

Current Status of Stem Cell Therapy for Thromboangiitis Obliterans (Buerger's Disease)

Hassan Peyvandi MD^{1#}, Biraja C. Dash PhD^{1#}, Sifon U. Ndon BA¹, Kyle Steven Gabrick MD¹, Athena Alipour Faz MD², John A. Persing MD¹, Henry C. Hsia* MD¹

1. Department of Surgery, Section of Plastic Surgery, Yale University School of Medicine, New Haven, Connecticut, USA
2. Hearing Disorders Research Center, Loghman Hakim Medical Center, Shahid Beheshti University of Medical Sciences, Tehran, Iran

Running title: Cell therapy development in Buerger's disease

Co-first Authors

*Address correspondence to:

Henry C. Hsia, MD
Department of Surgery
Section of Plastic Surgery
Yale School of Medicine
PO Box 208041
New Haven, CT 06520-8041

Phone: 203-785-2571; Fax: 203-785-3714
E-mail: henry.hsia@yale.edu

Abstract

Buerger's disease or Thromboangiitis Obliterans (TAO) is a nonatherosclerotic segmental vascular disease which affects small and medium arteries and veins in the upper and lower extremities. Based on pathological findings, TAO can be considered as a distinct form of vasculitis that is most prevalent in young male smokers. There is no definitive cure for this disease as therapeutic modalities are limited in number and efficacy. Surgical bypass has limited utility and 24% of patients will ultimately require amputation. Recently, studies have shown that therapeutic angiogenesis and immunomodulatory approaches through the delivery of cells to target tissues are potential options for ischemic lesion treatment. In this review, we summarize the current knowledge of TAO treatment and provide an overview of stem cell-based treatment modalities.

Key words: Thromboangiitis Obliterans; Inflammation; Angiogenesis; Immunomodulation; Pluripotent stem cell; Mesenchymal stem cell

Introduction

Thromboangiitis Obliterans (TAO) is caused by vasculitis and inflammation of the peripheral blood vessels leading to impaired blood circulation, coagulation, and critical limb ischemia (CLI) [1]. TAO has a global distribution but is most prevalent in Middle and Far Eastern nationalities. Although the precise etiology still remains unknown [1], it predominantly affects young male and female smokers [2-4][5]. Just less than 5% of cases might be caused by other non-smoking related factors such as frostbite, extremity trauma, or even sympathomimetic drug abuse [3, 4, 6, 7].

TAO is a rare form of vasculitis which is distinguished from other types by cellular inflammatory thrombus formation with relative sparing of the vessel wall. TAO has also shown to contain elevated level of pro- and anti-inflammatory cytokines and autoantibodies [8-10]. While the precise pathophysiology behind TAO is controversial, it is believed to be due to an IL-33-mediated immune response resulting in vascular abnormalities [8, 11]. The clinical pathogenesis of TAO begins with lower extremity pain during physical activity which progresses to pain while resting. Additionally, patients may experience Raynaud's phenomena. Many patients also develop ischemic ulcerations that progress to gangrene [2-4]. Despite considerable advances in treatment options, TAO is still associated with high morbidity [12]. While pharmacological approaches and surgical intervention remain generally palliative, novel therapeutic approaches such as gene and stem cell therapy to promote angiogenesis have been considered promising for the treatment of TAO [13-16]. The chief aim of this paper is to provide an overview of the current treatment modalities of TAO and outline approaches using stem cell therapy that may provide new therapeutic solutions to halt the pathogenesis of TAO.

Conventional Approaches

All patients with TAO are advised to stop smoking and avoid second-hand smoke exposure. However, cessation does not completely prevent disease development and progression [17]. Proper foot care is essential to monitor for and treat ischemic ulceration. Often emollient skin cream is helpful to prevent fissure formation.

Pharmacologic treatment:

Pharmacological treatment of TAO is focused on anticoagulation (Aspirin), vasodilators (calcium channel blockers), systemic anti-inflammatory drugs (prostacyclins analogs) and analgesics. [18, 19] Previous studies have demonstrated that iloprost is effective for analgesia and improved wound healing potential. These properties are superior with intravenous infusion. Additionally, thrombolytic therapy with streptokinase and urokinase have demonstrated utility for the treatment of toe and foot gangrene [20]. It has been also reported treatment with bosentan (endothelin receptor antagonists) could improve healing of the ulcers [21]. Calcium-channel blockers like nifedipine can increase distal blood flow due to peripheral vasodilation and improve circulation to the distal ischemic limb. It also can be beneficial in combination with antibiotics and iloprost [2, 22].

Surgical procedures:

Surgical modalities offer limited efficacy for the treatment of TAO. The absence of distal vascular targets makes surgical revascularization challenging. Endovascular treatment modalities have demonstrated utility lessening the progression of TAO [23]. In the setting of multi-level occlusion the stent puncture technique can help overcome vascular access challenges. [24]. Further, sympathectomy may be useful in relieving pain and promoting the

healing of ulcers in some patients; however, these effects were not consistent [25]. Spinal cord stimulation may also be used for relief or treat pain in these patients [26].

Gene therapy

In 1998, a benchmark publication demonstrated that vascular endothelial growth factor (VEGF) gene transfer shows utility in the treatment of TAO. Isner et al. revealed the feasibility of intramuscular gene transfer of naked plasmid DNA encoding - VEGF165 in six patients affected by TAO. Following this gene transfer, patients began to show marked improvement in healing of ischemic ulcers associated with increased blood flow in affected limbs. Patients also showed improvement in ankle-brachial index (ABI) in addition to demonstrating new vessel growth with magnetic resonance angiography (MRA) and serial contrast angiography [13]. Furthermore, phase I clinical trial data demonstrated the safety of intramuscular injections of plasmid DNA expressing two isoforms of hepatocyte growth factor (HGF) (VM202) for patients with critical limb ischemia (CLI). Following plasmid DNA injection, the median ABI and transcutaneous oxygen pressure (tcPO₂) values showed a positive shift, and patients responded clinically with a reduction in reported pain profile [27]. To demonstrate clinical efficacy of intramuscular plasmid injections, Belch and colleagues have evaluated intramuscular injections of non-viral 1 (NV1) fibroblast growth factor (FGF) in a phase 3 clinical trial in CLI patients. In this study, patients who were not considered suitable for revascularization were randomized to treatment with NV1FGF (naked DNA plasmid with gene encoding FGF1) or placebo. In a one-year follow-up study, the NV1FGF treated group didn't show a significant improvement in major amputation rates and mortality over the placebo group [28]. While advancements in gene therapy show hope for future therapeutic options, stem cell therapy may also play an important role in improving the quality of life in patients affected by TAO.

Stem cell-based approaches

Clinical trials have evaluated the potential benefits of stem cell therapy in CLI [29, 30]. Reported benefits include more rapid angiogenesis, reduced inflammation, increased temperature and perfusion of the ischemic limb, and overall increased healing rates as observed by the size of the wound. Clinically, patients have lower rates of surgical amputation and report lower rates of claudication. These treatment approaches are based on the stem cells' ability to stimulate immunomodulation [8, 31] and formation of new blood vessel formation (angiogenesis) and vessel growth (vasculogenesis) [15, 32, 33]. It has been recently reported cell therapy using mononuclear stem cells (MNCs), endothelial progenitor cells (EPCs), mesenchymal stem cells (MSCs), and pluripotent stem cells (PSCs) may have useful roles in prevention of the progression of disease and reduction in major amputation rates [15, 32].

Mononuclear Stem Cells (MNCs):

MNCs isolated from bone marrow (BM-MNCs) and peripheral blood (PB-MNCs) have widely been used for cardiovascular disease applications [34]. Both BM-MNCs and PB-MNCs consists of heterogenous populations of hematopoietic stem cells, MSCs, and EPCs and their cellular composition vary depending on the purification procedure. The MNC-based cell therapy is popular because of their ease in harvesting and the implantation can be done in short turnaround time [35].

Research studies on bone marrow mononuclear cells (BM-MNCs) implantation have shown that cell therapy could increase tissue angiogenesis, neovascularization, and collateral vessel formation in both experimental models and clinical trials. **(Table 1)** The first clinical therapeutic effect of cell transplantation in angiogenesis process was shown by Tateishi-

Yuyama et al. who evaluated therapeutic angiogenesis for patients with severe peripheral vascular disease and limb ischemia by autologous implantation of BM-MNCs. Their results revealed that use of BM-MNCs is a safe and effective method to achieve therapeutic angiogenesis [36]. In a study by Idei et al., long term clinical outcomes of BMMNC transplantation was assayed in patients with CLI including peripheral arterial disease (PAD) and TAO. Reduction of long-term major amputation risk was observed in patients with PAD who were treated with autologous BM-MNCs. In TAO patients, ABI and TcPO₂ were markedly enhanced at 1 month after cell therapy and these effects remain at a high level during the 3-year follow-up [37].

Moriya et al. have shown that treatment with PB-MNCs improves ischemic symptoms and amputation rates in TAO [38]. It has been demonstrated that some factors such as granulocyte colony stimulating factor (G-CSF), granulocyte/macrophage colony stimulating factor (GM-CSF), VEGF, and estrogen induce EPCs mobilization from BM into PB, and the mobilized EPCs then localize to the neovascularization site and contribute to the repair of damaged vessels [39]. In a phase I/IIa clinical trial, cell therapy outcome was evaluated in patients with PAD or TAO. It has been reported that following intramuscular injection of PB-MNCs containing G-CSF-mobilized CD34⁺ stem cells, ischemic signs and symptoms involving Wong-Baker FACES pain rating scale (WBS), toe brachial pressure index (TBPI), TcPO₂, total or pain-free walking distance, and size of ulcer improved in all patients. Furthermore, no mortality or major amputations were observed in this clinical study [40].

Endothelial Progenitor Cells (EPCs):

The stimulation of angiogenesis is critical to reversing the pathogenesis of TAO. Several factors are involved in the angiogenic process. It has been shown that growth factors,

angiogenic genes, and stem cells including EPCs, are involved in modulation of angiogenesis [41]. It has been hypothesized that stem cell therapy for ischemic limbs could promote vascular angiogenesis by supplying EPCs, cytokines, and angiogenic factors. EPCs refer to the cell population that carries the ability for differentiation into endothelial cells. CD34⁺ or CD133⁺ (AC133⁺) MNCs enriched EPCs can be derived from adult bone marrow (BM) or peripheral blood (PB) [42]. Human cord blood– derived CD133 progenitors implanted into the ischemic hind limbs of mice are capable of being incorporated into the capillary networks and improved neovascularization and vessel perfusion [43].

Mesenchymal Stem Cells (MSCs):

MSCs are multipotent stem cells and can be derived from various sources such as bone marrow, liver, adipose tissue, blood, and liver [44, 45]. As a multipotent stem cell, MSCs can be predominantly differentiated into cells of musculoskeletal and fat lineages and also to endothelial lineage (ref). Besides differentiating into cells of the endothelial lineage, MSCs have further contributed to immunomodulatory and angiogenic response by secreting paracrine factors and cytokines such as VEGF, basic fibroblast growth factor (bFGF), and platelet derived growth factor (PDGF), interleukin-10 (IL-10) [31, 45]. Investigations have been performed examining the potential of MSCs derived from bone marrow (BM-MSCs), umbilical cord blood (UC-MSCs), and adipose tissue (ADSCs).

BM-MSCs have widely been used to treat wounds and/or ischemic tissue due to their immunomodulatory and angiogenic effect [31, 34, 45]. In a current study Martin-Rufino et al., investigated the efficacy of sequential intravenous allogeneic MSC administration in treating TAO instead of usual local intramuscular injections [8]. The idea was to illicit a systemic anti-inflammatory effect in the vasculature and thus to modulate the immune

response. In this single patient clinical study, the patient with TAO and at a risk of amputation was treated with four sequential intravenous infusions of allogeneic BM-MSCs, a total of 3.4×10^8 cells, from a healthy donor. The infusion of BM-MSCs passed the safety issue and showed no allograft rejection. Six months after the infusions the result showed significantly healed foot ulcer accompanied with reduced rest pain. Furthermore, the infusion resulted in the improvement of Walking Impairment Questionnaire scores and quality of life and the patient did not need any amputation sixteen months after the infusion. The success of this therapy can be attributed to the immunomodulatory activity BM-MSCs against IL-33 mediated inflammation in TAO [8, 11].

Studies have revealed that paracrine factors secreted by UC-MSCs stimulate angiogenesis. A preclinical study has shown that UCMSCs therapy is better than BM-MSCs transplantation in improving angiogenesis in an ischemic limb disease (ILD) mouse model. UCMSCs cells can secrete high levels of HGF and upon stimulation with TNF α also produce higher amounts of VEGF compared to the BM-MSCs, which are key elements for angiogenesis process during ischemia [46]. Kim et al have also demonstrated UCMSCs transplantation can produce effective outcomes in TAO. Cell therapy improves ischemic symptoms including alleviation of ischemic rest pain, healing of necrotic skin lesions associated with an increase in the size and density of capillaries [47].

Studies have been recently reported that adipose tissue derived MSCs (ADSCs) can be differentiated into endothelial cells and enhance micro-vascularity and blood flow in CLI animal models. The mechanism by which ADSCs induce therapeutic angiogenesis is through the secretion of angiogenic factors such as VEGF [48-51]. Lee et al. determined the safety and efficacy of multiple intramuscular transplantations of ADSCs are safe and effective

demonstrating improved pain rating scales and pain-free walking distances and limb amputation rates. The collateral vascular network formation was also detected in the affected arteries using angiography [32][52].

Pluripotent Stem Cells (PSCs):

Pluripotent stem cells have the ability to self-renew infinitely and can be any cells of the body. These PSCs can be derived either from inner cell mass of the blastocyst to become embryonic stem cells (ESCs), or from somatic cells by expressing specific transcription factors (e.g. Oct4, Sox-2, Klf-4 and c-myc) to form induced pluripotent stem cells (iPSCs) [53, 54]. The use of iPSCs are advantageous as they avoid ethical concerns associated with ESCs. Both ESCs and iPSCs have already been used to derive vascular cells such as endothelial (EC) and vascular smooth muscle cells (VSMC) to induce neovascularization. In two separate studies Cho et al., [55] and Huang et al. [56] demonstrated efficacy of ESC-derived endothelial cells in inducing neovascularization in animal model of CLI. While intramuscular (IM) injection of ESC-ECs by Cho et al., resulted in improved blood perfusion and limb salvage, the study by Huang et al., compared various routes of cell delivery and found systemic route to be more efficient in improving blood perfusion and neovascularization than IM [55, 56]. In another study, Yamahara et al., used a combination of ESC-derived ECs and VSMCs to treat ischemic limb [57]. The result showed formation of mature vasculature compared to EC or SMC alone. These initial studies have led to the generation of clinical grade ESC-EC product developed using good manufacturing practice to use them to reperfuse ischemic limb in patients [58].

Similar to ESCs, iPSCs have been used to derive functional ECs. An initial study by Rufaihah et al. showed the efficacy of these cells in improving blood perfusion and

neovascularization in an animal model of limb ischemia [59]. In order to further establish PSCs as a functional source of ECs, Lai et al., compared ECs derived from various sources including bone marrow (BM-EC), ESCs, and iPSCs in their ability to improve neovascularization and secretion of paracrine factors [60]. The ESC- and iPSC-EC outperformed BM-EC in their ability to secrete paracrine factors and have similar in vivo efficiency in inducing neovascularization as compared to ECs derived from human umbilical vein. These data further support the feasibility of using human PSC-ECs in developing novel cell therapies for patients with CLI and TAO.

Conclusion

In the past two decades, stem cell-based therapy has demonstrated clinical efficacy in the form of therapeutic angiogenesis in peripheral vascular disease. Use of cell therapy for induction of angiogenesis as well as immunomodulation in the animal model has slowly progressed into human clinical trials. While the angiogenic and immunomodulatory potentials of stem cell-based therapy have been demonstrated in human clinical trials of both CLI and TAO, there are still many challenges. The factors that would determine the success of these cell therapies in near future are i) understanding the mechanism and therapeutic potentials of cell-based therapy; ii) establishing a large-scale and renewable source of autologous cells; iii) developing strategies to improve cell potency in vivo and iv) deciding optimal dose, efficient route of administration and frequency of application. Ultimately, we will also need to understand how the in vivo tissue microenvironmental affect the therapeutic activity.

Study	Year	Patients (n)	Cell Source	Cell Route	Follow up	Main Results	Ref.
Kim, D.I et al.	2006	27	BMMSCs	fenestration of the tibia bone	19.1 ± 3.5 months (range, 12.4 to ± 25 months)	inducing therapeutic angiogenesis	[9]
Heo et al.	2016	37	BMMSCs	IM	11.9 ± 7.2 months	Improvement of TBPI and healing of ischemic wounds, pain relief	[23]
Idei et al.	2011	26	BMMSCs	IM	4.8 years	Increase in ABI, TcPO ₂ In 3years follow up	[37]
Kim, S.W et al.	2006	4	UCBMSCs	IM	1&4 Months	Disappearance of Ischemic rest pain, healing of necrotic skin lesions, Increase in number and size of capillaries	[36]
Lee et al.	2012	15 male CLI patients Including TAO	ADSCs	IM	6 months	therapeutic angiogenesis	[24]
Ra et al.	2017	17	ADSCs	IM	2 year	increase in TWD, PFWD And decrease in rest pain.	[41]

Table 1: Stem cell therapy clinical studies in TAO patients

BMMSC, bone marrow mesenchymal stem cell; UCMSCs, umbilical cord blood derived mesenchymal stem cells; ADSCs, adipose tissue derived mesenchymal stem cells; TBPI, toe brachial pressure index ; TcPO₂, transcutaneous partial oxygen pressure; ABI, ankle-brachial index. TWD, total walking distance; PFWD, pain free walking distance. IM, Intramuscular.

Reference:

- [1] Olin JW. Thromboangiitis obliterans (Buerger's disease). *New England Journal of Medicine*. 2000;343:864-9.
- [2] Vijayakumar A, Tiwari R, Kumar Prabhuswamy V. Thromboangiitis obliterans (Buerger's disease)—current practices. *International journal of inflammation*. 2013;2013.
- [3] Hagen B, Lohse S. Clinical and radiologic aspects of Buerger's disease. *Cardiovascular and interventional radiology*. 1984;7:283-93.
- [4] Juergens J. Thromboangiitis obliterans (Buerger's disease, TAO). *Peripheral vascular diseases*. 1980:467-91.
- [5] Joviliano EE, Dellalibera-Joviliano R, Dalio M, Évora PR, Piccinato CE. Etiopathogenesis, clinical diagnosis and treatment of thromboangiitis obliterans—current practices. *International Journal of Angiology*. 2009;18:119-25.
- [6] Hill G. A rational basis for management of patients with the Buerger syndrome. *British Journal of Surgery*. 1974;61:476-81.
- [7] Busch K. Buerger's disease (thromboangiitis obliterans): clinical features and assessment by colour duplex ultrasound. *Australasian journal of ultrasound in medicine*. 2011;14:18-22.
- [8] Martin-Rufino JD, Lozano FS, Redondo AM, Villaron EM, Rueda R, Fernandez-Samos R, et al. Sequential intravenous allogeneic mesenchymal stromal cells as a potential treatment for thromboangiitis obliterans (Buerger's disease). *Stem Cell Res Ther*. 2018;9:150.
- [9] Klein-Weigel PF, Richter JG. Thromboangiitis obliterans (Buerger's disease). *Vasa*. 2014;43:337-46.
- [10] Klein-Weigel P, Volz TS, Zange L, Richter J. Buerger's disease: providing integrated care. *J Multidiscip Healthc*. 2016;9:511-8.
- [11] Sun XL, Law BY, de Seabra Rodrigues Dias IR, Mok SWF, He YZ, Wong VK. Pathogenesis of thromboangiitis obliterans: Gene polymorphism and immunoregulation of human vascular endothelial cells. *Atherosclerosis*. 2017;265:258-65.
- [12] Cooper LT, Tse TS, Mikhail MA, McBane RD, Stanson AW, Ballman KV. Long-term survival and amputation risk in thromboangiitis obliterans (Buerger's disease). *Journal of the American College of Cardiology*. 2004;44:2410-1.
- [13] Isner JM, Baumgartner I, Rauh G, Schainfeld R, Blair R, Manor O, et al. Treatment of thromboangiitis obliterans (Buerger's disease) by intramuscular gene transfer of vascular endothelial growth factor: preliminary clinical results. *Journal of vascular surgery*. 1998;28:964-75.

- [14] Kim DI, Kim MJ, Joh JH, Shin SW, Do YS, Moon JY, et al. Angiogenesis facilitated by autologous whole bone marrow stem cell transplantation for Buerger's disease. *Stem Cells*. 2006;24:1194-200.
- [15] Durdu S, Akar AR, Arat M, Sancak T, Eren NT, Ozyurda U. Autologous bone-marrow mononuclear cell implantation for patients with Rutherford grade II-III thromboangiitis obliterans. *Journal of vascular surgery*. 2006;44:732-9.
- [16] Kajiguchi M, Kondo T, Izawa H, Kobayashi M, Yamamoto K, Shintani S, et al. Safety and efficacy of autologous progenitor cell transplantation for therapeutic angiogenesis in patients with critical limb ischemia. *Circulation Journal*. 2007;71:196-201.
- [17] Lawrence PF, Lund OI, Jimenez JC, Muttalib R. Substitution of smokeless tobacco for cigarettes in Buerger's disease does not prevent limb loss. *Journal of vascular surgery*. 2008;48:210-2.
- [18] Bozkurt AK, Cengiz K, Arslan C, Mine DY, Oner S, Deniz DB, et al. A stable prostacyclin analogue (iloprost) in the treatment of Buerger's disease: a prospective analysis of 150 patients. *Annals of Thoracic and Cardiovascular Surgery*. 2013;19:120-5.
- [19] Cacione DG, Baptista-Silva J, Macedo CR. Pharmacological treatment for Buerger's disease. *Cochrane Database Syst Rev*. 2016;2.
- [20] Hussein E. Intra-arterial streptokinase as adjuvant therapy for complicated Buerger's disease: early trials. *International surgery*. 1993;78:54-8.
- [21] De Haro J, Bleda S, Acin F. An open-label study on long-term outcomes of bosentan for treating ulcers in thromboangiitis obliterans (Buerger's disease). *International journal of cardiology*. 2014;177:529-31.
- [22] Jorge VC, Araújo AC, Noronha C, Panarra A, Riso N, Riscado MV. Buerger's disease (Thromboangiitis obliterans): a diagnostic challenge. *BMJ case reports*. 2011;2011:bcr0820114621.
- [23] Ryu S-W, Jeon H-J, Cho S-S, Choi R-M, Yoon J-S, Ko H-S, et al. Treatment of digit ulcers in a patient with Buerger's disease by using cervical spinal cord stimulation-a case report. *Korean journal of anesthesiology*. 2013;65:167-71.
- [24] Lee J-H, Ko Y-G, Choi D. Endovascular Treatment of Multilevel Chronic Total Occlusion Using a Stent Puncture Technique in Buerger's Disease. *Korean circulation journal*. 2016;46:417-20.
- [25] Sasajima T, Kubo Y, Inaba M, Goh K, Azuma N. Role of infrainguinal bypass in Buerger's disease: an eighteen-year experience. *European Journal of Vascular and Endovascular Surgery*. 1997;13:186-92.

- [26] Donas KP, Schulte S, Ktenidis K, Horsch S. The role of epidural spinal cord stimulation in the treatment of Buerger's disease. *Journal of vascular surgery*. 2005;41:830-6.
- [27] Henry T, Hirsch A, Goldman J, Wang Y, Lips D, McMillan W, et al. Safety of a non-viral plasmid-encoding dual isoforms of hepatocyte growth factor in critical limb ischemia patients: a phase I study. *Gene therapy*. 2011;18:788-94.
- [28] Belch J, Hiatt WR, Baumgartner I, Driver IV, Nikol S, Norgren L, et al. Effect of fibroblast growth factor NV1FGF on amputation and death: a randomised placebo-controlled trial of gene therapy in critical limb ischaemia. *The Lancet*. 2011;377:1929-37.
- [29] Lee K-B, Kang E-S, Kim A-K, Kim M-H, Do Y-S, Park K-B, et al. Stem cell therapy in patients with thromboangiitis obliterans: assessment of the long-term clinical outcome and analysis of the prognostic factors. *International journal of stem cells*. 2011;4:88.
- [30] Heo S-H, Park Y-S, Kang E-S, Park K-B, Do Y-S, Kang K-S, et al. Early results of clinical application of autologous whole bone marrow stem cell transplantation for critical limb ischemia with Buerger's disease. *Scientific reports*. 2016;6.
- [31] Gao F, Chiu SM, Motan DA, Zhang Z, Chen L, Ji HL, et al. Mesenchymal stem cells and immunomodulation: current status and future prospects. *Cell Death Dis*. 2016;7:e2062.
- [32] Lee HC, An SG, Lee HW, Park J-S, Cha KS, Hong TJ, et al. Safety and effect of adipose tissue-derived stem cell implantation in patients with critical limb ischemia. *Circulation Journal*. 2012;76:1750-60.
- [33] Miyamoto K, Nishigami K, Nagaya N, Akutsu K, Chiku M, Kamei M, et al. Unblinded pilot study of autologous transplantation of bone marrow mononuclear cells in patients with thromboangiitis obliterans. *Circulation*. 2006;114:2679-84.
- [34] Brenes RA, Bear M, Jadowiec C, Goodwin M, Hashim P, Protack CD, et al. Cell-based interventions for therapeutic angiogenesis: review of potential cell sources. *Vascular*. 2012;20:360-8.
- [35] Goumans MJ, Maring JA, Smits AM. A straightforward guide to the basic science behind cardiovascular cell-based therapies. *Heart*. 2014;100:1153-7.
- [36] Tateishi-Yuyama E, Matsubara H, Murohara T, Ikeda U, Shintani S, Masaki H, et al. Therapeutic angiogenesis for patients with limb ischaemia by autologous transplantation of bone-marrow cells: a pilot study and a randomised controlled trial. *The Lancet*. 2002;360:427-35.
- [37] Idei N, Soga J, Hata T, Fujii Y, Fujimura N, Mikami S, et al. Autologous Bone-Marrow Mononuclear Cell Implantation Reduces Long-Term Major Amputation Risk in Patients With Critical Limb Ischemia. *Circulation: Cardiovascular Interventions*. 2011;4:15-25.

- [38] Moriya J, Minamino T, Tateno K, Shimizu N, Kuwabara Y, Sato Y, et al. Long-term outcome of therapeutic neovascularization using peripheral blood mononuclear cells for limb ischemia. *Circulation: Cardiovascular Interventions*. 2009;CIRCINTERVENTIONS. 108.799361.
- [39] Imanishi T, Tsujioka H, Akasaka T. Endothelial progenitor cells dysfunction and senescence: contribution to oxidative stress. *Current cardiology reviews*. 2008;4:275-86.
- [40] Kawamoto A, Katayama M, Handa N, Kinoshita M, Takano H, Horii M, et al. Intramuscular transplantation of G-CSF-mobilized CD34+ cells in patients with critical limb ischemia: a phase I/IIa, multicenter, single-blinded, dose-escalation clinical trial. *Stem Cells*. 2009;27:2857-64.
- [41] Huang NF, Li S. Mesenchymal stem cells for vascular regeneration. 2008.
- [42] Friedrich EB, Walenta K, Scharlau J, Nickenig G, Werner N. CD34-/CD133+/VEGFR-2+ endothelial progenitor cell subpopulation with potent vasoregenerative capacities. *Circulation research*. 2006;98:e20-e5.
- [43] Yang C, Zhang ZH, Li ZJ, Yang RC, Qian GQ, Han ZC. Enhancement of neovascularization with cord blood CD133⁺ cell-derived endothelial progenitor cell transplantation. *THROMBOSIS AND HAEMOSTASIS-STUTTGART*-. 2004;91:1202-12.
- [44] Pittenger MF, Mackay AM, Beck SC, Jaiswal RK, Douglas R, Mosca JD, et al. Multilineage potential of adult human mesenchymal stem cells. *Science*. 1999;284:143-7.
- [45] Galipeau J, Sensebe L. Mesenchymal Stromal Cells: Clinical Challenges and Therapeutic Opportunities. *Cell Stem Cell*. 2018;22:824-33.
- [46] Shetty P, Thakur AM, Ravindran G, Viswanathan C. Directed therapeutic angiogenesis by mesenchymal stem cells from umbilical cord matrix in preclinical model of ischemic limb disease. *Stem cell studies*. 2011;1:16.
- [47] Kim SW, Han H, Chae GT, Lee SH, Bo S, Yoon JH, et al. Successful stem cell therapy using umbilical cord blood-derived multipotent stem cells for Buerger's disease and ischemic limb disease animal model. *Stem cells*. 2006;24:1620-6.
- [48] Kondo K, Shintani S, Shibata R, Murakami H, Murakami R, Imaizumi M, et al. Implantation of adipose-derived regenerative cells enhances ischemia-induced angiogenesis. *Arteriosclerosis, thrombosis, and vascular biology*. 2009;29:61-6.
- [49] Sumi M, Sata M, Toya N, Yanaga K, Ohki T, Nagai R. Transplantation of adipose stromal cells, but not mature adipocytes, augments ischemia-induced angiogenesis. *Life sciences*. 2007;80:559-65.

- [50] Traktuev DO, Merfeld-Clauss S, Li J, Kolonin M, Arap W, Pasqualini R, et al. A population of multipotent CD34-positive adipose stromal cells share pericyte and mesenchymal surface markers, reside in a periendothelial location, and stabilize endothelial networks. *Circulation research*. 2008;102:77-85.
- [51] Nakagami H, Maeda K, Morishita R, Iguchi S, Nishikawa T, Takami Y, et al. Novel autologous cell therapy in ischemic limb disease through growth factor secretion by cultured adipose tissue-derived stromal cells. *Arteriosclerosis, thrombosis, and vascular biology*. 2005;25:2542-7.
- [52] Ra JC, Jeong EC, Kang SK, Lee SJ, Choi KH. A Prospective, Nonrandomized, no Placebo-Controlled, Phase I/II Clinical Trial Assessing the Safety and Efficacy of Intramuscular Injection of Autologous Adipose Tissue-Derived Mesenchymal Stem Cells in Patients with Severe Buerger's Disease. *Cell Medicine*. 2017;9:87-102.
- [53] Thomson JA, Itskovitz-Eldor J, Shapiro SS, Waknitz MA, Swiergiel JJ, Marshall VS, et al. Embryonic stem cell lines derived from human blastocysts. *Science*. 1998;282:1145-7.
- [54] Takahashi K, Tanabe K, Ohnuki M, Narita M, Ichisaka T, Tomoda K, et al. Induction of pluripotent stem cells from adult human fibroblasts by defined factors. *Cell*. 2007;131:861-72.
- [55] Cho SW, Moon SH, Lee SH, Kang SW, Kim J, Lim JM, et al. Improvement of postnatal neovascularization by human embryonic stem cell derived endothelial-like cell transplantation in a mouse model of hindlimb ischemia. *Circulation*. 2007;116:2409-19.
- [56] Huang NF, Niiyama H, Peter C, De A, Natkunam Y, Fleissner F, et al. Embryonic stem cell-derived endothelial cells engraft into the ischemic hindlimb and restore perfusion. *Arterioscler Thromb Vasc Biol*. 2010;30:984-91.
- [57] Yamahara K, Sone M, Itoh H, Yamashita JK, Yurugi-Kobayashi T, Homma K, et al. Augmentation of neovascularization [corrected] in hindlimb ischemia by combined transplantation of human embryonic stem cells-derived endothelial and mural cells. *PLoS One*. 2008;3:e1666.
- [58] MacAskill MG, Saif J, Condie A, Jansen MA, MacGillivray TJ, Tavares AAS, et al. Robust Revascularization in Models of Limb Ischemia Using a Clinically Translatable Human Stem Cell-Derived Endothelial Cell Product. *Mol Ther*. 2018;26:1669-84.
- [59] Rufaihah AJ, Huang NF, Jame S, Lee JC, Nguyen HN, Byers B, et al. Endothelial cells derived from human iPSCs increase capillary density and improve perfusion in a mouse model of peripheral arterial disease. *Arterioscler Thromb Vasc Biol*. 2011;31:e72-9.

[60] Lai WH, Ho JC, Chan YC, Ng JH, Au KW, Wong LY, et al. Attenuation of hind-limb ischemia in mice with endothelial-like cells derived from different sources of human stem cells. *PLoS One*. 2013;8:e57876.