

*Brief Report***First report and comparative genomics analysis of a *bla*<sub>OXA-244</sub>–harbouring *Escherichia coli* isolate recovered in American continent.**

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1 **Abstract:** The carbapenemase OXA-244 is a derivate of OXA-48, and its detection is very difficult in  
2 laboratories. Here we report the identification and genomic analysis of an *Escherichia coli* isolate  
3 (28Eco12) harbouring the *bla*<sub>OXA-244</sub> gene identified in Colombia, South America.  
4 The 28Eco12 isolate was identified during a retrospective study and it was recovered from a patient  
5 treated in Colombia. The complete nucleotide sequence was established using the PacBio platform.  
6 A comparative genomics analysis with other *bla*<sub>OXA-244</sub>–harbouring *Escherichia coli* strains was  
7 performed. The 28Eco12 isolate belonged to sequence type (ST) 38 and its genome was composed of  
8 two molecules, a chromosome of 5,343,367 bp and a plasmid of 92,027 bp, which belonged to the  
9 incompatibility group IncY and did not harbour resistance genes. The *bla*<sub>OXA-244</sub> gene was  
10 chromosomally-encoded and mobilized by an ISR1-related Tn6237 composite transposon. Notably,  
11 this transposon was inserted and located within a new genomic island. For our knowledge this is  
12 the first report of a *bla*<sub>OXA-244</sub>–harbouring *Escherichia coli* isolate in American continent.  
13 Our results suggest that the introduction of the OXA-244-producing *E. coli* isolate was through  
14 clonal expansion of the ST38 pandemic clone. Other isolates producing OXA-244 could be  
15 circulating silently on the American continent.

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17 **Keywords:** *bla*<sub>OXA-244</sub>; *Escherichia coli*; Carbapenems; Resistance; Colombia.

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## 21 1. Introduction

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23 The WHO has recognized carbapenem-resistant *Enterobacteriaceae* as pathogens with critical  
24 priority for the development of new antibiotics [1]. OXA-244 is a carbapenemase belonging to the  
25 Class D family, is a derivate of OXA-48, and is encoded by the *bla*<sub>OXA-244</sub> gene. Although there are  
26 multiple reports of OXA-48-producing isolates, reports of isolates harbouring OXA-244 are less  
27 frequent, perhaps as their detection is difficult due to their reduced carbapenem activity. The *bla*<sub>OXA-  
28 244</sub> gene was initially described in 2011, within a *Klebsiella pneumoniae* isolate, identified in Spain [2].  
29 It has already been identified in *Escherichia coli* isolates recovered from Germany [3], France [4,5],  
30 the United Kingdom [6], South East Asia [7], and Egypt [5]. The molecular characterization of some  
31 of these *E. coli* isolates have shown that the majority of them belong to sequence type (ST) 38,  
32 although recently other STs have been found (ST361, ST1722, and ST3541) [5]; and contain other  $\beta$ -  
33 lactamases such as TEM, CTX-M and CMY. The *bla*<sub>OXA-244</sub> gene is located in the chromosome within  
34 a truncated Tn1999.2 transposon, which is immersed into an ISR1-based Tn6237 transposon [4,8].  
35 Here, we provide a genomic analysis of an *Escherichia coli* isolate (28Eco12) contained the *bla*<sub>OXA-244</sub>  
36 gene recovered from a patient in Colombia, South America. For our knowledge this is the first  
37 report of a *bla*<sub>OXA-244</sub>-harbouring *Escherichia coli* isolate in American continent.

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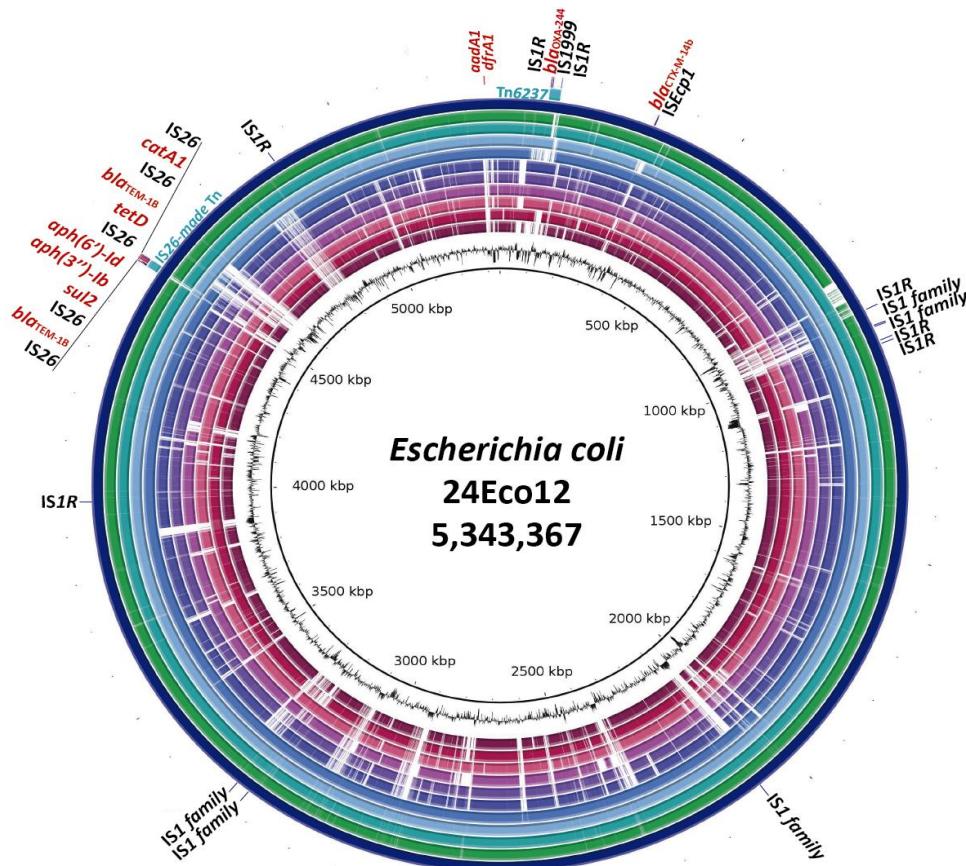
## 39 2. Results

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41 The 28Eco12 isolate was identified from a retrospective study in Bogotá, Colombia (see Materials  
42 and Methods) and we decided to establish its complete genome to determine its resistome and  
43 mobile genetic platform distribution (IS content). The genome was composed of two molecules, a  
44 chromosome of 5,343,367 bp and a plasmid of 92,027 bp, which did not harbour resistance genes  
45 and belonged to the incompatibility group IncY. The resistance genes arsenal of the isolate was  
46 composed of *aph*(3'')-Ib, *aph*(6)-Id, *aaaA1* (aminoglycosides), *bla*<sub>OXA-244</sub>, *bla*<sub>CTX-M-14b</sub>, *bla*<sub>TEM-1b</sub> (beta-  
47 lactams), *catA1* (chloramphenicol), *sul2* (sulphonamides), *dfrA1* (trimethoprim), and *tetD*  
48 (tetracycline) genes, all chromosomally-encoded (Figure 1). The 28Eco12 isolate belonged to ST38  
49 [9]. The "in silico" serotyping of the isolate was O102:H6.

50 Using the complete genome sequence, the 28Eco12 isolate was found to have a close genetic  
51 relationship with the *E. coli* strain 266917\_2 (ST38), described recently in the United Kingdom (90%  
52 coverage, 97% identity, GenBank accession number CP026723.1), which does not contain the *bla*<sub>OXA-  
53 244</sub> gene. The genomic comparative analysis revealed that the *bla*<sub>OXA-244</sub> gene was mobilized by the  
54 Tn6237 transposon, as it has previously been described in *Escherichia coli* strain VAL [4,8]. However,  
55 in the 28Eco12 isolate, the Tn6237 transposon was not inserted within the II<sub>536</sub> pathogenicity island,  
56 as was previously reported to *bla*<sub>OXA-48</sub> [8], but into a new putative genomic island, inserted within  
57 the *tRNA-sec* gene. Its insertion produced a 39 bp direct repeat sequence  
58 (TTCGACTCCTGTGATCTTCCGCCAATTAACATCTTCTGA). This event did not change the  
59 *tRNA-sec* gene sequence (Figure 2).

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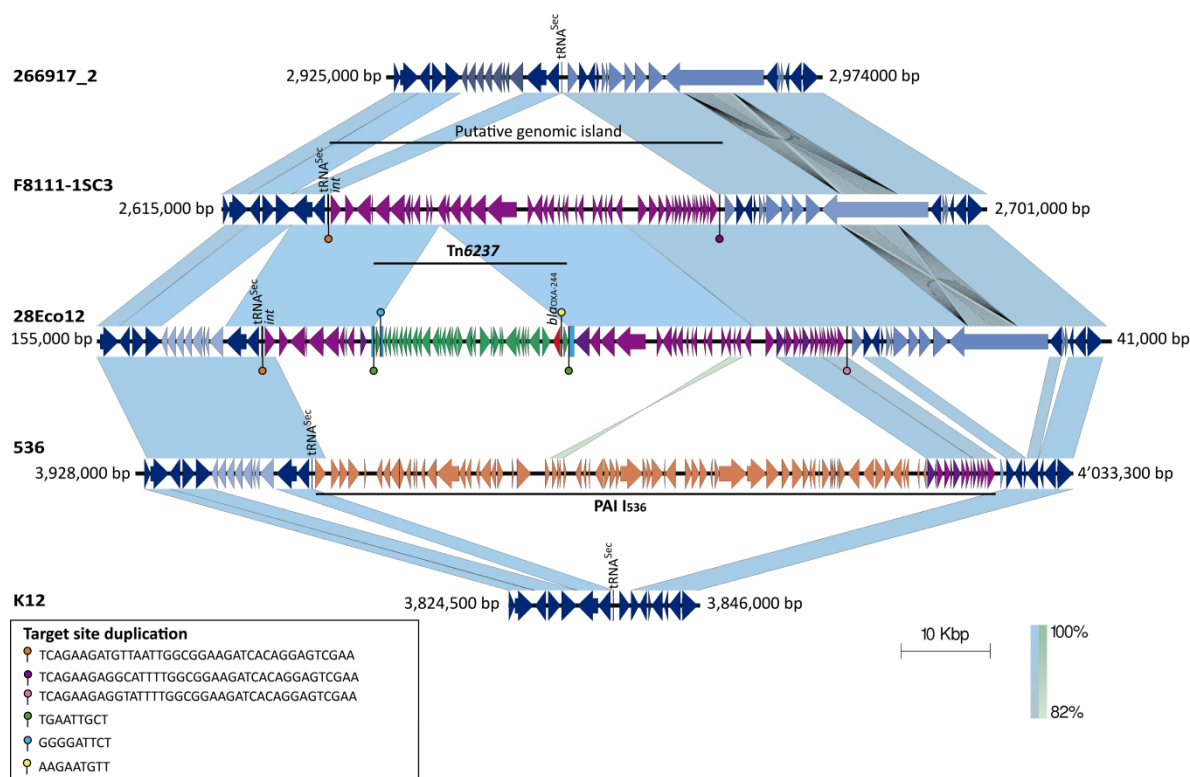


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62 **Figure 1.** BLASTn comparison of the *bla*<sub>OXA-244</sub>-containing *Escherichia coli* chromosomes. The K-12, F8111-1SC3  
 63 and 266917\_2 strains were used as reference. At the more external circle is shown the localization of the  
 64 resistance genes and their putative genetic platforms of mobilization. The positions of the seven identical *ISR1*  
 65 and five *IS1*-family (>89% of identity) sequences are also indicated. The strain positions on the figure are as  
 66 follow (internal to external) (sequence type/serotype): K12 (ST10/O16:H48), F8111-1SC3 (ST182/O169:H41),  
 67 86J1 (ST361/O9:H30), 62D3 (ST1722/O1:H25), 85H4 (ST3541/O53:H18), 73G4 (ST3541/O53:H18), 266917\_2  
 68 (ST38/O51:H30), 35J9 (ST38/O102:H6), 69E6 (ST38/O102:H6), 78B5 (ST38/O102:H6) and 28Eco12  
 69 (ST38/O102:H6).

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71 The putative island was also present in the *bla*<sub>OXA-244</sub>-negative enterotoxigenic *E. coli* F8111-1SC3  
 72 isolate (GenBank accession number NZ\_CP024269.1). Interestingly, the *tRNA-sec* gene is a hot spot  
 73 for DNA insertion because it also serves as the insertion site of the *I*<sub>536</sub> pathogenicity island in the  
 74 uropathogenic strain *E. coli* 536 [10]. These results suggest that the Tn6237 transposon is active, and  
 75 moves to different sites in the *E. coli* chromosome. In addition, the isolate harboured 69 ISs  
 76 belonging to 17 different IS families (Table 1). Some of these present as single copy, partial form or  
 77 multiple copies. The most frequent IS families were *IS1*, *IS200/IS605\_ssgr\_IS200*, and *IS3* with 13, 10,  
 78 and 8 IS copies, respectively. Target Site Duplications (TSD) are signatures of transposition events  
 79 and among the 69 ISs, 25 presented TSDs and none were present within the *E. coli* F8111-1SC3



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81 **Figure 2.** Comparison of the region where the *bla*<sub>OXA-244</sub> gene was inserted within *Escherichia coli* 28Eco12  
 82 isolate. The red arrow corresponds to the *bla*<sub>OXA-244</sub> gene. The mobile genetics elements are shown in different  
 83 colors. The putative genomic island is shown in purple and its insertion within the *tRNA-sec* gene is indicated  
 84 respect to the *E. coli* strain 266917\_2 (GenBank accession number CP026723.1). The blue rectangles correspond  
 85 to the gene where the Tn6237 transposon was inserted (green arrows). The pallets represent the target site  
 86 duplications. The *int* gene that encodes the phage integrase protein is shown. Blue shading between pairs of  
 87 sequences indicates >90% of identity in a window of 400 bp. The scale bar indicates sequence length.

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90 isolate indicating that were inserted by single copy transposition. The TSD pattern analysis also  
 91 revealed the presence of two composite transposons, the Tn6237 (mentioned previously) and a  
 92 15,730 bp IS26-made transposon, which was inserted within a gene that encodes a hypothetical  
 93 protein and mobilizes the *aph(3'')-Ib*, *aph(6)-Id*, *bla*<sub>TEM-1b</sub> (two copies), *catA1*, *sul2*, and *tetD* genes.  
 94 Notably, this IS26 transposon was also inserted within another putative genomic island, which was  
 95 inserted into the *tRNA-leu* gene. The comparative analysis suggested that this IS26 transposon was  
 96 mobilized from a plasmid because it harboured the *repA* gene that corresponds to the  
 97 incompatibility group IncQ-1 and possesses DNA fragments with a high percentage of identity to  
 98 pD90-1 and pEC141 plasmids, which were identified in *mcr-1*-containing *Salmonella enterica* and *E.*  
 99 *coli* strains, respectively [11]. With respect to the other resistance genes, the *bla*<sub>CTX-M</sub> gene was  
 100 mobilized by ISEcp1 and an IS26 remnant, which were inserted within a gene that encodes a  
 101 hypothetical protein.

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103 **Table 1.** Insertion sequences identified in 28Eco12 isolate. Target Site Duplications (TSD) are showed in bold  
 104 and underlined. In addition, the TSD of the composite transposons are colored.

IS family	IS	Position	Right and left flanking sequences		Comments
	IS1R	102025..102792	<b><u>TGAATTGCT</u></b>	AAGAATGTT	Composite transposon harboring the <i>bla<sub>OXA-24</sub></i> gene.
	IS1R	123120..123887	GGGGATTCT	<b><u>TGAATTGCT</u></b>	
	IS1R	936063..936830	<b><u>CAGACAAC</u></b> <b><u>G</u></b>	<b><u>CAGACAAC</u></b> <b><u>G</u></b>	Single IS transposition. IS inserted within a putative prophage
	IS1-like	975280..976060	GTCGCAACC	TACAACGTT	IS inserted within a putative prophage
	IS1-like	977300..978080	GACAATGTC	CAATCTGCT	IS inserted within a putative prophage
	IS1R	1007836..1008603	<b><u>TGCTTTTCT</u></b>	<b><u>TGCTTTTCT</u></b>	Single IS transposition. IS inserted within an intergenic region
IS1	IS1R	1015519..1016286	<b><u>GCCAATTC</u></b> <b><u>G</u></b>	<b><u>GCCAATTC</u></b> <b><u>G</u></b>	Single IS transposition. IS inserted within the <i>cmtB</i> gene
	IS1-like	2087231.. 2087998	CGGTTTTGG	GAAGAGTTC	IS inserted within the <i>hchA</i> gene
	IS1-like	3237236..3237910	-	GAAATCCCC	IS (truncated) inserted within a putative prophage
	IS1-like	3266386..3267153	CTGCAAATC	TACAACCGG	IS inserted within a putative prophage
	IS1R	3972674..3973441	<b><u>CTGCTCCTG</u></b>	<b><u>CTGCTCCTG</u></b>	Single IS transposition. IS inserted within a hypothetical gene
	IS1R	4845817..4846584	GACGGTATT	CGGATGCTG	IS inserted within the <i>adiA</i> gene
	IS1H	5066636..5067399	CCGGTAAAC	CTTCTGATG	IS inserted within an intergenic region
	IS200C	1127230..1127936	<b><u>TTTT</u></b>	<b><u>TTTT</u></b>	Single IS transposition. IS inserted within a T rich region
	IS200C	1690413..1691121	<b><u>TTTT</u></b>	<b><u>TTTT</u></b>	Single IS transposition. IS inserted within a T rich region
	IS200C	2442570..2443280	<b><u>TTAA</u></b>	<b><u>TTAA</u></b>	Single IS transposition. IS inserted within a T rich region
	IS200C	2481694..2482403	<b><u>TTTT</u></b>	<b><u>TTAT</u></b>	Single IS transposition. IS inserted within a T rich region
IS200/ IS605_s sgr_IS2 00	IS200C	2990220..2990930	<b><u>AAAA</u></b>	<b><u>AAAA</u></b>	Single IS transposition. IS inserted within a T rich region
	IS200C	3058643..3059351	<b><u>TAAA</u></b>	<b><u>AAAA</u></b>	Single IS transposition. IS inserted within a T rich region
	IS200C	3060222..3060929	<b><u>AAAA</u></b>	<b><u>AAAA</u></b>	Single IS transposition. IS inserted within a T rich region
	IS200C	3271558..3272271	GCAA	AAAA	IS inserted within a putative prophage
	IS200C	3939865..3940573	<b><u>CAAA</u></b>	<b><u>AAAA</u></b>	Single IS transposition. IS inserted within a T rich region
	IS200C	3994005..3994713	<b><u>AAAA</u></b>	<b><u>AAAA</u></b>	Single IS transposition. IS

					inserted within a T rich region
IS3	IS600	3254256..3255501	CAA	ACA	IS inserted within a genomic island
	ISSd1	949559..950499	CAGTT	-	IS (truncated) inserted within a putative prophage
	ISSd1	3267154..3267978	-	GGT	IS (truncated) inserted within a genomic island
	ISSfl10	951719..952045	-	GTT	IS (truncated) inserted within a putative prophage
	IS3	3259199..3260456	TCAT	TTTA	IS inserted within a genomic island
	IS3	3236998..3237235	-	CTTC	IS (truncated) inserted within a genomic island
	ISEc52	3249338..3250086	-	-	IS (truncated) inserted within a genomic island
	ISEc52	3246586..3247067	-	-	IS (truncated) inserted within a genomic island
ISAs1	ISEc1	369367..369900	-	CCCT	IS (truncated, formerly Rhs - Rearrangement hot spots- element)
	ISEc1	2456311..2456957	GATC	-	IS (truncated, formerly Rhs - Rearrangement hot spots- element)
	ISEc1	3675287..3676199	TGTTGTAG	TCCTTGGC	IS (formerly Rhs -Rearrangement hot spots- element)
	ISEc1	3815490..3816780	GATGTATA	CCTGCTCA	IS (formerly Rhs -Rearrangement hot spots- element)
	ISEc1	4160599..4161889	TTCCTTCC	CACTTCAC	IS (formerly Rhs -Rearrangement hot spots- element)
	ISEc1	5069737..5071026	AGACCAGT	GCATGTCA	IS (formerly Rhs -Rearrangement hot spots- element)
IS6	IS26	4500893..4501712	<u>AAATCATG</u>	ATATCAAG	Composite transposon harboring the <i>bla</i> <sub>TEM-1B</sub> (two copies), <i>catA1</i> , <i>aph(6')-id</i> , <i>aph(3'')-ib</i> , <i>sul2</i> , and <i>tetD</i> genes.
	IS26	4503629..4504448	ATATCGGC	GGTAAATC	
	IS26	4509192..4510011	CCGGCAAT	GTAAGCTG	
	IS26	4513665..4514484	ACCATTG	CGCTGCGG	
	IS26	4515814..4516633	CAACAGGG	<u>AAATCATG</u>	
IS200/ IS605	IS609	3978710..3980457	CTCA	ATAA	IS inserted within the <i>yajl</i> gene
	IS609	4689442..4691189	TGTG	ATAA	IS inserted within a intergenic region
	IS609	2110716..2111379	-	-	IS (truncated) inserted within the <i>yedK</i> gene
	ISEc46	2191062..2192824	TCAT	CTAA	IS inserted within a intergenic region
IS3 ssgr IS150	IS1397	1214273..1215704	<u>TCAA</u>	<u>TCAA</u>	Single IS transposition within a intergenic region
	IS1397	1368490..1369921	<u>TGGC</u>	<u>TGGC</u>	Single IS transposition within a intergenic region

	IS150	259853..261295	<u>AAG</u>	<u>AAG</u>	Single IS transposition within a intergenic region
	IS150	2414087..2415529	<u>GTT</u>	<u>GTT</u>	Single IS transposition. IS inserted within a genomic island
IS3_ssg r_IS2	IS2	937126..938456	GTGGT	TTGTC	IS inserted within a putative prophage
	IS2	966497..967827	CCGCC	ACGGT	IS inserted within a putative prophage
	IS2	2027528..2028858	<u>CCTT</u>	<u>CCTT</u>	Single IS transposition. IS inserted within a genomic island
	IS2	4799912..4800262	AAAAC	-	IS (truncated) inserted within a putative prophage
IS21	IS100Ky p	2015511..2017464	<u>TTTGT</u>	<u>TTTGT</u>	Single IS transposition. IS inserted within a genomic island
	IS100Ky p	3273162..3275115	GTGATAAC	GATAACAT	IS inserted within a genomic island
	IS100Ky p	4582722..4584675	TTCAGATG	AGATGTAT	IS inserted within a putative prophage
IS66	IS682	924827..926816	-	CATGTATC	IS (truncated) inserted within a putative prophage
	ISEc22	923252..924827	ACAGAAGG	-	IS (truncated) inserted within a putative prophage
	ISCro1	946022..948720	<u>TTTTATCT</u>	<u>TTTTATCT</u>	Single IS transposition. IS inserted within a putative prophage
IS3_ssg r_IS51	IS629	570569..571878	<u>ATT</u>	<u>ATT</u>	IS inserted within the <i>acrF</i> gene
	IS1203	971759..973068	GATTACTG	GTAATATC	IS inserted within a putative prophage
ISL3	ISKox3	970324..971101	-	ATGTATCA	IS (truncated) inserted within a putative prophage
	ISEc38	2022594..2024315	AAAAGT	ACTTTT	Single IS transposition. IS inserted within a genomic island (inverted TSD)
IS481	ISErp1	891175..892368	<u>TATAATG</u>	<u>TATAATG</u>	Single IS transposition. IS inserted within a putative prophage
IS30	IS30D	950498..951718	<u>GT</u>	<u>GT</u>	Single IS transposition. IS inserted within a putative prophage
IS4	IS10A	105162..106490	GGCCGAGC	GTGCTGAAC	IS inserted into IS1-composite transposon
IS1380	ISEcp1	326913..330008	<u>TTTA</u>	<u>TTTA</u>	Single IS transposition. IS inserted within a hypothetical gene
IS110	IS5075	1568363..1569689	<u>TT</u>	<u>TT</u>	Single IS transposition. IS inserted within a hypothetical gene

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### 109 3. Discussion

110 In this study, we perform the first report of an *Escherichia coli* isolate carrying the *bla*<sub>OXA-244</sub> gene  
111 in Colombia and the American continent. These *bla*<sub>OXA-244</sub>-positive isolates are less frequent (or  
112 perhaps they circulate but are not detected) by their difficult detection and clonal dissemination.  
113 The multiresistant 28Eco12 isolate harboured an unique plasmid without resistance genes and  
114 genetically related to the plasmids p266917\_2\_02 (88% coverage, 99% identity, GenBank accession  
115 number CP026725.1), p1303\_95 (91% coverage, 99% identity, GenBank accession number  
116 CP009168.1), p1 of *Salmonella enterica* strain ty3-243 (90% coverage, 93% identity, GenBank accession  
117 number LT905089.1), and pCRKP-59-KPC (89% coverage, 94% identity, GenBank accession number  
118 KX928752.1). All resistant genes were chromosomally located and mobilized by active composite  
119 transposons as such Tn6237, which has moved to different sites in the *E. coli* chromosome. In *E. coli*,  
120 the *bla*<sub>OXA-244</sub> gene has been disseminated mainly by ST38 clone in Europe and Asia [3-7]. However,  
121 non-ST38 *E. coli* isolates are starting to appear in other countries, showing some genetic differences  
122 (Figure 1).

123 As it is known that ISs have an important impact on genetic variability, genome structure and  
124 function, and foreign DNA acquisition, we try to decipher the potential of the 28Eco12 isolate to  
125 capture and move more resistance genes through an analysis of the IS content and their TSD and  
126 flanking sequences patterns. Notably, this isolate has suffered at least 69 IS incorporations,  
127 showing a IS massive expansion process [12]; the ISs belonging family IS1 were the most active  
128 with fifteen copies, which four copies probably were recently mobilized as single transposition  
129 events (unique copies) and two mobilized as a composite transposon and responsible of the *bla*<sub>OXA-  
130 244</sub> gene integration (Table 1). In spite of finding five IS26 copies, only two of these were mobilized  
131 as a composite transposon and transported seven resistance genes. A study conducted by He *et al*  
132 reported the IS26 participation in the plasmid reorganization from clinical strains [13]. The high IS  
133 content found in this multiresistant *E. coli* isolate indicates a high likelihood to acquire more  
134 resistance genes.

135 Finally, our institution searched for the presence of the *bla*<sub>OXA-244</sub> gene within other carbapenem-  
136 resistant *E. coli* isolates from 2013 to the present day, but none were positive. Considering the time  
137 of the identification of the isolate, we believe that the *E. coli* isolate could have been acquired in the  
138 remittent institution, suggesting an inter-institution dissemination. No additional information  
139 could be obtained from the other institution.

140

### 141 4. Materials and Methods

142 The 28Eco12 isolate was identified from a retrospective study, conducted to characterize the  
143 molecular mechanisms in carbapenem-resistant *Enterobacteriaceae* isolates, which were recovered  
144 between 2013 and 2017 from a health institution in Bogotá, Colombia. The 28Eco12 isolate was  
145 recovered from a male patient in September 2013, who was transferred from another health  
146 institution in the same city. The patient had suffered multiple traumas cause by a fall from a height  
147 of 20 meters, and required treatment in the intensive care unit for eleven days. The patient was  
148 transferred to our institution, however, on the next day; the patient had fever, dysuria, urethral  
149 pain, leukocytosis, and urethral purulent secretion, suggesting a possible catheter-associated  
150 urinary tract infection. From a urine sample, the carbapenem-resistant *Escherichia coli* isolate



151 28Eco12 was identified, which was also resistant to ampicillin/sulbactam, cefotaxime, ceftriaxone,  
152 cefepime and aztreonam. The Hodge Test was positive and synergy and double-disc tests with  
153 boronic acid and EDTA were negative. The patient was treated with meropenem (2 g every 8 hours)  
154 and colistin (100 mg every 8 hours) and thirteen days later, he was discharged alive. No history of  
155 travel by the patient or his parents was reported.

156 The complete genome sequence of the *bla*<sub>OXA-244</sub>-positive 28Eco12 isolate was obtained using the  
157 PacBio RS II platform (Pacific Biosciences, USA) and assembled through the previously reported  
158 procedure [14]. Briefly, sequencing reads were *de novo* assembled using the HGAP 3 protocol and  
159 manually verified using BWA-MEM (Burrows-Wheeler Aligner with maximal exact matches)[15]  
160 and Tablet v1.15.09.01[16]. Misassembled terminal repeat overlap sequences were identified with  
161 Gepard (Genome Pair Rapid Dotter)[17] and trimmed manually. The genome was annotated using  
162 Prokka v1.11[18], and the interesting regions were manually confirmed using BLASTn and BLASTp  
163 and edited in Artemis [19]. The resistance gene arsenal was identified using ARIBA  
164 (<https://github.com/sanger-pathogens/ariba/wiki>), ResFinder [20], CARD [21] and ARG-ANNOT  
165 databases [22]. The insertion sequences (IS) were found using ISsaga (<http://issaga.biotoul.fr/>) and  
166 their flanking sequences were manually determined.

167 The study was approved by the ethics committee of the Shaio Clinic. The 28Eco12 complete  
168 genome sequenced in this study is available in the DDBJ/EMBL/GenBank public databases under  
169 the accession numbers CP038505.1 and CP038506.1.

## 170 5. Conclusion

171 Our results suggest that the introduction of the OXA-244-producing *E. coli* isolate was through  
172 clonal expansion of the ST38 pandemic clone. In addition, isolates producing OXA-244 could be  
173 circulating since 2013 on the American continent and have not been identified, perhaps due to their  
174 very low frequency, very difficult detection, and weakness in antimicrobial resistance surveillance  
175 programs in some countries (such as Colombia). It is necessary to strengthen the surveillance of  
176 last-line antibiotic resistance, and to move towards the implementation of molecular tools for the  
177 detection of resistance genes in clinical settings.

178

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185

## 186 Author Contributions

187 JEP, IGBM and NVG designed research; IGBM, DFJM and ITM, identified the isolate; performed  
188 microbiological analysis, and interpreted the clinical characteristics of the patient; DA, RAMO, and ZLCR  
189 performed the molecular analysis and genome sequencing; DA, RAMO, JEP and ZLCR performed the  
190 bioinformatics analysis; DA, IGBM, RAMO, NVG and JEP interpreted the data; DA, IGBM and JEP wrote the  
191 paper.

192

193 **Conflicts of Interest**

194 None to declare.

195

196 **References**

- 197 1. Tacconelli, E.; Carrara, E.; Savoldi, A.; Harbarth, S.; Mendelson, M.; Monnet, D.L.; Pulcini, C.; Kahlmeter,  
198 G.; Kluytmans, J.; Carmeli, Y., et al. Discovery, research, and development of new antibiotics: the WHO  
199 priority list of antibiotic-resistant bacteria and tuberculosis. *Lancet Infect Dis* **2018**, *18*, 318-327,  
200 doi:10.1016/S1473-3099(17)30753-3.
- 201 2. Oteo, J.; Hernandez, J.M.; Espasa, M.; Fleites, A.; Saez, D.; Bautista, V.; Perez-Vazquez, M.; Fernandez-  
202 Garcia, M.D.; Delgado-Iribarren, A.; Sanchez-Romero, I., et al. Emergence of OXA-48-producing *Klebsiella*  
203 *pneumoniae* and the novel carbapenemases OXA-244 and OXA-245 in Spain. *J Antimicrob Chemother* **2013**, *68*,  
204 317-321, doi:10.1093/jac/dks383.
- 205 3. Valenza, G.; Nickel, S.; Pfeifer, Y.; Eller, C.; Krupa, E.; Lehner-Reindl, V.; Holler, C. Extended-spectrum-  
206 beta-lactamase-producing *Escherichia coli* as intestinal colonizers in the German community. *Antimicrob*  
207 *Agents Chemother* **2014**, *58*, 1228-1230, doi:10.1128/AAC.01993-13.
- 208 4. Potron, A.; Poirel, L.; Dortet, L.; Nordmann, P. Characterisation of OXA-244, a chromosomally-encoded  
209 OXA-48-like beta-lactamase from *Escherichia coli*. *Int J Antimicrob Agents* **2016**, *47*, 102-103,  
210 doi:10.1016/j.ijantimicag.2015.10.015.
- 211 5. Hoyos-Mallecot, Y.; Naas, T.; Bonnin, R.A.; Patino, R.; Glaser, P.; Fortineau, N.; Dortet, L. OXA-244-  
212 Producing *Escherichia coli* Isolates, a Challenge for Clinical Microbiology Laboratories. *Antimicrob Agents*  
213 *Chemother* **2017**, *61*, doi:10.1128/AAC.00818-17.
- 214 6. Findlay, J.; Hopkins, K.L.; Loy, R.; Doumith, M.; Meunier, D.; Hill, R.; Pike, R.; Mustafa, N.; Livermore,  
215 D.M.; Woodford, N. OXA-48-like carbapenemases in the UK: an analysis of isolates and cases from 2007 to  
216 2014. *J Antimicrob Chemother* **2017**, *72*, 1340-1349, doi:10.1093/jac/dkx012.
- 217 7. van Hattem, J.M.; Arcilla, M.S.; Bootsma, M.C.; van Genderen, P.J.; Goorhuis, A.; Grobusch, M.P.; Molhoek,  
218 N.; Oude Lashof, A.M.; Schultsz, C.; Stobberingh, E.E., et al. Prolonged carriage and potential onward  
219 transmission of carbapenemase-producing Enterobacteriaceae in Dutch travelers. *Future Microbiol* **2016**, *11*,  
220 857-864, doi:10.2217/fmb.16.18.
- 221 8. Beyrouthy, R.; Robin, F.; Delmas, J.; Gibold, L.; Dalmasso, G.; Dabboussi, F.; Hamze, M.; Bonnet, R. ISIR-  
222 mediated plasticity of IncL/M plasmids leads to the insertion of bla OXA-48 into the *Escherichia coli*  
223 Chromosome. *Antimicrob Agents Chemother* **2014**, *58*, 3785-3790, doi:10.1128/AAC.02669-14.
- 224 9. Wirth, T.; Falush, D.; Lan, R.; Colles, F.; Mensa, P.; Wieler, L.H.; Karch, H.; Reeves, P.R.; Maiden, M.C.;  
225 Ochman, H., et al. Sex and virulence in *Escherichia coli*: an evolutionary perspective. *Mol Microbiol* **2006**, *60*,  
226 1136-1151, doi:10.1111/j.1365-2958.2006.05172.x.
- 227 10. Brzuszkiewicz, E.; Bruggemann, H.; Liesegang, H.; Emmerth, M.; Olschlager, T.; Nagy, G.; Albermann, K.;  
228 Wagner, C.; Buchrieser, C.; Emody, L., et al. How to become a uropathogen: comparative genomic analysis  
229 of extraintestinal pathogenic *Escherichia coli* strains. *Proc Natl Acad Sci U S A* **2006**, *103*, 12879-12884,  
230 doi:10.1073/pnas.0603038103.
- 231 11. Wang, J.; Li, X.; Li, J.; Hurley, D.; Bai, X.; Yu, Z.; Cao, Y.; Wall, E.; Fanning, S.; Bai, L. Complete genetic  
232 analysis of a *Salmonella enterica* serovar Indiana isolate accompanying four plasmids carrying mcr-1, ESBL  
233 and other resistance genes in China. *Vet Microbiol* **2017**, *210*, 142-146, doi:10.1016/j.vetmic.2017.08.024.
- 234 12. Siguier, P.; Gournayre, E.; Chandler, M. Bacterial insertion sequences: their genomic impact and diversity.  
235 *FEMS Microbiol Rev* **2014**, *38*, 865-891, doi:10.1111/1574-6976.12067.
- 236 13. He, S.; Hickman, A.B.; Varani, A.M.; Siguier, P.; Chandler, M.; Dekker, J.P.; Dyda, F. Insertion Sequence  
237 IS26 Reorganizes Plasmids in Clinically Isolated Multidrug-Resistant Bacteria by Replicative Transposition.  
238 *MBio* **2015**, *6*, e00762, doi:10.1128/mBio.00762-15.

- 239 14. Marquez-Ortiz, R.A.; Haggerty, L.; Olarte, N.; Duarte, C.; Garza-Ramos, U.; Silva-Sanchez, J.; Castro, B.E.;  
240 Sim, E.M.; Beltran, M.; Moncada, M.V., et al. Genomic Epidemiology of NDM-1-Encoding Plasmids in  
241 Latin American Clinical Isolates Reveals Insights into the Evolution of Multidrug Resistance. *Genome Biol*  
242 *Evol* **2017**, *9*, 1725-1741, doi:10.1093/gbe/evx115.
- 243 15. Li, H.; Durbin, R. Fast and accurate long-read alignment with Burrows-Wheeler transform. *Bioinformatics*  
244 **2010**, *26*, 589-595, doi:10.1093/bioinformatics/btp698.
- 245 16. Milne, I.; Stephen, G.; Bayer, M.; Cock, P.J.; Pritchard, L.; Cardle, L.; Shaw, P.D.; Marshall, D. Using Tablet  
246 for visual exploration of second-generation sequencing data. *Brief Bioinform* **2013**, *14*, 193-202,  
247 doi:10.1093/bib/bbs012.
- 248 17. Krumsiek, J.; Arnold, R.; Rattei, T. Gepard: a rapid and sensitive tool for creating dotplots on genome scale.  
249 *Bioinformatics* **2007**, *23*, 1026-1028, doi:10.1093/bioinformatics/btm039.
- 250 18. Seemann, T. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* **2014**, btu153.
- 251 19. Rutherford, K.; Parkhill, J.; Crook, J.; Horsnell, T.; Rice, P.; Rajandream, M.A.; Barrell, B. Artemis: sequence  
252 visualization and annotation. *Bioinformatics* **2000**, *16*, 944-945.
- 253 20. Zankari, E.; Hasman, H.; Cosentino, S.; Vestergaard, M.; Rasmussen, S.; Lund, O.; Aarestrup, F.M.; Larsen,  
254 M.V. Identification of acquired antimicrobial resistance genes. *J Antimicrob Chemother* **2012**, *67*, 2640-2644,  
255 doi:10.1093/jac/dks261.
- 256 21. McArthur, A.G.; Waglechner, N.; Nizam, F.; Yan, A.; Azad, M.A.; Baylay, A.J.; Bhullar, K.; Canova, M.J.; De  
257 Pascale, G.; Ejim, L., et al. The comprehensive antibiotic resistance database. *Antimicrob Agents Chemother*  
258 **2013**, *57*, 3348-3357, doi:10.1128/AAC.00419-13.
- 259 22. Gupta, S.K.; Padmanabhan, B.R.; Diene, S.M.; Lopez-Rojas, R.; Kempf, M.; Landraud, L.; Rolain, J.M. ARG-  
260 ANNOT, a new bioinformatic tool to discover antibiotic resistance genes in bacterial genomes. *Antimicrob*  
261 *Agents Chemother* **2014**, *58*, 212-220, doi:10.1128/AAC.01310-13.
- 262