to the presence of introns in the 5-HT7R gene [38].

The 5-HT7R is a G protein-coupled receptor, that can activate $G_{\alpha s}$ and stimulate adenylate cyclase, leading to an increase in cyclic adenosine monophosphate (cAMP). The latter activates protein kinase A (PKA) leading to phosphorylation of various proteins such as the Mitogen-Activated Protein Kinase and Extracellular signal-Regulated Kinases (ERK), [39,40]. 5-HT7R signaling involves also changes in intracellular Ca^{2+} concentration and Ca^{2+} /calmodulin pathways [41,42], as well as PKA independent mechanisms which include EPAC (exchange protein directly activated by cAMP) signaling [43].

Furthermore, the 5-HT7R can interact with another member of the G protein family, the $G_{\alpha 12}$. Activation of this signaling pathway in fibroblasts and neurons leads to morphological modifications through stimulation of Rho GTPases, Cdc42 and RhoA, a class of intracellular signaling proteins critical for the regulation of cytoskeleton organization [44].

5-HT receptors signaling has been recently shown to depend also on their oligomerization. In particular the 5-HT7R can form homodimers, as well as heterodimers with 5-HT1AR [45]. The latter, when is in a monomeric conformation, causes a decrease in cAMP concentration through activation

of the Gi. Heterodimerization with 5-HT7R inhibits the 5-HT1AR cAMP signaling pathway, while homodimerization of both receptors do not influence the respective cAMP pathways. These findings suggest that oligomerization of G-protein-coupled-receptors may have profound functional consequences on the downstream signaling, thus triggering cellular and developmental-specific regulatory effects.

2.2. Role of the 5-HT7R in shaping neuronal circuits during development

The influence of the 5-HT7R on neuronal morphology has stimulated interest in studying its potential role in the establishment and maintenance of brain connectivity and in synaptic plasticity. The availability of selective agonists and antagonists, as well as that of genetically modified mice lacking the 5-HT7R, has shed light on the physio-pathological role of this receptor [40,46,47]. By using rodents primary cultures of hippocampal neurons and various 5-HT7R agonists in combination with selective antagonists, it was consistently shown that pharmacological stimulation of the endogenous 5-HT7R promotes pronounced extension of neurite length [44,48,49]. The morphogenic effects of 5-HT7R stimulation have been demonstrated also in cultured neurons from additional embryonic forebrain areas, such as the striatum and the cortex [50,51]. Neurite elongation was shown to rely on *de novo* protein synthesis and multiple signaling systems, such as ERK, Cdk5, the RhoGTPase Cdc42 and mTOR. These pathways converge to promote reorganization of the neuronal cytoskeleton through qualitative and quantitative changes of selected proteins, such as microtubule-associated proteins and cofilin [50,52]. In hippocampal neurons has been demonstrated that 5-HT7R finely modulates NMDA receptors activity [53,54]. Furthermore 5-HT7R activation increases phosphorylation of the GluA1 AMPA receptor subunit and AMPA receptor-mediated neurotransmission in the hippocampus [55,56]. Consistent with these findings, 5-HT7R-KO mice display reduced LTP in the hippocampus [57].

Chronic stimulation of the 5-HT7R/G α_{12} signaling pathway promotes dendritic spine formation, enhances the basal neuronal excitability and modulates LTP in organotypic slices preparation from the hippocampus of juvenile mice. Interestingly, 5-HT7R stimulation does not affect neuronal morphology, synaptogenesis, and synaptic plasticity in hippocampal slices from adult animals, probably due to decreased hippocampal expression of the 5-HT7R during later postnatal stages [58]. It has been recently hypothesized that this decline could be due to the simultaneous upregulation of the miR-29a in the developing hippocampus. Indeed 5-HT7R mRNA is downregulated by the miR-29a in cultured hippocampal neurons and miR-29a overexpression impairs the 5-HT7R-dependent neurite elongation [33].

Neuronal remodeling is highly influenced by the extracellular matrix. Accordingly, it has been shown that the physical interaction between the 5-HT7R and the hyaluronan receptor CD44, a main component of the extracellular matrix, plays a crucial role in synaptic remodeling. Briefly, stimulation of the 5-HT7R increases the activity of the metalloproteinase MMP-9, which, in turn, cleaves the extracellular domain of CD44. This signaling cascade promotes detachment from the extracellular matrix, thus triggering dendritic spine elongation in mouse hippocampal neurons [59].

In accordance with the influence of the 5-HT7R signaling pathways in remodeling developing forebrain neuron morphology, it was shown that prolonged stimulation of this receptor and the downstream activation of Cdk5 and Cdc42 increased the density of filopodia-like dendritic spines and synaptogenesis in cultured striatal and cortical neurons [60]. The crucial role of 5-HT7R in shaping developing synapses was confirmed by pharmacological inactivation of the receptor as well as by analysis of early postnatal neurons isolated from 5-HT7R-deficient mice. It is noteworthy that, when 5-HT7R was blocked pharmacologically, and in 5-HT7R-KO neurons, the number of dendritic spines decreased, suggesting that constitutive receptor activity is critically involved in dendritic spinogenesis. From this point of view, a detailed analysis of dendritic spine shape and density in the brain of 5-HT7R-KO mice at various ages would be crucial to assess the physiological effects of this receptor on neuronal cytoarchitecture.

The involvement of 5-HT7R in spinogenesis and synaptogenesis together with the demonstration that its activation is able to stimulate protein synthesis-dependent neurite elongation, as well as axonal elongation [50,52], suggests the intriguing possibility that the activation of this receptor may be linked to the axonal and synaptic system of protein synthesis. The local system of protein synthesis has been demonstrated to play a crucial role in synaptic plasticity, although its regulatory mechanisms are only partially understood [61-63], and 5-HT7R and its related pathways are good candidates to be part of this system.

2.2 Role of the 5-HT7R in remodeling neuronal circuits in the adult

Neuronal circuits remain able to reorganize in response to experience well into adulthood, continuing to exhibit robust plasticity along the entire life [64]. Consistently, the action of 5-HT7R on the modulation of neuronal plasticity is not restricted to embryonic and early postnatal development, but can also occur in later developmental stages and in adulthood.

Interestingly, it was shown that selective pharmacological stimulation of 5-HT7R during adolescence determines its persistent upregulation in adult rat forebrain areas [65]. Likewise, it has been hypothesized that 5-HT7R may underlie the persistent structural rearrangements of the brain reward pathways occurring during postnatal development, following exposure to methylphenidate, the elective drug for the treatment of Attention Deficit Hyperactivity Disorder [66]. Accordingly, stimulation of the 5-HT7R in adolescent rats leads to increased dendritic arborization in the nucleus accumbens, a limbic area involved in reward, as well as increased functional connectivity in different forebrain networks likely to be involved in anxiety-related behavior [67]. Changes in dendritic spine formation, turnover and shape occur during the entire life span in response to stimuli that trigger long-term alterations in synaptic efficacy, such as LTP and LTD [68-70]. Consistently, it has been shown that activation of 5-HT7R in hippocampal slices from wild type mice (as well as in Fragile X Syndrome mice, see next paragraph) reverses LTD mediated by metabotropic glutamate receptors (mGluR-LTD), a form of plasticity playing a crucial role in cognition and in behavioral flexibility [55]. Moreover, acute *in vivo* administration of a selective 5-HT7R agonist improved cognitive performance in mice [71]. These results are consistent with the hypothesis that long term changes of synaptic plasticity, which are a substrate of learning and memory formation, leads to neural network rewiring. Accordingly, the 5-HT7R-KO mice exhibit reduced hippocampal LTP, and specific impairments in contextual learning, seeking behavior and allocentric spatial memory [57,72].

Interestingly, the expression level of 5-HT7R in the hippocampal CA3 region, an area of the brain involved in allocentric navigation, decreases with age [73], suggesting that the spatial memory deficits associated with aging could be attributed to decreased 5-HT7R activity in this region of the brain. Conversely, another group reported that hippocampal expression of 5-HT7R does not change with age but exhibits 24 h rhythms [74]. This observation should be taken into account in the interpretation of previous findings, as well as in planning future experiments. Several other studies have produced contradictory results related to the involvement of 5-HT7R in memory and attention-related processes [75-77], probably due to experimental differences (animal strain, behavioral tests, compounds and doses, route of administration etc.). In conclusion, although the role of this receptor on cognitive functions needs to be fully elucidated, it is clear that it modulates various aspects of learning and memory processes.

Interestingly, the 5-HT7R is also involved in bidirectional modulation of cerebellar synaptic plasticity, since its activation induces LTD at the parallel fiber-Purkinje cell synapse, whereas it blocks LTP induced by parallel fiber stimulation [36]. These results suggest that the receptor might be involved in motor learning, a cognitive function depending on cerebellar circuits activity [78].

Altogether, these findings strongly suggest that the 5-HT7R plays a role in modulating synaptic plasticity and neuronal connectivity in both developing and mature brain circuits, although the molecular and cellular mechanisms underlying this modulation are only partially understood.

3. The 5-HT7R and neurological diseases

Numerous brain disorders, such as ASD, cognitive and mood dysfunctions, schizophrenia, depression, anxiety, impulsivity, epilepsy, migraine and neuropathic pain show altered 5-HT_{7R}-mediated signaling [37,79]. The potential involvement of 5-HT_{7R} in most of these diseases was discovered studying the effects of a broad range of antidepressant and antipsychotic drugs that interact with the receptor, displaying high affinity [40]. Recently, the importance of 5-HT_{7R} modulation was brought to the attention of psychiatric and pharmacological communities, since a novel very effective and atypical mood-stabilizing antipsychotic drug, lurasidone, predominantly blocks this receptor. This drug also acts as agonist of the 5-HT_{1A}R. Experiments on animal models indicate that chronic treatment with lurasidone enhances 5-HT transmission in dorsal raphe nuclei by coordinated 5-HT_{1A}R agonism and 5-HT_{7R} antagonism through modulation of GABAergic and glutamatergic pathways, thus contributing to the augmentation of drug's antidepressive effects [80,81].

In line with the possible involvement of 5-HT_{7R} in the mechanisms of action of antidepressants, genome-wide association studies in humans have suggested a relationship between 5-HT_{7R} genetic polymorphisms and schizophrenia [82]. Likewise, a very recent work showed that one single nucleotide polymorphisms located in the promoter region of the 5-HT_{7R} gene is associated with better response to two antidepressants, paroxetine and fluoxetine, that are selective 5-HT reuptake inhibitors. These data provide novel pharmacogenomic evidence to support the role of 5-HT_{7R} in antidepressant response [83].

However, pharmacological and genetic manipulation of 5-HT_{7R} in animal models of depression, anxiety and schizophrenia has often given inconsistent or conflicting results. Experimental differences (for instance animals' strain, behavioral tests, drugs and their doses, route of administration), as well as the use of non-selective drugs targeting other receptors in addition to the 5-HT_{7R}, might account for these mixed results. In addition, the interpretation of behavioral data from 5-HT_{7R}-KO mice is complicated by the indirect effects of the missing gene, such as changes in developmental processes or dysregulation of compensatory genes and pathways [84]. Very recently, mixed outcomes on various behavioral assays for anxiety, depression and psychosis, performed on mice treated with two selective 5-HT_{7R} antagonists [85], and on 5-HT_{7R}-KO mice [86], raised doubts on the role played by the receptor in these neuropsychiatric diseases. Ultimately, the available data suggest that additional research will be required to further evaluate and dissect the contribution of this receptor in anxiety/depression and schizophrenia, and its potential involvement for the treatment of these neuropsychiatric diseases.

Conversely, compelling evidence strongly suggest that the 5-HT_{7R} is involved in neurodevelopmental disorders characterized by intellectual disabilities and cognitive impairment, such as ASD, Rett syndrome (RTT) and Fragile X syndrome (FXS).

ASD includes a group of neurodevelopmental disorders characterized by impaired social interaction and communication, repetitive and stereotyped behaviors, often accompanied by intellectual disability [87]. Growing evidence indicate that the brain 5-HT neurotransmission system is altered in ASD patients, and in various animal models of the disease [88-90]. For instance, mice lacking brain 5-HT, in addition to several abnormal phenotypes (growth retardation, high aggressive behavior, maternal neglect), show selective deficits resembling ASD's symptoms, including impairments in social interactions and repetitive behavior [3,4]. Various pharmacological studies are providing evidence that targeting 5-HT_{7R} has the potential to treat the core symptoms of ASD and associated intellectual disabilities [91-94]. Recent evidence in animal models suggest that, among other subtypes, the 5-HT_{7R} might be one of the players involved in ASD (see below). In line with this hypothesis, the only two drugs that to date are approved for the treatment of behavioral manifestations of ASD, risperidone and aripiprazole, are 5-HT_{7R} antagonists [95], although their efficacy may be attributed also to their interactions with other receptors. Indeed, none of the approved ASD drugs are highly selective for 5-HT_{7R}, hampering our understanding of its potential as a target for pharmacological treatment of ASD in humans. Nevertheless, brain-permeant and selective agonists of the 5-HT_{7R},

have been successfully employed to rescue ASD dysfunctions in animal models of FXS, RTT and CDKL5 Deficiency (CDD).

FXS mice exhibit cognitive impairment and stereotyped behavior, accompanied by altered morphology and density of dendritic spines in the forebrain, and synapse malfunctioning in the hippocampus, with abnormal enhancement of mGluR-LTD. The activation of 5-HT₇R by a selective brain-permeant agonist in hippocampal slices from FXS mice is able to correct excessive mGluR-LTD through activation of cAMP/PKA pathway, bringing it back to its physiological level and thereby restoring synaptic plasticity. Noteworthy, acute *in vivo* administration of the agonist rescues learning and autistic-like behavior in 3/4 months-old FXS mice [71].

Beneficial effects of the same agonist, chronically administered, were also observed in adult mouse models of RTT. This syndrome is a severe X-linked neurological disorder characterized by deficits in autonomic, cognitive, motor functions and autistic features. *In vivo* systemic repeated stimulation of 5-HT₇R with a selective brain-permeant agonist was able to improve cognitive and motor coordination deficits, as well as spatial memory and synaptic plasticity in RTT mice. 5-HT₇R stimulation also restored the normal level of key molecules regulating actin cytoskeleton dynamics, such as Rho GTPases and mTOR signaling pathways that showed altered expression levels in the hippocampus of RTT mice [96,97]. The 5-HT₇R-mediated neurobehavioral and molecular changes were still present 2 months after the last injection, suggesting long-lasting beneficial effects on RTT-related impairments. Subsequent studies uncovered functional alterations of brain mitochondria in RTT mouse models, that were rescued by chronic pharmacological stimulation of the 5-HT₇R [98,99]. Similar promising preclinical results have been recently obtained in a mouse model of CDD, a rare neurodevelopmental syndrome characterized by severe neurobehavioral and motor deficits and stereotyped movements [100].

Altogether, the above findings provide compelling evidence that 5-HT₇R stimulation exerts a widespread beneficial effect on behavioral and molecular symptomatology in various mouse models of neurodevelopmental disorders, in particular ASD. Moreover, these results have important therapeutic implications, indicating that it is possible to reverse severe behavioral and molecular deficits in the animal models by pharmacological treatment at adult age. Intriguingly, all these diseases are accompanied by alteration of dendritic spines in forebrain areas involved in higher cognitive functions, suggesting altered connectivity. Although data on the 5-HT₇R-dependent remodeling of dendritic spines in the ASD animal models are still missing, it is possible to hypothesize that activation of 5-HT₇R may promote structural rearrangements of neural circuits also in adult brain, that in turn might underlie the rescue of long term synaptic plasticity.

4. The 5-HT₇R in the gut and in the immune system

Despite the vast repertoire of neurodevelopmental, behavioral and cognitive processes modulated by the brain 5-HT, only 5% of the total body content of 5-HT is located in the CNS, while the remaining part is synthesized and stored in peripheral tissues.

Outside the CNS, 5-HT is found in gastrointestinal epithelium, where it is mainly produced by enterochromaffin (EC) cells and only in small quantity (5%) by neurons of the Enteric Nervous System and by the resident gut microbiota. 5-HT released by EC cells is actively taken up and stored by blood platelets, and released upon their activation, modifying vascular smooth muscle tone and a variety of other functions controlled by peripheral organs [101]. 5-HT is also synthesized and released by cells of the immune system, expanding the range of tissues involved in its signaling [102]. Peripheral 5-HT₇R expression roughly mirrors peripheral 5-HT distribution, since it has been observed in the gastrointestinal tract, as well as in peripheral organs (kidney, liver, pancreas, spleen, and stomach), and in the cells of the immune system (lymphoid progenitor cells, mast cells, monocytes, macrophages, dendritic cells and T lymphocytes) [102,103].

Here, we briefly review recent studies showing that 5-HT₇R plays a crucial role in generation/perpetuation of intestinal inflammation, and in immune cell activation, suggesting its possible involvement in the bidirectional communication between the brain and gut. As a key

element of this axis, 5-HT signaling may link emotional and cognitive areas of the brain with peripheral gut activity. Accordingly, recent findings suggest that alteration of this two-way serotonergic system of communication between brain and gut may play a role in the pathogenesis of various diseases, including ASD [104]. Moreover, it is becoming increasingly evident that the resident gut microbiota, that produce tryptophan and 5-HT, is a critical component of the gut-brain communication, modulating brain development and behavioral responses [105,106]. The 5-HT effects in this complex microbiota-gut-brain communication are mediated by 5-HT receptors and, among other subtypes, the 5-HT₇R is a very interesting candidate, being expressed both in the gut and in the brain.

Various findings suggest that 5-HT₇R may have a crucial role in the pathogenesis of inflammatory disorders affecting the gastrointestinal tract, such as ulcerative colitis and Crohn's Disease. For instance, genetic or pharmacological silencing of 5-HT₇R in mouse models of ulcerative colitis reduced the severity of intestinal inflammation and decreased the production of inflammatory markers by gastrointestinal dendritic cells. These antigen-presenting cells initiate adaptive immune responses upon inflammation [107]. These results indicating that 5-HT₇R inhibition reduces inflammation symptoms in gut inflammatory disorders, are in contrast with other findings. Indeed, Guseva et al. [108] reported that pharmacological blockade or genetic ablation of 5-HT₇R resulted in increased severity of both acute and chronic mouse models of colitis, whereas receptor stimulation produced an anti-inflammatory effect. In addition, expression of 5-HT₇R significantly increased after induction of colitis in mice and in inflamed intestinal dendritic cells of patients with Crohn's disease. Novel epigenetic mechanisms regulating 5-HT₇R expression have been recently highlighted by studies on animal models and patients with Irritable Bowel Syndrome (IBS), a functional gastrointestinal disorder often associated to visceral hyperalgesia without inflammatory processes. It was shown that miR-29a modulates visceral hypersensitivity in a mouse model of IBS by directly targeting the 5-HT₇R and downregulating its expression. The authors found that intestinal tissues from mice and patients with IBS displayed increased levels of miR-29a and reduced levels of 5-HT₇R. Consistently, in mice with IBS, when miR-29a was knocked-out, 5-HT₇R was overexpressed and intestinal hyperalgesia was attenuated [34]. These findings suggest that colon hypersensitivity may be mediated by the endogenous interaction between miRNA-29a and 5-HT₇R, offering a potential promising therapeutic approach for reversing abdominal pain in IBS patients.

The discrepancies on the role of 5-HT₇R in gut disorders may depend on the notable differences in animal models and experimental design. In addition, it is possible that immune cells are differentially recruited depending on the experimental models of induced intestinal damage and human gut diseases, and that their response/activation is modulated by the level of the 5-HT₇R expression. Indeed, it has been demonstrated that the 5-HT₇R–Cdc42-mediated signaling regulates dendritic cell morphology and enhances chemotactic motility [109]. Likewise, a prominent role of 5HT₇R in regulating endothelial cell migration has been identified, suggesting that this receptor is a potential modulator of physiological and pathophysiological processes involving cell migration, adhesion [110] and inflammatory fibrotic infiltration [103,111]. Thus, 5-HT₇R-dependent activation and migration of dendritic cells might be significantly different in various intestinal inflammatory diseases, accounting - at least in part - for conflicting results on the role of 5-HT₇R in the pathogenesis of gut inflammatory disorders. As a conclusion, additional studies are required to precisely understand 5-HT₇R function and dysfunction in the intestine.

Nevertheless, these findings highlight the potential involvement of the 5-HT₇R in the physiology and pathology of the immune system. Indeed, it is well known that 5-HT plays a key role in inflammation and immunity by acting on 5-HT receptors that are differentially expressed in immune cells, both in rodents and humans. 5-HT₇R expression has been detected in lymphoid progenitor cells, mast cells, monocytes, macrophages, dendritic cells and T lymphocytes [102,103]. This receptor is also expressed by microglia, the brain resident macrophages, and its stimulation in human microglial cell lines leads to increased IL-6 expression [112]. Stimulation of 5-HT₇R by 5-HT enhances the proliferation and activation of mouse naive T cells through ERK signaling [113]. Likewise, in human macrophages, the anti-inflammatory and pro-fibrotic activity of 5-HT is primarily mediated by 5-HT₇R-PKA pathway

[111]. Notably, it has been recently shown that brain regulatory T (Treg) cells are distinct from those of other tissues since they express unique genes related to the Nervous System, including the 5-HT7R. The specific features and functions of brain Treg cells are poorly understood because their number is very low in the brain under normal conditions. Conversely, a large number of Treg cells infiltrated the mouse brain during the chronic phase of ischemic stroke, suppressing astrogliosis and potentiating neurological recovery [114]. Brain Treg cells, but not splenic Treg cells, responded to 5-HT by increased proliferation and this response was blocked by a selective antagonist of the 5-HT7R. Notably, 5-HT7R-deficient Treg cells do not expand correctly into the brain and do not promote neurological recovery after ischemic stroke. These findings demonstrated that 5-HT7R play a specialized role in Treg cells and suggest that the 5-HT signaling mediated by Treg cells might represent one of the mechanisms that contribute to the cross talk between immune system and brain inflammation.

Altogether, the reported findings on the 5-HT7R signaling in the brain, gut and immune system, suggest the involvement of this receptor in the pathways underlying bidirectional communication between gut and brain possibly with the immune system as a mediator among them. For instance, as mentioned above, two very recent studies showed that 5-HT7R is a direct target of miR-29a in brain, as well as in the intestine [33,34], suggesting that this miRNA might coordinately modulate 5-HT7R expression in both tissues.

5. Conclusions and future perspectives

The modulation of 5-HT7R expression using pharmacological and genetic tools, coupled to cellular, molecular, electrophysiological and behavioral approaches, has greatly increased our knowledge on the functions of this receptor in the brain, as well as in other organs.

The results highlighted here indicate that 5-HT7R is an important player involved in the modulation of synaptic and structural plasticity in both developing and mature brain circuits. However, the detailed molecular mechanisms and signaling pathways underlying 5-HT7R morphogenic effects are still object of intense investigation. We believe that conditional KO mouse models able to silence/overexpress the 5-HT7R gene during selected age windows and in specific cell types, would help to define more accurately the contribution of the 5-HT7R to brain physiology.

Altered 5-HT7R-mediated signaling is involved in numerous brain diseases. In particular, compelling evidence indicate that 5-HT7R stimulation reverts behavioral, molecular and functional deficits in various animal models of neurodevelopmental disorders. These 5-HT7R beneficial effects, at least in RTT, might operate through rescue of the mitochondrial dysfunctions associated to the disease. Thus, it would be of great interest to deepen our understanding on the mechanisms underlying the regulatory effects of 5-HT7R on mitochondrial function.

Finally, we briefly discussed recent findings highlighting a crucial role of the 5-HT7R in intestinal inflammation and immune cell activation, and suggested its possible involvement in the complex interaction between gut, immune cells and brain.

Importantly, the findings described here open new avenues in the development of selective drugs targeting 5-HT7R as novel potential therapeutic agents in many diseases so far considered incurable.

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Abbreviations

5-HT	Serotonin
5-HT7R	Serotonin Receptor subtype 7
5-HTP	5-hydroxytryptophan
ASD	Autism Spectrum Disorders
CDD	CDKL5 Deficiency
CNS	Central Nervous System
EC	enterochromaffin
FXS	Fragile X Syndrome
IBS	Irritable Bowel Disease
KO	Knockout
LTP	Long Term Potentiation
LTD	Long Term Depression
miR	microRNA
PKA	Protein Kinase A
RTT	Rett syndrome
TPH	Tryptophan-5-hydroxylase
Treg	regulatory T cells

References

- Pilowsky, P.M. *Serotonin The Mediator that Spans Evolution* 1st ed.; Elsevier: United States of America, 2019; pp. 1-420.
- Lillesaar, C.; Gaspar, P. Serotonergic Neurons in Vertebrate and Invertebrate Model Organisms (Rodents, Zebrafish, *Drosophila melanogaster*, *Aplysiacalifornica*, *Caenorhabditis elegans*). In *Serotonin The Mediator that Spans Evolution* 1st ed.; Pilowsky, P.M.; Elsevier: United States of America, 2019; pp. 49-80.
- Pratelli, M.; Pasqualetti, M. Serotonergic neurotransmission manipulation for the understanding of brain development and function: Learning from Tph2 genetic models. *Biochimie* **2019**, *161*, 3-14.
- Mosienko, V.; Alenina, N. Life Without Brain Serotonin: Phenotypes of Animals Deficient in Central Serotonin Synthesis. In *Serotonin The Mediator that Spans Evolution* 1st ed.; Pilowsky, P.M.; Elsevier: United States of America, 2019; pp. 405-420.
- Levitt, P.; Rakic, P. The time of genesis, embryonic origin and differentiation of the brain stem monoamine neurons in the rhesus monkey. *Brain Res* **1982**, *256*, 35-57, doi:10.1016/0165-3806(82)90095-5.
- Jacobs, B.L.; Azmitia, E.C. Structure and function of the brain serotonin system. *Physiol Rev* **1992**, *72*, 165-229.
- Gagnon, D.; Parent, M. Distribution of VGLUT3 in highly collateralized axons from the rat dorsal raphe nucleus as revealed by single-neuron reconstructions. *PLoS One* **2014**, *9*, e87709.
- Wirth, A.; Holst, K.; Ponimaskin, E. How serotonin receptors regulate morphogenic signalling in neurons. *Prog Neurobiol* **2017**, *151*, 35-56.
- Bonnin, A.; Goeden, N.; Chen, K.; Wilson, M.L.; King, J.; Shih, J.C.; Blakely, R.D.; Deneris, E.S.; Levitt, P. A transient placental source of serotonin for the fetal forebrain. *Nature* **2011**, *472*, 347-350.
- Shallie, P.D.; Naicker, T. The placenta as a window to the brain: A review on the role of placental markers in prenatal programming of neurodevelopment. *Int J Dev Neurosci* **2019**, *73*, 41-49.
- Goeden, N.; Velasquez, J.; Arnold, K.A.; Chan, Y.; Lund, B.T.; Anderson, G.M.; Bonnin, A. Maternal Inflammation Disrupts Fetal Neurodevelopment via Increased Placental Output of Serotonin to the Fetal Brain. *J Neurosci* **2016**, *36*, 6041-6049.
- Brummelte, S.; Mc Glanaghy, E.; Bonnin, A.; Oberlander, T.F. Developmental changes in serotonin signaling: Implications for early brain function, behavior and adaptation. *Neuroscience* **2017**, *342*, 212-231.
- Shah, R.; Courtiol, E.; Castellanos, F.X.; Teixeira, C.M. Abnormal Serotonin Levels During Perinatal Development Lead to Behavioral Deficits in Adulthood. *Front Behav Neurosci* **2018**, *12*, 114.
- Cases, O.; Vitalis, T.; Seif, I.; De Maeyer, E.; Sotelo, C.; Gaspar, P. Lack of barrels in the somatosensory cortex of monoamine oxidase A-deficient mice: role of a serotonin excess during the critical period. *Neuron* **1996**, *16*, 297-307.
- Persico, A.M.; Mengual, E.; Moessner, R.; Hall, F.S.; Revay, R.S.; Sora, I.; Arellano, J.; DeFelipe, J.; Gimenez-Amaya, J.M.; Conciatori, M.; Marino, R.; Baldi, A.; Cabib, S.; Pascucci, T.; Uhl, G.R.; Murphy, D.L.; Lesch, K.P.; Keller, F. Barrel pattern formation requires serotonin uptake by thalamocortical afferents, and not vesicular monoamine release. *J Neurosci* **2001**, *21*, 6862-6873.

16. Riccio, O.; Potter, G.; Walzer, C.; Vallet, P.; Szabó, G.; Vutskits, L.; Kiss, J.Z.; Dayer, A.G. Excess of serotonin affects embryonic interneuron migration through activation of the serotonin receptor 6. *Mol Psychiatry* **2009**, *14*, 280-290.
17. Ansorge, M.S.; Morelli, E.; Gingrich, J.A. Inhibition of serotonin but not norepinephrine transport during development produces delayed, persistent perturbations of emotional behaviors in mice. *J Neurosci* **2008**, *28*, 199-207.
18. Oberlander, T.F.; Gingrich, J.A.; Ansorge, M.S. Sustained neurobehavioral effects of exposure to SSRI antidepressants during development: molecular to clinical evidence. *Clin Pharmacol Ther* **2009**, *86*, 672-677.
19. Migliarini, S.; Pacini, G.; Pelosi, B.; Lunardi, G.; Pasqualetti, M. Lack of brain serotonin affects postnatal development and serotonergic neuronal circuitry formation. *Mol Psychiatry* **2013**, *18*, 1106-1118.
20. De Gregorio, R.; Chen, X.; Petit, E.I.; Dobrenis, K.; Sze, J.Y. Disruption of Transient SERT Expression in Thalamic Glutamatergic Neurons Alters Trajectory of Postnatal Interneuron Development in the Mouse Cortex. *Cereb Cortex* **2019**.
21. Marín, O. Developmental timing and critical windows for the treatment of psychiatric disorders. *Nat Med* **2016**, *22*, 1229-1238.
22. Maddaloni, G.; Bertero, A.; Pratelli, M.; Barsotti, N.; Boonstra, A.; Giorgi, A.; Migliarini, S.; Pasqualetti, M. Development of Serotonergic Fibers in the Post-Natal Mouse Brain. *Front Cell Neurosci* **2017**, *11*, 202.
23. Teissier, A.; Soiza-Reilly, M.; Gaspar, P. Refining the Role of 5-HT in Postnatal Development of Brain Circuits. *Front Cell Neurosci* **2017**, *11*, 139.
24. Pratelli, M.; Migliarini, S.; Pelosi, B.; Napolitano, F.; Usiello, A.; Pasqualetti, M. Perturbation of Serotonin Homeostasis during Adulthood Affects Serotonergic Neuronal Circuitry. *eNeuro* **2017**, *4*, e0376-16.2017.
25. Giorgi, A.; Migliarini, S.; Galbusera, A.; Maddaloni, G.; Mereu, M.; Margiani, G.; Gritti, M.; Landi, S.; Trovato, F.; Bertozzi, S.M., et al. Brain-wide Mapping of Endogenous Serotonergic Transmission via Chemogenetic fMRI. *Cell Rep* **2017**, *21*, 910-918.
26. Lesch, K.P.; Waider, J. Serotonin in the modulation of neural plasticity and networks: implications for neurodevelopmental disorders. *Neuron* **2012**, *76*, 175-191.
27. Dayer, A. Serotonin-related pathways and developmental plasticity: relevance for psychiatric disorders. *Dialogues Clin Neurosci* **2014**, *16*, 29-41.
28. Deneris, E.; Gaspar, P. Serotonin neuron development: shaping molecular and structural identities. *Wiley Interdiscip Rev Dev Biol* **2018**, *7*.
29. McCorvy, J.D.; Roth, B.L. Structure and function of serotonin G protein-coupled receptors. *Pharmacol Ther* **2015**, *150*, 129-142.
30. Bard, J.A.; Zgombick, J.; Adham, N.; Vaysse, P.; Branchek, T.A.; Weinshank, R.L. Cloning of a novel human serotonin receptor (5-HT7) positively linked to adenylate cyclase. *J Biol Chem* **1993**, *268*, 23422-23426.
31. Lovenberg, T.W.; Baron, B.M.; de Lecea, L.; Miller, J.D.; Prosser, R.A.; Rea, M.A.; Foye, P.E.; Racke, M.; Slone, A.L.; Siegel, B.W. A novel adenylyl cyclase-activating serotonin receptor (5-HT7) implicated in the regulation of mammalian circadian rhythms. *Neuron* **1993**, *11*, 449-458.
32. Ruat, M.; Traiffort, E.; Leurs, R.; Tardivel-Lacombe, J.; Diaz, J.; Arrang, J.M.; Schwartz, J.C. Molecular cloning, characterization, and localization of a high-affinity serotonin receptor (5-HT7) activating cAMP formation. *Proc Natl Acad Sci U S A* **1993**, *90*, 8547-8551.
33. Volpicelli, F.; Speranza, L.; Pulcrano, S.; De Gregorio, R.; Crispino, M.; De Sanctis, C.; Leopoldo, M.; Lacivita, E.; di Porzio, U.; Bellenchi, G.C., et al. The microRNA-29a Modulates Serotonin 5-HT7 Receptor Expression and Its Effects on Hippocampal Neuronal Morphology. *Mol Neurobiol* **2019**, *56*, 8617-8627.
34. Zhu, H.; Xiao, X.; Chai, Y.; Li, D.; Yan, X.; Tang, H. MiRNA-29a modulates visceral hyperalgesia in irritable bowel syndrome by targeting HTR7. *Biochem Biophys Res Commun* **2019**, *511*, 671-678.
35. Volpicelli, F.; Speranza, L.; di Porzio, U.; Crispino, M.; Perrone-Capano, C. The serotonin receptor 7 and the structural plasticity of brain circuits. *Front Behav Neurosci* **2014**, *8*, 318.
36. Lippiello, P.; Hoxha, E.; Speranza, L.; Volpicelli, F.; Ferraro, A.; Leopoldo, M.; Lacivita, E.; Perrone-Capano, C.; Tempia, F.; Miniaci, M.C. The 5-HT7 receptor triggers cerebellar long-term synaptic depression via PKC-MAPK. *Neuropharmacology* **2016**, *101*, 426-438.
37. Blattner, K.M.; Canney, D.J.; Pippin, D.A.; Blass, B.E. Pharmacology and Therapeutic Potential of the 5-HT. *ACS Chem Neurosci* **2019**, *10*, 89-119.
38. Gellynck, E.; Heynink, K.; Andressen, K.W.; Haegeman, G.; Levy, F.O.; Vanhoenacker, P.; Van Craenenbroeck, K. The serotonin 5-HT7 receptors: two decades of research. *Exp Brain Res* **2013**, *230*, 555-568.
39. Errico, M.; Crozier, R.A.; Plummer, M.R.; Cowen, D.S. 5-HT(7) receptors activate the mitogen activated protein kinase extracellular signal related kinase in cultured rat hippocampal neurons. *Neuroscience* **2001**, *102*, 361-367.

40. Leopoldo, M.; Lacivita, E.; Berardi, F.; Perrone, R.; Hedlund, P.B. Serotonin 5-HT₇ receptor agents: Structure-activity relationships and potential therapeutic applications in central nervous system disorders. *Pharmacol Ther* **2011**, *129*, 120-148.
41. Lenglet, S.; Louiset, E.; Delarue, C.; Vaudry, H.; Contesse, V. Activation of 5-HT₇ receptor in rat glomerulosa cells is associated with an increase in adenylyl cyclase activity and calcium influx through T-type calcium channels. *Endocrinology* **2002**, *143*, 1748-1760.
42. Johnson-Farley, N.N.; Kertesy, S.B.; Dubyak, G.R.; Cowen, D.S. Enhanced activation of Akt and extracellular-regulated kinase pathways by simultaneous occupancy of G_q-coupled 5-HT_{2A} receptors and G_s-coupled 5-HT_{7A} receptors in PC12 cells. *J Neurochem* **2005**, *92*, 72-82.
43. Fields, D.P.; Springborn, S.R.; Mitchell, G.S. Spinal 5-HT₇ receptors induce phrenic motor facilitation via EPAC-mTORC1 signaling. *J Neurophysiol* **2015**, *114*, 2015-2022.
44. Kvachnina, E.; Liu, G.; Dityatev, A.; Renner, U.; Dumuis, A.; Richter, D.W.; Dityateva, G.; Schachner, M.; Voyno-Yasenetskaya, T.A.; Ponimaskin, E.G. 5-HT₇ receptor is coupled to G_α subunits of heterotrimeric G12-protein to regulate gene transcription and neuronal morphology. *J Neurosci* **2005**, *25*, 7821-7830.
45. Prasad, S.; Ponimaskin, E.; Zeug, A. Serotonin receptor oligomerization regulates cAMP-based signaling. *J Cell Sci* **2019**, *132*.
46. Hedlund, P.B.; Danielson, P.E.; Thomas, E.A.; Slanina, K.; Carson, M.J.; Sutcliffe, J.G. No hypothermic response to serotonin in 5-HT₇ receptor knockout mice. *Proc Natl Acad Sci U S A* **2003**, *100*, 1375-1380.
47. Di Pilato, P.; Niso, M.; Adriani, W.; Romano, E.; Travaglini, D.; Berardi, F.; Colabufo, N.A.; Perrone, R.; Laviola, G.; Lacivita, E., et al. Selective agonists for serotonin 7 (5-HT₇) receptor and their applications in preclinical models: an overview. *Rev Neurosci* **2014**, *25*, 401-415.
48. Tajiri, M.; Hayata-Takano, A.; Seiriki, K.; Ogata, K.; Hazama, K.; Shintani, N.; Baba, A.; Hashimoto, H. Serotonin 5-HT₇ receptor blockade reverses behavioral abnormalities in PACAP-deficient mice and receptor activation promotes neurite extension in primary embryonic hippocampal neurons: therapeutic implications for psychiatric disorders. *J Mol Neurosci* **2012**, *48*, 473-481.
49. Rojas, P.S.; Neira, D.; Muñoz, M.; Lavandero, S.; Fiedler, J.L. Serotonin (5-HT) regulates neurite outgrowth through 5-HT_{1A} and 5-HT₇ receptors in cultured hippocampal neurons. *J Neurosci Res* **2014**, *92*, 1000-1009.
50. Speranza, L.; Chambery, A.; Di Domenico, M.; Crispino, M.; Severino, V.; Volpicelli, F.; Leopoldo, M.; Bellenchi, G.C.; di Porzio, U.; Perrone-Capano, C. The serotonin receptor 7 promotes neurite outgrowth via ERK and Cdk5 signaling pathways. *Neuropharmacology* **2013**, *67*, 155-167.
51. Lacivita, E.; Podlewska, S.; Speranza, L.; Niso, M.; Satała, G.; Perrone, R.; Perrone-Capano, C.; Bojarski, A.J.; Leopoldo, M. Structural modifications of the serotonin 5-HT₇ receptor agonist N-(4-cyanophenylmethyl)-4-(2-biphenyl)-1-piperazinehexanamide (LP-211) to improve in vitro microsomal stability: A case study. *Eur J Med Chem* **2016**, *120*, 363-379.
52. Speranza, L.; Giuliano, T.; Volpicelli, F.; De Stefano, M.E.; Lombardi, L.; Chambery, A.; Lacivita, E.; Leopoldo, M.; Bellenchi, G.C.; di Porzio, U., et al. Activation of 5-HT₇ receptor stimulates neurite elongation through mTOR, Cdc42 and actin filaments dynamics. *Front Behav Neurosci* **2015**, *9*, 62.
53. Vasefi, M.S.; Kruk, J.S.; Heikkila, J.J.; Beazely, M.A. 5-Hydroxytryptamine type 7 receptor neuroprotection against NMDA-induced excitotoxicity is PDGF β receptor dependent. *J Neurochem* **2013**, *125*, 26-36.
54. Vasefi, M.S.; Yang, K.; Li, J.; Kruk, J.S.; Heikkila, J.J.; Jackson, M.F.; MacDonald, J.F.; Beazely, M.A. Acute 5-HT₇ receptor activation increases NMDA-evoked currents and differentially alters NMDA receptor subunit phosphorylation and trafficking in hippocampal neurons. *Mol Brain* **2013**, *6*, 24.
55. Costa, L.; Spatuzza, M.; D'Antoni, S.; Bonaccorso, C.M.; Trovato, C.; Musumeci, S.A.; Leopoldo, M.; Lacivita, E.; Catania, M.V.; Ciranna, L. Activation of 5-HT₇ serotonin receptors reverses metabotropic glutamate receptor-mediated synaptic plasticity in wild-type and Fmr1 knockout mice, a model of Fragile X syndrome. *Biol Psychiatry* **2012**, *72*, 924-933.
56. Andreetta, F.; Carboni, L.; Grafton, G.; Jeggo, R.; Whyment, A.D.; van den Top, M.; Hoyer, D.; Spanswick, D.; Barnes, N.M. Hippocampal 5-HT₇ receptors signal phosphorylation of the GluA1 subunit to facilitate AMPA receptor mediated-neurotransmission in vitro and in vivo. *Br J Pharmacol* **2016**, *173*, 1438-1451.
57. Roberts, A.J.; Krucker, T.; Levy, C.L.; Slanina, K.A.; Sutcliffe, J.G.; Hedlund, P.B. Mice lacking 5-HT receptors show specific impairments in contextual learning. *Eur J Neurosci* **2004**, *19*, 1913-1922.
58. Kobe, F.; Guseva, D.; Jensen, T.P.; Wirth, A.; Renner, U.; Hess, D.; Müller, M.; Medrihan, L.; Zhang, W.; Zhang, M., et al. 5-HT_{7R}/G12 signaling regulates neuronal morphology and function in an age-dependent manner. *J Neurosci* **2012**, *32*, 2915-2930.
59. Bijata, M.; Labus, J.; Guseva, D.; Stawarski, M.; Butzlaff, M.; Dzwonek, J.; Schneeberg, J.; Böhm, K.; Michaluk, P.; Rusakov, D.A., et al. Synaptic Remodeling Depends on Signaling between Serotonin Receptors and the Extracellular Matrix. *Cell Rep* **2017**, *19*, 1767-1782.

60. Speranza, L.; Labus, J.; Volpicelli, F.; Guseva, D.; Lacivita, E.; Leopoldo, M.; Bellenchi, G.C.; di Porzio, U.; Bijata, M.; Perrone-Capano, C., et al. Serotonin 5-HT7 receptor increases the density of dendritic spines and facilitates synaptogenesis in forebrain neurons. *J Neurochem* **2017**, *141*, 647-661.
61. Crispino, M.; Cefaliello, C.; Kaplan, B.; Giuditta, A. Protein synthesis in nerve terminals and the glia-neuron unit. *Results Probl Cell Differ* **2009**, *48*, 243-267.
62. Crispino, M.; Chun, J.T.; Cefaliello, C.; Perrone Capano, C.; Giuditta, A. Local gene expression in nerve endings. *Dev Neurobiol* **2014**, *74*, 279-291.
63. Holt, C.E.; Martin, K.C.; Schuman, E.M. Local translation in neurons: visualization and function. *Nat Struct Mol Biol* **2019**, *26*, 557-566.
64. Hübener, M.; Bonhoeffer, T. Neuronal plasticity: beyond the critical period. *Cell* **2014**, *159*, 727-737.
65. Nativio, P.; Zoratto, F.; Romano, E.; Lacivita, E.; Leopoldo, M.; Pascale, E.; Passarelli, F.; Laviola, G.; Adriani, W. Stimulation of 5-HT7 receptor during adolescence determines its persistent upregulation in adult rat forebrain areas. *Synapse* **2015**, *69*, 533-542.
66. Leo, D.; Adriani, W.; Cavaliere, C.; Cirillo, G.; Marco, E.M.; Romano, E.; di Porzio, U.; Papa, M.; Perrone-Capano, C.; Laviola, G. Methylphenidate to adolescent rats drives enduring changes of accumbal Htr7 expression: implications for impulsive behavior and neuronal morphology. *Genes Brain Behav* **2009**, *8*, 356-368.
67. Canese, R.; Zoratto, F.; Altabella, L.; Porcari, P.; Mercurio, L.; de Pasquale, F.; Butti, E.; Martino, G.; Lacivita, E.; Leopoldo, M., et al. Persistent modification of forebrain networks and metabolism in rats following adolescent exposure to a 5-HT7 receptor agonist. *Psychopharmacology (Berl)* **2015**, *232*, 75-89.
68. Bosch, M.; Hayashi, Y. Structural plasticity of dendritic spines. *Curr Opin Neurobiol* **2012**, *22*, 383-388.
69. Chang, J.Y.; Parra-Bueno, P.; Laviv, T.; Szatmari, E.M.; Lee, S.R.; Yasuda, R. CaMKII Autophosphorylation Is Necessary for Optimal Integration of Ca. *Neuron* **2017**, *94*, 800-808.e804.
70. Lai, K.O.; Ip, N.Y. Structural plasticity of dendritic spines: the underlying mechanisms and its dysregulation in brain disorders. *Biochim Biophys Acta* **2013**, *1832*, 2257-2263.
71. Costa, L.; Sardone, L.M.; Bonaccorso, C.M.; D'Antoni, S.; Spatuzza, M.; Gulisano, W.; Tropea, M.R.; Puzzo, D.; Leopoldo, M.; Lacivita, E., et al. Activation of Serotonin 5-HT. *Front Mol Neurosci* **2018**, *11*, 353.
72. Beaudet, G.; Jozet-Alves, C.; Asselot, R.; Schumann-Bard, P.; Freret, T.; Boulouard, M.; Paizanis, E. Deletion of the serotonin receptor type 7 disrupts the acquisition of allocentric but not egocentric navigation strategies in mice. *Behav Brain Res* **2017**, *320*, 179-185.
73. Beaudet, G.; Bouet, V.; Jozet-Alves, C.; Schumann-Bard, P.; Dauphin, F.; Paizanis, E.; Boulouard, M.; Freret, T. Spatial memory deficit across aging: current insights of the role of 5-HT7 receptors. *Front Behav Neurosci* **2014**, *8*, 448.
74. Duncan, M.J.; Smith, J.T.; Franklin, K.M. Time of day but not aging regulates 5-HT. *Neurosci Lett* **2018**, *662*, 306-311.
75. Freret, T.; Paizanis, E.; Beaudet, G.; Gusmao-Montaigne, A.; Nee, G.; Dauphin, F.; Bouet, V.; Boulouard, M. Modulation of 5-HT7 receptor: effect on object recognition performances in mice. *Psychopharmacology (Berl)* **2014**, *231*, 393-400.
76. Meneses, A. 5-HT7 receptor stimulation and blockade: a therapeutic paradox about memory formation and amnesia. *Front Behav Neurosci* **2014**, *8*, 207.
77. Zareifopoulos, N.; Papatheodoropoulos, C. Effects of 5-HT-7 receptor ligands on memory and cognition. *Neurobiol Learn Mem* **2016**, *136*, 204-209.
78. D'Angelo, E. Physiology of the cerebellum. *Handb Clin Neurol* **2018**, *154*, 85-108.
79. Nikiforuk, A.; Hołuj, M.; Potasiewicz, A.; Popik, P. Effects of the selective 5-HT7 receptor antagonist SB-269970 on premature responding in the five-choice serial reaction time test in rats. *Behav Brain Res* **2015**, *289*, 149-156.
80. Okada, M.; Fukuyama, K.; Nakano, T.; Ueda, Y. Pharmacological Discrimination of Effects of MK801 on Thalamocortical, Mesothalamic, and Mesocortical Transmissions. *Biomolecules* **2019**, *9*.
81. Okada, M.; Fukuyama, K.; Okubo, R.; Shiroyama, T.; Ueda, Y. Lurasidone Sub-Chronically Activates Serotonergic Transmission via Desensitization of 5-HT1A and 5-HT7 Receptors in Dorsal Raphe Nucleus. *Pharmaceuticals (Basel)* **2019**, *12*.
82. Ikeda, M.; Iwata, N.; Kitajima, T.; Suzuki, T.; Yamanouchi, Y.; Kinoshita, Y.; Ozaki, N. Positive association of the serotonin 5-HT7 receptor gene with schizophrenia in a Japanese population. *Neuropsychopharmacology* **2006**, *31*, 866-871.
83. Wei, Y.B.; McCarthy, M.; Ren, H.; Carrillo-Roa, T.; Shekhtman, T.; DeModena, A.; Liu, J.J.; Leckband, S.G.; Mors, O.; Rietschel, M., et al. A functional variant in the serotonin receptor 7 gene (HTR7), rs7905446, is associated with good response to SSRIs in bipolar and unipolar depression. *Mol Psychiatry* **2019**.

84. Nelson, R.J.; Young, K.A. Behavior in mice with targeted disruption of single genes. *Neurosci Biobehav Rev* **1998**, *22*, 453-462.
85. Maxwell, J.; Gleason, S.D.; Falcone, J.; Svensson, K.; Balcer, O.M.; Li, X.; Witkin, J.M. Effects of 5-HT. *Behav Brain Res* **2019**, *359*, 467-473.
86. Balcer, O.M.; Seager, M.A.; Gleason, S.D.; Li, X.; Rasmussen, K.; Maxwell, J.K.; Nomikos, G.; Degroot, A.; Witkin, J.M. Evaluation of 5-HT. *Behav Brain Res* **2019**, *360*, 270-278.
87. Sztainberg, Y.; Zoghbi, H.Y. Lessons learned from studying syndromic autism spectrum disorders. *Nat Neurosci* **2016**, *19*, 1408-1417.
88. Muller, C.L.; Anacker, A.M.J.; Veenstra-VanderWeele, J. The serotonin system in autism spectrum disorder: From biomarker to animal models. *Neuroscience* **2016**, *321*, 24-41.
89. Montgomery, A.K.; Shuffrey, L.C.; Guter, S.J.; Anderson, G.M.; Jacob, S.; Mosconi, M.W.; Sweeney, J.A.; Turner, J.B.; Sutcliffe, J.S.; Cook, E.H., et al. Maternal Serotonin Levels Are Associated With Cognitive Ability and Core Symptoms in Autism Spectrum Disorder. *J Am Acad Child Adolesc Psychiatry* **2018**, *57*, 867-875.
90. Garbarino, V.R.; Gilman, T.L.; Daws, L.C.; Gould, G.G. Extreme enhancement or depletion of serotonin transporter function and serotonin availability in autism spectrum disorder. *Pharmacol Res* **2019**, *140*, 85-99.
91. Chugani, D.C.; Chugani, H.T.; Wiznitzer, M.; Parikh, S.; Evans, P.A.; Hansen, R.L.; Nass, R.; Janisse, J.J.; Dixon-Thomas, P.; Behen, M., et al. Efficacy of Low-Dose Buspirone for Restricted and Repetitive Behavior in Young Children with Autism Spectrum Disorder: A Randomized Trial. *J Pediatr* **2016**, *170*, 45-53.e41-44.
92. Amodeo, D.A.; Cuevas, L.; Dunn, J.T.; Sweeney, J.A.; Ragozzino, M.E. The adenosine A. *Autism Res* **2018**, *11*, 223-233.
93. Amodeo, D.A.; Rivera, E.; Dunn, J.T.; Ragozzino, M.E. M100907 attenuates elevated grooming behavior in the BTBR mouse. *Behav Brain Res* **2016**, *313*, 67-70.
94. de Bruin, E.I.; Graham, J.H.; Louwse, A.; Huizink, A.C. Mild dermatoglyphic deviations in adolescents with autism spectrum disorders and average intellectual abilities as compared to typically developing boys. *Autism Res Treat* **2014**, *2014*, 968134.
95. Ghanizadeh, A.; Sahraeizadeh, A.; Berk, M. A head-to-head comparison of aripiprazole and risperidone for safety and treating autistic disorders, a randomized double blind clinical trial. *Child Psychiatry Hum Dev* **2014**, *45*, 185-192.
96. De Filippis, B.; Nativio, P.; Fabbri, A.; Ricceri, L.; Adriani, W.; Lacivita, E.; Leopoldo, M.; Passarelli, F.; Fusco, A.; Laviola, G. Pharmacological stimulation of the brain serotonin receptor 7 as a novel therapeutic approach for Rett syndrome. *Neuropsychopharmacology* **2014**, *39*, 2506-2518.
97. De Filippis, B.; Chiodi, V.; Adriani, W.; Lacivita, E.; Mallozzi, C.; Leopoldo, M.; Domenici, M.R.; Fusco, A.; Laviola, G. Long-lasting beneficial effects of central serotonin receptor 7 stimulation in female mice modeling Rett syndrome. *Front Behav Neurosci* **2015**, *9*, 86.
98. De Filippis, B.; Valenti, D.; Chiodi, V.; Ferrante, A.; de Bari, L.; Fiorentini, C.; Domenici, M.R.; Ricceri, L.; Vacca, R.A.; Fabbri, A., et al. Modulation of Rho GTPases rescues brain mitochondrial dysfunction, cognitive deficits and aberrant synaptic plasticity in female mice modeling Rett syndrome. *Eur Neuropsychopharmacol* **2015**, *25*, 889-901.
99. Valenti, D.; de Bari, L.; Vigli, D.; Lacivita, E.; Leopoldo, M.; Laviola, G.; Vacca, R.A.; De Filippis, B. Stimulation of the brain serotonin receptor 7 rescues mitochondrial dysfunction in female mice from two models of Rett syndrome. *Neuropharmacology* **2017**, *121*, 79-88.
100. Vigli, D.; Rusconi, L.; Valenti, D.; La Montanara, P.; Cosentino, L.; Lacivita, E.; Leopoldo, M.; Amendola, E.; Gross, C.; Landsberger, N., et al. Rescue of prepulse inhibition deficit and brain mitochondrial dysfunction by pharmacological stimulation of the central serotonin receptor 7 in a mouse model of CDKL5 Deficiency Disorder. *Neuropharmacology* **2019**, *144*, 104-114.
101. Martin, A.M.; Sun, E.W.; Keating, D. Mechanisms controlling hormone secretion in human gut and its relevance to metabolism. *J Endocrinol* **2019**.
102. Roumier, A.; Bechade, C.; Maroteaux, L. Serotonin and the Immune System. In *Serotonin The Mediator that Spans Evolution* 1st ed.; Pilowsky, P.M.; Elsevier: United States of America, 2019; pp.181-196.
103. Polat, B.; Halici, Z.; Cadirci, E.; Karakus, E.; Bayir, Y.; Albayrak, A.; Unal, D. Liver 5-HT7 receptors: A novel regulator target of fibrosis and inflammation-induced chronic liver injury in vivo and in vitro. *Int Immunopharmacol* **2017**, *43*, 227-235.
104. Jenkins, T.A.; Nguyen, J.C.; Polglaze, K.E.; Bertrand, P.P. Influence of Tryptophan and Serotonin on Mood and Cognition with a Possible Role of the Gut-Brain Axis. *Nutrients* **2016**, *8*, doi:10.3390/nu8010056.
105. Israelyan, N.; Margolis, K.G. Serotonin as a link between the gut-brain-microbiome axis in autism spectrum disorders. *Pharmacol Res* **2018**, *132*, 1-6.
106. Banskota, S.; Ghia, J.E.; Khan, W.I. Serotonin in the gut: Blessing or a curse. *Biochimie* **2019**, *161*, 56-64.

107. Kim, J.J.; Bridle, B.W.; Ghia, J.E.; Wang, H.; Syed, S.N.; Manocha, M.M.; Rengasamy, P.; Shajib, M.S.; Wan, Y.; Hedlund, P.B., et al. Targeted inhibition of serotonin type 7 (5-HT₇) receptor function modulates immune responses and reduces the severity of intestinal inflammation. *J Immunol* **2013**, *190*, 4795-4804.
108. Guseva, D.; Holst, K.; Kaune, B.; Meier, M.; Keubler, L.; Glage, S.; Buettner, M.; Bleich, A.; Pabst, O.; Bachmann, O., et al. Serotonin 5-HT₇ receptor is critically involved in acute and chronic inflammation of the gastrointestinal tract. *Inflamm Bowel Dis* **2014**, *20*, 1516-1529.
109. Holst, K.; Guseva, D.; Schindler, S.; Sixt, M.; Braun, A.; Chopra, H.; Pabst, O.; Ponimaskin, E. The serotonin receptor 5-HT₇R regulates the morphology and migratory properties of dendritic cells. *J Cell Sci* **2015**, *128*, 2866-2880.
110. Profirovic, J.; Strelakova, E.; Urao, N.; Krbanjevic, A.; Andreeva, A.V.; Varadarajan, S.; Fukai, T.; Hen, R.; Ushio-Fukai, M.; Voyno-Yasenetskaya, T.A. A novel regulator of angiogenesis in endothelial cells: 5-hydroxytryptamine 4 receptor. *Angiogenesis* **2013**, *16*, 15-28.
111. Domínguez-Soto, Á.; Usategui, A.; Casas-Engel, M.L.; Simón-Fuentes, M.; Nieto, C.; Cuevas, V.D.; Vega, M.A.; Luis Pablos, J.; Corbí, Á. Serotonin drives the acquisition of a profibrotic and anti-inflammatory gene profile through the 5-HT₇R-PKA signaling axis. *Sci Rep* **2017**, *7*, 14761.
112. Mahé, C.; Loetscher, E.; Dev, K.K.; Bobirnac, I.; Otten, U.; Schoeffter, P. Serotonin 5-HT₇ receptors coupled to induction of interleukin-6 in human microglial MC-3 cells. *Neuropharmacology* **2005**, *49*, 40-47.
113. León-Ponte, M.; Ahern, G.P.; O'Connell, P.J. Serotonin provides an accessory signal to enhance T-cell activation by signaling through the 5-HT₇ receptor. *Blood* **2007**, *109*, 3139-3146.
114. Ito, M.; Komai, K.; Mise-Omata, S.; Iizuka-Koga, M.; Noguchi, Y.; Kondo, T.; Sakai, R.; Matsuo, K.; Nakayama, T.; Yoshie, O., et al. Brain regulatory T cells suppress astrogliosis and potentiate neurological recovery. *Nature* **2019**, *565*, 246-250.